1 Macroalgae morphological complexity affects the functional diversity of

2 epifaunal annelid assemblages

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2223 Abstract

Host structural complexity influences the diversity of associated epifaunal species, but 24 25 its role in shaping functional trait diversity remains underexplored. We developed a trait-based framework to assess whether macroalgal structural complexity significantly 26 27 influences the functional assembly of marine annelid epifauna in a sandstone reef 28 system at Enseada dos Corais Beach (NE Brazil). Sampling was conducted in 29 December 2018, February 2019, April 2019, and June 2019. Ten fronds from each of 30 four macroalgal species—Gelidiella acerosa and Palisada perforata (corticated), 31 Padina gymnospora and Ulva lactuca (foliose)-were collected to describe the 32 associated annelid fauna. Structural complexity was guantified using the interstitial 33 space index (ISI), height, and the fractal dimensions of frond area (Da) and perimeter 34 (Dp). Based on body length, feeding strategy, and larval development, the functional 35 trait diversity of annelid assemblages was analyzed using Rao's Quadratic Entropy 36 (Rao's Q) and RLQ analysis. Corticated algae species hosted more functionally dissimilar annelid assemblages than foliose ones. Moreover, morphological traits of 37 macroalgae influenced epifaunal functional trait composition, particularly during the 38 rainy season, when hydrodynamics are more intense. Our findings thus supported the 39 40 hypothesis that increased habitat complexity positively influences functional trait 41 diversity in marine macroalgal phytal communities.

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43 **Keywords:** Trait-Based Approach, Benthos, Polychaeta, Habitat complexity.

44 Introduction

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The critical role of habitat structure in shaping community diversity has been 46 47 recognized for a long time in ecological literature (Tokeshi & Araraki, 2012; Carvalho 48 & Barros, 2017). This concept encompasses both the qualitative and quantitative 49 aspects of spatial structuring, following the paradigm that greater habitat complexity provides more microhabitats and ecological niches, ultimately supporting higher 50 51 biodiversity (Tokeshi & Araraki, 2012; Stein et al., 2014; Carvalho & Barros, 2017; 52 LaRue et al., 2023). Habitat structure is typically described in terms of three key components: scale, heterogeneity, and complexity (Carvalho & Barros, 2017; LaRue 53 et al., 2023). Complexity refers to the multidimensional variation in structural attributes 54 within an environment, while heterogeneity represents a single facet of habitat 55 complexity (Carvalho & Barros, 2017; LaRue et al., 2023). Numerous studies, 56 especially from marine and freshwater systems, have quantified the influence of 57 58 habitat complexity and heterogeneity on species diversity at local scales, consistently revealing a strong positive effect (Dean & Connel, 1987; Christie et al., 2009; Stein et 59 al., 2014; Carvalho & Barros, 2017; Torres-Pulliza et al., 2020). 60

61 Phytal ecosystems, characterized by dense assemblages of macroalgae and 62 macrophytes in shallow coastal waters, play a crucial role in sustaining high ecological productivity and biodiversity (Christie et al., 2009; Stagnol et al., 2013). However, they 63 64 are increasingly threatened by climate change and various human-induced impacts (Stagnol et al., 2013). These vegetated habitats provide shelter and resources for 65 66 multiple animal species, acting as natural architects of habitat structure (Gee & Warwick, 1994; Christie et al., 2009). Such habitat engineers might exhibit a wide 67 68 variety of morphologies, which can be summarized into distinct functional groups 69 based on their chemical, reproductive, and morphological traits (Gee & Warwick, 1994; 70 Steneck & Dethier, 1994; Balata et al., 2011; Gan et al., 2019). Two recurrent 71 architectural groups have been recognized based on macroalgae frond morphology: (i) foliose species, with broad blades with none to few branching (Fig. 1a), and (ii) 72 corticated species, with stiff, highly ramified thalli (Fig. 1a). Corticated forms supply a 73 more intricate three-dimensional matrix than their foliose counterparts (Fig. 1a) 74 (Steneck & Dethier, 1994; McAbendroth, 2005; Dibble & Thomas, 2006; Gan et al., 75 2019; Craveiro & Rosa-Filho, 2024). 76

77 Dean and Connell (1987) proposed three non-exclusive mechanisms by which increasing algal complexity (Fig. 1a) can raise the diversity of resident epifauna: (i) the 78 79 protection effect – complex fronds block visual or tactile detection by predators, reducing predation-induced mortality; (ii) the sheltering effect - interstices dampen 80 physical stressors such as wave action; (iii) the filtering effect – intricate matrices slow 81 82 water flow, trapping larvae or suspended food particles and enhancing colonization. These mechanisms are linked to macroalgae morphology, playing key roles in the 83 assembly process of epifaunal communities, as they can buffer the effects of 84 85 environmental severity, such as hydrodynamics, and negative biological interactions, 86 including predation and resource competition (Dean & Connell, 1987; Christie et al., 2009). 87

Previous studies have shown that higher structural complexity of host 88 89 macroalgae supports greater epifaunal abundance, diversity, and biomass (Gee & 90 Warwick 1994; Veiga et al. 2014; Pérez-García et al. 2015; Gan et al. 2019; Waren et al. 2019; Duarte et al. 2020a, b; Craveiro & Rosa-Filho 2024). However, only a few 91 investigations have tested this prediction using a functional trait-based approach, and 92 93 those indicate that host complexity positively influences epifaunal functional diversity 94 (Barbosa et al., 2019; Duarte et al., 2020a; Katsiaras et al., 2022). Moreover, this 95 relationship may exhibit important nuances because a more complex corticated 96 architecture can impose spatial constraints on larger-bodied adult epifaunal species 97 (Dean & Connell, 1987; Gee & Warwick, 1994). Therefore, a well-designed study 98 employing specific functional traits linked to hypotheses about the role of structural complexity could enhance our understanding of how macroalgal morphology 99 100 influences the assembly of associated communities. By varying experimental designs, focal species, and geographic regions, such research would allow broader 101 102 generalization of patterns in this critical component of coastal marine ecosystems.

Marine annelids, particularly polychaetes, are among the most diverse groups inhabiting the ocean floor, performing many ecological functions and exhibiting a remarkable array of forms and life strategies (Rouse et al., 2022). Notably, these animals are prevalent in macroalgal epifaunal communities (Bailey-Brock et al., 1980; Rossbach et al., 2021). Their functional traits - defined as morphological, phenological, and physiological traits indirectly or directly related to fitness (Violle et al., 2007) - have been employed to discern ecological patterns across various environmental gradients,
contributing to the overall understanding of the assembly process in coastal systems
(Wouters et al., 2018; Morais et al., 2019; Nogueira et al., 2023; Medeiros et al., 2021;
Katsiaras et al., 2022; Mendes et al., 2025).

113 We analyzed a dataset of epifaunal annelid assemblages associated with four distinct macroalgal species from a tropical phytal ecosystem in a bedrock reef 114 115 formation called Enseada dos Corais (South Atlantic, NE Brazil). Building upon the well-documented roles of macroalgae architecture in providing "protection", "filtering", 116 117 and "sheltering" to their associated fauna (Dean & Connell, 1987; Christie et al., 2009). We hypothesize that macroalgae complexity will favour a more diverse set of epifaunal 118 functional traits configurations (Fig. 1). Specifically, we anticipate that structurally 119 120 complex corticated macroalgae will support annelid assemblages exhibiting a broader 121 range of sizes, feeding strategies, and reproductive traits when compared to the structurally simple foliose macroalgae, which we expect to favour narrower trait 122 123 configurations related to the greater exposure to the external environment and the 124 weaker capacity in retaining nutrients on their fronds (Fig. 1b). Hence, by examining 125 the relationship between the structural complexity of macroalgae and the functional 126 trait diversity of associated annelid assemblages, our study aims to contribute a

deeper understanding of how habitat complexity influences community structure inmarine ecosystems from a functional trait perspective.



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Figure 1 The morphological characteristics of corticated and foliose macroalgae 130 represent two extremes of a gradient of complexity and fractality, with significant 131 implications for the taxonomic diversity of associated epifaunal communities. 132 Corticated algae, with their more intricate and structurally complex features, support a 133 more diverse array of epifauna. This raises the question of whether a similar 134 relationship exists when viewed from the perspective of functional traits (a). We expect 135 that the functional trait diversity of marine annelids associated with corticated 136 macroalgae should display greater dissimilarity, as foliose algae offer fewer resources 137 and protection when compared to corticated counterparts (b) 138

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141 Material and methods

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- 143 Data collection
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145 Samples of two corticated macroalgae, Gelidiella acerosa and Palisada perforata, and two foliose macroalgae, Padina gymnospora and Ulva lactuca, were 146 randomly collected during four sampling periods: December 2018, February, April, and 147 June 2019. Collections were conducted at Enseada dos Corais (8°19'09.6" S, 148 34°56′53.7″ W) in northeastern Brazil (Fig. 2). This site is a 3-km-long coastal area 149 150 characterized by sandstone (beachrock) reefs parallel to the shoreline (Vasconcelos 151 et al., 2013). The region has a tropical climate with two distinct seasons: a dry season 152 from September to February and a rainy season from March to August. Environmental

- 153 conditions include a mean water temperature of 27 °C, salinity levels around 36, high
- dissolved oxygen concentrations, and low turbidity (Domingues et al., 2017).



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Figure 2 Study area map. (a) Overview of South America highlighting the location of
 Pernambuco State in northeastern Brazil. (b) Detailed map of Pernambuco showing
 the Enseada dos Corais sampling site (c), indicated by the yellow point where
 macroalgal fronds were collected

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At each sampling time, ten fronds from each macroalgae species were collected. Before detaching the algae from the substrate, fronds were enclosed in a plastic bag to prevent the escape of the motile fauna. The specimens were then preserved in 4% saline formalin buffered with sodium tetraborate. In the laboratory, the samples were rinsed in fresh water and shaken multiple times to dislodge associated organisms. The resulting water was passed through a 0.3 mm mesh sieve to capture the epifaunal annelids. The fronds were then placed on a sheet of white paper, spread out to their full extent, and pressed. The fronds were subsequently dried in an oven at 60 °C for 72 hours. After drying, each frond was removed from the botanical press and photographed using a Nikon Coolpix AW100 digital camera. The photographs were analyzed using the ImageJ software to measure the Interstitial Spatial Index (ISI), height (cm), fractal dimensions of the area (Da) and perimeter (Dp) on Image J software (Scheider et al., 2012).

The Interstitial Space Index (ISI) was calculated following the Dibble and 174 175 Thomaz (2006) method. Briefly, two vertical black dashed lines, one orange dotted 176 line, and three horizontal black dashed lines were superimposed on each image to 177 delineate the upper, middle, and lower sections of the frond, and the interstitial spaces 178 within the macroalgae were quantified along these lines (Craveiro & Rosa-Filho, 179 2024). Specifically, the index was calculated using the formula: ISI = fh/lh + fv/lv, where 180 fh is the average frequency of interstices intercepted per centimeter along the 181 horizontal axis, lh is the average length of interstices along the horizontal axis, fv is 182 the average frequency of interstices intercepted per centimeter along the vertical axis. 183 and lv is the average length of interstices along the vertical axis (Dibble & Thomas, 2006; Craveiro & Rosa-Filho, 2024). 184

Macroalgae height was calculated by setting a central line (base to apex) on 185 186 each image (Craveiro & Rosa-Filho, 2024). Finally, regarding the fractal dimensions, 187 Da represents the measure of the area covered by the macroalgae, which is an 188 estimate of area occupancy of its fronds, while Dp indicates the perimeter area of the 189 macroalgae, which means the degree of dissection of its fronds (Haley et al., 2004; 190 McAbendroth et al., 2005). Fractal dimensions were calculated following the methods 191 of McAbendroth et al. (2005) and Kovalenko et al. (2009), using the box-count 192 algorithm from Image J (Craveiro & Rosa-Filho, 2024).

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194 Functional traits

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A functional trait matrix was constructed through fuzzy-coding of body size, feeding strategy, and reproduction traits of annelid epifaunal genera (Table 1). The scores varied from 0 (no affinity), 1 (low affinity), 2 (high affinity), and 3 (absolute affinity, i.e., when all other modalities are 0 scored), following Oug et al. (2012) coding criteria. Annelid size was assessed following Jumars et al. (2015) body length trait 201 modalities, coded after a generic-level literature consult. Regarding the feeding 202 strategy trait, we follow the guidelines of Jumars et al. (2015) and Wouters et al. 203 (2018). Feeding trait modalities, whenever possible, were also coded based on the 204 generic-level literature. When diet information was unavailable at a generic level, 205 family-level literature was consulted for additional information. Larval development 206 was assessed based on Rouse (2000) and updated family- and genus-level literature 207 (Rouse et al., 2022).

208 The fuzzy-scores for each trait were calculated using the prep.fuzzy function 209 from the ade4 R package (Dray & Dufour, 2007). Then, a Gower distance matrix was 210 calculated based on annelid genera's fuzzy-coded traits. This matrix, alongside the 211 abundance of each genus per macroalgae frond matrix, was used to calculate Rao's 212 Quadratic Entropy (Rao's Q) index of each epifaunal assemblage using the "melodic" 213 function (de Bello et al., 2016). Rao's Q measures trait dispersion by quantifying the mean dissimilarity among epifaunal genera in each assemblage, summarizing the 214 expected differences between randomly selected species pairs with replacement 215 216 (Ricotta & Moretti, 2011; de Bello et al., 2016).

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218 Data analysis

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220 The macroalgae traits were compared between morpho-functional groups 221 (corticated vs foliose) and among months ("December/18", "February/19", "April/19", 222 and "June/19") using a Permutational Analysis of Variance (Permanova). Moreover, 223 for the linear modelling step described below, a Principal Component Analysis (PCA) 224 of scaled macroalgae traits and a Pearson correlation test were implemented to check 225 for multicollinearity. The Rao's Q of epifaunal communities was modelled against the 226 interaction between macroalgae functional groups (fixed factor with two levels: "corticated" and "foliose") and months (fixed factor with four levels: "December/18", 227 "February/19", "April/19" and "June/19"). 228

Macroalgae traits related to their morphological complexity (Da, Dp, Height, and ISI) were fitted as predictors of Rao's Q in a global model. A multimodel inference approach, combining model selection and model averaging, was applied to determine which macroalgae traits were included in the best-fitting models for explaining variation in epifaunal Rao's Q values, using the MuMIn package in R (Burnham & Anderson, 2002; Bartoń, 2024). The model with the lowest Akaike Information Criterion corrected 235 for small sample sizes (AICc) was considered the best approximating model for 236 predicting Rao's Q variation (Burnham & Anderson, 2002). To evaluate the importance 237 of each predictor (macroalgae trait) and estimate their averaged effects, we selected 238 all models within $\Delta AICc < 2$ units of the first-ranked model (Burnham & Anderson, 239 2002; Symonds & Moussalli, 2011; Tredennick et al., 2021). The importance of a given predictor was quantified as the sum of Akaike weights (AICw) across all models in 240 241 which it appeared, representing the probability that the predictor is part of the best approximating model (Burnham & Anderson, 2002; Galipaud et al., 2013). Each model 242 243 weight (AICwi) was calculated as the relative likelihood of the model "i" divided by the 244 sum of the likelihoods across all selected models (Burnham & Anderson, 2002; 245 Galipaud et al., 2013).

246 Separately, to investigate specific correlations between annelid and 247 macroalgae traits, RLQ and fourth-corner analysis were employed. RLQ and fourthcorner analyses were conducted using the ade4 package to investigate potential 248 249 associations between functional traits and macroalgae traits (Dray et al., 2014). The 250 RLQ analysis integrates three matrices: R (scaled macroalgae traits), L (genera 251 abundances), and Q (fuzzy-coded functional traits), enabling the identification of 252 multivariate relationships between environmental gradients (in our case macroalgae 253 traits) and annelid functional traits, mediated by annelid genera abundances (Dray et 254 al., 2014). The fourthcorner analysis complements this approach by testing the 255 significance of bivariate associations between annelid traits and macroalgae traits 256 (Dray et al., 2014). Each matrix was individually processed using appropriate 257 multivariate analyses. The Q matrix was analyzed using a Fuzzy Correspondence 258 Analysis (FCA), while the R and L matrices were subjected to Principal Component 259 Analysis (PCA) and Correspondence Analysis (CA), respectively. Finally, a Monte 260 Carlo permutation test, with 49999 repetitions within model 6, was implemented to assess the significance of correlations between macroalgae traits and annelid traits 261 with RLQ axes, controlling for p values using the false discovery rate (FDR) method 262 263 (Benjamini & Hochberg, 1995; Dray et al., 2014). All analyses were performed in R with R Studio (R Core Team, 2023). 264

- 265
- 266 Results
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268 Most morphological complexity traits of macroalgae (Da, Dp, Height, and ISI) 269 differed significantly between the two morpho-functional groups (Table S1; Figs. S1-270 S2) and also among months (Table S1; Fig. S3), with particularly marked differences 271 in June (Fig. S3). In most months, corticated and foliose macroalgae had distinct 272 values of Da and Dp, except in June, when both groups had maximum values and no 273 longer differed (Fig. S2a-b). ISI values remained relatively consistent throughout the 274 year, although the ISI of corticated macroalgae notably increased in June, becoming more subdivided than in previous months (Fig. S2c). Corticated and foliose algae had 275 276 similar height throughout most of the year, except in June, when corticated algae were 277 taller while foliose algae were shorter (Fig. S2d).

278 The two morpho-functional groups of macroalgae supported polychaete 279 epifaunal assemblages with distinct patterns of trait dissimilarity (Table 2, Fig. 3a). Corticated algae generally hosted a more diverse set of epifaunal annelids traits than 280 the foliose ones (Fig. 3a). The mean trait dissimilarity of the epifaunal assemblages 281 did not vary significantly among months (Table 2, Fig. 3a). In addition, model selection 282 283 indicated Da, ISI, and Height as predictors of Rao's Q in the best approximating model 284 (Table 3). The ISI was positively related to Rao's Q, whereas Da and Height were 285 negatively related.



286 Figure 3 Predicted values (95% Confidence Interval) of mean functional trait 287 dissimilarity of epifaunal annelid assemblages. Corticated algae supported more 288 dissimilar epifaunal assemblages than foliose algae (a). However, the mean trait 289 290 dissimilarity of epifaunal annelids did not vary significantly across months (a). The 291 macroalgal traits predictors present in the first-ranked model were Da, Height, and ISI 292 (b-d). Da and Height showed a negative relationship with Rao's Q, while ISI was positively associated with it, suggesting that the greater structural complexity of 293 macroalgae influences the trait diversity of its epifauna 294 295

The RLQ analysis demonstrated an evident influence of macroalgae functional 296 297 groups on the traits of epifaunal annelids, with corticated and foliose algae being 298 distinctly separated from each other (Fig. 4a). The first two axes accounted for 99,8% 299 of the variation (axis 1: 98.7%, axis 2: 1.7%) (Table S2). Axis one distinguished corticated macroalgae (mostly positively associated) from foliose macroalgae 300 301 (primarily negatively associated), but with both groups exhibiting some degree of overlap over time (L correlation = 0.435). However, observations from June (Fig. 4a) 302 formed a topological group apart from those of other months, a pattern more evident 303 304 along the second axis (L correlation = 0.24). Epifaunal annelid traits were significantly 305 associated only with the first RLQ axis, particularly suspension feeding, herbivory, and 306 predation (Table 4). In contrast, body size and larval development strategies were 307 weakly correlated with RLQ axes 1 and 2 and did not contribute significantly to the 308 observed multivariate pattern (Table 4). Importantly, fourth-corner analysis revealed 309 no significant bivariate correlations between annelid and macroalgal traits.

The macroalgal traits ISI and height were positively related to the first axis, 310 whereas Da and Dp were negatively related (Fig. 4b). Specifically, Da and height were 311 312 more strongly correlated with this axis than Dp and ISI, which were more closely associated with axis 2 (Table S3). Herbivore and facultative suspension-feeding 313 314 nereidids, such as *Platynereis* Kinberg, 1865, and *Pseudonereis* Kinberg, 1865, were 315 the dominant genera, showing a negative association with the first axis (Fig. 4c-d). In contrast, predatory genera, mostly syllids, were positively related (Fig. 4c-d). 316 317 Moreover, the first axis distinguished some foliose algae observations in June from 318 the others, occupying its negative extreme (Fig. 4a). The second axis was negatively correlated with all macroalgae traits, and also distinguished some samples collected 319 320 in June from others, as some observations of corticated algae exhibited high ISI and 321 height values, clearly separating them from their foliose counterparts (Fig. 4a-b). The 322 higher ISI and height associated with the June observations on the second axis 323 coincided with the opportunistic/scavenger and deposit-feeding strategies of epifaunal 324 annelids (Fig. 4c).



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Figure 4 RLQ ordinations illustrating changes in epifaunal assemblages associated 326 with corticated and foliose macroalgae (a), as revealed by the correlation of 327 macroalgal morphological traits (b) with the abundance and functional traits of 328 epifaunal annelid genera (c-d). Functional traits of epifaunal annelids that showed 329 330 significant correlations with RLQ axis 1 are highlighted in red. Abbreviations = Ddir: 331 Direct development; Dlc: Development through lecithotrophic larval stage; Dpl = Development through planktotrophic larval stage; Fso = Opportunist/scavenger 332 feeding strategy; Fsp = Predatory feeding strategy; Fsh = Herbivore feeding strategy; 333 Fssf = Suspension-feeding strategy; Lgl = Large body length; Lgm = medium body 334 335 length; Lgs = small body length

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338 Discussion

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340 It was hypothesized that the two distinct macroalgal functional groups would 341 provide contrasting habitats for their associated annelid assemblages, favouring 342 distinct epifaunal trait configurations. Our findings indeed showed that Rao's Q varied significantly between the two macroalgal functional groups, responding to differences 343 in their morphological complexity. In addition, as demonstrated by the RLQ analysis, 344 corticated and foliose macroalgae presented separate affinities to annelid traits and 345 346 genera, corroborating the initial expectations, as well. The assessed macroalgae morphological traits as predictors of Rao's Q of annelid assemblages captured 347 348 relevant aspects of the assembly process at the frond scale. Among the measured traits of macroalgae, Da, ISI, and height were selected in the best approximating 349 350 model, with Da and ISI presenting higher importances than height.

351 Since Da represents a measure of fractality, expressed as total frond 352 occupancy area, fronds with higher Da values are less subdivided than those with 353 lower Da values (McAbendroth et al., 2005). On the other hand, ISI represents how 354 much macroalgae's fronds are subdivided, with higher ISI values representing fronds with numerous interstitial spaces (Dibble & Thomas, 2006). Thus, the structural 355 356 complexity of macroalgae influenced the trait dispersion of their annelid assemblages, and this finding is consistent with the well-documented role of morphological 357 358 complexity of hosts in shaping epifaunal taxonomic diversity in freshwater and marine 359 ecosystems (Dean & Connell, 1987; Gee & Warwick, 1994; Chemello & Milazzo, 2002; Hansen et al., 2010; Hansen et al., 2011; Veiga et al., 2014; Gan et al., 2019; Fraser 360 361 et al., 2020; Duarte et al., 2020a,b; Craveiro & Rosa-Filho, 2024).

362 Under this paradigm, the combination of the RLQ approach with Rao's Q 363 modelling demonstrates an interesting tradeoff of epifaunal trait combinations as a 364 response to the fractal nature of host macroalgae. Although the more complex 365 corticated architecture is expected to impose spatial restrictions on adult epifaunal 366 species with larger bodies (Dean & Connel, 1987; Gee & Warwick, 1994), by 367 displaying higher ISI and lower Da values, they presented assemblages with more dissimilar feeding trait configurations, and not necessarily smaller body length. As for 368 369 foliose species, the lower Rao's Q values agree with prior expectations, as they do not 370 impose size restrictions to their epifauna, but are less effective in capturing suspended 371 material and in providing protection (Dean & Connel, 1987; Gee & Warwick, 1994), 372 ultimately leading to the observed narrower set of feeding trait affinities. A similar 373 situation was also observed in previous studies, relating to feeding strategies and body 374 size traits, with complex macroalgae presenting higher functional diversity of mollusk 375 assemblages (Barbosa et al., 2019; Duarte et al., 2020a).

376 Such pattern is expected because of the positive effects of macroalgae and 377 macrophytes structural complexity on the diversity of their epifaunal communities, 378 which act by influencing the space availability for foraging, colonization, and refuge 379 (Gregg & Rose, 1982; Dean & Connell, 1987; Hacker & Steneck, 1990; Gee & 380 Warwick, 1994; Christie et al., 2009; Barbosa et al., 2019; Ware et al., 2019; Duarte et al., 2020a). More structurally complex hosts are effective at accumulating organic 381 382 matter, facilitating the settlement and persistence of small-sized, detritivorous, and opportunistic species (Christie et al., 2009; Panyawai et al., 2019; Barbosa et al., 2019; 383 384 Duarte et al., 2020a), while also enhancing protection against predation and 385 hydrodynamics as interstitial spaces serve as refuges (Barbosa et al., 2019; Ware et 386 al., 2019).

387 Although Rao's Q did not vary significantly among months, the RLQ analysis 388 indicated a monthly affinity of both macroalgae and annelid traits in relation to June 389 observations. This apparent discrepancy between the mean dissimilarity of annelid 390 traits, as quantified by Rao's Q, and the RLQ output also contrasts with the findings of 391 Craveiro & Rosa-Filho (2024), who documented a monthly shift in epifaunal species 392 composition in response to changes in macroalgal morphological traits. Such changes 393 affected the dominance patterns of polychaetes within the same phytal system. A possible explanation lies in the strong correlation between Rao's Q and taxonomic 394 395 diversity, which reduces the index's sensitivity to changes driven by species relative 396 abundances in low-richness systems, where the dissimilarity matrix is "small" (de Bello 397 et al., 2016).

398 On the Pernambuco coast, winds, mainly driven by the semi-permanent high-399 pressure system over the South Atlantic Ocean, control rainfall and hydrodynamics 400 (Lira et al., 2010; Domingues et al., 2017). Winds tend to be predominant from the 401 east in austral summer and shift to the southeast in austral winter (Lira et al., 2010). This seasonal inversion in wind direction affects rainfall and hydrodynamics, effectively 402 dividing the year into two distinct climatic periods: a rainy season (March to August) 403 404 and a dry season (September to February) (Macêdo et al., 2004; Lira et al., 2010; Vasconcelos et al., 2013; Domingues et al., 2017). In the rainy season, high rainfall, 405 hydrodynamics, and turbidity stress intertidal marine plants (Domingues et al., 2017; 406 407 Bérgamo et al., 2022; Bérgamo et al., 2024). Macroalgae may respond to such 408 stressful conditions by altering their morphological traits, growth rates, and flexibility 409 (Madsen et al., 2001; Hurd, 2000). This pattern was observed by Craveiro & RosaFilho (2024) in the studied system, where macroalgae morphological complexity andbiomass were higher in more hydrodynamically stable months during the dry season.

412 In June, the multivariate distinction between algae morpho-functional groups 413 assemblages was influenced and polychaete traits by the affinity of 414 opportunistic/scavenger and deposit-feeding annelid trait modalities with corticated 415 algae observations positioned along the negative extremes of the second RLQ axis. 416 and herbivore and facultative suspension-feeding modalities with foliose species 417 observations positioned along the negative extremes of the first RLQ axis. As 418 corticated macroalgae can retain suspended material more efficiently (Dean et al., 419 1987), the superposition of deposit-feeding and opportunistic/scavenger annelid 420 genera with observations from corticated algae is expected. In contrast, the 421 suspension-feeding correlation with foliose algae observations is attributed to the 422 incidence of large herbivores and tube-building nereidids, which can secrete mucus within their tubes to capture the suspended material from the water column to be later 423 424 ingested (Daly, 1973; Toba & Sato, 2013).

425 Putting things together, the structural complexity of macroalgae hosts can be 426 interpreted to evaluate the assembly process of their epifauna through the lens of the filtering metaphor, especially in a niche selection context (Dean & Connell, 1987: 427 HilleRisLambers et al., 2012; Locke & Chisholm, 2023). Briefly, assembly theory 428 429 predicts that at fine spatial scales, biotic interactions exert a more decisive influence 430 than abiotic environmental filtering in modulating functional trait diversity within local 431 communities (Mayfield & Levine, 2010; Kraft et al., 2015; Boet et al., 2022; Gross et 432 al., 2022). The morphological traits of macroalgae can be considered as filters for the 433 traits of associated epifauna by mediating this process at the frond scale, as they 434 significantly affected the mean trait dissimilarity of annelid genera, leading to distinct 435 epifaunal trait affinities between the two host morpho-functional groups. For these reasons, the assembly of epifaunal communities on macroalgae is a multifaceted 436 ecological process mediated by host structural traits that can mitigate the effects of 437 negative interspecific interactions and buffer environmental severity, thereby creating 438 439 complex habitats that sustain their high biodiversity from both taxonomic and functional perspectives. 440

- 441
- 442 Conclusion

444 The relationship between habitat structure and functional trait diversity was examined, revealing that increased macroalgal architectural complexity positively 445 446 influences the trait dispersion of associated epifaunal assemblages. However, the 447 strength and nature of this relationship varied depending on the specific traits 448 considered, as different traits capture distinct dimensions of species' ecological niches (Spasojevic et al., 2012; Kraft et al., 2015). All annelid genera inhabiting macroalgae 449 450 were errant polychaetes, characterized by a shared set of morphological traits linked 451 to an epifaunal lifestyle and high mobility, typical of the Errantia clade (Rouse et al., 452 2022). Nonetheless, their traits varied primarily in body size, reproductive modes, and 453 feeding strategies, with the latter contributing most significantly to the observed 454 multivariate patterns of trait distribution across macroalgal morpho-functional groups 455 and months.

456 Finally, a major limitation to the advancement of more robust trait-based 457 approaches is the current paucity of information on the life-history traits of marine 458 invertebrates—a knowledge gap known as the Raunkiaeran shortfall (Hortal et al., 459 2015; Goncalves-Souza et al., 2023; Luza et al., 2023). To overcome this constraint, 460 future research should prioritize the characterization of functional traits in epifaunal 461 species, with particular emphasis on updating and expanding trait data for tropical taxa. It is also important to recognize that macroalgae interact with both their 462 463 environment and associated fauna not only through morphological traits, but also via 464 chemical and reproductive characteristics. Future research should place greater 465 emphasis on elucidating their role in shaping the functional, phylogenetic, and 466 taxonomic diversity of epifaunal assemblages. Such efforts may reveal a highly multidimensional structure of epifaunal biodiversity, underscoring the need for an 467 468 integrative, cross-taxa framework that encompasses multiple facets of biological 469 diversity. This comprehensive approach will be crucial for advancing our 470 understanding of how climate change and anthropogenic pressures impact marine 471 phytal ecosystems.

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Author contributions S.L.D.D.M. originally formulated the idea, analyzed the data and wrote the first manuscript draft under the supervision of R.L.N.. P.C.P. contributed to the statistical analysis routine supervision, data modelling insights, and theoretical refinement. J.S.R.F. and N.C. formulated the sampling design, conducted the fieldwork and provided the taxonomic identifications. All authors reviewed and edited the final text.

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498 **Data availability** The R script with the coding routines will become publicly available 499 after the reviewing process at S.L.D.D.M. github repository 500 (https://github.com/samuelmendes-polychaeta?tab=repositories).

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502 **Confict of interest** We have no conflict of interest to declare.

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