

1 The impact of the rhizocephalan parasitic barnacle on its host crab through
2 non-consumptive effects

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16

17 **Abstract**

18 Parasites can significantly influence ecological communities by altering host traits.
19 Rhizocephalans, a group closely related to thoracican barnacles within Cirripedia, are
20 common in marine ecosystems and profoundly impact their hosts, most notably by
21 inducing reproductive castration. However, their influence on other host traits,
22 particularly those related to predator defense, remains underexplored. The
23 rhizocephalan *Sacculina* sp. modifies the length of the cheliped propodus in the host
24 crab *Hemigrapsus sanguineus*. To determine whether parasitized crabs are more
25 vulnerable to predation, we conducted field tethering experiments comparing parasitized
26 and unparasitized individuals. The results showed that parasitism itself did not directly
27 increase the likelihood of crabs being lost (presumably due to predation) in either sex.
28 However, structural equation modeling revealed that in parasitized males, a reduced
29 cheliped propodus length indirectly increased predation risk. In females, parasitism
30 reduced carapace width, which in turn shortened the cheliped propodus and increased
31 vulnerability to predation. These findings demonstrate that *Sacculina* infection
32 heightens predation risk through non-consumptive effects by inducing host
33 morphological changes.

34
35 **Key words:** predator-host-parasite interaction, predation of the host crab, sacculinid,
36 morphological change, cheliped propodus length

37
38 **BACKGROUND**

39 Parasites play a key role in shaping marine ecosystems by altering the traits of their
40 hosts (Kuris 1974; Kuris and Lafferty 1992). Rhizocephalans (Thecostraca: Cirripedia)
41 parasitize a variety of crustaceans, including hermit crabs (Yoshida et al. 2014; Jung et
42 al. 2019; Kajimoto et al. 2022), crabs (Lützen et al. 2018; Toyota et al. 2023), and
43 shrimps (Lützen et al. 2016). These parasites are particularly notable for causing
44 parasitic castration, rendering hosts reproductively inactive (Hines et al. 1997).
45 Rhizocephalans are highly adapted parasites, with adult females developing root-like
46 internal structures (internae) for nutrient absorption and external reproductive structures
47 (externae). Males are drastically reduced in size and live inside females (Høeg 1995a, b;

48 Høeg and Lützen 1995; Høeg et al. 2019, 2020).

49 Several species, particularly in the family Sacculinidae, induce feminization in male
50 hosts, altering secondary sexual traits such as abdominal morphology, chela size, and
51 copulatory appendages (Alvarez and Calderon 1996; Kristensen et al. 2012; Waiho et al.
52 2017; Toyota et al. 2023). A well-documented example is the transformation of a male's
53 narrow, semicircular abdomen into a broader, more female-like shape, especially in
54 brachyuran crabs. This adaptation allows parasitized males to accommodate larger
55 externae, potentially increasing parasite reproductive success (Rees and Glenner 2014;
56 Nagler et al. 2017; Mouritsen et al. 2018). Other morphological changes include
57 reduced chela size, altered copulatory structures (Hartnoll 1962; Kristensen et al. 2012;
58 Toyota et al. 2023), and changes in pleopod number (Kristensen et al. 2012).

59 Although such parasite-induced morphological changes have been well
60 documented, their implications for host survival are not well understood. In particular,
61 no studies have investigated how changes in chela size—a key component of anti-
62 predator defense—might affect host vulnerability to predation. This study aims to
63 address this gap by examining whether parasitism by *Sacculina* sp. increases the
64 susceptibility of *Hemigrapsus sanguineus* to predation through a reduction in cheliped
65 propodus length. Field experiments were conducted along the coast of Osaka, Japan.
66
67

68 MATERIALS AND METHODS

69 **Ethical statement**

70 No specific permissions were required for this study, as it did not involve
71 endangered or protected invertebrate species in Japan. Vertebrates, humans, or human-
72 derived tissues were not used. All animal experiments were conducted in compliance
73 with the ARRIVE guidelines (Percie du Sert et al. 2020).
74

75 **Sample collection and rearing of host crabs**

76 Between July and November 2021 and April and August 2022, 10 to 30 individuals
77 of the Asian shore crab *Hemigrapsus sanguineus* were collected by hand at least once
78 per month from the coastline of Shin Town, Hannan City, Osaka, Japan (34.213°N,

79 135.135°E).

80 In the laboratory, each crab was housed individually in a round plastic container
81 (7.2 cm diameter × 16.7 cm height) filled with 150 mL of artificial seawater (Gex,
82 Osaka, Japan) and maintained at $25 \pm 1^\circ\text{C}$ under fluorescent room lighting. The
83 seawater was replaced every three days, and crabs were fed approximately 30 mg of
84 commercial crayfish feed (“Kyozaizarigani no esa,” Kyorin, Hyogo, Japan) with each
85 water change.

86

87 **Measurements of morphological traits**

88 The sex of each crab was determined by the presence of female pleopods or male
89 gonopods. We then recorded the presence of *Sacculina* sp. externa on the abdomen.
90 Carapace width (CW) and the length of the right and left cheliped propodus (CPL) were
91 measured using a digital caliper. The average of the right and left CPL (Ave CPL) was
92 calculated for each individual, based on the assumption that both claws are used
93 defensively against predators.

94

95 **Field predation assay**

96 Two crabs with carapace widths between 10 and 35 mm were randomly selected
97 from each of the following groups: parasitized males, unparasitized males, parasitized
98 females, and unparasitized females. The dorsal carapace of each crab was dried with a
99 towel, and a 30-cm monofilament fishing line was affixed using a droplet of quick-
100 drying glue. To accelerate bonding, 1 mL of seawater was applied to the glue. The other
101 end of the line was tied to a ring attached to a small rock (~20 cm diameter), allowing
102 the crab to hide beneath the rock but limiting its range of movement. Eight tethered
103 crabs (two per group) were randomly spaced at least 1 meter apart in the lower intertidal
104 zone. Crab loss (presumed predation) was monitored daily for three consecutive days.

105 To control for non-predatory losses, eight crabs were similarly prepared and placed
106 inside mesh cages (29.5 cm width × 23.3 cm depth × 17.0 cm height; 2.31 mm mesh
107 size). These cages allowed for water flow but prevented predator access. A total of 12
108 replicates were conducted at the same site between September–November 2021 and
109 May–September 2022, yielding 24 individuals per crab category.

110

111 **Statistical analysis**

112 Data from 10 crabs that molted during the experiment (identified by the presence of
113 intact exuviae with no feeding marks; see Fig. 1) were excluded from all analyses. To
114 test whether *Sacculina* sp. infection directly affected the likelihood of crab loss,
115 generalized linear models (GLMs) with a binomial distribution and logit link function
116 were applied separately for males and females. In this model, the presence of *Sacculina*
117 externa was the explanatory variable, and crab fate (intact or lost) was the dependent
118 variable. Analyses were conducted using SAS version 9.4 (SAS Institute Inc., Cary, NC,
119 USA). To examine whether CW or Ave CPL directly influenced crab loss, additional
120 GLMs (binomial, logit link) were performed separately for each sex, with CW or Ave
121 CPL as the explanatory variable.

122 The effect of rhizocephalan infection on relative Ave CPL was analyzed by testing
123 for equality of regression slopes using CW as a covariate. When no significant
124 interactions were detected, an ANCOVA was conducted using CW as a covariate. The
125 analysis included four groups: unparasitized males, unparasitized females, parasitized
126 males, and parasitized females.

127 To test the hypothesis that reduced Ave CPL due to *Sacculina* sp. infection
128 increases crab vulnerability, structural equation modeling (SEM) was performed
129 separately for each sex. SEMs included CW, Ave CPL, and crab fate as observed
130 variables. To improve model fit, paths from CW and infection status to crab fate were
131 excluded. Model fit was assessed using the goodness-of-fit index (GFI), comparative fit
132 index (CFI), and chi-square (χ^2) test. SEMs were constructed using the "lavaan"
133 package (Rosseel, 2012) in R version 4.3.0 (R Core Team, 2023), with the maximum
134 likelihood method used for parameter estimation. Standardized path coefficients
135 (ranging from -1 to 1) were used to represent the strength and direction of relationships
136 among variables, including both direct and indirect effects on crab loss.

137

138

RESULTS

139

140 There was no significant difference in loss rates between unparasitized and
parasitized male crabs (Table 1A), nor between unparasitized and parasitized female

141 crabs (Table 1B). Additionally, no significant difference in loss rates was found between
142 males and females within each CW category (Table 2). However, in male crabs, Ave
143 CPL was found to be associated with crab intactness (Table 2). This trend was not
144 observed in females (Table 2).

145 A significant difference in the slope of the regression of Ave CPL on CW was found
146 between unparasitized males and females (Table 3; Fig. 2). In males infected by
147 *Sacculina* sp., Ave CPL was significantly lower than in unparasitized males, and the
148 slope of the Ave CPL–CW regression was more similar to that of females (Table 3; Fig.
149 2). Additionally, the slope differed significantly between parasitized males and
150 unparasitized females (Table 3; Fig. 2). Among female crabs, no significant differences
151 in Ave CPL were found between parasitized and unparasitized individuals, as both the
152 slope and intercept of the Ave CPL–CW regression overlapped (Table 3; Fig. 2).

153 SEM showed a good fit to the observed data for both sexes (males: GFI = 0.961,
154 CFI = 0.989, $\chi^2/df = 1.63$, $p = 0.195$; females: GFI = 0.980, CFI = 1.000, $\chi^2/df = 0.703$,
155 $p = 0.495$). No crab loss occurred in the control treatment for males (Fig. 3A). In
156 contrast, in the tethering experiment, male crabs with an Ave CPL of 1.90 mm or less
157 experienced loss, regardless of infection status (Fig. 3B). The SEM for males revealed
158 that *Sacculina* infection was negatively associated with CW and Ave CPL, and CW was
159 positively related to Ave CPL (Fig. 3C). Furthermore, Ave CPL was negatively
160 associated with crab loss (Fig. 3C). Similarly, no crab loss was observed in the female
161 control group (Fig. 3D). In the tethering experiment, however, crab loss occurred among
162 individuals with an Ave CPL of 1.5 mm or less, regardless of parasitic infection (Fig.
163 3E). The SEM for females indicated that infection by *Sacculina* sp. negatively affected
164 CW, and CW was positively related to Ave CPL (Fig. 3F). Additionally, Ave CPL was
165 negatively related to crab loss (Fig. 3F).

166

167

DISCUSSION

168 In the field survey, no crab loss was observed in the control treatment, where
169 individuals were enclosed in cages, while loss occurred in the tethering experiment.
170 This indicates that the observed losses were likely due to predation. However, our
171 results provide no evidence that infection by the rhizocephalan parasite *Sacculina* sp.

172 directly increases predation risk in host crabs. In contrast, previous research has shown
173 that parasitized mud crabs *Eurypanopeus depressus* are more susceptible to predation
174 when infected with the rhizocephalan *Loxothylacus panopaei* (Gehman and Byers
175 2017). Gehman and Byers (2017) proposed that behavioral alterations in parasitized
176 crabs, such as slower escape responses, increase their likelihood of being preyed upon,
177 possibly by making them easier or more attractive targets for predators. Although our
178 study did not detect a direct link between *Sacculina* infection and predation, further
179 research is needed to examine potential behavioral changes in infected hosts. For
180 instance, measuring escape speeds of parasitized versus unparasitized crabs could
181 provide insight into subtle behavioral effects of parasitism that were not captured in the
182 present analysis.

183 The SEM analysis indicated that *Sacculina* sp. infection suppressed both CW and
184 Ave CPL in male crabs, consistent with morphological feminization previously reported
185 in this and other studies (Hartnoll 1962; Kristensen et al. 2012; Toyota et al. 2023).
186 SEM results further suggest that the parasite may reduce Ave CPL either directly or
187 indirectly by inhibiting CW growth, given the strong positive correlation between CW
188 and Ave CPL. Moreover, SEM showed that a one-unit increase in Ave CPL was
189 associated with a 0.35 decrease in the probability of crab loss. This indicates that
190 *Sacculina*-induced reductions in Ave CPL heighten vulnerability to predation in males.
191 In female crabs, ANCOVA detected no significant reduction in Ave CPL due to
192 infection. However, SEM revealed a negative effect of *Sacculina* sp. infection on CW
193 and a positive relationship between CW and Ave CPL. No direct effect of infection on
194 Ave CPL was observed in females, possibly due to their inherently smaller chelipeds.
195 Nevertheless, SEM results showed that a one-unit increase in Ave CPL reduced the
196 probability of loss by 0.30. This suggests that the infection-induced reduction in CW led
197 to decreased Ave CPL, indirectly increasing predation risk. Overall, these findings
198 demonstrate that *Sacculina* infection increases host susceptibility to predation through
199 indirect morphological changes—non-consumptive effects in the sense of Peacor et al.
200 (2020). This interpretation aligns with the GLM analysis of Ave CPL, which
201 demonstrated a significant association between Ave CPL and loss of male hosts. Thus,
202 the reduction in CPL caused by rhizocephalan infection may play a particularly

203 important role in increasing predation risk in male crabs.

204 Several studies have suggested that rhizocephalan parasites may increase predation
205 risk in their hosts through behavioral modifications. For example, in *E. depressus*
206 parasitized by *L. panopaei* and in the commercial sand crab *Portunus pelagicus*
207 parasitized by *Sacculina graniferae*, infection inhibits burying behavior and prolongs
208 the time spent on land for grooming of the externa (Bishop and Cannon 1979; Gehman
209 and Byers 2017). In contrast, relatively few studies have examined whether
210 rhizocephalan-induced morphological changes contribute to increased predation risk,
211 although such a possibility has been proposed (Gehman and Byers 2017; Galil and
212 Innocenti 2024). The present study provides the first empirical evidence that
213 morphological changes caused by rhizocephalan infection—specifically, reductions in
214 cheliped size—can weaken the defensive capabilities of the host and thereby increase its
215 vulnerability to predation. However, the current study did not directly observe predator
216 species or the defensive behavior of host crabs in response to predatory threats. To fully
217 assess the ecological impact of rhizocephalan infection on crab populations, it will be
218 essential to identify the main predators in the study area and investigate their
219 interactions with parasitized hosts.

220 Rhizocephalans play an important role in marine food webs by indirectly
221 influencing the dynamics of species associated with their hosts, such as predators and
222 prey, through alterations in host behavior, physiology, and morphology (Mouritsen and
223 Poulin 2002; Toscano et al. 2014; Gehman and Byers 2017). For example, the crab *E.*
224 *depressus* parasitized by *L. panopaei* has been shown to reduce its consumption of the
225 mussel *Brachidontes exustus*, a primary prey item (O’Shaughnessy et al. 2014; Toscano
226 et al. 2014). Toscano et al. (2014) proposed that the parasite’s invasion of internal
227 organs impairs digestive function, prolongs digestion time, and ultimately decreases
228 both foraging effort and prey intake. These feeding reductions in infected crabs may, in
229 turn, benefit conspecifics by increasing mussel abundance, which creates a predation
230 refuge and promotes the growth of unparasitized crab populations. Additionally,
231 Gehman and Byers (2017) reported that the predatory crab *Callinectes sapidus*
232 preferentially preys on *E. depressus* individuals infected by *L. panopaei*. According to
233 Gehman et al. (2017), the frequency of predation is positively correlated with the

234 prevalence of parasitized hosts, possibly due to reduced host mobility caused by
235 infection. In this study, we demonstrated that rhizocephalan infection increases host
236 susceptibility to predation through parasite-induced morphological changes. However,
237 to fully understand the ecological implications of rhizocephalan parasitism, long-term
238 monitoring of predator, prey, and parasite populations at the study site will be essential.
239 Such data would provide a clearer understanding of the complex, cascading interactions
240 mediated by rhizocephalans in marine ecosystems.

241

242

CONCLUSIONS

243 Rhizocephalans are key parasites of decapod crustaceans, known for inducing
244 parasitic castration, suppressing the development of secondary sexual characteristics,
245 feminizing male hosts, and altering host behavior. This study is the first to provide
246 empirical evidence that infection by the rhizocephalan *Sacculina* sp. increases the
247 vulnerability of the Asian shore crab (*Hemigrapsus sanguineus*) to predation by
248 reducing chela size, a critical defensive trait. Although the parasite did not directly
249 increase the likelihood of crab loss in either sex, parasitized males exhibited
250 significantly shorter chelipeds compared to unparasitized males, and smaller cheliped
251 size was associated with a higher risk of predation. In females, *Sacculina* sp. infection
252 negatively affected average cheliped propodus length indirectly through its suppression
253 of carapace width growth, which also contributed to increased predation risk. These
254 findings demonstrate that rhizocephalan parasites can increase host susceptibility to
255 predators through indirect morphological changes, constituting a non-consumptive
256 effect. This study offers new insight into the broader ecological impacts of
257 rhizocephalan infection and highlights their potential role in shaping marine community
258 dynamics through host modification.

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Acknowledgments

262 We thank the members of the Laboratory of Aquatic Ecology at Nara Women's
263 University for their valuable discussions and assistance. This study was supported by
264 the Japan Society for the Promotion of Science (JSPS) under Grant Numbers 19H03284

265 and 24K02100, and by a fellowship from the Japan Science and Technology Agency
266 (JST), Grant Number JPMJFS2127.

267

268 List of abbreviations

269 **ANCOVA**

270 analysis of covariance

271 **Ave CPL**

272 the average value of right and left cheliped propodus length

273 **CFI**

274 comparative fit index

275 **CPL**

276 cheliped propodus length

277 **CW**

278 carapace width

279 **GFI**

280 goodness-of-fit index

281 **SEM**

282 structural equation modeling

283

284 Footnotes

285 **Competing interests:** The authors declare that they have no competing interests.

286

287 **Author contributions:** AK and YY designed the research. AK conducted the sampling,
288 measurements of the morphological parameters, field survey, analysis of the data, and
289 writing the first draft. AK and KT analyzed morphological parameters. KT and YY
290 improved the manuscript.

291

292 **Consent for publication:** Not applicable.

293

294 **Ethics approval consent to participate:** Not applicable.

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385

FIGURE LEGENDS

386

387 Fig. 1. Examples of (A) molted and (B) lost (presumably preyed) host crabs *Hemigrapsus*
388 *sanguineus*.

389

390 Fig. 2. Effect of carapace width on the average cheliped propodus length (Ave CPL) of
391 *Hemigrapsus sanguineus* regarding sex and parasitic status.

392

393 Fig. 3. Fate of unparasitized (black circles) and parasitized (red diamonds) male crabs in
394 the control treatment (A) and tethering experiment (B), plotted against average
395 cheliped propodus length (Ave CPL). A value of 1.0 on the Y-axis indicates that the
396 crab was lost (presumably due to predation), while 0.0 indicates that it remained
397 intact until the end of the experiment. (C) Path diagram from structural equation
398 modeling showing the relationships among *Sacculina* sp. infection, carapace width
399 (CW), Ave CPL, and crab fate (lost or intact). (D), (E), and (F) present the same
400 information for female crabs.

401

TABLES AND FIGURES

402

403

404 Table 1. Fate of male and female *Hemigrapsus sanguineus* regarding parasitic status by

405 *Sacculina* sp. in tethering experiments (generalized linear model).

Host crab type	Intact	Lost	χ^2	df	<i>p</i> -value
A. Male					
Unparasitized	7	15	0.04	1	0.833
Parasitized	8	15			
B. Female					
Unparasitized	8	15	1.75	1	0.186
Parasitized	11	9			

406

407 Table 2. Fate of male and female *Hemigrapsus sanguineus* regarding carapace width
 408 (CW) and the average propodus length of right and left chelipeds (Ave CPL) in
 409 tethering experiments (generalized linear model).

The body parts compared	Intact	Lost	χ^2	df	<i>p</i> -value
Male					
CW	30	15	3.66	1	0.056
Ave CPL			4.73	1	0.030
Female					
CW	24	19	1.37	1	0.242
Ave CPL			3.34	1	0.069

410

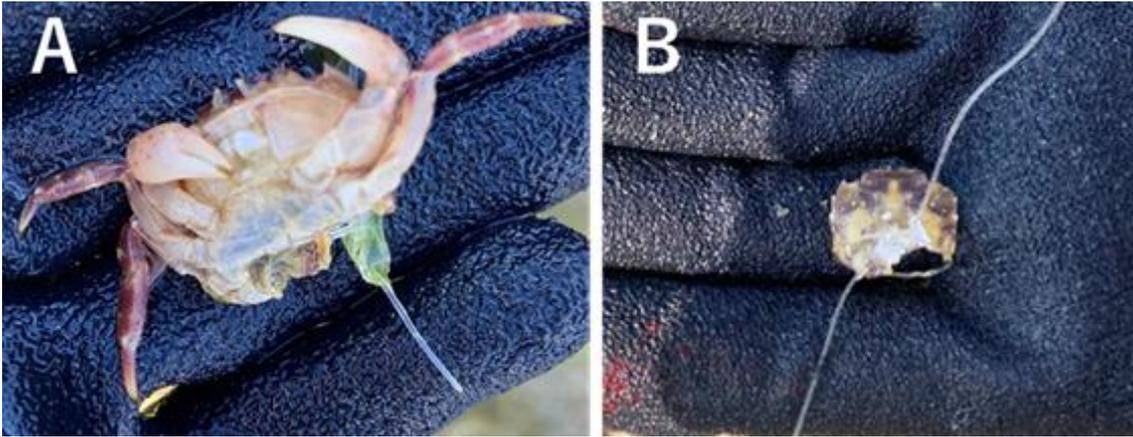
411

412 Table 3. Results on the effect of carapace width on the average cheliped propodus length
 413 (Ave CPL) of *Hemigrapsus sanguineus* concerning sex and parasitic status.

Comparison of the regression lines	Homogeneity of the slopes of the regression lines		Intercept in Ave CPL	
	F value	<i>p</i> -value	F value	<i>p</i> -value
Unparasitized male vs. Unparasitized female	9.66	0.003		
Unparasitized male vs. Parasitized male	0.62	0.435	18.23	< 0.001
Unparasitized female vs. Parasitized male	5.64	0.022		
Unparasitized female vs. Parasitized female	0.02	0.897	0.03	0.860

414

415

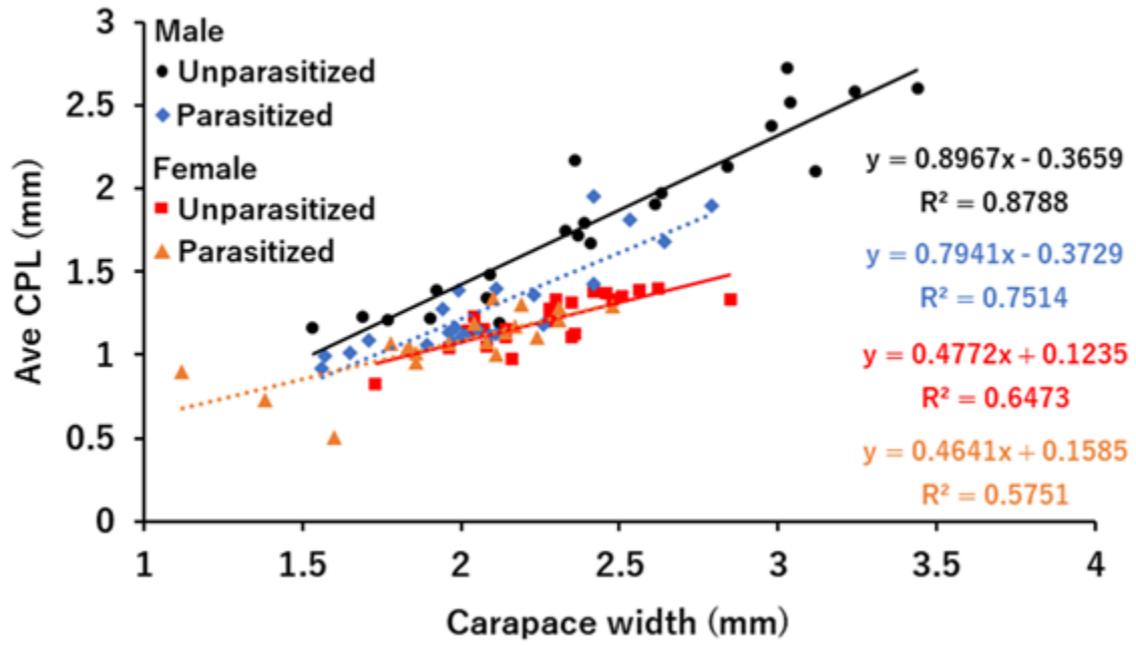


416

417 Fig. 1. Examples of (A) molted and (B) lost (presumably preyed) host crabs

418 *Hemigrapsus sanguineus*.

419

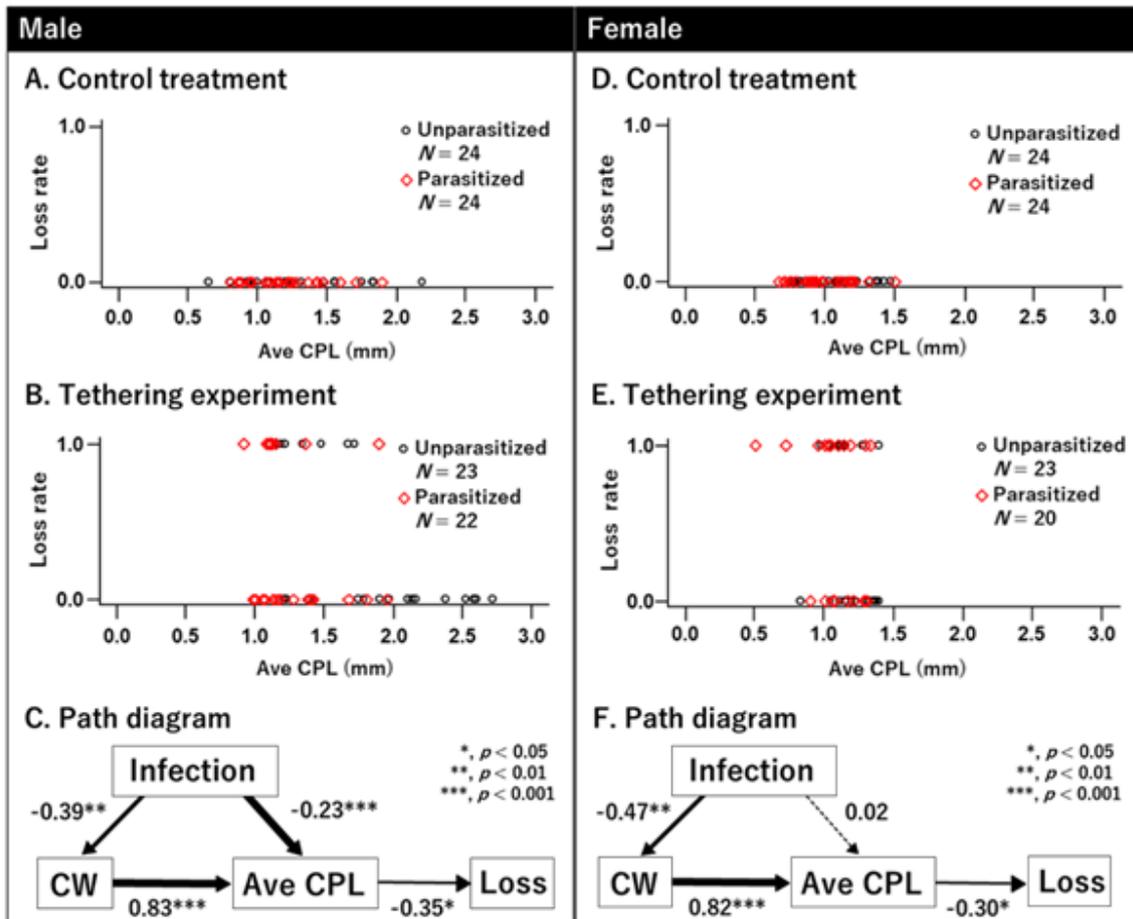


420

421 Fig. 2. Effect of carapace width on the average cheliped propodus length (Ave CPL) of

422 *Hemigrapsus sanguineus* regarding sex and parasitic status.

423



424

425 Fig. 3. Fate of unparasitized (black circles) and parasitized (red diamonds) male crabs in
 426 the control treatment (A) and tethering experiment (B), plotted against average cheliped
 427 propodus length (Ave CPL). A value of 1.0 on the Y-axis indicates that the crab was lost
 428 (presumably due to predation), while 0.0 indicates that it remained intact until the end
 429 of the experiment. (C) Path diagram from structural equation modeling showing the
 430 relationships among *Sacculina* sp. infection, carapace width (CW), Ave CPL, and crab
 431 fate (lost or intact). (D), (E), and (F) present the same information for female crabs.