- 1 The ecological relevance of fast-cycling mineral-associated organic matter a dynamic pool
- 2 of 'persistent' soil carbon and nitrogen
- 3

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- 40 Abstract
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42 Longstanding theories and models classify mineral-associated organic matter (MAOM) as the 43 large (~60%) but slow-cycling and persistent portion of the soil organic matter (SOM) pool. 44 Strong physico-chemical interactions and diffusion limitations restrict the turnover of MAOM, 45 allowing carbon and nitrogen bound therein to persist in soil for as long as centuries to millennia. 46 However, MAOM is a chemically and functionally diverse pool with a substantial portion 47 cycling at relatively fast (i.e., minutes to years) timescales. Despite a growing body of evidence 48 for the heterogenous and multi-pool nature of MAOM, we lack consensus on how to 49 conceptualize and directly quantify fast-cycling MAOM and its ecological significance. We 50 demonstrate the dynamic qualities of fast-cycling MAOM vary based on 1) the chemistry of the 51 mineral particles and organic matter, 2) the complex set of interactions between OM and the 52 mineral matrix, and 3) the presence and strength of destabilizing forces that lead to 53 decomposition or loss of MAOM (i.e., plant-microbe interactions, land use change, agricultural 54 intensification, and climate change). Finally, we discuss potential implications and research 55 opportunities for how we measure, manage, and model the dynamic subfraction of this otherwise 56 persistent pool of SOM. 57 58 59 1. Introduction 60 61 62 Soil organic matter (SOM) plays a central role in terrestrial ecosystem functioning, providing 63 nitrogen (N) and other nutrients to plants, and holding the largest pool of organic carbon (C) on

land. A portion of the SOM pool cycles slowly – with turnover time ranging from decades to 64 65 millennia. Our understanding of SOM persistence has evolved from an emphasis on chemical 66 complexity and "recalcitrance" towards a growing appreciation for the interactions between relatively simple plant and microbial inputs and reactive mineral surfaces as key controls on 67 SOM persistence¹. As such, there is considerable focus on characterizing and measuring the 68 69 mineral-associated organic matter (MAOM; see Box 1) pool, the fraction of SOM which holds 70 the majority of organic C and N in the terrestrial biosphere ($\sim 60\%$ of C and 75% of N)². Strong 71 physico-chemical interactions and diffusion limitations restrict the turnover of some MAOM, 72 allowing C within this pool to persist for centuries to millennia³.

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Box 1: MAOM definitions and characteristics

Mineral-associated organic matter (MAOM) is the term used to describe both a conceptuallydefined and methodologically-defined fraction of soil organic matter (SOM). Conceptually, MAOM is defined as a fraction of SOM where OM exists in some degree of association with soil minerals, such as via sorption, co-precipitation, encapsulation in fine, micro-aggregates sensu Lehmann et al.⁴, or via organo-organo interactions. MAOM is typically defined in contrast to sand-sized and less protected material known as particulate organic matter (POM). The methodological definition of MAOM is material collected from soil (generally, operationally defined as < 2mm) with aggregates dispersed by physical (i.e., glass beads), chemical (i.e., dispersing agents like sodium hexametaphosphate), or energetic (i.e., sonication) methods and then collected as the fraction passing through a 50-63 μ m sieve (i.e., the silt- and clay-sized particles) and/or with a density greater than 1.6-1.85 g cm⁻³ (i.e., the heavy fraction). Operational definitions of MAOM (e.g., as defined by a small particle size or heavier density) are not always fully aligned with conceptual definitions. Size separations can also capture partially decomposed plant and fungal components not associated with minerals in the fine fraction, and density separations may capture SOM associated with sand in the heavy fraction. Recent work has found that combining size and density separation methods can minimize these artifacts⁵.

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75 Particulate organic matter (POM) is considered a more bioavailable and faster cycling 76 pool of soil C and N, relative to MAOM. POM is dominated by larger fragments of plant and 77 microbial residues, which are at early stages of decomposition. The POM pool responds rapidly 78 to changes in tillage or plant inputs and often correlates strongly with whole soil measures of 79 decomposition⁶, contributing to the view of POM as an SOM pool that is particularly sensitive to 80 disturbances and physically accessible to decomposers. However, the decomposition of POM, in 81 addition to litter decomposition, is also needed to build MAOM⁷. The leaching and 82 depolymerization of POM and litter releases OM compounds into solution, which can then 83 adsorb to a reactive mineral surface before or after cycling through microbial biomass^{8–10}. While microaggregates provide further protection to MAOM¹¹, aggregate turnover and disturbance 84 85 promote MAOM formation as well¹². MAOM formation is also more efficient within the 86 rhizosphere relative to the bulk soil, which suggests MAOM forms under highly dynamic conditions¹³. MAOM formation is fast: transfer of ¹⁵N or ¹³C-labeled plant residues into MAOM 87 fractions can occur within days to months^{14–17}. 88 89 The abundance of fine mineral particles and mineral specific surface area are strong

predictors of soil C storage^{18,19}. As such, MAOM is often examined for its capacity to store and
sequester C and nutrients, with less attention on the heterogeneity underlying the MAOM

92 fraction and its implications for potential destabilization following disturbance (for exceptions, see Bailey et al. and Dynarski et al.)^{20,21}. Although the majority of MAOM is likely persistent 93 94 and long-lived, there is increasing evidence for a dynamic, fast-cycling MAOM pool. Given the vast quantity of C and N stored in MAOM, the presence of even a small active 95 96 MAOM pool can substantially influence ecosystem processes. For example, a hypothetical and 97 very conservative annual MAOM turnover rate of 1% in a grassland with 3,000 kg MAOM N ha-¹ in the top 10 cm of soil²² would supply 30 kg N ha⁻¹ per year, which is approximately a third of 98 the N uptake by grassland species²³. The fast-cycling MAOM pool may function as a distinct 99 100 SOM pool with essential roles in ecosystems, including retaining nutrients prone to loss and 101 supplying nutrients to plants and microbes. At the same time, this fast-cycling MAOM pool may 102 become a larger source of greenhouse gas emissions over time, especially under accelerated land 103 use and climate change, as we discuss below.

The idea of a fast-cycling MAOM pool is not new (e.g., Kleber et al. and Torn et al.)^{24,25}, 104 105 but the concept is rarely acknowledged in research and policies related to the management of 106 MAOM (e.g., natural climate solutions-oriented work) or in Earth system models. In this review, 107 we seek to clarify the existence and ecological importance of fast-cycling MAOM with a 108 targeted summary of research on this crucial SOM pool. Others have synthesized relevant geochemical mechanisms of mineral-organic interactions²⁶ and the land use and climate change 109 implications of POM versus MAOM²⁷ but have not focused explicitly on the fast-cycling 110 111 MAOM pool. Here, we synthesize recent studies that confirm the existence of a fast-cycling 112 MAOM pool across various ecosystem types and provide insight into its size, stability and 113 mechanisms of turnover. We describe its key role in ecosystem functions such as nutrient

provisioning, soil C cycling, and plant-microbe interactions, and suggest implications for how
we measure, manage, and model this dynamic subfraction of MAOM.

- 116
- 117 2. MAOM is chemically heterogeneous
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119 MAOM is a large and heterogenous pool of SOM, containing mineral particles and 120 organic molecules that vary widely in age, size, and reactivity. Variations in MAOM binding 121 mechanisms and chemical characteristics determine how this pool of OM cycles through 122 terrestrial ecosystems, by influencing its availability to microbes, turnover rate and potential to 123 serve as a plant nutrient source. At the scale of mineral surface-OM associations, the behavior of 124 MAOM and the chemical bonds therein will depend largely on characteristics of the interacting 125 organic compounds, mineral particles and soil solution^{26,28–31}(Figure 1). Minerals can have 126 permanent and/or variable charges of varying strength, distribution, and density and such 127 differences will impact the type and number of binding sites, adsorption-desorption behavior, 128 and hence, the bioavailability of associated molecules^{32,33}. For example, minerals with a 129 relatively high point-of-zero-charge that are positively charged under neutral and acidic 130 conditions, such as goethite, may be able to bind SOM more strongly via ligand exchange 131 reactions compared to negatively charged clay minerals that bind to OM via cation bridging³⁴. 132 Mineral types that bind compounds more weakly may harbor more vulnerable and accessible 133 forms of MAOM.

134 Characteristics of organic substrates expected to influence the mechanism and strength of
135 association include molecular size, hydrophobicity, acidity, and abundance of carboxylic,
136 phenolic and aromatic constituents, which interact with mineral surface chemistry to influence

137	the mechanism and reversibility of organo-mineral associations ^{35–37} . For example, stronger
138	covalent bonds tend to form between organic substrates enriched in aromatic acid and phenolic
139	groups and mineral fractions with a high abundance of reactive Fe and Al (oxy)hydroxide
140	phases, especially under acidic soil solution conditions ^{36,38} . Stoichiometry (e.g., the C:N ratio) of
141	organic matter will also influence its potential for adsorption or desorption ³⁹ . Empirical work and
142	direct imaging of organo-mineral interfaces at sub-micron scales indicate that N-rich organic
143	compounds (e.g., N-containing groups in proteins) bind preferentially to mineral surfaces ⁴⁰⁻⁴⁵ ,
144	and it is often suggested that MAOM-N is more stable than MAOM-C as a result.
145	The stability and turnover dynamics of MAOM are also shaped by its fine-scale spatial
146	and compositional heterogeneity ^{4,43,46–49} . Several lines of evidence support the existence of
147	spatially or chemically distinct portions of MAOM that are more vulnerable to mineralization.
148	Kleber et al. proposed a model suggesting that OM binds to mineral surfaces in a zonal
149	structure ²⁴ . In this multi-layer model, the outermost zone is referred to as the "kinetic zone" and
150	hosts organic molecules that are weakly bound via van der Waals forces, hydrogen bonding, and
151	cation bridging interactions. Secondary interactions among organic molecules (i.e., organo-
152	organic interactions ⁴³) are consequently an important mechanism within this kinetic zone, and
153	provide a mechanism for loading of new organic substrates to pre-existing MAOM ^{31,49} . In the
154	zonal model, the more weakly-associated kinetic material is hypothesized to exchange readily
155	with the soil solution. Similarly, Kaiser and Guggenberger argued that the potential
156	bioavailability of MAOM depends on the degree of OM loading on mineral surfaces ^{50,51} . As
157	more organic molecules occupy binding sites, a proportion of MAOM may become more
158	susceptible to degradation due to fewer functional groups being involved in sorption. Further, the
159	enrichment of N-rich OM at the mineral surface is consistent with a zonal model driven by

160 preferential retention of N-containing functional groups (e.g., through hydrogen bonding, cation bridges, or ligand exchange)²⁸. However, in batch adsorption experiments using dissolved 161 162 organic matter (DOM) and goethite, Coward et al. found that N-containing compounds were the 163 last to adsorb, populating the "kinetic" zone, which is hypothesized to constitute the most dynamic, exchangeable fraction⁵². Variations in mineral-organic interactions, including those 164 165 regulated by variation in chemical bonds, organic matter composition and fine-scale spatial 166 heterogeneity, warrants deeper engagement with the idea of MAOM containing a fast cycling 167 and bioavailable subfraction.

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169 3. MAOM contains young and mineralizable compounds

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171 MAOM is generally characterized as old and slow-cycling due to a consistent increase in SOC mean residence time (aka 'mean system age')⁵³ with decreasing particle size. Radiocarbon 172 173 analyses demonstrate that silt- and clay-associated OM has a longer estimated turnover time than 174 sand-associated OM⁵⁴. However, a small but fast-cycling portion of MAOM (i.e., days to years) 175 may be obscured in these estimates by much slower-cycling compounds (i.e., centuries to 176 millennia) that substantially shift the mean estimate of residence time in these observational 177 studies^{55,56}. Radiocarbon tracer studies can more accurately track the rate of incorporation of new 178 inputs to MAOM, though they are rarely employed. However, one site in an oak forest with 179 accidental radiocarbon release found that up to 37% of MAOM is replaced on an annual basis, implying a mean residence time of four years⁵⁷. In the same soils, the steady-state mean 180 181 residence time estimated from natural abundance radiocarbon measurements was 108 years,

182 which suggests that natural abundance-based averages may not be a reliable measure of typical
183 MAOM residence times in topsoil⁵⁷.

184 Recent work suggests that a primary driver of skewness in radiocarbon-based SOM and MAOM age data may be organic fossil C in sedimentary soil parent materials, including 185 186 kerogen, lignite, and coal, which could contribute substantially to MAOM C pools. The 187 presence of fossil C may skew radiocarbon ages in MAOM and other SOM fractions^{58–60}. The 188 use of ramped thermal analysis has proved an effective means of illustrating how the average 189 radiocarbon value of a MAOM fraction may obscure the strongly contrasting "ages" of its 190 constituent compounds, which may range from post-bomb (modern) to nearly radiocarbon dead $(\sim 60,000 \text{ years old or older})^{61}$. Thus, the reliance on radiocarbon dating alone to infer the mean 191 192 age or persistence of MAOM may lead to spurious conclusions regarding MAOM durability in 193 the context of climate change and its potential to store new C additions over long timescales. 194 Mineral surfaces provide MAOM with some protection from decomposition, but they are 195 also host to active microbial communities. Mineral surfaces support high microbial biomass and 196 accelerated rates of microbial turnover and enzyme production^{62–64}. The abundance of low-197 molecular weight and low C:N compounds support a high mineralization potential within MAOM ⁶⁵. The intrinsic bioavailability of MAOM is illustrated by several incubation-based 198 199 experiments. In a study of 156 diverse soils from across the United States, Yu et al. demonstrated 200 that both MAOM and POM are strong predictors of whole soil C decomposition dynamics⁶⁶. 201 They suggest that the large pool size of MAOM may compensate for its lower decomposition 202 rates in some soils creating similar overall contributions to CO_2 flux as POM. When SOM 203 fractions are incubated under ideal temperature and moisture conditions, MAOM can even

exhibit higher N and C mineralization potential compared to POM or light fractions ^{67–72}. This
 was recently observed for soils collected from forest, grassland, and cropland land uses ⁷³.

206 Two main methodological approaches provide direct estimates of the potentially 207 bioavailable and fast-cycling portion of the MAOM pool: (1) isotope tracer mineralization 208 experiments and (2) batch adsorption-desorption experiments. Isotope tracer mineralization 209 experiments, which trace the decomposition of clay-bound organic compounds over the course 210 of days or weeks-long incubations, demonstrate an intrinsic capacity for MAOM to supply C and 211 N (Table 1). Over half of adsorbed organic compounds in such experiments were bioavailable 212 and mineralizable, with the magnitude of decomposition often mediated by mineral type (Table 213 1).

214 Batch adsorption-desorption studies highlight how rapidly compounds may desorb from 215 mineral surfaces (Table 2). Such experiments quantify the equilibrium partitioning of a sorbate between solid and solution phases, in a liquid suspension over 24-48 hours⁷⁴ and using a wide 216 217 range in soil:solution ratios (1:4 to 1:10,000). Results demonstrate the rapid and largely 218 irreversible adsorption of organic compounds. However, approximately half of adsorbed OC can 219 be removed with water within a few hours, which highlights the high potential bioavailability of 220 MAOM. Reported rates of desorption vary widely due to the variety of experimental conditions 221 including mineral type, desorption agents, background electrolytes, and pH. Likewise, the short 222 duration, highly disturbed, and often artificial context limit our ability to interpret and translate 223 results to natural soil systems. Nonetheless, laboratory experiments tracing the desorption and 224 mineralization of OC from clays as in Tables 1 and 2 highlight the potential for microbes and 225 plant roots to access and decompose MAOM. As will be discussed below, there are certain

ecological contexts and processes that may allow for this mineralization potential to be realizedat the ecosystem scale.

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229 4. Ecological Drivers of MAOM Destabilization

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231 While there has been much emphasis on the controls of SOM formation and persistence, a 232 growing body of research has focused on the drivers of SOM destabilization²⁰. Forces of 233 destabilization have been categorized into those that disrupt soil physical structure (such as soil 234 aggregates), those that disrupt mineral-organic associations, and those that enhance C metabolism²⁰. Below, we focus on three proximal drivers of MAOM destabilization: plant-235 236 microbial interactions, climate change, and agricultural intensification. The transfer of OM from 237 mineral-associated to more bioavailable pools will depend on the presence and strength of 238 particular environmental conditions that favor MAOM destabilization. 239 240 4.1. Plant-microbial forces

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Rhizosphere environments provide conditions that can facilitate the destabilization and turnover of MAOM. MAOM in the rhizosphere is more likely to be released from mineral surfaces than MAOM in bulk soil due to a higher concentration of plant and microbial exudates in the soil near active roots^{75,76}. These exudates may stimulate specific extracellular enzymes or more generally stimulate decomposition (i.e. glucose, organic acids). Additionally, plant and microbial exudates can form associations with soil minerals directly, contributing to the MAOM pool⁷⁷. In fact, rootderived organic matter, particularly root exudates, is suggested to be a dominant source of 249 mineral-associated C and N^{13,78–81}. As a result, rhizosphere MAOM acts as both a source and
250 sink for organic compounds and is likely more dynamic compared to MAOM in the bulk soil.

251 MAOM may be particularly vulnerable to destabilization by plant exudates in ecosystems 252 with rapidly growing vegetation and high demand for soil nutrients. These conditions are 253 common in early successional forests, agricultural systems, and ecosystems recovering from disturbance or responding to global change^{82–84}. Some evidence for these patterns comes from 254 255 analyses of MAOM radiocarbon content where in the eastern U.S., MAOM turns over more quickly in ecosystems with higher root mass⁸⁵. This suggests that ecosystems with high plant 256 257 nutrient demand may have a higher proportion of fast-cycling MAOM compared to other 258 ecosystems, particularly in the rhizosphere.

259 The rhizosphere is also characterized by the presence and enrichment of mineral-260 weathering soil bacteria. Rhizosphere activity can accelerate the formation of metal oxides⁸⁶ or facilitate rock weathering and clay transformation processes^{87–89}. Phosphate-dissolving bacteria 261 have been reported in the rhizosphere of agricultural soils ⁹⁰ and around the roots of mangrove 262 263 trees ⁹¹. Similarly, the dissolution of clay particles was highest in rhizosphere of Norway spruce and oak compared to bulk soil ⁹². Although mineral weathering is generally seen as a slow 264 265 pedogenic process, it can occur over short time scales, especially in the microbially active zone 266 around roots. Processes occurring at the root-soil interface can significantly accelerate pedogenesis such that mineral dissolution or alteration can occur within 20 years^{92,93} or even 267 within a growing season⁹⁴. It is unclear how these localized, accelerated alterations to clay 268 particles in the rhizosphere alter the formation and cycling of MAOM specifically. 269

270 Mycorrhizal fungi, which receive energy-rich carbon compounds from plant hosts, are
271 equipped to destabilize and mobilize MAOM. The capacity for ectomycorrhizal fungi (ECM) to

272 liberate MAOM was recently reviewed by Tunlid et al⁹⁵. Many ECM fungi secrete low 273 molecular weight compounds like oxalic acid that can disrupt mineral-organic associations ⁹⁶, dissolve minerals ⁹⁷, release nutrients ⁹⁵ and facilitate the direct assimilation of N from MAOM⁹⁸. 274 275 Some ECM species also mediate the reductive dissolution of iron-bearing minerals and the subsequent generation of reactive oxygen species⁹⁹. MAOM was shown to be more sensitive to 276 277 hydroxyl radicals than POM, which was attributed to the enrichment in MAOM of low molecular weight compounds with lower activation energies¹⁰⁰. Thus, ECM-facilitated 278 279 dissolution of minerals may accelerate the decomposition of MAOM. Arbuscular mycorrhizal 280 fungi (AMF) also generate many of the same compounds with known weathering capacities^{101,102} 281 but may have limited capacity to mobilize MAOM. AMF rely on bacterial-mediated decomposition to access inorganic N¹⁰³ as their enzymes are not as effective at mining nutrients 282 283 directly from SOM. AMF-generated organic acids were shown to mobilize phosphorus bound to iron oxides¹⁰⁴, but this area of research remains highly understudied. Overall, evidence suggests 284 285 that the activities of roots, mycorrhizal and free-living microbes contribute substantially to 286 MAOM mobilization, likely making the rhizosphere a hotspot for dynamic and fast-cycling 287 MAOM.

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289 4.2. Climate-related forces

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Climate is a major driver of plant growth, microbial activity, and abiotic properties of soils.
Therefore, it is also likely to alter the formation and destabilization of MAOM, with direct
implications for the fast-cycling MAOM pool. Shifts in climate may disrupt and alter MAOM
formation and destabilization processes, potentially limiting MAOM accrual and/or accelerating

the loss of MAOM. Multiple studies report how changes in precipitation, temperature, and
atmospheric CO₂ may destabilize MAOM across varied ecosystems.

297 Given projected shifts in climate toward intensification of hydrologic cycles, it is 298 important to understand how shifting moisture regimes may alter fast-cycling MAOM. In arid 299 systems where moisture limitation is a physiological control on microbial mineralization, 300 changing precipitation patterns (and increased DOC fluxes to subsoil) may facilitate MAOM 301 losses⁸⁵. Moisture availability indices (e.g., MAP/PET) and correlated moisture-driven 302 mineralogy gradients (e.g., reactive metal vs. base cation-dominated mineralogy) have therefore 303 been applied as organizing concepts to predict differences in MAOM formation mechanisms^{105–} ¹⁰⁸ and susceptibility to destabilization under changing moisture availability across broad-scale 304 305 climatic regions¹⁰⁷.

306 In addition to the role of moisture regime at the ecosystem scale, MAOM destabilization 307 processes are influenced by moisture-driven changes in soil redox status at the pore scale. Most 308 notably, the reduction of iron-bearing minerals under saturated conditions can release mineral-309 bound C into soluble forms, leading to potential net losses of C following a return to oxic conditions^{109–113}. However, saturation may also cause more dynamic sorption-desorption events 310 in soils less dominated by iron-bearing minerals¹¹⁴. In environments experiencing frequent wet-311 312 dry cycles, shifting aerobic to anaerobic conditions has been shown to increase the 313 bioavailability of MAOM, possibly due to a loss of Fe-MAOM associations⁴³. Wet-dry cycling 314 was also shown to enhance the decomposition of MAOM and its vulnerability to simulated 315 exudates¹¹⁵. This suggests that increases in moisture variability due to climate intensification 316 may promote a larger fast-cycling MAOM pool.

317	Temperature is an important control on SOM decomposition rates ¹¹⁶ , but the sensitivity
318	of faster-cycling MAOM components to changing temperature is less well described. In general,
319	MAOM is expected to be less temperature-sensitive overall compared to POM ^{117,118} . However,
320	there have been few direct studies of the responses of different components of MAOM to
321	changes in temperature. Available evidence suggests larger and lighter components of MAOM
322	are more responsive to warming ^{119,120} . One possibility, which warrants further investigation, is
323	that larger and lighter portions of MAOM are held more loosely to mineral surfaces, making
324	them more susceptible to mineralization with increased microbial activity. In addition, Possinger
325	et al. found that OM protected by reactive metal associations was more vulnerable to warming
326	temperatures than OM protected by base cations, potentially due to the temperature sensitivity of
327	biological processes involved in overcoming stable organo-metal interaction mechanisms ¹⁰⁷ .
328	As the CO ₂ fertilization effect promotes higher rates of photosynthesis, plants require
329	more nutrients from soil, particularly N ¹²¹ . As a result, rising atmospheric CO ₂ concentrations
330	may indirectly promote MAOM desorption and turnover as a consequence of increasing plant
331	nutrient demand. In ecosystems without a large standing stock of bioavailable N, plants may rely
332	on MAOM as a source of limiting nutrients to fuel higher rates of productivity. This may occur
333	through greater root exudation ⁸⁴ , which could cause desorption of N-rich MAOM that would
334	subsequently be available for mineralization. However, rhizosphere-induced priming may also
335	act on the POM pool rather than the MAOM pool ^{122,123} . In support of minimal MAOM
336	responses, a meta-analysis of global change manipulations found no significant effect of elevated
337	CO_2 (eCO ₂) on MAOM-C concentrations ¹¹⁸ . However, in studies conducted for more than five
338	years, MAOM-C concentrations tended to decrease with eCO_2^{118} . This pattern suggests that the
339	effects of eCO ₂ may not be immediately apparent, but could become more evident over time as

the diminishing soil nutrient pool prompts plants and microbes to begin mining resources fromMAOM.

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344 4.3. Land use change and agricultural management

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346 In general, land management that alters the input and microbial processing of OM will influence 347 MAOM cycling¹²⁴. Agricultural management systems may shift disturbance regimes, external 348 inputs, and plant diversity, as well as the abiotic environment (e.g. temperature and moisture), with both biotic and abiotic drivers influencing MAOM^{115,125,126}. For example, the physical 349 350 disruption of tillage and changes in irrigation can disproportionately impact the MAOM protected within macro-aggregates¹²⁷. MAOM turnover also depends on the C:N ratio and 351 352 chemical composition of organic inputs. For example, a clover-rye mixed cover crop facilitated transfer of POM-C to MAOM-C more than a rye or clover crop alone¹²⁸. Manure additions have 353 354 been linked to faster MAOM turnover times than synthetic inputs¹²⁹, perhaps because inorganic fertilizer applications may suppress the biological mechanisms that mobilize MAOM ¹³⁰. 355 356 MAOM response to management depends on both MAOM formation and loss; 357 management that increases soil microbial activity can also enhance MAOM mineralization. The 358 balance between stabilization and destabilization processes is heavily context-dependent and may result in either increases or decreases in MAOM²⁰. For example, in a study on intensively 359 360 managed Mollisols, incorporating legume cover crops and adding manure did not result in increased MAOM as hypothesized¹³¹. The researchers attributed this finding to nutrient mining 361 362 of MAOM by the maize crop, which offset potential gains in MAOM due to enhanced microbial 363 efficiency and biomass production. Similarly, long-term organic fertilization increased MAOM-364 C turnover, but not accrual¹²⁹. Organic cropping with manure on an Alfisol also did not increase MAOM¹³², but adding a legume cover crop did increase MAOM in an arid wheat cropping 365 366 systems on silt loam soils¹³³. These studies highlight how measuring net changes overlooks 367 active-cycling MAOM, which may be a large flux in systems that experience high rates of both 368 MAOM loss and accrual. This fast-cycling MAOM may prove to be important in predicting N 369 available to crops, as N mineralization remains an important, and poorly predicted, element of crop yield^{130,134–136}. 370

371 In some cases, management may not alter POM or MAOM quantity, but rather cause 372 shifts in the chemical composition of one or both fractions. For example, MAOM chemical 373 composition shifted in response to contrasting cover crop functional types¹³⁷ and organic 374 cropping systems with cover crops and manure¹³², even though MAOM-C concentration was not 375 affected. Similarly, Teixeira et al. observed that with the shift from tropical savanna to pasture, losses of savanna-derived MAOM-C were offset by gains from pasture-derived C¹³⁸. These shifts 376 377 in MAOM chemical composition or quantity may correspond with faster-cycling MAOM, but it 378 is uncertain how to measure or manage plant access to this nutrient supply.

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381 5. Implications of the multi-pool and dynamic nature of MAOM

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383 The implications of a fast-cycling MAOM pool for both agricultural management and

384 representation within Earth system models have largely gone unexplored. Below, we discuss the

relevance of fast-cycling MAOM for these applications and highlight key research opportunitiesto improve our understanding and measurement of this MAOM pool's importance.

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388 5.1. Managing fast-cycling MAOM in managed landscapes

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390 While some management practices may destabilize MAOM, there are other practices that may be 391 harnessed to promote fast-cycling MAOM for multiple benefits such as soil nutrient availability 392 and accrual of soil carbon. A recent review by Daly et al. outlined several key hypotheses about 393 the potential role of the fast-cycling MAOM pool that require further testing¹³⁰. First, fast-394 cycling MAOM can provide key nutrients for crops, particularly in low input systems that 395 promote plant investment in root production and mycorrhizal symbioses¹³⁰. Furthermore, fast-396 cycling MAOM may play a key role in the accumulation of more persistent SOC. Movement of 397 compounds into and out of the fast-cycling MAOM fraction could help equilibrate the 398 concurrent, though seemingly opposed, processes of SOM accrual and SOM mobilization in 399 soils. Fast-cycling MAOM fraction could briefly stabilize soluble C and N compounds during 400 periods when their concentrations are high, thereby acting as a temporary buffer against losses of 401 C and N via leaching, microbial respiration, and transformations like denitrification. For 402 example, in agroecosystems, the rapid, early season movement of inorganic N fertilizers into 403 microbial biomass and MAOM can enhance its availability later, during periods of peak crop 404 growth. When soluble SOM pools are low, desorption and decomposition of fast-cycling MAOM 405 could supply plants and microbes with energy and nutrients, and thus may regulate short-term 406 nutrient availability. Over time, some fast-cycling MAOM may form stronger bonds with 407 mineral surfaces or be incorporated into larger SOM complexes like aggregates, thereby

transforming it to slower-cycling MAOM. This pipeline of loss-prone dissolved compounds
moving through fast-cycling MAOM into persistent SOM may be key to increasing retention of
C, N, and other nutrients in the soil system.

411 Ultimately, the ecosystem and management implications of rapidly cycling MAOM 412 depend on the fate(s) of MAOM following desorption. Fast-cycling MAOM may be functionally 413 similar to POM, but the lower C:N ratio of MAOM suggests that it should be utilized relatively 414 efficiently by microbes, resulting in less CO₂ losses and greater C and N incorporation into 415 microbial biomass and metabolites compared to POM. This cycle of SOM temporarily sorbing to 416 mineral surfaces, undergoing microbial consumption and transformation, and re-release into soil 417 solution as DOM, has been suggested to result in the "cascade theory" of SOM downward 418 translocation¹³⁹. This theory is supported by the observation of relatively older, more 419 microbially-processed SOM in deeper soil horizons, as well as by empirical observations of 420 DOM chemical composition changes during downward transport through grassland soil profiles 421 using ultrahigh-resolution mass spectrometry¹⁴⁰. Specific surface soil management strategies 422 promoting SOC transport to, and accumulation within, deeper soils remain a critical knowledge gap ¹⁴¹, and fast cycling MAOM may be key to this process. Research focusing specifically on 423 424 MAOM dynamics across soil depths could help clarify our understanding of the fate and 425 importance of fast-cycling MAOM in long-term C storage.

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427 5.2. Integrating fast-cycling MAOM into models of soil C and N dynamics

428

Earth system models typically conceptualize soil carbon as multiple pools that decompose via
first-order decay kinetics and differ in their turnover times¹⁴². In the last decade, models that

431 explicitly include nonlinear microbial-mineral interactions are becoming more prevalent but vary

432 widely in their process representations and parameterizations ^{143–146}. Mechanisms of

433 stabilization vary from DOC sorption only ^{147,148} to accumulation of microbial necromass^{146,149} or

434 chemically recalcitrant compounds 123 to a combination of the two $^{150-153}$. As a result of these

435 different model formulations, the predicted temporal dynamics of SOM, and the underlying

436 pools conceptualized as MAOM, can vary widely in response to disturbance^{143,154}.

437 The vast majority of SOM models, including all models used at global scales, assume that 438 modeled pools are homogenous and that any particle within a pool has an equal probability of entering or leaving as its neighbor¹⁴². Within this framework, turnover time and age is estimated 439 as a single value for each pool, such that representing MAOM of varying ages would require 440 441 increasing the number of pools and defining specific stabilization mechanisms or a combination 442 of mechanisms for each pool. However, when MAOM is explicitly represented in SOM models, it is most often defined as a single pool^{147,148,150,152}. Brunmayr et al. found that multiple of these 443 444 SOM models overestimate MAOM turnover and suggested further dividing the MAOM pool to better represent components with different turnover times¹⁵⁵. 445

446 Select models do include a second MAOM pool that is considered faster cycling – for example, the – for example, the 'exchangeable MAOM' pool in the MEMS v2 model¹⁵⁵, the 'Q' 447 pool in the MEND model¹⁵³, and the 'Q DOM' pool in the COMISSION model¹⁵¹, which all 448 449 represent DOC sorbed to minerals that can readily exchange with DOC in solution. However, 450 parameterizing sorption/desorption rates for these distinct MAOM pools, and benchmarking their 451 relative sizes, poses a significant challenge due to data limitations especially at larger scales. 452 Alternatively, other modeling frameworks exist that focus on a continuum of individual particles 453 traveling through model pools, with the ability to estimate the distribution of ages and transit

times within a single pool^{53,156,157}. Models can also represent the fast-cycling MAOM pool by
allowing for MAOM desorption at relatively high rates; a study using the SOMic model, which
represents microbial dynamics and allows for rapid MAOM desorption, found that this modeling
framework generated predictions of global SOC distribution and long-term SOC dynamics that
were closely aligned with empirical data¹⁵⁸. Regardless of the modeling framework, the existence
of fast-cycling MAOM suggests that modeling MAOM as a single homogenous, passively
cycling SOM pool is likely insufficient for accurately capturing temporal dynamics.

461

462 5.3 Measurement of fast-cycling MAOM

463

464 Although SOM fractionation methods drive our operational and conceptual understanding of 465 POM and MAOM pools, we lack a standard method for directly quantifying the fast-cycling 466 MAOM pool. MAOM can be further separated along physical (density, particle size) and 467 chemical (solubility in acid or base, oxidation, thermal lability) gradients, which may, in theory, 468 be able to isolate a more bioavailable subfraction. However, these methods also often require 469 further dispersion and disruption that may facilitate the inter-fraction transfer of C and N. 470 Common procedures for isolating SOM fractions may remove or obscure the fast-cycling portion 471 of MAOM. Fractionation procedures require a degree of mixing and dispersion in liquid that will 472 release and redistribute DOM between POM and MAOM. The quantity of DOM leached during fractionation can be significant¹⁵⁹ and it is currently not possible to distinguish between POM or 473 474 MAOM in leached compounds. MAOM that desorbs and enters dissolved pools during 475 fractionation may be an important component of fast-cycling MAOM. More studies are needed 476 to identify the extent of leaching under contrasting fractionation and dispersion conditions as it is possible that certain methods favor the transfer of POM C or N into the MAOM pool, potentiallyinflating the stabilization capacity of this fraction.

479 Sequential chemical extractions can isolate phases of MAOM that differ in solubility and 480 potentially bioavailability. Selective dissolutions with Na-pyrophosphate, hydroxylamine, and 481 dithionite-HCl can be used to target organo-metal complexes, short-range order Al and Fe hydroxides, and crystalline Fe hydroxides, respectively¹⁶⁰. These well-stabilized forms of 482 483 MAOM are still vulnerable to loss, such as the iron-associated carbon that is destabilized and released by low molecular weight organic acids or under saturated conditions^{111,161}. Physical and 484 485 chemical fractionation methods can also be combined with thermal and/or chemometric 486 approaches to estimate the chemical and biological stability of particle size fractions of SOM¹⁶². 487 Otherwise, a combined chemical and biological assay is a potentially promising method for 488 simulating destabilizing agents and the conditions that accelerate MAOM cycling. For example, 489 by incubating isolated MAOM fractions with organic acids and glucose to simulate root 490 exudates, one can estimate the destabilization potential of MAOM¹⁶³. However, this has not been 491 examined comprehensively across soil types and experimental conditions to generate a standard 492 method.

Radiocarbon and ¹³C-based experiments can provide estimates of pool turnover times.
However, these methods provide an average turnover time, which may obscure the smaller,
faster cycling subfraction, unless paired with ramped thermal analysis techniques that can
quantify an underlying distribution of turnover times. MAOM generally has a wide distribution
of radiocarbon mean ages, with considerable overlap with the distribution for POM ^{10,108}.
Measurements of CO₂ respiration in long-term incubations can be used to calculate the size and
turnover rate of active and slow pools of C¹⁶⁴ although calculated residence times may vary

500 based on experiment duration¹⁶⁵ and these incubations are unable to determine the origin of the 501 CO₂. Isotope tracer-based experiments in which POM and MAOM fractions are differentially 502 labelled would allow for the tracing of fast-cycling MAOM and provide critical, mechanistic 503 insights into the potential size of and controls on this pool. In general, more research is needed to 504 identify a consistent method and guidance for how to pair fractionation methods with isotope 505 tracer and/or spectroscopic/imaging methods to isolate the various MAOM sub-fractions. 506 507 508 6. Conclusion 509 510 For decades, the soil science community has acknowledged the existence of fast-cycling 511 MAOM. Although the majority of MAOM is highly persistent and cycles on decadal and 512 millennial time-scales, a portion is bioavailable, exchangeable, and an active contributor to C and 513 N fluxes. As we have summarized, the size of this pool will likely depend on the intrinsic 514 properties of the MAOM – namely, the availability and physicochemical properties of minerals 515 as well as the composition, quantity, and structure of the associated organic matter. In addition, 516 fast-cycling MAOM will likely respond readily to drivers of destabilization, such as plant-517 microbe interactions, climate change, agricultural intensification, and land use change. 518 Therefore, clarifying its size and responses in varying ecological scenarios is important to 519 improve model predictions and make better recommendations for land managers. While 520 counterintuitive to the goals of soil C sequestration, within certain contexts, it may be 521 desirable/beneficial to promote the existence of a fast-cycling MAOM pool. Accurate 522 quantification of this pool and its role in ecological processes will enhance our understanding of

523 these contexts and could inform agricultural management approaches. Likewise, our previous 524 understanding of MAOM as a persistent, slowly-aggrading pool is clearly oversimplified, and 525 including more realistic representations of fast-cycling MAOM in Earth system models is needed 526 to accurately quantify both the current and future size of this SOM pool. These applications are 527 currently hindered by methodological limitations that have made it challenging to characterize 528 this dynamic pool of MAOM. Continued development of methods to isolate and trace fast-529 cycling MAOM will enable better quantification of this pool and subsequent integration of these 530 insights into modeling and soil management practices. 531 532 7. Acknowledgements 533 Work by A.J. was funded by the National Science Foundation Award 2103187. K.G. was 534 supported by the LLNL-LDRD Program Project No. 24-LW-053 under the auspices of DOE 535 Contract DE-AC52-07NA27344. Work conducted at LLNL was supported by the U.S. 536 Department of Energy, Office of Biological and Environmental Research, Genomic Science 537 Program 'Microbes Persist' Scientific Focus Area (#SCW1632), and was conducted under the auspices of the U.S. Department of Energy under Contract DE-AC52-07NA27344. We thank 538 539 science illustrator Elena Hartley for the design of the figure. 540 541 8. Conflict of Interests Statement 542 The authors declare no conflicts of interest. 543 544 References 545 Grandy, A. S. & Neff, J. C. Molecular C dynamics downstream: The biochemical decomposition 1. 546 sequence and its impact on soil organic matter structure and function. Sci. Total Environ. 404, 297-

547 307 (2008).

549		abundance and persistence. Glob. Change Biol. 28, 1178-1196 (2022).
550	3.	Torn, M. S., Trumbore, S. E., Chadwick, O. A., Vitousek, P. M. & Hendricks, D. M. Mineral
551		control of soil organic carbon storage and turnover. Nature 389, 170-173 (1997).
552	4.	Lehmann, J., Kinyangi, J. & Solomon, D. Organic matter stabilization in soil microaggregates:
553		implications from spatial heterogeneity of organic carbon contents and carbon forms.
554		<i>Biogeochemistry</i> 85 , 45–57 (2007).
555	5.	Leuthold, S., Lavallee, J. M., Haddix, M. L. & Cotrufo, M. F. Contrasting properties of soil organic
556		matter fractions isolated by different physical separation methodologies. Geoderma 445, 116870
557		(2024).
558	6.	Bu, R. et al. Particulate organic matter affects soil nitrogen mineralization under two crop rotation
559		systems. <i>PLoS ONE</i> 10 , (2015).
560	7.	Angst, G. et al. Unlocking complex soil systems as carbon sinks: multi-pool management as the
561		key. Nat. Commun. 14, 2967 (2023).
562	8.	Chenu, C. & Plante, A. F. Clay-sized organo-mineral complexes in a cultivation chronosequence:
563		revisiting the concept of the 'primary organo-mineral complex'. Eur. J. Soil Sci. 57, 596-607
564		(2006).
565	9.	Cotrufo, F. M. et al. Formation of soil organic matter via biochemical and physical pathways of
566		litter mass loss. Nat. Geosci. (2015) doi:10.1038/NGEO2520.
567	10.	Cotrufo, M. F. & Lavallee, J. M. Chapter One - Soil organic matter formation, persistence, and
568		functioning: A synthesis of current understanding to inform its conservation and regeneration. in
569		Advances in Agronomy (ed. Sparks, D. L.) vol. 172 1-66 (Academic Press, 2022).
570	11.	Totsche, K. U. et al. Microaggregates in soils. J. Plant Nutr. Soil Sci. 181, 104-136 (2018).
571	12.	Even, R. J. & Francesca Cotrufo, M. The ability of soils to aggregate, more than the state of
572		aggregation, promotes protected soil organic matter formation. Geoderma 442, 116760 (2024).

Heckman, K. et al. Beyond bulk: Density fractions explain heterogeneity in global soil carbon

548

2.

- 573 13. Sokol, N. W., Sanderman, J. & Bradford, M. A. Pathways of mineral-associated soil organic matter
 574 formation: Integrating the role of plant carbon source, chemistry, and point of entry. *Glob. Change*575 *Biol.* 25, 12–24 (2019).
- 576 14. Poirier, V. *et al.* Organo-Mineral Interactions Are More Important for Organic Matter Retention in
 577 Subsoil Than Topsoil. *Soil Syst.* 4, 4 (2020).
- 578 15. Haddix, M. L., Paul, E. A. & Cotrufo, M. F. Dual, differential isotope labeling shows the
- 579 preferential movement of labile plant constituents into mineral-bonded soil organic matter. *Glob.*580 *Change Biol.* 22, 2301–2312 (2016).
- 581 16. Fulton-Smith, S., Even, R. & Cotrufo, M. F. Depth impacts on the aggregate-mediated mechanisms
 582 of root carbon stabilization in soil: Trade-off between MAOM and POM pathways. *Geoderma* 452,
- **583** 117078 (2024).
- 584 17. Shabtai, I. A. *et al.* Calcium promotes persistent soil organic matter by altering microbial
 585 transformation of plant litter. *Nat. Commun.* 14, 6609 (2023).
- 18. Angst, G. et al. Soil organic carbon stocks in topsoil and subsoil controlled by parent material,
- 587 carbon input in the rhizosphere, and microbial-derived compounds. *Soil Biol. Biochem.* 122, 19–30
 588 (2018).
- 589 19. Barré, P., Fernandez-Ugalde, O., Virto, I., Velde, B. & Chenu, C. Impact of phyllosilicate
- 590 mineralogy on organic carbon stabilization in soils: incomplete knowledge and exciting prospects.
 591 *Geoderma* 235–236, 382–395 (2014).
- 592 20. Bailey, V. L., Pries, C. H. & Lajtha, K. What do we know about soil carbon destabilization?
 593 *Environ. Res. Lett.* 14, 083004 (2019).
- 594 21. Dynarski, K. A., Bossio, D. A. & Scow, K. M. Dynamic Stability of Soil Carbon: Reassessing the
 595 "Permanence" of Soil Carbon Sequestration. *Front. Environ. Sci.* 8, (2020).
- 596 22. Piñeiro, G., Paruelo, J. M., Jobbágy, E., Jackson, R. B. & Oesterheld, M. Grazing effects on
- belowground C and N stocks along a network of cattle exclosures in temperate and subtropical
- 598 grasslands of South America. *Glob. Biogeochem. Cycles* 23, (2009).

- Woodmansee, R. G. & Duncan, D. A. Nitrogen and phosphorus dynamics and budgets in annual
 grasslands. *Ecology* 61, 893–904 (1980).
- 601 24. Kleber, M., Sollins, P. & Sutton, R. A conceptual model of organo-mineral interactions in soils:
- self-assembly of organic molecular fragments into zonal structures on mineral surfaces.
- 603 *Biogeochemistry* **85**, 9–24 (2007).
- 604 25. Torn, M. S. *et al.* A dual isotope approach to isolate soil carbon pools of different turnover times.
 605 *Biogeosciences* 10, 8067–8081 (2013).
- 606 26. Kleber, M. *et al.* Dynamic interactions at the mineral–organic matter interface. *Nat. Rev. Earth*607 *Environ.* 2, 402–421 (2021).
- Lavallee, J. M., Soong, J. L. & Cotrufo, M. F. Conceptualizing soil organic matter into particulate
 and mineral-associated forms to address global change in the 21st century. *Glob. Change Biol.*(2019) doi:10.1111/gcb.14859.
- 611 28. Kleber, M. *et al.* Mineral-organic associations: formation, properties, and relevance in soil
 612 environments. *Adv. Agron.* 130, 1–140 (2015).
- 613 29. Mikutta, R. *et al.* Biogeochemistry of mineral-organic associations across a long-term mineralogical
- 614 soil gradient (0.3-4100 kyr), Hawaiian Islands. *Geochim. Cosmochim. Acta* 73, 2034–2060 (2009).
- 615 30. Sanderman, J., Maddern, T. & Baldock, J. Similar composition but differential stability of mineral
- retained organic matter across four classes of clay minerals. *Biogeochemistry* **121**, 409–424 (2014).
- 617 31. Underwood, T. R., Bourg, I. C. & Rosso, K. M. Mineral-associated organic matter is heterogeneous
- and structured by hydrophobic, charged, and polar interactions. *Proc. Natl. Acad. Sci.* 121,
- e2413216121 (2024).
- 620 32. Kögel-Knabner, I. et al. Organo-mineral associations in temperate soils: Integrating biology,
- 621 mineralogy, and organic matter chemistry. J. Plant Nutr. Soil Sci. 171, 61–82 (2008).
- 622 33. Scheidegger, A. M. & Sparks, D. L. A critical assessment of sorption-desorption mechanisms at the
- 623 soil mineral/water interface. *Soil Sci.* 161, 813 (1996).

34. Bramble, D. S. E. *et al.* Formation of mineral-associated organic matter in temperate soils is
primarily controlled by mineral type and modified by land use and management intensity. *Glob.*

626 *Change Biol.* **30**, e17024 (2024).

- 627 35. Deng, Y. & Dixon, J. B. Soil Organic Matter and Organic-Mineral Interactions. in *Soil Mineralogy*628 *with Environmental Applications* 69–107 (John Wiley & Sons, Ltd, 2002).
- 629 doi:10.2136/sssabookser7.c3.
- 630 36. Lv, J. *et al.* Molecular-Scale Investigation with ESI-FT-ICR-MS on Fractionation of Dissolved
 631 Organic Matter Induced by Adsorption on Iron Oxyhydroxides. *Environ. Sci. Technol.* 50, 2328–
 632 2336 (2016).
- 633 37. Oren, A. & Chefetz, B. Sorptive and Desorptive Fractionation of Dissolved Organic Matter by
 634 Mineral Soil Matrices. *J. Environ. Qual.* 41, 526–533 (2012).
- 635 38. Kramer, M. C., Sanderman, J., Chadwick, O. A., Chorover, J. & Vitousek, P. M. Long term carbon
 636 storage through retention of dissolved aromatic acids by reactive particles in soil. *Glob. Change*637 *Biol.* 18, 2594–2605 (2012).
- 838 39. Rocci, K. S. *et al.* Impacts of nutrient addition on soil carbon and nitrogen stoichiometry and
 stability in globally-distributed grasslands. *Biogeochemistry* 159, 353–370 (2022).
- 640 40. Heckman, K. et al. Sorptive fractionation of organic matter and formation of organo-hydroxy-
- aluminum complexes during litter biodegradation in the presence of gibbsite. *Geochim. Cosmochim. Acta* 121, 667–683 (2013).
- 643 41. Inagaki, T. M. et al. Subsoil organo-mineral associations under contrasting climate conditions.
- 644 *Geochim. Cosmochim. Acta* **270**, 244–263 (2020).
- Keiluweit, M. *et al.* Nano-scale investigation of the association of microbial nitrogen residues with
 iron (hydr)oxides in a forest soil O-horizon. *Geochim. Cosmochim. Acta* 95, 213–226 (2012).
- 647 43. Possinger, A. R. *et al.* Organo-mineral interactions and soil carbon mineralizability with variable
- saturation cycle frequency. *Geoderma* **375**, 114483 (2020).

- 649 44. Rillig, M. C., Caldwell, B. A., W??sten, H. A. B. & Sollins, P. Role of proteins in soil carbon and
 650 nitrogen storage: Controls on persistence. *Biogeochemistry* 85, 25–44 (2007).
- 45. Spohn, M. Preferential adsorption of nitrogen- and phosphorus-containing organic compounds to
 minerals in soils: A review. *Soil Biol. Biochem.* 194, 109428 (2024).
- 46. Kinyangi, J. *et al.* Nanoscale Biogeocomplexity of the Organomineral Assemblage in Soil. *Soil Sci. Soc. Am. J.* **70**, 1708–1718 (2006).
- 47. Lehmann, J. *et al.* Spatial complexity of soil organic matter forms at nanometre scales. *Nat. Geosci*1, 238–242 (2008).
- 48. Steffens, M. *et al.* Identification of Distinct Functional Microstructural Domains Controlling C
 Storage in Soil. *Environ. Sci. Technol.* 51, 12182–12189 (2017).
- 49. Vogel, C. *et al.* Submicron structures provide preferential spots for carbon and nitrogen
 sequestration in soils. *Nat. Commun.* 5, 2947 (2014).
- 50. Kaiser, K. & Guggenberger, G. Mineral surfaces and soil organic matter. *Eur J Soil Sci.* 54, 219–
 236 (2003).
- 51. Kaiser, K. & Guggenberger, G. Sorptive stabilization of organic matter by microporous goethite:
 sorption into small pores vs. surface complexation. *Eur. J. Soil Sci.* 58, 45–59 (2007).
- 52. Coward, E. K., Ohno, T. & Sparks, D. L. Direct Evidence for Temporal Molecular Fractionation of
 Dissolved Organic Matter at the Iron Oxyhydroxide Interface. *Environ. Sci. Technol.* 53, 642–650
 (2019).
- 668 53. Sierra, C. A., Hoyt, A. M., He, Y. & Trumbore, S. E. Soil Organic Matter Persistence as a
- 669 Stochastic Process: Age and Transit Time Distributions of Carbon in Soils. *Glob. Biogeochem*.
- 670 *Cycles* **32**, 1574–1588 (2018).
- 671 54. Feng, W. *et al.* Methodological uncertainty in estimating carbon turnover times of soil fractions.
 672 *Soil Biol. Biochem.* 100, 118–124 (2016).

- 673 55. Hall, S. J., McNicol, G., Natake, T. & Silver, W. L. Large fluxes and rapid turnover of mineral-
- associated carbon across topographic gradients in a humid tropical forest: insights from paired 14C
 analysis. *Biogeosciences* 12, 2471–2487 (2015).
- 676 56. Schrumpf, M. & Kaiser, K. Large differences in estimates of soil organic carbon turnover in density
- 677 fractions by using single and repeated radiocarbon inventories. *Geoderma* **239–240**, 168–178
- 678 (2015).
- 679 57. Heckman, K. A. *et al.* Soil organic matter is principally root derived in an Ultisol under oak forest.
 680 *Geoderma* 403, 115385 (2021).
- 58. Fox, P. M. *et al.* Shale as a Source of Organic Carbon in Floodplain Sediments of a Mountainous
 Watershed. *J. Geophys. Res. Biogeosciences* 125, e2019JG005419 (2020).
- 683 59. Grant, K. E. *et al.* Diverse organic carbon dynamics captured by radiocarbon analysis of distinct
 684 compound classes in a grassland soil. *Biogeosciences* 21, 4395–4411 (2024).
- 685 60. Schrumpf, M. *et al.* Storage and stability of organic carbon in soils as related to depth, occlusion
 686 within aggregates, and attachment to minerals. *Biogeosciences* 10, 1675–1691 (2013).
- 687 61. Stoner, S. *et al.* Relating mineral–organic matter stabilization mechanisms to carbon quality and age
 688 distributions using ramped thermal analysis. *Philos. Trans. R. Soc. Math. Phys. Eng. Sci.* 381,
 689 20230139 (2023).
- 690 62. Allison, S. D. & Jastrow, J. D. Activities of extracellular enzymes in physically isolated fractions of
 691 restored grassland soils. *Soil Biol. Biochem.* 38, 3245–3256 (2006).
- 692 63. Breulmann, M. et al. Short-term bioavailability of carbon in soil organic matter fractions of
- different particle sizes and densities in grassland ecosystems. *Sci. Total Environ.* 497–498, 29–37
 (2014).
- 695 64. Kandeler, E. et al. The mineralo-sphere Succession and physiology of bacteria and fungi
- 696 colonising pristine minerals in grassland soils under different land-use intensities. *Soil Biol.*
- 697 Biochem. 136, 107534 (2019).

- 65. Jilling, A. *et al.* Minerals in the rhizosphere: overlooked mediators of soil nitrogen availability to
 plants and microbes. *Biogeochemistry* (2018) doi:10.1007/s10533-018-0459-5.
- 700 66. Yu, W., Huang, W., Weintraub-Leff, S. R. & Hall, S. J. Where and why do particulate organic
- 701 matter (POM) and mineral-associated organic matter (MAOM) differ among diverse soils? *Soil*
- 702 Biol. Biochem. 172, 108756 (2022).
- 703 67. Bailey, T. *et al.* Opposing patterns of carbon and nitrogen stability in soil organic matter fractions
 704 compared to whole soil. *Eur. J. Soil Sci.* 75, e13495 (2024).
- 70568. Bimüller, C. *et al.* Decoupled carbon and nitrogen mineralization in soil particle size fractions of a
- 706 forest topsoil. *Soil Biol. Biochem.* **78**, 263–273 (2014).
- Mueller, C. W. *et al.* Bioavailability and isotopic composition of CO2 released from incubated soil
 organic matter fractions. *Soil Biol. Biochem.* 69, 168–178 (2014).
- 709 70. Parfitt, R. L. & Salt, G. J. Carbon and nitrogen mineralisation in sand, silt, and clay fractions of soils
 710 under maize and pasture. *Aust. J. Soil Res. Aust J Soil Res* 39, 361–371 (2001).
- 711 71. Sollins, P., Spycher, G. & Glassman, C. A. Net nitrogen mineralization from light- and heavy712 fraction forest soil organic matter. *Soil Biol. Biochem.* 16, 31–37 (1984).
- 713 72. Whalen, J. K., Bottomley, P. J. & Myrold, D. D. Carbon and nitrogen mineralization from light- and
 714 heavy-fraction additions to soil. *Soil Biol. Biochem.* 32, 1345–1352 (2000).
- 715 73. Villarino, S. H. *et al.* A large nitrogen supply from the stable mineral-associated soil organic matter
 716 fraction. *Biol. Fertil. Soils* 59, 833–841 (2023).
- 717 74. Abramoff, R. Z. *et al.* How much carbon can be added to soil by sorption? *Biogeochemistry* 152,
 718 127–142 (2021).
- 719 75. Neurath, R. A. *et al.* Root Carbon Interaction with Soil Minerals Is Dynamic, Leaving a Legacy of
 720 Microbially Derived Residues. *Environ. Sci. Technol.* 55, 13345–13355 (2021).
- 721 76. Shabtai, I. A. *et al.* Root exudates simultaneously form and disrupt soil organo-mineral associations.
- 722 *Commun. Earth Environ.* 5, 1–12 (2024).

- 723 77. Chari, N. R. & Taylor, B. N. Soil organic matter formation and loss are mediated by root exudates
 724 in a temperate forest. *Nat. Geosci.* 15, 1011–1016 (2022).
- 725 78. Kaštovská, E. *et al.* Root but not shoot litter fostered the formation of mineral-associated organic
 726 matter in eroded arable soils. *Soil Tillage Res.* 235, 105871 (2024).
- 727 79. Sokol, N. W. *et al.* The path from root input to mineral-associated soil carbon is dictated by habitat728 specific microbial traits and soil moisture. *Soil Biol. Biochem.* 193, 109367 (2024).
- 80. Sokol, N. W. & Bradford, M. A. Microbial formation of stable soil carbon is more efficient from
 belowground than aboveground input. *Nat. Geosci.* 12, 46–53 (2019).
- 731 81. Villarino, S. H., Pinto, P., Jackson, R. B. & Piñeiro, G. Plant rhizodeposition: A key factor for soil
 732 organic matter formation in stable fractions. *Sci. Adv.* 7, eabd3176 (2021).
- 733 82. Bélanger, N., C^oté, B., Fyles, J. W., Courchesne, F. & Hendershot, W. H. Forest regrowth as the
- controlling factor of soil nutrient availability 75 years after fire in a deciduous forest of Southern
 Quebec. *Plant Soil* 262, 363–272 (2004).
- 736 83. Lovett, G. M. *et al.* Nutrient retention during ecosystem succession: a revised conceptual model.
 737 *Front. Ecol. Environ.* 16, 532–538 (2018).
- 73884.Phillips, R. P., Bernhardt, E. S. & Schlesinger, W. H. Elevated CO2 increases root exudation from
- 739 loblolly pine (Pinus taeda) seedlings as an N-mediated response. *Tree Physiol.* 29, 1513–1523
 740 (2009).
- 741 85. Heckman, K. A. *et al.* Moisture-driven divergence in mineral-associated soil carbon persistence.
 742 *Proc. Natl. Acad. Sci.* 120, e2210044120 (2023).
- 86. Seguin, V. *et al.* Mineral weathering in the rhizosphere of forested soils. in *Biogeochemistry of Trace Elements in the Rhizpshere* 22–55 (2005).
- 745 87. Hinsinger, P. & Jaillard, B. Root-induced release of interlayer potassium and vermiculitization of
 746 phlogopite as related to potassium depletion in the rhizosphere of ryegrass. *J. Soil Sci.* 44, 525–534
- 747 (1993).

- 88. Uroz, S. *et al.* Bacterial weathering and its contribution to nutrient cycling in temperate forest
 ecosystems. *Res. Microbiol.* 162, 821–831 (2011).
- 750 89. Zhang, Z., Huang, J., He, L. & Sheng, X. Distinct Weathering Ability and Populations of Culturable
- 751 Mineral-Weathering Bacteria in the Rhizosphere and Bulk Soils of Morus Alba. *Geomicrobiol. J.*
- **33**, 39–45 (2016).
- 753 90. Chabot, R., Antoun, H. & Cescas, M. P. Growth promotion of maize and lettuce by phosphate754 solubilizing Rhizobium leguminosarum biovar. phaseoli. *Plant Soil* **39**, 311–321 (1996).
- 755 91. Vazquez, P., Holguin, G., Puente, M. E., Lopez-Cortes, a. & Bashan, Y. Phosphate-solubilizing
- 756 microorganisms associated with the rhizosphere of mangroves in a semiarid coastal lagoon. *Biol.*757 *Fertil. Soils* 30, 460–468 (2000).
- 758 92. Calvaruso, C., Mareschal, L., Turpault, M.-P. & Leclerc, E. Rapid clay weathering in the
- rhizosphere of norway spruce and oak in an acid forest ecosystem. *Soil Sci. Soc. Am. J.* 73, 331
 (2009).
- 93. Mareschal, L., Turpault, M.-P., Bonnaud, P. & Ranger, J. Relationship between the weathering of
 clay minerals and the nitrification rate: a rapid tree species effect. *Biogeochemistry* 112, 293–309
 (2013).
- Paola, A., Pierre, B., Vincenza, C., Vincenzo, D. M. & Bruce, V. Short term clay mineral release
 and re-capture of potassium in a Zea mays field experiment. *Geoderma* 264, 54–60 (2016).
- 766 95. Tunlid, A., Floudas, D., Op De Beeck, M., Wang, T. & Persson, P. Decomposition of soil organic
 767 matter by ectomycorrhizal fungi: Mechanisms and consequences for organic nitrogen uptake and
 768 soil carbon stabilization. *Front. For. Glob. Change* 5, (2022).
- 769 96. Keiluweit, M. *et al.* Mineral protection of soil carbon counteracted by root exudates. *Nat. Clim.*770 *Change* 5, 588–595 (2015).
- 771 97. Bonneville, S. *et al.* Tree-mycorrhiza symbiosis accelerate mineral weathering: Evidences from
- nanometer-scale elemental fluxes at the hypha–mineral interface. *Geochim. Cosmochim. Acta* 75,
- **773** 6988–7005 (2011).

774	98.	Wang, T., Persson, P. & Tunlid, A. A widespread mechanism in ectomycorrhizal fungi to access
775		nitrogen from mineral-associated proteins. Environ. Microbiol. 23, 5837-5849 (2021).
776	99.	Krumina, L., Op De Beeck, M., Meklesh, V., Tunlid, A. & Persson, P. Ectomycorrhizal Fungal
777		Transformation of Dissolved Organic Matter: Consequences for Reductive Iron Oxide Dissolution
778		and Fenton-Based Oxidation of Mineral-Associated Organic Matter. Front. Earth Sci. 10, (2022).
779	100.	Liu, F. et al. Production of reactive oxygen species and its role in mediating the abiotic
780		transformation of organic carbon in sandy soil under vegetation restoration. Carbon Res. 2, 35
781		(2023).
782	101.	Verbruggen, E., Struyf, E. & Vicca, S. Can arbuscular mycorrhizal fungi speed up carbon
783		sequestration by enhanced weathering? PLANTS PEOPLE PLANET 3, 445-453 (2021).
784	102.	Wu, S. et al. Soil organic matter dynamics mediated by arbuscular mycorrhizal fungi – an updated
785		conceptual framework. New Phytol. 242, 1417-1425 (2024).
786	103.	Fernandez, C. W., Langley, J. A., Chapman, S., McCormack, M. L. & Koide, R. T. The
787		decomposition of ectomycorrhizal fungal necromass. Soil Biol. Biochem. 93, 38-49 (2016).
788	104.	Andrino, A. et al. Production of Organic Acids by Arbuscular Mycorrhizal Fungi and Their
789		Contribution in the Mobilization of Phosphorus Bound to Iron Oxides. Front. Plant Sci. 12, (2021).
790	105.	King, A. E. et al. A soil matrix capacity index to predict mineral-associated but not particulate
791		organic carbon across a range of climate and soil pH. <i>Biogeochemistry</i> 165, 1–14 (2023).
792	106.	Kramer, M. G. & Chadwick, O. A. Climate-driven thresholds in reactive mineral retention of soil
793		carbon at the global scale. Nat. Clim. Change 8, 1104–1108 (2018).
794	107.	Possinger, A. R. et al. Climate Effects on Subsoil Carbon Loss Mediated by Soil Chemistry.
795		Environ. Sci. Technol. 55, 16224–16235 (2021).
796	108.	Rasmussen, C. et al. Beyond clay: towards an improved set of variables for predicting soil organic
797		matter content. Biogeochemistry 137, 297–306 (2018).
798	109.	Adhikari, D. et al. Dynamics of ferrihydrite-bound organic carbon during microbial Fe reduction.
799		Geochim. Cosmochim. Acta 212, 221–233 (2017).

- 800 110. Huang, W. & Hall, S. J. Elevated moisture stimulates carbon loss from mineral soils by releasing
 801 protected organic matter. *Nat. Commun.* 8, 1774 (2017).
- 802 111. Patzner, M. S. *et al.* Iron mineral dissolution releases iron and associated organic carbon during
 803 permafrost thaw. *Nat. Commun.* 11, 6329 (2020).
- 804 112. Zhao, Q. *et al.* Coupled dynamics of iron and iron-bound organic carbon in forest soils during
 805 anaerobic reduction. *Chem. Geol.* 464, 118–126 (2017).
- 806 113. Zhao, Q. *et al.* Oxidation of soil organic carbon during an anoxic-oxic transition. *Geoderma* 377,
 807 114584 (2020).
- 808 114. Lieberman, H. P., Rothman, M., von Sperber, C. & Kallenbach, C. M. Experimental flooding shifts
- 809 carbon, nitrogen, and phosphorus pool distribution and microbial activity. *Biogeochemistry* 165, 75–
 810 90 (2023).
- 811 115. Jilling, A., Sokol, N. W., Morán-Rivera, K. & Stuart Grandy, A. Wet-dry cycling influences the
- 812 formation of mineral-associated organic matter and its sensitivity to simulated root exudates.
- 813 *Geoderma* 445, 116869 (2024).
- 814 116. von Lützow, M. & Kögel-Knabner, I. Temperature sensitivity of soil organic matter
- 815 decomposition—what do we know? *Biol. Fertil. Soils* **46**, 1–15 (2009).
- 816 117. Georgiou, K. *et al.* Emergent temperature sensitivity of soil organic carbon driven by mineral
 817 associations. *Nat. Geosci.* 17, 205–212 (2024).
- 818 118. Rocci, K. S., Lavallee, J. M., Stewart, C. E. & Cotrufo, M. F. Soil organic carbon response to global
 819 environmental change depends on its distribution between mineral-associated and particulate
- 820 organic matter: A meta-analysis. *Sci. Total Environ.* **793**, 148569 (2021).
- 821 119. He, N., Chen, Q., Han, X., Yu, G. & Li, L. Warming and increased precipitation individually
- 822 influence soil carbon sequestration of Inner Mongolian grasslands, China. *Agric. Ecosyst. Environ.*
- **823 158**, 184–191 (2012).

- Phillips, C. L., Murphey, V., Lajtha, K. & Gregg, J. W. Asymmetric and symmetric warming
 increases turnover of litter and unprotected soil C in grassland mesocosms. *Biogeochemistry* 128,
 217–231 (2016).
- 827 121. Luo, Y. *et al.* Progressive Nitrogen Limitation of Ecosystem Responses to Rising Atmospheric
 828 Carbon Dioxide. *BioScience* 54, 731–739 (2004).
- 829 122. Rocci, K. S. *et al.* Bridging 20 Years of Soil Organic Matter Frameworks: Empirical Support,
 830 Model Representation, and Next Steps. *J. Geophys. Res. Biogeosciences* 129, e2023JG007964
 831 (2024).
- 832 123. Sulman, B. N., Phillips, R. P., Oishi, a. C., Shevliakova, E. & Pacala, S. W. Microbe-driven
- 833 turnover offsets mineral-mediated storage of soil carbon under elevated CO2. *Nat. Clim. Change* 4,
 834 1099–1102 (2014).
- 835 124. Cotrufo, M. F., Wallenstein, M. D., Boot, C. M., Denef, K. & Paul, E. The Microbial Efficiency836 Matrix Stabilization (MEMS) framework integrates plant litter decomposition with soil organic
 837 matter stabilization: do labile plant inputs form stable soil organic matter? *Glob. Change Biol.* 19,
- 838 988–995 (2013).
- 839 125. Cates, A. M., Jilling, A., Tfaily, M. M. & Jackson, R. D. Temperature and moisture alter organic
 840 matter composition across soil fractions. *Geoderma* 409, 115628 (2022).
- 841 126. Leitner, Z. R., Daigh, A. L. M. & DeJong-Hughes, J. Temporal fluctuations of microbial
 842 communities within the crop growing season. *Geoderma* 391, 114951 (2021).
- 843 127. Fu, Z., Hu, W., Beare, M. & Baird, D. Soil macroaggregate-occluded mineral-associated organic
- carbon drives the response of soil organic carbon to land use change. *Soil Tillage Res.* 244, 106271
 (2024).
- 846 128. Connell, R. K., James, T. Y. & Blesh, J. A legume-grass cover crop builds mineral-associated
- organic matter across variable agricultural soils. *Soil Biol. Biochem.* **203**, 109726 (2025).

- 848 129. Mayer, M. *et al.* Dynamic stability of mineral-associated organic matter: enhanced stability and
 849 turnover through organic fertilization in a temperate agricultural topsoil. *Soil Biol. Biochem.* 184,
 850 109095 (2023).
- B51 130. Daly, A. B. *et al.* A holistic framework integrating plant-microbe-mineral regulation of soil
 bioavailable nitrogen. *Biogeochemistry* 154, 211–229 (2021).
- 853 131. Rui, Y. *et al.* Persistent soil carbon enhanced in Mollisols by well-managed grasslands but not
- annual grain or dairy forage cropping systems. *Proc. Natl. Acad. Sci.* **119**, e2118931119 (2022).
- 855 132. Kauer, K., Pärnpuu, S., Talgre, L., Eremeev, V. & Luik, A. Soil Particulate and Mineral-Associated
- 856 Organic Matter Increases in Organic Farming under Cover Cropping and Manure Addition.
- 857 *Agriculture* 11, 903 (2021).
- 858 133. van der Pol, L. K. et al. Addressing the soil carbon dilemma: Legumes in intensified rotations
- 859 regenerate soil carbon while maintaining yields in semi-arid dryland wheat farms. *Agric. Ecosyst.*860 *Environ.* 330, 107906 (2022).
- 861 134. Osterholz, W. R. *et al.* Predicting Gross Nitrogen Mineralization and Potentially Mineralizable
 862 Nitrogen using Soil Organic Matter Properties. *Soil Sci. Soc. Am. J.* 81, 1115–1126 (2017).
- 863 135. Spackman, J. A., Fernández, F. G., Paiao, G. D., Venterea, R. T. & Coulter, J. A. Corn 15N uptake
 864 and partitioning in response to fertilizer application rate and timing. *Agron. J.* 116, 1991–2006
 865 (2024).
- 866 136. White, C. M., Finney, D. M., Kemanian, A. R. & Kaye, J. P. Modeling the contributions of nitrogen
 867 mineralization to yield of corn. *Agron. J.* 113, 490–503 (2021).
- 868 137. Zhang, Z., Kaye, J. P., Bradley, B. A., Amsili, J. P. & Suseela, V. Cover crop functional types
- differentially alter the content and composition of soil organic carbon in particulate and mineralassociated fractions. *Glob. Change Biol.* 28, 5831–5848 (2022).
- 871 138. Teixeira, R. da S. *et al.* Land-use change with pasture and short rotation eucalypts impacts the soil C
- emissions and organic C stocks in the Cerrado biome. *Land Degrad. Dev.* **31**, 909–923 (2020).

- 873 139. Kaiser, K. & Kalbitz, K. Cycling downwards dissolved organic matter in soils. *Soil Biol. Biochem.*874 52, 29–32 (2012).
- 875 140. Roth, V.-N. *et al.* Persistence of dissolved organic matter explained by molecular changes during its
 876 passage through soil. *Nat. Geosci.* 12, 755–761 (2019).
- 877 141. Cagnarini, C. *et al.* Zones of influence for soil organic matter dynamics: A conceptual framework
 878 for data and models. *Glob. Change Biol.* 25, 3996–4007 (2019).
- 879 142. Todd-Brown, K. E. O. *et al.* Causes of variation in soil carbon simulations from CMIP5 Earth
 880 system models and comparison with observations. *Biogeosciences* 10, 1717–1736 (2013).
- 881 143. Sulman, B. N. *et al.* Multiple models and experiments underscore large uncertainty in soil carbon
 882 dynamics. *Biogeochemistry* 141, 109–123 (2018).
- 144. Todd-Brown, K. E. O., Hopkins, F. M., Kivlin, S. N., Talbot, J. M. & Allison, S. D. A framework
 for representing microbial decomposition in coupled climate models. *Biogeochemistry* 109, 19–33
 (2012).
- 886 145. Treseder, K. K. *et al.* Integrating microbial ecology into ecosystem models: challenges and
 887 priorities. *Biogeochemistry* 109, 7–18 (2012).
- 888 146. Wieder, W. R., Grandy, A. S., Kallenbach, C. M. & Bonan, G. B. Integrating microbial physiology
 889 and physio-chemical principles in soils with the MIcrobial-MIneral Carbon Stabilization (MIMICS)
 890 model. *Biogeosciences* 11, 3899–3917 (2014).
- 891 147. Ahrens, B., Braakhekke, M. C., Guggenberger, G., Schrumpf, M. & Reichstein, M. Contribution of
 892 sorption, DOC transport and microbial interactions to the 14C age of a soil organic carbon profile:
- Insights from a calibrated process model. *Soil Biol. Biochem.* **88**, 390–402 (2015).
- 148. Tang, J. & Riley, W. J. Weaker soil carbon–climate feedbacks resulting from microbial and abiotic
 interactions. *Nat. Clim. Change* 5, 56–60 (2015).
- 896 149. Wieder, W. R., Grandy, A. S., Kallenbach, C. M., Taylor, P. G. & Bonan, G. B. Representing life in
- the Earth system with soil microbial functional traits in the MIMICS model. *Geosci. Model Dev.* 8,
- 898 1789–1808 (2015).

- 899 150. Abramoff, R. et al. The Millennial model: in search of measurable pools and transformations for 900 modeling soil carbon in the new century. Biogeochemistry 137, 51-71 (2018).
- 901 151. Ahrens, B. et al. Combination of energy limitation and sorption capacity explains 14C depth 902 gradients. Soil Biol. Biochem. 148, 107912 (2020).
- 903 152. Robertson, A. D. et al. Unifying soil organic matter formation and persistence frameworks: the 904 MEMS model. *Biogeosciences* 16, 1225–1248 (2019).
- 905 153. Wang, G., Post, W. M. & Mayes, M. A. Development of microbial-enzyme-mediated 906 decomposition model parameters through steady-state and dynamic analyses. Ecol. Appl. 23, 255– 907
- 272 (2013).
- 908 154. Wieder, W. R. et al. Simulating Global Terrestrial Carbon and Nitrogen Biogeochemical Cycles
- 909 With Implicit and Explicit Representations of Soil Microbial Activity. J. Adv. Model. Earth Syst. 910 16, e2023MS004156 (2024).
- 911 155. Zhang, Y. et al. Simulating measurable ecosystem carbon and nitrogen dynamics with the 912 mechanistically defined MEMS 2.0 model. *Biogeosciences* 18, 3147–3171 (2021).
- 913 156. Agren, G. I. & Bosatta, N. Theoretical Analysis of the Long-Term Dynamics of Carbon and
- 914 Nitrogen in Soils. Ecology 68, 1181–1189 (1987).
- 915 157. Waring, B. G. et al. From pools to flow: The PROMISE framework for new insights on soil carbon 916 cycling in a changing world. Glob. Change Biol. (2020) doi:10.1111/gcb.15365.
- 917 158. Woolf, D. & Lehmann, J. Microbial models with minimal mineral protection can explain long-term 918 soil organic carbon persistence. Sci. Rep. 9, 6522 (2019).
- 919 159. Plaza, C., Giannetta, B., Benavente, I., Vischetti, C. & Zaccone, C. Density-based fractionation of
- 920 soil organic matter: effects of heavy liquid and heavy fraction washing. Sci. Rep. 9, 10146 (2019).
- 921 160. Heckman, K., Lawrence, C. R. & Harden, J. W. A sequential selective dissolution method to
- 922 quantify storage and stability of organic carbon associated with Al and Fe hydroxide phases.
- 923 Geoderma 312, 24–35 (2018).

- 924 161. Li, H. *et al.* Simple Plant and Microbial Exudates Destabilize Mineral-Associated Organic Matter
 925 via Multiple Pathways. *Environ. Sci. Technol.* 55, 3389–3398 (2021).
- 926 162. Schiedung, M., Barré, P. & Peoplau, C. Separating fast from slow cycling soil organic carbon A
 927 multi-method comparison on land use change sites. *Geoderma* 453, 117154 (2025).
- 928 163. Jilling, A., Keiluweit, M., Gutknecht, J. L. M. & Grandy, A. S. Priming mechanisms providing
- 929 plants and microbes access to mineral-associated organic matter. *Soil Biol. Biochem.* 158, 108265
 930 (2021).
- 931 164. Paul, E. A., Morris, S. J. & Böhm, S. The determination of soil C pool sizes and turnover rates:
- biophysical fractionation and tracers. in *Assessment Methods for Soil Carbon* 193–206 (CRC Press,
 2000).
- 934 165. Paul, E. A., Morris, S. J., Conant, R. & Plante, A. F. Does the Acid Hydrolysis–Incubation Method
 935 Measure Meaningful Soil Organic Carbon Pools? *Soil Sci. Soc. Am. J.* 70, 1023–1035 (2006).
- 166. Konrad, A. *et al.* Microbial carbon use efficiency of mineral-associated organic matter is related to
 its desorbability. *Soil Biol. Biochem.* 203, 109740 (2025).
- 938 167. Jagadamma, S., Steinweg, J. M., Mayes, M., Wang, G. & Post, W. Decomposition of added and
- native organic carbon from physically separated fractions of diverse soils. *Biol. Fertil. Soils* 1–9
- 940 (2013) doi:10.1007/s00374-013-0879-2.
- 941 168. Jones, D. L. & Edwards, A. C. Influence of sorption on the biological utilization of two simple
 942 carbon substrates. *Soil Biol. Biochem.* 30, 1895–1902 (1998).
- 943 169. McGhee, I., Sannino, F., Gianfreda, L. & Burns, R. G. Bioavailability of 2,4-D sorbed to a chlorite944 like complex. *Chemosphere* 39, 285–291 (1999).
- 945 170. Mikutta, R. *et al.* Biodegradation of forest floor organic matter bound to minerals via different
 946 binding mechanisms. *Geochim. Cosmochim. Acta* 71, 2569–2590 (2007).
- 947 171. Saidy, A. R., Smernik, R. J., Baldock, J. A., Kaiser, K. & Sanderman, J. Microbial degradation of
- 948 organic carbon sorbed to phyllosilicate clays with and without hydrous iron oxide coating. *Eur. J.*
- 949 Soil Sci. 66, 83–94 (2015).

- 950 172. Singh, N. *et al.* Bioavailability of an Organophosphorus Pesticide, Fenamiphos, Sorbed on an
 951 Organo Clay. *J. Agric. Food Chem.* 51, 2653–2658 (2003).
- 952 173. Gu, B., Schmitt, J., Chen, Z., Llang, L. & Mccarthyt, J. F. Adsorption and Desorption of Natural
 953 Organic Matter on Iron Oxide: Mechanisms and Models. *Env. Sci Technol* 28, 38–48 (1994).
- 954 174. Gu, B., Schmitt, J., Chen, Z., Liang, L. & McCarthy, J. F. Adsorption and desorption of different
- 955 organic matter fractions on iron oxide. *Geochim. Cosmochim. Acta* **59**, 219–229 (1995).
- 956 175. Joo, J. C., Shackelford, C. D. & Reardon, K. F. Association of humic acid with metal (hydr)oxide957 coated sands at solid–water interfaces. *J. Colloid Interface Sci.* 317, 424–433 (2008).
- 958 176. Kahle, M., Kleber, M. & Jahn, R. Retention of dissolved organic matter by phyllosilicate and soil

959 clay fractions in relation to mineral properties. Org. Geochem. 35, 269–276 (2004).

- 960 177. Kaiser, K. & Zech, W. Release of Natural Organic Matter Sorbed to Oxides and a Subsoil. *Soil Sci.*961 *Soc. Am. J.* 63, 1157–1166 (1999).
- 962 178. Konrad, A. *et al.* Microbial carbon use efficiency of mineral-associated organic matter is related to
 963 its desorbability. *Soil Biol. Biochem.* 203, 109740 (2025).
- 964 179. Navon, R., Hernandez-Ruiz, S., Chorover, J. & Chefetz, B. Interactions of Carbamazepine in Soil:
 965 Effects of Dissolved Organic Matter. *J. Environ. Qual.* 40, 942–948 (2011).
- 966 180. Saidy, A. R., Smernik, R. J., Baldock, J. A., Kaiser, K. & Sanderman, J. The sorption of organic
- 967 carbon onto differing clay minerals in the presence and absence of hydrous iron oxide. *Geoderma*968 209–210, 15–21 (2013).
- 969 181. Singh, M., Sarkar, B., Biswas, B., Churchman, J. & Bolan, N. S. Adsorption-desorption behavior of
 970 dissolved organic carbon by soil clay fractions of varying mineralogy. *Geoderma* 280, 47–56
- 971 (2016).
- 972 182. Singh, M. *et al.* Influence of physico-chemical properties of soil clay fractions on the retention of
 973 dissolved organic carbon. *Environ. Geochem. Health* 39, 1335–1350 (2017).

974	183. Yeasmin, S. et al. Influence of mineral characteristics on the retention of low molecular weight
975	organic compounds: A batch sorption-desorption and ATR-FTIR study. J. Colloid Interface Sci.
976	432 , 246–257 (2014).
977 978	
979	Figure Captions
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981	Figure 1. Conceptual figure illustrating the controls on MAOM chemical heterogeneity and three
982	known drivers of MAOM destabilization
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987 Table 1. Results of mineralization experiments that provide evidence for the fast-cycling MAOM pool. Such studies measure respiration of

988 isotope-labeled compounds adsorbed to a pure clay or MAOM fraction. The method used in each paper is described alongside the clay or soil type

989 studied or employed in each experiment, and the primary results of said experiment.

Reference	Method	Clay or soil type	Substrate mineralization / bioavailability
166	¹⁴ C-labeled organic monomers (e.g., citric acid) adsorbed to pure minerals, mixed with soil, and incubated for three weeks; ¹⁴ C traced into microbial biomass. ¹⁴ C- CO ₂ evolution was monitored during the first three weeks and for an additional three weeks after priming with glucose.	Kaolinite, illite, and goethite in loamy vs sandy arable topsoils	Substrate mineralization up to $> 80\%$ (citric acid on kaolinite and illite) and incorporation into microbial biomass up to ~65% (acetylglucosamine on kaolinite) over three weeks of incubation; subsequent glucose-induced priming, especially of organic acids, was highest in goethite
167	Particulate (POM) and mineral-associated (MAOM) fractions incubated with unlabeled or ¹⁴ C-labelled glucose for 150 days to measure total C and ¹⁴ C respiration.	Soils from five orders (Mollisol, Ultisol, Oxisol, Andisol, Gelisol) over four climate zones (temperate, tropics, sub- arctic, and arctic)	62–70% of glucose added to MAOM and 3–11% of native C from MAOM were respired over 150 days.
168	¹⁴ C-labeled citrate or glucose were equilibrated with minerals then incubated for 24 h in oxygenated suspension with or without bacteria cultured from grassland soil to quantify ¹⁴ C recovery in respired CO ₂ , in the solid phase (bacteria and minerals), and in supernatant	Synthetic illite-mica; pure kaolinite; synthetic ferric hydroxide (ferrihydrite); and a mixed clay subsoil containing kaolinite, illite/smectite, and goethite	40-60% of glucose C in all mineral treatments and 65% of citrate C sorbed to illite-mica were microbially respired within 6 h, but only 25%, 12%, and 1% of citrate C were respectively respired from kaolinite, mixed subsoil clay, and ferric hydroxide treatments
169	A ¹⁴ C-labeled organic compound (herbicide 2,4-D [dichloro-phenoxyacetic acid]) was sorbed to a mineral complex and incubated with cultured <i>Pseudomonas</i> sp.; ¹⁴ C in respired CO ₂ , solid, and liquid fractions measured after 28 days	An Al(OH)x-montmorillonite (chlorite- like) complex	Over 28 days, the bacteria desorbed or mineralized 80% of the sorbed 2,4-D, compared to 45% released by abiotic desorption alone

Reference	Method	Clay or soil type	Substrate mineralization / bioavailability
170	Organic matter (OM) extracted from forest floor O-horizon was sorbed onto minerals in the presence of different electrolytes to model the relative contributions of Ca^{2+} bridging and ligand exchange to organo-mineral binding mechanisms in solution, and aerobically incubated with O-horizon microbial inocula for 90 d to measure substrate respiration as CO2	Synthetic goethite (iron oxide); and purchased pyrophyllite and vermiculite	24-77% of OM was bioavailable after sorption to the minerals tested, and OM mineralization increased by 42-62%. Ca ²⁺ bridging slightly increased OM bioavailability in goethite, but decreased OM bioavailability by 40-55% in pyrophylite and vermiculite; ligand exchange reduced OM bioavailability by 12%, 46%, and 66% respectively in vermiculite, pyrophylite, and goethite
171	Dissolved organic carbon (DOC) from plant residue loaded onto minerals was inoculated with soil extracts and incubated for 120 d to assess OC mineralization as CO ₂	Kaolinite, illite, and smectite, with and without coatings of goethite, hematite, and ferrihydrite	13–24% of sorbed OC was mineralized over 120 days; mineral associations reduced OC bioavailability by 27-43%
172	An organo-clay complex of fenamiphos, an organophosphorus pesticide, was incubated in aqueous suspension in abiotic conditions or with added <i>Brevibacterium</i> sp., and pesticide desorption and hydrolysis was monitored by HPLC over 24 h	Cetyltrimethylammonium (CTMA)- exchanged montmorillonite clay	Bacteria hydrolyzed fenamiphos at a rate 77% greater than the abiotic desorption rate over 24 h, indicating high bioavailability despite strong interlayer sorption; 82% of the pesticide was hydrolyzed in inoculated samples versus 3% and 4.6% respectively degraded and desorbed in abiotic controls Added enzymes penetrated the mineral interlayer and retained hydrolytic activity even when sorbed

995 Table 2. Results of batch (ad)sorption-desorption experiments that highlight the wide-ranging rate of OM desorption from minerals. The mineral 996 type employed in each study is presented alongside the organic compounds that were sorbed/desorbed to said minerals, the experimental 997 conditions, and the primary results of each study. For studies that report a hysteresis coefficient: this metric refers to the ratio of the slope of the 998 sorption curve to the slope of the desorption curve. It is a measure of adsorption-desorption reversibility where an h of 0 indicates completely 999 reversible and an h of 1 indicates completely irreversible.

Reference	Mineral type	Organic compounds	Experimental conditions	Results
173	Commercial iron oxide powder	Natural organic matter (NOM), collected from wetland pond	Acid (HCl), base (NaOH), or inorganic salts (NaCl, Na ₂ SO ₄ , Na ₃ PO ₄) added before adsorption	Hysteresis coefficient (h) between 0.72 and 0.86 for pH 4.1 and between 0.78 and 0.92 for pH 6.0, indicating that desorption was very limited
174	Iron oxide powder	NOM from wetland pond, separated into hydrophobic and hydrophilic fractions	Acid (HCl), base (NaOH), or inorganic salts (NaCl, Na ₂ SO ₄ , Na ₃ PO ₄) added before adsorption	Hysteresis coefficient (h) between 0.81 and 0.91 for hydrophobic NOM, h between 0.821 and 0.984 for hydrophilic NOM, indicating desorption was very limited
175	Uncoated, FeO(OH)- coated, and Al ₂ O ₃ -coated sands	Terrestrial humic acid extracted from commercially available peat	Uncoated vs (hydr)oxide coated sands	At pH 4.1, hysteresis coefficient (h) between 0.70 and 0.96 for metal (hydr)oxide coated sands, indicating limited desorption. For uncoated sands, h was between 0.41 and 0.78 suggesting higher desorption potential
176	Soils (Vitric Phaeozem, Haplic Chernozem, Chromic Luvisol, Calcaric Phaeozem, and Chromic Cambisol) and phyllosilicate clay fractions (Ca- montmorillonite, kaolinite, and illite)	Dissolved organic carbon (DOC) extracted from pine forest floor	Near-neutral pH; phosphate treatment used to block reactive hydroxyl groups	In samples without phosphate treatment, between 13% and 50% of DOC was desorbed, with greater desorption in the phyllosilicate clay compared to the soil clay fractions
177	AmorphousAl(OH) ₃ , goethite, and low organic C subsoil	NOM, extracted from Oa horizon of an Entic Haplorthod	Desorption using solutions with a range of pH and inorganic anions (Cl ⁻ , $SO_4^{2^-}$, and $H_2PO_4^{-}$)	Under normal conditions, desorption of NOM was $<3\%$, desorption reached 60% in the presence of high concentrations of H ₂ PO ₄ ⁻
178	Kaolinite, illite, and geothite	14C-labelled monomers (glucose, acetylglucosamine,	Also measured microbial carbon use efficiency (CUE)	40-99% of monomers across all treatments were retained after desorption with NaN ₃ , the

Reference	Mineral type	Organic compounds	Experimental conditions	Results
		phenylalanine, salicylic acid, and citric acid)		range was reduced to $3-55\%$ after subsequent desorption using PO_4^{3-}
179	Vertisol soil from crop field (bulk soil and its clay size fraction)	Carbamazepine (CBZ)	No CBZ, CBZ co- introduced, or CBZ introduced after DOM pre-adsorption	The hysteresis coefficient (HI) for CBZ-clay was 0.91 versus 0.64 in CBZ-bulk soil, HI decreased following pre-adsorption of DOM
37	Organic matter-poor, alkaline soils (Fluvent, Rhodoxeralf, and Loess)	DOM extracted from mature composted biosolids	Four sequential desorption steps	Up to 83% of sorbed DOM was retained after desorption
180	Pure clays (kaolinite, illite, smectite) with and without Fe oxide (haematite, goethite, ferrihydrite) coatings	DOC extracted from medic shoot	Effect of goethite coating on different clay types, effects of different Fe oxide coatings on illite	Across all treatments, 5.7–14.4% of sorbed DOC was desorbed
181	Soil clay fractions (kaolinite-illite, smectite, and allophane)	DOC extracted from wheat straw	Adsorption measured under varying electrolyte conditions: 0.1M and 0.01M Ca(NO ₃) ₂ and NaNO ₃	6.4% to 55.3% of adsorbed DOC was desorbed
182	Soil clay fractions (kaolinite–illite, smectite, allophane)	DOC extracted from wheat straw	Untreated clay, C removed, and sesquioxide removed	30% to 71% of adsorbed DOC was desorbed across clay type; highest desorption in kaolinite-illite
183	Pure minerals (kaolinite, illite, montmorillonite, ferrihydrite, and goethite)	14C-labelled carboxylic acids and amino acids		For carboxylic acids: Fe oxides retained 83– 100%, while phyllosilicates retained 31– 85%. For amino acids: glutamic acid retention was 53–98% on Fe oxides versus 0–48% on phyllosilicates; lysine retention was 41–99% on phyllosilicates and 13–50% on Fe oxides



Controls on MAOM chemical heterogeneity

Ecological drivers of destabilization

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1001 Figure 1. Conceptual figure illustrating the controls on MAOM chemical heterogeneity and three known drivers of MAOM

1002 destabilization