

***AmphiTherm*: a comprehensive database of amphibian thermal tolerance and preference**

Authors

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Abstract

Thermal traits are crucial to our understanding of the ecology and physiology of ectothermic animals. While rising global temperatures have increasingly pushed research towards the study of upper thermal limits, lower thermal limits and thermal preferences are essential for defining the thermal niche of ectotherms. Through a systematic review of the literature in seven languages, we expanded an existing database of amphibian heat tolerance by adding 1,009 estimates of cold tolerance and 816 estimates of thermal preference across 375 species. *AmphiTherm* is a comprehensive and reproducible database that contains 4,899 thermal trait estimates from a diverse sample of 659 species (~7.5% of all described amphibians) spanning 38 families. Despite its broad geographic coverage, we report evident gaps across amphibian biodiversity hotspots in Africa, most regions of Asia, central South America, and Western Australia. By providing a more holistic understanding of amphibian thermal tolerance and preferences, *AmphiTherm* is a valuable resource for advancing research in evolutionary biology, ecophysiology, and biogeography of amphibians, offering insights that are increasingly needed in changing climates.

Background & Summary

Thermal trait data are crucial to our understanding of the biology and physiology of ectotherms. The recent increase in broad-scale syntheses of ectotherm thermal physiologies demonstrates recurring interest in how these organisms respond to changing thermal environments^{1–10}. Much of this work has focused on traits relating to heat tolerance, reflecting the urgency to predict the impacts of climate warming on natural populations^{11–17}. However, climate change also brings an increased probability of extreme weather events, including negative temperature anomalies^{18,19}. As such, a sole focus on heat tolerance provides an incomplete picture of ectotherm responses to climate change. A comprehensive understanding of heat tolerance, cold tolerance, and thermal preference is necessary to fully define ectotherm's thermal niches and predict their responses to climate change. Below, we briefly emphasise the importance of these thermal traits for amphibians (see ^{8,20,21} for more in depth discussion).

While the significance of heat tolerance in predicting species' responses to warming climates is well documented¹, data on lower thermal limits are equally vital yet often understudied, especially in amphibians, an at-risk, data-deficient group of ectotherms²². Lower thermal limits represents the lower boundary of an organism's thermal niche and have been included in several data syntheses^{5,6,8,10}. This trait provides key insights into how species might respond to increasing frequency of extreme cold weather events, which can lead to significant population reduction events known as winterkills^{18,19,23}. Gaining understanding of lower thermal limits can thus help predict the sensitivity and resilience of amphibian populations to extreme cold weather events. Moreover, data on lower thermal limits can inform conservation and management strategies, for instance, by identifying microhabitats buffering the effects of extreme cold on activity and survival²⁴.

Preferred body temperatures reflect the temperature optimising overall performance and the most favourable microhabitat in the absence of other biotic and abiotic factors^{25,26}. Knowledge of preferred body temperatures can thus help predict how climate change will affect species distributions and activity patterns^{24,27–29}. In particular, thermal preference data can be used to infer behavioural thermoregulation patterns and the microhabitats available for crucial physiological processes^{30–32}. While upper thermal limits can help predict acute survival in the face of extreme heat, gradual warming below the upper thermal limit thresholds can make some areas unsuitable for amphibians' activity needs²⁴. On the contrary, warming can benefit some amphibians in historically cooler climates by increasing activity windows or reducing hibernation times³³. As such, thermal preference data can help predict the sublethal effects of climate change. Thermal preferences also, for instance, affect susceptibility to pathogens^{34–36}, shape the composition of commensal microbes^{37,38}, and mediate interactions between host and microbial communities³⁹.

Although investigating thermal traits separately provides important knowledge, the study of a combination of thermal traits provides deeper and more comprehensive insights. A simultaneous analysis of upper and lower thermal limits is particularly interesting as it provides an estimation of thermal tolerance breadth²¹—a measure of the thermal envelope ectotherms can occupy in the absence of other abiotic or biotic factors (e.g., competition, resource availability). When thermal preference is integrated with upper and lower thermal limits, the thermal envelope gains shape, providing additional insights to parameterise models and predict activity and survival in changing environments. For instance, leveraging data on thermal limits and preferred temperatures can help infer past, current, and future distributions of ectothermic species, including assessing potential invasion risk^{15,40–43}. Parametrising biophysical models with data on thermal limits and preference now also allow more accurate predictions of overall performance, activity windows^{17,44} and microhabitat heterogeneity^{17,44,45}, strengthening our ability to predict the impacts of climate change on natural populations^{46,47}. From an

108 evolutionary perspective, the integration of different thermal traits can also advance our understanding
109 of the (co)evolution of these traits, and how climate change may shape evolutionary pressures on
110 thermal tolerance and preference^{6,29,48–51}.

111 Thermal trait data can also be used to inform conservation efforts. Comparing thermal niche envelopes
112 among amphibians, their microbiota, and potential pathogens can help predict changes in the
113 microbiome and disease risk^{52–57}, providing key information on the environment component of the host-
114 pathogen-environment interactions emphasized with the disease triangle concept⁵⁴. Amphibians are
115 often accessioned into captivity to establish assurance colonies for breeding and eventual
116 reintroduction⁵⁸. Knowledge of the thermal tolerance and preference of a broad range of species from
117 different habitats can help inform the design of enclosures that better simulate natural thermal
118 variability^{59–61}. It is well established that temperature influences captive breeding success of
119 amphibians⁶⁰. Amphibian husbandry guides recommend setting thermal environments near the
120 preferred temperature of a species, and when practical, providing thermal gradients and thermal
121 cycling^{60,62}. Exposure to environmental conditions via “soft release” or mesocosms prior to
122 reintroduction can also influence survival and success of reintroduction efforts^{63,64}. In addition,
123 understanding thermal constraints on activity, demography, and disease risk enhances our ability to
124 identify habitats suitable for repatriation and reintroduction efforts in endangered species^{58,65,66}.

125 Therefore, a holistic understanding of amphibian thermal tolerance limits and preferences is essential
126 for defining the fundamental thermal niche of ectotherms and to project their activity, distribution and
127 survival in rapidly changing environments. Here, we expand an existing database on amphibian upper
128 thermal limits¹, which has already facilitated global assessments of vulnerability to climate warming¹⁷.
129 We conducted systematic searches and aggregated lower thermal limits and thermal preference data
130 from the global literature published in seven languages. In doing so, we expanded thermal niche data
131 for 378 species, providing a stronger foundation for research on amphibian ecology, evolution, and
132 conservation.

133

Methods

Reporting

We reported the contributions of each author using the CRediT (Contributor Roles Taxonomy) statement⁶⁷, and MeRIT (Method Reporting with Initials for Transparency) guidelines⁶⁸. We also followed recommendations to maximise the indexing of titles, abstracts, and keywords in databases⁶⁹.

Literature Searches

We adapted methods from the previous version of the database on amphibian upper thermal limits¹ to search the literature on thermal physiological traits. We aimed to compile a comprehensive and representative sample of the experimental literature on lower thermal limits and thermal preference in amphibians, complementing the data on upper thermal limits compiled previously (see ¹ for methods specific to upper thermal limits). PPottier accessed Scopus, ISI Web of Science (core collection), Lens, and ProQuest (dissertation & theses) on 01 November 2022 using The University of New South Wales' institutional subscriptions (full search strings are available in Supplementary Information (Table S1). Briefly, the search strings were built using as a combination of relevant terms, including “temperature” (and synonyms) and “thermal tolerance or preference” (and synonyms) and “amphibian” (and synonyms) (see Table S1 for details). For studies in English, PPottier modified search strings to accommodate the structure of each database (Table S1) and performed backward searches of previously published reviews of amphibian thermal preference and tolerance^{1,4-6,10}. This resulted in a total of 1676 unique documents. We limited the timespan of our searches to 31 May 2021 to match with the timespan represented in the upper thermal limit data¹. This decision was made to normalise all searches to a single timespan to simplify future database updates. PPottier also performed Traditional and Simplified Chinese, French, Japanese, Portuguese, and Spanish searches in Google Scholar using search strings translated by native speakers (NR, PPottier, PPollo, SN, YY, and RRYO). The searches contained translated singular and plural forms of the following: “amphibian”, “frog”, “toad”, “salamander”, “newt”, “tadpole”, “preferred temperature”, “selected temperature”, “thermal preference”, “Tpref”, “Tsel”, or “CTmin”. Due to search string limitations in Google Scholar (256 characters), each term was assessed in its singular or plural form, and the search producing the largest number of results was selected. We performed searches following the format of (“preferred temperature” OR synonyms) AND (amphibian OR synonyms). We also performed separate searches with “CTmin” and (“Tpref” OR “Tsel”), as these terms are commonly used in the literature to refer to lower thermal limits and preferred temperatures. We (PPottier, NC) opted not to use “thermoregulation” as a synonym for “thermal preference” in our search strategy, as pilot searches revealed that this term often returned studies that did not present experimentally-derived thermal preference values, or studies that were already captured by the other search terms. PPottier used Publish or Perish⁷⁰ to extract bibliographic records from Google Scholar. We also reused studies (designated with ** in Fig. 1) from non-English searches conducted in ¹ as the key terms used successfully retrieved results on cold tolerance. However, we limited the inclusion of studies to those meeting our first two criteria (i.e., studies on amphibians, and published in the targeted language) to reduce the volume of screening. We acknowledge that our searches do not encompass all languages relevant to amphibian thermal physiology research and invite speakers of languages not represented in the current version to contribute to future updates of the database.

Eligibility criteria

We considered studies that empirically tested lower thermal limits or thermal preference in wild or laboratory amphibians. We included studies on larval, juvenile, metamorphic, and adult amphibians, and distinguished studies that acclimated both embryos and larvae to those acclimating single life

stages. We excluded studies solely on embryonic stages due to the lack of comparable methods in embryos—for example, embryos are not capable of movement in a thermal gradient to assess thermal preference, and it is not possible to assess the righting response of embryos. For lower thermal limits, we included studies that measured critical thermal minimum (CT_{min})⁷¹, median lethal temperature (LT₅₀)⁷², or presented data that were convertible to these metrics (e.g. % survival of cohorts tested at different temperatures). We distinguish these metrics in the database. CT_{min} represents the temperature at which a specific physiological or behavioural endpoint is observed—such as the loss of righting response or a lack of response to prodding—when an organism is exposed to progressively decreasing temperatures (e.g., 1°C/min). It does not represent the lowest temperature an organism can tolerate, but rather the onset of functional stasis, the point at which the organism is unable to move and incapable of essential survival behaviours such as thermoregulation or predator evasion⁷¹. This distinction is important, because many ectotherms can recover from temperatures below their CT_{min}. For instance, some species can recover from freezing to later resume normal function⁷³. In contrast, LT₅₀ is the temperature that is lethal to 50% of animals tested and is derived through statistical interpolation from survival rates across a range of temperatures⁷². For thermal preference, we included studies that empirically tested amphibian temperature selection in a thermal gradient or shuttlebox via measures of body temperature (or inferred body temperature from the position in the gradient or shuttlebox). We did not include data reported on amphibian body temperatures from uncontrolled (wild) conditions because available environmental temperatures were not standardised. We only included studies where thermal history (acclimatisation or acclimation temperature) was reported or could be inferred from the dates and coordinates of sampling. Detailed inclusion criteria and decision trees are presented in Supplementary Information (Fig. S1-2, Tables S3-4). PPottier, RRO, PPollo, ANRV, YY, SV, AVL, and NC screened articles for eligibility using Rayyan QCRI⁷⁴. This software facilitated the identification of key terms in titles, abstracts, and keywords to streamline the screening process for large volumes of literature. During data extraction, 47 papers were ultimately excluded for either lacking extractable data or for not complying with our inclusion criteria (43 English, 1 Traditional Chinese, 3 Simplified Chinese). A total of 184 studies were deemed eligible for inclusion in the database. Of these, 157 were identified through formal database searches (comprising one study published in simplified Chinese, and another study in French), while 20 non-English studies and an additional 7 English-language studies were retrieved through Google Scholar (Fig. 1). Therefore, nearly 15% (27/184) of the included studies were retrieved through non-English literature searches. Our literature search methods and screening process is summarised in a PRISMA flowchart⁷⁵ (Fig. 1).

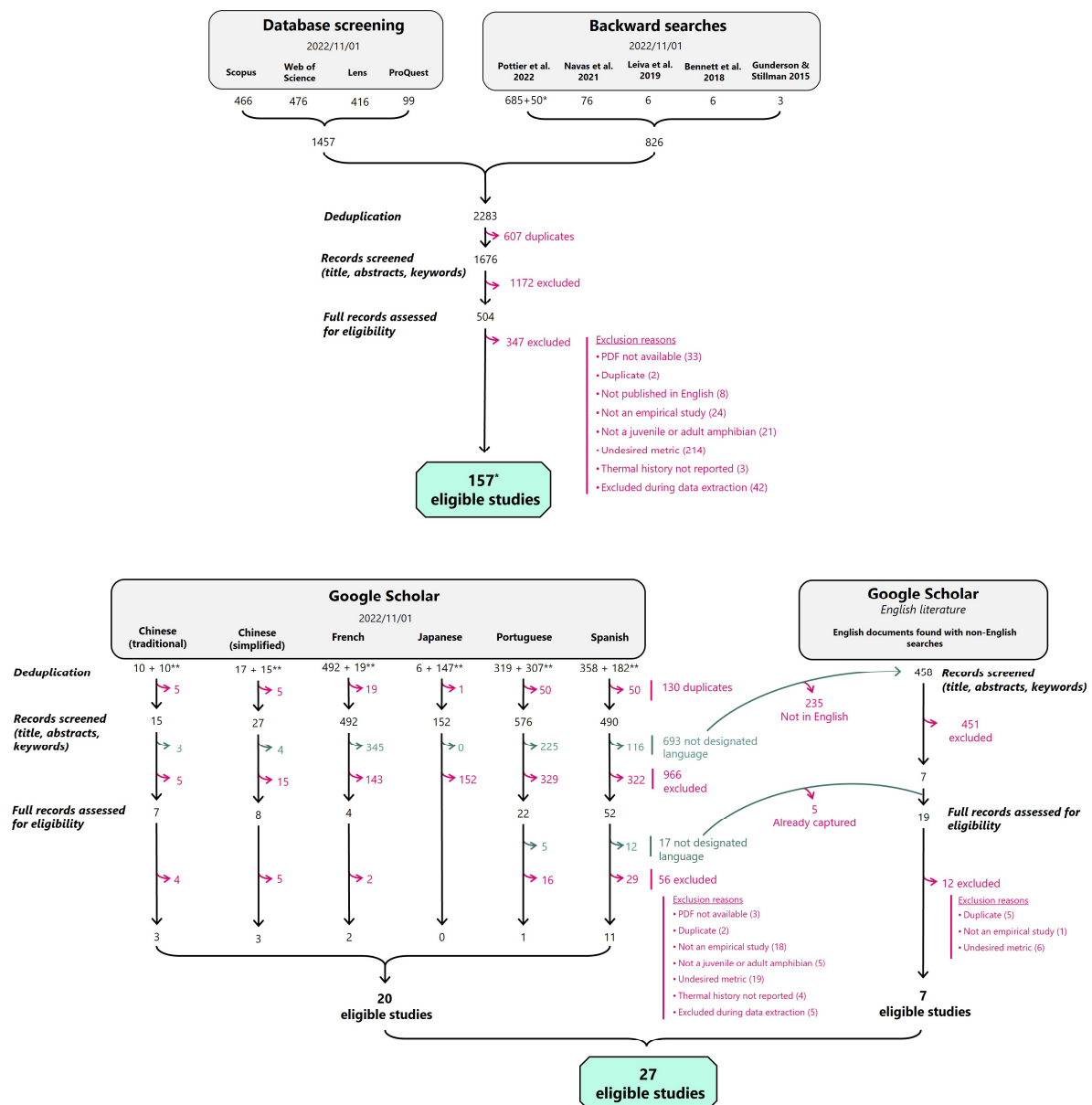


Figure 1 | PRISMA Flowchart delineating the databases used to retrieve studies on lower thermal limits and preferred body temperatures, the number of studies obtained at each stage of the screening process, and the reasons for excluding studies. * Two studies published in languages other than English (French, simplified Chinese) were retrieved through English searches. ** Studies from non-English searches done in Google Scholar from ¹. For the workflow used to obtain data on upper thermal limits, see ¹.

Data Extraction

Data extractions were performed by PPottier (7.7% of estimates extracted), RRYO (6.7%), PPollo (5.3%), ANRV (6.2%), YY (3.0%), AVL (11.0%), SV (20.5%), and NC (45.0%). Note that these values do not add to 100% because some data entries were extracted by two authors. Data were extracted following the protocols described in the previous version of the database¹. We extracted data directly from text and tables, and primarily used *metaDigitise*⁷⁶ (version 1.0.1) in R⁷⁷ to extract data presented in figures (although note that some authors have used WebPlotDigitizer⁷⁸ (version 4.7). When data were available in multiple formats (e.g., text and figure), we extracted it from the format with the highest resolution (e.g. data stratified by sex or location rather than aggregated across species). Where possible, we extracted measures of data dispersion (i.e. standard deviation, standard error) to accompany mean

estimates. In cases where the raw data was available, we calculated summary statistics (means, standard deviation, sample size) to enhance the accuracy of the analysis. For studies reporting survival rates at different temperatures, we predicted the temperature at which 50% mortality occurred using logistic regression from the *dose.p* function from the MASS package⁷⁹.

We also extracted all additional information presented in the studies to allow investigations of the sources of variation in the data and account for non-independence. We assigned identification numbers to each study, and assigned unique identifiers within each study for each estimate, species, population (individuals of the same species sampled from the same geographical location), and cohort (independent group of individuals within a study). Additional variables included sampling coordinates, acclimation temperatures, ramping rates, life stages, endpoints used to infer cold tolerance, or the duration of exposure to experimental treatments. Additional notes were also taken by each researcher extracting the data to facilitate technical validation. The full list of variables is described in Supplementary Information (Table S2). Species names were standardised during the extraction to match AmphibiaWeb⁸⁰ and further standardised to match the most comprehensive phylogenetic tree to date⁸¹ (see *Data Curation*).

Data Curation

To ensure consistency in data extraction across all studies, PPottier and NC extensively reviewed the extracted data to correct typological errors and resolve uncertainties identified during the extraction process. PPottier then curated the data in R⁷⁷ (version 4.4.2), merging the newly extracted data with the previously compiled dataset from ¹. This process involved standardising publication information (publication year, source name) and other variables (e.g., geographical coordinates, IUCN threat status⁸²) to ensure uniformity across both datasets. PPottier also standardised species names and taxonomy with phylogenetic information from ⁸¹, which is primarily based on AmphibiaWeb⁸⁰. The combined dataset comprises 324 publications^{83–406}. Note that 53 of these publications were taken from university dissertations, and some of this work may have now been published^{e.g., 407, 408}.

We also provide a curated version of the database (n = 4,401 estimates), where PPottier excluded data with procedural inconsistencies (e.g., additional stressor, data collected from a single individual), incomplete species information (e.g., *Hyloscirtus sp.*), and studies involving animals exposed to toxicants, hormones, high levels of UV radiations, or infected with a pathogen. A script detailing the data curation steps is available at <https://github.com/p-pottier/AmphiTherm>. This data curation step removed 498 estimates from 15 studies and 45 species. However, we believe that this curated dataset offers broader usability. Nevertheless, we also provide the uncurated version of *AmphiTherm* for users interested in addressing more specific questions (e.g., how toxicants affect thermal tolerance and preference) or identifying existing research gaps within the field.

Data Records

AmphiTherm encompasses 4,899 thermal physiological trait estimates, derived from 324 studies and covering 659 species across 38 families across a broad geographical coverage (Fig. 2-3). This sample represents ~7.5% of all described amphibian species to date⁸⁰ (Fig. 4). According to the IUCN red list⁸², most species (79.2%) are either not threatened or data-deficient (Fig. 2), yet 47 species are classified as near threatened (NC), 43 as vulnerable (VU), 29 as endangered (EN), 14 as critically endangered (CR), and one species now extinct. Considering that over 40% of amphibians are globally threatened²², this suggests that research on amphibian thermal physiology is predominantly conducted on non-threatened species, likely due to the invasive (or lethal) nature of some thermal tolerance

experiments and the associated conservation concerns for threatened species. This database contains substantial within-species variation, with an average of 7.43 ± 19.5 (mean $\pm$ s.d.) estimates per species, spanning a range of 1 to 292 estimates, with species sampled from an average of 2.51 ± 3.29 populations. Approximately 81% of these estimates include a measure of statistical dispersion (standard deviation, standard error), facilitating their use in weighted (meta-)analyses⁴⁰⁹.

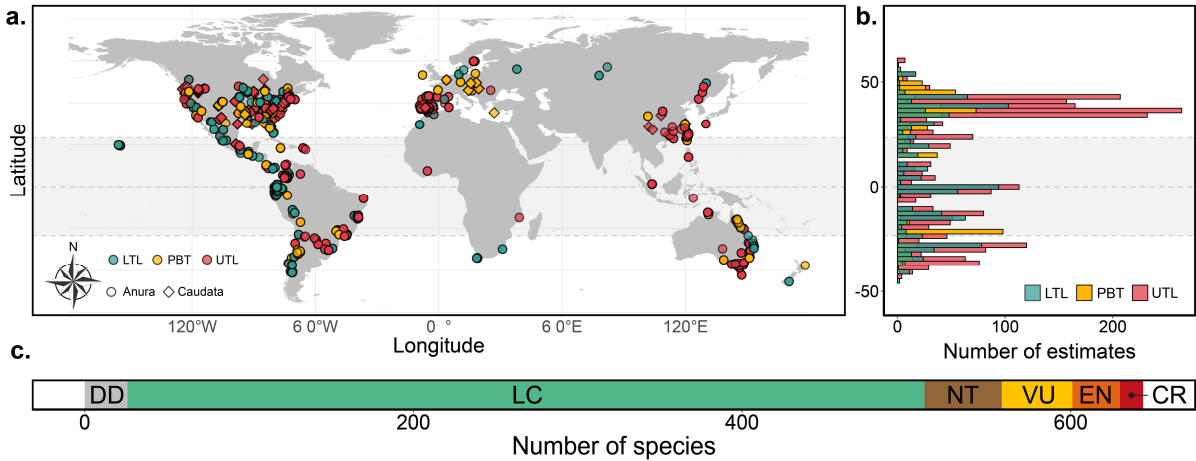


Figure 2 | Geographic distribution of thermal tolerance and preference data. a) World map showing the distribution of lower thermal limits (LTL), preferred body temperatures (PBT), and upper thermal limits (UTL) for anurans (circles) and salamanders (triangles). The shaded area represents the tropics. Note that coordinates were unavailable for 775 (15.8%) estimates. b) Latitudinal distribution of estimates for LTL, PBT, and UTL. c) Threat status of species, classified according to the International Union for the Conservation of Nature (IUCN⁸²). One species (not displayed) is now extinct.

This database update adds thermal data for 375 species, including lower thermal limits for 300 amphibian species and thermal preference data for 137 amphibian species ($n = 1,825$ estimates; Fig 3-4). The majority (98%) of lower thermal limit data are derived from CTmin estimates (990 estimates), with roughly 2% of estimates (19 estimates) derived from lethal limits (LT50). Thermal preference data represent 44% of the database update (816 estimates). This update has a relatively broad phylogenetic coverage, spanning 32 families, with 19.2% of records from salamanders (Fig, 3-4).

Approximately 62.7% of this database is comprised of upper thermal limit estimates (3074 estimates from 616 species and 212 studies; Fig. 3-4), highlighting a significant bias towards responses to heat extremes relative to lower thermal limits (1,009 estimates, 300 species, 88 studies) and thermal preferences (816 estimates, 137 species, 114 studies). We found that only 16 studies measured all three thermal traits, covering 59 species (~9% of the species in the dataset; Fig. 3-4). Upper and lower thermal limits were studied more frequently in tandem (60 studies), allowing to calculate the thermal tolerance breadth (i.e., difference between upper and lower thermal limits) of 276 species (~42% of the species in the dataset; Fig. 3-4).

Geographically, data were collected on all continents where amphibians occur yet exhibit a strong bias towards Nearctic and European regions (Fig. 2). Large geographic gaps in thermal data are evident across Africa, most regions of Asia, Western Australia, and central South America—regions that are biodiversity hotspots for amphibians (Fig. 2). This is particularly concerning as they constrain our understanding of how species from these underrepresented yet extremely diverse regions⁴¹⁰ might respond to climate change. We also identified taxonomic gaps in existing sampling where an entire order of amphibians, Gymnophiona, remained unrepresented in the database (Fig. 4). In addition, 1 of

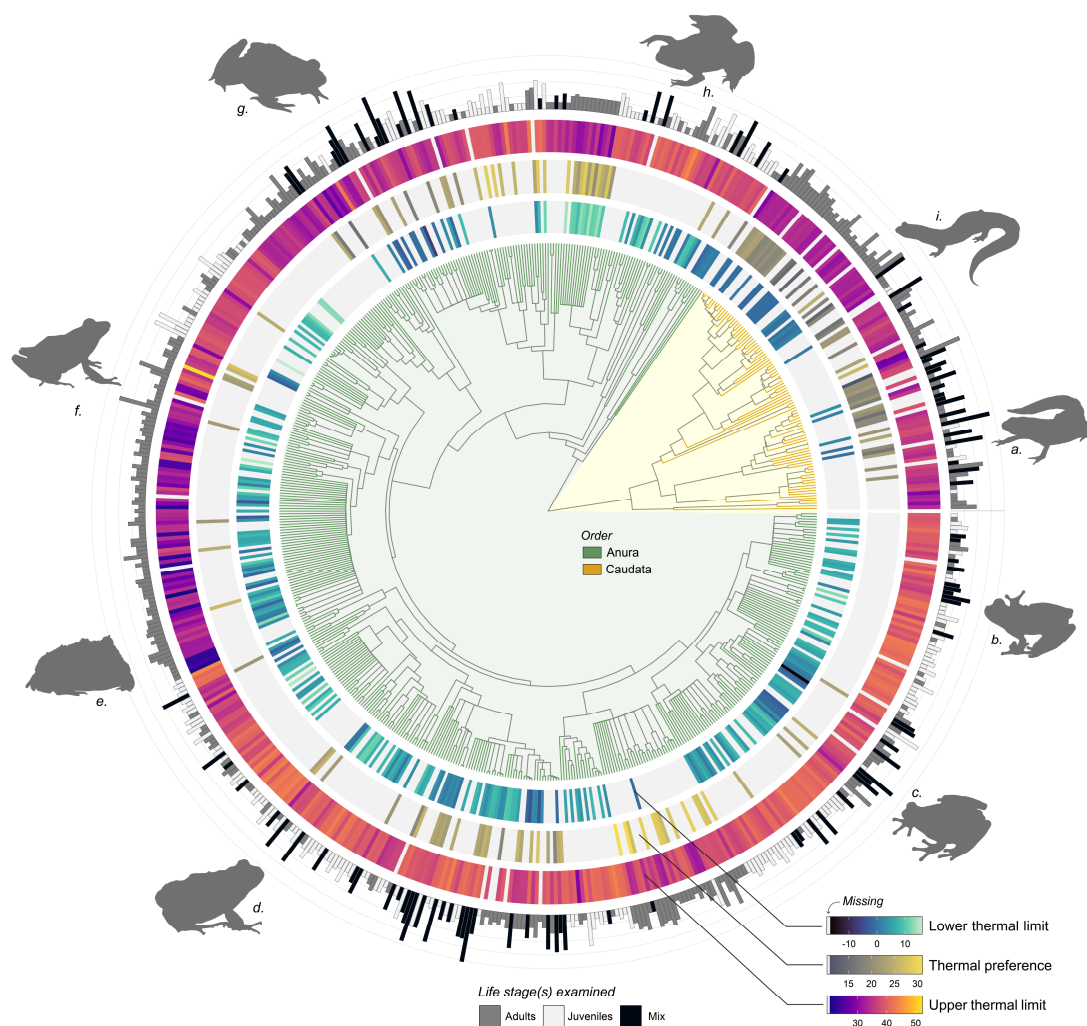
10 families of Caudata and 7 of 36 families of Anura lack thermal limits or preferred body temperature estimates (Fig. 4). This suggests that further efforts are needed to broaden the research scope and better represent the thermal niche of amphibians.

We found that the majority (88.7%) of the literature on amphibian thermal physiological traits was published in English (4,343 estimates). However, non-English language literature contributes a notable and important portion of the knowledge base, accounting for approximately 11.3% of the data. Notably, this includes 289 estimates from publications in traditional Chinese (23 species, 7 studies), 131 estimates from Spanish (40 species, 11 studies), 82 estimates from simplified Chinese (12 species, 10 studies), 28 estimates from Portuguese (10 species, 3 studies), and 26 estimates from French publications (4 species, 3 studies). Including more languages, such as Afrikaans, Arabic, Bengali, Dutch, German, Hindu-Urdu, Korean, Russian, or Swahili in the screening process may help fill some gaps in future updates to the database⁴¹¹. Given the historical bias of higher impact publishing outlets against studies on herpetofauna⁴¹², there are likely a number of studies in non-indexed journals or regional journals in local languages that were not retrieved using our methods.

The *AmphiTherm* database is stored at <https://github.com/p-pottier/AmphiTherm>, and regularly archived to Zenodo⁴¹³. The repository is organised into four main folders: “data”, “R”, “references”, and “shiny_app”. These contain the metadata (.xlsx), raw, cleaned, and curated data (.csv), code for data curation and for producing the figures (.Rmd), code for deploying the Shiny web application (.R), supplementary data (.csv) and phylogenetic tree (.tre) for producing the figures, and bibliographic files (.ris and .bib) with all the references in the database. The metadata file describes all columns (n = 84) of the *AmphiTherm* database in detail. We have also launched a Shiny web application to help navigate and visualise the database. The Shiny app is available at <https://p-pottier.shinyapps.io/AmphiTherm-Explorer/>.

Data records are under a CC-BY license, enabling reuse with attribution. Therefore, database users must cite this study as well as the primary data sources to attribute the original authors and comply with copyright regulations.

We aim to conduct updates at regular five-year intervals, following the same systematic methods, to maintain the database as an up-to-date resource for amphibian thermal envelope data. We encourage researchers who possess relevant thermal data not included in the current version to contact us so that the database can be updated to continuously reflect the most comprehensive and current body of knowledge.



333

334 **Figure 3** | Distribution of mean estimates for three key thermal traits: lower thermal limits (inner heat
 335 map), thermal preference (central heat map), and upper thermal limits (outer heat map). The number of
 336 estimates for each species is displayed as histograms, scaled on a $\log_2(x+1)$ axis for clarity. The
 337 histograms are colour-coded according to the life stage assessed in the experiments, with the category
 338 “juveniles” comprising larvae, metamorphic, and juvenile stages. Gray colour represents missing data.
 339 The phylogeny relationships are based on the consensus tree from ⁸¹. a. *Notophthalmus viridescens*, b.
 340 *Dendropsophus ebraccatus*, c. *Hyla cinerea*, d. *Pleurodema thaul*, e. *Ceratophrys cranwelli*, f.
 341 *Craugastor longirostris*, g. *Rana pipiens*, h. *Xenopus laevis*, i. *Plethodon cylindraceus*.

348 **Technical Validation**

349 We employed a transparent and reproducible workflow to systematically review over 4,000 studies from
350 five databases and across seven languages. The potential limitations of this database are similar to those
351 described in ¹. First, the methods used for indexing and retrieving studies in Google Scholar are not
352 publicly disclosed⁴¹⁴, which may undermine reproducibility. However, given the limited coverage of
353 non-English studies in other databases (with 95% and 93% of references in Scopus and Web of Science
354 indexed in English, respectively), Google Scholar remains one of the most suitable tools to synthesise
355 across multiple non-English languages at present^{415,416}. Second, different authors extracted data from
356 the original studies, introducing the possibility of individual errors. To ensure consistency and accuracy,
357 all extracted data were subsequently cross-checked and standardised by NC and PPottier (see *Data*
358 *Curation*) to minimise the risk of bias and strengthen the reliability of the dataset.

359 **Usage Notes**

360 We anticipate that this database will facilitate a wide range of novel analyses and investigations that
361 may be difficult to foresee at this time, but we are excited to see how these data advance research in
362 amphibian biology and conservation. Our recommendations for using this resource are straightforward:
363 we encourage researchers to have strong foundations in thermal ecology and amphibian biology, and to
364 carefully consider the best approaches for integrating these data into their own investigations.

365 The database represents a comprehensive compilation of studies employing diverse approaches and
366 experimental designs. Given that we cannot anticipate the full scope of research applications, we have
367 made the entire dataset available to allow users to filter and customise the data as needed. We strongly
368 encourage users to clearly document their analytical steps to ensure reproducibility. However, we
369 emphasise that this database version includes data from animals tested under atypical conditions (e.g.,
370 amputations, chemical exposure), or from experiments without replication (e.g. data from a single
371 individual). To accommodate most research needs, we therefore also provide a curated version of the
372 database where we excluded data with procedural inconsistencies, incomplete species-level
373 information, and data involving animals exposed to toxicants, hormones, excessive UV radiation, or
374 pathogens. This curated version of the database is likely more suited for research in ecophysiology,
375 though users with more specialised research questions may find value in the complete dataset. Scripts
376 detailing the data cleaning and curation processes are available at [https://github.com/p-](https://github.com/p-pottier/AmphiTherm)
377 [pottier/AmphiTherm](https://github.com/p-pottier/AmphiTherm) and should provide further guidance for researchers in tailoring the dataset to their
378 specific research needs. We also provide a Shiny web application ([https://p-](https://p-pottier.shinyapps.io/AmphiTherm-Explorer/)
379 [pottier.shinyapps.io/AmphiTherm-Explorer/](https://p-pottier.shinyapps.io/AmphiTherm-Explorer/)) to facilitate data exploration and filtering.

380 As described in the first iteration of this database¹, the data contain inherent sources of non-
381 independence as multiple estimates were extracted from each study, species, population (multiple
382 sampling locations from each species), and cohort (e.g., repeated measures on the same individuals).
383 We recommend that users use phylogenetically-informed statistical models with hierarchical random-
384 effect structures to account for and partition sources of variation^{409,417}. Users should also account for
385 variations in sampling effort (sample size differences), for instance, by weighting estimates by the
386 inverse of their sampling variance⁴⁰⁹. Employing hierarchical models that incorporate sampling
387 variance can help address issues of biological and methodological non-independence, enabling more
388 accurate inferences of the factors driving variation in the data⁴⁰⁹. Most (81%) estimates are associated
389 with a measure of dispersion (standard deviation or standard error), species information is standardised
390 to published phylogenetic information⁸¹, and unique identifiers have been assigned to each study,

species, population, and cohort. These features make *AmphiTherm* readily applicable for use in complex statistical models aimed at uncovering the drivers of thermal tolerance and preferences in amphibians.

As described in previous studies, thermal traits in amphibians are influenced by multiple variables, including acclimation temperature, acclimation time, ramping rate, endpoint metrics, body size, sex, assay duration, and geographic origin, among others^{8,20,21}. We recommend careful attention to these variables, with consideration of incorporating sources of methodological or biological variation as covariates in statistical models, to better capture the complexities of amphibian thermal ecology. Finally, it is important to note that due to the data gaps in hotspots of amphibian diversity, the data herein represent only a subset of total amphibian diversity, and subsequent analyses should acknowledge this limitation. Formally assessing the extent to which geographic and taxonomic biases may influence ecological inferences is an important avenue for research.

Code Availability

The code used to process the data and produce the figures for this manuscript is available at <https://github.com/p-pottier/AmphiTherm>, which is archived permanently in Zenodo⁴¹³.

Author Contributions

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Competing Interests

The authors declare no conflict of interest or competing interests.

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445 **References**

- 446 1. Pottier, P. *et al.* A comprehensive database of amphibian heat tolerance. *Sci. Data* **9**, 600 (2022).
- 447 2. Pinsky, M. L., Eikeset, A. M., McCauley, D. J., Payne, J. L. & Sunday, J. M. Greater
- 448 vulnerability to warming of marine versus terrestrial ectotherms. *Nature* **569**, 108–111 (2019).
- 449 3. Morley, S. A., Peck, L. S., Sunday, J. M., Heiser, S. & Bates, A. E. Physiological acclimation
- 450 and persistence of ectothermic species under extreme heat events. *Global Ecol. Biogeogr.* **28**,
- 451 1018–1037 (2019).
- 452 4. Navas, C. A., Gouveia, S. F., Solano-Iguarán, J. J., Vidal, M. A. & Bacigalupe, L. D. Amphibian
- 453 responses in experimental thermal gradients: Concepts and limits for inference. *Comp. Biochem.*
- 454 *Physiol. B* **254**, 110576 (2021).
- 455 5. Leiva, F. P., Calosi, P. & Verberk, W. C. E. P. Scaling of thermal tolerance with body mass and
- 456 genome size in ectotherms: a comparison between water- and air-breathers. *Phil. Trans. R. Soc.*
- 457 *B* **374**, 20190035 (2019).
- 458 6. Bennett, J. M. *et al.* GlobTherm, a global database on thermal tolerances for aquatic and
- 459 terrestrial organisms. *Sci. Data* **5**, 180022 (2018).
- 460 7. Rohr, J. R. *et al.* The complex drivers of thermal acclimation and breadth in ectotherms. *Ecol.*
- 461 *Lett.* **21**, 1425–1439 (2018).
- 462 8. DuBose, T. P. *et al.* Thermal Traits of Anurans Database for the Southeastern United States
- 463 (TRAD): A Database of Thermal Trait Values for 40 Anuran Species. *Ichthyol. Herpetol.* **112**,
- 464 21–30 (2024).
- 465 9. Bayat, H. S. *et al.* Global thermal tolerance of freshwater invertebrates and fish. Preprint at
- 466 <https://doi.org/10.1101/2024.07.08.602306> (2024).
- 467 10. Gunderson, A. R. & Stillman, J. H. Plasticity in thermal tolerance has limited potential to buffer
- 468 ectotherms from global warming. *Proc. R. Soc. B* **282**, 20150401 (2015).
- 469 11. Urban, M. C. Accelerating extinction risk from climate change. *Science* **348**, 571–573 (2015).
- 470 12. Wiens, J. J. Climate-Related Local Extinctions Are Already Widespread among Plant and
- 471 Animal Species. *PLoS Biol.* **14**, e2001104 (2016).
- 472 13. Carey, C. & Alexander, M. A. Climate change and amphibian declines: is there a link? *Divers.*
- 473 *Distrib.* **9**, 111–121 (2003).
- 474 14. Sinervo, B. *et al.* Erosion of Lizard Diversity by Climate Change and Altered Thermal Niches.
- 475 *Science* **328**, 894–899 (2010).
- 476 15. Anderson, R. O., Meiri, S. & Chapple, D. G. The biogeography of warming tolerance in lizards.
- 477 *J. Biogeogr.* **49**, 1274–1285 (2022).
- 478 16. Biber, M. F., Voskamp, A. & Hof, C. Potential effects of future climate change on global reptile
- 479 distributions and diversity. *Global Ecol. Biogeogr.* **32**, 519–534 (2023).
- 480 17. Pottier, P. *et al.* Vulnerability of amphibians to global warming. *Nature* **954–961** (2025).
- 481 18. Turner, M. G. *et al.* Climate change, ecosystems and abrupt change: science priorities. *Phil.*
- 482 *Trans. R. Soc. B* **375**, 20190105 (2020).
- 483 19. Walsh, J. E. *et al.* Extreme weather and climate events in northern areas: A review. *Earth-Sci.*
- 484 *Rev.* **209**, 103324 (2020).
- 485 20. Bodensteiner, B. L. *et al.* Thermal adaptation revisited: How conserved are thermal traits of
- 486 reptiles and amphibians? *J. Exp. Zool. A* **335**, 173–194 (2021).
- 487 21. Taylor, E. N. *et al.* The thermal ecology and physiology of reptiles and amphibians: A user’s
- 488 guide. *J. Exp. Zool. A* **335**, 13–44 (2021).
- 489 22. Luedtke, J. A. *et al.* Ongoing declines for the world’s amphibians in the face of emerging
- 490 threats. *Nature* **622**, 308–314 (2023).
- 491 23. Hatch, K. A. & Kroft, K. L. Winterkill in Lotic Systems May Be an Important Driver of
- 492 Amphibian Population Declines. *Ichthyol. Herpetol.* **110**, 575–584 (2022).
- 493 24. Enriquez-Urzelai, U., Kearney, M. R., Nicieza, A. G. & Tingley, R. Integrating mechanistic and
- 494 correlative niche models to unravel range-limiting processes in a temperate amphibian. *Global*
- 495 *Change Biol.* **25**, 2633–2647 (2019).
- 496 25. Huey, R. B. & Slatkin, M. Cost and Benefits of Lizard Thermoregulation. *Q. Rev. Biol.* (1976).

- 497 26. Dawson, W. R. On the Physiological Significance of the Preferred Body Temperatures of
498 Reptiles. In: Gates, D. M. & Schmerl, R. B. (eds) *Perspectives of Biophysical Ecology*, 443–473
499 (Springer, 1975).
- 500 27. Buckley, L. B. Linking Traits to Energetics and Population Dynamics to Predict Lizard Ranges
501 in Changing Environments. *Am. Nat.* **171**, E1–E19 (2008).
- 502 28. Buckley, L. B. *et al.* Can mechanism inform species' distribution models? *Ecol. Lett.* **13**, 1041–
503 1054 (2010).
- 504 29. Clusella-Trullas, S., Blackburn, T. M. & Chown, S. L. Climatic Predictors of Temperature
505 Performance Curve Parameters in Ectotherms Imply Complex Responses to Climate Change.
506 *Am. Nat.* **177**, 738–751 (2011).
- 507 30. Caetano, G. H. O. *et al.* Time of activity is a better predictor of the distribution of a tropical
508 lizard than pure environmental temperatures. *Oikos* **129**, 953–963 (2020).
- 509 31. Claunch, N. M. *et al.* Commonly collected thermal performance data can inform species
510 distributions in a data-limited invader. *Sci. Rep.* **13**, 15880 (2023).
- 511 32. Ivey, K. *et al.* Temperature-based activity estimation accurately predicts surface activity, but not
512 microhabitat use, in the endangered heliothermic lizard *Gambelia sila*. *Amphib. Reptile Conserv.*
513 **16**, 10 (2022).
- 514 33. Üveges, B. *et al.* Experimental evidence for beneficial effects of projected climate change on
515 hibernating amphibians. *Sci. Rep.* **6**, 26754 (2016).
- 516 34. Rollins-Smith, L. A. Amphibian immunity–stress, disease, and climate change. *Dev. Comp.*
517 *Immunol.* **66**, 111–119 (2017).
- 518 35. Beukema, W. *et al.* Microclimate limits thermal behaviour favourable to disease control in a
519 nocturnal amphibian. *Ecol. Lett.* **24**, 27–37 (2021).
- 520 36. Waddle, A. W. *et al.* Hotspot shelters stimulate frog resistance to chytridiomycosis. *Nature* **631**,
521 344–349 (2024).
- 522 37. Greenspan, S. E. *et al.* Warming drives ecological community changes linked to host-associated
523 microbiome dysbiosis. *Nat. Clim. Chang.* **10**, 1057–1061 (2020).
- 524 38. Zhu, L. *et al.* Environmental Temperatures Affect the Gastrointestinal Microbes of the Chinese
525 Giant Salamander. *Front. Microbiol.* **12** (2021).
- 526 39. Robak, M. J., Saenz, V., de Cortie, E. & Richards-Zawacki, C. L. Effects of temperature on the
527 interaction between amphibian skin bacteria and *Batrachochytrium dendrobatidis*. *Front.*
528 *Microbiol.* **14** (2023).
- 529 40. Sunday, J. M., Bates, A. E. & Dulvy, N. K. Thermal tolerance and the global redistribution of
530 animals. *Nat. Clim. Chang.* **2**, 686–690 (2012).
- 531 41. Moore, N. A. *et al.* Temperate species underfill their tropical thermal potentials on land. *Nat.*
532 *Ecol. Evol.* **7**, 1993–2003 (2023).
- 533 42. Bennett, J. M. *et al.* The evolution of critical thermal limits of life on Earth. *Nat. Commun.* **12**,
534 1198 (2021).
- 535 43. Harishchandra, A., Xue, H., Salinas, S. & Jayasundara, N. Thermal physiology integrated
536 species distribution model predicts profound habitat fragmentation for estuarine fish with ocean
537 warming. *Sci. Rep.* **12**, 21781 (2022).
- 538 44. Zlotnick, O. B., Musselman, K. N. & Levy, O. Deforestation poses deleterious effects to tree-
539 climbing species under climate change. *Nat. Clim. Chang.* **14**, 289–295 (2024).
- 540 45. Stark, G., Ma, L., Zeng, Z.-G., Du, W.-G. & Levy, O. Cool shade and not-so-cool shade: How
541 habitat loss may accelerate thermal stress under current and future climate. *Global Change Biol.*
542 **29**, 6201–6216 (2023).
- 543 46. Briscoe, N. J. *et al.* Mechanistic forecasts of species responses to climate change: The promise
544 of biophysical ecology. *Global Change Biol.* **29**, 1451–1470 (2023).
- 545 47. Kearney, M. *et al.* Modelling species distributions without using species distributions: the cane
546 toad in Australia under current and future climates. *Ecography* **31**, 423–434 (2008).
- 547 48. Araújo, M. B. *et al.* Heat freezes niche evolution. *Ecol. Lett.* **16**, 1206–1219 (2013).
- 548 49. Hoffmann, A. A., Chown, S. L. & Clusella-Trullas, S. Upper thermal limits in terrestrial
549 ectotherms: how constrained are they? *Funct. Ecol.* **27**, 934–949 (2013).
- 550 50. Chown, S. L. Physiological variation in insects: hierarchical levels and implications. *J. Insect*
551 *Physiol.* **47**, 649–660 (2001).

- 552 51. Rubalcaba, J. G., Gouveia, S. F., Villalobos, F., Olalla-Tárraga, M. Á. & Sunday, J. Climate
553 drives global functional trait variation in lizards. *Nat. Ecol. Evol.* **7**, 524–534 (2023).
- 554 52. Longo, A. V. & Zamudio, K. R. Temperature variation, bacterial diversity and fungal infection
555 dynamics in the amphibian skin. *Mol. Ecol.* **26**, 4787–4797 (2017).
- 556 53. Xu, L. *et al.* The Behavior of Amphibians Shapes Their Symbiotic Microbiomes. *mSystems* **5**,
557 e00626-20 (2020).
- 558 54. Bernardo-Cravo, A. P., Schmeller, D. S., Chatzinotas, A., Vredenburg, V. T. & Loyau, A.
559 Environmental Factors and Host Microbiomes Shape Host–Pathogen Dynamics. *Trends*
560 *Parasitol.* **36**, 616–633 (2020).
- 561 55. Sonn, J. M., Porter, W. P., Mathewson, P. D. & Richards-Zawacki, C. L. Predictions of Disease
562 Risk in Space and Time Based on the Thermal Physiology of an Amphibian Host–Pathogen
563 Interaction. *Front. Ecol. Evol.* **8** (2020).
- 564 56. Neely, W. J. *et al.* Synergistic effects of warming and disease linked to high mortality in cool-
565 adapted terrestrial frogs. *Biol. Conserv.* **245**, 108521 (2020).
- 566 57. Fontaine, S. S., Mineo, P. M. & Kohl, K. D. Experimental manipulation of microbiota reduces
567 host thermal tolerance and fitness under heat stress in a vertebrate ectotherm. *Nat. Ecol. Evol.* **6**,
568 405–417 (2022).
- 569 58. Crump, P. & Grow, S. Action plan for ex situ amphibian conservation in the AZA community.
570 Association of Zoos & Aquariums, Amphibian Taxon Advisory Group (2007).
- 571 59. Beaupre, S., Jacobson, E., Lillywhite, H. & Zamudio, K. Guidelines for use of live amphibians
572 and reptiles in field and laboratory research. (2004).
- 573 60. Poole, V. A. & Grow, S. Amphibian Husbandry Resource Guide, Edition 2.0. (Association of
574 Zoos & Aquariums, Silver Spring, Maryland, USA, 2017).
- 575 61. Pough, H. F. Amphibian biology and husbandry. *ILAR J.* **48**, 203–213 (2007).
- 576 62. ASIH (American Society of Ichthyologists and Herpetologists). Guidelines for Use of Live
577 Amphibians and Reptiles in Field and Laboratory Research. (Lawrence, 2004).
- 578 63. Linhoff, L. J. & Donnelly, M. A. Assessing release strategies for reintroductions of endangered
579 Wyoming toads. *Wildl. Soc. Bull.* **46**, e1341 (2022).
- 580 64. Klocke, B. *et al.* Movement and survival of captive-bred *Atelopus* harlequin frogs (*Atelopus*
581 *limosus*) released into the wild. *Front. Amphib. Reptile Sci.* **1** (2023).
- 582 65. Greenspan, S. E. *et al.* Realistic heat pulses protect frogs from disease under simulated
583 rainforest frog thermal regimes. *Funct. Ecol.* **31**, 2274–2286 (2017).
- 584 66. Sinervo, B. *et al.* Climate change and collapsing thermal niches of desert reptiles and
585 amphibians: Assisted migration and acclimation rescue from extirpation. *Sci. Total Environ.* **908**,
586 168431 (2024).
- 587 67. McNutt, M. K. *et al.* Transparency in authors’ contributions and responsibilities to promote
588 integrity in scientific publication. *Proc. Natl. Acad. Sci. U.S.A.* **115**, 2557–2560 (2018).
- 589 68. Nakagawa, S. *et al.* Method Reporting with Initials for Transparency (MeRIT) promotes more
590 granularity and accountability for author contributions. *Nat. Commun.* **14**, 1788 (2023).
- 591 69. Pottier, P. *et al.* Title, abstract and keywords: a practical guide to maximize the visibility and
592 impact of academic papers. *Proc. R. Soc. B* **291**, 20241222 (2024).
- 593 70. Harzing, A. Publish or perish. (2007).
- 594 71. Cowles, R. B. & Bogert, C. M. A preliminary study of the thermal requirements of desert
595 reptiles. *Bull. Am. Mus. Nat. Hist.* **83**, 261–296 (1944).
- 596 72. Fry, F. E. J. Effects of the environment on animal activity. *Publ. Ont. Fish. Res. Lab.* **55**, 1–62
597 (1947).
- 598 73. Costanzo, J. P., Reynolds, A. M., Amaral, M. C. F. do, Rosendale, A. J. & Jr, R. E. L.
599 Cryoprotectants and Extreme Freeze Tolerance in a Subarctic Population of the Wood Frog.
600 *PLoS ONE* **10**, e0117234 (2015).
- 601 74. Ouzzani, M., Hammady, H., Fedorowicz, Z. & Elmagarmid, A. Rayyan—a web and mobile app
602 for systematic reviews. *Syst. Rev.* **5**, 210 (2016).
- 603 75. O’Dea, R. E. *et al.* Preferred reporting items for systematic reviews and meta-analyses in
604 ecology and evolutionary biology: a PRISMA extension. *Biol. Rev.* **96**, 1695–1722 (2021).

- 605 76. Pick, J. L., Nakagawa, S. & Noble, D. W. A. Reproducible, flexible and high-throughput data
606 extraction from primary literature: The metaDigitise r package. *Methods Ecol. Evol.* **10**, 426–
607 431 (2019).
- 608 77. R Core Team. R: A language and environment for statistical computing. R Foundation for
609 Statistical Computing (2019).
- 610 78. Rohatgi, A. WebPlotDigitizer. (2024).
- 611 79. Ripley, B. *et al.* Package ‘mass’. *CRAN R* **538**, 822 (2013).
- 612 80. AmphibiaWeb. <https://amphibiaweb.org>. University of California, Berkeley, California, USA
613 (2025).
- 614 81. Jetz, W. & Pyron, R. A. The interplay of past diversification and evolutionary isolation with
615 present imperilment across the amphibian tree of life. *Nat. Ecol. Evol.* **2**, 850–858 (2018).
- 616 82. IUCN. The IUCN Red List of Threatened Species. (2021).
- 617 83. Agudelo-Cantero, G. A. & Navas, C. A. Interactive effects of experimental heating rates,
618 ontogeny and body mass on the upper thermal limits of anuran larvae. *J. Therm. Biol.* **82**, 43–51
619 (2019).
- 620 84. Alveal, N. *et al.* Relationship between thermal behavior and sex of a population of *Pleurodema*
621 *thaul* (Amphibia: Leiuperidae) of the commune of Antuco, Biobío region. *Gayana (Concepción)*
622 **83**, 93–101 (2019).
- 623 85. Alveal Riquelme, N. Relaciones entre la fisiología térmica y las características bioclimáticas de
624 *Rhinella spinulosa* (Anura: Bufonidae) en Chile a través del enlace mecanicista de nicho
625 (Universidad de Concepción, 2015).
- 626 86. Alves, M. Tolerância térmica em espécies de anuros neotropicais do gênero *Dendropsophus*
627 Fitzinger, 1843 e efeito da temperatura na resposta à predação. (Universidade Estadual de Santa
628 Cruz, 2016).
- 629 87. Anderson, R. C. O. & Andrade, D. V. Trading heat and hops for water: Dehydration effects on
630 locomotor performance, thermal limits, and thermoregulatory behavior of a terrestrial toad.
631 *Ecol. Evol.* **7**, 9066–9075 (2017).
- 632 88. Aponte Gutiérrez, A. Endurecimiento térmico en *Pristimantis medemi* (Anura: Craugastoridae),
633 en coberturas boscosas del Municipio de Villavicencio (Meta). (Universidad Nacional de
634 Colombia, 2020).
- 635 89. Arrigada García, K. Conductas térmica en dos poblaciones de *Batrachyla taeniata* provenientes
636 de la localidad de Ucúquer en la región de O’Higgins y de la localidad de Hualpén en la
637 (Universidad de Concepción, 2019).
- 638 90. Azambuja, G., Martins, I. K., Franco, J. L. & Santos, T. G. dos. Effects of mancozeb on heat
639 Shock protein 70 (HSP70) and its relationship with the thermal physiology of *Physalaemus*
640 *henselii* tadpoles (Anura: Leptodactylidae). *J. Therm. Biol.* **98**, 102911 (2021).
- 641 91. Bacigalupe, L. D. *et al.* Natural selection on plasticity of thermal traits in a highly seasonal
642 environment. *Evol. Appl.* **11**, 2004–2013 (2018).
- 643 92. Bakewell, L., Kelehear, C. & Graham, S. P. Impacts of temperature on immune performance in a
644 desert anuran (*Anaxyrus punctatus*). *J. Zool.* **315**, 49–57 (2021).
- 645 93. Balogová, M. & Gvoždík, L. Can Newts Cope with the Heat? Disparate Thermoregulatory
646 Strategies of Two Sympatric Species in Water. *PLoS ONE* **10**, e0128155 (2015).
- 647 94. Barria, A. M. & Bacigalupe, L. D. Intraspecific geographic variation in thermal limits and
648 acclimatory capacity in a wide distributed endemic frog. *J. Therm. Biol.* **69**, 254–260 (2017).
- 649 95. Bazin, Y., Wharton, D. A. & Bishop, P. J. Cold tolerance and overwintering of an introduced
650 New Zealand frog, the brown tree frog (*Litoria ewingii*). *Cryo Lett.* **28**, 347–358 (2007).
- 651 96. Beltrán, I., Ramírez-Castañeda, V., Rodríguez-López, C., Lasso, E. & Amézquita, A. Dealing
652 with hot rocky environments: critical thermal maxima and locomotor performance in
653 *Leptodactylus lithonaetes* (Anura: Leptodactylidae). *Herpetol. J.* **29**, 155–161 (2019).
- 654 97. Berkhouse, C. & Fries, J. Critical thermal maxima of juvenile and adult San Marcos
655 salamanders (*Eurycea nana*). *Southwest. Nat.* **40**, 430–434 (1995).
- 656 98. Berman, D. I., Meshcheryakova, E. N. & Bulakhova, N. A. The Japanese tree frog (*Hyla*
657 *japonica*), one of the most cold-resistant species of amphibians. *Dokl. Biol. Sci.* **471**, 276–279
658 (2016).

- 659 99. Berman, D. I., Bulakhova, N. A., Meshcheryakova, E. N. & Shekhovtsov, S. V. Overwintering
660 and cold tolerance in the Moor Frog (*Rana arvalis*) across its range. *Can. J. Zool.* **98**, 705–714
661 (2020).
- 662 100. Bernal Castro, E. A. Influence of environment on thermal ecology of direct-developing frogs
663 (Anura: Craugastoridae: *Pristimantis*) in the eastern Andes of Colombia. (Universidad de los
664 Andes, 2019).
- 665 101. Berner, N. J. & Puckett, R. E. Phenotypic flexibility and thermoregulatory behavior in the
666 eastern red-spotted newt (*Notophthalmus viridescens viridescens*). *J. Exp. Zool. A Ecol. Genet.*
667 *Physiol.* **313**, 231–239 (2010).
- 668 102. Beukema, W. *et al.* Microclimate limits thermal behaviour favourable to disease control in a
669 nocturnal amphibian. *Ecol. Lett.* **24**, 27–37 (2021).
- 670 103. Bicego, K. C. & Branco, L. G. S. Discrete electrolytic lesion of the preoptic area prevents LPS-
671 induced behavioral fever in toads. *J. Exp. Biol.* **205**, 3513–3518 (2002).
- 672 104. Bicego-Nahas, K. C., Steiner, A. A., Carnio, E. C., Antunes-Rodrigues, J. & Branco, L. G. S.
673 Antipyretic effect of arginine vasotocin in toads. *Am. J. Physiol. Regul. Integr. Comp. Physiol.*
674 **278**, R1408–R1414 (2000).
- 675 105. Bicego-Nahas, K. C., Gargaglioni, L. H. & Branco, L. G. S. Seasonal changes in the preferred
676 body temperature, cardiovascular, and respiratory responses to hypoxia in the toad, *Bufo*
677 *paracnemis*. *J. Exp. Zool.* **289**, 359–365 (2001).
- 678 106. Blem, C. R., Ragan, C. A. & Scott, L. S. The thermal physiology of two sympatric treefrogs
679 *Hyla cinerea* and *Hyla chrysoscelis* (Anura; Hylidae). *Comp. Biochem. Physiol. A* **85**, 563–570
680 (1986).
- 681 107. Bliss, M. M. & Cecala, K. K. Terrestrial Salamanders Alter Antipredator Behavior Thresholds
682 Following Tail Autotomy. *Herpetologica* **73**, 94–99 (2017).
- 683 108. Bonino, M. F., Cruz, F. B. & Perotti, M. G. Does temperature at local scale explain thermal
684 biology patterns of temperate tadpoles? *J. Therm. Biol.* **94** (2020).
- 685 109. Bovo, R. P. Fisiologia térmica e balanço hídrico em anfíbios anuros. (Universidade Estadual
686 Paulista, 2015).
- 687 110. Branco, L. G. Effects of 2-deoxy-D-glucose and insulin on plasma glucose levels and behavioral
688 thermoregulation of toads. *Am. J. Physiol.* **272**, R1–R5 (1997).
- 689 111. Branco, L. G. & Malvin, G. M. Thermoregulatory effects of cyanide and azide in the toad, *Bufo*
690 *marinus*. *Am. J. Physiol.* **270**, R169–173 (1996).
- 691 112. Branco, L. G., Steiner, A. A., Tattersall, G. J. & Wood, S. C. Role of adenosine in the hypoxia-
692 induced hypothermia of toads. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **279**, R196–201
693 (2000).
- 694 113. Branco, L. G. & Wood, S. C. Role of central chemoreceptors in behavioral thermoregulation of
695 the toad, *Bufo marinus*. *Am. J. Physiol.* **266**, R1483–1487 (1994).
- 696 114. Branco, L. G. S. & Steiner, A. A. Central thermoregulatory effects of lactate in the toad *Bufo*
697 *paracnemis*. *Comp. Biochem. Physiol. A* **122**, 457–461 (1999).
- 698 115. Brattstrom, B. H. Thermal acclimation in Australian amphibians. *Comp. Biochem. Physiol.* **35**,
699 69–103 (1970).
- 700 116. Brattstrom, B. H. & Regal, P. Rate of thermal acclimation in the Mexican salamander
701 *Chiropterotriton*. *Copeia* **1965**, 514–515 (1965).
- 702 117. Brattstrom, B. H. A Preliminary Review of the Thermal Requirements of Amphibians. *Ecology*
703 **44**, 238–255 (1963).
- 704 118. Brattstrom, B. H. Thermal acclimation in Anuran amphibians as a function of latitude and
705 altitude. *Comp. Biochem. Physiol.* **24**, 93–111 (1968).
- 706 119. Brattstrom, B. H. & Lawrence, P. The Rate of Thermal Acclimation in Anuran Amphibians.
707 *Physiol. Zool.* **35**, 148–156 (1962).
- 708 120. Brown, H. A. The heat resistance of some anuran tadpoles (Hylidae and Pelobatidae). *Copeia*
709 **1969**, 138 (1969).
- 710 121. Burke, E. M. & Pough, F. H. The role of fatigue in temperature resistance of salamanders. *J.*
711 *Therm. Biol.* **1**, 163–167 (1976).
- 712 122. Burrowes, P. A., Navas, C. A., Jiménez-Robles, O., Delgado, P. & De La Riva, I. Climatic
713 heterogeneity in the Bolivian Andes: Are frogs trapped? *South Am. J. Herpetol.* **18**, 1–12 (2020).

- 714 123. Bury, R. B. Low thermal tolerances of stream amphibians in the Pacific Northwest: Implications
715 for riparian and forest management. *Appl. Herpetol.* **5**, 63–74 (2008).
- 716 124. Carey, C. Factors affecting body temperatures of toads. *Oecologia* **35**, 197–219 (1978).
- 717 125. Carvajalino-Fernández, J. M., Gomez, M. A. B., Giraldo-Gutiérrez, L. & Navas, C. A. Freeze
718 tolerance in neotropical frogs: an intrageneric comparison using *Pristimantis* species of high
719 elevation and medium elevation. *J. Trop. Ecol.* **37**, 118–125 (2021).
- 720 126. Castellanos García, L. A. Days of futures past: integrating physiology, microenvironments, and
721 biogeographic history to predict response of frogs in neotropical dry-forest to global warming.
722 (Universidad de los Andes, 2017).
- 723 127. Catenazzi, A., Lehr, E. & Vredenburg, V. T. Thermal physiology, disease, and amphibian
724 declines on the eastern slopes of the Andes. *Conserv. Biol.* **28**, 509–517 (2014).
- 725 128. Cecala, K. K., Noggle, W. & Burns, S. Negative Phototaxis Results from Avoidance of Light
726 and Temperature in Stream Salamander Larvae. *J. Herpetol.* **51**, 263–269 (2017).
- 727 129. Chang, L.-W. Heat tolerance and its plasticity in larval *Bufo bankorensis* from different
728 altitudes. (National Cheng Kung University, 2002).
- 729 130. Chavez Landi, P. A. Fisiología térmica de un depredador *Dasythemis* sp.(Odonata: Libellulidae)
730 y su presa *Hypsiboas pellucens* (Anura: Hylidae) y sus posibles implicaciones frente al
731 (Pontificia Universidad Católica Del Ecuador, 2017).
- 732 131. Chen, T.-C., Kam, Y.-C. & Lin, Y.-S. Thermal physiology and reproductive phenology of
733 *Buergeria japonica* (Rhacophoridae) breeding in a stream and a geothermal hot spring in
734 Taiwan. *Zool. Sci.* **18**, 591–596 (2001).
- 735 132. Cheng, C.-B. A study of warming tolerance and thermal acclimation capacity of tadpoles in
736 Taiwan. (Tunghai University, 2017).
- 737 133. Cheng, Y.-J. Effect of salinity on the critical thermal maximum of tadpoles living in brackish
738 water. (Tunghai University, 2017).
- 739 134. Christian, K. A., Nunez, F., Clos, L. & Diaz, L. Thermal relations of some tropical frogs along
740 an altitudinal gradient. *Biotropica* **20**, 236–239 (1988).
- 741 135. Churchill, T. A. & Storey, K. B. Dehydration tolerance in wood frogs: a new perspective on
742 development of amphibian freeze tolerance. *Am. J. Physiol.* **265**, R1324–R1332 (1993).
- 743 136. Claussen, D. L. The thermal relations of the tailed frog, *Ascaphus truei*, and the pacific treefrog,
744 *Hyla regilla*. *Comp. Biochem. Physiol. A* **44**, 137–153 (1973).
- 745 137. Claussen, D. L. Thermal acclimation in ambystomatid salamanders. *Comp. Biochem. Physiol. A*
746 **58**, 333–340 (1977).
- 747 138. Clemente, A. C. Resposta termofílica pós-prandial do sapo-cururu, *Rhinella diptycha*. (UNESP,
748 2019).
- 749 139. Cohen, J. M. *et al.* The thermal mismatch hypothesis explains host susceptibility to an emerging
750 infectious disease. *Ecol. Lett.* **20**, 184–193 (2017).
- 751 140. Contreras Cisneros, J. Temperatura crítica máxima, tolerancia al frío y termopreferendum del
752 tritón del Montseny (*Calotriton arnoldii*). (Universitat de Barcelona, 2019).
- 753 141. Contreras López, J. M. Requerimientos térmicos de *Craugastor loki* (Anura: Craugastoridae) en
754 la Sierra Madre de Chiapas, México. (Universidad de Ciencias y Artes de Chiapas, 2021).
- 755 142. Contreras Oñate, S. Posible efecto de las temperaturas de aclimatación sobre las respuestas
756 térmicas en temperaturas críticas máximas (TC_{más}) y mínimas (TC_{mín}) de una población de
757 (Universidad de Concepción, 2016).
- 758 143. Cooper, R. D. & Shaffer, H. B. Allele-specific expression and gene regulation help explain
759 transgressive thermal tolerance in non-native hybrids of the endangered California tiger
760 salamander (*Ambystoma californiense*). *Mol. Ecol.* **30**, 987–1004 (2021).
- 761 144. Cortes, P. A., Puschel, H., Acuña, P., Bartheld, J. L. & Bozinovic, F. Thermal ecological
762 physiology of native and invasive frog species: do invaders perform better? *Conserv. Physiol.* **4**,
763 cow056 (2016).
- 764 145. Crawshaw, L. I., Rausch, R. N., Wollmuth, L. P. & Bauer, E. J. Seasonal Rhythms of
765 Development and Temperature Selection in Larval Bullfrogs, *Rana catesbeiana* Shaw. *Physiol.*
766 *Zool.* **65**, 346–359 (1992).

- 767 146. Crow, J. C., Forstner, M. R. J., Ostr, K. G. & Tomasso, J. R. The role of temperature on survival
768 and growth of the Barton Springs salamander (*Eurycea sosorum*). *Herpetol. Conserv. Biol.* **11**,
769 328–334 (2016).
- 770 147. Cupp, P. V. Thermal Tolerance of Five Salientian Amphibians during Development and
771 Metamorphosis. *Herpetologica* **36**, 234–244 (1980).
- 772 148. Dabruzzi, T. F., Wygoda, M. L. & Bennett, W. A. Some Like it Hot: Heat Tolerance of the Crab-
773 Eating Frog, *Fejervarya cancrivora*. *Micronesica* **43**, 101–106 (2012).
- 774 149. Dainton, B. H. Heat tolerance and thyroid activity in developing tadpoles and juvenile adults of
775 *Xenopus laevis* (Daudin). *J. Therm. Biol.* **16**, 273–276 (1991).
- 776 150. Daniel, N. J. J. Impact of climate change on Singapore amphibians. (National University of
777 Singapore, 2013).
- 778 151. Davies, S. J., McGeoch, M. A. & Clusella-Trullas, S. Plasticity of thermal tolerance and
779 metabolism but not water loss in an invasive reed frog. *Comp. Biochem. Physiol. A* **189**, 11–20
780 (2015).
- 781 152. de Oliveira Anderson, R. C., Bovo, R. P. & Andrade, D. V. Seasonal variation in the thermal
782 biology of a terrestrial toad, *Rhinella icterica* (Bufonidae), from the Brazilian Atlantic Forest. *J.*
783 *Therm. Biol.* **74**, 77–83 (2018).
- 784 153. de Vlaming, V. L. & Bury, R. B. Thermal Selection in Tadpoles of the Tailed-Frog, *Ascaphus*
785 *truei*. *J. Herpetol.* **4**, 179–189 (1970).
- 786 154. Delgado-Suazo, P. & Burrowes, P. A. Response to thermal and hydric regimes point to
787 differential inter- and intraspecific vulnerability of tropical amphibians to climate warming. *J.*
788 *Therm. Biol.* **103**, 103148 (2022).
- 789 155. Delson, J. & Whitford, W. G. Critical Thermal Maxima in Several Life History Stages in Desert
790 and Montane Populations of *Ambystoma tigrinum*. *Herpetologica* **29**, 352–355 (1973).
- 791 156. Dohm, M. R., Mautz, W. J., Looby, P. G., Gellert, K. S. & Andrade, J. A. Effects of Ozone on
792 Evaporative Water Loss and Thermoregulatory Behavior of Marine Toads (*Bufo marinus*).
793 *Environ. Res.* **86**, 274–286 (2001).
- 794 157. Drakulić, S. *et al.* Local differences of thermal preferences in European common frog (*Rana*
795 *temporaria*) tadpoles. *Zool. Anz.* **268**, 47–54 (2017).
- 796 158. Duarte, H. *et al.* Can amphibians take the heat? Vulnerability to climate warming in subtropical
797 and temperate larval amphibian communities. *Glob. Change Biol.* **18**, 412–421 (2012).
- 798 159. Duarte, H. S. A comparative study of the thermal tolerance of tadpoles of Iberian anurans.
799 (Universidade de Lisboa, 2011).
- 800 160. Duclaux, R., Fantino, M. & Cabanac, M. Comportement thermoregulateur chez *Rana esculenta*.
801 *Pflugers Arch.* **342**, 347–358 (1973).
- 802 161. Dunlap, D. Evidence for a daily rhythm of heat resistance in cricket frogs, *Acris crepitans*.
803 *Copeia* **852** (1969).
- 804 162. Dunlap, D. G. Critical Thermal Maximum as a Function of Temperature of Acclimation in Two
805 Species of Hylid Frogs. *Physiol. Zool.* **41**, 432–439 (1968).
- 806 163. Easton, L. Determining the feasibility of a translocation by investigating the ecology and
807 physiology of the threatened Hochstetter's frog (*Leiopelma hochstetteri*). (University of Otago,
808 2015).
- 809 164. Elwood, J. R. L. Variation in hsp70 levels and thermotolerance among terrestrial salamanders of
810 the *Plethodon glutinosus* complex. (Drexel University, 2003).
- 811 165. Enriquez-Urzelai, U. *et al.* Ontogenetic reduction in thermal tolerance is not alleviated by earlier
812 developmental acclimation in *Rana temporaria*. *Oecologia* **189**, 385–394 (2019).
- 813 166. Enriquez-Urzelai, U. *et al.* The roles of acclimation and behaviour in buffering climate change
814 impacts along elevational gradients. *J. Anim. Ecol.* **89**, 1722–1734 (2020).
- 815 167. Enriquez-Urzelai, U., Palacio, A. S., Merino, N. M., Sacco, M. & Nicieza, A. G. Hindered and
816 constrained: limited potential for thermal adaptation in post-metamorphic and adult *Rana*
817 *temporaria* along elevational gradients. *J. Evol. Biol.* **31**, 1852–1862 (2018).
- 818 168. Erskine, D. J. & Hutchison, V. H. Reduced thermal tolerance in an amphibian treated with
819 melatonin. *J. Therm. Biol.* **7**, 121–123 (1982).

- Escobar Serrano, D. Acclimation scope of the critical thermal limits in *Agalychnis spurrelli* (Hylidae) and *Gastrotheca pseustes* (Hemiphractidae) and their implications under climate change scenarios. (Pontificia Universidad Católica Del Ecuador, 2016).
- Familiar Lopez, M. Distribution, Ecology, Disease and Physiology of a Mountain-Top Endemic Frog in the Face of Climate Change: A Study on *Phyllorhina loveridgei*. (Griffith University, 2016).
- Fan, X., Lei, H. & Lin, Z. Ontogenetic shifts in selected body temperature and thermal tolerance of the tiger frog, *Hoplobatrachus chinensis*. *Acta Ecol. Sin.* **32**, 5574–5580 (2012).
- Fan, X. L., Lin, Z. H. & Scheffers, B. R. Physiological, developmental, and behavioral plasticity in response to thermal acclimation. *J. Therm. Biol.* **97** (2021).
- Fernández-Loras, A. *et al.* Infection with *Batrachochytrium dendrobatidis* lowers heat tolerance of tadpole hosts and cannot be cleared by brief exposure to CTmax. *PLoS ONE* **14** (2019).
- Finnerty, P. B., Shine, R. & Brown, G. P. Survival of the feces: Does a nematode lungworm adaptively manipulate the behavior of its cane toad host? *Ecol. Evol.* **8**, 4606–4618 (2018).
- Floyd, R. B. Ontogenetic change in the temperature tolerance of larval *Bufo marinus* (Anura: Bufonidae). *Comp. Biochem. Physiol. A* **75**, 267–271 (1983).
- Floyd, R. B. Variation in Temperature Preference with Stage of Development of *Bufo marinus* Larvae. *J. Herpetol.* **18**, 153–158 (1984).
- Floyd, R. B. Effects of Photoperiod and Starvation on the Temperature Tolerance of Larvae of the Giant Toad, *Bufo marinus*. *Copeia* **1985**, 625–631 (1985).
- Fong, S.-T. Thermal tolerance of adult Asiatic painted frog *Kaloula pulchra* from different populations. (National University of Tainan, 2014).
- Fontenot, C. L. & Lutterschmidt, W. I. Thermal selection and temperature preference of the aquatic salamander, *Amphiuma tridactylum*. *Herpetol. Conserv. Biol.* **6**, 395–399 (2011).
- Freidenburg, L. K. & Skelly, D. K. Microgeographical variation in thermal preference by an amphibian. *Ecol. Lett.* **7**, 369–373 (2004).
- Frishkoff, L. O., Hadly, E. A. & Daily, G. C. Thermal niche predicts tolerance to habitat conversion in tropical amphibians and reptiles. *Glob. Change Biol.* **21**, 3901–3916 (2015).
- Frost, J. S. & Martin, E. W. A Comparison of Distribution and High Temperature Tolerance in *Bufo americanus* and *Bufo woodhousii fowleri*. *Copeia* **1971**, 750 (1971).
- Galindo, C. A., Cruz, E. X. & Bernal, M. H. Evaluation of the combined temperature and relative humidity preferences of the Colombian terrestrial salamander *Bolitoglossa ramosi* (Amphibia: Plethodontidae). *Can. J. Zool.* **96**, 1230–1235 (2018).
- Gatten, R. E. & Hill, C. J. Social influence on thermal selection by *Hyla crucifer*. *J. Herpetol.* **18**, 87–88 (1984).
- Gatz, A. J. Critical Thermal Maxima of *Ambystoma maculatum* (Shaw) and *Ambystoma jeffersonianum* (Green) in Relation to Time of Breeding. *Herpetologica* **27**, 157–160 (1971).
- Gatz, A. J. Intraspecific Variations in Critical Thermal Maxima of *Ambystoma maculatum*. *Herpetologica* **29**, 264–268 (1973).
- Geise, W. & Linsenmair, K. E. Adaptations of the reed frog *Hyperolius viridiflavus* (Amphibia, Anura, Hyperoliidae) to its arid environment - IV. Ecological significance of water economy with comments on thermoregulation and energy allocation. *Oecologia* **77**, 327–338 (1988).
- Gélinas, N. Rôle du comportement thermorégulateur chez la grenouille du nord (*Rana septentrionalis*, Baird) adulte et le ouaouaron (*Rana catesbeiana*, Shaw) juvénile en rapport avec le besoin alimentaire. (Université du Québec à Trois-Rivières, 1996).
- Goldstein, J. A., Hoff, K. von S. & Hillyard, S. D. The effect of temperature on development and behaviour of relict leopard frog tadpoles. *Conserv. Physiol.* **5**, cow075 (2017).
- González-del-Piego, P. *et al.* Thermal tolerance and the importance of microhabitats for Andean frogs in the context of land use and climate change. *J. Anim. Ecol.* **89**, 2451–2460 (2020).
- Gouveia, S. F. *et al.* Climatic niche at physiological and macroecological scales: The thermal tolerance-geographical range interface and niche dimensionality. *Glob. Ecol. Biogeogr.* **23**, 446–456 (2014).
- Gray, R. Lack of physiological differentiation in three color morphs of the cricket frog (*Acris crepitans*) in Illinois. *Trans. Ill. State Acad. Sci.* **70**, 73–79 (1977).
- Greenspan, S. E. *et al.* Infection increases vulnerability to climate change via effects on host thermal tolerance. *Sci. Rep.* **7** (2017).

- 875 194. Guevara-Molina, E. C., Gomes, F. R. & Camacho, A. Effects of dehydration on
876 thermoregulatory behavior and thermal tolerance limits of *Rana catesbeiana* (Shaw, 1802). *J.*
877 *Therm. Biol.* **93** (2020).
- 878 195. Gutiérrez Pesquera, L. Una valoración macrofisiológica de la vulnerabilidad al calentamiento
879 global. Análisis de los límites de tolerancia térmica en comunidades de anfibios en gradientes
880 latitudinales y altitudinales. (Pontificia Universidad Católica Del Ecuador, 2015).
- 881 196. Gutiérrez Pesquera, M. Thermal tolerance across latitudinal and altitudinal gradients in tadpoles.
882 (Universidad de Sevilla, 2016).
- 883 197. Gutiérrez-Pesquera, L. M. *et al.* Testing the climate variability hypothesis in thermal tolerance
884 limits of tropical and temperate tadpoles. *J. Biogeogr.* **43**, 1166–1178 (2016).
- 885 198. Gvoždík, L., Puky, M. & Šugerková, M. Acclimation is beneficial at extreme test temperatures
886 in the Danube crested newt, *Triturus dobrogicus* (Caudata, Salamandridae). *Biol. J. Linn. Soc.*
887 **90**, 627–636 (2007).
- 888 199. Gvoždík, L. Does reproduction influence temperature preferences in newts? *Can. J. Zool.* **83**,
889 1038–1044 (2005).
- 890 200. Gvoždík, L. Mismatch Between Ectotherm Thermal Preferenda and Optima for Swimming: A
891 Test of the Evolutionary Pace Hypothesis. *Evol. Biol.* **42**, 137–145 (2015).
- 892 201. Gvoždík, L. & Kristín, P. Economic thermoregulatory response explains mismatch between
893 thermal physiology and behaviour in newts. *J. Exp. Biol.* **220**, 1106–1111 (2017).
- 894 202. Gvoždík, L. Postprandial thermophily in the Danube crested newt, *Triturus dobrogicus*. *J.*
895 *Therm. Biol.* **28**, 545–550 (2003).
- 896 203. Hadamová, M. & Gvoždík, L. Seasonal acclimation of preferred body temperatures improves
897 the opportunity for thermoregulation in newts. *Physiol. Biochem. Zool.* **84**, 166–174 (2011).
- 898 204. Haggerty, J. Thermal tolerance of the common coqui frog (*Eleutherodactylus coqui*) in East
899 Hawaii along an elevation gradient. ProQuest Dissertations and Theses (University of Hawai'i at
900 Hilo, 2016).
- 901 205. Hanna, A. The effects of temperature on physiology in frogs and their dispersal of the temperate
902 zone from the tropics. (Truman State University, 2019).
- 903 206. He, J. *et al.* Influence of High Temperatures and Heat Wave on Thermal Biology, Locomotor
904 Performance, and Antioxidant System of High-Altitude Frog *Nanorana pleskei* Endemic to
905 Qinghai-Tibet Plateau. *Front. Ecol. Evol.* **9** (2021).
- 906 207. Heath, A. G. Behavioral Thermoregulation in High Altitude Tiger Salamanders, *Ambystoma*
907 *tigrinum*. *Herpetologica* **31**, 84–93 (1975).
- 908 208. Heatwole, H., De Austin, S. B. & Herrero, R. Heat tolerances of tadpoles of two species of
909 tropical anurans. *Comp. Biochem. Physiol.* **27**, 807–815 (1968).
- 910 209. Heatwole, H., Mercado, N. & Ortiz, E. Comparison of Critical Thermal Maxima of Two Species
911 of Puerto Rican Frogs of the Genus *Eleutherodactylus*. *Physiol. Zool.* **38**, 1–8 (1965).
- 912 210. Holzman, N. & McManus, J. J. Effects of acclimation on metabolic rate and thermal tolerance in
913 the carpenter frog, *Rana vergatipes*. *Comp. Biochem. Physiol. A* **45**, 833–842 (1973).
- 914 211. Hoppe, D. M. Thermal Tolerance in Tadpoles of the Chorus Frog *Pseudacris triseriata*.
915 *Herpetologica* **34**, 318–321 (1978).
- 916 212. Hou, P.-C. Thermal tolerance and preference in the adult amphibians from different altitudinal
917 LTER sites. (National Cheng Kung University, 2003).
- 918 213. Howard, J. H., Wallace, R. L. & Stauffer Jr, J. R. Critical thermal maxima in populations of
919 *Ambystoma macrodactylum* from different elevations. *J. Herpetol.* **17**, 400–402 (1983).
- 920 214. Hutchison, V. H. & Ritchart, J. P. Annual cycle of thermal tolerance in the salamander, *Necturus*
921 *maculosus*. *J. Herpetol.* **23**, 73–76 (1989).
- 922 215. Hutchison, V. H. & Murphy, K. Behavioral thermoregulation in the salamander *Necturus*
923 *maculosus* after heat shock. *Comp. Biochem. Physiol. A* **82**, 391–394 (1985).
- 924 216. Hutchison, V. H. The Distribution and Ecology of the Cave Salamander, *Eurycea lucifuga*. *Ecol.*
925 *Monogr.* **28**, 2–20 (1958).
- 926 217. Hutchison, V. H. Critical Thermal Maxima in Salamanders. *Physiol. Zool.* **34**, 92–125 (1961).
- 927 218. Hutchison, V. H., Engbretson, G. & Turney, D. Thermal Acclimation and Tolerance in the
928 Hellbender, *Cryptobranchus alleganiensis*. *Copeia* **1973**, 805–807 (1973).

- 929 219. Hutchison, V. H. & Rowland, S. D. Thermal Acclimation and Tolerance in the Mudpuppy,
930 *Necturus maculosus*. *J. Herpetol.* **9**, 367–368 (1975).
- 931 220. Hutchison, V. H. & Spriestersbach, K. K. Diel and Seasonal Cycles of Activity and Behavioral
932 Thermoregulation in the Salamander *Necturus maculosus*. *Copeia* **1986**, 612–618 (1986).
- 933 221. Jara Méndez, D., Krumel Castillo, M. & San Martín Venegas, E. Estudio de las preferencias
934 térmicas y niveles de profundidad en tres estadios larvales de la especie *Pleurodema thaul*
935 (Lesson, 1826) del Parque Nacional Laguna del Laja. (Universidad de Concepción, 2020).
- 936 222. Jiang, S., Yu, P. & Hu, Q. A study on the critical thermal maxima of five species of salamanders
937 of China. *Acta Herpetol. Sin.* **6**, 56–62 (1987).
- 938 223. John-Alder, H. B., Morin, P. J. & Lawler, S. Thermal Physiology, Phenology, and Distribution of
939 Tree Frogs. *Am. Nat.* **132**, 506–520 (1988).
- 940 224. Johnson, C. R. Daily variation in the thermal tolerance of *Litoria caerulea* (Anura: Hylidae).
941 *Comp. Biochem. Physiol. A* **40**, 1109–1111 (1971).
- 942 225. Johnson, C. R. Thermal relations and water balance in the day frog, *Taudactylus diurnus*, from
943 an Australian rain forest. *Aust. J. Zool.* **19**, 35–39 (1971).
- 944 226. Johnson, C. R. Diel variation in the thermal tolerance of *Litoria gracilentia* (Anura: Hylidae).
945 *Comp. Biochem. Physiol. A* **41**, 727–730 (1972).
- 946 227. Johnson, C. R. & Prine, J. E. The effects of sublethal concentrations of organophosphorus
947 insecticides and an insect growth regulator on temperature tolerance in hydrated and dehydrated
948 juvenile western toads, *Bufo boreas*. *Comp. Biochem. Physiol. A* **53**, 147–149 (1976).
- 949 228. Johnson, C. R. Observations on body temperatures, critical thermal maxima and tolerance to
950 water loss in the Australian hylid, *Hyla caerulea* (White). *Proc. R. Soc. Queensland* **82**, 47–50
951 (1970).
- 952 229. Johnson, C. R. Thermal Relations and Daily Variation in the Thermal Tolerance in *Bufo*
953 *marinus*. *J. Herpetol.* **6**, 35 (1972).
- 954 230. Johnson, C. Thermal relations in some southern and eastern Australian anurans. *Proc. R. Soc.*
955 *Queensland* **82**, 87–94 (1971).
- 956 231. Johnson, C. The effects of five organophosphorus insecticides on thermal stress in tadpoles of
957 the Pacific tree frog, *Hyla regilla*. *Zool. J. Linn. Soc.* **69**, 143–147 (1980).
- 958 232. Katzenberger, M., Duarte, H., Relyea, R., Beltrán, J. F. & Tejedo, M. Variation in upper thermal
959 tolerance among 19 species from temperate wetlands. *J. Therm. Biol.* **96** (2021).
- 960 233. Katzenberger, M. *et al.* Swimming with predators and pesticides: How environmental stressors
961 affect the thermal physiology of tadpoles. *PLoS ONE* **9** (2014).
- 962 234. Katzenberger, M., Hammond, J., Tejedo, M. & Relyea, R. Source of environmental data and
963 warming tolerance estimation in six species of North American larval anurans. *J. Therm. Biol.*
964 **76**, 171–178 (2018).
- 965 235. Katzenberger, M. Thermal tolerance and sensitivity of amphibian larvae from Palearctic and
966 Neotropical communities. (Universidade de Lisboa, 2013).
- 967 236. Katzenberger, M. Impact of global warming in Holarctic and Neotropical communities of
968 amphibians. (Universidad de Sevilla, 2014).
- 969 237. Keen, W. H. & Schroeder, E. E. Temperature Selection and Tolerance in Three Species of
970 *Ambystoma* Larvae. *Copeia* **1975**, 523–530 (1975).
- 971 238. Kern, P., Cramp, R. L. & Franklin, C. E. Temperature and UV-B-insensitive performance in
972 tadpoles of the ornate burrowing frog: An ephemeral pond specialist. *J. Exp. Biol.* **217**, 1246–
973 1252 (2014).
- 974 239. Kern, P., Cramp, R. L., Seebacher, F., Ghanizadeh Kazerouni, E. & Franklin, C. E. Plasticity of
975 protective mechanisms only partially explains interactive effects of temperature and UVR on
976 upper thermal limits. *Comp. Biochem. Physiol. A* **190**, 75–82 (2015).
- 977 240. Kern, P., Cramp, R. L. & Franklin, C. E. Physiological responses of ectotherms to daily
978 temperature variation. *J. Exp. Biol.* **218**, 3068–3076 (2015).
- 979 241. Kirsch, D. R., Fix, S., Davenport, J. M., Cecala, K. K. & Ennen, J. R. Body Size Is Related to
980 Temperature Preference in *Hyla chrysoscelis* Tadpoles. *J. Herpetol.* **55**, 21–25 (2021).
- 981 242. Köhler, A. *et al.* Staying warm or moist? Operative temperature and thermal preferences of
982 common frogs (*Rana temporaria*), and effects on locomotion. *Herpetol. J.* **21**, 17–26 (2011).

- 983 243. Kolbe, J. J., Kearney, M. & Shine, R. Modeling the consequences of thermal trait variation for
984 the cane toad invasion of Australia. *Ecol. Appl.* **20**, 2273–2285 (2010).
- 985 244. Komaki, S., Igawa, T., Lin, S.-M. & Sumida, M. Salinity and thermal tolerance of Japanese
986 stream tree frog (*Buergeria japonica*) tadpoles from island populations. *Herpetol. J.* **26**, 207–
987 211 (2016).
- 988 245. Komaki, S., Lau, Q. & Igawa, T. Living in a Japanese onsen: Field observations and
989 physiological measurements of hot spring amphibian tadpoles, *Buergeria japonica*. *Amphibia-
990 Reptilia* **37**, 311–314 (2016).
- 991 246. Krakauer, T. Tolerance limits of the toad, *Bufo marinus*, in South Florida. *Comp. Biochem.
992 Physiol.* **33**, 15–26 (1970).
- 993 247. Kurabayashi, A. *et al.* Improved transport of the model amphibian, *Xenopus tropicalis*, and its
994 viable temperature for transport. *Curr. Herpetol.* **33**, 75–87 (2014).
- 995 248. Lange, L. Influences environnementales précoques et plasticité phénotypique : étude d'un modèle
996 amphibien avec soins parentaux prénataux, l'Alyte accoucheur. (Université de La Rochelle,
997 2020).
- 998 249. Lange, Z. Thermal quality explains shift in habitat association from forest to clearings for
999 terrestrial-breeding frogs along an elevation gradient in Colombia. (John Carroll University,
1000 2019).
- 1001 250. Lau, E. T. C., Leung, K. M. Y. & Karraker, N. E. Native amphibian larvae exhibit higher upper
1002 thermal limits but lower performance than their introduced predator *Gambusia affinis*. *J. Therm.
1003 Biol.* **81**, 154–161 (2019).
- 1004 251. Layne, J. R. Freeze tolerance and cryoprotectant mobilization in the gray treefrog (*Hyla
1005 versicolor*). *J. Exp. Zool.* **283**, 221–225 (1999).
- 1006 252. Layne, J. R. & Claussen, D. L. Seasonal variation in the thermal acclimation of critical thermal
1007 maxima (CTMax) and minima (CTMin) in the salamander *Eurycea bislineata*. *J. Therm. Biol.* **7**,
1008 29–33 (1982).
- 1009 253. Layne, J. R. & Claussen, D. L. The time courses of CTMax and CTMin acclimation in the
1010 salamander *Desmognathus fuscus*. *J. Therm. Biol.* **7**, 139–141 (1982).
- 1011 254. Layne, J. R. & Claussen, D. L. Time courses of thermal acclimation for critical thermal minima
1012 in the salamanders *Desmognathus quadramaculatus*, *D. monticola*, *D. ochrophaeus*, and
1013 *Plethodon jordani*. *Comp. Biochem. Physiol. A* **87**, 895–898 (1987).
- 1014 255. Layne, J. R. Seasonal variation in the cryobiology of *Rana sylvatica* from Pennsylvania. *J.
1015 Therm. Biol.* **20**, 349–353 (1995).
- 1016 256. Layne, J. R. & Romano, M. A. Critical Thermal Minima of *Hyla chrysoscelis*, *H. cinerea*, *H.
1017 gratiosa* and Natural Hybrids (*H. cinerea* × *H. gratiosa*). *Herpetologica* **41**, 216–221 (1985).
- 1018 257. Lee, P.-T. Acidic effect on tadpoles living in container habitats. (Tunghai University, 2019).
- 1019 258. Leger, J. P. & Mathieson, W. B. Effects of Bombesin on Behavioral Thermoregulation in the
1020 Bullfrog. *Brain Behav. Evol.* **50**, 304–312 (2008).
- 1021 259. Li, Z. *et al.* Landscape Connectivity Limits the Predicted Impact of Fungal Pathogen Invasion.
1022 *J. Fungi* **6**, 205 (2020).
- 1023 260. Lillywhite, H. B. Temperature selection by the bullfrog, *Rana catesbeiana*. *Comp. Biochem.
1024 Physiol. A* **40**, 213–227 (1971).
- 1025 261. Litmer, A. R. & Murray, C. M. Critical Thermal Capacities of *Hyla chrysoscelis* in Relation to
1026 Season. *J. Herpetol.* **54**, 413–417 (2020).
- 1027 262. Llewellyn, D., Brown, G. P., Thompson, M. B. & Shine, R. Behavioral Responses to Immune-
1028 System Activation in an Anuran (the Cane Toad, *Bufo marinus*): Field and Laboratory Studies.
1029 *Physiol. Biochem. Zool.* **84**, 77–86 (2011).
- 1030 263. Longhini, L. S., De Almeida Prado, C. P., Bicego, K. C., Zena, L. A. & Gargaglioni, L. H.
1031 Measuring cardiorespiratory variables on small tadpoles using a non-invasive methodology. *Rev.
1032 Cubana Invest. Biomed.* **38** (2019).
- 1033 264. López Rosero, A. C. Ontogenetic variation of thermal tolerance in two anuran species of
1034 Ecuador: *Gastrotheca pseustes* (Hemiphractidae) and *Smilisca phaeota* (Hylidae) and their
1035 relative vulnerability to environmental temperature change. (Pontificia Universidad Católica Del
1036 Ecuador, 2015).

- 1037 265. Lotshaw, D. P. Temperature adaptation and effects of thermal acclimation in *Rana sylvatica* and
1038 *Rana catesbeiana*. *Comp. Biochem. Physiol. A* **56**, 287–294 (1977).
- 1039 266. Lu, H.-L., Wu, Q., Geng, J. & Dang, W. Swimming performance and thermal resistance of
1040 juvenile and adult newts acclimated to different temperatures. *Acta Herpetol.* **11**, 189–195
1041 (2016).
- 1042 267. Lu, H. L., Geng, J., Xu, W., Ping, J. & Zhang, Y. P. Physiological response and changes in
1043 swimming performance after thermal acclimation in juvenile Chinese fire-belly newts, *Cynops*
1044 *orientalis*. *Acta Ecol. Sin.* **37**, 1603–1610 (2017).
- 1045 268. Lutterschmidt, W. I. & Hutchison, V. H. The critical thermal maximum: Data to support the
1046 onset of spasms as the definitive end point. *Can. J. Zool.* **75**, 1553–1560 (1997).
- 1047 269. Madalozzo, B. Variação latitudinal nos limites de tolerância e plasticidade térmica em anfíbios
1048 em um cenário de mudanças climáticas: efeito dos micro-habitats, sazonalidade e filogenia.
1049 (Universidade Federal de Santa Maria, 2018).
- 1050 270. Mahoney, J. J. & Hutchison, V. H. Photoperiod acclimation and 24-hour variations in the critical
1051 thermal maxima of a tropical and a temperate frog. *Oecologia* **2**, 143–161 (1969).
- 1052 271. Malvin, G. M. & Wood, S. C. Behavioral thermoregulation of the toad, *Bufo marinus*: Effects of
1053 air humidity. *J. Exp. Zool.* **258**, 322–326 (1991).
- 1054 272. Maness, J. D. & Hutchison, V. H. Acute adjustment of thermal tolerance in vertebrate
1055 ectotherms following exposure to critical thermal maxima. *J. Therm. Biol.* **5**, 225–233 (1980).
- 1056 273. Manis, M. L. & Claussen, D. L. Environmental and genetic influences on the thermal
1057 physiology of *Rana sylvatica*. *J. Therm. Biol.* **11**, 31–36 (1986).
- 1058 274. Markle, T. M. & Kozak, K. H. Low acclimation capacity of narrow-ranging thermal specialists
1059 exposes susceptibility to global climate change. *Ecol. Evol.* **8**, 4644–4656 (2018).
- 1060 275. Markle, T. Ecology and Evolution of Geographic Range Size Variation in North American
1061 Plethodontid Salamanders: Perspectives from Thermal Physiology. (University of Minnesota
1062 Twin Cities, 2015).
- 1063 276. Marshall, E. & Grigg, G. C. Acclimation of CTM, LD50, and Rapid Loss of Acclimation of
1064 Thermal Preferendum in Tadpoles of *Limnodynastes peronii* (Anura, Myobatrachidae). *Austral.*
1065 *Zool.* **20**, 447–456 (1980).
- 1066 277. Mathias, J. H. The Comparative Ecologies of Two Species of Amphibia (*B. bufo* and *B.*
1067 *calamita*) on the Ainsdale Sand Dunes National Nature Reserve. (The University of Manchester,
1068 1971).
- 1069 278. McCann, S., Greenlees, M. J., Newell, D. & Shine, R. Rapid acclimation to cold allows the cane
1070 toad to invade montane areas within its Australian range. *Funct. Ecol.* **28**, 1166–1174 (2014).
- 1071 279. McCann, S. M., Kosmala, G. K., Greenlees, M. J. & Shine, R. Physiological plasticity in a
1072 successful invader: rapid acclimation to cold occurs only in cool-climate populations of cane
1073 toads (*Rhinella marina*). *Conserv. Physiol.* **6**, cox072 (2018).
- 1074 280. McManus, J. J. & Nellis, D. W. The critical thermal maximum of the marine toad, *Bufo marinus*.
1075 *Caribb. J. Sci.* **15**, 67–70 (1975).
- 1076 281. Menke, M. E. & Claussen, D. L. Thermal acclimation and hardening in tadpoles of the bullfrog,
1077 *Rana catesbeiana*. *J. Therm. Biol.* **7**, 215–219 (1982).
- 1078 282. Merino-Viteri, A. R. The vulnerability of microhylid frogs, *Cophixalus* spp., to climate change
1079 in the Australian Wet Tropics. (James Cook University, 2018).
- 1080 283. Messerman, A. F. Tales of an ‘Invisible’ Life Stage: Survival and Physiology Among Terrestrial
1081 Juvenile Ambystomatid Salamanders. (University of Missouri, 2019).
- 1082 284. Meza-Parral, Y., García-Robledo, C., Pineda, E., Escobar, F. & Donnelly, M. A. Standardized
1083 ethograms and a device for assessing amphibian thermal responses in a warming world. *J.*
1084 *Therm. Biol.* **89** (2020).
- 1085 285. Miller, K. & Packard, G. C. Critical thermal maximum: Ecotypic variation between montane
1086 and piedmont chorus frogs (*Pseudacris triseriata*, Hylidae). *Experientia* **30**, 355–356 (1974).
- 1087 286. Miller, K. & Packard, G. C. An Altitudinal Cline in Critical Thermal Maxima of Chorus Frogs
1088 (*Pseudacris triseriata*). *Am. Nat.* **111**, 267–277 (1977).
- 1089 287. Mittan, C. S. & Zamudio, K. R. Rapid adaptation to cold in the invasive cane toad *Rhinella*
1090 *marina*. *Conserv. Physiol.* **7**, coy075 (2019).

- 1091 288. Moretti, E. H., Ortega Chinchilla, J. E., Marques, F. S., Fernandes, P. A. C. M. & Gomes, F. R.
1092 Behavioral fever decreases metabolic response to lipopolysaccharide in yellow Cururu toads
1093 (*Rhinella icterica*). *Physiol. Behav.* **191**, 73–81 (2018).
- 1094 289. Mueller, C. A., Bucsky, J., Korito, L. & Manzanares, S. Immediate and persistent effects of
1095 temperature on oxygen consumption and thermal tolerance in embryos and larvae of the Baja
1096 California chorus frog, *Pseudacris hypochondriaca*. *Front. Physiol.* **10** (2019).
- 1097 290. Mullens, D. P. & Hutchison, V. H. Diel, seasonal, postprandial and food-deprived
1098 thermoregulatory behaviour in tropical toads (*Bufo marinus*). *J. Therm. Biol.* **17**, 63–67 (1992).
- 1099 291. Navas, C. A., Antoniazzi, M. M., Carvalho, J. E., Suzuki, H. & Jared, C. Physiological basis for
1100 diurnal activity in dispersing juvenile *Bufo granulosus* in the Caatinga, a Brazilian semi-arid
1101 environment. *Comp. Biochem. Physiol. A* **147**, 647–657 (2007).
- 1102 292. Navas, C. A., Úbeda, C. A., Logares, R. & Jara, F. G. Thermal Tolerances in Tadpoles of Three
1103 Species of Patagonian Anurans. *South Am. J. Herpetol.* **5**, 89–96 (2010).
- 1104 293. Nietfeldt, J. W., Jones, S. M., Droge, D. L. & Ballinger, R. E. Rate of thermal acclimation of
1105 larval *Ambystoma tigrinum*. *J. Herpetol.* **14**, 209–211 (1980).
- 1106 294. Nol, R. & Ultsch, G. R. The Roles of Temperature and Dissolved Oxygen in Microhabitat
1107 Selection by the Tadpoles of a Frog (*Rana pipiens*) and a Toad (*Bufo terrestris*). *Copeia* **1981**,
1108 645–652 (1981).
- 1109 295. Novarro, A. J. Thermal Physiology in a Widespread Lungless Salamander. (University of
1110 Maryland, 2018).
- 1111 296. Nowakowski, A. J. *et al.* Thermal biology mediates responses of amphibians and reptiles to
1112 habitat modification. *Ecol. Lett.* **21**, 345–355 (2018).
- 1113 297. Nowakowski, A. J. *et al.* Tropical amphibians in shifting thermal landscapes under land-use and
1114 climate change. *Conserv. Biol.* **31**, 96–105 (2017).
- 1115 298. O'Connor, M. P. & Tracy, C. R. Thermoregulation by Juvenile Toads of *Bufo woodhousei* in the
1116 Field and in the Laboratory. *Copeia* **1992**, 865–876 (1992).
- 1117 299. Orille, A. C., McWhinnie, R. B., Brady, S. P. & Raffel, T. R. Positive Effects of Acclimation
1118 Temperature on the Critical Thermal Maxima of *Ambystoma mexicanum* and *Xenopus laevis*. *J.*
1119 *Herpetol.* **54**, 289–292 (2020).
- 1120 300. Oyamaguchi, H. M. *et al.* Thermal sensitivity of a Neotropical amphibian (*Engystomops*
1121 *pustulosus*) and its vulnerability to climate change. *Biotropica* **50**, 326–337 (2018).
- 1122 301. Paez Vacas, M. I. Mechanisms of population divergence along elevational gradients in the
1123 tropics. (Colorado State University, 2016).
- 1124 302. Paulson, B. K. & Hutchison, V. H. Blood changes in *Bufo cognatus* following acute heat stress.
1125 *Comp. Biochem. Physiol. A* **87**, 461–466 (1987).
- 1126 303. Paulson, B. & Hutchison, V. Origin of the stimulus for muscular spasms at the critical thermal
1127 maximum in anurans. *Copeia* 810–813 (1987).
- 1128 304. Percino-Daniel, R. *et al.* Environmental heterogeneity shapes physiological traits in tropical
1129 direct-developing frogs. *Ecol. Evol.* (2021).
- 1130 305. Perotti, M. G., Bonino, M. F., Ferraro, D. & Cruz, F. B. How sensitive are temperate tadpoles to
1131 climate change? The use of thermal physiology and niche model tools to assess vulnerability.
1132 *Zoology* **127**, 95–105 (2018).
- 1133 306. Piasečná, K., Pončová, A., Tejedo, M. & Gvoždík, L. Thermoregulatory strategies in an aquatic
1134 ectotherm from thermally-constrained habitats: An evaluation of current approaches. *J. Therm.*
1135 *Biol.* **52**, 97–107 (2015).
- 1136 307. Pintanel, P., Tejedo, M., Almeida-Reinoso, F., Merino-Viteri, A. & Gutiérrez-Pesquera, L. M.
1137 Critical thermal limits do not vary between wild-caught and captive-bred tadpoles of *Agalychnis*
1138 *spurrelli* (Anura: Hylidae). *Diversity* **12** (2020).
- 1139 308. Pintanel, P., Tejedo, M., Ron, S. R., Llorente, G. A. & Merino-Viteri, A. Elevational and
1140 microclimatic drivers of thermal tolerance in Andean *Pristimantis* frogs. *J. Biogeogr.* **46**, 1664–
1141 1675 (2019).
- 1142 309. Pintanel, P. Thermal adaptation of amphibians in tropical mountains. Consequences of global
1143 warming. (Universitat de Barcelona, 2018).

- 1144 310. Pintanel, P., Tejedo, M., Salinas-Ivanenko, S., Jervis, P. & Merino-Viteri, A. Predators like it hot:
1145 Thermal mismatch in a predator-prey system across an elevational tropical gradient. *J. Anim.*
1146 *Ecol.* (2021).
- 1147 311. Pough, F. H. Natural daily temperature acclimation of eastern red efts, *Notophthalmus v.*
1148 *viridescens* (Rafinesque) (amphibia: caudata). *Comp. Biochem. Physiol. A* **47**, 71–78 (1974).
- 1149 312. Pough, F. H., Stewart, M. M. & Thomas, R. G. Physiological basis of habitat partitioning in
1150 Jamaican *Eleutherodactylus*. *Oecologia* **27**, 285–293 (1977).
- 1151 313. Quiroga, L. B., Sanabria, E. A., Fornés, M. W., Bustos, D. A. & Tejedo, M. Sublethal
1152 concentrations of chlorpyrifos induce changes in the thermal sensitivity and tolerance of anuran
1153 tadpoles in the toad *Rhinella arenarum*? *Chemosphere* **219**, 671–677 (2019).
- 1154 314. Rausch, C. M., Starkweather, P. L. & van Breukelen, F. One year in the life of *Bufo punctatus*:
1155 annual patterns of body temperature in a free-ranging desert anuran. *Naturwissenschaften* **95**,
1156 531–535 (2008).
- 1157 315. Rausch, C. The thermal ecology of the Red-spotted toad, *Bufo punctatus*, across life history.
1158 (University of Nevada, Las Vegas, 2007).
- 1159 316. Reichenbach, N. & Brophy, T. R. Natural history of the peaks of otter salamander (*Plethodon*
1160 *hubrichti*) along an elevational gradient. *Herpetol. Bull.* 7–15 (2017).
- 1161 317. Reider, K. E., Larson, D. J., Barnes, B. M. & Donnelly, M. A. Thermal adaptations to extreme
1162 freeze–thaw cycles in the high tropical Andes. *Biotropica* **53**, 296–306 (2021).
- 1163 318. Richter-Boix, A. *et al.* Local divergence of thermal reaction norms among amphibian
1164 populations is affected by pond temperature variation. *Evolution* **69**, 2210–2226 (2015).
- 1165 319. Riquelme, N. A., Díaz-Páez, H. & Ortiz, J. C. Thermal tolerance in the Andean toad *Rhinella*
1166 *spinulosa* (Anura: Bufonidae) at three sites located along a latitudinal gradient in Chile. *J.*
1167 *Therm. Biol.* **60**, 237–245 (2016).
- 1168 320. Ritchart, J. P. & Hutchison, V. H. The effects of ATP and cAMP on the thermal tolerance of the
1169 mudpuppy, *Necturus maculosus*. *J. Therm. Biol.* **11**, 47–51 (1986).
- 1170 321. Rivera-Burgos, A. C. Habitat suitability for *Eleutherodactylus* frogs in Puerto Rico: indexing
1171 occupancy, abundance and reproduction to climatic and habitat characteristics. (North Carolina
1172 State University, 2019).
- 1173 322. Rivera-Ordóñez, J. M., Nowakowski, A. J., Manansala, A., Thompson, M. E. & Todd, B. D.
1174 Thermal niche variation among individuals of the poison frog, *Oophaga pumilio*, in forest and
1175 converted habitats. *Biotropica* **51**, 747–756 (2019).
- 1176 323. Rocha, P. L. & Branco, L. G. S. Physiological significance of behavioral hypothermia in
1177 hypoglycemic frogs (*Rana catesbeiana*). *Comp. Biochem. Physiol. A* **119**, 957–961 (1998).
- 1178 324. Rodríguez, C. Y., Bustos, D. A. & Sanabria, E. A. Adaptation of the Andean Toad *Rhinella*
1179 *spinulosa* (Anura: Bufonidae) at low temperatures: The role of glucose as cryoprotectant.
1180 *Physiol. Biochem. Zool.* **92**, 473–480 (2019).
- 1181 325. Romero Barreto, P. Requerimientos fisiológicos y microambientales de dos especies de anfibios
1182 (*Scinax ruber* e *Hyloxalus yasuni*) del bosque tropical de Yasuní y sus implicaciones
1183 (Pontificia Universidad Católica Del Ecuador, 2013).
- 1184 326. Ruiz-Aravena, M. *et al.* Impact of global warming at the range margins: Phenotypic plasticity
1185 and behavioral thermoregulation will buffer an endemic amphibian. *Ecol. Evol.* **4**, 4467–4475
1186 (2014).
- 1187 327. Ruthsatz, K. *et al.* Thyroid hormone levels and temperature during development alter thermal
1188 tolerance and energetics of *Xenopus laevis* larvae. *Conserv. Physiol.* **6** (2018).
- 1189 328. Ruthsatz, K. *et al.* Post-metamorphic carry-over effects of altered thyroid hormone level and
1190 developmental temperature: physiological plasticity and body condition at two life stages in
1191 *Rana temporaria*. *J. Comp. Physiol. B* **190**, 297–315 (2020).
- 1192 329. Rutledge, P. S., Spotila, J. R. & Easton, D. P. Heat hardening in response to two types of heat
1193 shock in the lungless salamanders *Eurycea bislineata* and *Desmognathus ochrophaeus*. *J.*
1194 *Therm. Biol.* **12**, 235–241 (1987).
- 1195 330. Sanabria, E. *et al.* Effect of salinity on locomotor performance and thermal extremes of
1196 metamorphic Andean toads (*Rhinella spinulosa*) from Monte Desert, Argentina. *J. Therm. Biol.*
1197 **74**, 195–200 (2018).

- 1198 331. Sanabria, E. A., González, E., Quiroga, L. B. & Tejedo, M. Vulnerability to warming in a desert
1199 amphibian tadpole community: the role of interpopulational variation. *J. Zool.* **313**, 283–296
1200 (2021).
- 1201 332. Sanabria, E. A. & Quiroga, L. B. Change in the thermal biology of tadpoles of *Odontophrynus*
1202 *occidentalis* from the Monte desert, Argentina: Responses to photoperiod. *J. Therm. Biol.* **36**,
1203 288–291 (2011).
- 1204 333. Sanabria, E. A., Quiroga, L. B., González, E., Moreno, D. & Cataldo, A. Thermal parameters
1205 and locomotor performance in juvenile of *Pleurodema nebulosum* (Anura: Leptodactylidae)
1206 from the Monte Desert. *J. Therm. Biol.* **38**, 390–395 (2013).
- 1207 334. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal changes in the thermal tolerances of
1208 the toad *Rhinella arenarum* (Bufonidae) in the Monte Desert of Argentina. *J. Therm. Biol.* **37**,
1209 409–412 (2012).
- 1210 335. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal Changes in the thermal tolerances of
1211 *Odontophrynus occidentalis* (BERG, 1896) (Anura: Cycloramphidae). *Belg. J. Zool.* **143**, 23–29
1212 (2013).
- 1213 336. Sanabria, E. A. *et al.* Thermal ecology of the post-metamorphic Andean toad (*Rhinella*
1214 *spinulosa*) at elevation in the Monte Desert, Argentina. *J. Therm. Biol.* **52**, 52–57 (2015).
- 1215 337. Sanabria, E. A., Vaira, M., Quiroga, L. B., Akmentins, M. S. & Pereyra, L. C. Variation of
1216 thermal parameters in two different color morphs of a diurnal poison toad, *Melanophryniscus*
1217 *rubriventris* (Anura: Bufonidae). *J. Therm. Biol.* **41**, 1–5 (2014).
- 1218 338. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal changes in the thermoregulatory
1219 strategies of *Rhinella arenarum* in the Monte desert, Argentina. *J. Therm. Biol.* **36**, 23–28
1220 (2011).
- 1221 339. Sanabria, E. A., Vergara, S. C., Rodríguez, C. Y. & Quiroga, L. B. Thermophilic response post
1222 feeding in *Pleurodema nebulosum* (Anura: Leptodactylidae) from Monte desert, Argentina. *J.*
1223 *Therm. Biol.* **90**, 102605 (2020).
- 1224 340. Sanabria, E. A. & Quiroga, L. B. Thermal parameters changes in males of *Rhinella arenarum*
1225 (Anura: Bufonidae) related to reproductive periods. *Rev. Biol. Trop.* **59**, 347–353 (2011).
- 1226 341. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Variation in the Thermal Parameters of
1227 *Odontophrynus occidentalis* in the Monte Desert, Argentina: Response to the Environmental
1228 Constraints. *J. Exp. Zool. A Ecol. Genet. Physiol.* **317**, 185–193 (2012).
- 1229 342. Sauer, E. L. *et al.* Variation in individual temperature preferences, not behavioural fever, affects
1230 susceptibility to chytridiomycosis in amphibians. *Proc. R. Soc. B* **285**, 20181111 (2018).
- 1231 343. Sauer, E. L., Trejo, N., Hoverman, J. T. & Rohr, J. R. Behavioural fever reduces ranaviral
1232 infection in toads. *Funct. Ecol.* **33**, 2172–2179 (2019).
- 1233 344. Sauer, E. L. Behavioral Thermoregulation and Thermal Mismatches Influence Disease
1234 Dynamics in Amphibians. (University of South Florida, 2018).
- 1235 345. Scheffers, B. R. *et al.* Thermal buffering of microhabitats is a critical factor mediating warming
1236 vulnerability of frogs in the Philippine biodiversity hotspot. *Biotropica* **45**, 628–635 (2013).
- 1237 346. Scheffers, B. R., Edwards, D. P., Diesmos, A., Williams, S. E. & Evans, T. A. Microhabitats
1238 reduce animal's exposure to climate extremes. *Glob. Change Biol.* **20**, 495–503 (2014).
- 1239 347. Schmid, W. D. Survival of frogs in low temperature. *Science* **215**, 697–698 (1982).
- 1240 348. Schmid, W. D. High Temperature Tolerances of *Bufo hemiophrys* and *Bufo cognatus*. *Ecology*
1241 **46**, 559–560 (1965).
- 1242 349. Seal, J. A. & West, B. W. Critical Thermal Maxima of Some Arkansas Salamanders in Relation
1243 to Thermal Acclimation. *Herpetologica* **25**, 122–124 (1969).
- 1244 350. Seibel, R. V. Variables Affecting the Critical Thermal Maximum of the Leopard Frog, *Rana*
1245 *pipiens* Schreber. *Herpetologica* **26**, 208–213 (1970).
- 1246 351. Sherman, E. Ontogenetic change in thermal tolerance of the toad *Bufo woodhousii fowleri*.
1247 *Comp. Biochem. Physiol. A* **65**, 227–230 (1980).
- 1248 352. Sherman, E. Thermal biology of newts (*Notophthalmus viridescens*) chronically infected with a
1249 naturally occurring pathogen. *J. Therm. Biol.* **33**, 27–31 (2008).
- 1250 353. Sherman, E., Baldwin, L., Fernández, G. & Deurell, E. Fever and thermal tolerance in the toad
1251 *Bufo marinus*. *J. Therm. Biol.* **16**, 297–301 (1991).

- 1252 354. Sherman, E. & Levitis, D. Heat hardening as a function of developmental stage in larval and
1253 juvenile *Bufo americanus* and *Xenopus laevis*. *J. Therm. Biol.* **28**, 373–380 (2003).
- 1254 355. Shi, L., Zhao, L., Ma, X. & Ma, X. Selected body temperature and thermal tolerance of tadpoles
1255 of two frog species (*Fejervarya limnocharis* and *Microhyla ornata*) acclimated under different
1256 thermal conditions. *Acta Ecol. Sinica* **32**, 0465–0471 (2012).
- 1257 356. Siddons, S. R. & Searle, C. L. Exposure to a fungal pathogen increases the critical thermal
1258 minimum of two frog species. *Ecol. Evol.* **11**, 9589–9598 (2021).
- 1259 357. Sievert, L. M. Thermoregulatory behaviour in the toads *Bufo marinus* and *Bufo cognatus*. *J.*
1260 *Therm. Biol.* **16**, 309–312 (1991).
- 1261 358. Sievert, L. M. & Andreadis, P. T. Differing Diel Patterns of Temperature Selection in Two
1262 Sympatric *Desmognathus*. *Copeia* 2002, 62–66 (2002).
- 1263 359. Simon, M. N., Ribeiro, P. L. & Navas, C. A. Upper thermal tolerance plasticity in tropical
1264 amphibian species from contrasting habitats: Implications for warming impact prediction. *J.*
1265 *Therm. Biol.* **48**, 36–44 (2015).
- 1266 360. Simon, M. Plasticidade fenotípica em relação à temperatura de larvas de *Rhinella* (Anura:
1267 Bufonidae) da caatinga e da floresta Atlântica. (Universidade de Sao Paulo, 2010).
- 1268 361. Skelly, D. K. & Freidenburg, L. K. Effects of beaver on the thermal biology of an amphibian.
1269 *Ecol. Lett.* **3**, 483–486 (2000).
- 1270 362. Smolinský, R. & Gvoždík, L. The ontogenetic shift in thermoregulatory behaviour of newt
1271 larvae: testing the ‘enemy-free temperatures’ hypothesis. *J. Zool.* **279**, 180–186 (2009).
- 1272 363. Smolinský, R. & Gvoždík, L. Interactive influence of biotic and abiotic cues on the plasticity of
1273 preferred body temperatures in a predator–prey system. *Oecologia* **170**, 47–55 (2012).
- 1274 364. Sos, T. Thermoconformity even in hot small temporary water bodies: a case study in yellow-
1275 bellied toad (*Bombina v. variegata*). *Herpetol. Romanica* **1**, 1–11 (2007).
- 1276 365. Spotila, J. R. Role of Temperature and Water in the Ecology of Lungless Salamanders. *Ecol.*
1277 *Monogr.* **42**, 95–125 (1972).
- 1278 366. Strickland, J. C., Pinheiro, A. P., Cecala, K. K. & Dorcas, M. E. Relationship between
1279 Behavioral Thermoregulation and Physiological Function in Larval Stream Salamanders. *J.*
1280 *Herpetol.* **50**, 239–244 (2016).
- 1281 367. Swanson, D. L., Graves, B. M. & Koster, K. L. Freezing tolerance/intolerance and
1282 cryoprotectant synthesis in terrestrially overwintering anurans in the Great Plains, USA. *J.*
1283 *Comp. Physiol. B* **166**, 110–119 (1996).
- 1284 368. Tattersall, G. J. & Boutilier, R. G. Balancing Hypoxia and Hypothermia in Cold-Submerged
1285 Frogs. *J. Exp. Biol.* **200**, 1031–1038 (1997).
- 1286 369. Tattersall, G. J., Tyson, T. M., Lenchyshyn, J. R. & Carlone, R. L. Temperature Preference
1287 During Forelimb Regeneration in the Red-Spotted Newt *Notophthalmus viridescens*. *J. Exp.*
1288 *Zool. A* **317**, 248–258 (2012).
- 1289 370. Toufarová, E. & Gvoždík, L. Do female newts modify thermoregulatory behavior to manipulate
1290 egg size? *J. Therm. Biol.* **57**, 72–77 (2016).
- 1291 371. Tracy, C. R., Christian, K. A., O’Connor, M. P. & Tracy, C. R. Behavioral Thermoregulation by
1292 *Bufo americanus*: The Importance of the Hydric Environment. *Herpetologica* **49**, 375–382
1293 (1993).
- 1294 372. Tracy, C. R., Christian, K. A., Betts, G. & Tracy, C. R. Body temperature and resistance to
1295 evaporative water loss in tropical Australian frogs. *Comp. Biochem. Physiol. A* **150**, 102–108
1296 (2008).
- 1297 373. Tracy, C. R. & Christian, K. A. Preferred Temperature Correlates with Evaporative Water Loss
1298 in Hylid Frogs from Northern Australia. *Physiol. Biochem. Zool.* **78**, 839–846 (2005).
- 1299 374. Trochet, A. *et al.* Variation of preferred body temperatures along an altitudinal gradient: A multi-
1300 species study. *J. Therm. Biol.* **77**, 38–44 (2018).
- 1301 375. Turriago, J. L., Parra, C. A. & Bernal, M. H. Upper thermal tolerance in anuran embryos and
1302 tadpoles at constant and variable peak temperatures. *Can. J. Zool.* **93**, 267–272 (2015).
- 1303 376. Vidal, M. A., Novoa-Muñoz, F., Werner, E., Torres, C. & Nova, R. Modeling warming predicts a
1304 physiological threshold for the extinction of the living fossil frog *Calyptocephalella gayi*. *J.*
1305 *Therm. Biol.* **69**, 110–117 (2017).

- 1306 377. Viertel, B. The reaction of suspension feeding anuran larvae to temperature, hyperbaric and
1307 hyperoxic waters. *Zool. Jahrb. Syst.* **120**, 1–11 (1993).
- 1308 378. Vimercati, G. Exploring the invasion of the guttural toad *Sclerophrys gutturalis* in Cape Town
1309 through a multidisciplinary approach. (Stellenbosch University, 2017).
- 1310 379. Vinšálková, T. & Gvoždík, L. Mismatch between temperature preferences and morphology in F1
1311 hybrid newts (*Triturus carnifex* × *T. dobrogicus*). *J. Therm. Biol.* **32**, 433–439 (2007).
- 1312 380. Vo, P. & Gridi-Papp, M. Low temperature tolerance, cold hardening and acclimation in tadpoles
1313 of the neotropical túngara frog (*Engystomops pustulosus*). *J. Therm. Biol.* **66**, 49–55 (2017).
- 1314 381. Voituron, Y., Paaschburg, L., Holmstrup, M., Barré, H. & Ramløv, H. Survival and metabolism
1315 of *Rana arvalis* during freezing. *J. Comp. Physiol. B* **179**, 223–230 (2009).
- 1316 382. von May, R. *et al.* Divergence of thermal physiological traits in terrestrial breeding frogs along a
1317 tropical elevational gradient. *Ecol. Evol.* **7**, 3257–3267 (2017).
- 1318 383. von May, R. *et al.* Thermal physiological traits in tropical lowland amphibians: Vulnerability to
1319 climate warming and cooling. *PLoS ONE* **14** (2019).
- 1320 384. Wagener, C., Kruger, N. & Measey, J. Progeny of *Xenopus laevis* from altitudinal extremes
1321 display adaptive physiological performance. *J. Exp. Biol.* **224** (2021).
- 1322 385. Wang, H. & Wang, L. Thermal adaptation of the common giant toad (*Bufo gargarizans*) at
1323 different earlier developmental stages. *J. Agric. Univ. Hebei* **31**, 79–83 (2008).
- 1324 386. Wang, L. The effects of constant and variable thermal acclimation on thermal tolerance of the
1325 common giant toad tadpoles (*Bufo gargarizans*). *Acta Ecol. Sinica* **34**, 1030–1034 (2014).
- 1326 387. Wang, L.-Z. & Li, X.-C. Effect of temperature on incubation and thermal tolerance of the
1327 Chinese forest frog. *Chin. J. Zool.* (2007).
- 1328 388. Wang, L. & Li, X.-C. Effects of constant thermal acclimation on thermal tolerance of the
1329 Chinese forest frog (*Rana chensinensis*). *Acta Hydrobiol. Sinica* **31**, 748–750 (2007).
- 1330 389. Wang, L.-Z., Li, X.-C. & Sun, T. Preferred temperature, avoidance temperature and lethal
1331 temperature of tadpoles of the common giant toad (*Bufo gargarizans*) and the Chinese forest
1332 frog (*Rana chensinensis*). *Chin. J. Zool.* **40**, 23–27 (2005).
- 1333 390. Warburg, M. R. On the water economy of Israel amphibians: The anurans. *Comp. Biochem.*
1334 *Physiol. A* **40**, 911–924 (1971).
- 1335 391. Warburg, M. R. The water economy of Israel amphibians: The urodeles *Triturus vittatus*
1336 (Jenyns) and *Salamandra salamandra* (L.). *Comp. Biochem. Physiol. A* **40**, 1055–1063 (1971).
- 1337 392. Whitehead, P. J., Puckridge, J. T., Leigh, C. M. & Seymour, R. S. Effect of temperature on jump
1338 performance of the frog *Limnodynastes tasmaniensis*. *Physiol. Zool.* **62**, 937–949 (1989).
- 1339 393. Willhite, C. & Cupp, P. V. Daily rhythms of thermal tolerance in *Rana clamitans* (Anura:
1340 Ranidae) tadpoles. *Comp. Biochem. Physiol. A* **72**, 255–257 (1982).
- 1341 394. Williams, A. A. & Wygoda, M. L. Dehydration stimulates behavioral hypothermia in the gulf
1342 coast toad, *Bufo valliceps*. *J. Therm. Biol.* **18**, 223–227 (1993).
- 1343 395. Winterová, B. & Gvoždík, L. Influence of interspecific competitors on behavioral
1344 thermoregulation: developmental or acute plasticity? *Behav. Ecol. Sociobiol.* **72**, 169 (2018).
- 1345 396. Witters, L. R. & Sievert, L. Feeding causes thermophily in the woodhouse's toad (*Bufo*
1346 *woodhousii*). *J. Therm. Biol.* **26**, 205–208 (2001).
- 1347 397. Wollmuth, L. P., Crawshaw, L. I., Forbes, R. B. & Grahn, D. A. Temperature selection during
1348 development in a montane anuran species, *Rana cascadae*. *Physiol. Zool.* **60**, 472–480 (1987).
- 1349 398. Wu, C.-S. & Kam, Y.-C. Thermal tolerance and thermoregulation by Taiwanese rhacophorid
1350 tadpoles (*Buergeria japonica*) living in geothermal hot springs and streams. *Herpetologica* **61**,
1351 35–46 (2005).
- 1352 399. Wu, Q.-H. & Hsieh, C.-H. Thermal tolerance and population genetics of *Hynobius fuca*. (2016).
- 1353 400. Wu, Q.-X. Study on the temperature preference and temperature acclimation ability of tadpoles
1354 in different microhabitats. (2021).
- 1355 401. Xu, X. The effect of temperature on body temperature and thermoregulation in different
1356 geographic populations of *Rana dybowskii*. (Harbin Normal University, Harbin, China, 2017).
- 1357 402. Yandún Vela, M. C. Capacidad de aclimatación en renacuajos de dos especies de anuros:
1358 *Rhinella marina* (Bufonidae) y *Gastrotheca riobambae* (Hemiphractidae) y su vulnerabilidad al
1359 cambio climático. (Pontificia Universidad Católica Del Ecuador, Quito, Ecuador, 2017).

1360 403. Young, V. K. H. & Gifford, M. E. Limited capacity for acclimation of thermal physiology in a
1361 salamander, *Desmognathus brimleyorum*. *J. Comp. Physiol. B* **183**, 409–418 (2013).
1362 404. Yu, Z., Dickstein, R., Magee, W. E. & Spotila, J. R. Heat shock response in the salamanders
1363 *Plethodon jordani* and *Plethodon cinereus*. *J. Therm. Biol.* **23**, 259–265 (1998).
1364 405. Zheng, R.-Q. & Liu, C.-T. Giant spiny-frog (*Paa spinosa*) from different populations differ in
1365 thermal preference but not in thermal tolerance. *Aquat. Ecol.* **44**, 723–729 (2010).
1366 406. Zweifel, R. G. Studies on the Critical Thermal Maxima of Salamanders. *Ecology* **38**, 64–69
1367 (1957).
1368 407. Pintanel, P. *et al.* Elevational and local climate variability predicts thermal breadth of mountain
1369 tropical tadpoles. *Ecography* 2022, e05906 (2022).
1370 408. Gutiérrez-Pesquera, L. M. *et al.* Phenology and plasticity can prevent adaptive clines in thermal
1371 tolerance across temperate mountains: The importance of the elevation-time axis. *Ecol. Evol.* **12**,
1372 e9349 (2022).
1373 409. Pottier, P. *et al.* New horizons for comparative studies and meta-analyses. *Trends Ecol. Evol.* **39**,
1374 435–445 (2024).
1375 410. Jenkins, C. N., Pimm, S. L. & Joppa, L. N. Global patterns of terrestrial vertebrate diversity and
1376 conservation. *Proc. Natl. Acad. Sci. U.S.A.* **110**, E2602–E2610 (2013).
1377 411. Asubiaro, T. V. & Onaolapo, S. A comparative study of the coverage of African journals in Web
1378 of Science, Scopus, and CrossRef. *J. Assoc. Inf. Sci. Technol.* **74**, 745–758 (2023).
1379 412. Bonnet, X., Shine, R. & Lourdais, O. Taxonomic chauvinism. *Trends Ecol. Evol.* **17**, 1–3 (2002).
1380 413. Pottier, P. *et al.* AmphiTherm v1.0.1. Zenodo <https://doi.org/10.5281/zenodo.16751427> (2025).
1381 414. Giustini, D. & Boulos, M. N. K. Google Scholar is not enough to be used alone for systematic
1382 reviews. *Online J. Public Health Inform.* **5**, e61265 (2013).
1383 415. Haddaway, N. R., Collins, A. M., Coughlin, D. & Kirk, S. The Role of Google Scholar in
1384 Evidence Reviews and Its Applicability to Grey Literature Searching. *PLoS ONE* **10**, e0138237
1385 (2015).
1386 416. Amano, T. *et al.* Tapping into non-English-language science for the conservation of global
1387 biodiversity. *PLoS Biol.* **19**, e3001296 (2021).
1388 417. Noble, D. W. A. *et al.* Meta-analytic approaches and effect sizes to account for ‘nuisance
1389 heterogeneity’ in comparative physiology. *J. Exp. Biol.* **225**, jeb243225 (2022).
1390

Supplementary Information

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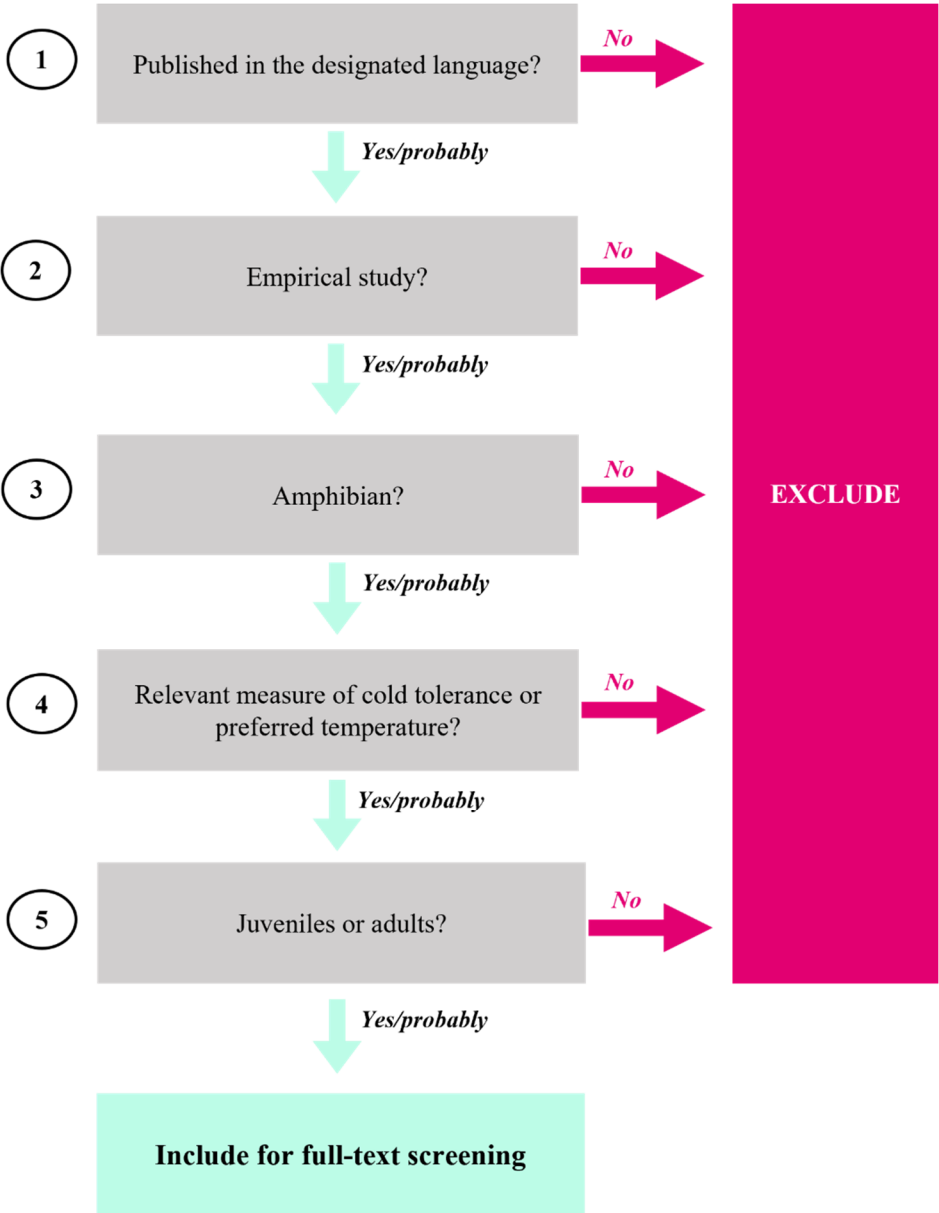


Figure S1: Decision tree used to screen titles, abstracts, and keywords. Additional details can be found in Table S3.

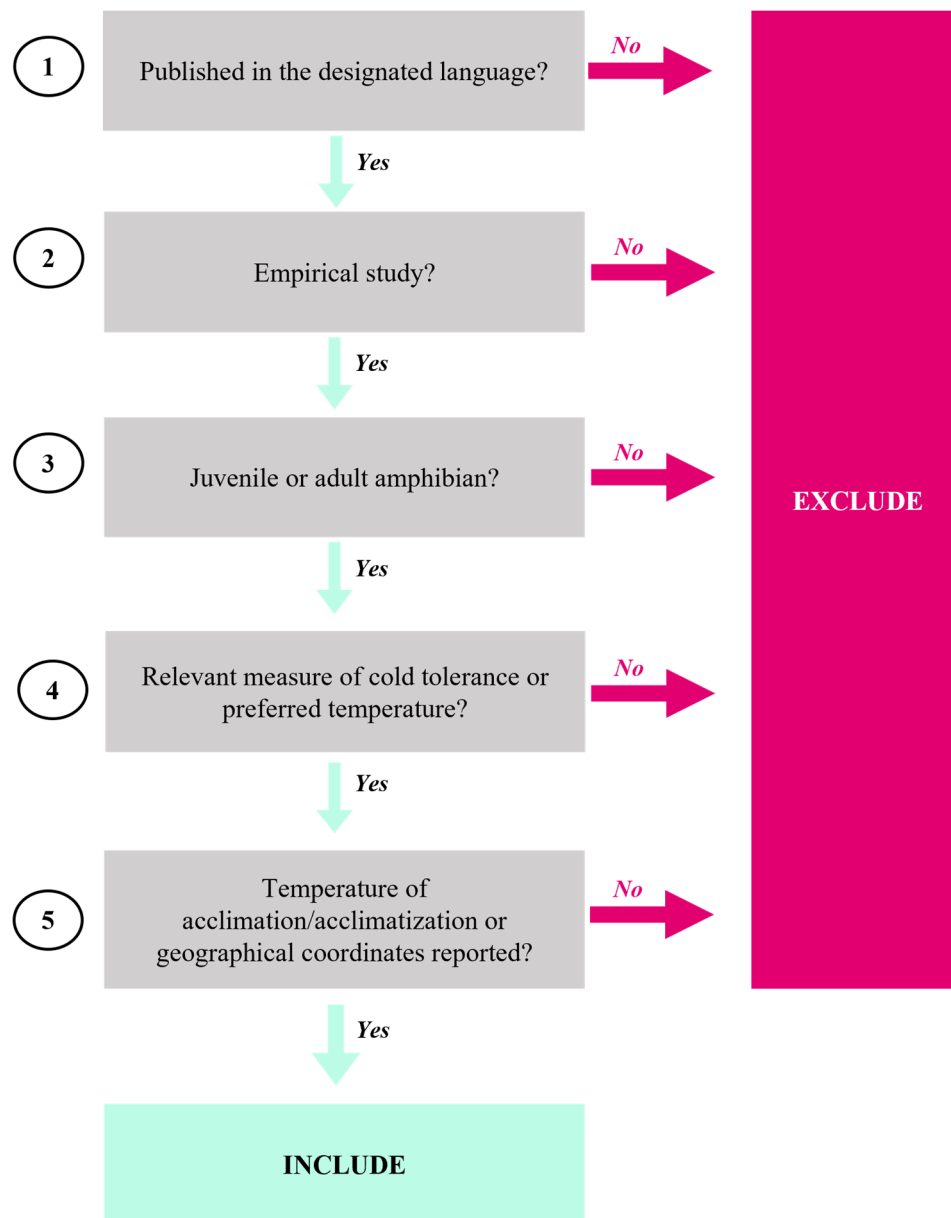


Figure S2: Decision tree used to assess full articles for eligibility. Additional details can be found in Table S4.

Table S1: Search strings used for the different databases.

Database	Search strings
Scopus	TITLE-ABS-KEY("temperature*" OR "thermal" OR "cold*" OR "cool*") AND TITLE-ABS-KEY("cold tolerance*" OR "tolerance* to cold" OR "thermal min*" OR "CTmin" OR "CT min" OR "chill coma" OR "cold stress tolerance*" OR "tolerance to cold stress" OR "cold stupor" OR "cold resistance" OR "resistance to cold stress" OR "supercooling point" OR "SCP" OR "crystal* temperature*" OR "cold hardness" OR "freez* tolerance" OR "tolerance to freezing" OR "preferred temperature*" OR "preferred body temperature*" OR "temperature preference*" OR "selected temperature*" OR "selected body temperature*" OR "thermal prefer*" OR "temperature* prefer*" OR "temperature* select*" OR "thermal selection") AND TITLE-ABS-KEY("amphibia*" OR "frog*" OR "toad*" OR "salamand*" OR "newt" OR "newts" OR "tadpole*" OR "metamorph" OR "metamorphs" OR "caecili*" OR "rhinatrema*" OR "ichthyophi*" OR "scolecomorph*" OR "chikil*" OR "herpelidae" OR "typhlonect*" OR "indotyphlid*" OR "dermophi*" OR "siphonop*" OR "caudata" OR "urodela" OR "cryptobranch*" OR "hynobiid*" OR "sirenidae" OR "ambystoma*" OR "dicamptodon*" OR "proteidae" OR "rhyacotriton*" OR "amphium*" OR "plethodon*" OR "anura*" OR "ascaph*" OR "leiopelma*" OR "bombina*" OR "alyt*" OR "rhinophryn*" OR "pipidae" OR "xenopus" OR "scaphiop*" OR "pelodyt*" OR "megophry*" OR "pelobat*" OR "heleophryn*" OR "calyptocephalell*" OR "myobatrach*" OR "rhinoderma*" OR "alsod*" OR "hylod*" OR "batrachyl*" OR "cycloramph*" OR "telmatob*" OR "ceratophry*" OR "hemiphract*" OR "hyla*" OR "hylidae" OR "bufo*" OR "leptodactyl*" OR "odontophryn*" OR "allophryn*" OR "centrolen*" OR "dendrobat*" OR "ceuthomanti*" OR "eleutherodactyl*" OR "brachycephalidae" OR "craugastor*" OR "strabomantidae" OR "pristimantis" OR "nasikabatrach*" OR "soogloss*" OR "microhyl*" OR "arthroleptid*" OR "hyperol*" OR "brevicipitidae" OR "hemisus" OR "odontobatrach*" OR "phrynobatrach*" OR "ptychaden*" OR "conraua" OR "petropedet*" OR "pyxicephal*" OR "micrixalus" OR "nyctibatrach*" OR "ranixalidae" OR "ceratobatrach*" OR "dicroglossidae" OR "rana" OR "ranidae" OR "rhacophor*" OR "mantellidae") AND (EXCLUDE(PUBYEAR, 2022))
Web of Science (core collection)	TS=("temperature*" OR "thermal" OR "cold*" OR "cool*") AND TS=("cold tolerance*" OR "tolerance* to cold" OR "thermal min*" OR "CTmin" OR "CT min" OR "chill coma" OR "cold stress tolerance*" OR "tolerance to cold stress" OR "cold stupor" OR "cold resistance" OR "resistance to cold stress" OR "supercooling point" OR "SCP" OR "crystal* temperature*" OR "cold hardness" OR "freez* tolerance" OR "tolerance to freezing" OR "preferred temperature*" OR "preferred body temperature*" OR "temperature preference*" OR "selected temperature*" OR "selected body temperature*" OR "thermal prefer*" OR "temperature* prefer*" OR "temperature* select*" OR "thermal selection") AND TS=("amphibia*" OR "frog*" OR "toad*" OR "salamand*" OR "newt" OR "newts" OR "tadpole*" OR "metamorph" OR "metamorphs" OR "caecili*" OR "rhinatrema*" OR "ichthyophi*" OR "scolecomorph*" OR "chikil*" OR "herpelidae" OR "typhlonect*" OR "indotyphlid*" OR "dermophi*" OR "siphonop*" OR "caudata" OR "urodela" OR "cryptobranch*" OR "hynobiid*" OR "sirenidae" OR "ambystoma*" OR "dicamptodon*" OR "proteidae" OR "rhyacotriton*" OR "amphium*" OR "plethodon*" OR "anura*" OR "ascaph*" OR "leiopelma*" OR "bombina*" OR "alyt*" OR "rhinophryn*" OR "pipidae" OR "xenopus" OR "scaphiop*" OR "pelodyt*" OR "megophry*" OR "pelobat*" OR "heleophryn*" OR "calyptocephalell*" OR "myobatrach*" OR "rhinoderma*" OR "alsod*" OR "hylod*" OR "batrachyl*" OR "cycloramph*" OR "telmatob*" OR "ceratophry*" OR "hemiphract*" OR "hyla*" OR "hylidae" OR "bufo*" OR "leptodactyl*" OR "odontophryn*" OR "allophryn*" OR "centrolen*" OR

	"dendrobat*" OR "ceuthomanti*" OR "eleutherodactyl*" OR "brachycephalidae" OR "craugastor*" OR "strabomantidae" OR "pristimantis" OR "nasikabatrach*" OR "soogloss*" OR "microhyl*" OR "arthroleptid*" OR "hyperol*" OR "brevicipitidae" OR "hemisus" OR "odontobatrach*" OR "phrynobatrach*" OR "ptychaden*" OR "conraua" OR "petropedet*" OR "pyxicephal*" OR "micrixalus" OR "nyctibatrach*" OR "ranixalidae" OR "ceratobatrach*" OR "dicroglossidae" OR "rana" OR "ranidae" OR "rhacophor*" OR "mantellidae") NOT PY=(2022)
Lens	("temperature*" OR "thermal" OR "cold*" OR "cool*") AND ("cold tolerance*" OR "tolerance* to cold" OR "thermal min*" OR "CTmin" OR "CT min" OR "chill coma" OR "cold stress tolerance*" OR "tolerance to cold stress" OR "cold stupor" OR "cold resistance" OR "resistance to cold stress" OR "supercooling point" OR "SCP" OR "crystal* temperature*" OR "cold hardiness" OR "freez* tolerance" OR "tolerance to freezing" OR "preferred temperature*" OR "preferred body temperature*" OR "temperature preference*" OR "selected temperature*" OR "selected body temperature*" OR "thermal prefer*" OR "temperature* prefer*" OR "temperature* select*" OR "thermal selection") AND ("amphibia*" OR "frog*" OR "toad*" OR "salamand*" OR "newt" OR "newts" OR "tadpole*" OR "metamorph" OR "metamorphs" OR "caecili*" OR "rhinatrema*" OR "ichthyophi*" OR "scolecomorph*" OR "chikil*" OR "herpelidae" OR "typhlonect*" OR "indotyphlid*" OR "dermophi*" OR "siphonop*" OR "caudata" OR "urodela" OR "cryptobranch*" OR "hynobiid*" OR "sirenidae" OR "ambystoma*" OR "dicamptodon*" OR "proteidae" OR "rhyacotriton*" OR "amphium*" OR "plethodon*" OR "anura*" OR "ascaph*" OR "leiopelma*" OR "bombina*" OR "alyt*" OR "rhinophryn*" OR "pipidae" OR "xenopus" OR "scaphiop*" OR "pelodyt*" OR "megophry*" OR "pelobat*" OR "heleophryn*" OR "calyptocephalell*" OR "myobatrach*" OR "rhinoderma*" OR "alsod*" OR "hylod*" OR "batrachyl*" OR "cycloramph*" OR "telmatob*" OR "ceratophry*" OR "hemiphract*" OR "hyla*" OR "hylidae" OR "bufo*" OR "leptodactyl*" OR "odontophryn*" OR "allophryn*" OR "centrolen*" OR "dendrobat*" OR "ceuthomanti*" OR "eleutherodactyl*" OR "brachycephalidae" OR "craugastor*" OR "strabomantidae" OR "pristimantis" OR "nasikabatrach*" OR "soogloss*" OR "microhyl*" OR "arthroleptid*" OR "hyperol*" OR "brevicipitidae" OR "hemisus" OR "odontobatrach*" OR "phrynobatrach*" OR "ptychaden*" OR "conraua" OR "petropedet*" OR "pyxicephal*" OR "micrixalus" OR "nyctibatrach*" OR "ranixalidae" OR "ceratobatrach*" OR "dicroglossidae" OR "rana" OR "ranidae" OR "rhacophor*" OR "mantellidae") Year Published = (1900 - 2021) Field of Study = (excl Botany , excl Physics , excl Geology , excl Materials science , excl Context (language use) , excl Geochemistry , excl Internal medicine , excl Mutant , excl Arabidopsis , excl Condensed matter physics , excl Computer science , excl Biophysics , excl Biotechnology , excl Computational biology , excl Horticulture , excl Aquaporin , excl Crop , excl Mineralogy , excl Nanotechnology , excl Arabidopsis thaliana , excl Chemical physics)
Proquest (Dissertation and Theses)	(noft(cold tolerance*) OR noft(CTmin*) OR noft(preferred temperature*) OR noft(selected temperature*)) AND (noft(amphibia*) OR noft(frog*) OR noft(toad*) OR noft(anura*) OR noft(tadpole*) OR noft(salamand*) OR noft(newts))
Google Scholar (French)	("température préférée" OR "température sélectionnée" OR "température choisie" OR "préférences thermiques") AND (amphibiens OR grenouille OR crapaud OR salamandres OR triton OR têtards OR Amphibia OR Caudata OR Anura OR batracien OR anoure) CTmin AND ("amphibiens" OR grenouille OR crapaud OR "salamandres" OR triton OR têtards OR batracien OR anoure) (Tpref OR Tsel) AND ("amphibiens" OR grenouille OR crapaud OR "salamandres" OR triton OR têtards OR batracien OR anoure)

Google Scholar (Japanese)	(好適温度 OR 選択温度 OR 温度嗜好性) AND (両生類 OR カエル OR ヒキガエル OR サンショウウオ OR イモリ OR おたまじゃくし OR “Amphibia” OR “Caudata” OR “Anura”) CTmin AND (両生類 OR カエル OR ヒキガエル OR サンショウウオ OR イモリ OR オタマジャクシ) (Tpref OR Tsel) AND (両生類 OR カエル OR ヒキガエル OR サンショウウオ OR イモリ OR オタマジャクシ)
Google Scholar (Portuguese)	(“temperatura preferida” OR “temperatura selecionada” OR “preferência termal”) AND (anfíbio OR “rã” OR sapos OR salamandra OR tritão OR girino OR Amphibia OR Caudata OR Anura OR anuros) CTmin AND (anfíbio OR “rã” OR sapos OR salamandra OR tritão OR girino) (Tpref OR Tsel) AND (anfíbio OR “rã” OR sapos OR salamandra OR tritão OR girino)
Google Scholar (simplified Chinese)	(合适温度 OR 选温度 OR 耐热程度) AND (两栖动物 OR 青蛙 OR 蛤蟆 OR 蝾螈 OR 蝌蚪 OR 小鲵 OR 大鲵 OR Amphibia OR Caudata OR Anura) CTmin AND (两栖动物 OR 青蛙 OR 蛤蟆 OR 蝾螈 OR 蝌蚪 OR 小鲵 OR 大鲵) (Tpref OR Tsel) AND (两栖动物 OR 青蛙 OR 蛤蟆 OR 蝾螈 OR 蝌蚪 OR 小鲵 OR 大鲵)
Google Scholar (traditional Chinese)	(偏好温度 OR 溫度選擇 OR 熱偏好) AND (兩棲類 OR 蛙青蛙 OR 蟾蜍蝾螈 OR 蝾螈 OR 蝌蚪 OR 小鲵 OR 大鲵 OR Amphibia OR Caudata OR Anura) CTmin AND (兩棲類 OR 蛙青蛙 OR 蟾蜍蝾螈 OR 蝾螈 OR 蝌蚪 OR 小鲵 OR 大鲵 OR 小鲵山椒魚 OR 大鲵娃娃魚) (Tpref OR Tsel) AND (兩棲類 OR 蛙青蛙 OR 蟾蜍蝾螈 OR 蝾螈 OR 蝌蚪 OR 小鲵 OR 大鲵 OR 小鲵山椒魚 OR 大鲵娃娃魚)
Google Scholar (Spanish)	(“temperatura preferida” OR “temperatura seleccionada” OR “preferencias térmicas”) AND (anfíbio OR rana OR sapo OR salamandra OR tritón OR renacuajo OR Amphibia OR Caudata OR Anura OR anuros) CTmin AND (anfíbio OR rana OR sapo OR salamandra OR tritón OR renacuajo or Anuros) (Tpref OR Tsel) AND (anfíbio OR rana OR sapo OR salamandra OR tritón OR renacuajo or Anuros)

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45 **Table S2: Metadata table and column descriptions.**

Data	Description
unique_ID	Unique identifier for each row in the data.
name	Name of the researcher who performed the data extraction.
ref	Abbreviated reference for the study.
title	Title of the paper or thesis.
pub_year	Publication year of the paper or thesis.
thesis_chapter	If the study is a thesis, the chapter the data is taken from (e.g., 2).
chapter_title	The title of the thesis chapter the data is taken from.
peer-reviewed	Whether the study was peer-reviewed or not (i.e., thesis). Factor with two levels: “peer-reviewed”, “not_peer-reviewed”.
doi	DOI of the paper.
language	Language of the paper (main text). Factor with seven levels: “English”, “traditional Chinese”, “simplified Chinese”, “French”, “Japanese”, “Portuguese”, “Spanish”.
population_ID	Unique identifier for each population. Note that populations were considered individuals of the same species taken from different geographical locations. For studies without geographical coordinates, populations were assigned based on descriptions made by the authors (e.g., “Northern population” vs. “Southern population”). Note that when multiple data points were extracted from the same population, they shared the same population_ID. The population_ID is structured as a combination of the species name and the population number (e.g., <i>Eupemphix nattereri</i> _pop1).
cohort_ID	Unique identifier for each cohort. By “cohort”, we refer to independent groups of animals. In some cases, traits were measured multiple times on the same cohort of animals (e.g., using different endpoints, or at different life stages). As such, the same cohort_ID was assigned to repeated measures. Note that cohort_ID was assigned at the trait-level; as it was not always possible to assign whether multiple traits (e.g., CTmin and CTmax) were measured with the same, or independent groups of animals.
notes_ID	General notes related to population_ID and cohort_ID.
order	Species order, according to Jetz and Pyron (2018).
family	Species family, according to Jetz and Pyron (2018).
species	Species name, according to Jetz and Pyron (2018).
strain	The strain, variety, subspecies, or morph of the species, as reported in the study.

IUCN_status	International Union for the Conservation of Nature (IUCN) threat status. Factor with 7 levels: “DD”, “LC”, “NT”, “VU”, “EN”, “CR” and “EX”, for “data-deficient”, “least-concern”, “near threatened”, “vulnerable”, “endangered”, “critically endangered” and “extinct”, respectively.
origin	Origin of studied animals. Factor with four levels: recently collected from the wild (i.e., “wild”), eggs laid in the laboratory (i.e., “lab”), animals provided from a supplier (i.e., “supplier”) or “unclear”. For studies collecting eggs from the wild and testing the same generation of animals, animals were considered as “wild”.
n_generations_lab	Number of generations spent in the laboratory, if reported in the study. For “wild” animals, this was left as NA.
latitude	Latitude from which animals were collected (decimal degrees). Latitudes presented in degrees/minutes/seconds were converted to decimal degrees. When geographical coordinates were not presented, the coordinates were estimated using Google Maps.
longitude	Longitude from which animals were collected (decimal degrees). Longitudes presented in degrees/minutes/seconds were converted to decimal degrees. When geographical coordinates were not presented, the coordinates were estimated using Google Maps.
elevation	Elevation from which animals were collected (meters above sea level), as reported in the study. When not reported, elevation was estimated using latitude and longitude and freemaptools.com.
date_sampling	Date at which the animals were sampled (format YEAR/MONTH/DAY, e.g. “2020/07/26”). If the precise day is not reported, the month was reported in “month_sampling”. If a range of dates was presented (e.g., “June to August 2020”), details were reported in the “start_range_sampling_dates” and “end_range_sampling_dates” columns.
month_sampling	Month from which the animals were collected. If the sampling spans two or more months, details were reported in the “start_range_sampling_dates” and “end_range_sampling_dates” columns.
year_sampling	Year from which the animals were collected. If the sampling spanned two years, details were reported in the “start_range_sampling_dates” and “end_range_sampling_dates” columns.
start_range_sampling_dates	The beginning of the range of dates over which animals were collected. Indicated are both the month and the year of collection (e.g., “January_2015”).

end_range_sampling_dates	The end of the range of dates over which animals were collected. Indicated are both the month and the year of collection (e.g., “September_2015”).
notes_sampling	General notes regarding the sampling of the animals.
ambient_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean ambient temperature (°C) in the month of collection, if reported in the study. If animals were collected over a range of months, the mean temperature across this sampling period was reported.
substrate_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean temperature of the substrate (°C) in the month of capture. If animals were collected over a range of months, the mean temperature across this sampling period was reported.
water_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean water temperature (°C) in the month of collection. If animals were collected over a range of months, the mean temperature across this sampling period was reported.
field_body_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean body temperature (°C) measured in the field when animals were collected. If animals were collected over a range of months, the mean temperature across this sampling period was reported.
notes_env_temp	General notes regarding the sampling of animals in the field.
acclimated	Whether the animals were maintained in the laboratory for >12h or tested shortly after collection. Factor with two levels: “acclimated” or “field-fresh”.
incubation_temp	For animals born in the laboratory, the mean temperature (°C) at which the embryos were incubated, if reported in the study. For the temperature at which the larvae and adults were acclimated, see “acclimation_temp”.
sd_incubation_temp	Variability (e.g., +/- 1C) in incubation_temp (°C), if reported.
life_stage_acclimated	For acclimated animals, the life stage acclimated prior to the upper thermal limit assessment. Factor with five levels: “embryos_and_larvae”, “larvae”, “juveniles”, “metamorphs” or “adults”. Larval stages of salamanders and tadpoles were referred to as “larvae”. Animals between Gosner stages 42 and 45 were considered “metamorphs”, while those between Gosner stage 45 and sexual maturity were considered “juveniles”.

gosner_acclimated	For acclimated animals, the Gosner stage when the acclimation started, if reported in the study. If a range of Gosner stage was presented (24-27), the mean of the range was taken.
acclimation_temp	For acclimated animals, the mean temperature of acclimation (°C). Note that “acclimation” refers to a prolonged (>12h) exposure to a new temperature. Therefore, cold/heat shocks or housing conditions just prior to assessing thermal tolerance or preference (e.g., 2 hours at 25°C) were not considered as “acclimation” conditions. If animals were exposed to multiple acclimation conditions (e.g., 15°C for 1 month, and then re-acclimated to 25°C for 7 days), we took the latest acclimation condition as the “acclimation_temp”.
sd_acclimation_temp	Variability (e.g., +/- 1C) in acclimation_temp (°C), if reported in the study.
acclimation_time	The duration of acclimation (days).
notes_acclimation	General notes regarding the laboratory acclimation of animals.
life_stage_tested	The life stage tested for thermal tolerance or preference. Factor with four levels: “larvae”, “metamorphs”, “juveniles” or “adults”. Larval stages of salamanders and tadpoles were referred to as “larvae”. Animals between Gosner stages 42 and 45 were considered “metamorphs”, while those between Gosner stage 45 and sexual maturity were considered “juveniles”.
gosner_tested	Gosner stage when the animals were assessed for heat tolerance, if reported in the study. If a range of Gosner stage was presented (24-27), the mean of the range was taken.
SVL	Mean snout-vent length of the animals (mm) when assessed for thermal tolerance or preference, if reported in the study. Note that SVL data was often taken from Rohr et al. (2018).
body_mass	Mean body mass of the animals (g) when assessed for thermal tolerance or preference.
age_tested	The age (days-post-hatching) at which the animals were tested for thermal tolerance or preference.
sex	The sex of the animals. Factor with four levels: “male”, “female”, “mixed”, “unknown”. The “mixed” category was used when authors clearly stipulate that they mixed males and females.
metric	The metric used to assess thermal tolerance (CTmax, LT50_hot, CTmin, LT50_cold) or preference (Tpref). Factor with five levels: “CTmax”, “LT50_hot”, “CTmin”, “LT50_cold”, “Tpref”.
endpoint	The endpoint that was used for assessing thermal tolerance (loss or righting response, loss of equilibrium, onset of spasms, no response to prodding, supercooling point, death). Factor with seven levels:

	“LRR”, “LOE”, “OS”, “prodding”, “SCP”, “death”, “other”. If “other”, details are reported in “notes_test” (see below).
medium_test_temp	Whether the temperature measured during the test was the ambient, the water, the substrate, or the body temperature. Factor with three levels: “ambient”, “substrate”, “water”, “body”.
start_temp	If the metric was CTmin or CTmax, the starting temperature used in the thermal tolerance assay (°C).
ramping	If the metric was CTmin or CTmax, the ramping (cooling or heating) rate applied to the animals (°C/min).
set_time	If the metric was LT50, the time the animals spent at the test temperature (the time after which the animals the survival was assessed, in hours). If the authors report e.g., 96h-LT50, then set_time would be 96.
n_test_temp	If the metric was LT50, the number of temperatures tested to assess upper thermal limits. E.g., if authors measured survival at 36, 38, 39, and 41°C, n_test_temp = 4.
n_replicates_per_temp	If the metric was LT50, the number of replicates used at each test temperatures. E.g., if authors used 5 test temperatures and measured the survival of three independent cohorts of animals at each test temperature, then n_replicates_per_temp = 3.
n_animals_per_replicate	If the metric was LT50, the number of animals in each replicate.
duration_measurement	If the metric was Tpref, the duration of the assay to measure thermal preference (hours).
rate_measurement	If the metric was Tpref, the rate at which body temperature was measured (measurements/hour).
gradient_type	If the metric was Tpref, the type of thermal gradient used. Factor with two levels: “linear”, “shuttlebox”.
gradient_low_temp	Lowest temperature in the thermal gradient (°C).
gradient_high_temp	Highest temperature in the thermal gradient (°C).
notes_test	General notes regarding the thermal tolerance or preference assays.
humidity	Humidity at which animals were acclimated or tested (% relative humidity). If the humidity during the acclimation and the test were different, priority was given to the conditions of the test.
oxygen	Oxygen at which animals were acclimated or tested (mg.L ⁻¹ dissolved oxygen). If the oxygen concentration during the acclimation and the test were different, priority was given to the conditions of the test.
salinity	Salinity at which animals were acclimated or tested (parts per thousands). If the salinity during the acclimation and the test were different, priority was given to the conditions of the test.

pH	pH at which animals were acclimated. If the pH during the acclimation and the test were different, priority was given to the conditions of the test.
photoperiod	Photoperiod at which animals were acclimated (number of hours of light per day).
chemical	If any, which chemical (e.g., pollutant, toxin) was added to the animals' environment. If animals were in a control group (i.e., only supplemented with a solvent), "control" was indicated.
hormone	If any, which hormone (e.g., corticosterone, thyroid hormone) was added to the animals' environment. If animals were in a control group (i.e., only supplemented with a solvent), "control" was indicated.
concentration_chemical_hormone	If any, the concentration of the hormones or chemicals used. If animals were in a control group, "0" was indicated.
unit_chemical_hormone	The unit used to quantify the chemical or hormonal concentration administered (e.g., g/L, ng/g of sediment).
infected	Whether the animals were infected with a pathogen. Infected animals were marked as "infected". Otherwise, this field was left blank.
pathogen	If the animals were infected with a pathogen, the name of the pathogen (e.g., <i>Batrachochytrium dendrobatidis</i>).
notes_supplements	General notes regarding the addition of chemicals, hormones, or pathogens.
data_source	Where the upper thermal limit data is reported (main text, table, figure, published data).
data_url	If the data was published in a repository, the url link to the repository containing the data.
flag	Whether the data presents procedural concerns. The exact issue is listed in this column. Note that these observations were removed from the curated version of the database.
mean_trait	Mean thermal tolerance or preference (°C).
error_trait	Standard deviation or standard error of mean_trait (see error_type)
n_trait	Sample size of mean_trait. When the metric was LT50, the sample size was taken as the number of test temperatures ("n_test_temp") * the number of replicates per test temperature (n_replicates_per_temp).
error_type	Whether the error is presented as standard deviations (i.e., "sd") or standard errors (i.e., "se").
notes_trait	General notes about thermal tolerance or preference estimates

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TABLE S3: Inclusion criteria used to screen abstracts, titles and keywords. Numbers match those used in in Figure S1 (decision tree).

	Description
1	Studies not published in French, Japanese, Portuguese, simplified Chinese, traditional Chinese or Spanish were excluded.
2	We only included empirical studies presenting original data. Therefore, we excluded reviews, syntheses, simulations, theoretical studies, and conference abstracts, unless supplemented with original data.
3	“Amphibians” refer to frogs, toads, salamanders, newts, and caecilians. We only included studies on whole organisms.
	Desired measures of cold tolerance included the:
	i) critical thermal minimum (CT _{min}), where animals are subject to incremental decreases in temperature until an endpoint (e.g. loss of righting response, onset of spasms, supercooling point, crystallisation) is reached;
	ii) the temperature lethal for 50% of the animals (LT ₅₀ ; sometimes referred to as the “incipient lethal temperature”), where survival is recorded after animals are abruptly transferred to a set of cold temperatures for a given period of time (e.g. 24 hours) and LT ₅₀ is interpolated from the survival curve; and
	iii) the death time or chill coma time where animals are abruptly transferred to cold temperatures and the time needed for animals to reach an endpoint (e.g. immobilisation, death) is recorded as the response. With the latter measure, the thermal tolerance limit can be inferred from the relationship between chill coma time and the temperature of the knockdown assay. Therefore, cold knockdown times must have been measured at >2 temperatures (e.g. chill coma times at 10, 15, and 17°C).
4	We exclude alternative measures of cold tolerance which cannot be converted to the temperature scale (e.g. chill coma recovery time) or CT _{min} extrapolated from physiological performance curves (e.g. critical temperature for ATPase activity).
	Desired measures of preferred temperature were:
	i) where animals are placed in a temperature gradient and the body temperature of the animals is measured at regular intervals, or inferred from photography. The mode or median body temperatures animal select is usually defined as the preferred (or selected) body temperature.
	ii) where animals are placed in an experimental set up with levers that trigger the warming or cooling of the surface or experimental chamber (shuttlebox). Similarly to above, the body temperatures of animals is tracked, and the mode or median body temperature animals experienced is usually defined as the preferred (or selected) body temperature.
5	We focused our search on juveniles (i.e., tadpole, metamorph, froglet) or adults. Hence, we excluded studies only measuring the cold tolerance of embryos.

TABLE S4: Inclusion criteria used to assess full articles for eligibility. Numbers match those used in in Figure S2 (decision tree).

	Description
1	Studies not published in French, Japanese, Portuguese, simplified Chinese, traditional Chinese or Spanish were excluded.
2	We only included empirical studies presenting original data. Therefore, we excluded reviews, syntheses, simulations, theoretical studies, and conference abstracts, unless supplemented with original data.
3	Amphibians” refer to frogs, toads, salamanders, newts and caecilians. We only included studies on juveniles (i.e. tadpole, metamorph, froglet) or adults. Hence, we exclude studies only measuring the cold tolerance of embryos.
	Desired measures of cold tolerance included the: iv) critical thermal minimum (CT_{min}), where animals are subject to incremental decreases in temperature until an endpoint (e.g. loss of righting response, onset of spasms, supercooling point, crystallisation) is reached; v) the temperature lethal for 50% of the animals (LT₅₀; sometimes referred to as the “incipient lethal temperature”), where survival is recorded after animals are abruptly transferred to a set of cold temperatures for a given period of time (e.g. 24 hours) and LT₅₀ is interpolated from the survival curve; and vi) the death time or chill coma time where animals are abruptly transferred to cold temperatures and the time needed for animals to reach an endpoint (e.g. immobilisation, death) is recorded as the response. With the latter measure, the thermal tolerance limit can be inferred from the relationship between chill coma time and the temperature of the knockdown assay. Therefore, cold knockdown times must have been measured at >2 temperatures (e.g. chill coma times at 10, 15, and 17°C). We exclude alternative measures of cold tolerance which cannot be converted to the temperature scale (e.g. chill coma recovery time) or CT_{min} extrapolated from physiological performance curves (e.g. critical temperature for ATPase activity). Desired measures of preferred temperature were: iii) where animals are placed in a temperature gradient and the body temperature of the animals is measured at regular intervals, or inferred from photography. The mode or median body temperatures animal select is usually defined as the preferred (or selected) body temperature. iv) where animals are placed in an experimental set up with levers that trigger the warming or cooling of the surface or experimental chamber (shuttlebox). Similarly to above, the body temperatures of animals is tracked, and the mode or median body temperature animals experienced is usually defined as the preferred (or selected) body temperature.
5	To be included, the study must have reported the temperature at which animals were maintained in the laboratory (i.e. temperature of acclimation), the temperature of the environment from which animals were captured (i.e. temperature of acclimatization), or the geographical coordinates and dates of capture.

TABLE S5: Summary of procedural concerns found in some studies. Note that estimates having procedural concerns were excluded during the data curation (see main text).

Procedural concerns	Number of estimates concerned
Data from a single individual	70
Uncommon or inconsistent methodology	55
Unclear/uncommon acclimation conditions	53
Animals were starved prior to testing	34
Animals underwent surgery or amputation	16
Highly uncertain estimates	14
Animals were dehydrated prior to testing	14
Animals were exposed to hypoxic or hypercapnic conditions	13
Animals were exposed to high levels to UV radiation	5
Animals were perfused with pH solution	3
Statistical dispersion and sample sizes not reported	3
Animals were exposed to predators	2