

No evidence for assortative mating in the Atlantic puffin

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46

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49

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52 images, statistical analyses, and writing the manuscript.

53 **Abstract**

54 Assortative mating occurs when individuals with similar phenotypes mate together more often than by
55 chance and can contribute to increases in homozygosity, linkage disequilibrium between loci, and
56 premating isolation in a phenotypically divergent population. While this phenomenon has been well
57 documented in many avian species, evidence is relatively scarce in seabirds. Most seabirds are long-lived,
58 monogamous, and form multi-year pair bonds, so assortative mating is hypothesized to occur if
59 individuals pair early in life and change predictably over time. Furthermore, most seabirds are sexually
60 monomorphic, which may in part result from mutual mate choice on the same traits. In this study, we
61 looked for evidence of assortative mating in Atlantic Puffins across 11 traits: current body condition,
62 structural size (wing length), bill size (depth and culmen length), bill shape (three measures from
63 geometric morphometric analyses), and bill colouration (four regions). Despite a large sample size, we
64 detected only weak support for non-random pairing. Specifically, we found evidence for positive
65 assortative mating on cere colour, but no other trait under investigation. We discuss these results in the
66 context of poorly understood mate choice behaviours in Atlantic Puffins and argue that determining the
67 individual age of these long-lived animals would help resolve many unanswered questions.

68

69

70 **Keywords:** Seabird, *Fratercula arctica*, visual ecology, mate choice, ornamentation, elaborate
71 monomorphism

72 **Introduction**

73 Assortative mating is the non-random pairing of breeding individuals based on one or more
74 phenotypic traits (Burley, 1983, Jiang et al., 2013). Pairing between individuals with similar trait values
75 (i.e., positive assortative mating) can promote locus homozygosity and linkage disequilibrium between
76 loci, as well as contribute to premating isolation (Coyne and Orr 2004; Janicke et al., 2019). In contrast,
77 pairing between individuals with dissimilar trait values (i.e., negative assortative mating) could increase
78 heterozygosity and limit inbreeding depression (Pusey and Wolf 1996).

79 Correlations between phenotypic trait values such as size, colour, behaviour, and/or diet in mated
80 individuals have been detected in a wide range of taxa including amphibians (Lu et al., 2009; Swierk and
81 Langkilde, 2021), fish (Snowberg and Bolnick, 2008; Ward et al., 2004), and birds (Caro et al., 2021;
82 Indykiewicz et al., 2017; Pryke and Griffith, 2007; Tamin and Doligez, 2021). Because of the
83 mechanisms that lead to non-random pairing, positive assortative mating has been reported more often
84 than negative assortative mating (Jiang et al., 2013; also called “disassortative mating”, Partridge, 1983).

85 Assortative mating often occurs through passive mechanisms, where individuals with similar traits
86 mate based on convenience or availability rather than explicit mate choice (Newton et al., 1981). Indeed,
87 individuals with similar temporal and/or spatial requirements may passively assortatively mate. For
88 example, individuals with similar traits may arrive to the breeding grounds around the same time (Dittrich
89 et al., 2018; Village, 1985; Wiebe, 2000), resulting in population segregation where individuals must
90 choose mates that are similar to themselves (Jiang et al., 2013). This has been well described in migratory
91 species in which high quality breeders are the first to arrive at the breeding site (Blums et al., 2005;
92 Limmer and Becker, 2007; Matyjasiak, 2013) and pair immediately with available mates (Dittrich et al.,
93 2018; Moiron et al., 2020). Territorial defence or site fidelity can also lead to passive assortative mating if
94 individuals with similar traits occupy or prefer the same space (e.g., Ferrer and Penteriani, 2003). For
95 example, ladybird beetle races (*Henosepilachna diekei*) do not mate assortatively under lab conditions in
96 the absence of host plants but exhibit almost total assortative mating by race when presented with a
97 choice to occupy one of two host plant species (Matsubayashi et al., 2013). Similarly, intrasexual

98 competition for nest sites may also lead to passive assortative mating. High quality individuals generally
99 can access and defend higher quality breeding sites, thus attracting similarly high quality mates (Bitton et
100 al., 2008; Harris and Siefferman, 2014). Finally, certain phenotypes change predictably as individuals age
101 (e.g., colour, Bitton et al., 2008; size, Simmons, 1995; behaviour, Class and Brommer, 2016), so positive
102 assortative mating may persist in the population if monogamously mated individuals age together
103 (Woodman et al., 2022).

104 Only mutual mate choice for the same phenotype can be considered an active mechanism of
105 assortative mating (Wiley and Poston, 1996). When phenotypic traits honestly advertise quality and lead
106 to reproductive or survival advantages in both sexes, active assortative mating may occur because higher
107 quality individuals preferentially choose to mate with one another. While it can be challenging to separate
108 passive and active mechanisms solely based on observations of pairing in the wild, experiments have
109 provided strong evidence for mutual mate choice in birds based on various traits. Some of these traits are
110 structural (e.g., tail length in bearded tit *Panurus biarmicus*, Romero-Pujante et al., 2002), but most
111 studies have focused on elaborate, often colourful, ornaments (e.g., crest length, Jones and Hunter 1993;
112 UV colour, Nolan et al., 2010). Therefore, active assortative mating has been most actively sought and
113 found in mutually ornamented species.

114 Atlantic Puffins (*Fratercula arctica*) are elaborate monomorphic seabirds. While sub-adults
115 between age one and three display relatively drab features, adults display a bright red-orange bill, orange
116 rosette, and orange feet during the breeding season. During the winter months, Atlantic Puffins shed their
117 bill sheath and regrow it before the start of each breeding season, such that the colourful bill could reflect
118 nonbreeding condition or serve another signalling role (Lowther et al., 2020). They are long-lived with
119 high adult survival (0.909–0.965 in Sandvik et al. 2008, > 0.915 in five colonies over ~10 years in Harris
120 et al. 2005), are socially and genetically monogamous, form multi-year pair bonds, and obligately share
121 parental care to raise a single chick each year (Anker-Nilssen et al., 2008; Creelman and Storey, 1991;
122 Lowther et al., 2020). Breeding attempts in this species are particularly energetically costly and can
123 negatively impact body condition and survival (Erikstad et al., 2009), so mutual choice for a high-quality

124 partner could be critical to ensure successful breeding (Henderson et al., 2013). However, little is known
125 about pair formation and the criteria by which Atlantic Puffins choose their mates (Lowther et al., 2020).

126 The mutual ornamentation of Atlantic Puffins along with their life-history strategies suggests that
127 assortative mating through passive and/or active mechanisms could be expected. Because of their high
128 adult survival and low divorce rate (~7% in long term study by Harris and Wanless, 2011), most newly
129 formed pairs should be composed of young first-time breeding birds, and pairs should be assortatively
130 mated by age at the population level. Since some aspects of their morphology are expected to change
131 predictably with age (e.g., mass in Bond et al., 2016), age-based pairing should passively lead to evidence
132 of assortative mating in these traits. Other traits known to change as individuals age in avian species
133 include structural size (e.g., longer wing length with age, Piliczewski et al., 2018), bill size and shape
134 (wider bill with age in Alisauskas, 1987; longer with age in Petersen, 1976; Piliczewski et al., 2018), and
135 ornament size (longer/larger with age, Adámková et al. 2022; Evans et al. 2011; Laucht and Dale, 2012).
136 Thus far, Bond et al. (2016) only found weak evidence for assortative mating in Atlantic puffins based on
137 culmen length in a Newfoundland, Canada, population and head + bill length in a New Brunswick,
138 Canada, population. It is unknown if Atlantic Puffins mate assortatively based on other untested
139 morphological traits like bill shape.

140 Alternatively, individuals that are new breeders or have lost their previous mate could actively
141 choose a new mate of high quality among those available. In many study systems, carotenoid-based
142 colouration in plumage and fleshy ornaments provide information on aspects of individual quality such as
143 foraging ability and overall health and condition (Caro et al., 2021; Kirschel et al., 2020). In Atlantic
144 Puffins, bill and rosette colouration has been found to correlate with body condition (Doutrelant et al.
145 2013, but see Kochvar et al. 2024), and chick growth rates (Kochvar and Bitton, 2025). However, no
146 studies to date have searched for evidence of assortative mating in Atlantic Puffins based on the shape
147 and colour of their ornaments.

148 In this study we investigated whether Atlantic Puffins breeding in Newfoundland and Labrador,
149 Canada, mate assortatively. Premised on the two mechanisms described above, we hypothesized that adult

150 puffins would show evidence of positive assortative mating based on body condition, structural traits, and
151 bill characteristics.

152

153 **Methods**

154 **Study site and subjects**

155 This study was conducted on Gull Island in the Witless Bay Ecological Reserve of Newfoundland and
156 Labrador, Canada (47.26N, 52.77W) during the chick rearing phases (July and August) of 2019, 2020,
157 2021. The reserve supports over 300,000 breeding pairs of Atlantic Puffins, 120,000 of which breed on
158 Gull Island (Wilhelm et al., 2015). Our full dataset consisted of 351 individuals including 75 confirmed
159 mated adult pairs, as determined by a genetically sexed male and female occupying the same burrow
160 containing a chick within the same breeding season. All 75 pairs were used to investigate assortative
161 mating by morphometrics, and a subsample of 57 mated pairs were used to investigate assortative mating
162 by bill colouration.

163

164 **Field methods**

165 Individuals were captured in active burrows between the hours of 22:00-03:00. Upon capture, adults were
166 placed inside a small breathable cloth bag and transferred to a banding station to be processed. We banded
167 each puffin with a uniquely numbered Canadian Wildlife Service steel band, measured their weight (600
168 g Pesola spring scale to the nearest 5 g) and wing chord (flat wing ruler to the nearest mm), and collected
169 a small blood sample (less than 75 μ l) from the brachial vein. Puffins were then taken into a blind where
170 ultraviolet (UV) and visual spectrum RAW images (20 megapixel) were taken of the left and right sides
171 of the head with a UV converted Samsung NX1000 (following instructions from Troscianko, 2018) using
172 two 2-inch Baader lens filters (UV: 300-400 nm; visual: 400-700 nm; Figure S1). The puffins were
173 illuminated with a full-spectrum metal halide bulb (150 W; Iwasaki eyeColour MT70D E27 6500K)
174 placed behind a light diffuser to avoid shadows, and all photos included optic grade white (99 %
175 reflectance) and dark (10 % reflectance) standards, a ruler, and a small white board to record the date and

176 band number of the individual. The bill was cleaned of debris with a toothbrush prior to photo capture and
177 was held in place in a standardized position using a custom-built wooden stabilizer (Figure S1). Three
178 pictures in the visible range and three pictures in the UV range were obtained using the camera bracketing
179 function set at -0.5, 0, and 0.5.

180

181 **Molecular sexing**

182 DNA was isolated from blood samples collected in the field on #2 filter paper (Whatman™, Cytiva,
183 Maidstone, UK) and stored at room temperature. A ~1 cm² section of the saturated filter paper was
184 removed with sterilized scissors and placed into a 1.5 ml collection tube. A DNeasy Blood & Tissue kit
185 (Qiagen Inc., Toronto, ON, CA) was used to extract DNA following standard protocols. All samples
186 were stored in a freezer at -20 °C for later use.

187 A polymerase chain reaction (PCR) was run on an Eppendorf Mastercycler® ep gradient S for
188 determining sex of the individuals based on the amplification of the chromo-helicase-DNA 1 (CHD1)
189 gene found on the W and Z chromosomes of birds. This protocol is the standard used for sexing seabirds
190 (Fridolfsson and Ellegren, 1999). For each sample, this included the use of 12.5 µl Thermo Fisher
191 Promega PCR Master Mix, 2 µl each of primers 2550F and 2718R, and 2 µl of extracted DNA. Each
192 batch also included a no-template control. The PCR was run on a program of initial denaturation at 95 °C
193 for 5 minutes, then 35 cycles of denaturing, annealing, and extension at 94 °C for 30 seconds, 50 °C for
194 30 seconds, 72 °C for 60 seconds, followed by a final extension at 72 °C for 7 minutes and a cooling
195 period of 4 °C for 10 minutes. Samples were stored in a research freezer at -20 °C if not used
196 immediately.

197 The amplified samples were run on a RedSafe™ agarose gel (iNtRON Biotechnology, Inc.,
198 South Korea) along with 100 base pair reference ladders and a no template control on a Thermo
199 Scientific™ EC 300 XL for 50 minutes at 130 amps. The gels were then imaged and stored digitally.
200 Individuals associated with one band on the gel were assigned as males and individuals with two bands
201 were assigned as females.

202

203 **Bill size and shape through geometric morphometrics**

204 Bill size measurements were obtained directly from the visual spectrum images of the left side of puffin
205 bills using ImageJ (Rasband, 2021). After setting the scale using the ruler included in the image, we
206 extracted the culmen length (distance from points 2 to 4; Figure 1) and bill depth (distance from points 2
207 to 6; Figure 1). To quantify the shape of the bill we used geometric morphometric methods (Zelditch et al.
208 2012) applied to our complete dataset of 150 females and 201 males. The inclusion of individuals other
209 than the 150 of interest (75 mated pairs) improves the reliability of the metrics extracted by capturing
210 variation that is more representative of the whole population. Basic operations (i.e., file management,
211 positioning of landmarks and semi-landmarks) were conducted using the series of tps software (i.e.,
212 tpsUtil, tpsDig2, tpsRelw; Rohlf, 2015) and multivariate analyses on the landmarks were performed in
213 MorphoJ (Klingenberg, 2011). We scaled the images using the ruler in each photograph and placed
214 landmarks and semi-landmarks at locations on the bill that best represented their geometric form and were
215 easily recognisable and repeatable (Figure 1). Landmarks were positioned at: (1) the point where the
216 forehead meets the uppermost part of the cere, (2) the uppermost edge of the cere, (3) the point where the
217 upper mandibles' orange colouration dissipates into the lamella, (4) the tip of the nail, (5) the point where
218 the mandibular tomium straightens laterally after its curvature from the nail, (6) the most proximal point
219 of the lower mandible, (7) the commissure, (8) the lowermost part of the cere, (9) the closest point of the
220 rosette to the cere, and (10) the outermost dorsal and (11) outermost posterior edge of the nare (Badikova
221 and Dzerzhynsky, 2015; Lockley, 1962; Petersen, 1976). Semi-landmarks were positioned: (12-14) along
222 the curve of the upper mandible, (15) at the most concave and (16) convex area of the lower mandible, at
223 the middle of the (17) lower and (18) upper lamella, (19) the middle of the cere, and (20) where the
224 lowest point of the lamella meets the cere (Badikova and Dzerzhynsky, 2015; Burnham et al., 2020).

225 Raw landmark coordinates were made comparable using tpsRelw software by removing the
226 variation in digitizing orientation, location and scale, and superimposing them in a common coordinate
227 system (Adams et al., 2004). Distances between landmarks were treated as multivariate data and

228 processed with a Procrustes analysis in MorphoJ (Adams et al., 2004), which finds the transformations
229 that best superimpose the coordinates of all individuals. This was followed by a principal component
230 analysis (PCA) to assess the magnitude of shape variation among the landmarks. The resulting PCs
231 described shape variation with values near zero associated with the average shape of the bills, and high
232 values, both negative and positive, associated with strong deviations from the average shape (see Results
233 for details, Klingenberg, 2016; Slice, 2007).

234

235 **Bill colour analysis**

236 For each individual, a multispectral image of the left side of the bill was generated using the plugin
237 micaToolbox version 2.2 in ImageJ (Troscianko and van den Berg, 2021; van den Berg et al. 2020). One
238 visual spectrum and one UV RAW photo were chosen for alignment using the ‘Photoscreening’ tool,
239 selecting the most illuminated sample without evidence of RGB channel oversaturation. Visual spectrum
240 and UV images were aligned and merged with the ‘Affine align’ tool by selecting four landmarks along
241 the bill of each image. To evaluate the success of the alignment, we created a false colour image with the
242 ‘Make Presentation Image’ tool, using yellow for the normalised red channel and blue for the normalised
243 uvB channel. False colour images with little to no blue colouration visible around the bill were deemed
244 effectively aligned and suitable for analysis. From the aligned multispectral images, we generated cone
245 catch images using the spectral sensitivity of a violet-sensitive (VS) avian visual system with the peak
246 sensitivity of the four photoreceptors set at 410 nm (SWS1), 450 nm (SWS2), 505 nm (MWS), and 565
247 nm (LWS) (Endler and Mielke, 2005; Ödeen et al., 2010), and standard illuminant D65 as a typical
248 ambient light spectrum. While no data on the photoreceptor sensitivities of Atlantic Puffins are available,
249 closely related species (i.e., common murre *Uria aalge* and razorbills *Alca torda*), do not possess a UV
250 sensitive photoreceptor (based on SWS1 sequencing, Ödeen et al. 2010).

251 The average quantum catch values for four regions of interest (ROIs) on the bill were extracted
252 from the cone catch images, each of which consisted of a 30x30 pixel circle. The locations of the four
253 ROIs included the tips of the upper and lower mandible, the semi-fleshy cere, and the fleshy rosette

254 (Figure 1). The colours represented by the average quantum catch values from the ROIs were modelled in
255 a tetrachromatic colourspace to obtain Cartesian coordinates (x,y,z) with the ‘colspace’ function in the R
256 package pavo (Maia et al., 2013, 2019).

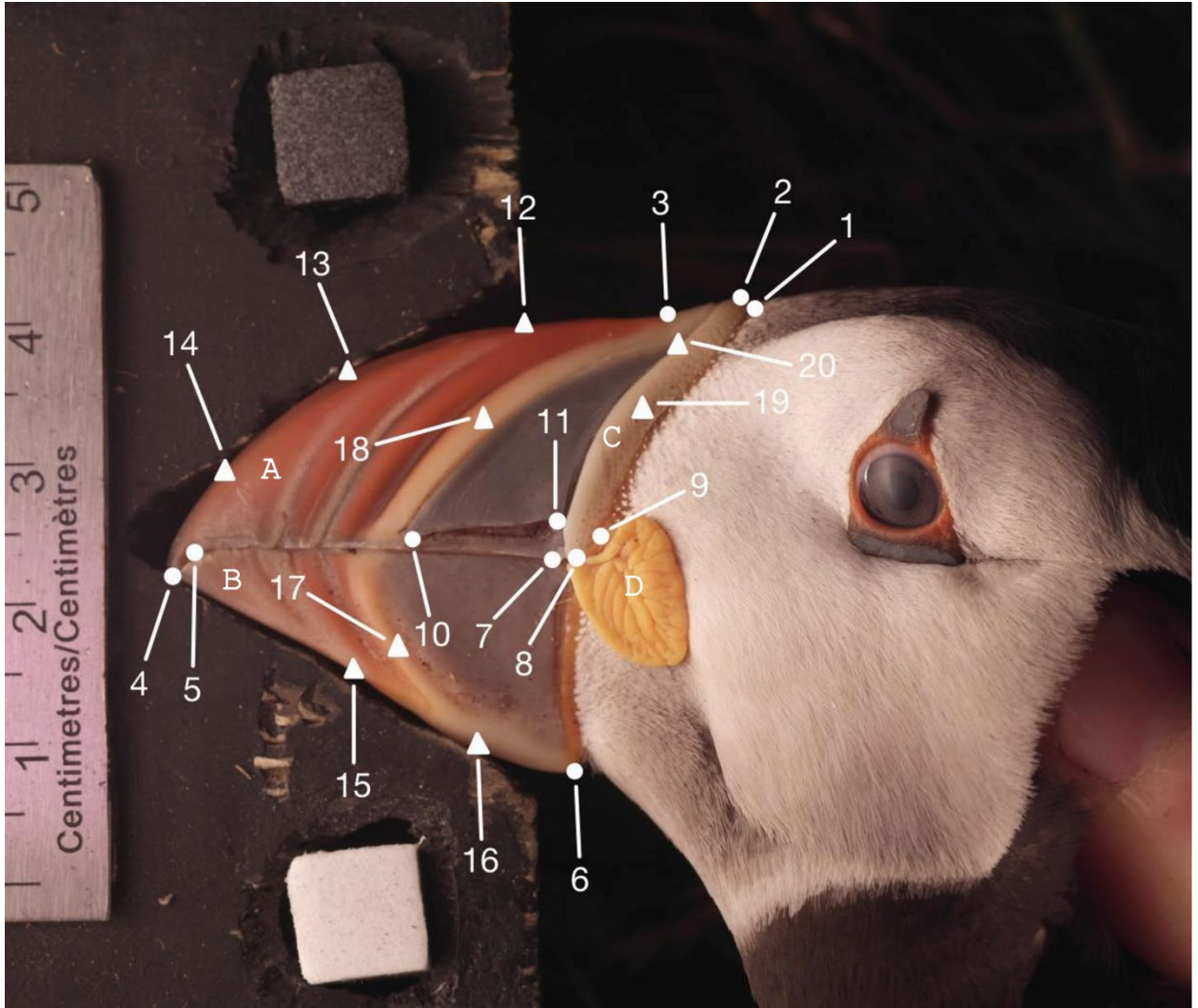
257

258 **Statistical analysis**

259 The condition of a bird is often calculated using a major axis regression of mass over size (Labocha and
260 Hayes, 2012). However, the mass of Atlantic Puffins is different between the sexes (Lowther et al. 2020)
261 and decreases over the course of the breeding season (unpublished data). Therefore, we calculated an
262 index of condition using ordinary least squares regression residuals obtained from a linear model with
263 mass as the dependent variable and sex, Julian date, and wing chord length as independent variables. We
264 initially included all two-way interactions in a full model and performed stepwise removal of non-
265 significant terms to achieve a final model. We assessed the assumptions of multiple regression (no
266 multicollinearity, homoscedasticity, no outliers, linear relation) on the initial full model and final model.

267 For morphological traits, we assessed the presence or absence of assortative mating using Pearson
268 correlations between the male and female trait values (i.e., direct measurements on the bill, principal
269 components from the geometric morphometric analyses, and body condition). To assess assortative
270 mating based on colouration, we first assessed the pairwise differences between the colours of mated
271 individuals by calculating the Euclidean distance between their Cartesian coordinates in a tetrahedral
272 colourspace. We calculated this separately for each ROI to assess the similarity in colour between mated
273 individuals across different regions. We compared the mean Euclidean distance between mated
274 individuals to a null distribution of expected mean values generated by randomly pairing males and
275 females over 1000 iterations. We determined whether assortative mating occurred based on the position
276 of the observed mean distance compared to the null distribution; positive assortative mating was defined
277 as observed values smaller than the 25th smallest value in the null distribution (i.e., two-tailed test at $p =$
278 0.05), and negative assortative mating was defined as observed values greater than the 25th largest value

279 in the null distribution. P values were then adjusted with the false discovery rate correction to reduce the
280 probability of type 1 error (Benjamini and Hochberg 1995).



281

282 Figure 1. Exemplar puffin bill image with landmarks (solid circles) and semi-landmarks (solid triangles)

283 for geometric morphometric analysis. Points 2 and 4 were used to calculate bill length and points 2 and 6

284 were used to calculate bill depth. Letters A through D represent regions of interest from which colour data

285 were obtained: A) upper mandible, B) lower mandible, C) cere, and D) rosette.

286

287 Results

288 Our sample size for assortative mating analyses was determined by identifying all the mated pairs of

289 puffins sampled within the same year, resulting in an initial sample size of 84 pairs. Pairs that were

290 sampled in multiple years were only used once in the analysis. In cases where one member of the pair

291 changed in a subsequent year, both pairs were included in the analysis. Additional pairs were excluded
292 from the analyses because one or both individuals were missing a blood sample (4 pairs) or missing
293 photos (2 pairs), as well as in unusual cases where three adults were sampled from the same burrow (2
294 pairs) or two individuals of the same sex only were found (1 pair). After the removal of these individuals,
295 the final sample size was 75 pairs for the morphological analysis and 57 pairs for colour analyses
296 (available UV and VIS images).

297

298 **Body condition**

299 All main effect variables (i.e., sex, wing length, Julian date), but none of the interactions, were retained in
300 our final model of body condition (adjusted $R^2 = 0.56$; Table S1). Mass was significantly greater in males
301 than females (parameter estimate: 39.94 ± 3.77 SE, $F_{1,145} = 112.11$, $p < 0.001$), in individuals with longer
302 wing lengths (parameter estimate: 1.55 ± 0.42 , $F_{1,145} = 13.41$, $p < 0.001$), and in individuals sampled
303 earlier in the breeding season (parameter estimate: -0.50 ± 0.17 , $F_{1,145} = 8.40$, $p = 0.004$). We retained the
304 residuals of the full model as the index of the individuals' body condition.

305

306 **Geometric morphometrics**

307 The first seven principal components retrieved from the Procrustes analysis captured 83 % of the
308 variation in bill morphologies. However, the first three, which captured over 50 % of the variation, were
309 the most easily interpretable based on a visual inspection of the mesh planes (Figure S2, S3, and S4).

310 The first principal component (PC1; Figure S2) identified variance in the relationship between the
311 depth and culmen length of the bill: individuals with large negative values had a relatively long and
312 shallow bill while individuals with large positive values had relatively short and deep bills. The second
313 principal component (PC2; Figure S3) identified differences in the way the lamella converged on the cere:
314 individuals with large negative values displayed a thick connection between the lamella and the cere
315 whereas individuals with large positive values had a thin (if any) connection between the lamella and the
316 cere. Finally, the third principal component (PC3, Figure S4) pointed to differences in the posterior

317 curvature of the bill (where the bill meets the face): individuals with large negative values displayed a
318 relatively straight posterior bill curvature, whereas individuals with large positive values had a more
319 pronounced posterior bill curvature.

320

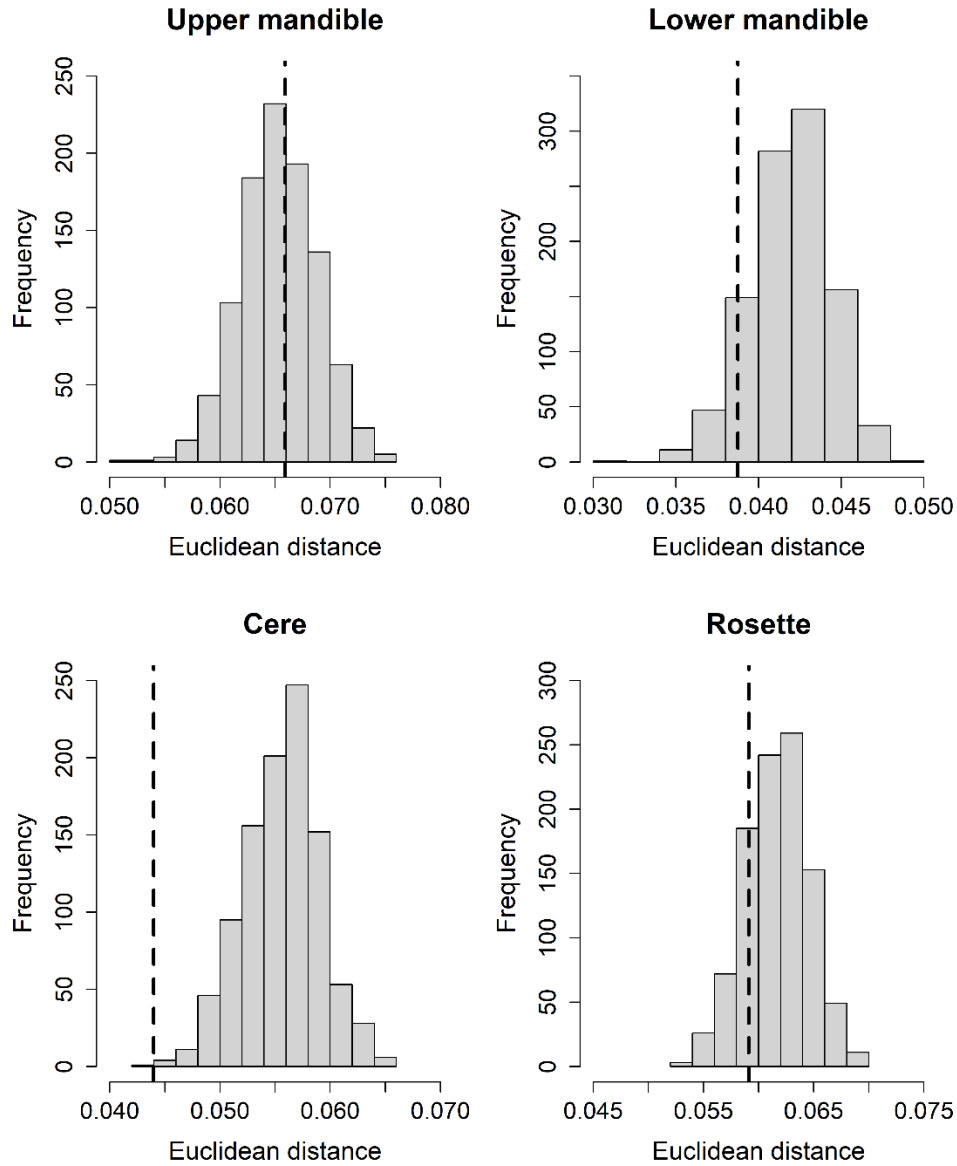
321 **Assortative mating by morphometrics**

322 We did not find any evidence for assortative mating based on the morphological traits measured. Mated
323 individuals did not pair according to body condition ($n = 74$, $r = -0.065$, 95% CI: $-0.289 - 0.166$, $p =$
324 0.58), body size (wing length: $n = 74$, $r = -0.346$, 95% CI: $-0.255 - 0.202$, $p = 0.81$), or bill size (bill
325 depth: $n = 75$, $r = -0.130$, 95% CI: $-0.346 - 0.100$, $p = 0.27$; bill length: $n = 75$, $r = -0.087$, 95% CI: -0.308
326 $- 0.143$, $p = 0.46$). We also did not find evidence for assortative mating based on the three retained
327 descriptors of bill shape (PC1: $n = 75$, $r = 0.103$, 95% CI: $-0.127 - 0.322$, $p = 0.38$; PC2: $n = 75$, $r =$
328 0.063 , 95% CI: $-0.167 - 0.285$, $p = 0.59$; PC3: $n = 75$, $r = -0.197$, 95% CI: $-0.406 - 0.031$, $p = 0.09$).

329

330 **Assortative mating by bill colour**

331 We found evidence for positive assortative mating by cere colour only. The mean distance between male
332 and female cere colour was 0.046 (unitless), significantly below the mean of the null distribution ($0.056 \pm$
333 0.003 SD, $p = 0.012$; Figure 2). For all other regions, there was no sign of positive or negative assortative
334 mating (upper mandible: observed mean = 0.060; null distribution mean = 0.062, $p = 0.67$; lower
335 mandible: observed mean = 0.039; null distribution mean = 0.042, $p = 0.31$; rosette: observed mean =
336 0.057 ; null distribution mean = 0.061, $p = 0.31$; Figure 2).



337

338 Figure 2. Pairs of Atlantic Puffins mated assortatively by cere colour, but not by any other bill colour
 339 trait. Each panel presents a null frequency distribution of Euclidean colour distances between randomly
 340 paired males and females ($n = 1000$ iterations) in a tetrahedral colourspace. Smaller distances indicate
 341 greater similarity between the colours. The dashed line marks the observed colour distance between
 342 mated individuals.

343 **Discussion**

344 Evidence of assortative mating has been found in a few Alcid species, including Lesser Auklets [*Aethia*
345 *pusilla*; plumage colour (positive), tarsus length (positive), and facial plume size (negative); Jones and
346 Montgomerie, 1991], Crested Auklets [*Aethia cristatella*; crest size (positive); Jones and Hunter, 1993],
347 and Little Auks [*Alle alle*; wing length (positive), extent of white area on upper eyelid (positive),
348 hormonal stress response (positive); Wojczulanis-Jakubas et al., 2018]. Despite a relatively large sample
349 we found only weak evidence for assortative mating. Of the 11 characteristics we explored, we found no
350 relationships between male and female morphology (bill size and shape, wing length), body condition
351 (corrected for covariates), or bill mandible and rosette colours. Only the colour of the fleshy cere was
352 significantly correlated between mated individuals. Thus, our hypotheses predicting passive or active
353 assortative mating are inconsistent with our findings. More detailed investigation of mating behaviours
354 across the lifespan is necessary to understand reproductive strategies and emergent mating patterns in this
355 species.

356 Because of the low adult mortality rate and low reported divorce rate in this species, we
357 hypothesized that Atlantic Puffins pairing for the first time would do so mostly with other young
358 individuals. Furthermore, because puffins form life-long pair bonds, we hypothesized that these mated
359 pairs would age together. Detecting assortative mating under these conditions would further require that
360 traits predictably change over time. One or all premises of this passive assortative mating mechanism
361 could be incorrect. Adult puffins disperse to multiple locations during the winter, and individuals that
362 breed together do not necessarily overwinter at the same sites (Guilford et al., 2011, Burnham et al., 2021,
363 Baran et al., 2022, Charrier et al., 2024). While the precise mechanism of how previously mated
364 individuals find each other at the breeding grounds is unclear, some have hypothesized that they reunite
365 on the water close to the colony (Haris and Wanless 2011). When, where, and how individuals pairing for
366 the first time initially encounter and court is undocumented (Lowther et al., 2022). Individuals that have
367 lost their mate through mortality or divorce could favour a breeding location, potentially at or near their
368 previous burrow, rather than actively seek a mate that displays specific traits. Fewer than 0.3 % of tracked

369 breeding adults have been found to reneest further than 100 m away from a previously used burrow (Harris
370 and Wanless, 2011). Furthermore, an annual mortality rate of ~ 3 % combined with a divorce rate of ~ 7
371 % would lead to 10 % reshuffling of relationships every year, which could be sufficient to eliminate
372 evidence of assortative mating even with the initial pairing of young individuals. Whether or not
373 morphological traits change predictably with age is also uncertain. While the size and shape of the bill of
374 Atlantic Puffins has been found to change in an expected way over the first six years of life (Petersen
375 1976), this period does not include most adults of breeding age considering their long lifespan (oldest
376 North American puffin aged at 33 years; USGS, 2023). Subsequent research using repeated measures
377 analyses on birds of known age did not find evidence of age-related change continuing later in life (Bond
378 et al. 2016). If morphological traits do not change predictably in older adults, assortative mating on these
379 traits would not necessarily be maintained after initial age-based pairing.

380 Our second hypothesis was premised under the mutual mate choice paradigm and predicted
381 assortative mating on traits typically associated with condition and/or quality in individuals. While
382 Atlantic puffin bill colouration is dynamic within the breeding season (Kochvar et al., 2024), evidence
383 that bill colour is correlated with condition is mixed (no relationship, Kochvar et al., 2024; positive
384 relationship stronger in female than males, Doutrelant et al., 2013). Moreover, there is evidence that
385 female colouration, but not male colouration, is positively related to offspring growth rate constants (mass
386 and 10th primary; Kochvar, 2025). For assortative mating to be achieved through mutual mate choice, the
387 same traits need to signal some measure of quality in both males and females (Johnstone, 1997), and most
388 pairings at the population level should occur relatively synchronously. Of the 11 traits we evaluated, only
389 cere colouration was more similar in mated pairs than expected by chance. In females only, cere hue has
390 been associated with offspring mass at fledging (Kochvar, 2025) but was not associated with body
391 condition in either males or females (Kochvar et al., 2024). The opportunity for mutual mate choice on
392 this trait seems limited, but bare part ornament colouration such as the cere and rosette are dynamic and
393 honest signals of current condition in many species (Iverson and Karubian 2017). Since our
394 measurements were obtained from adults caring for nestlings, measurements earlier in the breeding

395 season when pairing typically occurs may provide stronger evidence for mutual mate choice. Basic
396 studies of mating behaviours when Atlantic Puffins first arrive at the breeding grounds are needed to
397 better understand the factors that influence individual pairing.

398 Several of the premises underlying our hypotheses could be resolved with an accurate way to age
399 Atlantic Puffins. For instance, we could assess if paired individuals are generally of similar age and
400 whether divorced individuals choose a mate of similar age. Combined with our multispectral bill images,
401 it would also be possible to determine if the traits we measured (e.g., bill shape, size, and colour)
402 predictably change after the age of six. Ultimately, this information would give us insight into the
403 breeding ecology and mating behaviours of Atlantic Puffins and potentially clarify why there is no
404 evidence of assortative mating in this species.

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