

Relative High Fitness and Genome-wide Diversity May Facilitate Plastic and Active Foragers' Diversification

Dylan J. Padilla Perez^{1,2}, Martha M. Muñoz³, David K. Skelly²

¹Yale Institute for Biospheric Studies, Yale University, New Haven, CT 06511, USA.

²School of the Environment, Yale University, New Haven, CT 06511, USA.

³Department of Ecology and Evolutionary Biology, Yale University, New Haven, CT 06511, USA.

Corresponding author: dylan.padilla@yale.edu

Article type: Research Article

Conflict of interest: We declare we have no conflict of interests.

Keywords: Genetic diversity, genome size, foraging mode, reproductive effort.

This manuscript includes:

Abstract: 234 words

Main Text: ~ 5000 words, excluding references

Figures: 4

References: 62

18 **Abstract**

19 Explaining the variation in diversification rates across the Tree of Life is an important
20 challenge for evolutionary biologists. Growing evidence suggests that key innovations
21 or historical contingency give rise to high diversification rates, but the genetic mechanisms through which this process may occur remain poorly investigated. Based on
22 fitness landscapes, a high diversification is predicted to result from local adaptation as
23 species traverse along genotype space. To test this prediction, we conducted a comparative analysis of 997 reptile species that vary in their locomotion while foraging.
24 The species ranged from those that travel long distances to acquire food to those that
25 barely move and acquire food in nearby sites or those that adopt a plastic strategy. We
26 found that plastic foragers and active foragers not only have high diversification rates
27 but may also have higher fitness compared to sit-and-wait foragers. While traversing
28 among heterogeneous environments, plastic foragers and active foragers could accelerate the pace of evolution by exposing cryptic genetic variation to selection. This is
29 possible in plastic foragers because their larger genomes potentially facilitate variation
30 in gene expression. By contrast, higher genome-wide nucleotide diversity among active
31 foragers could make up for the small size of their genomes, allowing natural selection to
32 operate effectively to the point where divergence by ecological speciation could occur.
33 We used emerging genomic data and macroevolutionary observations supported by
34 microevolutionary processes to provide key insights into mechanisms of diversification.

35 **Keywords:** Genetic diversity, genome size, foraging mode, reproductive effort.

39 **Significance statement**

40 The ways by which organisms seek food vary along a continuum from stationary
41 to highly mobile foraging modes, although many species plastically switch between
42 modes. This range of locomotion may influence the pace of evolution based on the
43 extent to which novel genetic combinations arise as organisms encounter and colonize
44 new environments, leading to variation in diversification rates. Our observations sug-
45 gest that as plastic and active foragers colonize new environments, local adaptation
46 could take place by exposing cryptic genetic variation to selection. As such, high
47 diversification rates may follow through ecological speciation. By contrast, a rela-
48 tively low genetic diversity and fitness among sit-and-wait foragers might lead to low
49 diversification rates through stochastic processes like genetic drift and mutation.

50 **Introduction**

51 The process of evolution requires differences in fitness among individuals within species
52 given variation in their genomes and environments (1). Often, such variation in fitness
53 arises from plastic responses pushing populations into the realm of attraction of new
54 fitness peaks, which can lead to genetic differentiation (2). Accordingly, the relation-
55 ship between genes and fitness is of fundamental importance to better understand the
56 evolutionary history of organisms. Ideally, a theory that links microevolutionary pro-
57 cesses responsible for changes in fitness (e.g., genetic drift, mutation, selection) with
58 macroevolutionary patterns (e.g., speciation, diversification) should provide the basis
59 for describing how biodiversity evolves on Earth. To develop such a theory, researchers
60 rely on the notion of “fitness landscapes”, which enables one to analyze how the process
61 of diversification can occur (3).

62 A prevailing prediction emerging from consideration of fitness landscapes is that
63 many speciation events, and indeed whole adaptive radiations, result from local adap-
64 tation as species colonize new environments (4; 3). This prediction emerges from
65 several sources, including the introduction of novel genetic combinations as organ-
66 isms explore large areas of the genotype space, and the increased likelihood of spatial
67 sorting, where organisms with distinct traits accumulate at the leading edge of a pop-
68 ulation’s expansion (5; 6). To test this prediction, one could focus on species that vary
69 in their locomotor ability while foraging. The foraging behaviors of organisms lie along
70 a continuum of locomotion, ranging from species that travel long distances to acquire

71 food (“active foragers”) to species that barely move and acquire food in nearby sites
72 (“sit-and-wait foragers”; 7; 8). Importantly, the ability of organisms to change their
73 foraging behavior in response to environmental variations (“plastic foragers”) can be
74 specially observed in colonizing species inhabiting heterogeneous environments, where
75 encountering a new environment may result in selection pressures favoring divergence
76 from the ancestor (2; 6).

77 If populations can attain high fitness in heterogeneous environments as a conse-
78 quence of plastic foraging behavior, then genetic differentiation is expected as gene
79 combinations favored under the locally prevailing conditions are not useful in distant
80 environments (9). As populations colonize new environments, however, plasticity can
81 be hindered by a lack of genetic variation, extensive gene flow, and genetic correlation
82 between genes for one trait and genes for plasticity of another trait (10). Further-
83 more, a plastic response may come at the cost of carrying around additional genetic
84 machinery (11). Such a cost can be assessed if one analyzes the relationship between
85 the foraging behaviors of species and the sizes of their genomes. In this context, the
86 cost may be evident if plastic foragers have larger genomes but lower fitness than ac-
87 tive foragers and sit-and-wait foragers. One way to account for differences in fitness
88 among species is to compare their annual reproductive output, which is a good pre-
89 dictor of lifetime fitness (12). Intriguingly, a comparison between foraging behavior,
90 genome size, and reproductive output has not been previously evaluated among verte-
91 brate species, motivating us to conduct a thorough investigation that may enable us
92 to make general conclusions about the ecology and diversification of species across the
93 Tree of Life.

94 Here, we examined the relationship between foraging behaviors and diversification
95 rates among 997 reptile species representing 56 families. In doing so, we considered
96 the predictions of fitness landscapes to assess the idea that effectively traversing the
97 genotype space might result in high diversification rate through local adaptation (13).
98 Because both plastic foragers and active foragers may more effectively explore hetero-
99 geneous environments than sit-and-wait foragers, local adaptation may take place if
100 populations exploit new fitness peaks, leading to a relatively high diversification rate.
101 By contrast, restricted locomotion by sit-and-wait foragers could lead to relatively slow
102 diversification via stochastic processes such as population bottlenecks. These expect-
103 tations are supported by the idea that local adaptation driven by natural selection to
104 suit specific environments generally leads to faster diversification than stochastic pro-
105 cesses (14). Our study uses emerging genomic data and presents macroevolutionary
106 observations supported by microevolutionary processes to provide key insights into the
107 mechanisms of diversification.

108 Results

109 A state-dependent diversification framework indicated that a model in which both the
110 rates of speciation and extinction depend on foraging behavior was strongly supported
111 ($AIC_c = 10076.030, w = 1.000$). Overall, net diversification was indistinguishable
112 among active and plastic foragers, but this rate was higher than that of sit-and-wait
113 foragers (Figure 1C). Character state reconstruction revealed that active foraging ap-
114 pears to be the ancestral state of all reptiles, with a posterior probability of 0.639 at the
115 root of the tree (Figure 1A). Although two major transitions from active foraging to

116 sit-and-wait foraging occurred in Gekkota and Iguania, bursts of frequent transitions
117 immediately followed within each of these clades (Figure 1B). Specifically, the highest
118 number of transitions took place from sit-and-wait to active foraging (~ 64), succeeded
119 by a similar number of transitions from sit-and-wait to plastic foraging (~ 57). The
120 accumulation of lineages from the root of the tree to the present showed that active
121 foragers dominated the landmasses for the first 200 million years since the origin of
122 reptiles in the Tree of Life. However, sit-and-wait foragers subsequently took over
123 (Figure 2A).

124 **Figure 1.**

125 A phylogenetic-informed model revealed that the evolution of annual reproductive
126 output among reptiles is underlain by an interaction between body mass and for-
127 aging behavior (Figure 2B). In general, annual reproductive output increased with
128 body mass, but the highest rate of increase is observed in plastic foragers ($\beta =$
129 $0.134, Std.Error = 0.051, t = 2.603, p = 0.009$).

130 **Figure 2.**

131 As with annual reproductive output, genome size also evolved in response to the
132 interaction between body mass and foraging behavior across species (Figure 3). While
133 genome size decreased with body mass in sit-and-wait foragers ($\beta = -0.042, Std.Error =$
134 $0.017, t = -2.363, p = 0.020$) and plastic foragers ($\beta = -0.073, Std.Error = 0.021, t =$
135 $-3.403, p < 0.001$), the opposite pattern was evident in active foragers ($\beta = 9.188, Std.Error =$

136 0.037, $t = 246.673$, $p < 0.001$). However, plastic foragers had the largest genomes
137 on average ($\mu = 9.271$, $\sigma = 0.133$, $n = 12$) followed by sit-and-wait foragers ($\mu =$
138 9.254, $\sigma = 0.089$, $n = 29$).

139 **Figure 3.**

140 Importantly, active foragers have evolved the smallest genomes on average ($\mu =$
141 9.204, $\sigma = 0.073$, $n = 58$), but their genome-wide nucleotide diversity potentially ex-
142 ceeds that of plastic and sit-and-wait foragers (Figure 4).

143 **Figure 4.**

144 **Discussion**

145 Based on an analysis of nearly one thousand reptile species, we found that plastic
146 foraging and foraging actively are associated with higher diversification rates (Figure
147 1C). Previous hypotheses suggest that historical contingency has been a major de-
148 terminant of the diversification pattern that we observe in modern-day reptiles (15).
149 The early evolution of specialized feeding-related traits in active foragers, such as jaw
150 prehension to capture larger prey, may have enabled them to dominate for almost 200
151 million years since the origin of reptiles (Figure 2A). The subsequent rise may be linked
152 to the emergence and spread of angiosperms (flowering plants) in the past 100 million
153 years (16; 17). The habitat created by large sizes in angiosperms potentially conferred
154 a competitive advantage to sit-and-wait foragers adopting arboreal lifestyles where

155 limited movements suited the restrictions that arboreality exerts on the locomotion
156 of organisms (18; 19). The observation that many sit-and-wait foragers are arboreal
157 while most active foragers are terrestrial supports this claim (15). While the number
158 of active foragers and sit-and-wait foragers may be stabilizing in the present, plastic-
159 foraging lineages continue to grow monotonically, reflecting the diversifying effect of
160 plasticity (Figure 2A).

161 Explaining variation in diversification rates across the Tree of Life is an important
162 challenge for evolutionary biologists. Most of the work on this topic has focused on
163 associations between key innovations or historical factors and diversification rates (20).
164 But the role of variation in fitness and its effects on diversification rates have received
165 less attention. According to theoretical models, the reproductive effort of organisms
166 should influence their lifetime fitness (21; 22; 12). Recent empirical evidence supports
167 this idea; for example, (23) showed that the fitness of an iteroparous species was
168 the highest at its peak annual reproductive output. In this study, not only was the
169 net diversification of plastic foragers relatively high, but they also evolved relatively
170 high reproductive effort (Figure 1C and Figure 2). Foraging plasticity, the ability of an
171 organism to change its foraging behavior in response to environmental variations, could
172 then accelerate the pace of evolution, in turn accelerating diversification rates. This
173 flexibility may be crucial for colonizing species, such as plastic and active foragers,
174 to survive and reproduce above maintenance levels and, hence, for the persistence
175 of species (24). Often, such plasticity is adaptive in that organisms that show a
176 plastic response tend to have higher fitness than those that do not (2). There are
177 many examples where animals respond to heterogenous environments with immediate

178 behavioral changes. Many bird species show a realm of exploratory foraging behaviors,
179 occasionally resulting in quite innovative foraging techniques (25; 26). These feeding
180 innovations are correlated with species numbers (27), reflecting the contribution of
181 plasticity to lineage diversification. But the question of how fitness variation through
182 phenotypic plasticity can lead to species diversification remains puzzling.

183 To elucidate a potential answer to this question, let us consider the ability of plas-
184 tic foragers to colonize and survive in new environments (e.g., 28). In the process of
185 colonizing new environments, local adaptation could take place by exposing cryptic ge-
186 netic variation to selection (2). This is possible in plastic foragers because their large
187 genomes potentially contain more genes, more and longer introns, and more trans-
188 posable elements (Figure 3). Transposable elements often facilitate gene duplication
189 and variation in gene expression (29; 30). As variation in gene expression is expected
190 to underlie plasticity in higher order traits, including fitness, genetic changes in se-
191 quences regulating gene expression are likely to have a key role in lineage divergence
192 (31; 32; 33). If selection acts on the new genetic variation supplied by gene expression,
193 then different lineages might become adapted to and simultaneously develop genetic
194 preferences for different ecological niches (e.g., 34). Eventually, species utilizing differ-
195 ent ecological niches evolve differences in mating preferences by a process analogous
196 to reinforcement (for compelling reviews, see 35; 36). By contrast, the relatively small
197 genomes of active foragers may limit the variation in gene expression or function given
198 the restricted capacity to acquire new genes by gene duplication (37). Yet, a high
199 genome-wide nucleotide diversity among active foragers could compensate for the rel-
200 atively small size of their genomes (Figure 4A). Because nucleotide diversity (π) is

201 directly related to the effective population size (N_e), active-foraging species with high
202 nucleotide diversity are expected to have large effective population sizes (e.g., 37). As
203 the efficiency of selection increases with effective population size, speciation by nat-
204 ural selection could occur through ecological speciation (38; 39). Under this process,
205 natural selection acts in contrasting directions between environments, which drives
206 the fixation of different alleles, each advantageous in one environment but not in the
207 other, potentially causing populations to diverge into new species (40). However, if the
208 species happen to have relatively low nucleotide diversity, as in the case of sit-and-wait
209 foragers (Figure 4B), abundant resources and a lack of competitors might enable them
210 to seed populations that exploit different niches at low densities, potentially leading to
211 the diversification of species by stochastic processes like random mutation and genetic
212 drift (35; 41).

213 Overall, we provide a set of potential scenarios by which the process of diversifi-
214 cation could occur in reptiles. Our framework places emphasis on the ways in which
215 variation in foraging behavior alters the vagility of organisms, allowing them to effec-
216 tively traverse along the genotype space and explore different fitness peaks. Although
217 the mechanisms underlying our hypotheses sound appealing, alternative views should
218 also be considered. For instance, some evidence suggests that a high dispersal ability
219 is expected to favor higher rates of gene flow (42). In contrast to the predictions of
220 our hypotheses, over an evolutionary timescale gene flow is expected to suppress spe-
221 ciation events and thus clade level diversification (43; 44). However, recent work has
222 shown that speciation with gene flow is also possible (44; 45). In the past decades,
223 the emerging field of speciation genomics has enabled us to transition from individual

gene to whole genome, improving our understanding of speciation with gene flow. Our observations that plastic foragers have larger genomes and that active foragers have a relatively high genome-wide genetic diversity pave the way for others to investigate the issue of the relative importance of divergence hitchhiking and genome hitchhiking for facilitating speciation with gene flow among reptiles. Although annual reproductive output can be considered a good proxy for fitness, species survival must also be examined. In essence, fitness is rather intricately associated to the reproduction and survival of organisms. Future directions should address the question of whether foraging behavior and dispersal capacity are actually linked to each other. To our knowledge, studies comparing dispersal among reptile species varying in foraging behavior are scarce in the literature. Therefore, more compelling evidence is required to conduct meaningful comparative analyses of dispersal and its connection to diversification rates among reptiles.

Materials and Methods

Ecological data source

We used a comprehensive database for integrating a diverse range of physiological, behavioral, and life history data to explore patterns of diversification among reptiles (e.g., 46). Specifically, our analyses focused on predicting diversification rates based on the foraging behaviors of species. We classified the foraging behaviors based on whether the species have been reported as active forgers, sit-and-wait foragers, or using a plastic strategy (e.g., 47); a categorization that, albeit crude, remains useful to

245 biologists for defining the extremes of a continuum. Our investigation included data of
246 foraging behavior for 997 species of squamate reptiles distributed among 56 families.

247 The foraging behavior of organisms has become a paradigm in evolutionary biology
248 because of its connection to many traits including the locomotion of species, which
249 potentially affects their dispersal capacity (8). For example, while active foragers
250 disperse over long distances to acquire food, sit-and-wait species barely disperse and
251 forage in nearby sites. If these premises are met, then investigating the relationship
252 between foraging behaviors and the fitnesses of species enables the assessment of a
253 prevailing view of fitness landscapes. That is, effectively traversing along the genotype
254 space might result in high diversification rates through local adaptation (13; 4; 3).
255 To assess this prediction, we first collected data of annual reproductive output among
256 reptile species. The annual reproductive output can be considered a good proxy for
257 fitness because it is generally a good predictor of the long-term fitness of organisms (12;
258 22). We defined the annual reproductive output as the product between the average
259 clutch sizes of species and their average number of clutches per year. We then regressed
260 this quantity on the maximum body mass of the species (g). By regressing the product
261 between the average clutch size and the average number of clutches per year, the slope
262 of the linear relationship can be interpreted as reproductive effort—proportion of mass
263 allocated to reproduction—which enabled us to avoid statistical issues associated with
264 the analysis of ratios. To account for species relatedness, the regression was informed
265 by a time-calibrated phylogeny of squamate reptiles (48). The regression model was
266 fitted in the free software for statistical computing R (49), using the function *gls* from
267 the library “nlme” (50).

268 *Ancestral state reconstruction*

269 Because historical contingency may have played an important role in the diversification
270 of species that we observe today (51), we inferred the evolutionary history of foraging
271 behaviors among reptiles. To do so, we fitted a set of continuous-time, discrete-state
272 Markov chain models to sample the character histories from their posterior probability
273 distribution (52), across a time-calibrated phylogeny of squamate reptiles (48). The
274 models consisted of a an equal-rates (ER) model, in which the rate of change between
275 the three states of the character were assumed to be equivalent. We also fitted an
276 all-rates-different model (ARD), which enables transitions among states to occur at
277 different rates. Lastly, we fitted a symmetrical model, which enables pairs of states
278 to change at different rates but changes among all states are theoretically possible.
279 To fit the models, we used the default arguments of the function *make.simmap* from
280 the “phytools” library of R (53), and simulated 1×10^5 character maps. We then
281 summarized the number of state changes and the posterior probabilities of each internal
282 node generated from the character map simulations. We selected the most likely model
283 based on an information-theoretic approach such as the Akaike Information Criterion
284 (*AIC*).

285 *State-dependent diversification framework*

286 To explore whether the foraging behavior of species influenced speciation and extinc-
287 tion rates, we relied on state-dependent speciation and extinction models (SSE). These
288 models are a birth-death process where the diversification rates are dependent on the
289 state of an evolving character (54). Because the data of foraging behavior consisted of

290 a discrete character with 3 levels, we used the MuSSE method—a Multi-State Charac-
 291 ter extension of the Binary State Speciation and Extinction Model (BiSSE). In doing
 292 so, we first defined a likelihood function, and then optimized it as required by the
 293 library *diversitree* of R (55). The likelihood function requires a phylogenetic tree (i.e.,
 294 48), a vector of numbers ranging from 1 to 3 (where 1 = active foraging, 2 = plastic
 295 foraging, and 3 = sit-and-wait foraging), the number of states ($k = 3$), and a vec-
 296 tor specifying the proportion of species in each character state. We computed this
 297 proportion based on the ratio of the number of species we had data for within each
 298 foraging state to the total number of squamate species currently reported on Reptile
 299 Database (see <http://www.reptile-database.org>). Subsequently, we constrained
 300 this general likelihood function to fit different competing models. We started with a
 301 null model, in which all birth and death rates are equal between states. Next, we fitted
 302 the most complex model in which all rates of speciation and extinction depended on
 303 the character state for our multi-state character. Also, we fitted models in which only
 304 the speciation rate (λ) varied between states, only the extinction rate (μ) varied, and
 305 one in which neither λ nor μ varied, but the transition rates differed between types of
 306 transitions (e.g., ordered, unordered, etc.). As previously described, we compared the
 307 models' goodness of fit based on AIC_c and selected the most likely one for inferences.
 308 Finally, we used the most likely model to run a Bayesian Markov chain Monte Carlo
 309 simulation (MCMC) with 1×10^5 steps to take, an exponential prior distribution, and
 310 the control parameter (w) suggested by (54).

311 We complemented the state-dependent diversification framework with a lineage-
 312 through-time plot, which consists of a visual representation of how the number of

313 lineages within clades changed over time, essentially tracing the diversification history
314 of the clades (56). Importantly, the interpretation of this analysis remains inconclusive
315 because a simple comparison of the total number of species between clades of differ-
316 ent ages does not necessarily reflect consequences of species interactions even though
317 species numbers differed impressively.

318 *Genomic data source*

319 Because genetic diversity changes as organisms move their alleles around the genotype
320 space, important variation in the genetic makeup among species should be observed
321 (57). Accordingly, we first examined the association between genome size and the
322 foraging behavior of species. To do this, we obtained data of genome size from The
323 National Center for Biotechnology Information (NCBI; [https://www.ncbi.nlm.nih.](https://www.ncbi.nlm.nih.gov/)
324 [gov/](https://www.ncbi.nlm.nih.gov/)) and The Animal Genome Size Database (58). The genome size dataset that we
325 compiled included 99 squamate species distributed among 29 families. To model the
326 association between foraging behavior and genome size among species, we fitted a set
327 of phylogenetic-corrected models and evaluated their goodness of fit based on AIC_c
328 values. In addition to accounting for the effect of relatedness between species, we also
329 accounted for potential confounding factors such as body mass. To do so, we used the
330 function *gls* from the library “nlme” of R (50).

331 Furthermore, we compared the genome-wide genetic diversity of an active forager
332 (*Podarcis muralis*) with that of a sit-and-wait forager (*Anolis carolinensis*). To do
333 this, we obtained whole genomes from one population of each species ($n = 5$). The ge-
334 nomic sequences obtained for *P. muralis* and *A. carolinensis* are available on NCBI

under the Bioproject numbers PRJNA715201 and PRJNA533001, respectively. We performed a quality-control check of the samples (paired-end sequences) with FastQC (59), and filtered out reads of low quality with Trimmomatic (60). After quality control, we aligned the reads to the reference genome of *P. muralis* (PodMur_1.0) and *A. carolinensis* (rAnoCar3.1.pri) using *bwa* from samtools (61). We then ran the *HaplotypeCaller* algorithm from the software GATK4 (62) to identify single nucleotide polymorphisms (SNPs) across the genomes of the study species. This pipeline generated the variant-calling format files (VCF) that we later used to compute the nucleotide diversity (π). The nucleotide diversity is a measure of genetic variation within a population, which is calculated as the average number of nucleotide differences per site in pairwise comparisons of DNA sequences. To accomplish this task, we used *vcftools* to quantify the nucleotide diversity over 10kb (1×10^4bp) windows of the genome. Finally, we presented the average nucleotide diversity at a chromosome level for each species with the associated standard deviation.

Acknowledgements

We thank Laura Alencar, David Klings, and Raúl Araya-Donoso for their feedback on previous versions of this manuscript.

Data Accessibility Statement

A fully reproducible workflow of the data analyses, including R scripts and additional supporting material, is available in the following repositories: Github <https://dylan-padilla.github.io/speciation-foraging/>. A dryad link will be avail-

356 able upon acceptance: .

357 Conflict of interest

358 The authors have declared no competing interests.

359 References

- 360 [1] Orr HA (2009) Fitness and its role in evolutionary genetics. *Nature Reviews*
361 *Genetics* 10(8):531–539.
- 362 [2] Price TD, Qvarnström A, Irwin DE (2003) The role of phenotypic plasticity in
363 driving genetic evolution. *Proceedings of the Royal Society of London. Series B:*
364 *Biological Sciences* 270(1523):1433–1440.
- 365 [3] Wright S (1931) Evolution in mendelian populations. *Genetics* 16(2):97.
- 366 [4] Gavrillets S (1997) Evolution and speciation on holey adaptive landscapes. *Trends*
367 *in ecology & evolution* 12(8):307–312.
- 368 [5] Ochocki BM, Miller TE (2017) Rapid evolution of dispersal ability makes biolog-
369 ical invasions faster and more variable. *Nature communications* 8(1):14315.
- 370 [6] Shine R, Baeckens S (2023) Rapidly evolved traits enable new conservation tools:
371 perspectives from the cane toad invasion of australia. *Evolution* 77(8):1744–1755.
- 372 [7] Pianka ER (1966) Convexity, desert lizards, and spatial heterogeneity. *Ecology*
373 47(6):1055–1059.
- 374 [8] Reilly SM, McBrayer LD, Miles DB (2007) *Lizard ecology*. (Cambridge University
375 Press).
- 376 [9] Waddington CH (1961) Genetic assimilation. *Advances in genetics* 10:257–293.
- 377 [10] Murren CJ, et al. (2015) Constraints on the evolution of phenotypic plasticity:
378 limits and costs of phenotype and plasticity. *Heredity* 115(4):293–301.
- 379 [11] Knight CA, Molinari NA, Petrov DA (2005) The large genome constraint hypoth-
380 esis: evolution, ecology and phenotype. *Annals of Botany* 95(1):177–190.
- 381 [12] Alif Ž, Dunning J, Chik HYJ, Burke T, Schroeder J (2022) What is the best
382 fitness measure in wild populations? a case study on the power of short-term
383 fitness proxies to predict reproductive value. *PLoS One* 17(4):e0260905.

- [13] Patton AH, Richards EJ, Gould KJ, Buie LK, Martin CH (2022) Hybridization alters the shape of the genotypic fitness landscape, increasing access to novel fitness peaks during adaptive radiation. *Elife* 11:e72905.
- [14] García-Pintos LP (2024) Limits on the evolutionary rates of biological traits. *Scientific Reports* 14(1):11314.
- [15] Vitt LJ, Pianka ER, Cooper, Jr WE, Schwenk K (2003) History and the global ecology of squamate reptiles. *The American Naturalist* 162(1):44–60.
- [16] van der Kooi CJ, Ollerton J (2020) The origins of flowering plants and pollinators. *Science* 368(6497):1306–1308.
- [17] Jud NA, et al. (2018) A new fossil assemblage shows that large angiosperm trees grew in north america by the turonian (late cretaceous). *Science advances* 4(9):eaar8568.
- [18] Astley HC, Jayne BC (2007) Effects of perch diameter and incline on the kinematics, performance and modes of arboreal locomotion of corn snakes (*Elaphe guttata*). *Journal of Experimental Biology* 210(21):3862–3872.
- [19] Hyams SE, Jayne BC, Cameron GN (2012) Arboreal habitat structure affects locomotor speed and perch choice of white-footed mice (*Peromyscus leucopus*). *Journal of Experimental Zoology Part A: Ecological Genetics and Physiology* 317(9):540–551.
- [20] Ricklefs RE (2004) A comprehensive framework for global patterns in biodiversity. *Ecology letters* 7(1):1–15.
- [21] Roff DA, , et al. (2002) Life history evolution.
- [22] Stearns SC (2000) Life history evolution: successes, limitations, and prospects. *Naturwissenschaften* 87:476–486.
- [23] Hämäläinen A, et al. (2017) Fitness consequences of peak reproductive effort in a resource pulse system. *Scientific Reports* 7(1):9335.
- [24] Fusco G, Minelli A (2010) Phenotypic plasticity in development and evolution: facts and concepts. *Philosophical Transactions of the Royal Society B: Biological Sciences* 365(1540):547–556.
- [25] Lefebvre L (2000) Feeding innovations and their cultural transmission in bird populations in *The Evolution of Cognition*. (The MIT Press).
- [26] Lefebvre L, Juretic N, Nicolakakis N, Timmermans S (2001) Is the link between forebrain size and feeding innovations caused by confounding variables? a study of australian and north american birds. *Animal Cognition* 4:91–97.
- [27] Nicolakakis N, Sol D, Lefebvre L (2003) Behavioural flexibility predicts species richness in birds, but not extinction risk. *Animal Behaviour* 65(3):445–452.

- [28] Sol D, Timmermans S, Lefebvre L (2002) Behavioural flexibility and invasion success in birds. *Animal behaviour* 63(3):495–502.
- [29] Krasileva KV (2019) The role of transposable elements and dna damage repair mechanisms in gene duplications and gene fusions in plant genomes. *Current Opinion in Plant Biology* 48:18–25.
- [30] Marino A, Debaecker G, Fiston-Lavier AS, Haudry A, Nabholz B (2024) Effective population size does not explain long-term variation in genome size and transposable element content in animals. *bioRxiv* pp. 2024–02.
- [31] Siddiq MA, Duveau F, Wittkopp PJ (2024) Plasticity and environment-specific relationships between gene expression and fitness in *saccharomyces cerevisiae*. *Nature Ecology & Evolution* pp. 1–11.
- [32] Meyer A (1987) Phenotypic plasticity and heterochrony in *cichlasoma managuense* (pisces, cichlidae) and their implications for speciation in cichlid fishes. *Evolution* 41(6):1357–1369.
- [33] Kappeler PM, Fichtel C (2015) Eco-evo-devo of the lemur syndrome: did adaptive behavioral plasticity get canalized in a large primate radiation? *Frontiers in zoology* 12:1–16.
- [34] Alatalo RV, Gustafsson L (1988) Genetic component of morphological differentiation in coal tits under competitive release. *Evolution* pp. 200–203.
- [35] Gavrillets S (2010) High-dimensional fitness landscapes and speciation. *Evolution: the extended synthesis* pp. 45–79.
- [36] Gavrillets S, Losos JB (2009) Adaptive radiation: contrasting theory with data. *Science* 323(5915):732–737.
- [37] Charlesworth B, Barton N (2004) Genome size: does bigger mean worse? *Current Biology* 14(6):R233–R235.
- [38] Schluter D (2009) Evidence for ecological speciation and its alternative. *Science* 323(5915):737–741.
- [39] Nosil P (2012) *Ecological speciation*. (Oxford Ecology and Evolution).
- [40] Schluter D, Conte GL (2009) Genetics and ecological speciation. *Proceedings of the National Academy of Sciences* 106(supplement_1):9955–9962.
- [41] Gavrillets S (2014) Models of speciation: where are we now? *Journal of heredity* 105(S1):743–755.
- [42] Suárez D, Arribas P, Jiménez-García E, Emerson BC (2022) Dispersal ability and its consequences for population genetic differentiation and diversification. *Proceedings of the Royal Society B* 289(1975):20220489.

- [43] Claramunt S, Derryberry EP, Renssen Jr J, Brumfield RT (2012) High dispersal ability inhibits speciation in a continental radiation of passerine birds. *Proceedings of the Royal Society B: Biological Sciences* 279(1733):1567–1574.
- [44] Weeks BC, Claramunt S (2014) Dispersal has inhibited avian diversification in australasian archipelagoes. *Proceedings of the Royal Society B: Biological Sciences* 281(1791):20141257.
- [45] Feder JL, Egan SP, Nosil P (2012) The genomics of speciation-with-gene-flow. *Trends in genetics* 28(7):342–350.
- [46] Oskyrko O, Mi C, Meiri S, Du W (2024) Repttraits: a comprehensive dataset of ecological traits in reptiles. *Scientific Data* 11(1):243.
- [47] Meiri S (2018) Traits of lizards of the world: Variation around a successful evolutionary design. *Global ecology and biogeography* 27(10):1168–1172.
- [48] Zheng Y, Wiens JJ (2016) Combining phylogenomic and supermatrix approaches, and a time-calibrated phylogeny for squamate reptiles (lizards and snakes) based on 52 genes and 4162 species. *Molecular phylogenetics and evolution* 94:537–547.
- [49] R Core Team (2023) *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, Vienna, Austria).
- [50] Pinheiro J, et al. (2007) Linear and nonlinear mixed effects models. *R package version 3*(57):1–89.
- [51] Blount ZD, Lenski RE, Losos JB (2018) Contingency and determinism in evolution: Replaying life’s tape. *Science* 362(6415):eaam5979.
- [52] Huelsenbeck JP, Nielsen R, Bollback JP (2003) Stochastic mapping of morphological characters. *Systematic biology* 52(2):131–158.
- [53] Revell LJ (2012) phytools: an r package for phylogenetic comparative biology (and other things). *Methods in ecology and evolution* (2):217–223.
- [54] FitzJohn RG, Maddison WP, Otto SP (2009) Estimating trait-dependent speciation and extinction rates from incompletely resolved phylogenies. *Systematic biology* 58(6):595–611.
- [55] FitzJohn RG (2012) Diversitree: comparative phylogenetic analyses of diversification in r. *Methods in Ecology and Evolution* 3(6):1084–1092.
- [56] Helmstetter AJ, et al. (2022) Pulled diversification rates, lineages-through-time plots, and modern macroevolutionary modeling. *Systematic Biology* 71(3):758–773.
- [57] Waters J, Emerson BC, Arribas P, McCulloch G (2020) Dispersal reduction: causes, genomic mechanisms, and evolutionary consequences. *Trends in Ecology & Evolution* 35(6):512–522.

- 491 [58] Gregory TR, et al. (2007) Eukaryotic genome size databases. *Nucleic acids re-*
492 *search* 35(suppl_1):D332–D338.
- 493 [59] Andrews S, et al. (2012) FastQC (Babraham Institute).
- 494 [60] Bolger AM, Lohse M, Usadel B (2014) Trimmomatic: a flexible trimmer for illu-
495 mina sequence data. *Bioinformatics* 30(15):2114–2120.
- 496 [61] Danecek P, et al. (2021) Twelve years of SAMtools and BCFtools. *GigaScience*
497 10(2).
- 498 [62] Van der Auwera GA, O'Connor BD (2020) *Genomics in the cloud: using Docker,*
499 *GATK, and WDL in Terra*. (O'Reilly Media).

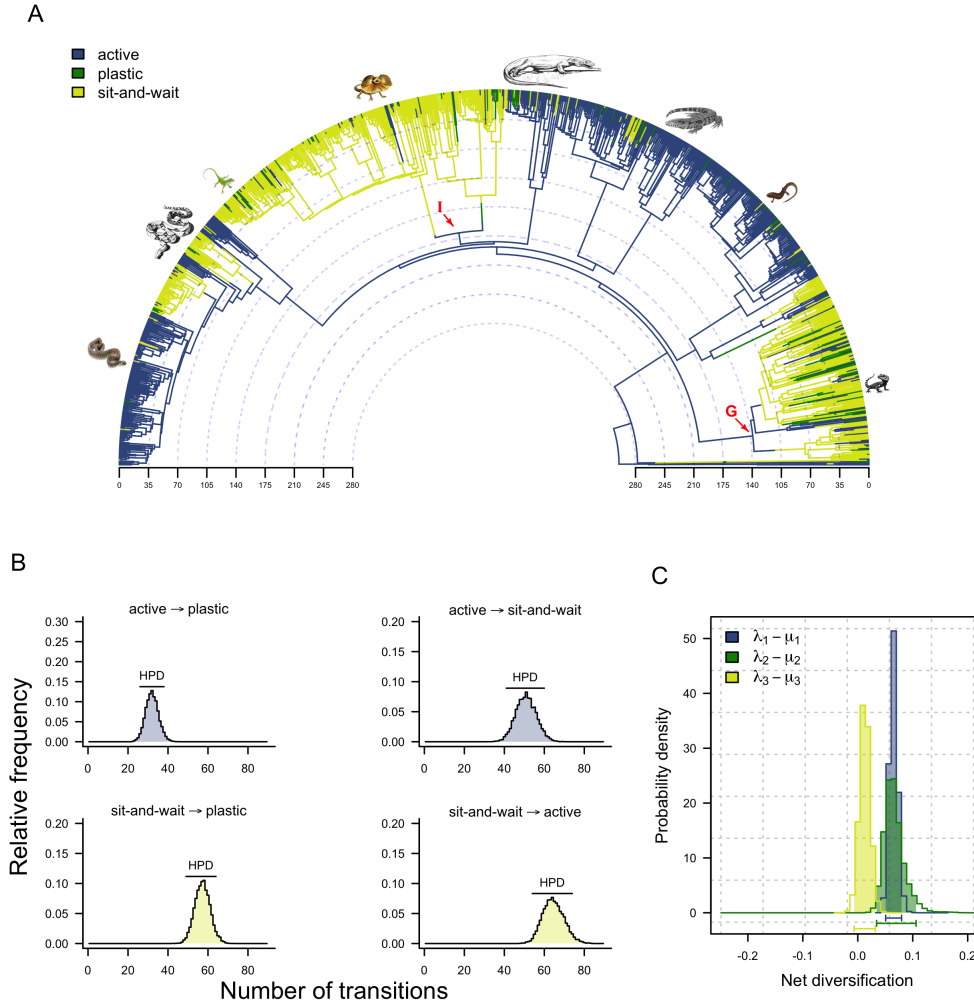


Figure 1: A) Random sample of 1×10^5 simulated character maps depicting the evolution of foraging behaviors among 997 reptile species. Clades where major transitions have occurred are indicated as follows: G = Gekkota, I = Iguania. B) Expected number of changes between the states of the character under the most likely model. The high probability density (HPD) reflects the variance of changes between states given the assumed model. C) Net diversification rates between clades defined by the foraging behavior of species. For each clade, the net diversification was computed as the difference between speciation rates (λ) and extinction rates (μ).

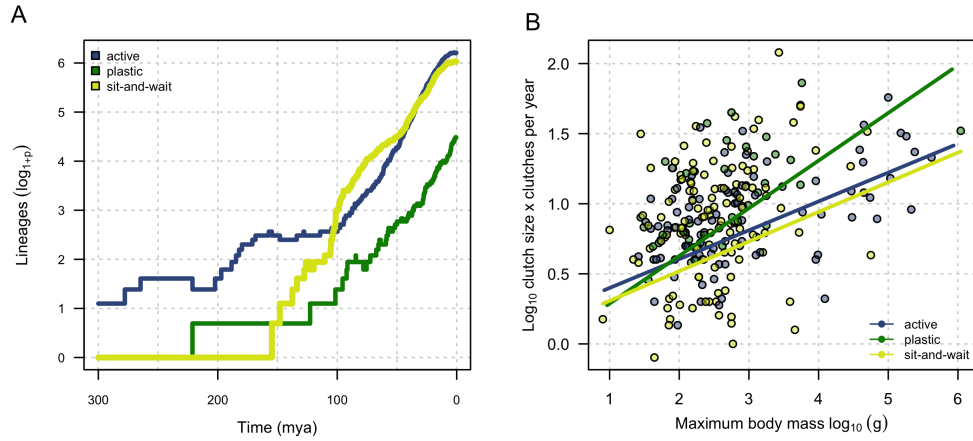


Figure 2: A) Lineage-through-time plot illustrating the accumulation of lineages within clades defined by the foraging behavior of species. B) Relationship between the annual reproductive output of species as a function of their body mass and foraging behavior.

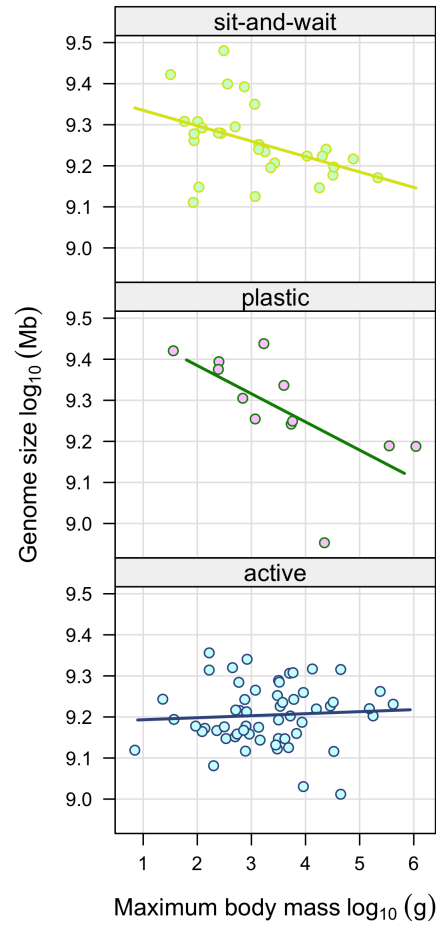


Figure 3: Evolution of genome size in response to the interaction between body mass and foraging behavior among reptile species.

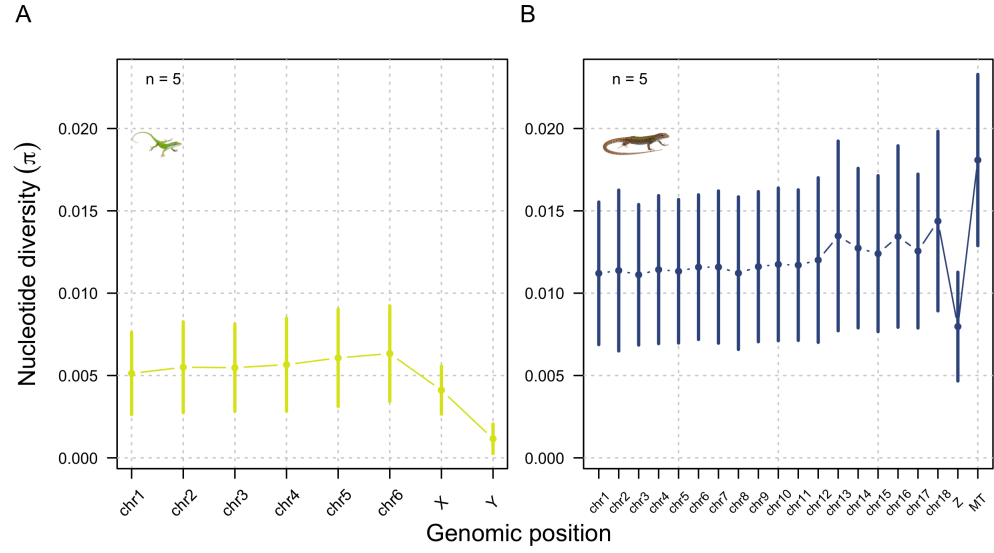


Figure 4: A) Genome-wide nucleotide diversity estimated from a population of a sit-and-wait forager (*Anolis carolinensis*). B) Genome-wide nucleotide diversity estimated from a population of an active forager (*Podarcis muralis*). Solid dots represent the average nucleotide diversity per chromosome and across the mitochondrial genome, which is abbreviated as MT. The bars associated indicate the standard deviation.