

Topological equivalence of stomata distribution patterns across vascular plants

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Summary

Stomata are ancient anatomical structures on leaves that regulate the exchange of water vapor, oxygen, and carbon dioxide between plants and the atmosphere. Acting as valve-like gateways between internal tissues and the external environment, stomata may function as locally interacting networks. Theoretical and experimental evidence suggests that local interactions among neighboring stomata influence their function and spatial arrangement. From this perspective, analyzing stomatal distributions as networks may yield novel insights into observed spatial patterns and their generative mechanisms. We hypothesize that variability in stomatal arrangements arises from shared underlying rules, with observed diversity reflecting an epiphenomenon. To test this, we employed a multi-species, multi-site common garden approach to assess potential convergences in stomatal distribution. A network-based framework enabled us to reduce individual-level variability and analyze stomatal patterns as interacting systems. Our results show that, across species and environments, stomatal spatial configurations consistently align with a null model linking minimum spanning tree (MST) length to stomatal density. Although a variety of patterns were present, over-dispersed arrangements predominated. These findings suggest that physical constraints during stomatal development impose strong limits on the range of viable spatial configurations that can evolve.

Keywords: stomata patterning, minimum spanning tree, scaling, leaf architecture, spatial networks

Introduction

Stomata are ancient anatomical structures on leaves that regulate the exchange of gases—water vapor, oxygen, and carbon dioxide—between plants and the atmosphere (Edwards et al., 1998; Croxdale, 2001; Hetherington & Woodward, 2003; Bergmann, 2004; Matkowski & Daszkowska-Golec, 2023). Traits such as stomatal size, density, shape, and spatial distribution vary widely among individuals, species, and environments. For example, stomatal size (often measured as pore length) has been found to correlate weakly with latitude, although the specific environmental drivers behind this pattern remain unclear (McKown et al., 2014). Results from common garden experiments suggest that pore size may co-vary with climatic conditions (Carlson et al., 2016). These findings support the idea that stomatal traits exhibit a degree of plasticity, with potentially important implications for plant physiological performance (Drake et al., 2013).

Stomatal density is the simplest and most commonly used descriptor of their spatial distribution on the leaf surface. For instance, McKown et al. (2014) reported that stomatal density in *Populus* species is negatively correlated with both temperature and precipitation across a latitudinal gradient. In contrast, Fontana et al. (2017) observed a slightly positive relationship between stomatal density and rainfall in *Salix miyabeana*, highlighting the complexity of environmental influences. Beyond temperature and precipitation, other factors also appear to modulate stomatal density. Antunes de Almeida Filho et al. (2017) found that the distance between neighboring stomata, likely influenced by overall density, increased with light exposure, suggesting that light availability can also shape stomatal spatial patterns.

Stomatal spatial arrangement has traditionally been described as uniform, with distinct “stomata-free” regions surrounding each stoma (Sachs, 1991). However, Naulin et al. (2017) demonstrated that this perception may be an artifact of the point-pattern analysis methods commonly employed in such studies. By applying a more realistic disc-pattern analysis, they showed that stomatal distributions can span the full theoretical spectrum—from uniform to random to clustered patterns. To fully understand stomatal organization and its underlying mechanisms, spatial patterning must be explicitly analyzed. Various biological (e.g., genetic) and physico-chemical mechanisms, such as activation–inhibition processes, appear to regulate stomatal spacing to avoid configurations that impair stomatal function (Serna & Fenoll, 1997; Illian et al., 2008; Dow et al., 2013; Fanourakis et al., 2015; Fiorin et al., 2015).

Thus, stomatal distribution can be considered an anatomical trait subject to strong ecological and evolutionary pressures (e.g., Fiorin et al., 2015; Xiong & Flexas, 2020).

Variability in stomatal spatial arrangements may—or may not—have an evolutionary basis. Evidence from common garden experiments indicates that intraspecific variation in stomatal density can be adaptive. For example, Carlson et al. (2016) identified a positive selection gradient favoring individuals with stomatal traits suited to drier and hotter environments. At the interspecific level, Yang et al. (2023) analyzed 90 species and found that, depending on the spatial scale, both aggregated and uniform (i.e., over-dispersed) stomatal distributions exhibit weak phylogenetic signals, suggesting a partial influence of evolutionary history. While these studies offer valuable insights, the relative contributions of evolutionary and environmental factors in shaping stomatal density and distribution remain unclear. Zhang et al. (2012), for instance, reported species-specific responses: in some species, stomatal traits were predominantly shaped by environmental conditions, whereas in others, genetic control was more pronounced.

The spatial and temporal context in which a leaf develops plays a crucial role in shaping stomatal density and distribution. Even today, variation in these traits is used to infer past environmental conditions, particularly in palaeobotanical records (e.g., Uhl & Kerp, 2005). Conceptually, stomata function as windows that connect internal leaf tissues with the external environment. Each stoma can be thought of as a valve-like structure that regulates gas exchange within a specific region of the leaf surface (Clark et al., 2022). Both theoretical and experimental studies have shown that stomatal function is influenced by neighboring stomata (Haefner et al., 1997; Mott et al., 1997). These local interactions suggest that stomatal arrangements operate as locally interacting networks. If this is the case, analyzing stomatal distributions and their structural properties through a network-based framework could offer novel insights into the spatial patterns observed and the underlying generative mechanisms. We therefore hypothesize that the apparent variability in stomatal distributions arises from shared generative rules, and that the diversity of observed patterns may simply be an epiphenomenon.

In this study, we apply a multi-species, multi-site common garden approach to identify potential commonalities in observed stomatal distribution patterns. By using a network-based

framework, we minimize the confounding effects of individual variability and treat stomatal arrangements as interacting biological systems.

Methods

Biological data

Sixty-six plant species, spanning four classes, 27 orders, and 41 families, were sampled from two arboreta: the Antumapu Arboretum (hereafter Antumapu; 33.57°S, 70.63°W) and the Frutillar Arboretum (hereafter Frutillar; 41.12°S, 73.03°W), both managed by the Faculty of Forestry and Nature Conservation, University of Chile (Table S1). These sites are approximately 980 km apart. Antumapu is located in a warm-summer Mediterranean climate (Köppen classification: Csb), with average summer and winter temperatures of 19°C and 9.5°C, respectively, and an annual precipitation of 371 mm (Santibañez, 2017). In contrast, Frutillar is situated in southern Chile within a temperate oceanic climate (Köppen classification: Cfb), characterized by average summer and winter temperatures of 14.3°C and 7°C, respectively, and an annual precipitation of 1516 mm (Santibañez, 2017).

For each species, three fully developed leaves were collected. Leaves were cleared following the protocol described by Castellaro et al. (2007) and mounted on glass slides for microscopic examination and imaging. After processing, a total of 180 samples were available for analysis. Anatomical measurements were performed using images of approximately 1 mm² from the central third of the abaxial leaf surface. To estimate spatial properties, the centroid of each stomatal complex was determined using Cartesian coordinates in ImageJ software (Rasband, 1997; Schneider et al., 2012). Additionally, the maximum and minimum diameters of 30 stomata were measured per sample, and the total number of stomata was recorded (Table S1).

Network reconstruction

As noted earlier, local interactions between stomata suggest that their spatial arrangements function as locally interacting networks. Our goal, therefore, is to define a method for constructing a graph that best represents these interactions among stomata. Based on the idea that each stoma connects a specific region of the leaf surface to the atmosphere, we estimated

164 this area using Voronoi polygons (Aurenhammer et al., 2013). In a Voronoi diagram, each
165 polygon is centered on a stoma, and all points within that polygon are closer to the central
166 stoma than to any other in the sampled area.

167
168 To represent potential local interactions, we connected neighboring polygons by linking the
169 centroids of adjacent stomata. This process generates a Delaunay triangulation—the dual
170 graph of the Voronoi diagram—which connects each stoma to its nearest neighbors, forming
171 a dense network in which each node (stoma) typically has a degree of approximately six
172 (Aurenhammer et al., 2013).

173
174 Within the Delaunay triangulation lies a key subgraph known as the Minimum Spanning
175 Tree (MST). The MST is a loop-free graph that connects all nodes while minimizing the total
176 edge length (Prim, 1957). In the context of complex spatial patterns, the MST is often used to
177 characterize dominant connectivity structures. The MST has a long history of applications in
178 biology and ecology, and its topological properties are well-documented (Dussert et al., 1986,
179 1987; Cantwell & Forman, 1993; Jones et al., 1996; Wallet & Dussert, 1997; Bunn et al.,
180 2000; Urban et al., 2001; Dry et al., 2012). More recently, Liu et al. (2021) applied MST-
181 based methods to the analysis of stomatal arrangements, making it an ideal starting point for
182 identifying common structural features of stomatal distribution across plant species—
183 particularly in light of the trade-off between epidermal space use for photosynthesis and
184 water conservation (Lawson & McElwain, 2016).

185
186 MSTs exhibit a range of topological properties that can be used to classify spatial
187 arrangements of objects. Metrics such as total MST length, mean edge length, and the
188 standard deviation of edge lengths are commonly used to characterize spatial configurations
189 (Hoffman & Jain, 1983; Dussert et al., 1989). Under the assumption of complete spatial
190 randomness (i.e., a Poisson process) in two dimensions, the expected MST length approaches
191 the following limit:

$$192 \quad L_{mts} = \beta (N^*A)^{1/2} \quad (1)$$

193
194
195 where N is the number of points (stomata), A is the area of observation, and β is a constant
196 that depends on the spatial structure of the distribution. Empirical estimates place β between
197 0.63 and 0.64 under random conditions (Beardwood et al., 1959; Bertsimas & Van Ryzin,

1990; Avram & Bertsimas, 1992; Jaillet, 1993; Cortina-Borja & Robinson, 2000). Values of β lower than this range indicate clustering, while higher values suggest over-dispersion.

In this study, we calculated the MST for each sample as a representation of the stomatal network. For each MST, we measured the total length and evaluated deviations from the null model (Equation 1) to infer differences in spatial arrangement.

To contextualize each sample, we compared its MST length to three simulated benchmark patterns: clustered, random (Poisson), and over-dispersed. Simulated patterns used the same number of stomata and observation area as the empirical sample. Clustered patterns were generated using four spatial clusters, while over-dispersed patterns applied a minimum inhibition distance equal to the mean stomatal diameter of the sample. Each scenario was simulated 1,000 times.

All analyses were conducted in R (2024), using the packages *stpp* (Gabriel et al., 2024), *netgen* (Bossek, 2025), and *emstreeR* (Quadros, 2025).

Results

Stomatal density across samples ranged from 3 to 777 stomata per mm². The lowest density was observed in a sample of the fern *Asplenium trilobum*, while the highest was recorded in *Cryptocarya alba* (Lauraceae). In general, higher stomatal densities were found in samples from the Mediterranean site (Antumapu), whereas stomatal diameter showed no consistent pattern between the two sites (Fig. 1; Table S2).

The same samples that exhibited the lowest and highest stomatal densities also showed the minimum and maximum MST lengths, with values ranging from 858 μm to 20,894 μm . As expected, there was a strong positive correlation between the number of stomata per sample and the total MST length (Pearson's $r = 0.945$; 95% CI: [0.927, 0.959]). Additionally, stomatal number was significantly negatively correlated with average stomatal diameter ($r = -0.568$; 95% CI: [-0.659, -0.459]).

Despite the three orders of magnitude variation in stomatal density, MST length across all samples was accurately predicted as a function of stomatal density, with a high coefficient of

determination ($R^2 = 0.94$; Fig. 1). However, the estimated value of the scaling parameter β was significantly higher than expected under complete spatial randomness. Specifically, we obtained a mean β of 0.7492 (95% CI: [0.7393, 0.7591]), which exceeds the typical range reported for Poisson-distributed patterns (0.63–0.64), suggesting a consistent tendency toward over-dispersion in stomatal spatial arrangements.

When comparing observed MST lengths against simulated distributions, 2.8% of the samples (5 out of 180) were classified as clustered, 27.2% (49 samples) as random, and 32.8% (59 samples) as over-dispersed. A total of 49.4% of the samples (89 samples) did not fall exclusively within any single category. This is due to overlapping confidence intervals, which allowed some samples to be classified into multiple categories.

Among the unclassified samples, 10% (18 samples) fell between the confidence intervals of the random and over-dispersed simulations, while 39.4% (71 samples) exceeded the upper limit of the over-dispersed range. In total, 72% of the samples (130 out of 180) either fell within or above the over-dispersed range, and 82% (148 samples) fell above the random distribution range.

No clear taxonomic pattern was observed in relation to the type of spatial arrangement. Full classification results for each sample are provided in Figures 2 and 3 and Table S2.

Discussion

Trade-offs and relationships among stomatal size, density, and spatial arrangement have garnered increasing attention over the past decade (e.g., Beerling & Franks, 2010; Brodribb et al., 2014). The diverse patterns reported in the literature appear to result from a still poorly understood interplay of genetic and environmental factors (Zhang et al., 2012). In this study, we collected data from a broad range of species growing in two ecologically contrasting environments. The stomatal density values observed in our samples were consistent with previous findings (Franks & Beerling, 2009; Haworth et al., 2023). Notably, the highest stomatal density and MST length were recorded in samples from the Mediterranean site (Antumapu) (Fig. 1), supporting the hypothesis that increased stomatal density under higher temperatures may represent an adaptive strategy to enhance gas exchange and evaporative cooling (Hill et al., 2015).

The strong positive correlation between stomatal density and MST length aligns with theoretical expectations from our model (Eq. 1): a greater number of nodes necessarily results in a longer minimum spanning tree (Beardwood et al., 1959; Cortina-Borja & Robinson, 2000). Furthermore, our findings are consistent with prior studies reporting a significant negative correlation between stomatal density and stomatal size across species (Franks & Beerling, 2009; Haworth et al., 2023).

However, when we focused on the subset of species present at both sites, we observed a more nuanced pattern (Table S2). With only one exception (*A. chilensis*), all species exhibited higher stomatal densities in the Mediterranean climate (Antumapu) compared to the temperate oceanic climate (Frutillar). In contrast, stomatal size did not follow a consistent trend across environments. These results suggest that, at the intraspecific level, stomatal density may be more responsive to environmental conditions than stomatal size—indicating a greater degree of plasticity. Nevertheless, this interpretation is based on a limited sample of seven species, and further studies with larger species pools are needed to confirm this pattern.

Although our dataset includes numerous species from diverse biogeographical origins and two ecologically distinct environments, the spatial configuration of stomata across all samples was well-described by the theoretical model (Eq. 1; Fig. 1). The strong fit of this model suggests that, across a wide range of lineages and taxonomic groups, stomatal arrangements adhere to a shared generative rule—one in which MST length scales with the square root of stomatal density and a constant parameter (β) representing spatial inhibition or repulsion among stomata. This result implies a form of topological equivalence across species and environments: the observed spatial patterns can be interpreted as scale transformations—shrinking or stretching—of a common underlying configuration.

At the spatial resolution of our analysis ($\sim 1 \text{ mm}^2$), no major deviations from this model were observed between species or sites. The estimated β value (>0.74) points to consistent repulsion or inhibition among stomata, aligning with both the "one-cell spacing" rule (Lawson & McElwain, 2016) and the finite-size constraint imposed by the physical dimensions of stomata themselves (Naulin et al., 2017). Our simulation results further support this interpretation. Although we examined a limited set of point process scenarios (single configurations for both clustered and over-dispersed patterns), the majority of

300 samples (72%) exhibited levels of over-dispersion equal to or greater than those generated by
301 the over-dispersed simulations (Figs. 2–3).

302
303 As previously discussed, two main components likely contribute to this over-dispersion. First,
304 stomata are not point-like objects—their finite size prevents overlap and inherently induces
305 spatial repulsion (Naulin et al., 2017). Since our simulations are based on idealized point
306 processes, part of the observed over-dispersion likely reflects this geometric constraint.
307 Second, and more importantly, 39% of samples displayed levels of over-dispersion that
308 exceeded our simulation range even after accounting for stomatal size. This suggests the
309 presence of active biological mechanisms, likely of genetic origin, that enforce spatial
310 inhibition during stomatal development—further supporting the view that stomatal patterns
311 are shaped by endogenous regulatory processes in addition to biophysical constraints.

312
313 If our model accurately reflects the mechanisms governing stomatal spatial organization, it
314 carries several important implications—both quantitative and qualitative—for understanding
315 how stomata can be distributed on the epidermis. At low densities, there are potentially
316 thousands of viable spatial configurations that achieve a functional balance in the trade-off
317 between maximizing photosynthetic area and minimizing water loss. However, evolutionary
318 increases in stomatal density not only force reductions in stomatal size (Franks & Beerling,
319 2009; Haworth et al., 2023), but also dramatically constrain the number of feasible spatial
320 arrangements. Beyond a certain threshold, only over-dispersed configurations remain viable.

321
322 This constraint arises from fundamental geometric principles: the number of possible
323 arrangements of finite-sized objects in a fixed area is far more limited than for point-like
324 objects (Simberloff, 1979; Wu et al., 1987). As stomata cannot be infinitely small, increasing
325 their number while preserving function requires increasingly regular spacing. In the
326 theoretical limit, this constraint leads to a near-regular grid-like distribution.

327
328 It is important to note that these findings apply specifically to the spatial scale analyzed in
329 this study ($\sim 1 \text{ mm}^2$). At broader spatial scales, additional structural constraints—such as
330 those imposed by leaf venation networks—may further shape or override local stomatal
331 arrangements (Matos et al., 2024). Future studies incorporating multiscale analyses and
332 biomechanical modeling may provide deeper insights into how such hierarchical constraints
333 interact across spatial levels.

Stomatal size and density are key determinants of CO₂ conductance in leaves (Franks & Beerling, 2009) and are therefore subject to strong selective pressures to balance mesophyll demand for carbon assimilation with efficient water vapor diffusion (Lawson & McElwain, 2016; Haworth et al., 2023). In a scenario of rising atmospheric CO₂ concentrations, species that already exhibit high stomatal densities may face increasingly limited options for allocating additional epidermal space to stomatal complexes due to geometric constraints. Our results suggest that further increases in stomatal density are only feasible through heightened over-dispersion, progressively driving stomatal arrangements toward grid-like spatial patterns across the leaf surface.

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Author contributions

PIN and SAE conceived and planned the work. PIN performed sample preparation and digitalization. PIN and SAE analyzed the data. SAE wrote the manuscript and both authors commented on the draft.

Competing interests: Authors have no competing interests.

Data availability: all data is available as supporting information

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Figure captions

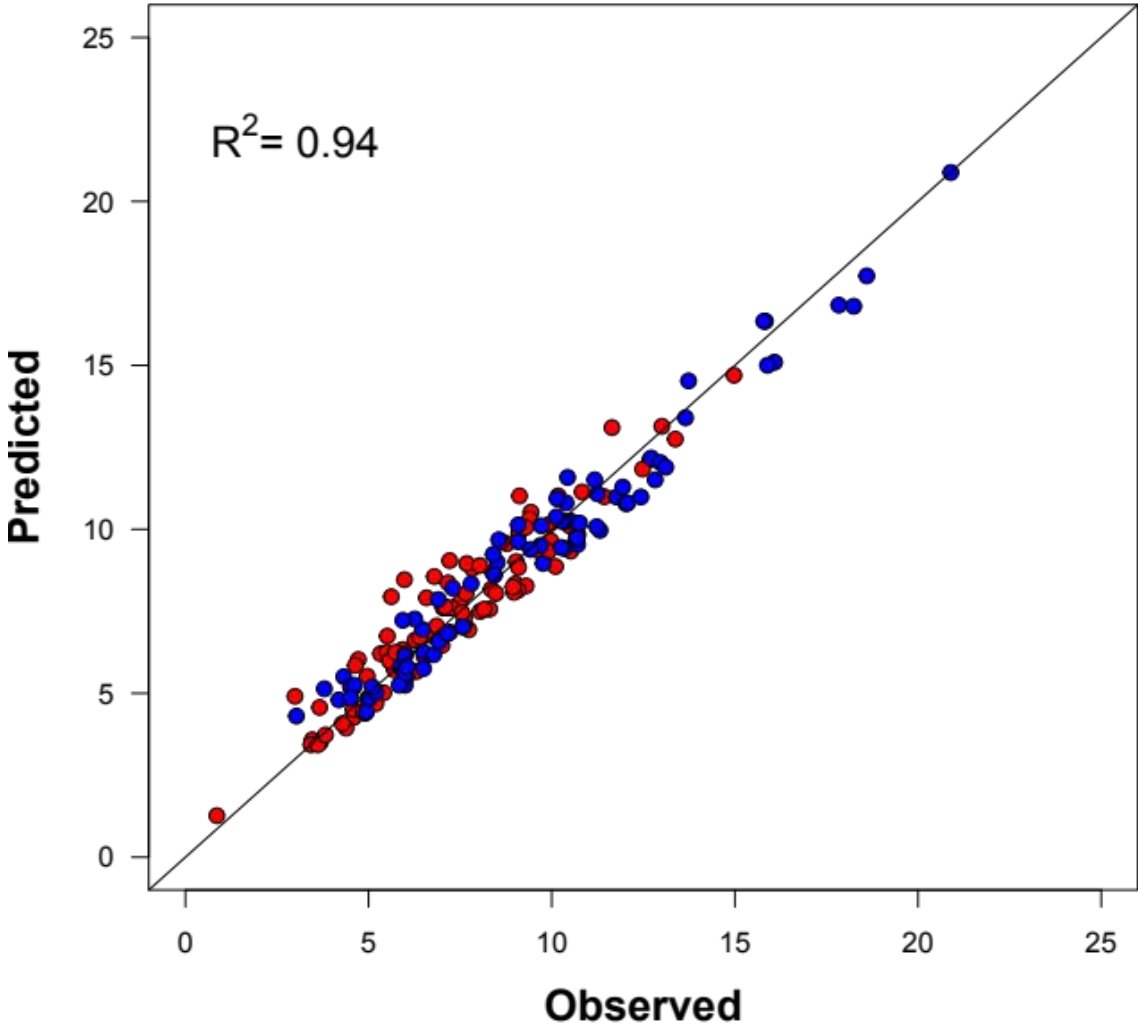
Figure 1. Relationship between observed and predicted MST lengths across all samples.

MST length was rescaled to millimeters. Each point represents a sample, with blue indicating Antumapu and red indicating Frutillar. The diagonal line denotes the 1:1 relationship, representing a perfect fit between observed and predicted values based on the theoretical model.

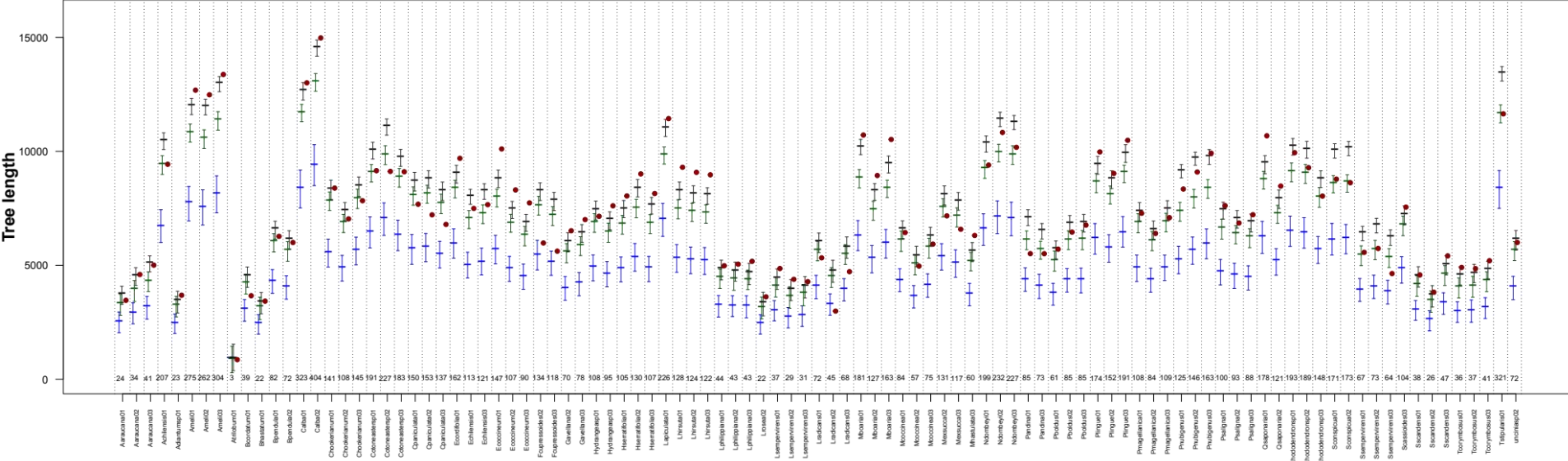
Figure 2. Comparison of observed MST length values (red points) with simulated distributions under clustered (blue), random (green), and over-dispersed (black) spatial scenarios for each sample from the Frutillar site. The x-axis shows sample identifiers, with the number of stomata in each sample indicated by small numbers above the axis. Bars represent the median and 95% confidence interval of the MST length from 1,000 simulations per spatial pattern scenario.

Figure 3. Comparison of observed MST length values (red points) with simulated distributions under clustered (blue), random (green), and over-dispersed (black) spatial scenarios for each sample from the Antumapu site. The x-axis shows sample identifiers, with the number of stomata in each sample indicated by small numbers above the axis. Bars represent the median and 95% confidence interval of the MST length from 1,000 simulations per spatial pattern scenario.

Figure 1.



614 **Figure 2.**
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625 **Figure 3.**

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