

# Location–scale models in ecology: heteroscedasticity in continuous, count and proportion data

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## 1 **Abstract**

2 Ecological data seldom meet the assumption of constant variance. Yet patterns of heteroscedasticity often  
3 reflect biologically meaningful variation, such as differences in plasticity or variable responses to  
4 environmental stresses. However, most studies model only the mean, treating variance as statistical noise.  
5 Here, we describe location–scale regression modeling, which estimates mean (location) as well as variance  
6 (scale) coefficients. We introduce three increasingly flexible formulations: (1) fixed-effect location–scale  
7 models, (2) models with random effects on the mean, and (3) double-hierarchical models with random  
8 effects on both mean and variance. We extend location–scale models from Gaussian to non-Gaussian data,  
9 including over-dispersed counts, proportions, and zero-inflated outcomes, features common to ecological  
10 datasets. Beyond overdispersion, we address underdispersion in count data and one-inflation in continuous  
11 proportions, providing a flexible framework for complex variance structures. We show that location–scale  
12 models can uncover informative variance patterns with minimal additional code. To support  
13 implementation, we provide an online tutorial ([link](#)), model selection workflow, and diagnostic guidance.  
14 Finally, we refer to new frontiers including multivariate, meta-analytic, phylogenetic, and  
15 location-scale-*shape* models. By treating variance as a biological response, instead of a nuisance,  
16 location–scale models enrich our understanding of organism and ecosystem dynamics in a changing world.

## 17 **1 Introduction**

18 Ecologists and evolutionary biologists strive to explain and account for variation in nature; this is usually  
19 done by statistically modeling target traits or measurements with hypothesized causal factors (e.g., a  
20 particular environmental factor accounts for 8% of the variance). In contrast, they rarely test whether  
21 variation changes across an environmental gradient or between groups (Cleasby and Nakagawa, 2011).

22 Although ecological data often exhibit non-constant variance, this variation is commonly considered a mere  
23 nuisance that violates the model's assumption of homogeneity (i.e., homoscedasticity). In reality, patterns  
24 in variance, or heteroscedasticity, can signal ecological, evolutionary, and environmental processes. For  
25 example, environmental stress (e.g., temperature increases) can not only change the mean but can also  
26 generate more variance in organismal responses (e.g., Buckley and Huey, 2016; O'Dea et al., 2016). On the  
27 other hand, plasticity, such as learning, can canalize variability because most individuals uniformly reach  
28 the behavioral optimum (e.g., Baldwin, 1896; Crispo, 2007).

29 More than a decade ago, Cleasby and Nakagawa (2011) surveyed and reported that over 95% of published  
30 studies in behavioral ecology ignored heteroscedasticity. Such neglect can yield incorrect standard errors  
31 (SE) of regression coefficients (e.g., Type I error) and, critically, overlook biological insights in dispersion  
32 patterns. Therefore, they recommended two practical solutions. First, they suggested the use of  
33 heteroscedasticity-consistent ("sandwich") estimators of SE, which resolve the statistical issues such as  
34 inflated Type I error (Hayes and Cai, 2007). Second, one can model different residual variances for different  
35 groups or across a continuous predictor (i.e., heteroscedasticity). This approach, however, does not directly  
36 provide inferential statistics – whether changes in variance are statistically significant or not. In their paper,  
37 Cleasby and Nakagawa (2011) neglected the third option: location–scale regression modeling, which  
38 provides statistical inference on both mean (*location*) and variance (*scale*, also known as *dispersion*) and  
39 thus resolves all issues at once. Statistically, location–scale models remove bias in SE and test statistics  
40 under heteroscedasticity (Carroll and Ruppert, 1988; Zuur et al., 2009). Biologically, these models can

41 reveal when and how both mean and variance respond to environmental and other drivers.

42 The most flexible forms of location-scale models are double-hierarchical with random effects in both mean  
43 (*location*) and variance (*scale*)(Lee and Nelder, 1996, 2006; Rönnegård and Lee, 2013). However, these  
44 models are computationally complex and require Bayesian implementation, which may have hindered  
45 wider adoption. However, simpler location-scale models, which can only include random effects in the  
46 location part, are straightforward to implement in widely used statistical software. For example, these  
47 variants can be implemented readily in `glmmTMB` (Brooks et al., 2017) with minimal coding.

48 Therefore, we aim to reintroduce the utility of location–scale regression models. To facilitate broader use,  
49 we focus on two simpler, practical formulations sufficient for many applications. In the following sections,  
50 we first introduce location–scale models with only fixed effects on both mean (*location*) and variance  
51 (*scale*) (Model 1). Next, we extend these to include random effects on the *location* part (Model 2) and, for  
52 completeness, describe the double-hierarchical framework with random effects on both *location* and *scale*  
53 (Model 3). We then expand these models (mainly Model 2) to non-Gaussian responses, namely count and  
54 proportion data; although such data are common, modeling over-dispersion of count and proportion seems  
55 to be rare in ecology, evolution, and environmental sciences (cf., Bolker et al., 2009). These non-Gaussian  
56 location-scale models can handle zero-inflation, and we refer to the issues of under-dispersion and  
57 one-inflation. We provide a range of examples illustrating biological insights obtained from location-scale  
58 models with both frequentist and Bayesian implementations using `glmmTMB` and `brms` (Bürkner, 2017),  
59 respectively (see the online tutorial ([link](#))). We also suggest a practical workflow to guide model selection.

60 Finally, we discuss broader applications of location-scale models (e.g., meta-analytic location-scale  
61 models; Nakagawa et al., 2025) and related advanced models, which are potentially even more flexible and  
62 biologically informative (Rigby and Stasinopoulos, 2005).

## 63 2 From simple to location-scale regression (Model 1)

### 64 2.1 Model and motivation

65 We begin with the familiar simple regression models (only with fixed effects), where we assume constant  
66 residual variance as well as data independence:

$$y_i = \beta_0 + \sum_{k=1}^K \beta_k x_{ik} + e_i, \quad (1)$$

$$e_i \sim \mathcal{N}(0, \sigma^2), \quad (2)$$

67 where  $y_i$  is the response for observation  $i$ ,  $x_{ik}$  ( $k = 1, \dots, K$ ) are the fixed covariates (predictors),  
68  $\{\beta_0, \beta_1, \dots, \beta_K\}$  are the regression coefficients, and the residual  $e_i$  is normally (Gaussian) distributed with  
69 mean zero and variance  $\sigma^2$ . Note that the predictor  $x_{ik}$  can be either a continuous or categorical variable.  
70 More accurately, for the latter case, when a categorical predictor has  $H$  levels, it becomes  $H - 1$  ‘dummy’  
71 variables or predictors. That is, a categorical variable becomes  $(H - 1)$  binary variables in the model, and  
72 corresponding regression coefficients represent contrasts (differences) between a reference level (the  
73 intercept  $\beta_0$ ) and another level.

74 Equivalently, we can write the model in its distributional form:

$$y_i \sim \mathcal{N}(\mu_i, \sigma^2), \quad (3)$$

$$\mu_i = \beta_0 + \sum_{k=1}^K \beta_k x_{ik}, \quad (4)$$

75 where  $\mu_i$  denotes the expected value of  $y_i$  given the covariates, and  $\sigma^2$  remains the constant variance.

76 This basic regression treats any heteroscedasticity as a nuisance. To turn it into biological/ecological  
77 signals, we allow the residual standard deviation to vary with predictors. The location–scale regression then  
78 comprises two linked submodels which can be written as (Model 1; Jorgensen, 1997; Lee et al., 2006;

79 Cleasby et al., 2015):

$$y_i \sim \mathcal{N}(\mu_i, \sigma_i^2), \quad (5)$$

$$\mu_i = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ik}, \quad (\text{location submodel}) \quad (6)$$

$$\ln(\sigma_i) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ik}, \quad (\text{scale submodel}) \quad (7)$$

80 where  $\mu_i$  is its expectation, modeled by the location submodel coefficients  $\beta^{(l)}$  and covariates  $x_{ik}$ , and  $\sigma_i$  is  
 81 the residual standard deviation, modeled on the log (ln) scale by the scale submodel coefficients  $\beta^{(s)}$  and  
 82 the same covariates.

83 This fixed effects location-scale regression, by linking predictors to both the mean and the ln-variance,  
 84 allows us to test if an environmental gradient or experimental treatment shifts not just the average response,  
 85 but also its individual variability. In other words, if a predictor ( $x_{ik}$ ) influences the mean, its corresponding  
 86 regression coefficient ( $\beta_k^{(l)} \neq 0$ ) will be non-zero (significant). If a predictor influences variance, the  
 87 associated regression coefficient for the scale component, represented as ( $\beta_p^{(s)} \neq 0$ ), will also be non-zero.  
 88 This suggests that the heterogeneity in the data varies in relation to that predictor, a phenomenon referred to  
 89 as heteroscedasticity. Translating a variance signals into regression coefficients formalizes heterogeneity  
 90 analysis and makes it accessible to researchers already familiar with interpreting regression coefficients for  
 91 the location part (Fig. 1).

92 The syntax for writing location-scale models in R builds off familiar modeling syntax in R. To fit a  
 93 (location-only) regression model on the relationship between the location (mean) of  $y$  by  $x$ , we would write  
 94 the following:

```
95 library("glmmTMB")
96
97 location_model <- glmmTMB(y~x, data = dt)
98
```

99 To explicitly model the 'scale' as well as 'location', we simply add the same formula (without the response

100 variable) to the `dispformula` argument:  
 101

```
102 location_scale_model <- glmmTMB(y~x, dispformula = ~x, data = dt)
```

104 This second model returns two regression tables, one (referred to as the `Conditional` by `glmmTMB`)  
 105 describes the relationship between `x` and mean `y`, while the second table (referred to as `Dispersion`)  
 106 describes the relationship between `x` and the variance of `y`.

## 107 2.2 Illustrative example

108 In the following illustrative examples, we report representative model results using the R packages  
 109 `glmmTMB` and/or `brms`, selected based on model type and functionality. Full model specifications, code, and  
 110 detailed explanations of datasets and interpretations are available in our online tutorial ([link](#)).

111 We analyzed whether early-life food supplementation had sex-specific effects on body size variability, using  
 112 adult tarsus length as an indicator, in a wild population of house sparrows (*Passer domesticus*) on Lundy  
 113 Island, England (Cleasby and Nakagawa, 2011). The model's location component showed no significant  
 114 effect of sex, treatment, or their interaction on mean adult tarsus length. However, the scale (dispersion)  
 115 component revealed a significant negative interaction between sex and treatment (`glmmTMB`:  $\beta_{[interaction]}^{(s)} =$   
 116  $-0.95$ , 95% CI =  $-1.66$  to  $-0.24$ ). This indicates a significant reduction in adult tarsus length variance among  
 117 supplemented males. Neither treatment nor sex alone significantly influenced variance. This suggests  
 118 early-life food supplementation can canalize trait development, leading to more uniform adult male  
 119 morphology under favorable nutritional conditions.

## 120 3 Adding random effects in the location part only (Model 2)

### 121 3.1 Model and motivation

122 Ecological and environmental datasets often violate both the homoscedasticity and non-independence  
 123 assumptions. The latter is common due to clustered or grouped data, such as multiple measurements *per*  
 124 site or individual. Consequently, 'mixed-effects' models are widely used in ecology and evolution, as they

125 incorporate both fixed and random effects to model these clustering and grouping structures (Bolker et al.,  
126 2009; Nakagawa and Schielzeth, 2013).

127 Introducing a random effect (intercept in the location submodel) allows each group  $j$  to have a  
128 group-specific mean, while keeping the scale model fixed-effects only. Such models can be written as  
129 (Model 2; Jorgensen, 1997; Lee et al., 2006; Cleasby et al., 2015):

$$y_{ij} \sim \mathcal{N}(\mu_{ij}, \sigma_{ij}^2), \quad (8)$$

$$\mu_{ij} = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (9)$$

$$\ln(\sigma_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk}, \quad (\text{scale submodel}) \quad (10)$$

130 where the random intercept  $u_j^{(l)}$  is distributed as  $u_j^{(l)} \sim \mathcal{N}(0, \sigma_u^2)$ . Here  $y_{ij}$  is the  $i$ -th response in group  $j$ ,  
131  $\mu_{ij}$  its expected value including the group-specific shift  $u_j^{(l)}$ , and  $\sigma_{ij}$  the residual standard deviation driven  
132 by the scale covariates alone. This (mixed-effects) location–scale model tests whether predictors affect both  
133 the mean across and within groups, while allowing groups to differ in their overall mean level.

134 Notably, we can easily extend Model 2’s location submodel to have more than one random effect (intercept)  
135 and random slopes. Indeed, such models with multi-random effects may be the rule rather than an  
136 exception in ecological and evolutionary data (? , e.g., site and year, or individuals nested in  
137 sites)[[schielzeth2013nested.

### 138 **3.2 Illustrative example**

139 We examined the difference in fledging scaled mass index (SMI), i.e., mass corrected by body size, between  
140 first and second hatched blue-footed booby (*Sula nebouxii*) chicks (Drummond et al., 2025). This Gaussian  
141 location-scale model included nest identity ( $\sigma_{NestID(l)}$ ) and hatching year ( $\sigma_{hatching.year(l)}$ ) as random  
142 effects in the location submodel, and hatching order in both submodels. We found a conclusive mean

143 ln(SMI) difference between first and second hatched chicks (brms:  $\beta_{[\text{first-second}]}^{(l)} = -0.02$ , 95% CI = -0.02 to  
 144 -0.01). Moreover, second hatched chicks exhibited greater ln(SMI) variability compared to their first  
 145 hatched counterparts (brms:  $\beta_{[\text{first-second}]}^{(s)} = 0.13$ , 95% CI = 0.08 to 0.18). Random effects in the location  
 146 component also showed that average ln(SMI) differed between nests (brms:  $\sigma_{NestID} = 0.05$ , 95% CI = 0.04  
 147 to 0.05) and hatching years (brms:  $\sigma_{hatching.year} = 0.10$ , 95% CI = 0.07 to 0.14). These results suggest that  
 148 second hatched chicks not only have a slightly lower average ln(SMI) but also exhibit greater variability in  
 149 their SMI compared to first hatched chicks.

## 150 4 Double-hierarchical model (Model 3)

### 151 4.1 Model and motivation

152 Model 2 naturally begs a question: why do not we add random effects in the scale part? Indeed,  
 153 “double-hierarchical” models were the first to arrive in ecology and evolution nearly a decade ago (e.g.,  
 154 Westneat et al., 2013). The double-hierarchical formulation jointly models how each group  $j$  shifts its mean  
 155 and its standard deviation on the natural logarithm scale (Model 3; Lee and Nelder, 1996, 2006; Cleasby  
 156 et al., 2015; O’Dea et al., 2022):

$$y_{ij} \sim \mathcal{N}(\mu_{ij}, \sigma_{ij}^2), \quad (11)$$

$$\mu_{ij} = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (12)$$

$$\ln(\sigma_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk} + u_j^{(s)}, \quad (\text{scale submodel}) \quad (13)$$

157 with the bivariate random-effect vector  $(u_j^{(l)}, u_j^{(s)})^\top$  following

$$\begin{pmatrix} u_j^{(l)} \\ u_j^{(s)} \end{pmatrix} \sim \mathcal{N} \left( \mathbf{0}, \begin{pmatrix} \sigma_{u^{(l)}}^2 & \rho_u \sigma_{u^{(l)}} \sigma_{u^{(s)}} \\ \rho_u \sigma_{u^{(l)}} \sigma_{u^{(s)}} & \sigma_{u^{(s)}}^2 \end{pmatrix} \right). \quad (14)$$

158 Here, each group  $j$  has its own intercept in the mean ( $u_j^{(l)}$ ) and in the ln-standard deviation ( $u_j^{(s)}$ ), with their  
 159 covariance governed by  $\rho_u$ . A positive  $\rho_u$  implies that groups with higher means also exhibit greater  
 160 variability, whereas a negative  $\rho_u$  indicates that high-mean groups are more tightly canalized. This full  
 161 double-hierarchical model thus allows simultaneous inference on fixed effects and group-level  
 162 mean–variance associations. An extension of this model with a random slope in both location and scale  
 163 parts in the context of uni- and multi-variate cases is well described in O’Dea et al. (2022). For example,  
 164 when the cluster  $u_j$  represents individuals ( $y_{ij}$  is repeated behavioral measures of an individual), the  
 165 parameter  $\rho_u$  is referred to as the personality-predictability association. This is because  $\sigma_{u^{(l)}}^2$  reflects  
 166 between-individual differences in mean behavior (personality), while  $\sigma_{u^{(s)}}^2$  captures differences in  
 167 behavioral variance (predictability). For instance, a positive correlation would indicate that more  
 168 aggressive individuals are also more unpredictable in the intensity of their aggression at one time point.

169 As described, our focus in this article is to highlight Model 2 (and Model 1). Therefore, even if one is  
 170 interested in  $\sigma_{u^{(s)}}^2$  and  $\rho_u$ , one should start with Model 2 as a robust baseline. One can fit Model 3, and  
 171 compare Models 2 and 3 using information criteria or likelihood-ratio tests, if sample size permits (more  
 172 than 10 repeats or observations per group may be required to model  $\sigma_{u^{(s)}}^2$  reliably; O’Dea et al., 2022);  
 173 indeed, a simple simulation reveals that one requires 20 observations to get unbiased variance estimates  
 174 (see the online tutorial (link)). Such a modeling strategy leverages the stability of Model 2 while allowing  
 175 the richer inferences of Model 3 when data permit (for more on model selection, see Section 7).

## 176 4.2 Illustrative example

177 Building upon the previous example of fledging scaled mass index (SMI) (Drummond et al., 2025), we  
 178 fitted a Double-hierarchical Gaussian location-scale model. This extended Model 2 by incorporating nest  
 179 identity ( $\sigma_{NestID}$ ) as a correlated random effect in both the location and scale submodels. This allowed us  
 180 to assess how average  $\ln(\text{SMI})$  and its variability differed across nests, and if these nest-specific variations  
 181 were related. Average  $\ln(\text{SMI})$  differ between nests (brms:  $\sigma_{NestID(l)} = 0.05$ , 95% CI = 0.04 to 0.05), and  
 182 some nests showed greater  $\ln(\text{SMI})$  variability (brms:  $\sigma_{NestID(s)} = 0.36$ , 95% CI = 0.32 to 0.40). Notably,  
 183 a negative correlation between location and scale random effects within nests (brms:  $\rho_{NestID} = -0.46$ , 95%  
 184 CI = -0.58 to -0.33) indicated that nests with higher average  $\ln(\text{SMI})$  tended to exhibit lower variability.  
 185 Fixed effects for hatching order remained consistent with our previous model, further supporting that  
 186 second hatched chicks have slightly lower mean  $\ln(\text{SMI})$  and greater variability.

## 187 5 Beyond Gaussian I: over-dispersed count data

188 In this and the next section, we turn from Gaussian responses to non-Gaussian data common in the natural  
 189 world. Our focus is deliberately selective: we concentrate on count and proportion responses, omitting  
 190 ordinal outcomes despite their feasibility with location-scale models (e.g., Martin et al., 2017). For these  
 191 two response variable types, we develop three practical formulations for researchers. Because structural  
 192 zeros (and ones for proportions) are common in ecological and environmental datasets, some count and  
 193 proportion models include zero- or zero/one-inflation components (submodels). To keep the description  
 194 clear, we present each model with the single random-intercept structure introduced in Model 2, though  
 195 Models 1 and 3 forms are also applicable.

### 196 5.1 Negative-binomial location–scale model

197 Many ecological questions involve integer counts: fledglings per nest, insect colony size, or the number of  
 198 eco- or endo-parasites. While Poisson regression is the usual starting point, real data rarely meet its  
 199 assumption that mean equals variance (i.e.,  $E[y] = Var[y]$ ). Indeed, as many researchers know, count data

200 often exhibit over-dispersion ( $E[y] < Var[y]$ ). Negative-binomial regression offers a solution because the  
 201 negative-binomial (NB) distribution (family) has an extra parameter to model this over-dispersion (Stoklosa  
 202 et al., 2022).

203 A negative-binomial location scale model – in the form of Model 2 (a random effect only in the scale part) –  
 204 can be written as (Jorgensen, 1997; Lee and Nelder, 1996, 2006):

$$y_{ij} \sim \text{NB}(\mu_{ij}, \theta_{ij}), \quad (15)$$

$$\ln(\mu_{ij}) = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (16)$$

$$\ln(\theta_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk}, \quad (\text{scale submodel}) \quad (17)$$

205 where  $y_{ij}$  is the count for observation  $i$  in group  $j$ ,  $\mu_{ij}$  is its mean, linked via a log (ln) link to fixed  
 206 covariates  $x_{ijk}$  and a group-level random intercept  $u_j^{(l)}$ ,  $\theta_{ij}$  is the dispersion parameter, linked on the ln  
 207 scale to the same covariates but no random effect,  $u_j^{(l)} \sim \mathcal{N}(0, \sigma_u^2)$  captures group-level shifts in the mean,  
 208 and the log links ensure  $\mu_{ij}, \theta_{ij} > 0$ . The parameter  $\theta_{ij}$  is analogous to the Gaussian dispersion parameter  
 209  $\sigma_{ij}$  but is quite different; it calibrates over-dispersion, and a larger value of  $\theta_{ij}$  represents less variation.  
 210 This role becomes clear when one sees the formula for variance for the negative-binomial distribution.  
 211  $\text{Var}(Y_{ij}) = \mu_{ij} + \mu_{ij}^2/\theta_{ij}$  so that as  $\theta_{ij} \rightarrow \infty$ , the term  $\mu_{ij}^2/\theta_{ij}$  vanishes and the distribution approaches  
 212 the Poisson mean-variance expectation ( $E[y] = Var[y]$ ); conversely, smaller  $\theta_{ij}$  produces increasingly  
 213 strong over-dispersion relative to the Poisson expectation.

## 214 **5.2 Zero-inflated negative-binomial location-scale model**

215 Ecological and evolutionary applications frequently encounter count data with both an excess of true  
 216 absences alongside over-dispersed counts (cf., Zuur et al., 2009). For example, surveying soil invertebrates  
 217 across patchy habitats might yield samples with zero individuals (structural zeros) and others with wildly  
 218 varying densities. Similarly, parasite counts in wildlife often include hosts with no infection and others with

219 heavy (Taylor et al., 2017, e.g.). To model these dual processes while allowing for distinct underlying  
 220 distributions across populations or sites, we embed a single random intercept in the location submodel of a  
 221 zero-inflated negative-binomial location-scale framework:

$$y_{ij} \sim \begin{cases} 0, & \text{with probability } \pi_{ij}, \\ \text{NB}(\mu_{ij}, \theta_{ij}), & \text{with probability } 1 - \pi_{ij}, \end{cases} \quad (18)$$

$$\text{logit}(\pi_{ij}) = \beta_0^{(0)} + \sum_{k=1}^K \beta_k^{(0)} x_{ijk}, \quad (\text{zero-inflation submodel}) \quad (19)$$

$$\ln(\mu_{ij}) = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (20)$$

$$\ln(\theta_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk}, \quad (\text{scale submodel}) \quad (21)$$

222 where  $y_{ij}$  is the count for observation  $i$  in group  $j$ . The zero-inflation submodel predicts the probability  $\pi_{ij}$   
 223 of a guaranteed zero via a logit link and fixed covariates  $x_{ijk}$ . Here,  $\beta_0^{(0)}$  is the baseline log-odds of an  
 224 excess zero when all covariates  $x_{ijk} = 0$ , and each  $\beta_k^{(0)}$  represents the change in log-odds of a guaranteed  
 225 zero per unit increase in covariate  $x_{ijk}$ . A positive  $\beta_k^{(0)}$  thus indicates that higher values of  $x_k$  increase the  
 226 probability of structural absence, whereas a negative  $\beta_k^{(0)}$  decreases it. The location submodel predicts  
 227  $\mu_{ij} > 0$  via a log link, including the group-specific random intercept  $u_j^{(l)} \sim \mathcal{N}(0, \sigma_u^2)$ , which captures  
 228 unobserved differences among groups. The scale submodel with fixed covariates alone governs the  
 229 dispersion parameter  $\theta_{ij} > 0$ , so larger  $\theta_{ij}$  yields variance closer to the mean, as described above.

230 This model formulation allows researchers to simultaneously investigate how habitat characteristics and  
 231 evolutionary history influence (1) the chance of encountering no individuals at all, (2) the expected  
 232 abundance when presence occurs, and (3) the degree of overdispersion beyond the Poisson expectation.  
 233 Notably, Stoklosa et al. (2022), in their review of negative-binomial modeling, advocate for  
 234 negative-binomial models as a default for count data in ecology and biodiversity, given their

235 near-ubiquitous over-dispersion.

### 236 **5.3 Conway–Maxwell–Poisson location–scale model**

237 Under-dispersion ( $Var(Y) < E[Y]$ ) is probably less common but potentially important in ecological and  
 238 environmental datasets. For example, stabilizing selection and biological ceiling (floor) effects could  
 239 canalize count data. The Conway–Maxwell–Poisson (CMP) family (distribution) spans over- and  
 240 under-dispersion with a parameter  $\nu$  (variance drops as  $\nu \uparrow$ ) (Sellers and Shmueli, 2010):

$$y_{ij} \sim \text{CMP}(\mu_{ij}, \nu_{ij}), \quad (22)$$

$$\ln(\mu_{ij}) = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (23)$$

$$\ln(\nu_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk}, \quad (\text{scale submodel}) \quad (24)$$

241 where  $y_{ij}$  is the count for observation  $i$  in group  $j$ ;  $\mu_{ij} > 0$  is the CMP “rate” (mean, often denoted as  $\lambda$ ),  
 242 on the log scale linked to predictors  $x_{ijk}$  and a random intercept  $u_j^{(l)} \sim \mathcal{N}(0, \sigma_u^2)$ ;  $\nu_{ij} > 0$  represents  
 243 under-dispersion:  $\nu = 1$  recovers the Poisson  $Var(Y) = E[Y]$  yields over-dispersion, and  $\nu > 1$   
 244 under-dispersion.

245 By fitting this mixed-effects location–scale CMP model, ecologists and environmental biologists can probe  
 246 not only how drivers such as resource availability, temperature stress, or habitat fragmentation shift the  
 247 average count of organisms or events, but also whether these same forces tighten or loosen the Poisson  
 248 expectation on variability. Notably, Brooks et al. (2019) points out the dual ability of CMP to deal with  
 249 both overdispersion and underdispersion. Moreover, they introduce zero-inflated CMP models (ZICMP)  
 250 using ‘glmmTMB’ (Brooks et al., 2019). As we mentioned earlier, its capability to model underdispersion  
 251 is important, because this cannot be done by negative-binomial models. For example, under strong  
 252 stabilizing selection on clutch size, many bird species have evolved canalized brood counts, often producing  
 253 almost exactly the same number of eggs each year, a pattern of under-dispersion captured by  $\nu > 1$  (e.g.,

254 Boyce and Perrins, 1987; Liou et al., 1993; Santos and Nakagawa, 2013).

## 255 **5.4 Illustrative example**

256 We analyzed visual preference in Estrildid finches by measuring gaze frequency to dot stimuli under  
 257 food-supplied and food-deprived conditions (Mizuno and Soma, 2023). To account for overdispersed count  
 258 data, we used a negative-binomial location-scale model (corresponding to Model 2), with species and  
 259 individual (nested within species) as random effects in the location component. Birds gazed significantly  
 260 less at dots when food was supplied ( $\text{glmmTMB: } \beta^{(l)}_{\text{deprived-supplied}} = -0.85, 95\% \text{ CI} = -1.08 \text{ to } -0.61$ ).  
 261 The scale component revealed greater individual-level variability under food-supplied conditions, indicated  
 262 by a negative effect of this condition ( $\theta: \beta^{(s)}_{\text{deprived-supplied}} = -0.66, 95\% \text{ CI} = -1.15 \text{ to } -0.18$ ).  
 263 Species-level variation in average gaze frequency ( $\text{SD} = 0.55, 95\% \text{ CI} = 0.31 \text{ to } 0.99$ ) exceeded  
 264 within-species individual variation ( $\text{SD} = 0.34, 95\% \text{ CI} = 0.17 \text{ to } 0.68$ ). Thus, food deprivation increased  
 265 average gazing, while availability reduced gazing but amplified individual variability.

## 266 **6 Beyond Gaussian II: over-dispersed proportion data**

267 Proportions come in two flavors. Discrete (binomial) proportions arise as “successes out of trials”, for  
 268 example, the number of germinated seeds out of 20, the tally of infected hosts in a sample, the abundance of  
 269 a certain taxon in microbial communities. They are naturally modeled with binomial regression (e.g.,  
 270 Bolker et al., 2009; Zuur et al., 2009). Continuous proportions, in contrast, are already measured as rates on  
 271 the unit interval,  $[0, 1]$  – leaf-area loss, percent cover, the fraction of time an animal spends foraging.  
 272 Continuous proportions are usually analyzed with Beta regression, which takes values between 0 and 1  
 273 (Ferrari and Cribari-Neto, 2004; Douma and Weedon, 2019).

274 Boundary values (i.e., 0 and 1) complicate matters differently for the two types of proportion. Because the  
 275 binomial distribution already includes zero and  $n$  (the number of ‘trials’), discrete counts can generate  
 276 observed proportions of exactly 0 or 1; yet in practice, true absences (e.g., empty traps and seeds that could  
 277 never germinate) often occur more frequently than a binomial distribution can allow (cf., Warton, 2005). A

278 zero-inflation component, therefore, captures a separate “structural-zero” process. In contrast, structural  
 279 ones (a one-inflation component) are seldom, if ever, needed because excess of perfect ‘successes’ are  
 280 unlikely to occur in nature (e.g., Zuur et al., 2009). Beta models, by construction, exclude the boundaries of  
 281 the unit interval, so when continuous proportions include any zeros or ones – for example, sprayed plots  
 282 with 0 % damage, or quadrats that are completely vegetated – both zeros and ones must be modeled via  
 283 zero- and one-inflation submodels respectively (Ospina and Ferrari, 2012). Bearing this in mind, we  
 284 introduce three location-scale models for proportion data below.

### 285 **6.1 Beta-binomial location–scale model**

286 For discrete proportions (e.g., seedling emergence, infection prevalence), one usually starts modeling by  
 287 assuming a binomial distribution:

$$y_{ij} \sim \text{Binomial}(n_{ij}, \mu_{ij}), \quad (25)$$

288 where  $y_{ij}$  is the number of successes out of  $n_{ij}$  trials in group  $j$  and  $\mu_{ij} \in (0, 1)$  is the underlying success  
 289 probability (often denoted  $p$ ). Yet, a binomial distribution ‘fixes’ the variance at  $n_{ij}\mu_{ij}(1 - \mu_{ij})$  (i.e., the  
 290 binomial-variance expectation) and therefore cannot accommodate the extra-binomial dispersion that is  
 291 common in field data.

292 However, if we assume that the success probability itself varies among observational units according to a  
 293 Beta distribution,  $\mu_{ij} \sim \text{Beta}(\alpha_{ij}, \beta_{ij})$ , we can combine these two distributions to yield a Beta–binomial  
 294 distribution:

$$y_{ij} \sim \text{Beta-binomial}(n_{ij}, \mu_{ij}, \phi_{ij}), \quad (26)$$

295 where the Beta distribution’s parameters are reparameterized as  $\alpha_{ij} = \mu_{ij} \phi_{ij}$  and  $\beta_{ij} = (1 - \mu_{ij}) \phi_{ij}$ .

296 Here  $\phi_{ij} > 0$  is a precision (inverse–dispersion or inverse–variance) term. For the resulting Beta–binomial

297 the variance is  $\text{Var}(y_{ij}) = n_{ij} \mu_{ij} (1 - \mu_{ij}) ((n_{ij} + \phi_{ij}) / (1 + \phi_{ij}))$ . When  $\phi_{ij} \rightarrow \infty$ , the fraction

298  $(n_{ij} + \phi_{ij}) / (1 + \phi_{ij})$  to 1; the variance collapses to the binomial-variance expectation  $n_{ij} \mu_{ij} (1 - \mu_{ij})$  and

299 there is no over-dispersion. Therefore,  $\phi$  has the same role as the  $\theta$  over-dispersion parameter in the  
 300 negative binomial distribution. Given this property of a Beta-binomial distribution, we can let predictors  
 301 explain both the mean success probability and the amount of extra dispersion, while allowing for  
 302 group-level shifts in the mean (Jorgensen, 1997; Lee and Nelder, 1996, 2006):

$$\text{logit}(\mu_{ij}) = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (27)$$

$$\ln(\phi_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk}. \quad (\text{scale submodel}) \quad (28)$$

303 In the location submodel, the random intercept  $u_j^{(l)} \sim \mathcal{N}(0, \sigma_u^2)$  captures baseline differences among sites  
 304 or populations. The scale submodel links the ln-precision to the same (or different) covariates, so predictors  
 305 can inflate ( $\phi_{ij} \downarrow$ ) or dampen ( $\phi_{ij} \uparrow$ ) the variation beyond the binomial-variance expectation. Relatedly,  
 306 Martin et al. (2020) introduced the use of the Beta-binomial location-scale model to quantify the relative  
 307 abundance of a specific taxon in microbial communities (genetic sequencing of microbiome samples results  
 308 in discrete proportion data). They indeed emphasized the importance of its ability to model dispersion.

## 309 **6.2 Zero-inflated Beta–binomial location–scale model**

310 In many ecological discrete proportion data (e.g., seedling emergence, infection prevalence), counts of  
 311 “successes” out of  $n_{ij}$  trials show both structural zeros (true absences) and extra-binomial scatter. A  
 312 zero-inflated Beta–binomial location–scale model accommodates: 1) a point-mass at zero, 2) group-level

313 shifts in the mean, and 3) over-dispersion beyond the binomial expectation, all within a single framework:

$$y_{ij} \sim \begin{cases} 0, & \text{with probability } \pi_{ij}, \\ \text{Beta-binomial}(n_{ij}, \mu_{ij}, \phi_{ij}), & \text{with probability } 1 - \pi_{ij}, \end{cases} \quad (29)$$

$$\text{logit}(\pi_{ij}) = \beta_0^{(0)} + \sum_{k=1}^K \beta_k^{(0)} x_{ijk}, \quad (\text{zero-inflation submodel}) \quad (30)$$

$$\text{logit}(\mu_{ij}) = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (31)$$

$$\ln(\phi_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk}, \quad (\text{scale submodel}) \quad (32)$$

314 Here  $y_{ij}$  is the number of successes in  $n_{ij}$  trials for observation  $i$  in group  $j$ . The zero-inflation submodel  
 315 predicts the probability  $\pi_{ij}$  of a “structural” zero via a logit link and covariates  $x_{ijk}$ . Conditional on  
 316 non-zero counts, the Beta–binomial component arises by assuming the success probability itself follows  
 317  $\text{Beta}(\mu_{ij} \phi_{ij}, (1 - \mu_{ij}) \phi_{ij})$ . The location submodel – with its random intercept  $u_j^{(l)}$  – captures baseline  
 318 differences among sites or populations, while the scale submodel lets covariates modulate the precision  $\phi_{ij}$ .  
 319 Similar to Martin et al. (2020), Hu et al. (2018) proposed zero-inflated beta-binomial models for  
 320 microbiome data. While not full location-scale models, their examples underscore the importance of  
 321 modeling zeros in such data.

### 322 **6.3 Zero-and-one-inflated Beta location–scale model**

323 Continuous proportions often include exact zeros or ones (e.g., complete absence or saturation), which  
 324 standard Beta regressions cannot accommodate. Zero-and-one-inflated Beta models resolve this by mixing  
 325 three submodels to estimate coefficients for point mass at 0, point mass at 1, and the Beta-distributed  
 326 interior (Ospina and Ferrari, 2012). This approach models the occurrence of boundary outcomes and the  
 327 variability of intermediate proportions in a single, interpretable framework, without dropping or adjusting

328 boundary data:

$$y_{ij} \sim \begin{cases} 0, & \text{with probability } \pi_{0,ij}, \\ 1, & \text{with probability } \pi_{1,ij}, \\ \text{Beta}(\mu_{ij} \phi_{ij}, (1 - \mu_{ij}) \phi_{ij}), & \text{with probability } 1 - \pi_{0,ij} - \pi_{1,ij}, \end{cases} \quad (33)$$

$$\text{logit}(\pi_{0,ij}) = \beta_0^{(0)} + \sum_{k=1}^K \beta_k^{(0)} x_{ijk}, \quad (\text{zero-inflation submodel}) \quad (34)$$

$$\text{logit}(\pi_{1,ij}) = \beta_0^{(1)} + \sum_{k=1}^K \beta_k^{(1)} x_{ijk}, \quad (\text{one-inflation submodel}) \quad (35)$$

$$\text{logit}(\mu_{ij}) = \beta_0^{(l)} + \sum_{k=1}^K \beta_k^{(l)} x_{ijk} + u_j^{(l)}, \quad (\text{location submodel}) \quad (36)$$

$$\ln(\phi_{ij}) = \beta_0^{(s)} + \sum_{k=1}^K \beta_k^{(s)} x_{ijk}, \quad (\text{scale submodel}) \quad (37)$$

329 Here  $\pi_{0,ij}$  and  $\pi_{1,ij}$  are the structural-zero and structural-one probabilities;  $\mu_{ij}$  and  $\phi_{ij}$  govern the  
 330 continuous Beta component; and  $u_j^{(l)}$  is the lone random intercept in the location submodel, allowing group  
 331  $j$  to differ in its baseline mean proportion. The parameters  $\beta_0^{(0)}$  and  $\beta_k^{(0)}$  set the log-odds of an exact zero,  
 332 while  $\beta_0^{(1)}$  and  $\beta_k^{(1)}$  set the log-odds of an exact one; each as a function of covariates. The variance of the  
 333 Beta-distributed interior is  $\text{Var}(y_{ij}) = \mu_{ij}(1 - \mu_{ij})/(1 + \phi_{ij})$ .

334 When  $\phi_{ij} \rightarrow \infty$  the dispersion shrinks to zero and data distribution concentrates around its mean, whereas  
 335 as  $\phi_{ij}$  approaches zero, the variance approaches its maximum  $\mu_{ij}(1 - \mu_{ij})$ . Thus, lower  $\phi_{ij}$  inflates and  
 336 higher  $\phi_{ij}$  deflates variability around the mean, and the scale submodel lets predictors modulate dispersion  
 337 separately from the mean process. Note that when data does not include zeros and ones, one can remove  
 338 corresponding submodels (i.e., Beta location-scale models).

339 Burke et al. (2023) used a zero-inflated Beta location-scale model – without one-inflation as their dataset  
 340 did not have ones — to examine patterns and drivers of coral diseases (measured by percentage areas of

341 diseased corals) in a meta-analytic context (see Section 8). They found that when sea surface temperature  
 342 increases, not only did the mean percentage of coral disease increase, but so did its variability, and,  
 343 surprisingly, the observations of zero-percent disease, too.

#### 344 **6.4 Illustrative example**

345 Lundgren et al. (2022) investigated whether mountain lion predation reduced feral donkey impacts on  
 346 desert wetlands. We re-analyzed some of these data with a Beta location-scale model. We included a  
 347 zero-one inflation and conditional one-inflation submodels to account for exact 0 and 1 values. The model  
 348 revealed that on average, the log-odds of the mean percentage of trampled bare ground were lower in areas  
 349 with high predation risk ( $\text{brms: } \beta_{[\text{predation-no predation}]}^{(l)} = -1.22, 95\%CI[-2.27, -0.71]$ ). The scale  
 350 component showed that log-precision ( $\phi$ ) was lower at sites with predation ( $\phi$ :  
 351  $\beta_{[\text{predation-no predation}]}^{(s)} = -1.07, 95\%CI[-2.01, -0.04]$ ), indicating more variation in trampling in areas with  
 352 predation. See our tutorial (link for the R code and interpretation of zero and one-inflated submodels).

#### 353 **7 Proposed workflow and diagnostics**

354 The workflow in Fig. 2 begins with data visualization. Scatterplots of the raw response against each  
 355 predictor often reveal variance fans or funnels, indicating heteroscedasticity. For categorical predictors,  
 356 some groups may show greater data spread (Cleasby and Nakagawa, 2011; Nakagawa et al., 2025).  
 357 However, these patterns are sometimes hard to discern from raw count and proportion data. In these  
 358 situations, residuals from a baseline ‘location-only’ model provide the most informative starting point. This  
 359 baseline may take the form of a linear or generalized linear model, or a random-intercept mixed model in  
 360 the presence of clustering. While residual Q–Q plots effectively identify heteroscedasticity in Gaussian  
 361 data, they are not suitable for other data types. Randomized quantile residual Q–Q plots generalize this  
 362 concept to any exponential-family response (e.g., Poisson or binomial), and they can quickly reveal  
 363 violations of homoscedasticity or other distributional assumptions (e.g., zero inflation) (Dunn and Smyth,  
 364 1996). These residuals can be calculated by the R package DHARMA (Hartig, 2022). Together, these graphics

365 indicate whether variance is stable enough to justify a location-only model.

366 If heteroscedasticity is observed, we extend the model to include a scale submodel. With generous sample  
367 sizes per cluster (e.g.,  $> 5$ ), random effects can enter the scale part, and a correlation can be modeled  
368 between the location and scale part of the same random effects (i.e., a double-hierarchical model). Classical  
369 information criteria, such as AIC for frequentist implementations (Anderson and Burnham, 2004) and  
370 WAIC or LOO-CV (leave-one-out cross validation) for Bayesian implementations (Vehtari et al., 2017) can  
371 help simplify the model. However, these scores should be treated as heuristic filters, flagging candidate  
372 models for closer inspection. Ultimately, the model must address the motivating ecological or evolutionary  
373 question, not merely minimize an index. Alternatively, if one has clear predictions for both the scale part  
374 and the location part, one can create such a model with all relevant predictors, bypassing model selection or  
375 simplification.

## 376 **8 Further extensions and future perspectives**

377 Location–scale thinking invites a broader re-imagination of data analysis. To assist this, we describe four  
378 extensions that expand the analytical capability to understand variability and quantify heteroscedasticity.  
379 First, ecological and environmental traits/measurements rarely act in isolation. Multivariate location–scale  
380 models analyze suites of traits simultaneously, estimating covariances not only among means but also  
381 among variances, and even mean–variance cross-links among traits. Such models can test, for instance,  
382 whether life-history ‘syndromes’ involve coordinated changes in both average values and trait predictability,  
383 or whether plasticity in one dimension buffers variability in another (O’Dea et al., 2022).

384 Second, Blowes (2024) and Nakagawa et al. (2025) have introduced and highlighted that bringing  
385 location–scale thinking into ecological and evolutionary meta-analysis would allow evidence syntheses to  
386 ask when and why heterogeneity among effect sizes change along environmental gradients and  
387 methodological differences. Meta-analytic location–scale models treat heterogeneity, which dominates  
388 ecological and evolutionary meta-analyses (Senior et al., 2016), as a parameter to be explained rather than

389 tolerated. As such, these models can uncover hidden structure in the “noise” of published effect sizes.  
390 Indeed, using several datasets from community ecology, Blowes (2024) showed that location-scale  
391 meta-regression can significantly improve model fit compared to location-only meta-regression.

392 Third, Halliwell (2025) and Nakagawa et al. (2025) have introduced phylogenetic location-scale models,  
393 emphasizing that variance itself can evolve and should be a part of macro-evolutionary and  
394 community-ecological investigation. Embedding phylogenetic covariance structures in both mean and  
395 variance sub-models opens new terrain for comparative biology. A phylogenetic location–scale model can  
396 reveal whether evolutionary shifts in trait means are accompanied by shifts in trait variability, and whether  
397 certain clades are consistently more (or less) variable than expected. By quantifying “phylogenetic  
398 heritability” for variance and means, researchers gain a fuller picture of evolutionary constraints,  
399 innovations and trade-offs.

400 Fourth, responses not only have location and scale but also have ‘shape’. Extending the framework to  
401 include a shape component (e.g., skewness, kurtosis or heavy tails) would ask how entire distributions shift  
402 under ecological, evolutionary and environmental change (Stemkovski et al., 2023; Cornwell and Ackerly,  
403 2009). ‘Location–scale–shape’ models are already feasible in generalized additive or flexible Bayesian  
404 settings (Rigby and Stasinopoulos, 2005; Corrales and Cepeda-Cuervo, 2022; Stasinopoulos and Rigby,  
405 2008; Umlauf et al., 2021). Such models promise insights into the frequency of extreme events, asymmetric  
406 risks, stabilizing selection, and bet-hedging strategies (Pick et al., 2022; Starrfelt and Kokko, 2012; Pollo  
407 et al., 2025).

408 Collectively, these extensions remind us that mean responses are only the tip of the statistical iceberg.  
409 Embracing location, scale and (eventually) shape as joint products of ecological and evolutionary processes  
410 will deepen our understanding of how organisms and ecosystems respond to an increasingly variable world.

## 411 **9 Conclusions**

412 Location-scale models provide a powerful lens through which ecologists and evolutionary biologists can  
413 interpret different types of data (i.e., continuous, count and proportion data). Building on the call from  
414 Cleasby and Nakagawa (2011) to treat heteroscedasticity as a biological clue and process, these approaches  
415 offer both conceptual and practical tools for richer inference. As datasets grow larger and more complex,  
416 studying variance as well as the mean should be standard practice in our analytical workflow in ecology,  
417 evolution, and environmental sciences. Let's re-imagine heterogeneity.

## 418 **10 Author Statement**

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## 429 **References**

- 430 Anderson, D. and K. Burnham (2004). Model selection and multi-model inference. *Second. NY:*  
431 *Springer-Verlag* 63(2020), 10.
- 432 Baldwin, J. M. (1896). A new factor in evolution. *American Naturalist* 30(354), 441–451.
- 433 Blowes, S. A. (2024). Known unknowns and model selection in ecological evidence synthesis. *bioRxiv*,

- 434 <https://doi.org/10.1101/2024.12.18.629303>.
- 435 Bolker, B. M., M. E. Brooks, C. J. Clark, S. W. Geange, J. R. Poulsen, M. H. H. Stevens, and J.-S. S. White  
436 (2009). Generalized linear mixed models: a practical guide for ecology and evolution. *Trends in ecology*  
437 *& evolution* 24(3), 127–135.
- 438 Boyce, M. S. and C. Perrins (1987). Optimizing great tit clutch size in a fluctuating environment.  
439 *Ecology* 68(1), 142–153.
- 440 Brooks, M. E., K. Kristensen, M. R. Darrigo, P. Rubim, M. Uriarte, E. Bruna, and B. M. Bolker (2019).  
441 Statistical modeling of patterns in annual reproductive rates. *Ecology* 100(7), e02706.
- 442 Brooks, M. E., K. Kristensen, K. J. Van Benthem, A. Magnusson, C. W. Berg, A. Nielsen, H. J. Skaug,  
443 M. Mächler, and B. M. Bolker (2017). glmmTMB balances speed and flexibility among packages for  
444 zero-inflated generalized linear mixed modelling. *The R Journal* 9(2), 378–400.
- 445 Buckley, L. B. and R. B. Huey (2016). How extreme temperatures impact organisms and the evolution of  
446 their thermal tolerance. *Integrative and comparative biology* 56(1), 98–109.
- 447 Burke, S., P. Pottier, M. Lagisz, E. L. Macartney, T. Ainsworth, S. M. Drobniak, and S. Nakagawa (2023).  
448 The impact of rising temperatures on the prevalence of coral diseases and its predictability: A global  
449 meta-analysis. *Ecology letters* 26(8), 1466–1481.
- 450 Bürkner, P.-C. (2017). brms: An r package for bayesian multilevel models using stan. *Journal of Statistical*  
451 *Software* 80, 1–28.
- 452 Carroll, R. J. and D. Ruppert (1988). *Transformation and Weighting in Regression*. New York: Chapman  
453 and Hall.
- 454 Cleasby, I. R. and S. Nakagawa (2011). Neglected biological patterns in the residuals: a behavioural  
455 ecologist’s guide to co-operating with heteroscedasticity. *Behavioral Ecology and Sociobiology* 65,  
456 2361–2372.
- 457 Cleasby, I. R., S. Nakagawa, and H. Schielzeth (2015). Quantifying the predictability of behaviour:  
458 statistical approaches for the study of between-individual variation in the within-individual variance.

- 459 *Methods in Ecology and Evolution* 6(1), 27–37.
- 460 Cornwell, W. K. and D. D. Ackerly (2009). Community assembly and shifts in plant trait distributions  
461 across an environmental gradient in coastal california. *Ecological monographs* 79(1), 109–126.
- 462 Corrales, M. L. and E. Cepeda-Cuervo (2022). Bayesian modeling of location, scale, and shape parameters  
463 in skew-normal regression models. *Statistical Analysis and Data Mining: The ASA Data Science*  
464 *Journal* 15(1), 98–111.
- 465 Crispo, E. (2007). The baldwin effect and genetic assimilation: Revisiting two mechanisms of evolutionary  
466 change mediated by phenotypic plasticity. *Evolution* 61(11), 2469–2479.
- 467 Douma, J. C. and J. T. Weedon (2019). Analysing continuous proportions in ecology and evolution: A  
468 practical introduction to beta and dirichlet regression. *Methods in Ecology and Evolution* 10(9),  
469 1412–1430.
- 470 Drummond, H., C. Rodríguez, and S. Ortega (2025). Long-term insights into who benefits from brood  
471 reduction. *Behavioral Ecology* 36(4), araf050.
- 472 Dunn, P. K. and G. K. Smyth (1996). Randomized quantile residuals. *Journal of Computational and*  
473 *graphical statistics* 5(3), 236–244.
- 474 Ferrari, S. and F. Cribari-Neto (2004). Beta regression for modelling rates and proportions. *Journal of*  
475 *applied statistics* 31(7), 799–815.
- 476 Halliwell, B. (2025). Rethinking niche conservatism with phylogenetic location-scale models. *bioRxiv*,  
477 2025–03.
- 478 Hartig, F. (2022). Dharma: residual diagnostics for hierarchical (multi-level/mixed) regression models. r  
479 package version 0.4. 6.
- 480 Hayes, A. F. and L. Cai (2007). Using heteroskedasticity-consistent standard error estimators in ols  
481 regression: An introduction and software implementation. *Behavior research methods* 39, 709–722.
- 482 Hu, T., P. Gallins, and Y.-H. Zhou (2018). A zero-inflated beta-binomial model for microbiome data  
483 analysis. *Stat* 7(1), e185.

- 484 Jorgensen, B. (1997). *The theory of dispersion models*. CRC Press.
- 485 Lee, Y. and J. A. Nelder (1996). Hierarchical generalized linear models. *Journal of the Royal Statistical*  
486 *Society Series B: Statistical Methodology* 58(4), 619–656.
- 487 Lee, Y. and J. A. Nelder (2006). Double hierarchical generalized linear models (with discussion). *Journal*  
488 *of the Royal Statistical Society Series C: Applied Statistics* 55(2), 139–185.
- 489 Lee, Y., J. A. Nelder, and Y. Pawitan (2006). *Generalized linear models with random effects: unified*  
490 *analysis via H-likelihood*. Chapman & Hall/CRC.
- 491 Liou, L. W., T. Price, M. S. Boyce, and C. M. Perrins (1993). Fluctuating environments and clutch size  
492 evolution in great tits. *The American Naturalist* 141(3), 507–516.
- 493 Lundgren, E. J., D. Ramp, O. S. Middleton, E. I. Wooster, E. Kusch, M. Balisi, W. J. Ripple, C. D.  
494 Hasselerharm, J. N. Sanchez, M. Mills, et al. (2022). A novel trophic cascade between cougars and feral  
495 donkeys shapes desert wetlands. *Journal of Animal Ecology* 91(12), 2348–2357.
- 496 Martin, B. D., D. Witten, and A. D. Willis (2020). Modeling microbial abundances and dysbiosis with  
497 beta-binomial regression. *The annals of applied statistics* 14(1), 94.
- 498 Martin, J. G., E. Pirota, M. B. Petelle, and D. T. Blumstein (2017). Genetic basis of between-individual  
499 and within-individual variance of docility. *Journal of Evolutionary Biology* 30(4), 796–805.
- 500 Mizuno, A. and M. Soma (2023). Pre-existing visual preference for white dot patterns in estrildid finches: a  
501 comparative study of a multi-species experiment. *Royal Society Open Science* 10(10), 231057.
- 502 Nakagawa, S., A. Mizuno, K. Morrison, L. Ricolfi, C. Williams, S. M. Drobniak, M. Lagisz, and Y. Yang  
503 (2025). Location-scale meta-analysis and meta-regression as a tool to capture large-scale changes in  
504 biological and methodological heterogeneity: A spotlight on heteroscedasticity. *Global Change*  
505 *Biology* 31(5), e70204.
- 506 Nakagawa, S., A. Mizuno, C. Williams, M. Lagisz, Y. Yang, and S. M. Drobniak (2025). Quantifying  
507 macro-evolutionary patterns of trait mean and variance with phylogenetic location-scale models.  
508 *EcoEvoRxiv*.

- 509 Nakagawa, S. and H. Schielzeth (2013). A general and simple method for obtaining  $r^2$  from generalized  
510 linear mixed-effects models. *Methods in ecology and evolution* 4(2), 133–142.
- 511 O’Dea, R. E., D. W. Noble, and S. Nakagawa (2022). Unifying individual differences in personality,  
512 predictability and plasticity: a practical guide. *Methods in Ecology and Evolution* 13(2), 278–293.
- 513 Ospina, R. and S. L. Ferrari (2012). A general class of zero-or-one inflated beta regression models.  
514 *Computational Statistics & Data Analysis* 56(6), 1609–1623.
- 515 O’Dea, R. E., D. W. Noble, S. L. Johnson, D. Hesselson, and S. Nakagawa (2016). The role of non-genetic  
516 inheritance in evolutionary rescue: epigenetic buffering, heritable bet hedging and epigenetic traps.  
517 *Environmental epigenetics* 2(1), dvv014.
- 518 Pick, J. L., H. E. Lemon, C. E. Thomson, and J. D. Hadfield (2022). Decomposing phenotypic skew and its  
519 effects on the predicted response to strong selection. *Nature Ecology & Evolution* 6(6), 774–785.
- 520 Pollo, P., S. M. Drobniak, H. Haselimashhadi, M. Lagisz, A. Mizuno, D. W. Noble, L. A. Wilson, and  
521 S. Nakagawa (2025). Beyond sex differences in mean: meta-analysis of differences in skewness,  
522 kurtosis, and correlation. *EcoEvoRxiv*. EcoEvoRxiv preprint.
- 523 Rigby, R. A. and D. M. Stasinopoulos (2005). Generalized additive models for location, scale and shape.  
524 *Journal of the Royal Statistical Society Series C: Applied Statistics* 54(3), 507–554.
- 525 Rönnegård, L. and Y. Lee (2013). Exploring the potential of hierarchical generalized linear models in  
526 animal breeding and genetics. *Journal of Animal Breeding & Genetics* 130(6).
- 527 Santos, E. S. and S. Nakagawa (2013). Breeding biology and variable mating system of a population of  
528 introduced dunnocks (*prunella modularis*) in new zealand. *PLoS One* 8(7), e69329.
- 529 Sellers, K. F. and G. Shmueli (2010). A flexible regression model for count data. *The Annals of Applied*  
530 *Statistics*, 943–961.
- 531 Senior, A. M., C. E. Grueber, T. Kamiya, M. Lagisz, K. O’Dwyer, E. S. A. Santos, and S. Nakagawa  
532 (2016). Heterogeneity in ecological and evolutionary meta-analyses: its magnitude and implications.  
533 *Ecology* 97(12), 3293–3299.

- 534 Starrfelt, J. and H. Kokko (2012). Bet-hedging—a triple trade-off between means, variances and  
535 correlations. *Biological Reviews* 87(3), 742–755.
- 536 Stasinopoulos, D. M. and R. A. Rigby (2008). Generalized additive models for location scale and shape  
537 (gamlss) in r. *Journal of Statistical Software* 23, 1–46.
- 538 Stemkovski, M., R. G. Dickson, S. R. Griffin, B. D. Inouye, D. W. Inouye, G. L. Pardee, N. Underwood, and  
539 R. E. Irwin (2023). Skewness in bee and flower phenological distributions.
- 540 Stoklosa, J., R. V. Blakey, and F. K. Hui (2022). An overview of modern applications of negative binomial  
541 modelling in ecology and biodiversity. *Diversity* 14(5), 320.
- 542 Taylor, R. A., S.-J. Park, and P. S. Grewal (2017). Nematode spatial distribution and the frequency of zeros  
543 in samples. *Nematology* 19(3), 263–270.
- 544 Umlauf, N., N. Klein, T. Simon, and A. Zeileis (2021). bamlss: a lego toolbox for flexible bayesian  
545 regression (and beyond). *Journal of Statistical Software* 100, 1–53.
- 546 Vehtari, A., A. Gelman, and J. Gabry (2017). Practical bayesian model evaluation using leave-one-out  
547 cross-validation and waic. *Statistics and computing* 27, 1413–1432.
- 548 Warton, D. I. (2005). Many zeros does not mean zero inflation: comparing the goodness-of-fit of  
549 parametric models to multivariate abundance data. *Environmetrics: The official journal of the*  
550 *International Environmetrics Society* 16(3), 275–289.
- 551 Westneat, D. F., M. Schofield, and J. Wright (2013). Parental behavior exhibits among-individual variance,  
552 plasticity, and heterogeneous residual variance. *Behavioral Ecology* 24(3), 598–604.
- 553 Zuur, A. F., E. N. Ieno, N. Walker, A. A. Saveliev, G. M. Smith, A. F. Zuur, E. N. Ieno, N. J. Walker, A. A.  
554 Saveliev, and G. M. Smith (2009). Zero-truncated and zero-inflated models for count data. *Mixed effects*  
555 *models and extensions in ecology with R*, 261–293.
- 556 Zuur, A. F., E. N. Ieno, N. J. Walker, A. A. Saveliev, G. M. Smith, et al. (2009). *Mixed effects models and*  
557 *extensions in ecology with R*, Volume 574. Springer.

558 **Figure and figure captions**

559 **Figure 1.** Homoscedasticity and heteroscedasticity patterns in common data distributions. Examples for  
560 (a)continuous, (b) count, and (c) proportion data. Top panels show homoscedasticity; bottom panels show  
561 heteroscedasticity. (a) Continuous responses demonstrate how continuous and categorical predictors can  
562 exhibit constant or varying variance across. (b) Count data inherently links the mean and variance, if counts  
563 follow Poisson distributions ( $E[y] < Var[y]$ ). Thus, variance increases with expected value. The bottom  
564 panel, though visually uniform, represents the heteroscedasticity with larger dispersion at higher means. (c)  
565 Proportion data (proportion 0 - 1) shows heteroscedasticity (bottom) as inflated frequencies at the  
566 boundaries (0 - 1), reflecting overdispersion. This is often modeled by a Beta-binomial distribution, where  
567 the success probability varies across observations.

568 **Figure 2.** Practical workflow for detecting and modeling heteroscedasticity with location-scale models.  
569 This diagram outlines a step-by-step guide for applying location-scale models to identify and interpret  
570 non-constant variance in continuous, count, or proportion data. The workflow progresses from initial data  
571 visualization and distribution identification (steps 1 and 2) to fitting a location-only baseline model and  
572 conducting residual diagnostics for variance patterns (steps 3 and 4). If heteroscedasticity is detected, a  
573 location-scale model is fitted (step 5) and compared against other possible models (e.g., ones with fewer or  
574 more fixed effects or random effects) using information criteria such as AIC (frequentist) or WAIC/LOO  
575 (Bayesian) (step 6). Then, we clearly report both mean and variance effects as final results. Note that the  
576 table in step 5 summarizes key variance-related parameters (e.g.,  $\sigma^2$ ,  $\theta$ ,  $\phi$ ,  $\nu$ ) and their corresponding  
577 interpretations (for more details, see the main text).

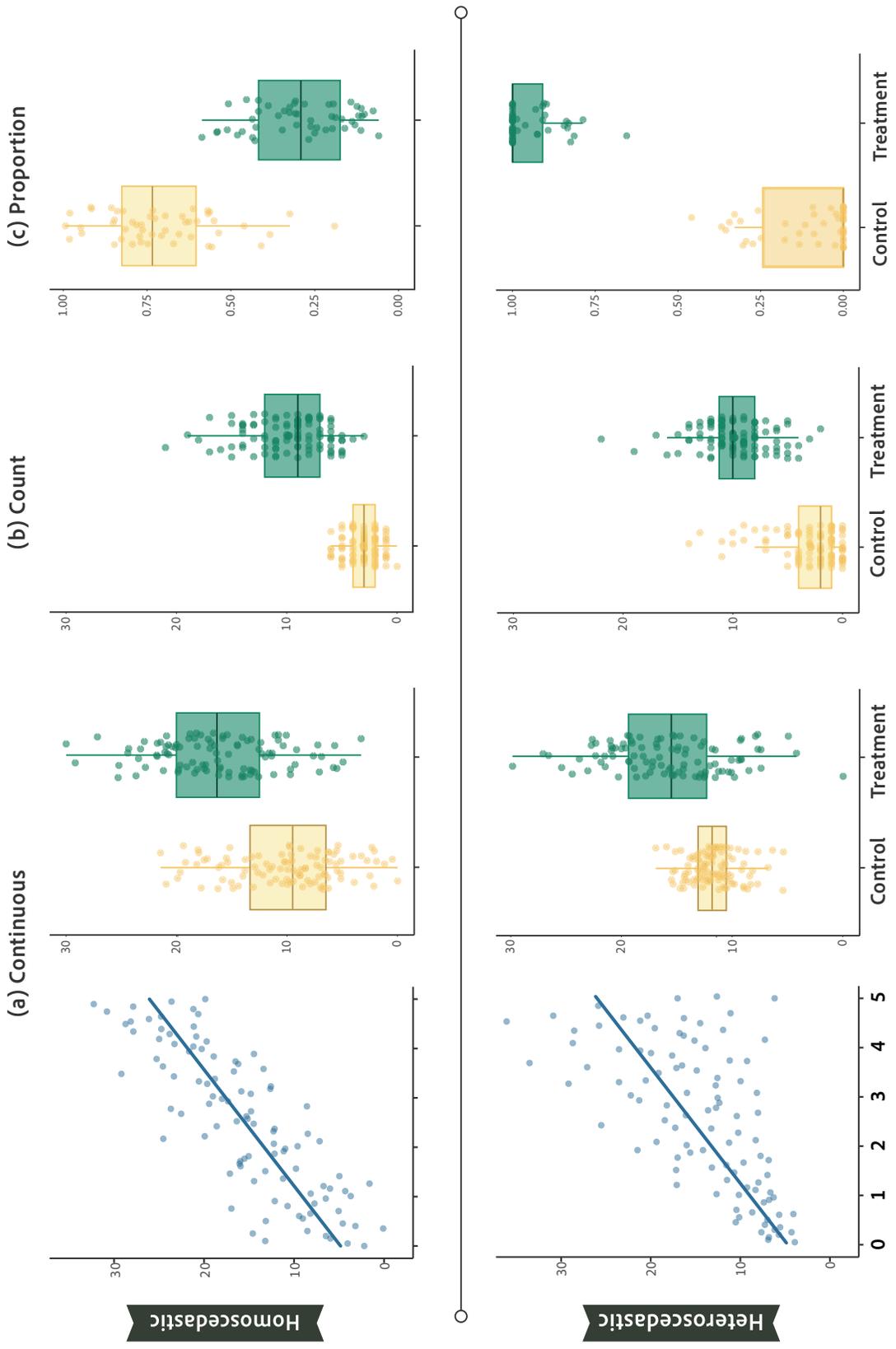


Figure 1

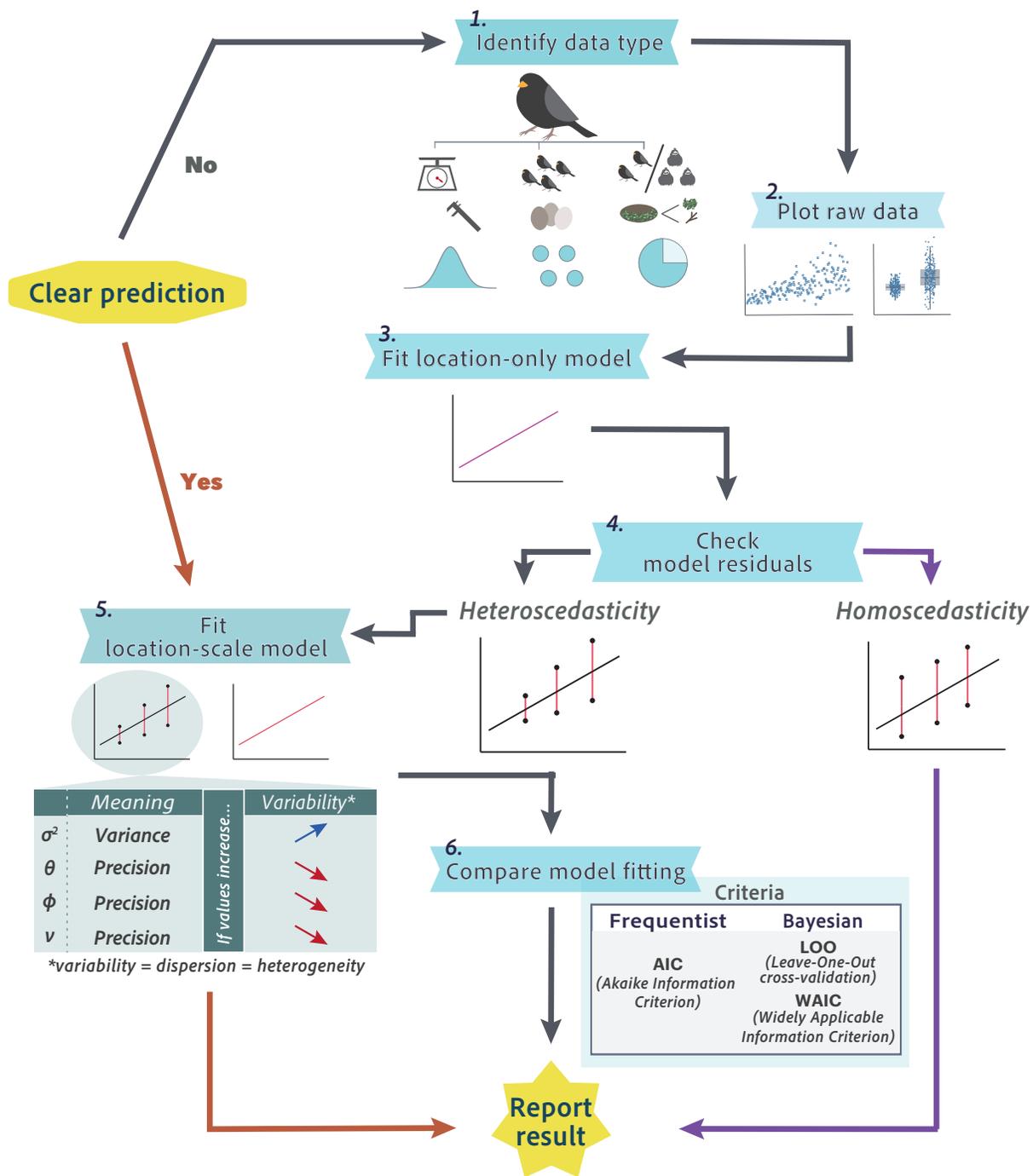


Figure 2