



28 **ORCID:** NCW (0000-0002-7130-1279), ROA (0000-0002-7220-0621), AB (0000-0003-1093-  
29 677X), SB (0000-0002-4128-5482), MD (0000-0002-5083-624X), SD (0000-0003-1771-0087),  
30 CAN (0000-0002-9859-0568), DSO (0000-0002-1480-1648), MO (0000-0002-5937-6585), SAS  
31 (0000-0002-8656-7168), BJS (0000-0002-7318-6059), SJS (0000-0002-0110-8078), RPB (0000-  
32 0003-4345-6430).

33 **Author contribution:** NCW, RPB conceived the study. SB and NCW developed the figures. All  
34 authors contributed to the development, write up and revisions. Authors are listed in alphabetical  
35 order by last name except for the lead authors NCW and RPB.

36 **Acknowledgement:** We thank Prof. Emily Taylor for encouraging us to write this paper, and the  
37 conference organisers of the 10<sup>th</sup> World Congress of Herpetology in Kuching, Malaysia for providing  
38 the opportunity for us to all gather and conceive this paper in person during the symposium  
39 ‘*Hydroregulation: adaptations, mechanisms, and climate risks*’.

40 **Competing interests:** The authors declare no competing interests.

41 **Funding information:** AB was supported by the Research Fund for International Scientists (RFIS)  
42 from the National Natural Science Foundation of China (NSFC; W2432021), SD was supported by  
43 the Azrieli Graduate Studies Fellowship, CAN and RPB were supported by the State of São Paulo  
44 Science Foundation (FAPESP #18/18900-1 and #19/04637-0, respectively), and BJS was supported  
45 by the National Key Research Development Program of China (#2022YFF0802300), the NSFC  
46 (#32271572), and the Youth Innovation Promotion Association CAS (#Y2023021).

## 47 **ABSTRACT**

48 Human impacts on ecosystems have intensified water variability for terrestrial life, thus challenging  
49 the maintenance of water balance, or hydroregulation. The accelerated development and  
50 accessibility of technologies and computational models over the past decade have enabled  
51 researchers to predict changes in animal hydroregulation and environmental water with greater  
52 spatial and temporal precision. Focusing on reptiles and amphibians, we discuss current methods,  
53 limitations and advances for quantifying ecologically relevant metrics of environmental water  
54 stressors and organismal responses to both acute and long-term water stress that are applicable for  
55 conservation and management. We also highlight approaches that integrate environmental water  
56 data with an organism's water balance and physiological, behavioural and life-history traits to  
57 predict the limits of species' responses and assess their vulnerability to climate change. Finally, we  
58 outline promising future directions and opportunities in hydroregulation studies with a conservation  
59 focus, including broader inferences about acclimation responses, linking gene expression to  
60 functional changes, and exploring inter- and transgenerational plasticity and adaptive evolution.

61 Advances in these fields will facilitate more accurate assessments of species' capacities and the  
62 limits of hydoregulation in response to a more variable and unpredictable future climate.

63 **Keywords:** dehydration, drought, ectotherm, exposure, sensitivity, vulnerability, water balance

## 64 **MAIN TEXT**

65 The colonisation of land presented substantial physiological and morphological challenges  
66 associated with water balance for early terrestrial animals (Gray, 1928; Dial et al., 2015), yet  
67 allowed opportunities for novel evolutionary strategies that enabled an explosion of animal diversity  
68 on land (Minter et al., 2017). Nevertheless, environmental dryness continues to pose a major  
69 challenge for land animals, with biodiversity being highest in humid tropical rainforests and lowest  
70 in desert environments (Owen, 1989; Biber et al., 2023; Coelho et al., 2023). Global trends towards  
71 aridification can therefore compromise biodiversity, a major conservation concern given accelerated  
72 changes in climate and land use, leading to unpredictable changes in water variability and  
73 availability (Moustakis et al., 2021; Moss et al., 2024; Zhang et al., 2024). Understanding 1) how  
74 environmental water changes over time and space, 2) how animals respond to water variability, and  
75 3) how they differ in resilience and response capacity is necessary to assess vulnerability, and it is a  
76 first step in managing extinction risk amid the current global biodiversity crisis. Progress towards  
77 these answers has been notable, due to enhanced computational power, novel statistical models, and  
78 more temporal and spatially detailed climate data (Brun et al., 2022; Klinges et al., 2024).  
79 Additionally, our understanding of hydoregulation strategies across biological levels and species  
80 has become more comprehensive (Navas and Carvalho, 2010; Lillywhite, 2017; Rozen-Rechels et  
81 al., 2019; Riddell et al., 2021). A key challenge for the management and conservation of species at  
82 risk due to climate change is to predictively link ecologically relevant water stressors (exposure  
83 risk) with the capacity of animals to maintain water balance (species sensitivity) (**Fig. 1**).

84 This paper reviews current knowledge on terrestrial water availability for assessing  
85 environmental exposure risk, species' short- and long-term responses to water deficits for  
86 evaluating sensitivity, and models predicting vulnerability to environmental drying. We examine  
87 key topics and conclude proposing future directions for refining predictions of species' vulnerability  
88 in a drying world. Our focus on amphibians and reptiles highlights their role in the aquatic-  
89 terrestrial transition and the contrasting hydoregulation strategies that enable them to thrive and  
90 reproduce despite dehydration challenges.

### 91 **1. Environmental exposure risk**

#### 92 *1.1 Water availability on land*

93 Water availability can be quantified and interpreted in many ways, and it is heavily influenced by  
94 the stochastic nature of the water cycle (Chahine, 1992; Oki and Kanae, 2006) and by the local

95 environment (Kinlaw, 1999; Geiger et al., 2003). Broadly, water enters a terrestrial environment  
96 through precipitation, and exits through evapotranspiration and runoff (Oki and Kanae, 2006). The  
97 ratio between precipitation and evapotranspiration can be used to calculate the aridity index (Zomer  
98 et al., 2022), broadly define climate classifications (Beck et al., 2018), and quantify annual site  
99 water balance. In climates with strong seasonal rainfall, precipitation can predict the phenology of  
100 breeding events (Gould et al., 2022; Thompson et al., 2022) and the seasonal primary productivity  
101 of ecosystems (Lieth, 1973). Once in the environment, water can be stored in the ground, in the air,  
102 or accumulated in water bodies (**Table 1**). Water content in the air is typically measured as the  
103 pressure of gaseous water, or water vapor pressure (Gates, 1980) which serves as a basis for  
104 calculating relative humidity (RH), a common meteorological metric. However, RH is often less  
105 relevant for assessing physiological responses in organisms compared to water vapor pressure  
106 (Anderson, 1936; Kurta, 2014; Wu, N. C. et al., 2024b). Finally, water vapor pressure and  
107 temperature are inherently linked (Campbell and Norman, 2000), and incorporating temperature  
108 allows for the calculation of vapor pressure deficit (VPD), a key driver of physiological processes in  
109 plants and animals (Adolph, 1932; Novick et al., 2024; Wu, N. C. et al., 2024a). For example, in  
110 two environments with the same humidity, that with higher air temperatures will increase VPD (**Fig.**  
111 **1**).

112 Water in the soil matter most for species that rely on underground microrefugia (Wu et al.,  
113 2015; Giacometti and Tattersall, 2023), or for many amphibians, which obtain water directly from  
114 the substrate (Hillman et al., 2009; Comanns et al., 2017). Water fluxes depend on soil properties  
115 (Campbell and Norman, 2000), with extreme examples in sand and clay. Wet sands have an open  
116 texture and dry quickly, whereas wet clays exhibit high soil moisture tension and dry slowly. In  
117 comparison, wet peats dry rapidly and are difficult to rehydrate. However, a valid generalisation is  
118 that below-ground climates maintain higher humidity than surface environments, reducing  
119 dehydration risk to animals (**Fig. 1**), for example in species that shelter underground (Carvalho et  
120 al., 2010), and especially desert dwellers (Woodbury, 1954; Bulova, 2002).

### 121 *1.2 Quantifying water variability and drought indices*

122 Quantifying spatiotemporal shifts in hydric patterns can be approached in various ways to  
123 determine whether an environment is drier than usual. Precipitation and moisture levels, whether in  
124 the air or soil, can be measured long-term via weather stations, or short-term using miniature  
125 environmental data loggers, with Bramer et al. (2018) and De Frenne et al. (2024) providing  
126 examples of commercially available loggers and field deployment considerations. When long-term  
127 datasets are available, various hydrological extreme metrics can be calculated (Pisor et al., 2023).  
128 For instance, using a monthly rainfall example from Sydney, Australia (**Fig. 2**), one can calculate

129 the duration (D) of high rainfall (above 90<sup>th</sup> percentile; high likelihood of flooding) and low rainfall  
130 events (below 10<sup>th</sup> percentile; high likelihood of drought), frequency of extreme rainfall events, and  
131 measures of intensity (I)—the average of extreme rainfall events—magnitude (M)—the maximum  
132 rainfall event—and severity (S)—the cumulative total of extreme rainfall events. Changes in  
133 permanent water sources such as lakes, ponds, streams, and rivers can be quantified via drones  
134 (Spence and Mengistu, 2016; Woodget et al., 2017), satellites (Nath and Deb, 2010; Zhou et al.,  
135 2021), or directly using standard environmental monitoring tools, enabling spatiotemporal  
136 quantification of water body dynamics, particularly relevant for species reliant on temporary or  
137 permanent aquatic habitats (**Table 1**).

138 Relevant for policy makers, quantifying environmental drying risk requires simplification of  
139 complex metrics and variables. One option is relying on meteorological drought indices reflecting  
140 the interplay between the climate variables mentioned in the previous section (**Table 1**). However,  
141 these indexes have no absolute value, meaning that changing indexes relate to species-specific  
142 effects that may change among individuals, populations and communities. The simplest drought  
143 index is the Standardised Precipitation Index (SPI), which relies only on precipitation data (McKee  
144 et al., 1993). Complex counterparts include the Palmer Drought Severity Index (PDSI), which  
145 incorporates the hydrological cycle (Palmer, 1965; Wells et al., 2004), and the Normalized  
146 Difference Vegetation Index (NDVI), which relies on satellite imaging to quantify vegetation  
147 ‘greenness’ (Rouse Jr et al., 1974). Each index has strengths and weaknesses (Zargar et al., 2011),  
148 and all have been used for predicting drought risks, but they are more valuable for long-term  
149 appraisals. The utility of drought indices in analysing short-term biological impacts, particularly in  
150 animals capable of behavioural and physiological adjustments, remains uncertain. However,  
151 integrating drought indices with other environmental processes presents a promising research  
152 direction, as shown by a recent assessment proposed by Crausbay et al. (2024), which integrates  
153 drought indices with vegetation types, canopy cover, slope, time since fire, and other environmental  
154 features, and develops region-specific management actions for decreasing further exposure. These  
155 actions include managing environmental water, restoring sites affected by deforestation and  
156 urbanisation, and promoting ecosystem persistence under drought conditions (Mathwin et al.,  
157 2021). Despite the enormous potential of such analyses, caution is required before implementing  
158 actions, especially those related to water supplies to landscapes, which might damage existing  
159 habitats or favour the accumulation of predators (Mathwin et al., 2021).

160 The analytical context discussed here requires moving from quantifying environmental  
161 drying risk to calculating case-specific exposure risks to drought. This process is challenging,  
162 partially because hydrological variables such as rainfall, evapotranspiration, and soil moisture

163 content exhibit greater uncertainty than temperature trends, due to the stochastic nature of  
164 atmospheric processes (Wu, Y. et al., 2024). Nonetheless, the expanding availability of independent  
165 hydrological models (**Table 1**) offers a timely opportunity to refine predictions of how reptiles and  
166 amphibians will respond to environmental water availability. Once ecologically-relevant water  
167 variables are quantified, numerous approaches may integrate them explicitly with biological traits  
168 across different levels of organization—from molecules to phenotypes to communities. The  
169 interplay between exposure—typically to stressors such as water shortages or drought periods—and  
170 biological response is key to understanding structural and functional consequences, providing  
171 indicators of sensitivity and vulnerability. The following section discuss sensitivity and  
172 vulnerability assessments, with a particular focus on the central theme of this review:  
173 hydroregulation.

### 174 *1.3 Linking Exposure, Sensitivity, and Vulnerability*

175 The vocabulary of ecological climate change research has become complex, with many terms  
176 defining interrelated, yet different concepts, including *Sensitivity*, *Vulnerability*, *Risk*, and  
177 *Resilience*. These terms have gained prominence across scientific and political discussion, as they  
178 are embedded in global agendas on sustainable development, disaster risk reduction, and climate  
179 change, and biodiversity loss (Williams et al., 2008; Scholz et al., 2012; Birkmann and McMillan,  
180 2020). While we acknowledge this conceptual variability, our focus is on identifying the most  
181 relevant information for predicting future trends and anticipating species declines. To this end,  
182 various methodological tools and conceptual frameworks have been employed to assess how  
183 organisms and species respond to climate changes. However, quantifying responses is inherently  
184 complex, as environmental variability operates across all possible spatial and temporal scales, and  
185 science requires operationalisation to specific cases. For instance, both exposure to climate change  
186 and species sensitivity to environmental shifts can differ dramatically within reptiles and  
187 amphibians. Linking exposure, sensitivity, and vulnerability is a goal benefitted by advances in  
188 computational power, more sophisticated statistical models, and large databases, which promote  
189 conceptual and disciplinary bridges. For example, connecting environmental changes not only with  
190 physiology and behaviour but also with conservation biology and ecosystem ecology (Cooke et al.,  
191 2013; Madliger et al., 2018). Methodological approaches have also evolved rapidly, and the most  
192 advanced tools explicitly link climate variables with organismal response by considering the  
193 underlying physiological and behavioural mechanisms that govern their survival and distribution  
194 (see ‘4. *Assessing vulnerability: integrating exposure and sensitivity*’ section).

195 Porter et al. (1973) and Tracy (1976) developed general microclimate models for  
196 quantifying the energy, heat and water budgets of organisms that have been revisited taking into

197 consideration current computing power. Nowadays, it is possible to calculate microclimate at any  
198 location, and with fine temporal resolutions (reviewed in Meyer et al., 2023; Kemppinen et al.,  
199 2024). Some programs even integrate microclimate (**Table 1**) with the calculated heat and water  
200 budgets of organisms (Kearney and Porter, 2020; Kearney and Enriquez-Urzelai, 2023) to estimate  
201 tolerance and distribution limits of organisms under real or any simulated climate (Kearney et al.,  
202 2018; Cheng et al., 2023). Relative to correlational models (Elith and Leathwick, 2009; Peterson et  
203 al., 2011), these developments have added capacity for mechanistic predictions based on  
204 physiological limits of vulnerability to climate change (Riddell et al., 2021; Briscoe et al., 2023;  
205 Pottier et al., 2025), bringing physiological data into the equation.

## 206 **2. Species sensitivity risk: short-term impacts**

207 Many acute and long-term responses to environmental drying are parallel to those triggered by other  
208 generalised stressors (e.g., temperature, pollutants, food restriction), including altered metabolism,  
209 cardiovascular responses, growth, cellular oxidative stress, neuroendocrine pathways, and gene  
210 expression. These common biomarkers are well-documented in the literature, both in terms of  
211 methodology and interpretation (Bustin et al., 2009; Ribou, 2016; Moretti et al., 2017; Madliger et  
212 al., 2018; Lighton, 2019). In this section, we highlight some common responses to drying, with a  
213 focus on water-specific responses and minimally invasive methods, outlined in **Table 2**. Most  
214 comparative studies compare arid adapted and non-arid adapted species, while experimental studies  
215 often involve subjecting animals to restricted water sources or increased environmental dryness.

### 216 *2.1 Genetic responses*

217 Genetic responses to environmental stress are broad across the literature. Here, we focus on three  
218 areas of hydroregulation: (1) the skin barrier, (2) water reabsorption, and (3) cellular repair and  
219 immunity (**Fig. 3**). At the site of evaporation, the skin barrier of reptiles is regulated by the  
220 epidermal differentiation complex (EDC) gene cluster, which encodes proteins essential for  
221 keratinised cells in amniote skin. Among these, corneous beta-proteins (CBPs) genes are crucial for  
222 forming the outer layer of the skin, the stratum corneum (Holthaus et al., 2024), while the *Loricrin*  
223 gene supports alpha-keratinisation in lizard epidermis (Holthaus and Eckhart, 2024). Under arid  
224 conditions, the upregulated expression of CBPs and *Loricrin* increases the thickness and strength of  
225 the stratum corneum, enhancing resistance to dehydration. Comparative genomics between the  
226 desert tortoise (*Gopherus agassizii*) and the temperate aquatic western painted turtle (*Chrysemys*  
227 *picta bellii*) have identified multiple positively selected genes associated with drought resistance.  
228 These genes include *CSTA* and *SDR16C5* (**Fig. 3**), which are involved in keratin formation and  
229 lipid-based waterproofing, respectively (Tollis et al., 2017). In contrast, amphibians rely on mucous  
230 secretions to minimise water loss, along with other functional roles. For example, *Xenopus laevis*

231 upregulates genes (e.g., *grp94* and *grp75*) related to glucose-regulated protein during dehydration,  
232 promoting the synthesis and secretion of protective glycoproteins to reduce water loss while  
233 preserving skin moisture (Malik et al., 2023).

234 Animals adapted to arid conditions exhibit a strong capacity for water reabsorption, enabling  
235 them to produce highly concentrated urine while maintaining salt balance. Several genes are  
236 essential for cellular water reabsorption, particularly those encoding membrane proteins forming  
237 water channels such as aquaporins (AQP). Aquaporin genes, widely present in amphibians and  
238 reptiles (**Fig. 3**), and the proteins they encode are crucial for osmoregulation, including transcellular  
239 water and solute transport (Suzuki and Tanaka, 2009; Shibata et al., 2014; Chen et al., 2019; Wu et  
240 al., 2019). On the other end of osmoregulation, water balance can also be regulated by moving  
241 chloride along with sodium or potassium, creating osmotic gradients between cells and their  
242 surrounding environment, which drive water reabsorption through osmosis. This process requires  
243 ion transport proteins, including those in the Solute Carrier 12 (*Slc12*) family of cation-coupled  
244 chloride cotransporters (**Fig. 3**) (Motoshima et al., 2023). The proteins encoded by these genes  
245 facilitate sodium reabsorption in renal structures like the distal convoluted tubule and thick  
246 ascending limb, generating a hyperosmotic environment that drives water reabsorption via osmosis  
247 (Marra et al., 2012).

248 The kidney is the epitomic organ in water homeostasis, filtering waste while regulating water  
249 and ion balance, making it particularly vulnerable to dehydration stress. Thus, impaired kidney  
250 function figures among the various forms of stress imposed by chronic water deprivation in  
251 amphibians and reptiles. Systems of protection have evolved in lizards and turtles adapted to arid  
252 environments, a condition thought to positively select genes associated with kidney repair, such as  
253 *DCHS2*, related to cell adhesion, and *CD34*, linked to vascular repair (**Fig. 3**) (Tollis et al., 2017;  
254 Araya-Donoso et al., 2022). Another physiological challenge caused by dehydration is disruption of  
255 cellular homeostasis, leading to oxidative stress traduced in the accumulation of reactive oxygen  
256 species, causing tissue damage (Dupoué et al., 2020c; Ritchie and Friesen, 2022). However,  
257 amphibians and reptiles adapted to arid environments activate antioxidant defence mechanisms to  
258 mitigate dehydration-induced oxidative stress (Moreira et al., 2020; de Amaral et al., 2024). For  
259 instance, numerous antioxidant and detoxification genes are regulated by the transcription factor  
260 erythroid 2-related factor 2 (*Nrf2*), a key regulator of oxidative stress responses under elevated  
261 reactive oxygen species levels (**Fig. 3**) (Malik and Storey, 2009). With declining genetic sequencing  
262 costs and expanding analytical capacity, the coming years hold promise for advancing our  
263 understanding of the genetic responses of amphibians and reptiles to hydric stress.



264 *2.2 Neuroendocrine responses*

265 One of the most well-documented effects of pond drying are neuroendocrine responses, which have  
266 been extensively studied in amphibians. Water-dependent tadpoles can adjust their developmental  
267 rate and, therefore the timing of metamorphosis in response to environmental changes (Newman,  
268 1988; Lai et al., 2002; Benard, 2004; Wu and Kam, 2009; Higginson and Ruxton, 2010; Thompson  
269 and Popescu, 2021; Sinai et al., 2022). These environmental cues stimulate the central nervous  
270 system, activating the hypothalamo-pituitary-interrenal/adrenal axis to initiate survival mechanisms.  
271 During pond drying, the hypothalamus increases the production of a corticotropin-releasing  
272 hormone (CRH), stimulating the release of the adrenocorticotrophic hormone (ACTH) and thyroid-  
273 stimulating hormone (TSH) from the pituitary. This, in turn, activates the thyroid and interrenal  
274 glands, elevating thyroid hormones (THs) and corticosterone (CORT) in the bloodstream, which  
275 help manage stress, metabolism, and developmental transitions (Kikuyama et al., 1993; Denver,  
276 1997; Kirschman et al., 2017; Ruthsatz et al., 2020). Increased hormone production accelerates  
277 metamorphosis, thus shortening the larval period and improving survival as aquatic habitats shrink  
278 (Denver, 2013). This hormonal plasticity highlights the resilience of some amphibians, enabling  
279 them to cope with environmental fluctuations and complete their life cycle under harsh conditions.  
280 Some hormones, such as CORT, are released into surrounding waters through various mechanisms  
281 (e.g., secretion and diffusion), and remain stable long enough to be quantified. Therefore, it can be  
282 measured non-invasively from water samples, allowing researchers and managers to monitor stress  
283 in both laboratory and field settings (Ruthsatz et al., 2023a; Ruthsatz et al., 2023b). However,  
284 waterborne and plasma CORT levels may vary across species (Millikin et al., 2019) and depend on  
285 environmental contexts (Mausbach et al., 2022).

286 In terrestrial amphibians and reptiles, acute dehydration triggers fluid balance responses, via  
287 mineralocorticoid hormones, including arginine vasopressin, angiotensin, and aldosterone, all of  
288 which play key roles in water metabolism, helping organisms retain water and maintain circulatory  
289 stability under dehydration stress (McCormick and Bradshaw, 2006; Uchiyama and Konno, 2006;  
290 Dantzler and Bradshaw, 2008; Hillman et al., 2009). CORT also contributes to hydroregulation  
291 through its mineralocorticoid actions (McCormick and Bradshaw, 2006; Dupoué et al., 2016;  
292 Bruschi et al., 2020), though it may not consistently correlate with plasma osmolality within a  
293 species (Dezetter et al., 2022b). Elevated CORT may mobilise energy reserves via muscle  
294 catabolism, reallocating bound water to maintain hydration (Brusch et al., 2018; Dezetter et al.,  
295 2021). These hormones also regulate other interrelated processes, including energy metabolism,  
296 reproduction, social behaviour, and thermoregulation (Ladyman et al., 2006; Bleu et al., 2013;  
297 Carsia et al., 2023; Crino et al., 2024). Therefore, when evaluating hormonal responses to

298 dehydration, researchers should consider these overlapping physiological functions and assess  
299 additional traits linked to hydration and water balance.

### 300 *2.3 Physiological responses*

301 Physiological responses and regulation to acute environmental drying include osmoregulation,  
302 cardiovascular function, metabolism, immunity, and the renin-angiotensin-aldosterone system (as  
303 mentioned above) are well documented in the literature for amphibians (Feder and Burggren, 1992;  
304 Hillman et al., 2009) and reptiles (Pough and Gans, 1982; Dantzler and Bradshaw, 2008; Bradshaw,  
305 2012). Here, we focus on traits commonly measured with conservation relevance, emphasising  
306 minimally invasive protocols (**Table 2**). These can be broadly classified into (1) water loss through  
307 evaporation, (2) hydration state, and (3) daily water flux.

308 Evaporative water loss (EWL) is of considerable interest because it responds immediately to  
309 low air humidity (Mautz, 1980; Hillman et al., 2009). Evaporation depends on both biophysics  
310 (Foley and Spotila, 1978; Campbell and Norman, 2000) and hydration state (Anderson et al., 2017;  
311 Senzano and Andrade, 2018; Weaver et al., 2022), but also on physiology, so that the rate of water  
312 loss tends to be lower in comparable counterparts from more arid environments, across populations  
313 and species (Bentley and Schmidt-Nielsen, 1966; Roberts and Lillywhite, 1983; Cox and Cox,  
314 2015; Salazar and Miles, 2024). EWL mainly occurs through respiratory and cutaneous pathways,  
315 with some influence from ocular and cloacal pathways (**Fig. 1**)(Hillman et al., 2009; Pirtle et al.,  
316 2019), and the combination of these pathways (total EWL, or TEWL) can be measured simply by  
317 the mass loss of the animal (or mass gain of a desiccant) over time, or by respirometry methods  
318 (Hillman et al., 2009; Lighton and Halsey, 2011). Physical models (Senzano et al., 2022) or  
319 mathematical approaches (Riddell et al., 2017) can be used to quantify EWL, but special  
320 consideration of boundary layers is required. These thin layers of fluid (air or water) that form at the  
321 interface between an organism's body and its surrounding environment, usually affect heat, water,  
322 and gas exchange. Experimentally, respiratory and cutaneous EWL (REWL and CEWL,  
323 respectively) can be distinguished by placing a mask with separate airflow for the lungs and skin  
324 (Withers, 1977; Senzano and Andrade, 2018) or by using an impervious membrane to isolate body  
325 regions (Dmiel, 2001). CEWL can also be measured directly with an evaporimeter in a flux  
326 chamber (Lillywhite et al., 2009; Tingley et al., 2012; Oufiero and Van Sant, 2018), a method that  
327 has the added benefit of focusing on specific body regions (Weaver et al., 2022; Weaver et al.,  
328 2023). For broader comparisons, global databases of EWL for frogs and squamates are available to  
329 statistically tease out environmental and phylogenetic drivers of EWL (Cox and Cox, 2015; Le  
330 Galliard, J. F. et al., 2021; Wu, N. C. et al., 2024a). However, EWL datasets for other herpetofauna

331 groups, such as Crocodylia, Testudines, Caudata, and Gymnophiona, remain relatively scarce  
332 compared to those for frogs and squamates.

333 Animals in dry environments survive by efficiently storing water, producing metabolic  
334 water, mobilising water from tissues, and tolerating low body water content (Cloudsley-Thompson,  
335 1999; Hillman et al., 2009; Lillywhite, 2017). The water content in the body of an animal is  
336 expressed as a percentage of whole mass or dry mass and has historically been measured by fully  
337 desiccating specimens (Thorson, 1955; Pough et al., 1983; Taigen et al., 1984). Amphibians  
338 typically have 77–83% water content by body weight (Hillman et al., 2009), while reptiles range  
339 from 63–74% (Thorson, 1968). Shifts in body water allocation may support water balance when  
340 facing dehydration, and the nature of such pathways varies across lineages. For example,  
341 amphibians can absorb water from their bladders (Sawyer and Schisgall, 1956; Schmuck and  
342 Linsenmair, 1997; Suzuki et al., 2015), while snakes and lizards rely on CORT-mediated muscle  
343 catabolism to release water originally associated with proteins (bound water), and may obtain water  
344 as a byproduct of lipid metabolism (Brusch et al., 2018; Dezetter et al., 2021). Internal water  
345 mobilisation can be tracked through changes in blood nutrients, proteins, triglycerides, uric acid,  
346 mineralocorticoid hormones, as well as transcriptome and proteome changes in blood and tissue  
347 samples (Suzuki et al., 2015; Brusch et al., 2018). Given recently established pathways for muscle  
348 catabolism in snakes and lizards, non-invasive methods, such as specimeters, now quantify muscle  
349 changes as proxies for water balance in reptiles (Lourdais et al., 2005; Dezetter et al., 2021). In  
350 amphibian research, a method to assess hydration states involves measuring the body mass of field-  
351 captured individuals, then allowing them to fully rehydrate in a field lab, and recording the  
352 subsequent mass. The difference between the initial field mass and the fully hydrated mass indicates  
353 the degree of dehydration experienced in their environment. For instance, a study on tropical frog  
354 species found that hydration behaviours and voluntary tolerance of dehydration varied with habitat  
355 use, even among closely related species within the same family (Tracy et al., 2014). This approach  
356 provides insights into species-specific water balance strategies and their adaptability to varying  
357 environmental conditions.

358 Blood biochemistry parameters, such as plasma osmolality and haematocrit, serve as  
359 indirect measures of hydration status (**Table 1**) in three dominant contexts, field studies (Capehart et  
360 al., 2016; Moeller et al., 2017; Brischoux and Cheron, 2019; Weaver et al., 2024), laboratory  
361 experiments (Dupoué et al., 2017; Wu et al., 2017; Dezetter et al., 2022b; Chabaud et al., 2023), and  
362 veterinary applications (Perry et al., 2020; Cameron et al., 2024). Plasma osmolality is a direct  
363 indicator of hydration, and is best measured using vapor pressure or freezing-point depression  
364 osmometers (Nevarez et al., 2012; Wright et al., 2013; Buchmiller et al., 2024), as formulas based

365 on solute concentrations often show poor agreement with direct measurements (Dallwig et al.,  
366 2010; Nevarez et al., 2012; Perry et al., 2020). Haematocrit is determined by centrifuging blood in  
367 microcapillary tubes, and it is frequently used as a proxy for hydration status, although it responds  
368 to multiple influencing factors such as blood oxygen-carrying capacity (Brischoux et al., 2011;  
369 Lourdais et al., 2014; Bodensteiner et al., 2021) and does not consistently correlate with plasma  
370 osmolality (Dupoue et al., 2015; Dezetter et al., 2021). Therefore, interpreting haematocrit changes  
371 as indicators of hydration requires caution, considering additional factors affecting blood viscosity  
372 and oxygen transport. Ecologically relevant osmolality measurements should incorporate species-  
373 specific normosmotic values, tolerance to variation, temporal dynamics of osmolality shifts, and  
374 threshold effects on physiological and behavioural water balance regulation (Dessauer, 1970).  
375 Notably, species from xeric environments tolerate greater osmolality fluctuations than those from  
376 mesic habitats, underscoring the importance of species-specific considerations in hydration studies  
377 (Nagy and Medica, 1986; Bruschi and DeNardo, 2017).

378 Whole-animal water flux, encompassing influx, storage, and efflux (**Fig. 1**), can be  
379 quantified using doubly labelled water (DLW), which estimates field metabolic rate and water flux  
380 over extended periods (**Table 1**) (Nagy, 1989). DLW has been widely applied to measure daily  
381 water flux in reptiles across diverse field conditions (Beaupre, 1996; Christian et al., 1999;  
382 Christian et al., 2007; Roe et al., 2008; Harden et al., 2014). For instance, velvet geckos from arid  
383 zones exhibit lower water flux rates year-round compared to those in tropical regions, reflecting  
384 adaptive water conservation strategies (Christian et al., 1998). The ability to estimate both field  
385 metabolic rate and water flux makes DLW a powerful tool for field-based physiological research.  
386 However, certain assumptions in DLW-derived metabolic rates can introduce measurement errors  
387 (Nagy, 1980). For example, high humidity can overestimate metabolic rates due to excessive water  
388 vapor exchange through cutaneous and respiratory surfaces, whereas total water flux aligns more  
389 reliably with gravimetric estimates of TEWL (Anderson et al., 2003). Additionally, DLW is  
390 unsuitable for species with high water flux, such as semi-aquatic reptiles, because rapid water  
391 turnover depletes isotopes too quickly, preventing accurate measurements (Booth, 2002; Jones et  
392 al., 2009). The method has limited use for amphibians due to their high water fluxes, but if the  
393 primary objective is to assess water turnover, the method could be applied to more terrestrial  
394 amphibian species. This would offer valuable insights into the water cost of activity and dispersal  
395 under field conditions.

#### 396 *2.4 Behavioural responses*

397 Terrestrial amphibians and reptiles employ diverse behavioural strategies to regulate water balance,  
398 which can be broadly classified into (1) water-conserving behaviours, (2) water-seeking behaviours,

399 and (3) moisture-harvesting behaviours. These strategies mitigate EWL, optimise hydration, and  
400 enhance survival in desiccating environments. Water-conserving behaviours are those minimising  
401 exposure to drying conditions. When avoidance of dehydration is no longer viable, animals may  
402 engage in water-seeking behaviours, actively locating and consuming water to restore hydration, or  
403 actively collecting and absorbing water from their surroundings. A most universal water-conserving  
404 behaviour involves reducing or shifting diel activity and selecting microhabitats that provide  
405 moisture, such as burrows, and this type of behaviour have been observed in the field (Daltry et al.,  
406 1998; Davis and DeNardo, 2010; Kearney et al., 2018; Moore et al., 2018) and experimentally  
407 (Navas et al., 2002; Székely et al., 2018; Rozen-Rechels et al., 2020; Dezetter et al., 2023). Also,  
408 water-conserving behaviours relate to body temperatures, which usually enhances rates of water  
409 loss (Tracy et al., 2008; Dupoué et al., 2015; Lourdais et al., 2017). Therefore, shifts in  
410 thermoregulatory behaviour, including thermal depression, can contribute to water-conserving  
411 strategies (Ladyman and Bradshaw, 2003; Anderson and Andrade, 2017; Le Galliard, J.-F. et al.,  
412 2021; Camacho et al., 2023). Although the interplay between water and heat budgets complicates  
413 the disentangling of hydoregulation and thermoregulation mechanisms (Pintor et al., 2016; Rozen-  
414 Rechels et al., 2019), recent modelling approaches considering both joint mechanisms and  
415 microclimatic data are improving our understanding of behavioural responses to drying and heating  
416 (Kearney et al., 2018; Moore et al., 2018; Encarnación-Luévano et al., 2021). By strictly controlling  
417 for temperature, experimental studies have demonstrated hydoregulation behaviours through the  
418 active selection of moister microclimate in both wet-skinned amphibians (Mitchell and Bergmann,  
419 2016) and dry-skinned reptiles (Dezetter et al., 2023). This behaviour mitigates the acute effects of  
420 desiccating conditions. These findings suggest that both resistance to water loss and hydric  
421 performance response curves may influence the timing of behavioural responses to drying in  
422 reptiles and amphibians.

423         Some animals can reduce water loss by modifying body posture and preferring those that  
424 reduce the exposed area to the environment (**Table 2**). Placing limbs against the body and using  
425 skin folds to cover ventral surfaces against the substrate, as in Anura (Pough et al., 1983) or coiling  
426 in Caudata (Cohen, 1952) greatly reduce TEWL (Spotila and Berman, 1976). In addition to postural  
427 changes, several species of arboreal frogs use limbs to spread waxy films over their body surfaces  
428 during dry seasons or produce cocoons to reduce CEWL (Lillywhite, 2006). By manipulating the  
429 hydration state via moisture gradients and assessing postural adjustments, experimental studies can  
430 examine the determinants of these behaviours and their benefits for maintaining hydration (Navas et  
431 al., 2002; Mitchell and Bergmann, 2016). In reptiles, behaviours such as coiling in snakes or  
432 adopting tucked-in postures in lizards may also confer water-saving benefits. However, this aspect

433 has received comparatively limited attention, mostly restricted to studies on egg-brooding behaviour  
434 in snakes, where subtle postural shifts can reduce egg surface exposure and limit water loss from  
435 the egg clutch (Lourdais et al., 2007; Stahlschmidt et al., 2008; Stahlschmidt and DeNardo, 2010).  
436 Finally, the simple closure of eyes can help reduce water loss through the permeable eye membrane  
437 in lizards (Pirtle et al., 2019).

438         When avoiding and restricting drying is no longer possible, reptiles and at least some  
439 amphibians will seek water to restore their hydration state (**Table 2**). Experimental systems called  
440 “olfactometers” designed by Grubb (1973), and follow-up studies with maze designs, have  
441 demonstrated that frogs and lizards can detect and locate free-standing water via olfactory cues  
442 (Navas et al., 2002; Madelaire et al., 2020; Ouellet et al., 2020; Lorrain-Soligon et al., 2022;  
443 Northrop, 2024). However, generalising is not possible for amphibians. Finding generic water for  
444 hydration and finding specific waters for reproduction seem to be independent processes, and both  
445 have been identified in some species. For example, telemetric studies show that poison frogs rely on  
446 odour cues from stagnant water to find new breeding pools (Serrano-Rojas and Pašukonis, 2021).  
447 However, this ability varies across species (Reshetnikov, 1998; Maia, 2014) and may relate to  
448 drying tolerance and habitat aridity (Cruz-Piedrahita et al., 2018; Galindo et al., 2024). Particularly,  
449 some anuran species rely on structured water search strategies, while others find water by erratic  
450 exploration (Maia, 2014). Finally, drinking matters for some species only as others will rely on a  
451 specialized, richly vascularized region of the pelvic skin (Willumsen et al., 2007). It has also been  
452 reported for snakes subjected to field experimental dehydration or rehydration after capture  
453 (Brischoux et al., 2017; Dezetter et al., 2022b) and is triggered by physiological thresholds such as  
454 hydration status (Sandfoss and Lillywhite, 2019; Edwards et al., 2021). Quantifying water-seeking  
455 behaviour (e.g. time to find water source) should be of consideration for habitat restoration  
456 managers when optimising water resources for herpetofauna to persist and flourish in a given  
457 habitat (Mathwin et al., 2021).

458         Moisture and rain-harvesting behaviours are also observed in reptiles (Sherbrooke, 1993;  
459 Joel et al., 2017), These behaviours include snakes coiling and flattening their bodies, lizards  
460 flattening their bodies, and both lizards and tortoises raising their abdomen and lowering their heads  
461 and tails (Repp and Schuett, 2008; Glaudas, 2009; Yenmiş et al., 2024). Similarly, some postural  
462 adjustments in amphibians can facilitate moisture and water uptake through the skin, particularly  
463 through the pelvic patch (McClanahan Jr and Baldwin, 1969; Bentley and Main, 1972; Hillyard et  
464 al., 1998; Word and Hillman, 2005). Overall, water-searching behaviours and related adaptations  
465 are critical for understanding species sensitivity and resilience to aridification. For instance,

466 invasive frogs at the forefront of their invasion show distinct water-searching tendencies, with stress  
467 differentially affecting this behaviour (Madelaire et al., 2020).

### 468 *2.5 Life history responses*

469 Water stress can impact life history by influencing (1) growth and the rate of development, (2) body  
470 size, and (3) reproduction. Animals can adjust their rate of development under different  
471 environmental conditions (see hormonal control under the '2.3 Neuroendocrine responses' section).  
472 This developmental plasticity can be either adaptive or maladaptive, depending on whether the  
473 developmental environment matches the conditions an individual experiences later in life  
474 (Monaghan, 2008; Beaman et al., 2016). For the larval stages of amphibians, reduced water  
475 availability, such as pond drying, can accelerate larval development, leading to smaller body sizes  
476 or incomplete metamorphosis due to resource constraints, crowding, poorer water quality, and  
477 increased predation risk (Márquez-García et al., 2010; Gomez-Mestre et al., 2013; Albecker et al.,  
478 2023). Some species, however, do not show changes in developmental rate nor exhibit delayed  
479 larval development under drying conditions (Richter-Boix et al., 2011), promoting the importance  
480 of species-specific responses. It is also clear that developmental plasticity to pond drying can have  
481 carry-over effects on post-metamorph individuals and adults. Under pond drying conditions,  
482 metamorphs have lower thermal tolerance, are less exploratory and more stressed, and have lower  
483 jumping performance and lower immunity (Gervasi and Foufopoulos, 2008; Crespi and Warne,  
484 2013; Charbonnier et al., 2018; Brannelly et al., 2019; Ohmer et al., 2023; Nolan et al., 2025; Wu et  
485 al., 2025). Size is particularly important because larger individuals are associated with increased  
486 survival rate, performance (Cabrera-Guzmán et al., 2013), and lower risk to disease progression for  
487 the same pathogen load (Brannelly et al., 2018; Wu et al., 2018).

488 Water availability also plays a critical role in the reproductive success of egg-laying reptiles,  
489 influencing both egg survival and offspring development. For species that lay eggs on land, eggshell  
490 thickness and composition are key determinants of desiccation risk. Flexible-shelled eggs, which  
491 lack or have minimal calcareous layers (most squamates and some chelonians), are more porous and  
492 susceptible to water loss compared to rigid-shelled eggs with a well-developed calcareous layer  
493 (crocodilians, some chelonians, and a few squamates) (Legendre et al., 2022). A meta-analysis  
494 showed that substrate moisture had a small but significant effect on hatchling length and mass for  
495 reptiles, as well as on sex ratios specifically for chelonians, but not on incubation duration (Bell et  
496 al., 2025). However, this meta-analysis did not consider differences in eggshell type due to  
497 phylogenetic biases in categorising shell type. Species that nest in arid environments tend to have  
498 highly absorbent eggshells and thicker shells, suggesting that species with flexible eggshells may be  
499 more vulnerable to environmental drying (D'Alba et al., 2021; Debruyn et al., 2023). At the other

500 extreme, excessive moisture can also be detrimental, leading to reduced oxygen availability in nests,  
501 lower hatchling success, and high embryo mortality (Marco and Díaz-Paniagua, 2008; Gatto and  
502 Reina, 2022; Warner et al., 2023). These findings underscore that water management in nesting  
503 habitats of reptiles is as crucial for egg and juvenile survival as it is for adult life history strategies  
504 in response to environmental dryness.

505 Drying stress during reproduction and early life can impact reproductive output and  
506 offspring phenotypes in reptiles (Dupoué et al., 2018; Dupoué et al., 2020b; Dezetter et al., 2021).  
507 Successful reproduction requires substantial water investment, particularly during gravidity in  
508 oviparous reptiles (Brusch et al., 2019; Dupoué et al., 2020a), and even more so in viviparous  
509 species, where pregnant females experience increasing hydration demands as embryos develop in  
510 utero (Dupoué et al., 2015; Lourdais et al., 2015; Lourdais et al., 2017). To reduce water loss, gravid  
511 females may adjust their behaviour, seeking moister microhabitats (Lourdais et al., 2017). However,  
512 under limited water availability, they face a trade-off between self-maintenance and offspring  
513 investment, often prioritizing embryonic water allocation at their own physiological expense  
514 (Dupoué et al., 2015; Dupoué et al., 2020a; Dezetter et al., 2021). Maternal dehydration can have  
515 severe reproductive consequences, including follicular resorption at early stages (Capehart et al.,  
516 2016; Zani and Stein, 2018), reduced investment in eggs, and thinner eggshells with altered immune  
517 function (Brusch et al., 2019). In later stages, maternal water deprivation increases embryonic  
518 mortality (Dezetter et al., 2021), potentially contributing to drought-driven population decline  
519 (Madsen et al., 2023). These demographic costs may be exacerbated by fecundity trade-offs, as  
520 larger females carrying more embryos experience greater physiological stress (Dupoué et al., 2015;  
521 Lourdais et al., 2015; Dezetter et al., 2021). However, the generality of maternal-offspring water  
522 trade-offs remains uncertain, as some species, such as *Anolis sagrei*, exhibit no observable effects of  
523 maternal dehydration on fecundity, egg size, or egg hydration (Wayne et al., 2025).

### 524 **3. Species sensitivity risk: long-term impacts**

525 Beyond the immediate effects of drying, animals must also cope with longer periods of desiccation  
526 for populations to survive. Understanding the long-term implications of water limitation requires an  
527 integrative approach that incorporates adaptation, developmental plasticity, and demographic shifts  
528 through experimental and field-based studies. This section explores three key aspects of long-term  
529 water deficit impacts: (1) the role of heritability and acclimation in hydroregulation traits, and (2)  
530 the influence of water availability on body size evolution.

#### 531 *3.1 Heritability and acclimation of hydroregulation traits*

532 Repeatability and heritability experiments are key to determining whether hydroregulation traits are  
533 targets of natural selection. Although often labour-intensive, repeatability measures the consistency



534 of a trait within individuals under similar physiological conditions, whereas heritability assesses its  
535 genetic transmission across generations (Wolak et al., 2012). However, our understanding of the  
536 repeatability and heritability of hydroregulation traits remains limited. Empirical studies provide  
537 some insights into the genetic and phenotypic basis of these traits. For instance, covariance between  
538 thermal traits and skin resistance to water loss has been observed in *Plethodon metcalfi* (McTernan  
539 and Sears, 2022), significant repeatability of EWL in *Sceloporus consobrinus* (Oufiero and Van  
540 Sant, 2018), and the moderate heritability of desiccation tolerance in *Lampropholis* skinks (Martins  
541 et al., 2019). While these studies suggest that hydroregulation traits exhibit some degree of  
542 plasticity, large-scale assessments of frog populations across natural climatic gradients indicate low  
543 variation in EWL rates, suggesting potential constraints on plasticity (Davies et al., 2015; Bovo et  
544 al., 2023). These findings emphasize the need for broader geographic and taxonomic coverage  
545 (White et al., 2021; Herrando-Pérez et al., 2023) to clarify the magnitude of variation in  
546 hydroregulation traits and how they scale over time to shape long-term responses to environmental  
547 change.

548         Acclimation, the ability of organisms to adjust to changing environmental conditions, is  
549 another key aspect of survival in fluctuating climates, especially in water-scarce environments. Our  
550 understanding of the acclimation potential of hydroregulation traits is limited compared to thermal  
551 physiological counterpart traits (Seebacher et al., 2015), although some studies have explored how  
552 organisms modify hydroregulation in response to thermal acclimation (Davies et al., 2015). For  
553 example, thermal acclimation during development can lead to changes in TEWL that persist until  
554 adulthood in snakes (Dezetter et al., 2022a), while the TEWL of lizards decreased in response to  
555 warmer temperatures (Vicenzi et al., 2021). Riddell et al. (2019) highlighted that temperature is an  
556 important cue for developing a desiccation-resistant phenotype, by regulating water loss through the  
557 regression and regeneration of capillary beds in the skin. The growing literature on disentangling  
558 the differences in acclimation effects of temperature and drying exposure on hydroregulation  
559 provides a promising area for understanding long-term water restrictions or simulated drying to  
560 assess the plasticity of these traits across different species (Kobayashi et al., 1983; Kattan and  
561 Lillywhite, 1989; Moen et al., 2005; Riddell et al., 2018a; Rozen-Rechels et al., 2020; Weaver et al.,  
562 2023).

### 563 3.2 *Water availability and body size evolution*

564 One notable potential long-term effect of changes in precipitation is altered body size (Gouveia and  
565 Correia, 2016; Guo et al., 2019; Pincheira-Donoso et al., 2019). Two contrasting mechanisms have  
566 been proposed to explain this relationship: (1) the ‘resource hypothesis’, where higher rainfall  
567 boosts primary productivity, supporting larger individuals due to greater food availability

568 (Rosenzweig, 1968), and the ‘water conservation hypothesis’, where arid environments favour  
569 larger individuals because lower surface-area-to-volume ratios reduce evaporative water loss  
570 relative to smaller individuals (Nevo, 1973; Gouveia and Correia, 2016). Evidence from reptile  
571 communities supports the resource hypothesis, with some species increasing in size as precipitation  
572 rises (Stanley et al., 2020). However, amphibians show a more complex pattern, with body size  
573 correlating with higher precipitation in cooler climates but with lower precipitation in warmer  
574 regions, possibly indicating a transition from resource-driven to desiccation-resistance-driven  
575 selection (Sheridan et al., 2022). Despite these findings, body size responses to climatic water  
576 balance remain debated (Servino et al., 2022). In contrast with reptiles, the permeable skin of  
577 amphibians makes them particularly vulnerable to desiccation. This key difference may contribute  
578 to diverging size trends between reptiles and amphibian communities in response to water  
579 availability. To clarify these patterns across sites, long-term body size monitoring in conjunction  
580 with environmental data—including analyses of museum specimens with historical climate  
581 records—can help elucidate the drivers of body size evolution. Further research is needed to assess  
582 long-term changes in skin permeability and their potential correlation with body size variation.

#### 583 **4. Assessing vulnerability: integrating exposure and sensitivity**

##### 584 *4.1 Vulnerability indices and organismal traits*

585 A number of vulnerability indices of physiological stress, extinction risk, activity time constraints,  
586 habitat suitability, or range shifts have been proposed depending on the question of interest  
587 (Deutsch et al., 2008; Kearney and Porter, 2009; Sinervo et al., 2010; Lertzman-Lepofsky et al.,  
588 2020; Souza et al., 2023). These indices are projected across space and time and are based on the  
589 experimental estimation of fitness-related traits. Some indices relate environmental variables with  
590 physiological thresholds (e.g., desiccation tolerance, performance curves; Greenberg and Palen,  
591 2021; Anderson et al., 2023), whereas more complex counterparts are based on biophysical models  
592 designed to reflect energy and water exchanges between animals and their microclimatic  
593 environments (Kearney et al., 2013; Kearney et al., 2018; Briscoe et al., 2023). Importantly, thermal  
594 biology information characterises most indexes (Taylor et al., 2020) despite the high relevance of  
595 hydroregulation for water-sensitive groups such as amphibians (Lertzman-Lepofsky et al., 2020;  
596 Greenberg and Palen, 2021; Wu, N. C. et al., 2024a).

597 Models and indexes have been used to predict biological constraints on fitness, using as proxies’  
598 development, growth, activity, reproduction, and survival (Sinervo et al., 2010; Kearney et al.,  
599 2018). This is because fitness-related traits are key to informing vulnerability to a given source of  
600 physiological stress. Yet disagreements exist on whether traits and what traits are good predictors  
601 for informing causal links of environmental changes on populations and species (Calosi et al., 2008;

602 Beissinger and Riddell, 2021). For example, common modelling variables related to  
603 hydroregulation include hydration level, rates of water loss, measures of water acquisition (seeking  
604 out water sources or specific microhabitats that enhances water uptake or maintenance), and the  
605 concentration of body fluids (**Table 2**). Hydroregulation traits are integrated with various functions  
606 related to gas exchange, energetics, thermoregulation, and reproduction as previously highlighted in  
607 section ‘2.1 *Species sensitivity risk: short-term impacts*’. Thus, and according to the physiology of  
608 the target groups, models exclusively based on hydroregulation may under- or overestimate  
609 vulnerability to climate change (Riddell et al., 2018a; Rozen-Rechels et al., 2019). The use of  
610 multiple physiological thresholds such as thermal tolerance, reproduction, and growth with  
611 hydroregulation through experimental manipulation of environmental stressors or inputting  
612 appropriate parameters to mechanistic models will provide more holistic estimations of  
613 vulnerability to climate change.

#### 614 *4.2 Challenges in predicting vulnerability*

615 Predicting vulnerability to environmental stressors and how this may scale-up to population or  
616 species-level responses remains a key challenge for the conservation of biodiversity (Bovo et al.,  
617 2018). Practical limitations include characterising with appropriate data species-specific  
618 microclimates, both temporally and spatially (Briscoe et al., 2023). These limitations extend to  
619 single-population estimates, and the sometimes-related use of an average value to represent a  
620 whole-species. The validity of such approaches is context-specific, but they may not reflect across-  
621 population variation in sensitivity to thermal (Herrando-Pérez et al., 2019; Senior et al., 2019; Bovo  
622 et al., 2023) and/or drying as well as population plasticity/adaptation to drying condition. The same  
623 principle applies to studies using closely related species to represent threatened counterparts  
624 (Reemeyer et al., 2024). Mechanistic models that embrace population variability and plasticity in  
625 response to environmental drying will allow more explicit predictions of vulnerability across a  
626 species range (see ‘Future directions’). Validating these predictions is essential, particularly for  
627 models that estimate survival, reproduction, and activity, which should be tested against  
628 observational data to ensure accuracy. Natural history observations (Greene, 2005), and large-scale  
629 longitudinal field and laboratory studies, particularly when there are geographically biased data  
630 (White et al., 2021; Herrando-Pérez et al., 2023), can help validate mechanistic models when  
631 predicting biological impacts of climate change across a species range or communities (Kearney et  
632 al., 2018; Enriquez-Urzelai et al., 2019; Riddell et al., 2019; Briscoe et al., 2023).

633 **5. Future directions**

634 *5.1 Linking gene expressions to functional changes in response to environmental drying*

635 Whole genome sequencing is becoming increasingly affordable and accessible for researchers and  
636 conservationist (Theissinger et al., 2023; Hogg, 2024). As we previously highlighted, several genes  
637 have been identified that are linked to an animal's hydroregulation. Understanding how the  
638 expression of these genes translates into functional changes in an animal's water balance is key to  
639 uncovering the genetic mechanisms underlying plasticity in response to environmental drying  
640 (Somero, 2010). Riddell et al. (2019) identified, in salamanders, over 500 genes in response to  
641 acclimation to different temperatures and vapour pressure deficits. Network analysis of these genes  
642 revealed suites of gene networks associated with the plasticity of skin resistance and the regulation  
643 of skin blood vessel growth. For example, the expression of *hydroperoxide isomerase (ALOXE3)*, a  
644 gene involved in regulating transepidermal water loss, was highlighted. This study underscores an  
645 important research direction for identifying which genes are targets of selection when inferring the  
646 adaptive potential of species to warming and drying environments. Epigenome-wide association  
647 studies represent a promising approach for establishing causal relationships between changes in the  
648 epigenome and phenotypic plasticity (Fanter et al., 2022).

649 *5.2 Inter- and transgenerational plasticity in response to environmental drying*

650 Parental environments can shape offspring phenotype via epigenetic mechanisms such as DNA  
651 methylation, histone modifications, and non-coding RNAs (Galloway and Eттerson, 2007; Beaman  
652 et al., 2016; Loughland et al., 2021; Husby, 2022). Intergenerational and transgenerational plasticity,  
653 which describe epigenetic inheritance across one or multiple generations, could buffer populations  
654 against environmental change, particularly if parental and offspring environments match (Shama  
655 and Wegner, 2014; Pettersen et al., 2024). However, despite growing interest in these mechanisms,  
656 few studies have explored these processes in amphibians and reptiles. One promising research  
657 direction is to experimentally test whether epigenetic modifications induced by water stress persist  
658 across generations and whether they enhance desiccation resistance. This could involve controlled  
659 desiccation experiments, tracking epigenetic changes and hydroregulation traits across multiple  
660 generations (Dupoue et al., 2015; Dupoué et al., 2018), or comparing populations from  
661 environments with different hydric regimes to assess whether ancestral exposure to aridity  
662 influences offspring water balance. Finally, we identify a critical question remaining unanswered:  
663 do hydroregulation strategies have an evolutionary limit? Studies on thermal tolerance suggest that  
664 plasticity alone may not be enough to ensure survival under extreme climate shifts (Morgan et al.,  
665 2020), but we lack similar insights for hydroregulation. Addressing this could inform conservation  
666 strategies, helping predict whether species can adjust to future drying events or if their  
667 physiological flexibility has constraints.

668 *5.3 Modelling plasticity and adaptation in response to environmental drying*

669 Animals can remodel their phenotype (physiology, morphology, and behaviour) to maintain optimal  
670 performance across a broad range of environments. This plastic response is a well-recognised  
671 phenomenon in predicting adaptive responses to climate change (Seebacher et al., 2015; Urban et  
672 al., 2016). Models that incorporate plasticity or adaptation tend to better predict a species' extinction  
673 risk or range contraction (Riddell et al., 2018b; Kellermann et al., 2020). Therefore, we encourage  
674 modelling studies to explicitly incorporate plasticity to provide realistic assessments of vulnerability  
675 to climate change (Bush et al., 2016; Gallegos et al., 2024). There is a substantial body of literature  
676 on plastic responses to pond drying in tadpoles (Gomez-Mestre et al., 2013; Székely et al., 2017;  
677 Delgadillo Méndez et al., 2024; Wu et al., 2025) and responses to soil moisture during embryo  
678 development in reptiles (reviewed in Bell et al., 2025). However, in studies of terrestrial drying,  
679 many acclimation experiments aimed at quantifying plastic responses have primarily focused on  
680 temperature effects on hydoregulation traits. This can confound causal inferences between the  
681 effects of temperature and drying (see '3. *Species sensitivity risk: long-term impacts*' section). For  
682 instance, the temperature effects on EWL may partially arise from the temperature-dependent nature  
683 of the metabolic rate (MR), as MR and the rate of gas exchange are closely linked to respiratory  
684 EWL (Woods and Smith, 2010), but see Riddell et al. (2024). It is possible that plasticity to water  
685 restrictions may differ from plasticity to temperature changes, potentially altering model predictions  
686 of extinction risk. Further studies across a broader range of species are needed to make biologically  
687 meaningful statements about the generality of within- and across-generation plasticity to drying,  
688 and to improve inferences in modelling vulnerability to future environmental drying scenarios.  
689 Finally, models explicitly testing adaptive evolution of traits should be more widespread used  
690 (Hansen, 2012; Moen et al., 2022).

691 *5.4 Translating knowledge for managing habitats*

692 Incorporating knowledge on hydoregulation strategies with projected changes in environmental  
693 water into land management and conservation planning, particularly at the microhabitat scale, could  
694 help mitigate the impacts of habitat modification and climate change on reptiles and amphibians  
695 (Nowakowski et al., 2018; Pottier et al., 2025). Complex microhabitats, such as heterogeneous  
696 vegetation patches and burrows, provide hydric refuges during droughts, which reptiles and  
697 amphibians can exploit through behavioural hydoregulation to avoid desiccation (see '2.4  
698 *Behavioural responses*' section). Conservation efforts should thus prioritise the protection,  
699 restoration, or creation of such microhabitats within the range of the focal species to support  
700 persistence under increasingly drying conditions (Moore et al., 2018; Weaver et al., 2024). For  
701 example, protecting swamps from groundwater loss and surface water contamination has also been  
702 recommended for conserving *Eulamprus leuraensis*, an endangered swamp-specialist skink in

703 Australia (Gorissen et al., 2017). Interventions aimed at extending hydroperiods may protect some  
704 amphibians from larval desiccation and enhance population viability by increasing recruitment  
705 (Hamer et al., 2016; Mathwin et al., 2021; Mathwin et al., 2023). Likewise, habitat water  
706 supplementation through mist irrigation can extend activity time in reptiles and amphibians under  
707 arid conditions (Ackley et al., 2015; Mathwin et al., 2021) and support reproduction and dispersal in  
708 amphibians (Mitchell, 2001; Channing et al., 2006; Hoffmann and Mitchell, 2022). Providing  
709 supplemental hydration, such as drinking water for targeted individuals, could also be an effective  
710 conservation strategy for small, range-limited species (Weaver et al., 2024), particularly benefiting  
711 gravid or pregnant females by mitigating physiological and reproductive costs during severe  
712 droughts (Capehart et al., 2016; Dezetter et al., 2021; Bedard et al., 2025). However, the feasibility  
713 of such interventions is questionable (Mathwin et al., 2021; Weaver et al., 2024) and further  
714 research is needed to assess the effectiveness, potential negative, and species-specific outcomes of  
715 hydrological manipulation and micro-habitat scale management as a conservation tool. To facilitate  
716 the global implementation of conservation, informed by recent and emerging hydroregulation  
717 research, publications are also needed in more accessible and taxa- and region-specific journals and  
718 government reports (Amano and Berdejo-Espinola, 2024; Choi et al., 2024).

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**TABLES**

**Table 1.** Environmental water variables and example indices of environmental dryness with definitions and calculations of the variables, their interpretation for reptiles and amphibians, the temporal resolution scale available, and non-exhaustive examples of online global datasets to extract environmental water variables and indices.

Name	Definition and calculation	Interpretation	Resolution	Online database
<b>Environmental water</b>				
Precipitation ( <i>P</i> )	Amount of rainfall per unit area (mm or kg m <sup>2</sup> ).	Less rainfall = less water available for animals to rehydrate.	Hourly to yearly	<i>CHIRPS</i> : Global daily rainfall from 1981 to near-present (Funk et al., 2015). <i>WorldClim 2</i> : Average yearly precipitation and seasonality from 1970 to 2000 (Fick and Hijmans, 2017). <i>CHELSA</i> : Precipitation, and potential evapotranspiration (1981–2010) with future scenarios at three future time periods (2011–2040, 2041–2070, and 2071–2100) under three shared socioeconomic pathways (SSP126, SSP370, SSP585) and across five Earth system models (Brun et al., 2022). <i>CHELSA-EarthEnv</i> : Global daily rainfall from 2003 to near-present (Karger et al., 2021). <i>TerraClim</i> : Average (1958–2019) global rainfall with future scenarios of +2°C and +4°C (Abatzoglou et al., 2018).
Atmospheric moisture content (RH or <i>e</i> )	Amount of moisture (water vapour) the air holds. Typically expressed as relative humidity (%), absolute humidity (g cm <sup>3</sup> ), or actual vapour pressure ( <i>e<sub>a</sub></i> ; kPa)	Less moisture in the air = higher evaporation of water through evaporative surfaces.	Seconds to yearly	<i>CHELSA</i> : Near-surface relative humidity (1981–2010) with future scenarios at three future time periods (2011–2040, 2041–2070, and 2071–2100) under three shared socioeconomic pathways (SSP126, SSP370, SSP585) and across five Earth system models (Brun et al., 2022). <i>TerraClim</i> : Average (1958–2019) global vapor pressure with future scenarios of +2°C and +4°C (Abatzoglou et al., 2018). <i>MODIS</i> : 5-minute interval global water vapour data ( <a href="https://ladsweb.modaps.eosdis.nasa.gov/missions-and-measurements/science-domain/water-vapor/#modis">https://ladsweb.modaps.eosdis.nasa.gov/missions-and-measurements/science-domain/water-vapor/#modis</a> ). <i>MODIS</i> : 8-day to annual interval global evapotranspiration data ( <a href="https://ladsweb.modaps.eosdis.nasa.gov/missions-and-measurements/science-domain/evapotranspiration/">https://ladsweb.modaps.eosdis.nasa.gov/missions-and-measurements/science-domain/evapotranspiration/</a> ).
Soil moisture content ( $\Phi$ )	Amount of water the soil holds. Expressed as volume (m <sup>3</sup> ), weight (kg) or water potential (kPa).	Relevant for animals that use burrows to acquire (from the soil/substrate), conserve (no	Seconds to yearly	<i>NicheMapR</i> : Above and below-ground microclimate from various sources (Kearney and Porter, 2017). <i>Microclimc</i> : Above and below-ground microclimate (Maclean and Klinges, 2021).

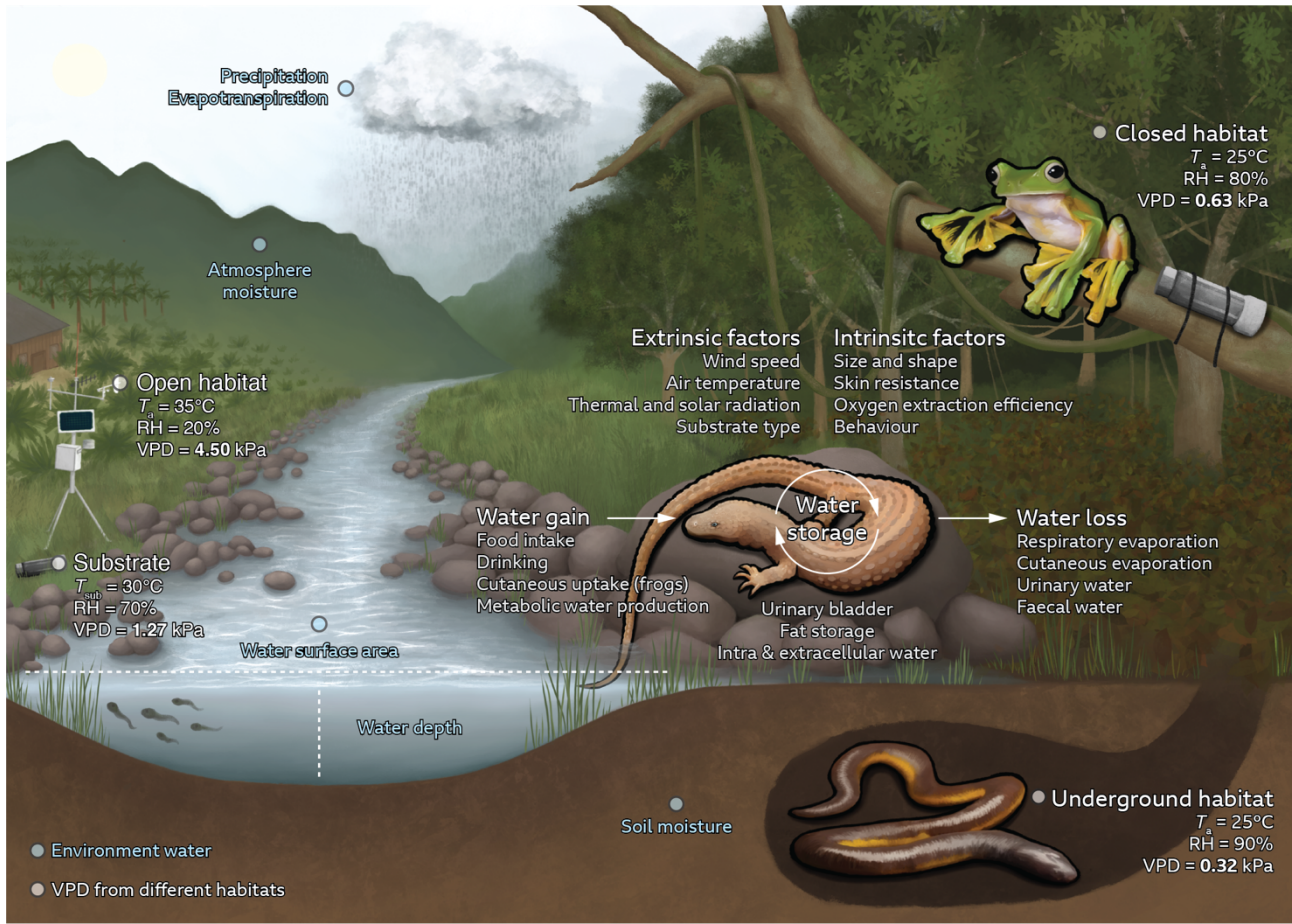
		water exchange) and reduce water loss.		
Water surface area	Land area covered by freshwater (%) e.g. lakes, rivers.	Amount of large-bodied freshwater sources for animals to rehydrate and/or breed.	Average over set years.	<i>HYDROSHEDS</i> : Global hydrographic products such as catchment boundaries, river networks, and lakes at multiple resolutions and scales ( <a href="http://www.hydrosheds.org">www.hydrosheds.org</a> ).
<b>Environmental dryness indices</b>				
Aridity index (AI) or climate moisture index (CMI)	$AI = P/PET$ $CMI (mm \text{ or } kg \text{ m}^2 \text{ month}^{-1}) = P - PET$  The difference (CMI) or ratio (AI) between the average annual precipitation ( $P$ ) and potential evapotranspiration ( $PET$ ).	Indicator of the degree of dryness of the climate.	Depending on $P$ and $PET$ resolution, but typically monthly to yearly average.	<i>CHELSA</i> : CMI (1981–2010) with future scenarios at three future time periods (2011–2040, 2041–2070, and 2071–2100) under three shared socioeconomic pathways (SSP126, SSP370, SSP585) and across five Earth system models (Brun et al., 2022). <i>Global-AI_PET_v3</i> : Global hydro-climatic data averaged (1970–2000) monthly and yearly (Zomer et al., 2022).
Drought index	Standardised index representing meteorological drought based on different formulas: <ul style="list-style-type: none"> <li>• Palmer Drought Severity Index (PDSI)</li> <li>• Standardised Precipitation Index (SPI)</li> <li>• Normalized Difference Vegetation Index (NDVI)</li> </ul>	Indicator of change in environmental dryness relative to ‘normal’ conditions of the location. The intensity, frequency, and duration of drought events can be calculated from these indices.	Monthly to decades.	<i>TerraClim</i> : Average (1958–2019) global PDSI with future scenarios of +2°C and +4°C (Abatzoglou et al., 2018). <i>Dai_et_al_2004</i> : global PDSI under three shared socioeconomic pathways: 1870–2002, SSP245, and SSP585 (Dai et al., 2004). <i>MODIS</i> : 16-day and monthly interval global NDVI ( <a href="https://modis.gsfc.nasa.gov/data/dataproduct/mod13.php">https://modis.gsfc.nasa.gov/data/dataproduct/mod13.php</a> ).
Vapour pressure deficit (VPD)	$VPD (kPa) = e_s - e_a$ The difference between the amount of moisture in the air ( $e_a$ ) and how much moisture the air can hold when it is saturated at known temperature ( $e_s$ ).	Determines desiccation risk and relates to the primary productivity of ecosystems (plant growth, food availability).	Depending on $e_s$ and $e_a$ resolution, but typically monthly to yearly average.	<i>CHELSA</i> : VPD (1981–2010) with future scenarios at three future time periods (2011–2040, 2041–2070, and 2071–2100) under three shared socioeconomic pathways (SSP126, SSP370, SSP585) and across five Earth system models (Brun et al., 2022). <i>TerraClim</i> : Average (1958–2019) global VPD with future scenarios of +2°C and +4°C (Abatzoglou et al., 2018).

1468 **Table 2.** Example measurements to estimate of an animal's water loss/balance with definitions and calculations, and  
 1469 their interpretation for reptiles and amphibians.

Name	Definition/calculation	Interpretation
Evaporative water loss	Water loss through evaporative surfaces. Typically expressed as rate of water loss per unit time ( $\text{g h}^{-1}$ ), or resistance to water loss ( $\text{s cm}^{-1}$ ) Measurements of EWL can be whole-body, regional (e.g. ocular, dorsal, ventral, cloacal), exposed cutaneous surface area, or respiratory.	Indicator of the animals' risk of drying to the environment.
Water content	Whole-body mass: The amount of water in the animal. Typically expressed as percentage of whole mass or dry mass (%) in relation to standard (hydrated) body mass. Muscle: The amount of water in a sample of muscle tissue.	How much water is stored and available for the animal to use. Note that fat storage is another source of water through aerobic metabolism.
Blood biochemistry	Osmolality: Biomarker that measures the concentration of dissolved solutes in the blood. Typically expressed as milliosmoles per kilogram or litre of solvent ( $\text{mosmol kg}^{-1}$ , $\text{mosmol l}^{-1}$ ). Haematocrit: Proportion of blood volume occupied by red blood cells. Expressed as percentage of blood volume (%).	Indicator of dehydration status.
Water flux	Isotopic analysis of doubly labelled water which traces the movement of water molecules between the organism and environment. Typically expressed as $\text{ml kg}^{-1} \text{day}^{-1}$ .	Estimate of daily water flux from free-ranging animals. Usually not suitable for semi-aquatic and aquatic species.
Water-seeking or conserving behaviour	Behaviours associated with seeking water (directional movement) and/or saving water (posture to diminish exposed body surface areas, shelter seeking, inactivity).	Indicator of behavioural focus on water balance.

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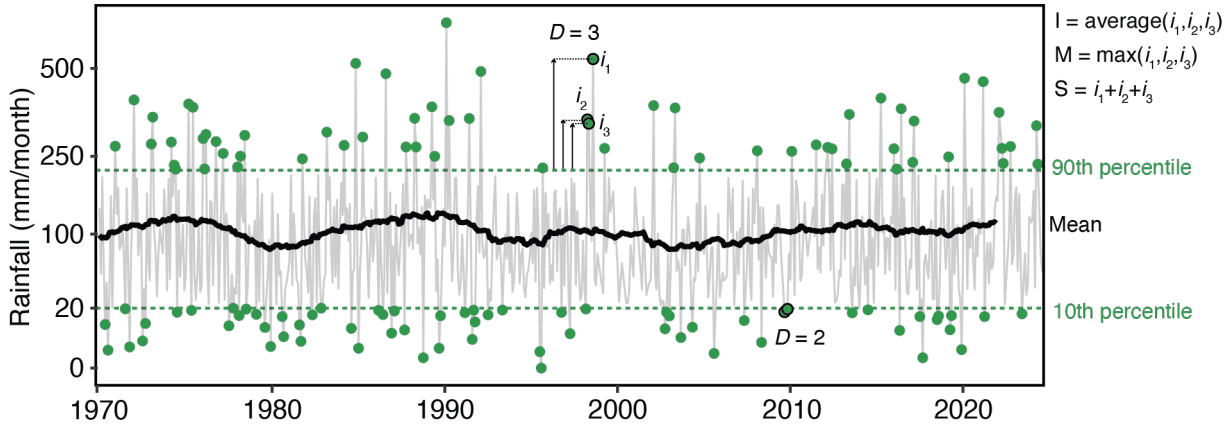


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1473 **Fig. 1 | Overview of the landscape hydrology and animal hydroregulation.** Blue text indicates environmental water  
 1474 that can influence hydroregulation such as precipitation and evapotranspiration, atmospheric and soil moisture content,  
 1475 water bodies (outlined in Table 1), and their interaction with external factors such as wind speed, temperature, thermal  
 1476 radiation, and substrate composition. The landscape includes habitats with different water stressors represented by vapour  
 1477 pressure deficit (VPD in kPa), which is calculated from measured air temperature ( $^{\circ}\text{C}$ ) and atmospheric moisture content  
 1478 (e.g. relative humidity in %). Hydroregulation includes water gain/loss, water storage and their interaction with extrinsic  
 1479 and intrinsic factors. Representative landscape and animals are based on Borneo's ecosystem. The representative  
 1480 terrestrial lizard is the earless monitor lizard (*Lanthanotus borneensis*), the representative arboreal frog is the Wallace  
 1481 flying frog (*Rhacophorus nigropalmatus*), and the representative subterranean caecilian is the Metang caecilian  
 1482 (*Ichthyophis biangularis*). Illustration by S. Buttmer.

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Sydney, Australia  
Humid subtropical climate (1970–2024)

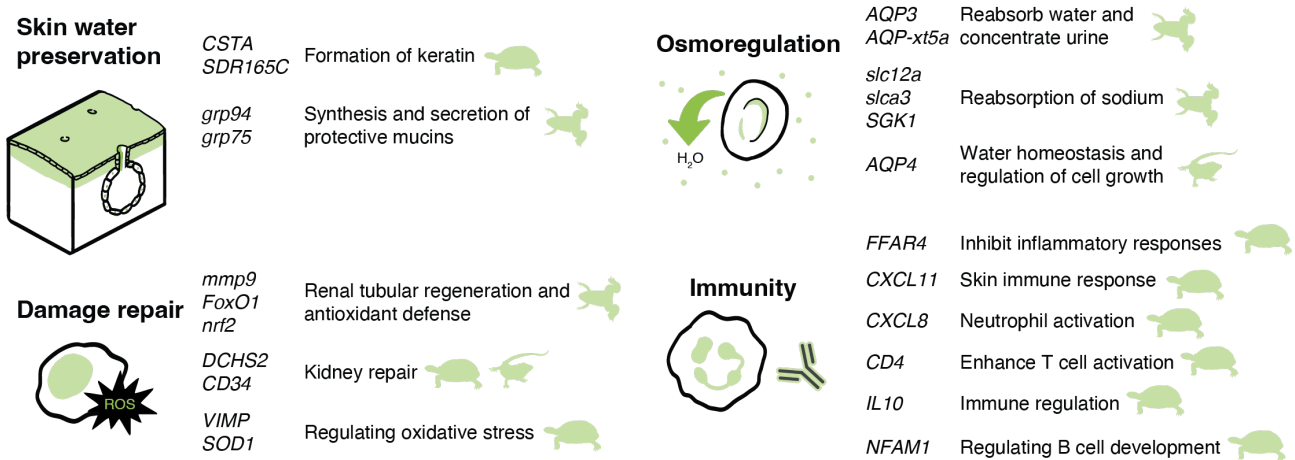


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1485 **Fig. 2 | Example variation in monthly precipitation in Sydney, Australia from 1970–2024.** Monthly precipitation  
 1486 data represented by thin grey lines from the Australian Government Bureau of Meteorology, with the 5-year rolling  
 1487 mean in thick black lines. The 10<sup>th</sup> and 90<sup>th</sup> month-specific percentiles represent dry and wet thresholds, respectively.  
 1488 Example durations ( $D$ ) for extremely dry (red points) or wet (blue points) months are shown which is calculated as the  
 1489 number of months consecutive months above the wet and below the dry thresholds. Example calculations of intensity  
 1490 ( $I$ ), magnitude ( $M$ ), and severity ( $S$ ) is also shown for a 3-month wet event ( $D = 3$ ) with a departure of  $i_1$ ,  $i_2$  and  $i_3$   
 1491 from the threshold. Frequency can be calculated as the number of times the monthly precipitation is above the wet and below  
 1492 the dry thresholds.

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1496 **Fig. 3 | Example genes identified in response to water stress.** Genes are grouped by the following functions: skin  
 1497 water preservation, osmoregulation, damage repair, and immunity. Specific functions of the genes are described on the  
 1498 right side with the taxon in which the function has been demonstrated.

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