

1 ***AmphiTherm: a comprehensive database of amphibian thermal tolerance***
2 ***and preference***

3 **Authors**

4 Patrice Pottier^{1,2*}, Rachel R.Y. Oh^{3,4,5}, Pietro Pollo^{1,6}, A. Nayelli Rivera-Villanueva^{5,7,8}, Yefeng Yang¹,
5 Sarah Varon⁹, Ana V. Longo⁹, Samantha Burke¹, Hsien-Yung Lin¹⁰, José O. Valdebenito^{11,12}, Tatsuya
6 Amano^{5,8}, Szymon M. Drobniak^{1,13}, Shinichi Nakagawa^{1,14}, and Natalie Claunch^{9,15,16*}

7 *Corresponding authors

8 Corresponding authors: Pottier P. (patrice.pottier37@gmail.com), Claunch N.

9 (nat.claunch@gmail.com)

10 **Affiliations**

11 ¹Evolution & Ecology Research Centre, School of Biological, Earth and Environmental Sciences, The
12 University of New South Wales, Sydney, New South Wales, Australia

13 ²Division of Ecology and Evolution, Research School of Biology, The Australian National University,
14 Canberra, Australian Capital Territory, Australia

15 ³German Centre for Integrative Biodiversity Research, Halle-Jena-Leipzig, Leipzig, Germany

16 ⁴Helmholtz Centre for Environmental Research (UFZ), Leipzig, Germany

17 ⁵Centre for Biodiversity and Conservation Science, The University of Queensland, Brisbane,
18 Queensland, Australia

19 ⁶School of Environmental and Life Sciences, University of Newcastle, Callaghan, New South Wales,
20 Australia

21 ⁷Laboratorio de Biología de la Conservación y Desarrollo Sustentable de la Facultad de Ciencias
22 Biológicas, Universidad Autónoma de Nuevo León, Monterrey, México

23 ⁸School of the Environment, The University of Queensland, Brisbane, Queensland, Australia

24 ⁹Department of Biology, University of Florida, Gainesville, Florida, USA

25 ¹⁰Canadian Wildlife Service, Environment and Climate Change Canada, Gatineau, Quebec, Canada.

26 ¹¹Facultad de Medicina Veterinaria y Agronomía, Campus Chacabuco, Universidad de Las Américas,
27 Chile

28 ¹²Instituto Milenio Biodiversidad de Ecosistemas Antárticos y Subantárticos (BASE), Chile

29 ¹³Institute of Environmental Sciences, Faculty of Biology, Jagiellonian University, Krakow, Poland

30 ¹⁴Department of Biological Sciences, University of Alberta, Edmonton, Canada

31 ¹⁵Department of Natural History, Florida Museum of Natural History, Gainesville, Florida, USA

32 ¹⁶USDA APHIS WS National Wildlife Research Center, Florida Field Station, Gainesville, Florida,
33 USA

34 **ORCID**

- 35 Patrice Pottier <https://orcid.org/0000-0003-2106-6597>
- 36 Rachel R. Y. Oh <https://orcid.org/0000-0003-2716-7727>
- 37 Pietro Pollo <https://orcid.org/0000-0001-6555-5400>
- 38 A. Nayelli Rivera-Villanueva <https://orcid.org/0000-0002-9190-4317>
- 39 Yefeng Yang <https://orcid.org/0000-0002-8610-4016>
- 40 Ana V. Longo <https://orcid.org/0000-0002-5112-1246>
- 41 Samantha Burke <https://orcid.org/0000-0001-6902-974X>
- 42 Hsien-Yung Lin <https://orcid.org/0000-0002-2564-3593>
- 43 José O. Valdebenito <https://orcid.org/0000-0002-6709-6305>
- 44 Tatsuya Amano <https://orcid.org/0000-0001-6576-3410>
- 45 Szymon M. Drobniak <https://orcid.org/0000-0001-8101-6247>
- 46 Shinichi Nakagawa <https://orcid.org/0000-0002-7765-5182>
- 47 Natalie Claunch <https://orcid.org/0000-0003-3144-4192>
- 48

49 **Abstract**

50 Thermal traits are crucial to our understanding of the ecology and physiology of ectothermic animals.
51 While rising global temperatures have increasingly pushed research towards the study of upper thermal
52 limits, lower thermal limits and thermal preferences are essential for defining the thermal niche of
53 ectotherms. Through a systematic review of the literature in seven languages, we expanded an existing
54 database of amphibian heat tolerance by adding 1,009 estimates of cold tolerance and 816 estimates of
55 thermal preference across 375 species. *AmphiTherm* is a comprehensive and reproducible database that
56 contains 4,899 thermal trait estimates from a diverse sample of 659 species (~7.5% of all described
57 amphibians) spanning 38 families. Despite its broad geographic coverage, we report evident gaps across
58 amphibian biodiversity hotspots in Africa, most regions of Asia, central South America, and Western
59 Australia. By providing a more holistic understanding of amphibian thermal tolerance and behavioural
60 preferences, *AmphiTherm* is a valuable resource for advancing research in evolutionary biology,
61 ecophysiology, and biogeography, offering insights that are increasingly needed in a changing climate.

62

63 **Background & Summary**

64 Thermal trait data are crucial to our understanding of the biology and physiology of ectotherms. The
65 recent increase in broad-scale syntheses of ectotherm thermal physiologies demonstrates recurring
66 interest in how these organisms respond to changing thermal environments^{1–10}. Much of this work has
67 focused on traits relating to heat tolerance, reflecting the urgency to predict the impacts of climate
68 warming on natural populations^{11–17}. However, climate change also brings an increased probability of
69 extreme weather events, including negative temperature anomalies^{18,19}. As such, a sole focus on heat
70 tolerance provides an incomplete picture of ectotherm responses to climate change. A comprehensive
71 understanding of heat tolerance, cold tolerance, and thermal preference is necessary to fully define
72 ectotherm's thermal niches and predict their responses to climate change. Below, we briefly emphasise
73 the importance of these thermal traits for amphibians (see ^{8,20,21} for more in depth discussion).

74 While the significance of heat tolerance in predicting species' responses to warming climates
75 is well documented¹, data on lower thermal limits are equally vital yet often understudied, especially in
76 amphibians, an at-risk, data-deficient group of ectotherms²². Lower thermal limits represents the lower
77 boundary of an organism's thermal niche and have been included in several data syntheses^{5,6,8,10}. This
78 trait provides key insights into how species might respond to increasing frequency of extreme cold
79 weather events, which can lead to significant population reduction events known as winterkills^{18,19,23}.
80 Gaining understanding of lower thermal limits can thus help predict the sensitivity and resilience of
81 amphibian populations to extreme cold weather events. Moreover, data on lower thermal limits can
82 inform conservation and management strategies, for instance, by identifying microhabitats buffering
83 the effects of extreme cold on activity and survival²⁴.

84 Preferred body temperatures reflect the temperature optimising overall performance and the
85 most favourable microhabitat in the absence of other biotic and abiotic factors^{25,26}. Knowledge of
86 preferred body temperatures can thus help predict how climate change will affect species distributions
87 and activity patterns^{24,27–29}. In particular, thermal preference data can be used to infer behavioural
88 thermoregulation patterns and the microhabitats available for crucial physiological processes^{30–32}. While
89 upper thermal limits can help predict acute survival in the face of extreme heat, gradual warming below
90 the upper thermal limit thresholds can make some areas unsuitable for amphibians' activity needs²⁴. On
91 the contrary, warming can benefit some amphibians in historically cooler climates by increasing activity
92 windows or reducing hibernation times³³. As such, thermal preference data can help predict the sublethal
93 effects of climate change. Thermal preferences also, for instance, affect susceptibility to pathogens^{34–36},
94 shape the composition of commensal microbes^{37,38}, and mediate interactions between host and microbial
95 communities³⁹.

96 Although investigating thermal traits separately provides important knowledge, the study of a
97 combination of thermal traits provides deeper and more comprehensive insights. A simultaneous
98 analysis of upper and lower thermal limits is particularly interesting as it provides an estimation of
99 thermal tolerance breadth²¹—a measure of the thermal envelope ectotherms can occupy in the absence
100 of other abiotic or biotic factors (e.g., competition, resource availability). When thermal preference is
101 integrated with upper and lower thermal limits, the thermal envelope gains shape, providing additional
102 insights to parameterise models and predict activity and survival in changing environments. For
103 instance, leveraging data on thermal limits and preferred temperatures can help infer past, current, and
104 future distributions of ectothermic species^{15,40–43}. Parametrising biophysical models with data on
105 thermal limits and preference now also allow more accurate predictions of overall performance, activity
106 windows^{17,44} and microhabitat heterogeneity^{17,44,45}, strengthening our ability to predict the impacts of
107 climate change on natural populations^{46,47}. From an evolutionary perspective, the integration of different

108 thermal traits can also advance our understanding of the (co)evolution of these traits, and how climate
109 change may shape evolutionary pressures on thermal tolerance and preference^{6,29,48–51}.

110 Thermal trait data can also be used to inform conservation efforts. Comparing thermal niche envelopes
111 among amphibians, their microbiota, and potential pathogens can help predict changes in the
112 microbiome and disease risk^{52–57}. Amphibians are often accessioned into captivity to establish assurance
113 colonies for breeding and eventual reintroduction⁵⁸. Exposure to environmental conditions via “soft
114 release” or mesocosms prior to reintroduction may influence survival and success of reintroduction
115 efforts^{59,60}. Knowledge of the thermal tolerance and preference of a broad range of species from different
116 habitats can help inform the design of enclosures that better simulate natural thermal variability^{61–63}. In
117 addition, understanding thermal constraints on activity, demography, and disease risk enhances our
118 ability to identify habitats suitable for repatriation and reintroduction efforts in endangered
119 species^{58,64,65}.

120 Therefore, a holistic understanding of amphibian thermal tolerance limits and preferences is essential
121 for defining the fundamental thermal niche of ectotherms and to project their activity, distribution and
122 survival in rapidly changing environments. Here, we expand an existing database on amphibian upper
123 thermal limits¹. We aggregated lower thermal limits and thermal preference data from the global
124 literature published in seven languages. In doing so, we expanded thermal niche data for 378 species,
125 providing a stronger foundation for research on amphibian ecology, evolution, and conservation.

126 Methods

127 *Reporting*

128 We reported the contributions of each author using the CRediT (Contributor Roles Taxonomy)
129 statement⁶⁶, and MeRIT (Method Reporting with Initials for Transparency) guidelines⁶⁷. We also
130 followed recommendations to maximise the indexing of titles, abstracts, and keywords in databases⁶⁸.

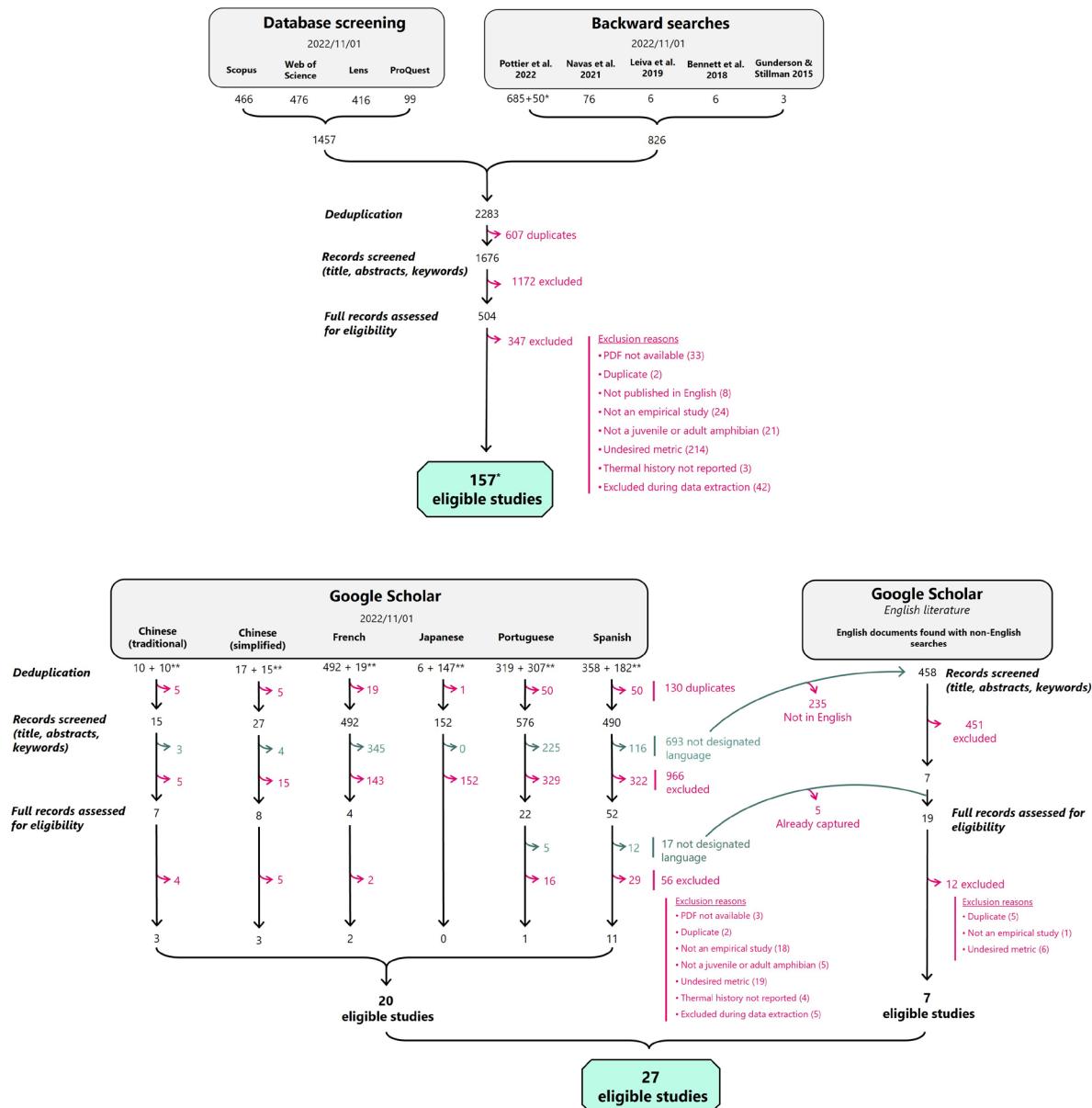
131 *Literature Searches*

132 We adapted methods from ¹ to search the literature on thermal physiological traits. We aimed to compile
133 a comprehensive and representative sample of the experimental literature on lower thermal limits and
134 thermal preference in amphibians, complementing the data on upper thermal limits compiled previously
135 (see ¹ for methods specific to upper thermal limits). PPottier accessed Scopus, ISI Web of Science (core
136 collection), Lens, and ProQuest (dissertation & theses) on 01 November 2022 using The University of
137 New South Wales’ institutional subscriptions (full search strings are available in Supplementary
138 Information (Table S1). For studies in English, PPottier modified search strings to accommodate the
139 structure of each database (Table S1) and performed backward searches of previously published reviews
140 of amphibian thermal preference and tolerance^{1,4–6,10}. This resulted in a total of 1676 unique documents.
141 We limited the timespan of our searches to 31 May 2021 to match with the timespan from ¹. This
142 decision was made to normalise all searches to a single timespan to simplify future database updates.
143 PPottier also performed Traditional and Simplified Chinese, French, Japanese, Portuguese, and Spanish
144 searches in Google Scholar using search strings translated by native speakers (NR, PPottier, PPollo,
145 SN, YY, and RRYO). The searches contained translated singular and plural forms of the following:
146 “amphibian”, “frog”, “toad”, “salamander”, “newt”, “tadpole”, “preferred temperature”, “selected
147 temperature”, “thermal preference”, “Tpref”, “Tsel”, or “CTmin”. Due to search string limitations in
148 Google Scholar (256 characters), each term was assessed in its singular or plural form, and the search
149 producing the largest number of results was selected. We performed searches following the format of
150 (“preferred temperature” OR synonyms) AND (amphibian OR synonyms). We also performed separate

151 searches with “CTmin” and (“Tpref” OR “Tsel”), as these terms are commonly used in the literature to
152 refer to lower thermal limits and preferred temperatures. We (PPottier, NC) opted not to use
153 “thermoregulation” as a synonym for “thermal preference” in our search strategy, as pilot searches
154 revealed that this term often returned studies that did not present experimentally-derived thermal
155 preference values, or studies that were already captured by the other search terms. PPottier used Publish
156 or Perish⁶⁹ to extract bibliographic records from Google Scholar. We also reused studies (** in Fig. 1)
157 from non-English searches conducted in ¹ as the key terms used successfully retrieved results on cold
158 tolerance. However, we limited the inclusion of studies to those meeting our first two criteria (i.e.,
159 studies on amphibians, and published in the targeted language) to reduce the volume of screening. We
160 acknowledge that our searches do not encompass all languages relevant to amphibian thermal
161 physiology research and invite speakers of languages not represented in the current version to contribute
162 to future updates of the database.

163 ***Eligibility criteria***

164 We considered studies that empirically tested lower thermal limits or thermal preference in wild or
165 laboratory amphibians. We only included studies on larval, juvenile, and adult amphibians, excluding
166 studies on embryonic stages due to the lack of comparable methods in embryos. For lower thermal
167 limits, we included studies that measured critical thermal minimum (CTmin)⁷⁰, median lethal
168 temperature (LT50)⁷¹, or presented data that were convertible to these metrics (e.g. % survival of cohorts
169 tested at different temperatures). CTmin represents the temperature at which a specific physiological or
170 behavioural endpoint is observed—such as the loss of righting response or a lack of response to
171 prodding—when an organism is exposed to progressively decreasing temperatures (e.g., 1°C/min). It
172 does not represent the lowest temperature an organism can tolerate, but rather the onset of functional
173 stasis, the point at which the organism is unable to move and incapable of essential survival behaviours
174 such as thermoregulation or predator evasion⁷⁰. This distinction is important, because many ectotherms
175 can recover from temperatures below their CTmin. For instance, some species can recover from freezing
176 to later resume normal function⁷². In contrast, LT50 is the temperature that is lethal to 50% of animals
177 tested and is derived through statistical interpolation from survival rates across a range of
178 temperatures⁷¹. For thermal preference, we included studies that empirically tested amphibian
179 temperature selection in a thermal gradient or shuttlebox via measures of body temperature (or inferred
180 body temperature from the position in the gradient or shuttlebox). We did not include data reported on
181 amphibian body temperatures from uncontrolled (wild) conditions because available environmental
182 temperatures were not standardised. We only included studies where thermal history (acclimatisation
183 or acclimation temperature) was reported or could be inferred from the dates and coordinates of
184 sampling. Detailed inclusion criteria and decision trees are presented in Supplementary Information
185 (Fig. S1-2, Tables S3-4). PPottier, RRO, PPollo, ANRV, YY, SV, AVL, and NC screened articles for
186 eligibility using Rayyan QCRI⁷³. This software facilitated the identification of key terms in titles,
187 abstracts, and keywords to streamline the screening process for large volumes of literature. During data
188 extraction, 47 papers were ultimately excluded for either lacking extractable data or for not complying
189 with our inclusion criteria (43 English, 1 Traditional Chinese, 3 Simplified Chinese). A total of 184
190 studies were deemed eligible for inclusion in the database. Of these, 157 were identified through formal
191 database searches (comprising one study published in simplified Chinese, and another study in French),
192 while 20 non-English studies and an additional 7 English-language studies were retrieved through
193 Google Scholar (Fig. 1). Therefore, nearly 15% (27/184) of the included studies were retrieved through
194 non-English literature searches. Our literature search methods and screening process is summarised in
195 a PRISMA flowchart⁷⁴ (Fig. 1).



196

197 **Figure 1 | PRISMA Flowchart** delineating the databases used to retrieve studies on lower thermal limits
198 and preferred body temperatures, the number of studies obtained at each stage of the screening process,
199 and the reasons for excluding studies. * Two studies published in languages other than English (French,
200 simplified Chinese) were retrieved through English searches. ** Studies from non-English searches
201 done in Google Scholar from¹. For the workflow used to obtain data on upper thermal limits, see¹.

202 **Data Extraction**

203 Data extractions were performed by PPottier (7.7% of estimates extracted), R.R.Y.O. (6.7%), PPollo
204 (5.3%), A.N.R.V (6.2%), Y.Y. (3.0%), A.V.L. (11.0%), S.V. (20.5%), and NC (45.0%). Note that these
205 values do not add to 100% because some data entries were extracted by two authors. Data were extracted
206 following the protocols described in¹. We extracted data directly from text and tables, and primarily
207 used *metaDigitise*⁷⁵ (version 1.0.1) in R⁷⁶ to extract data presented in figures (although note that some
208 authors have used WebPlotDigitizer⁷⁷ (version 4.7). When data were available in multiple formats (e.g.,
209 text and figure), we extracted it from the format with the highest resolution (e.g. data stratified by sex
210 or location rather than aggregated across species). Where possible, we extracted measures of data
211 dispersion (i.e. standard deviation, standard error) to accompany mean estimates. In cases where the

212 raw data was available, we calculated summary statistics (means, standard deviation, sample size) to
213 enhance the accuracy of the analysis. For studies reporting survival rates at different temperatures, we
214 predicted the temperature at which 50% mortality occurred using logistic regression from the *dose.p*
215 function from the MASS package⁷⁸.

216 We also extracted all additional information presented in the studies to allow investigations of the
217 sources of variation in the data and account for non-independence. We assigned identification numbers
218 to each study, and assigned unique identifiers within each study for each estimate, species, population
219 (individuals of the same species sampled from the same geographical location), and cohort (independent
220 group of individuals within a study). Additional variables included sampling coordinates, acclimation
221 temperatures, ramping rates, life stages, endpoints used to infer cold tolerance, or the duration of
222 exposure to experimental treatments. Additional notes were also taken by each researcher extracting the
223 data to facilitate technical validation. The full list of variables is described in Supplementary
224 Information (Table S2). Species names were standardised during the extraction to match
225 AmphibiaWeb⁷⁹ and further standardised to match the most comprehensive phylogenetic tree to date⁸⁰
226 (see *Data Curation*).

227 **Data Curation**

228 To ensure consistency in data extraction across all studies, PPottier and NC extensively
229 reviewed the extracted data to correct typological errors and resolve uncertainties identified during the
230 extraction process. PPottier then curated the data in R⁷⁶ (version 4.4.2), merging the newly extracted
231 data with the previously compiled dataset from ¹. This process involved standardising publication
232 information (publication year, source name) and other variables (e.g., geographical coordinates, IUCN
233 threat status⁸¹) to ensure uniformity across both datasets. PPottier also standardised species names and
234 taxonomy with phylogenetic information from ⁸⁰, which is primarily based on AmphibiaWeb⁷⁹. The
235 combined dataset comprises 324 publications⁸²⁻⁴⁰⁵. Note that 53 of these publications were taken from
236 university dissertations, and some of this work may have now been published^{e.g.,406,407}.

237 We also provide a curated version of the database (n = 4,401 estimates), where PPottier excluded data
238 with procedural inconsistencies (e.g., additional stressor, data collected from a single individual),
239 incomplete species information (e.g., *Hyloscirtus* sp.), and studies involving animals exposed to
240 toxicants, hormones, high levels of UV radiations, or infected with a pathogen. A script detailing the
241 data curation steps is available at <https://github.com/p-pottier/AmphiTherm>. This data curation step
242 removed 498 estimates from 15 studies and 45 species. However, we believe that this curated dataset
243 offers broader usability. Nevertheless, we also provide the uncurated version of *AmphiTherm* for users
244 interested in addressing more specific questions (e.g., how toxicants affect thermal tolerance and
245 preference) or identifying existing research gaps within the field.

246 **Data Records**

247 *AmphiTherm* encompasses 4,899 thermal physiological trait estimates, derived from 324
248 studies and covering 659 species across 38 families across a broad geographical coverage (Fig. 2-3).
249 This sample represents ~7.5% of all described amphibian species to date⁷⁹ (Fig. 4). According to the
250 IUCN red list⁸¹, most species (79.2%) are either not threatened or data-deficient (Fig. 2), yet 47 species
251 are classified as near threatened (NC), 43 as vulnerable (VU), 29 as endangered (EN), 14 as critically
252 endangered (CR), and one species now extinct. Considering that over 40% of amphibians are globally
253 threatened²², this suggests that research on amphibian thermal physiology is predominantly conducted
254 on non-threatened species, likely due to the invasive (or lethal) nature of some thermal tolerance

255 experiments and the associated conservation concerns for threatened species. This database contains
256 substantial within-species variation, with an average of 7.43 ± 19.5 (mean \pm s.d.) estimates per species,
257 spanning a range of 1 to 292 estimates, with species sampled from an average of 2.51 ± 3.29 populations.
258 Approximately 81% of these estimates include a measure of statistical dispersion (standard deviation,
259 standard error), facilitating their use in weighted (meta-)analyses⁴⁰⁸.

260 This database update adds thermal data for 375 species, including lower thermal limits for 300
261 amphibian species and thermal preference data for 137 amphibian species ($n = 1,825$ estimates; Fig 3-
262 4). The majority (98%) of lower thermal limit data are derived from CTmin estimates (990 estimates),
263 with roughly 2% of estimates (19 estimates) derived from lethal limits (LT50). Thermal preference data
264 represent 44% of the database update (816 estimates). This update has a relatively broad phylogenetic
265 coverage, spanning 32 families, with 19.2% of records from salamanders (Fig. 3-4).

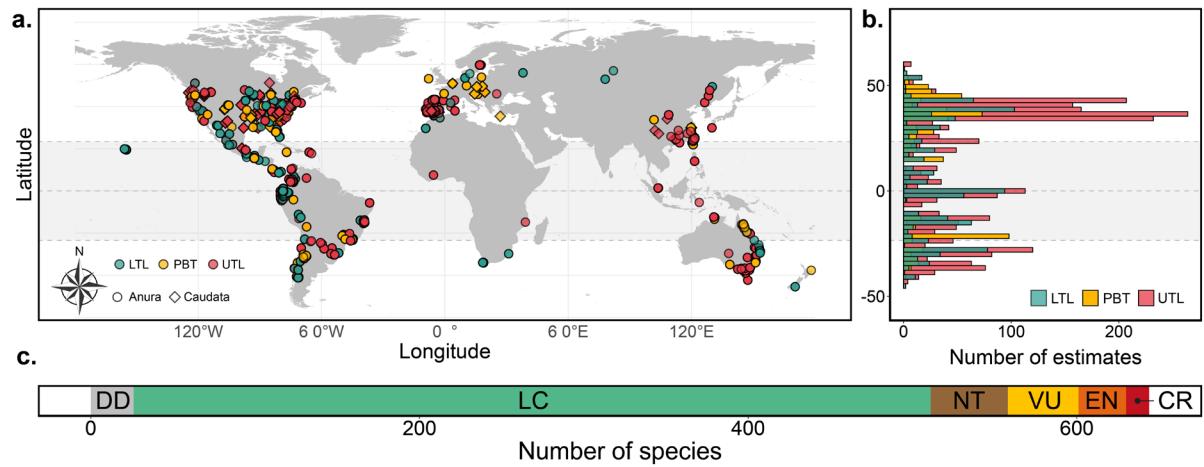
266 Approximately 62.7% of this database is comprised of upper thermal limit estimates (3074
267 estimates from 616 species and 212 studies; Fig. 3-4), highlighting a significant bias towards responses
268 to heat extremes relative to lower thermal limits (1,009 estimates, 300 species, 88 studies) and thermal
269 preferences (816 estimates, 137 species, 114 studies). We found that only 16 studies measured all three
270 thermal traits, covering 59 species (~9% of the species in the dataset; Fig. 3-4). Upper and lower thermal
271 limits were studied more frequently in tandem (60 studies), allowing to calculate the thermal tolerance
272 breadth (i.e., difference between upper and lower thermal limits) of 276 species (~42% of the species
273 in the dataset; Fig. 3-4).

274 Geographically, data were collected on all continents where amphibians occur yet exhibit a
275 strong bias towards Nearctic and European regions (Fig. 2). Large geographic gaps in thermal data are
276 evident across Africa, most regions of Asia, Western Australia, and central South America—regions that
277 are biodiversity hotspots for amphibians (Fig. 2). This is particularly concerning as they constrain our
278 understanding of how species from these underrepresented yet extremely diverse regions⁴⁰⁹ might
279 respond to climate change. We also identified taxonomic gaps in existing sampling where an entire
280 order of amphibians, Gymnophiona, remained unrepresented in the database (Fig. 4). In addition, 1 of
281 10 families of Caudata and 7 of 36 families of Anura lack thermal limits or preferred body temperature
282 estimates (Fig. 4). This suggests that further efforts are needed to broaden the research scope and better
283 represent the thermal niche of amphibians.

284 We found that the majority (88.7%) of the literature on amphibian thermal physiological traits
285 was published in English (4,343 estimates). However, non-English language literature contributes a
286 notable and important portion of the knowledge base, accounting for approximately 11.3% of the
287 data. Notably, this includes 289 estimates from publications in traditional Chinese (23 species, 7
288 studies), 131 estimates from Spanish (40 species, 11 studies), 82 estimates from simplified Chinese (12
289 species, 10 studies), 28 estimates from Portuguese (10 species, 3 studies), and 26 estimates from French
290 publications (4 species, 3 studies). Including more languages, such as Afrikaans, Arabic, Bengali,
291 Dutch, German, Hindu-Urdu, Korean, Russian, or Swahili in the screening process may help fill some
292 gaps in future updates to the database⁴¹⁰. Given the historical bias of higher impact publishing outlets
293 against studies on herpetofauna⁴¹¹, there are likely a number of studies in non-indexed journals or
294 regional journals in local languages that were not retrieved using our methods.

295 The *AmphiTherm* database is stored and archived at <https://github.com/p-pottier/AmphiTherm>.
296 These contain the metadata (.csv), raw, cleaned, and curated data (.csv), code for data curation and for
297 producing the figures (.Rmd), supplementary data (.csv) and phylogenetic tree (.tre) for producing the
298 figures, and bibliographic files (.ris and .bib) with all the references in the database. Data records are
299 under a CC-BY license, enabling reuse with attribution. Therefore, database users must cite this study

300 as well as the primary data sources to attribute the original authors and comply with copyright
301 regulations.

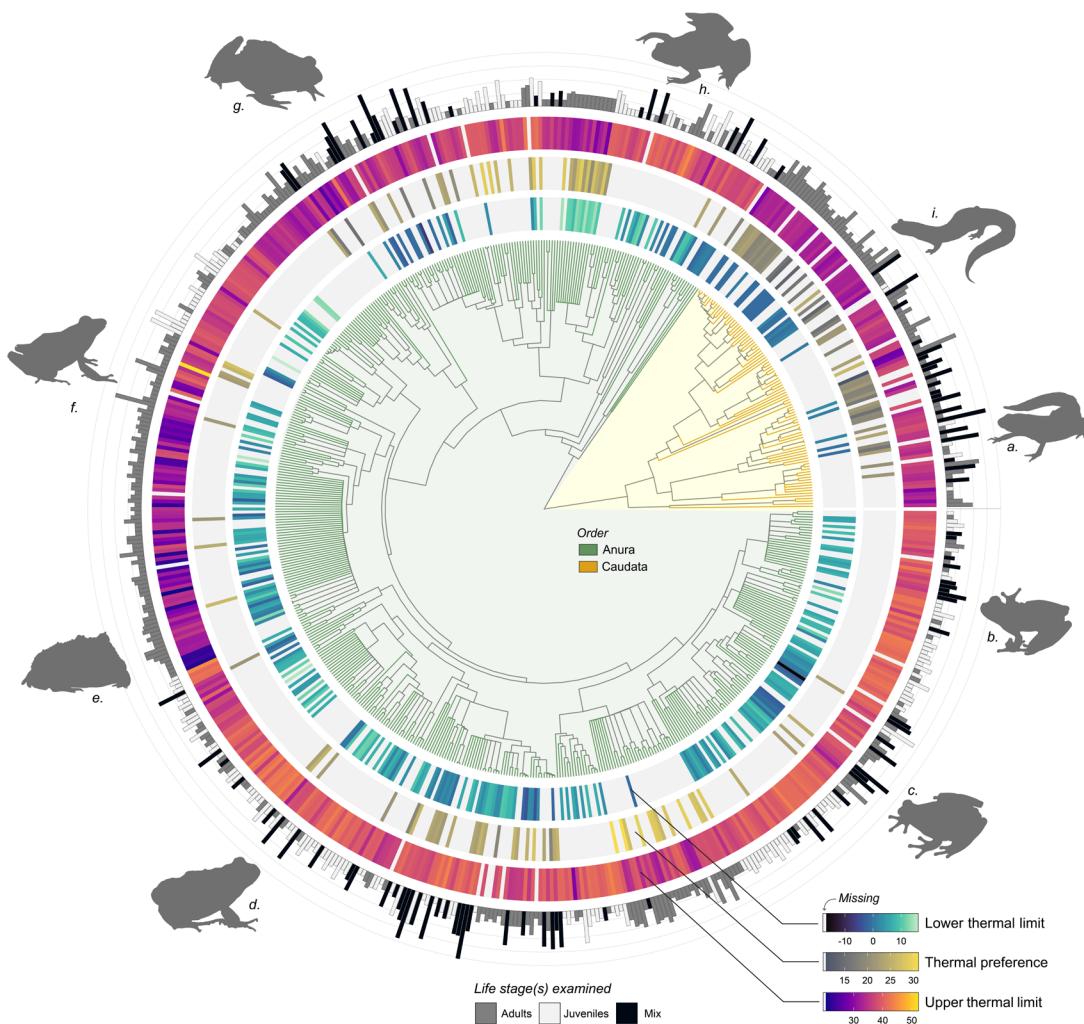


302

303 **Figure 2 |** Geographic distribution of thermal tolerance and preference data. a) World map showing the
304 distribution of lower thermal limits (LTL), preferred body temperatures (PBT), and upper thermal limits
305 (UTL) for anurans (circles) and salamanders (triangles). The shaded area represents the tropics. Note
306 that coordinates were unavailable for 775 (15.8%) estimates. b) Latitudinal distribution of estimates for
307 LTL, PBT, and UTL. c) Threat status of species, classified according to the International Union for the
308 Conservation of Nature (IUCN⁸¹). One species (not displayed) is now extinct.

309

310

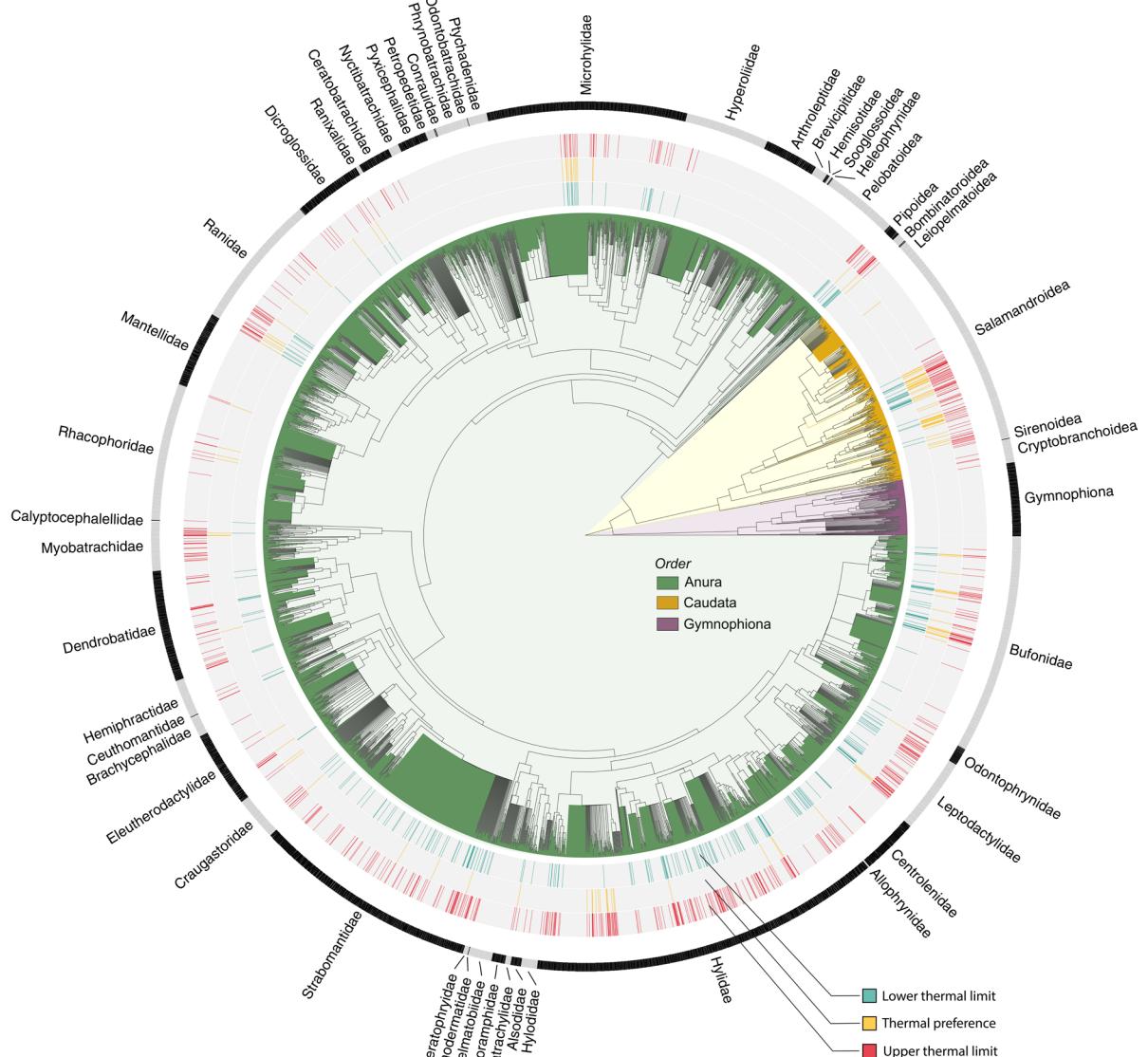


311

312 **Figure 3** | Distribution of mean estimates for three key thermal traits: lower thermal limits (inner heat
 313 map), thermal preference (central heat map), and upper thermal limits (outer heat map). The number of
 314 estimates for each species is displayed as histograms, scaled on a $\log_2(x+1)$ axis for clarity. The
 315 histograms are colour-coded according to the life stage assessed in the experiments, with the category
 316 “juveniles” comprising larvae, metamorphic, and juvenile stages. Gray colour represents missing data.
 317 The phylogeny relationships are based on the consensus tree from⁸⁰. a. *Notophthalmus viridescens*, b.
 318 *Dendropsophus ebraccatus*, c. *Hyla cinerea*, d. *Pleurodema thaul*, e. *Ceratophrys cranwelli*, f.
 319 *Craugastor longirostris*, g. *Rana pipiens*, h. *Xenopus laevis*, i. *Plethodon cylindraceus*.

320

321



322

323 **Figure 4** | Distribution of thermal trait estimates across the phylogeny of most extant amphibians.
 324 Thermal limits and preferences are mapped onto a comprehensive phylogeny of 7,238 species
 325 (consensus tree, ^{cf.80}) to identify taxonomic biases in existing knowledge. The outer circle presents
 326 family names, adapted from ⁸⁰.

327 Technical Validation

328 We employed a transparent and reproducible workflow to systematically review over 4,000 studies from
 329 five databases and across seven languages. The potential limitations of this database are similar to those
 330 described in ¹. First, the methods used for indexing and retrieving studies in Google Scholar are not
 331 publicly disclosed⁴¹², which may undermine reproducibility. However, given the limited coverage of
 332 non-English studies in other databases (with 95% and 93% of references in Scopus and Web of Science
 333 indexed in English, respectively), Google Scholar remains one of the most suitable tools to synthesise
 334 across multiple non-English languages at present^{413,414}. Second, different authors extracted data from
 335 the original studies, introducing the possibility of individual errors. To ensure consistency and accuracy,
 336 all extracted data were subsequently cross-checked and standardised by NC and PPottier (see *Data
 337 Curation*) to minimise the risk of bias and strengthen the reliability of the dataset.

338 We aim to conduct updates at regular five-year intervals, following the same systematic methods, to
339 maintain the database as an up-to-date resource for amphibian thermal envelope data. We encourage
340 researchers who possess relevant thermal data not included in the current version to contact us so that
341 the database can be updated to continuously reflect the most comprehensive and current body of
342 knowledge.

343 **Usage Notes**

344 We anticipate that this database will facilitate a wide range of novel analyses and investigations that
345 may be difficult to foresee at this time, but we are excited to see how these data advance research in
346 amphibian biology and conservation. Our recommendations for using this resource are straightforward:
347 we encourage researchers to have strong foundations in thermal ecology and amphibian biology, and to
348 carefully consider the best approaches for integrating these data into their own investigations.

349 The database represents a comprehensive compilation of studies employing diverse approaches and
350 experimental designs. Given that we cannot anticipate the full scope of research applications, we have
351 made the entire dataset available to allow users to filter and customise the data as needed. We strongly
352 encourage users to clearly document their analytical steps to ensure reproducibility. However, we
353 emphasise that this database version includes data from animals tested under atypical conditions (e.g.,
354 amputations, chemical exposure), or from experiments without replication (e.g. data from a single
355 individual). To accommodate most research needs, we therefore also provide a curated version of the
356 database where we excluded data with procedural inconsistencies, incomplete species-level
357 information, and data involving animals exposed to toxicants, hormones, excessive UV radiation, or
358 pathogens. This curated version of the database is likely more suited for research in ecophysiology,
359 though users with more specialised research questions may find value in the complete dataset. Scripts
360 detailing the data cleaning and curation processes are available at <https://github.com/p-pottier/AmphiTherm> and should provide further guidance for researchers in tailoring the dataset to their
362 specific research needs.

363 As described in the first iteration of this database¹, the data contain inherent sources of non-
364 independence as multiple estimates were extracted from each study, species, population (multiple
365 sampling locations from each species), and cohort (e.g., repeated measures on the same individuals).
366 We recommend that users use phylogenetically-informed statistical models with hierarchical random-
367 effect structures to account for and partition sources of variation^{408,415}. Users should also account for
368 variations in sampling effort (sample size differences), for instance, by weighting estimates by the
369 inverse of their sampling variance⁴⁰⁸. Employing hierarchical models that incorporate sampling
370 variance can help address issues of biological and methodological non-independence, enabling more
371 accurate inferences of the factors driving variation in the data⁴⁰⁸. Most (81%) estimates are associated
372 with a measure of dispersion (standard deviation or standard error), species information is standardised
373 to published phylogenetic information⁸⁰, and unique identifiers have been assigned to each study,
374 species, population, and cohort. These features make *AmphiTherm* readily applicable for use in complex
375 statistical models aimed at uncovering the drivers of thermal tolerance and preferences in amphibians.

376 As described in previous studies, thermal traits in amphibians are influenced by multiple variables,
377 including acclimation temperature, acclimation time, ramping rate, endpoint metrics, body size, sex,
378 assay duration, and geographic origin, among others^{8,20,21}. We recommend careful attention to these
379 variables, with consideration of incorporating sources of methodological or biological variation as
380 covariates in statistical models, to better capture the complexities of amphibian thermal ecology. Finally,
381 it is important to note that due to the data gaps in hotspots of amphibian diversity, the data herein

382 represent only a subset of total amphibian diversity, and subsequent analyses should acknowledge this
383 limitation.

384 **Code Availability**

385 The code used to process the data and produce the figures for this manuscript is available at
386 <https://github.com/p-pottier/AmphiTherm>, and the repository will be archived to Zenodo upon
387 acceptance.

388 **Author Contributions**

389 Conceptualisation: PPottier (lead), NC, SB, TA, SMD, SN.

390 Methodology: PPottier (lead), NC, TA, SMD, SN

391 Software: PPottier

392 Formal Analysis: PPottier

393 Investigation: NC (lead), PPottier, RRYO, PPollo, ANRV, YY, SV, AVL, SB, H-YL, JOV, SV

394 Data Curation: PPottier (lead), NC

395 Visualisation: PPottier

396 Writing (Original Draft): NC (lead), PPottier

397 Writing (Review and Editing): All authors

398 Project administration: NC (lead), PPottier

399 Supervision: SMD, SN

400 All authors gave final approval for publication.

401 **Competing Interests**

402 The authors declare no conflict of interest or competing interests.

403 **Acknowledgements**

404 We thank Emily Rypina, Valentina Oller, and Robert Guralnick for their assistance screening articles
405 for eligibility. We thank Lauren Augustine for providing insight into captive amphibian husbandry
406 techniques. We thank the authors of the original studies for their important contributions to the study of
407 amphibian thermal biology. We thank Miguel Tejedo for pointing to our attention thesis chapters that
408 have been published in journal articles. We pay our respects to the Bedegal people, the traditional
409 custodians of the land on which this work was primarily conducted. The findings and conclusions in
410 this publication have not been formally disseminated by the U.S. Department of Agriculture and should
411 not be construed to represent any agency determination or policy.

412 This study was supported by National Science Foundation Postdoctoral Research Fellowships in
413 Biology Program under Grant No. 2109663 to NC. We also acknowledge financial support to PPottier,
414 SB, and PPollo through the University of New South Wales' Scientia PhD scholarship. PPottier was
415 also supported by the Australian Research Council (ARC) Discovery Project DP230101248 and Future
416 Fellowship FT220100276. SN was supported by the ARC Discovery Project (DP230101248). SMD

417 was supported by the ARC through the DECRA fellowship (DE180100202). TA is supported by ARC
418 Future Fellowship FT180100354. RRYO acknowledges funding from the German Research Foundation
419 (DFG-FZT 118, 202548816), which supports the German Centre for Integrative Biodiversity Research
420 (iDiv). JOV was funded by FONDECYT Postdoctorado (3220722) and ANID – Millennium Science
421 Initiative Program – ICN2021_002. AVL received support from the National Science Foundation (IOS-
422 2011278) and startup funds from the University of Florida’s CLAS Research Office and the Department
423 of Biology.

424 References

- 425 1. Pottier, P. *et al.* A comprehensive database of amphibian heat tolerance. *Sci Data* **9**, 600 (2022).
- 426 2. Pinsky, M. L., Eikeset, A. M., McCauley, D. J., Payne, J. L. & Sunday, J. M. Greater
427 vulnerability to warming of marine versus terrestrial ectotherms. *Nature* **569**, 108–111 (2019).
- 428 3. Morley, S. A., Peck, L. S., Sunday, J. M., Heiser, S. & Bates, A. E. Physiological acclimation
429 and persistence of ectothermic species under extreme heat events. *Global Ecology and
430 Biogeography* **28**, 1018–1037 (2019).
- 431 4. Navas, C. A., Gouveia, S. F., Solano-Iguarán, J. J., Vidal, M. A. & Bacigalupe, L. D. Amphibian
432 responses in experimental thermal gradients: Concepts and limits for inference. *Comparative
433 Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* **254**, 110576 (2021).
- 434 5. Leiva, F. P., Calosi, P. & Verberk, W. C. E. P. Scaling of thermal tolerance with body mass and
435 genome size in ectotherms: a comparison between water- and air-breathers. *Philosophical
436 Transactions of the Royal Society B: Biological Sciences* **374**, 20190035 (2019).
- 437 6. Bennett, J. M. *et al.* GlobTherm, a global database on thermal tolerances for aquatic and
438 terrestrial organisms. *Sci Data* **5**, 180022 (2018).
- 439 7. Rohr, J. R. *et al.* The complex drivers of thermal acclimation and breadth in ectotherms. *Ecology
440 Letters* **21**, 1425–1439 (2018).
- 441 8. DuBose, T. P. *et al.* Thermal Traits of Anurans Database for the Southeastern United States
442 (TRAD): A Database of Thermal Trait Values for 40 Anuran Species. *Ichthyology &
443 Herpetology* **112**, 21–30 (2024).
- 444 9. Bayat, H. S. *et al.* Global thermal tolerance of freshwater invertebrates and fish.
445 2024.07.08.602306 Preprint at <https://doi.org/10.1101/2024.07.08.602306> (2024).
- 446 10. Gunderson, A. R. & Stillman, J. H. Plasticity in thermal tolerance has limited potential to buffer
447 ectotherms from global warming. *Proceedings of the Royal Society B: Biological Sciences* **282**,
448 20150401 (2015).
- 449 11. Urban, M. C. Accelerating extinction risk from climate change. *Science* **348**, 571–573 (2015).
- 450 12. Wiens, J. J. Climate-Related Local Extinctions Are Already Widespread among Plant and
451 Animal Species. *PLOS Biology* **14**, e2001104 (2016).
- 452 13. Carey, C. & Alexander, M. A. Climate change and amphibian declines: is there a link? *Diversity
453 and Distributions* **9**, 111–121 (2003).
- 454 14. Sinervo, B. *et al.* Erosion of Lizard Diversity by Climate Change and Altered Thermal Niches.
455 *Science* **328**, 894–899 (2010).
- 456 15. Anderson, R. O., Meiri, S. & Chapple, D. G. The biogeography of warming tolerance in lizards.
457 *Journal of Biogeography* **49**, 1274–1285 (2022).
- 458 16. Biber, M. F., Voskamp, A. & Hof, C. Potential effects of future climate change on global reptile
459 distributions and diversity. *Global Ecology and Biogeography* **32**, 519–534 (2023).
- 460 17. Pottier, P. *et al.* Vulnerability of amphibians to global warming. Preprint at
461 <https://doi.org/10.32942/X2T02T> (2024).
- 462 18. Turner, M. G. *et al.* Climate change, ecosystems and abrupt change: science priorities.
463 *Philosophical Transactions of the Royal Society B: Biological Sciences* **375**, 20190105 (2020).
- 464 19. Walsh, J. E. *et al.* Extreme weather and climate events in northern areas: A review. *Earth-
465 Science Reviews* **209**, 103324 (2020).

- 466 20. Bodensteiner, B. L. *et al.* Thermal adaptation revisited: How conserved are thermal traits of
467 reptiles and amphibians? *Journal of Experimental Zoology Part A: Ecological and Integrative*
468 *Physiology* **335**, 173–194 (2021).
- 469 21. Taylor, E. N. *et al.* The thermal ecology and physiology of reptiles and amphibians: A user's
470 guide. *Journal of Experimental Zoology Part A: Ecological and Integrative Physiology* **335**, 13–
471 44 (2021).
- 472 22. Luedtke, J. A. *et al.* Ongoing declines for the world's amphibians in the face of emerging
473 threats. *Nature* **622**, 308–314 (2023).
- 474 23. Hatch, K. A. & Kroft, K. L. Winterkill in Lotic Systems May Be an Important Driver of
475 Amphibian Population Declines. *Ichthyology & Herpetology* **110**, 575–584 (2022).
- 476 24. Enriquez-Urzelai, U., Kearney, M. R., Nicieza, A. G. & Tingley, R. Integrating mechanistic and
477 correlative niche models to unravel range-limiting processes in a temperate amphibian. *Global*
478 *Change Biology* **25**, 2633–2647 (2019).
- 479 25. Huey, R. B. & Slatkin, M. Cost and Benefits of Lizard Thermoregulation. *The Quarterly Review*
480 *of Biology* (1976) doi:10.1086/409470.
- 481 26. Dawson, W. R. On the Physiological Significance of the Preferred Body Temperatures of
482 Reptiles. in *Perspectives of Biophysical Ecology* (eds. Gates, D. M. & Schmerl, R. B.) 443–473
483 (Springer, Berlin, Heidelberg, 1975). doi:10.1007/978-3-642-87810-7_25.
- 484 27. Buckley, L. B. Linking Traits to Energetics and Population Dynamics to Predict Lizard Ranges
485 in Changing Environments. *The American Naturalist* **171**, E1–E19 (2008).
- 486 28. Buckley, L. B. *et al.* Can mechanism inform species' distribution models? *Ecology Letters* **13**,
487 1041–1054 (2010).
- 488 29. Clusella-Trullas, S., Blackburn, T. M. & Chown, S. L. Climatic Predictors of Temperature
489 Performance Curve Parameters in Ectotherms Imply Complex Responses to Climate Change.
490 *The American Naturalist* **177**, 738–751 (2011).
- 491 30. Caetano, G. H. O. *et al.* Time of activity is a better predictor of the distribution of a tropical
492 lizard than pure environmental temperatures. *Oikos* **129**, 953–963 (2020).
- 493 31. Clauch, N. M. *et al.* Commonly collected thermal performance data can inform species
494 distributions in a data-limited invader. *Sci Rep* **13**, 15880 (2023).
- 495 32. Ivey, K. *et al.* Temperature-based activity estimation accurately predicts surface activity, but not
496 microhabitat use, in the endangered heliothermic lizard *Gambelia sila*. *Amphibian and Reptile*
497 *Conservation* **16**, 10 (2022).
- 498 33. Üveges, B. *et al.* Experimental evidence for beneficial effects of projected climate change on
499 hibernating amphibians. *Sci Rep* **6**, 26754 (2016).
- 500 34. Rollins-Smith, L. A. Amphibian immunity–stress, disease, and climate change. *Developmental*
501 & *Comparative Immunology* **66**, 111–119 (2017).
- 502 35. Beukema, W. *et al.* Microclimate limits thermal behaviour favourable to disease control in a
503 nocturnal amphibian. *Ecology Letters* **24**, 27–37 (2021).
- 504 36. Waddle, A. W. *et al.* Hotspot shelters stimulate frog resistance to chytridiomycosis. *Nature* **631**,
505 344–349 (2024).
- 506 37. Greenspan, S. E. *et al.* Warming drives ecological community changes linked to host-associated
507 microbiome dysbiosis. *Nat. Clim. Chang.* **10**, 1057–1061 (2020).
- 508 38. Zhu, L. *et al.* Environmental Temperatures Affect the Gastrointestinal Microbes of the Chinese
509 Giant Salamander. *Front. Microbiol.* **12**, (2021).
- 510 39. Robak, M. J., Saenz, V., de Cortie, E. & Richards-Zawacki, C. L. Effects of temperature on the
511 interaction between amphibian skin bacteria and *Batrachochytrium dendrobatidis*. *Front.*
512 *Microbiol.* **14**, (2023).
- 513 40. Sunday, J. M., Bates, A. E. & Dulvy, N. K. Thermal tolerance and the global redistribution of
514 animals. *Nature Clim Change* **2**, 686–690 (2012).
- 515 41. Moore, N. A. *et al.* Temperate species underfill their tropical thermal potentials on land. *Nat*
516 *Ecol Evol* **7**, 1993–2003 (2023).
- 517 42. Bennett, J. M. *et al.* The evolution of critical thermal limits of life on Earth. *Nat Commun* **12**,
518 1198 (2021).

- 519 43. Harishchandra, A., Xue, H., Salinas, S. & Jayasundara, N. Thermal physiology integrated
520 species distribution model predicts profound habitat fragmentation for estuarine fish with ocean
521 warming. *Sci Rep* **12**, 21781 (2022).
- 522 44. Zlotnick, O. B., Musselman, K. N. & Levy, O. Deforestation poses deleterious effects to tree-
523 climbing species under climate change. *Nat. Clim. Chang.* **14**, 289–295 (2024).
- 524 45. Stark, G., Ma, L., Zeng, Z.-G., Du, W.-G. & Levy, O. Cool shade and not-so-cool shade: How
525 habitat loss may accelerate thermal stress under current and future climate. *Global Change
526 Biology* **29**, 6201–6216 (2023).
- 527 46. Briscoe, N. J. *et al.* Mechanistic forecasts of species responses to climate change: The promise
528 of biophysical ecology. *Global Change Biology* **29**, 1451–1470 (2023).
- 529 47. Kearney, M. *et al.* Modelling species distributions without using species distributions: the cane
530 toad in Australia under current and future climates. *Ecography* **31**, 423–434 (2008).
- 531 48. Araújo, M. B. *et al.* Heat freezes niche evolution. *Ecology Letters* **16**, 1206–1219 (2013).
- 532 49. Hoffmann, A. A., Chown, S. L. & Clusella-Trullas, S. Upper thermal limits in terrestrial
533 ectotherms: how constrained are they? *Functional Ecology* **27**, 934–949 (2013).
- 534 50. Chown, S. L. Physiological variation in insects: hierarchical levels and implications. *Journal of
535 Insect Physiology* **47**, 649–660 (2001).
- 536 51. Rubalcaba, J. G., Gouveia, S. F., Villalobos, F., Olalla-Tárraga, M. Á. & Sunday, J. Climate
537 drives global functional trait variation in lizards. *Nat Ecol Evol* **7**, 524–534 (2023).
- 538 52. Longo, A. V. & Zamudio, K. R. Temperature variation, bacterial diversity and fungal infection
539 dynamics in the amphibian skin. *Molecular Ecology* **26**, 4787–4797 (2017).
- 540 53. Xu, L. *et al.* The Behavior of Amphibians Shapes Their Symbiotic Microbiomes. *mSystems* **5**,
541 10.1128/msystems.00626-20 (2020).
- 542 54. Bernardo-Cravo, A. P., Schmeller, D. S., Chatzinotas, A., Vredenburg, V. T. & Loyau, A.
543 Environmental Factors and Host Microbiomes Shape Host–Pathogen Dynamics. *Trends in
544 Parasitology* **36**, 616–633 (2020).
- 545 55. Sonn, J. M., Porter, W. P., Mathewson, P. D. & Richards-Zawacki, C. L. Predictions of Disease
546 Risk in Space and Time Based on the Thermal Physiology of an Amphibian Host–Pathogen
547 Interaction. *Front. Ecol. Evol.* **8**, (2020).
- 548 56. Neely, W. J. *et al.* Synergistic effects of warming and disease linked to high mortality in cool-
549 adapted terrestrial frogs. *Biological Conservation* **245**, 108521 (2020).
- 550 57. Fontaine, S. S., Mineo, P. M. & Kohl, K. D. Experimental manipulation of microbiota reduces
551 host thermal tolerance and fitness under heat stress in a vertebrate ectotherm. *Nat Ecol Evol* **6**,
552 405–417 (2022).
- 553 58. Crump, P. & Grow, S. Action plan for ex situ amphibian conservation in the AZA community.
554 *Association of Zoos & Aquariums, Amphibian Taxon Advisory Group* (2007).
- 555 59. Linhoff, L. J. & Donnelly, M. A. Assessing release strategies for reintroductions of endangered
556 Wyoming toads. *Wildlife Society Bulletin* **46**, e1341 (2022).
- 557 60. Klocke, B. *et al.* Movement and survival of captive-bred Limosa harlequin frogs (*Atelopus*
558 *limosus*) released into the wild. *Front. Amphib. Reptile Sci.* **1**, (2023).
- 559 61. Beaupre, S., Jacobson, E., Lillywhite, H. & Zamudio, K. Guidelines for use of live amphibians
560 and reptiles in field and laboratory research. (2004).
- 561 62. Poole, V. A. & Grow, S. *Amphibian Husbandry Resource Guide, Edition 2.0.* (Association of
562 Zoos & Aquariums, Silver Spring, Maryland, USA, 2017).
- 563 63. Pough, H. F. Amphibian biology and husbandry. *ILAR journal* **48**, 203–213 (2007).
- 564 64. Greenspan, S. E. *et al.* Realistic heat pulses protect frogs from disease under simulated
565 rainforest frog thermal regimes. *Functional Ecology* **31**, 2274–2286 (2017).
- 566 65. Sinervo, B. *et al.* Climate change and collapsing thermal niches of desert reptiles and
567 amphibians: Assisted migration and acclimation rescue from extirpation. *Science of The Total
568 Environment* **908**, 168431 (2024).
- 569 66. McNutt, M. K. *et al.* Transparency in authors' contributions and responsibilities to promote
570 integrity in scientific publication. *Proceedings of the National Academy of Sciences* **115**, 2557–
571 2560 (2018).
- 572 67. Nakagawa, S. *et al.* Method Reporting with Initials for Transparency (MeRIT) promotes more
573 granularity and accountability for author contributions. *Nat Commun* **14**, 1788 (2023).

- 574 68. Pottier, P. *et al.* Title, abstract and keywords: a practical guide to maximize the visibility and
575 impact of academic papers. *Proceedings of the Royal Society B: Biological Sciences* **291**,
576 20241222 (2024).
- 577 69. Harzing, A. Publish or perish. (2007).
- 578 70. Cowles, R. B. & Bogert, C. M. A preliminary study of the thermal requirements of desert
579 reptiles. *Bull. Am. Mus. Nat. Hist.* **83**, 261–296 (1944).
- 580 71. Fry, F. E. J. Effects of the environment on animal activity. *Publ. Out. Fish. Res. Lab.* **55**, 1–62
581 (1947).
- 582 72. Costanzo, J. P., Reynolds, A. M., Amaral, M. C. F. do, Rosendale, A. J. & Jr, R. E. L.
583 Cryoprotectants and Extreme Freeze Tolerance in a Subarctic Population of the Wood Frog.
584 *PLOS ONE* **10**, e0117234 (2015).
- 585 73. Ouzzani, M., Hammady, H., Fedorowicz, Z. & Elmagarmid, A. Rayyan—a web and mobile app
586 for systematic reviews. *Syst Rev* **5**, 210 (2016).
- 587 74. O'Dea, R. E. *et al.* Preferred reporting items for systematic reviews and meta-analyses in
588 ecology and evolutionary biology: a PRISMA extension. *Biological Reviews* **96**, 1695–1722
589 (2021).
- 590 75. Pick, J. L., Nakagawa, S. & Noble, D. W. A. Reproducible, flexible and high-throughput data
591 extraction from primary literature: The metaDigitise r package. *Methods in Ecology and*
592 *Evolution* **10**, 426–431 (2019).
- 593 76. R Core Team. R: A language and environment for statistical computing. R Foundation for
594 Statistical Computing (2019).
- 595 77. Rohatgi, A. WebPlotDigitizer. (2024).
- 596 78. Ripley, B. *et al.* Package ‘mass’. *Cran r* **538**, 822 (2013).
- 597 79. AmphibiaWeb. <https://amphibiaweb.org>. University of California, Berkeley, California, USA
598 (2025).
- 599 80. Jetz, W. & Pyron, R. A. The interplay of past diversification and evolutionary isolation with
600 present imperilment across the amphibian tree of life. *Nat Ecol Evol* **2**, 850–858 (2018).
- 601 81. IUCN. The IUCN Red List of Threatened Species. (2021).
- 602 82. Agudelo-Cantero, G. A. & Navas, C. A. Interactive effects of experimental heating rates,
603 ontogeny and body mass on the upper thermal limits of anuran larvae. *Journal of Thermal*
604 *Biology* **82**, 43–51 (2019).
- 605 83. Alveal, N. *et al.* Relationship between thermal behavior and sex of a population of Pleurodematahul
606 (Amphibia: Leiuperidae) of the commune of Antuco, Biobío region. *Gayana (Concepción)*
607 **83**, 93–101 (2019).
- 608 84. Alveal Riquelme, N. Relaciones entre la fisiología térmica y las características bioclimáticas de
609 Rhinella spinulosa (Anura: Bufonidae) en Chile a través del enlace mecanicista de nicho
610 (Universidad de Concepción, Concepción, Chile, 2015).
- 611 85. Alves, M. Tolerância térmica em espécies de anuros neotropicais do gênero Dendropsophus
612 Fitzinger, 1843 e efeito da temperatura na resposta à predação. (Universidade Estadual de Santa
613 Cruz, Santa Cruz, Brazil, 2016).
- 614 86. Anderson, R. C. O. & Andrade, D. V. Trading heat and hops for water: Dehydration effects on
615 locomotor performance, thermal limits, and thermoregulatory behavior of a terrestrial toad.
616 *Ecology and Evolution* **7**, 9066–9075 (2017).
- 617 87. Aponte Gutiérrez, A. Endurecimiento térmico en Pristimantis medemi (Anura: Craugastoridae),
618 en coberturas boscosas del Municipio de Villavicencio (Meta). (Universidad Nacional de
619 Colombia, Bogotá, Colombia, 2020).
- 620 88. Arrigada García, K. Conductas térmica en dos poblaciones de Batrachyla taeniata provenientes
621 de la localidad de Ucúquer en la región de O'Higgins y de la localidad de Hualpén en la
622 (Universidad de Concepción, Concepción, Chile, 2019).
- 623 89. Azambuja, G., Martins, I. K., Franco, J. L. & Santos, T. G. dos. Effects of mancozeb on heat
624 Shock protein 70 (HSP70) and its relationship with the thermal physiology of Physalaemus
625 henselii (Peters, 1872) tadpoles (Anura: Leptodactylidae). *Journal of Thermal Biology* **98**,
626 102911 (2021).
- 627 90. Bacigalupe, L. D. *et al.* Natural selection on plasticity of thermal traits in a highly seasonal
628 environment. *Evolutionary Applications* **11**, 2004–2013 (2018).

- 629 91. Bakewell, L., Kelehear, C. & Graham, S. p. Impacts of temperature on immune performance in
630 a desert anuran (*Anaxyrus punctatus*). *Journal of Zoology* **315**, 49–57 (2021).
- 631 92. Balogová, M. & Gvoždík, L. Can Newts Cope with the Heat? Disparate Thermoregulatory
632 Strategies of Two Sympatric Species in Water. *PLOS ONE* **10**, e0128155 (2015).
- 633 93. Barria, A. M. & Bacigalupi, L. D. Intraspecific geographic variation in thermal limits and
634 acclimatory capacity in a wide distributed endemic frog. *Journal of Thermal Biology* **69**, 254–
635 260 (2017).
- 636 94. Bazin, Y., Wharton, D. A. & Bishop, P. J. Cold tolerance and overwintering of an introduced
637 New Zealand frog, the brown tree frog (*Litoria ewingii*). *Cryo Letters* **28**, 347–358 (2007).
- 638 95. Beltrán, I., Ramírez-Castañeda, V., Rodríguez-López, C., Lasso, E. & Amézquita, A. Dealing
639 with hot rocky environments: critical thermal maxima and locomotor performance in
640 leptodactylus lithonaetes (anura: Leptodactylidae). *Herpetological Journal* **29**, 155–161 (2019).
- 641 96. Berkhouse, C. & Fries, J. Critical thermal maxima of juvenile and adult San Marcos
642 salamanders (*Eurycea nana*). *Southwestern Naturalist* **40**, 430–434 (1995).
- 643 97. Berman, D. I., Meshcheryakova, E. N. & Bulakhova, N. A. The Japanese tree frog (*Hyla*
644 *japonica*), one of the most cold-resistant species of amphibians. *Dokl Biol Sci* **471**, 276–279
645 (2016).
- 646 98. Berman, D. I., Bulakhova, N. A., Meshcheryakova, E. N. & Shekhovtsov, S. V. Overwintering
647 and cold tolerance in the Moor Frog (*Rana arvalis*) across its range. *Can. J. Zool.* **98**, 705–714
648 (2020).
- 649 99. Bernal Castro, E. A. Influence of environment on thermal ecology of direct-developing frogs
650 (Anura: craugastoridae: pristimantis) in the eastern Andes of Colombia. (Universidad de los
651 Andes, Bogotá, Colombia, 2019).
- 652 100. Berner, N. J. & Puckett, R. E. Phenotypic flexibility and thermoregulatory behavior in the
653 eastern red-spotted newt (*Notophthalmus viridescens viridescens*). *J Exp Zool A Ecol Genet
654 Physiol* **313**, 231–239 (2010).
- 655 101. Beukema, W. *et al.* Microclimate limits thermal behaviour favourable to disease control in a
656 nocturnal amphibian. *Ecology Letters* **24**, 27–37 (2021).
- 657 102. Bicego, K. C. & Branco, L. G. S. Discrete electrolytic lesion of the preoptic area prevents LPS-
658 induced behavioral fever in toads. *Journal of Experimental Biology* **205**, 3513–3518 (2002).
- 659 103. Bicego-Nahas, K. C., Steiner, A. A., Carnio, E. C., Antunes-Rodrigues, J. & Branco, L. G. S.
660 Antipyretic effect of arginine vasotocin in toads. *American Journal of Physiology-Regulatory,
661 Integrative and Comparative Physiology* **278**, R1408–R1414 (2000).
- 662 104. Bicego-Nahas, K. C., Gargaglioni, L. H. & Branco, L. G. S. Seasonal changes in the preferred
663 body temperature, cardiovascular, and respiratory responses to hypoxia in the toad, *Bufo*
664 *paracnemis*. *Journal of Experimental Zoology* **289**, 359–365 (2001).
- 665 105. Blem, C. R., Ragan, C. A. & Scott, L. S. The thermal physiology of two sympatric treefrogs
666 *Hyla cinerea* and *Hyla chrysoscelis* (Anura; Hylidae). *Comparative Biochemistry and
667 Physiology -- Part A: Physiology* **85**, 563–570 (1986).
- 668 106. Bliss, M. M. & Cecala, K. K. Terrestrial Salamanders Alter Antipredator Behavior Thresholds
669 Following Tail Autotomy. *herp* **73**, 94–99 (2017).
- 670 107. Bonino, M. F., Cruz, F. B. & Perotti, M. G. Does temperature at local scale explain thermal
671 biology patterns of temperate tadpoles? *Journal of Thermal Biology* **94**, (2020).
- 672 108. Bovo, R. P. Fisiología térmica e balanço hídrico em anfíbios anuros. (Universidad Estadual
673 Paulista, Rio Claro, Brazil, 2015).
- 674 109. Branco, L. G. Effects of 2-deoxy-D-glucose and insulin on plasma glucose levels and behavioral
675 thermoregulation of toads. *Am J Physiol* **272**, R1-5 (1997).
- 676 110. Branco, L. G. & Malvin, G. M. Thermoregulatory effects of cyanide and azide in the toad, *Bufo*
677 *marinus*. *Am J Physiol* **270**, R169-173 (1996).
- 678 111. Branco, L. G., Steiner, A. A., Tattersall, G. J. & Wood, S. C. Role of adenosine in the hypoxia-
679 induced hypothermia of toads. *Am J Physiol Regul Integr Comp Physiol* **279**, R196-201 (2000).
- 680 112. Branco, L. G. & Wood, S. C. Role of central chemoreceptors in behavioral thermoregulation of
681 the toad, *Bufo marinus*. *Am J Physiol* **266**, R1483-1487 (1994).

- 682 113. Branco, L. G. S. & Steiner, A. A. Central thermoregulatory effects of lactate in the toad *Bufo*
683 *paracnemis*. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative*
684 *Physiology* **122**, 457–461 (1999).
- 685 114. Brattstrom, B. H. Thermal acclimation in Australian amphibians. *Comparative Biochemistry*
686 *And Physiology* **35**, 69–103 (1970).
- 687 115. Brattstrom, B. H. & Regal, P. Rate of thermal acclimation in the Mexican salamander
688 *Chiropterotriton*. *Copeia* **1965**, 514–515 (1965).
- 689 116. Brattstrom, B. H. A Preliminary Review of the Thermal Requirements of Amphibians. *Ecology*
690 **44**, 238–255 (1963).
- 691 117. Brattstrom, B. H. Thermal acclimation in Anuran amphibians as a function of latitude and
692 altitude. *Comparative Biochemistry and Physiology* **24**, 93–111 (1968).
- 693 118. Brattstrom, B. H. & Lawrence, P. The Rate of Thermal Acclimation in Anuran Amphibians.
694 *Physiological Zoology* **35**, 148–156 (1962).
- 695 119. Brown, H. A. The heat resistance of some anuran tadpoles (Hylidae and Pelobatidae). *Copeia*
696 **1969**, 138 (1969).
- 697 120. Burke, E. M. & Pough, F. H. The role of fatigue in temperature resistance of salamanders.
698 *Journal of Thermal Biology* **1**, 163–167 (1976).
- 699 121. Burrowes, P. A., Navas, C. A., Jiménez-Robles, O., Delgado, P. & De La Riva, I. Climatic
700 heterogeneity in the bolivian andes: Are frogs trapped? *South American Journal of Herpetology*
701 **18**, 1–12 (2020).
- 702 122. Bury, R. B. Low thermal tolerances of stream amphibians in the Pacific Northwest: Implications
703 for riparian and forest management. *Applied Herpetology* **5**, 63–74 (2008).
- 704 123. Carey, C. Factors affecting body temperatures of toads. *Oecologia* **35**, 197–219 (1978).
- 705 124. Carvajalino-Fernández, J. M., Gomez, M. A. B., Giraldo-Gutiérrez, L. & Navas, C. A. Freeze
706 tolerance in neotropical frogs: an intrageneric comparison using *Pristimantis* species of high
707 elevation and medium elevation. *Journal of Tropical Ecology* **37**, 118–125 (2021).
- 708 125. Castellanos García, L. A. Days of futures past : integrating physiology, microenvironments, and
709 biogeographic history to predict response of frogs in neotropical dry-forest to global warming.
710 (Universidad de los Andes, Bogotá, Colombia, 2017).
- 711 126. Catenazzi, A., Lehr, E. & Vredenburg, V. T. Thermal physiology, disease, and amphibian
712 declines on the eastern slopes of the andes. *Conservation Biology* **28**, 509–517 (2014).
- 713 127. Cecala, K. K., Noggle, W. & Burns, S. Negative Phototaxis Results from Avoidance of Light
714 and Temperature in Stream Salamander Larvae. *Journal of Herpetology* **51**, 263–269 (2017).
- 715 128. Chang, L.-W. Heat tolerance and its plasticity in larval *Bufo bankorensis* from different
716 altitudes. (National Cheng Kung University, Tainan, Taiwan, 2002).
- 717 129. Chavez Landi, P. A. Fisiología térmica de un depredador *Dasythemis* sp.(Odonata: Libellulidae)
718 y su presa *Hypsiboas pellucens* (Anura: Hylidae) y sus posibles implicaciones frente al
719 (Pontificia Universidad Católica Del Ecuador, Quito, Ecuador, 2017).
- 720 130. Chen, T.-C., Kam, Y.-C. & Lin, Y.-S. Thermal physiology and reproductive phenology of
721 *Buergeria japonica* (rhacophoridae) breeding in a stream and a geothermal hotspring in Taiwan.
722 *Zoological Science* **18**, 591–596 (2001).
- 723 131. Cheng, C.-B. A study of warming tolerance and thermal acclimation capacity of tadpoles in
724 Taiwan. (Tunghai University, Taichung, Taiwan, 2017).
- 725 132. Cheng, Y.-J. Effect of salinity on the critical thermal maximum of tadpoles living in brackish
726 water. (Tunghai University, Taichung, Taiwan, 2017).
- 727 133. Christian, K. A., Nunez, F., Clos, L. & Diaz, L. Thermal relations of some tropical frogs along
728 an altitudinal gradient. *Biotropica* **20**, 236–239 (1988).
- 729 134. Churchill, T. A. & Storey, K. B. Dehydration tolerance in wood frogs: a new perspective on
730 development of amphibian freeze tolerance. *Am J Physiol* **265**, R1324-1332 (1993).
- 731 135. Claussen, D. L. The thermal relations of the tailed frog, *Ascaphus truei*, and the pacific treefrog,
732 *Hyla regilla*. *Comparative Biochemistry and Physiology -- Part A: Physiology* **44**, 137–153
733 (1973).
- 734 136. Claussen, D. L. Thermal acclimation in ambystomatid salamanders. *Comparative Biochemistry*
735 *and Physiology -- Part A: Physiology* **58**, 333–340 (1977).

- 736 137. Clemente, A. C. [UNESP. Resposta termofílica pós-prandial do sapo-cururu, *Rhinella diptycha*.
737 (São Paulo State University, São Paulo, Brazil, 2019).
738 138. Cohen, J. M. *et al.* The thermal mismatch hypothesis explains host susceptibility to an emerging
739 infectious disease. *Ecol Lett* **20**, 184–193 (2017).
740 139. Contreras Cisneros, J. Temperatura crítica máxima, tolerancia al frío y termopreferendum del
741 tritón del Montseny (*Calotriton arnoldii*). (Universitat de Barcelona, Barcelona, Spain, 2019).
742 140. Contreras López, J. M. Requerimientos térmicos de *Craugastor loki* (Anura: Craugastoridae) en
743 la Sierra Madre de Chiapas, México. (Universidad de Ciencias y Artes de Chiapas, 2021).
744 141. Contreras Oñate, S. Posible efecto de las temperaturas de aclimatación sobre las respuestas
745 térmicas en temperaturas críticas máximas (TCmáx) y mínimas (TCmín) de una población de
746 (Universidad de Concepción, Concepción, Chile, 2016).
747 142. Cooper, R. D. & Shaffer, H. B. Allele-specific expression and gene regulation help explain
748 transgressive thermal tolerance in non-native hybrids of the endangered California tiger
749 salamander (*Ambystoma californiense*). *Molecular Ecology* **30**, 987–1004 (2021).
750 143. Cortes, P. A., Puschel, H., Acuña, P., Bartheld, J. L. & Bozinovic, F. Thermal ecological
751 physiology of native and invasive frog species: do invaders perform better? *Conservation
752 Physiology* **4**, cow056 (2016).
753 144. Crawshaw, L. I., Rausch, R. N., Wollmuth, L. P. & Bauer, E. J. Seasonal Rhythms of
754 Development and Temperature Selection in Larval Bullfrogs, *Rana catesbeiana* Shaw.
755 *Physiological Zoology* **65**, 346–359 (1992).
756 145. Crow, J. C., Forstner, M. R. J., Ostr, K.G. & Tomasso, J. R. The role of temperature on survival
757 and growth of the barton springs salamander (*Eurycea sosorum*). *Herpetological Conservation
758 and Biology* **11**, 328–334 (2016).
759 146. Cupp, P. V. Thermal Tolerance of Five Salientian Amphibians during Development and
760 Metamorphosis. *Herpetologica* **36**, 234–244 (1980).
761 147. Dabruzzi, T. F., Wygoda, M. L. & Bennett, W. A. Some Like it Hot: Heat Tolerance of the Crab-
762 Eating Frog, *Fejervarya cancrivora*. *Micronesica* **43**, 101–106 (2012).
763 148. Dainton, B. H. Heat tolerance and thyroid activity in developing tadpoles and juvenile adults of
764 *Xenopus laevis* (Daudin). *Journal of Thermal Biology* **16**, 273–276 (1991).
765 149. Daniel, N. J. J. Impact of climate change on Singapore amphibians. (National University of
766 Singapore, Singapore, 2013).
767 150. Davies, S. J., McGeoch, M. A. & Clusella-Trullas, S. Plasticity of thermal tolerance and
768 metabolism but not water loss in an invasive reed frog. *Comparative Biochemistry and
769 Physiology -Part A : Molecular and Integrative Physiology* **189**, 11–20 (2015).
770 151. de Oliveira Anderson, R. C., Bovo, R. P. & Andrade, D. V. Seasonal variation in the thermal
771 biology of a terrestrial toad, *Rhinella icterica* (Bufonidae), from the Brazilian Atlantic Forest.
772 *Journal of Thermal Biology* **74**, 77–83 (2018).
773 152. de Vlaming, V. L. & Bury, R. B. Thermal Selection in Tadpoles of the Tailed-Frog, *Ascaphus
774 truei*. *Journal of Herpetology* **4**, 179–189 (1970).
775 153. Delgado-Suazo, P. & Burrowes, P. A. Response to thermal and hydric regimes point to
776 differential inter- and intraspecific vulnerability of tropical amphibians to climate warming. *J
777 Therm Biol* **103**, 103148 (2022).
778 154. Delson, J. & Whitford, W. G. Critical Thermal Maxima in Several Life History Stages in Desert
779 and Montane Populations of *Ambystoma tigrinum*. *Herpetologica* **29**, 352–355 (1973).
780 155. Dohm, M. R., Mautz, W. J., Looby, P. G., Gellert, K. S. & Andrade, J. A. Effects of Ozone on
781 Evaporative Water Loss and Thermoregulatory Behavior of Marine Toads (*Bufo marinus*).
782 *Environmental Research* **86**, 274–286 (2001).
783 156. Drakulić, S. *et al.* Local differences of thermal preferences in European common frog (*Rana
784 temporaria* Linnaeus, 1758) tadpoles. *Zoologischer Anzeiger* **268**, 47–54 (2017).
785 157. Duarte, H. *et al.* Can amphibians take the heat? Vulnerability to climate warming in subtropical
786 and temperate larval amphibian communities. *Global Change Biology* **18**, 412–421 (2012).
787 158. Duarte, H. S. A comparative study of the thermal tolerance of tadpoles of iberian anurans.
788 (Universidade de Lisboa, Lisboa, Portugal, 2011).
789 159. Duclaux, R., Fantino, M. & Cabanac, M. Comportement thermoregulateur chez *Rana esculenta*.
790 *Pflugers Arch.* **342**, 347–358 (1973).

- 791 160. Dunlap, D. Evidence for a daily rhythm of heat resistance in cricket frogs, *Acris crepitans*.
792 *Copeia* 852- (1969).
- 793 161. Dunlap, D. G. Critical Thermal Maximum as a Function of Temperature of Acclimation in Two
794 Species of Hylid Frogs. *Physiological Zoology* **41**, 432–439 (1968).
- 795 162. Easton, L. Determining the feasibility of a translocation by investigating the ecology and
796 physiology of the threatened Hochstetter's frog (*Leiopelma hochstetteri*). (University of Otago,
797 Otago, New Zealand, 2015).
- 798 163. Elwood, J. R. L. Variation in hsp70 levels and thermotolerance among terrestrial salamanders of
799 the *Plethodon glutinosus* complex. (Drexel University, Philadelphia, Pennsylvania, USA, 2003).
- 800 164. Enriquez-Urzelai, U. *et al.* Ontogenetic reduction in thermal tolerance is not alleviated by earlier
801 developmental acclimation in *Rana temporaria*. *Oecologia* **189**, 385–394 (2019).
- 802 165. Enriquez-Urzelai, U. *et al.* The roles of acclimation and behaviour in buffering climate change
803 impacts along elevational gradients. *Journal of Animal Ecology* **89**, 1722–1734 (2020).
- 804 166. Enriquez-Urzelai, U., Palacio, A. S., Merino, N. M., Sacco, M. & Nicieza, A. G. Hindered and
805 constrained: limited potential for thermal adaptation in post-metamorphic and adult *Rana*
806 *temporaria* along elevational gradients. *Journal of Evolutionary Biology* **31**, 1852–1862 (2018).
- 807 167. Erskine, D. J. & Hutchison, V. H. Reduced thermal tolerance in an amphibian treated with
808 melatonin. *Journal of Thermal Biology* **7**, 121–123 (1982).
- 809 168. Escobar Serrano, D. Acclimation scope of the critical thermal limits in *Agalychnis spurrelli*
810 (Hylidae) and *Gastrotheca pseustes* (Hemiphractidae) and their implications under climate
811 change scenarios. (Pontificia Universidad Católica Del Ecuador, Quito, Ecuador, 2016).
- 812 169. Familiar Lopez, M. Distribution, Ecology, Disease and Physiology of a Mountain-Top Endemic
813 Frog in the Face of Climate Change: A Study on *Philoria loveridgei*. (Griffith University,
814 Australia, 2016).
- 815 170. Fan, X., Lei, H. & Lin, Z. Ontogenetic shifts in selected body temperature and thermal tolerance
816 of the tiger frog, *Hoplobatrachus chinensis*. *Acta Ecologica Sinica* **32**, 5574–5580 (2012).
- 817 171. Fan, X. L., Lin, Z. H. & Scheffers, B. R. Physiological, developmental, and behavioral plasticity
818 in response to thermal acclimation. *Journal of Thermal Biology* **97**, (2021).
- 819 172. Fernández-Loras, A. *et al.* Infection with *Batrachochytrium dendrobatidis* lowers heat tolerance
820 of tadpole hosts and cannot be cleared by brief exposure to CTmax. *PLoS ONE* **14**, (2019).
- 821 173. Finnerty, P. B., Shine, R. & Brown, G. P. Survival of the feces: Does a nematode lungworm
822 adaptively manipulate the behavior of its cane toad host? *Ecol Evol* **8**, 4606–4618 (2018).
- 823 174. Floyd, R. B. Ontogenetic change in the temperature tolerance of larval *Bufo marinus* (Anura:
824 bufonidae). *Comparative Biochemistry and Physiology -- Part A: Physiology* **75**, 267–271
825 (1983).
- 826 175. Floyd, R. B. Variation in Temperature Preference with Stage of Development of *Bufo marinus*
827 Larvae. *Journal of Herpetology* **18**, 153–158 (1984).
- 828 176. Floyd, R. B. Effects of Photoperiod and Starvation on the Temperature Tolerance of Larvae of
829 the Giant Toad, *Bufo marinus*. *Copeia* **1985**, 625–631 (1985).
- 830 177. Fong, S.-T. Thermal tolerance of adult Asiatic painted frog *Kaloula pulchra* from different
831 populations. (National University of Tainan, Tainan, Taiwan, 2014).
- 832 178. Fontenot, C. L. & Lutterschmidt, W. I. Thermal selection and temperature preference of the
833 aquatic salamander, *Amphiuma tridactylum*. *Herpetological Conservation and Biology* **6**, 395–
834 399 (2011).
- 835 179. Freidenburg, L. K. & Skelly, D. K. Microgeographical variation in thermal preference by an
836 amphibian. *Ecology Letters* **7**, 369–373 (2004).
- 837 180. Frishkoff, L. O., Hadly, E. A. & Daily, G. C. Thermal niche predicts tolerance to habitat
838 conversion in tropical amphibians and reptiles. *Global Change Biology* **21**, 3901–3916 (2015).
- 839 181. Frost, J. S. & Martin, E. W. A Comparison of Distribution and High Temperature Tolerance in
840 *Bufo americanus* and *Bufo woodhousii fowleri*. *Copeia* **1971**, 750 (1971).
- 841 182. Galindo, C. A., Cruz, E. X. & Bernal, M. H. Evaluation of the combined temperature and
842 relative humidity preferences of the Colombian terrestrial salamander *Bolitoglossa ramosi*
843 (Amphibia: Plethodontidae). *Can. J. Zool.* **96**, 1230–1235 (2018).
- 844 183. Gatten, R. E. & Hill, C. J. Social influence on thermal selection by *Hyla crucifer*. *Journal of*
845 *Herpetology* **18**, 87–88 (1984).

- 846 184. Gatz, A. J. Critical Thermal Maxima of *Ambystoma maculatum* (Shaw) and *Ambystoma*
- 847 *jeffersonianum* (Green) in Relation to Time of Breeding. *Herpetologica* **27**, 157–160 (1971).
- 848 185. Gatz, A. J. Intraspecific Variations in Critical Thermal Maxima of *Ambystoma maculatum*.
Herpetologica **29**, 264–268 (1973).
- 849 186. Geise, W. & Linsenmair, K. E. Adaptations of the reed frog *Hyperolius viridiflavus* (Amphibia,
850 Anura, Hyperoliidae) to its arid environment - IV. Ecological significance of water economy
851 with comments on thermoregulation and energy allocation. *Oecologia* **77**, 327–338 (1988).
- 852 187. Gélinas, N. Rôle du comportement thermorégulateur chez la grenouille du nord (*Rana*
853 *septentrionalis*, Baird) adulte et le ouaouaron (*Rana catesbeiana*, Shaw) juvénile en rapport avec
854 le besoin alimentaire. (Université du Québec à Trois-Rivières, Trois-Rivières, Québec, Canada,
855 1996).
- 856 188. Goldstein, J. A., Hoff, K. von S. & Hillyard, S. D. The effect of temperature on development
857 and behaviour of relict leopard frog tadpoles. *Conservation Physiology* **5**, cow075 (2017).
- 858 189. González-del-Pliego, P. *et al.* Thermal tolerance and the importance of microhabitats for Andean
859 frogs in the context of land use and climate change. *Journal of Animal Ecology* **89**, 2451–2460
860 (2020).
- 861 190. Gouveia, S. F. *et al.* Climatic niche at physiological and macroecological scales: The thermal
862 tolerance-geographical range interface and niche dimensionality. *Global Ecology and*
863 *Biogeography* **23**, 446–456 (2014).
- 864 191. Gray, R. Lack of physiological differentiation in three color morphs of the cricket frog (*Acris*
865 *crepitans*) in Illinois. *Transactions of the Illinois State Academy of Science* **70**, 73–79 (1977).
- 866 192. Greenspan, S. E. *et al.* Infection increases vulnerability to climate change via effects on host
867 thermal tolerance. *Scientific Reports* **7**, (2017).
- 868 193. Guevara-Molina, E. C., Gomes, F. R. & Camacho, A. Effects of dehydration on
869 thermoregulatory behavior and thermal tolerance limits of *Rana catesbeiana* (Shaw, 1802).
870 *Journal of Thermal Biology* **93**, (2020).
- 871 194. Gutiérrez Pesquera, L. Una valoración macrofisiológica de la vulnerabilidad al calentamiento
872 global. Análisis de los límites de tolerancia térmica en comunidades de anfibios en gradientes
873 latitudinales y altitudinales. (Pontificia Universidad Católica Del Ecuador, Quito, Ecuador,
874 2015).
- 875 195. Gutiérrez Pesquera, M. Thermal tolerance across latitudinal and altitudinal gradients in tadpoles.
876 (Universidad de Sevilla, Sevilla, Spain, 2016).
- 877 196. Gutiérrez-Pesquera, L. M. *et al.* Testing the climate variability hypothesis in thermal tolerance
878 limits of tropical and temperate tadpoles. *Journal of Biogeography* **43**, 1166–1178 (2016).
- 879 197. Gvoždík, L., Puky, M. & Šugerková, M. Acclimation is beneficial at extreme test temperatures
880 in the Danube crested newt, *Triturus dobrogicus* (Caudata, Salamandridae). *Biological Journal
881 of the Linnean Society* **90**, 627–636 (2007).
- 882 198. Gvoždík, L. Does reproduction influence temperature preferences in newts? *Can. J. Zool.* **83**,
883 1038–1044 (2005).
- 884 199. Gvoždík, L. Mismatch Between Ectotherm Thermal Preferenda and Optima for Swimming: A
885 Test of the Evolutionary Pace Hypothesis. *Evol Biol* **42**, 137–145 (2015).
- 886 200. Gvoždík, L. & Kristín, P. Economic thermoregulatory response explains mismatch between
887 thermal physiology and behaviour in newts. *Journal of Experimental Biology* **220**, 1106–1111
888 (2017).
- 889 201. Gvoždík, L. Postprandial thermophily in the Danube crested newt, *Triturus dobrogicus*. *Journal
890 of Thermal Biology* **28**, 545–550 (2003).
- 891 202. Hadamová, M. & Gvoždík, L. Seasonal acclimation of preferred body temperatures improves
892 the opportunity for thermoregulation in newts. *Physiol Biochem Zool* **84**, 166–174 (2011).
- 893 203. Haggerty, J. Thermal tolerance of the common coqui frog (*Eleutherodactylus coqui*) in East
894 Hawaii along an elevation gradient. *ProQuest Dissertations and Theses* (University of Hawai'i
895 at Hilo, United States -- Hawaii, 2016).
- 896 204. Hanna, A. The effects of temperature on physiology in frogs and their dispersal of the temperate
897 zone from the tropics. (Truman State University, Kirksville, Missouri, USA, 2019).
- 898

- 899 205. He, J. *et al.* Influence of High Temperatures and Heat Wave on Thermal Biology, Locomotor
900 Performance, and Antioxidant System of High-Altitude Frog *Nanorana pleskei* Endemic to
901 Qinghai-Tibet Plateau. *Front. Ecol. Evol.* **9**, (2021).
- 902 206. Heath, A. G. Behavioral Thermoregulation in High Altitude Tiger Salamanders, *Ambystoma*
903 *tigrinum*. *Herpetologica* **31**, 84–93 (1975).
- 904 207. Heatwole, H., De Austin, S. B. & Herrero, R. Heat tolerances of tadpoles of two species of
905 tropical anurans. *Comparative Biochemistry And Physiology* **27**, 807–815 (1968).
- 906 208. Heatwole, H., Mercado, N. & Ortiz, E. Comparison of Critical Thermal Maxima of Two Species
907 of Puerto Rican Frogs of the Genus *Eleutherodactylus*. *Physiological Zoology* **38**, 1–8 (1965).
- 908 209. Holzman, N. & McManus, J. J. Effects of acclimation on metabolic rate and thermal tolerance in
909 the carpenter frog, *Rana vergatipes*. *Comparative Biochemistry and Physiology -- Part A:*
910 *Physiology* **45**, 833–842 (1973).
- 911 210. Hoppe, D. M. Thermal Tolerance in Tadpoles of the Chorus Frog *Pseudacris triseriata*.
912 *Herpetologica* **34**, 318–321 (1978).
- 913 211. Hou, P.-C. Thermal tolerance and preference in the adult amphibians from different altitudinal
914 LTER sites. (National Cheng Kung University, Tainan, Taiwan, 2003).
- 915 212. Howard, J. H., Wallace, R. L. & Stauffer Jr, J. R. Critical thermal maxima in populations of
916 *Ambystoma macrodactylum* from different elevations. *Journal of Herpetology* **17**, 400–402
917 (1983).
- 918 213. Hutchison, V. H. & Ritchart, J. P. Annual cycle of thermal tolerance in the salamander, *Necturus*
919 *maculosus*. *Journal of Herpetology* **23**, 73–76 (1989).
- 920 214. Hutchison, V. H. & Murphy, K. Behavioral thermoregulation in the salamander *Necturus*
921 *maculosus* after heat shock. *Comparative Biochemistry and Physiology Part A: Physiology* **82**,
922 391–394 (1985).
- 923 215. Hutchison, V. H. The Distribution and Ecology of the Cave Salamander, *Eurycea lucifuga*.
924 *Ecological Monographs* **28**, 2–20 (1958).
- 925 216. Hutchison, V. H. Critical Thermal Maxima in Salamanders. *Physiological Zoology* **34**, 92–125
926 (1961).
- 927 217. Hutchison, V. H., Engbretson, G. & Turney, D. Thermal Acclimation and Tolerance in the
928 Hellbender, *Cryptobranchus alleganiensis*. *Copeia* **1973**, 805–807 (1973).
- 929 218. Hutchison, V. H. & Rowlan, S. D. Thermal Acclimation and Tolerance in the Mudpuppy,
930 *Necturus maculosus*. *Journal of Herpetology* **9**, 367–368 (1975).
- 931 219. Hutchison, V. H. & Spiestersbach, K. K. Diel and Seasonal Cycles of Activity and Behavioral
932 Thermoregulation in the Salamander *Necturus maculosus*. *Copeia* **1986**, 612–618 (1986).
- 933 220. Jara Méndez, D., Krumel Castillo, M. & San Martín Venegas, E. Estudio de las preferencias
934 térmicas y niveles de profundidad en tres estadios larvales de la especie *Pleurodema thaul*
935 (Lesson, 1826) del Parque Nacional Laguna del Laja. (Universidad de Concepción, Los
936 Ángeles, Chile, 2020).
- 937 221. Jiang, S., Yu, P. & Hu, Q. A study on the critical thermal maxima of five species of salamanders
938 of China. *Acta Herpetologica Sinica* **6**, 56–62 (1987).
- 939 222. John-Alder, H. B., Morin, P. J. & Lawler, S. Thermal Physiology, Phenology, and Distribution of
940 Tree Frogs. *The American Naturalist* **132**, 506–520 (1988).
- 941 223. Johnson, C. R. Daily variation in the thermal tolerance of *Litoria caerulea* (Anura: Hylidae).
942 *Comparative Biochemistry and Physiology -- Part A: Physiology* **40**, 1109–1111 (1971).
- 943 224. Johnson, C. R. Thermal relations and water balance in the day frog, *Taudactylus diurnus*, from
944 an Australian rain forest. *Australian Journal of Zoology* **19**, 35–39 (1971).
- 945 225. Johnson, C. R. Diel variation in the thermal tolerance of *Litoria gracilenta* (Anura: Hylidae).
946 *Comparative Biochemistry and Physiology -- Part A: Physiology* **41**, 727–730 (1972).
- 947 226. Johnson, C. R. & Prine, J. E. The effects of sublethal concentrations of organophosphorus
948 insecticides and an insect growth regulator on temperature tolerance in hydrated and dehydrated
949 juvenile western toads, *Bufo boreas*. *Comparative Biochemistry and Physiology -- Part A:*
950 *Physiology* **53**, 147–149 (1976).
- 951 227. Johnson, C. R. Observations on body temperatures, critical thermal maxima and tolerance to
952 water loss in the Australian hylid, *Hyla caerulea* (White). *Proceedings of the Royal Society of*
953 *Queensland* **82**, 47–50 (1970).

- 954 228. Johnson, C. R. Thermal Relations and Daily Variation in the Thermal Tolerance in *Bufo*
955 *marinus*. *Journal of Herpetology* **6**, 35 (1972).
- 956 229. Johnson, C. Thermal relations in some southern and eastern Australian anurans. *Proceedings of*
957 *the Royal Society of Queensland* **82**, 87–94 (1971).
- 958 230. Johnson, C. The effects of five organophosphorus insecticides on thermal stress in tadpoles of
959 the Pacific tree frog, *Hyla regilla*. *Zoological Journal of the Linnean Society* **69**, 143–147
960 (1980).
- 961 231. Katzenberger, M., Duarte, H., Relyea, R., Beltrán, J. F. & Tejedo, M. Variation in upper thermal
962 tolerance among 19 species from temperate wetlands. *Journal of Thermal Biology* **96**, (2021).
- 963 232. Katzenberger, M. *et al.* Swimming with predators and pesticides: How environmental stressors
964 affect the thermal physiology of tadpoles. *PLoS ONE* **9**, (2014).
- 965 233. Katzenberger, M., Hammond, J., Tejedo, M. & Relyea, R. Source of environmental data and
966 warming tolerance estimation in six species of North American larval anurans. *Journal of*
967 *Thermal Biology* **76**, 171–178 (2018).
- 968 234. Katzenberger, M. Thermal tolerance and sensitivity of amphibian larvae from Palearctic and
969 Neotropical communities. (Universidade de Lisboa, Lisboa, Portugal, 2013).
- 970 235. Katzenberger, M. Impact of global warming in holarctic and neotropical communities of
971 amphibians. (Universidad de Sevilla, Sevilla, Spain, 2014).
- 972 236. Keen, W. H. & Schroeder, E. E. Temperature Selection and Tolerance in Three Species of
973 *Ambystoma* Larvae. *Copeia* **1975**, 523–530 (1975).
- 974 237. Kern, P., Cramp, R. L. & Franklin, C. E. Temperature and UV-B-insensitive performance in
975 tadpoles of the ornate burrowing frog: An ephemeral pond specialist. *Journal of Experimental*
976 *Biology* **217**, 1246–1252 (2014).
- 977 238. Kern, P., Cramp, R. L., Seebacher, F., Ghanizadeh Kazerouni, E. & Franklin, C. E. Plasticity of
978 protective mechanisms only partially explains interactive effects of temperature and UVR on
979 upper thermal limits. *Comparative Biochemistry and Physiology -Part A : Molecular and*
980 *Integrative Physiology* **190**, 75–82 (2015).
- 981 239. Kern, P., Cramp, R. L. & Franklin, C. E. Physiological responses of ectotherms to daily
982 temperature variation. *Journal of Experimental Biology* **218**, 3068–3076 (2015).
- 983 240. Kirsch, D. R., Fix, S., Davenport, J. M., Cecala, K. K. & Ennen, J. R. Body Size Is Related to
984 Temperature Preference in *Hyla chrysoscelis* Tadpoles. *Journal of Herpetology* **55**, 21–25
985 (2021).
- 986 241. Köhler, A. *et al.* Staying warm or moist? Operative temperature and thermal preferences of
987 common frogs (*Rana temporaria*), and effects on locomotion. *The Herpetological Journal* **21**,
988 17–26 (2011).
- 989 242. Kolbe, J. J., Kearney, M. & Shine, R. Modeling the consequences of thermal trait variation for
990 the cane toad invasion of Australia. *Ecological Applications* **20**, 2273–2285 (2010).
- 991 243. Komaki, S., Igawa, T., Lin, S.-M. & Sumida, M. Salinity and thermal tolerance of Japanese
992 stream tree frog (*Buergeria japonica*) tadpoles from island populations. *Herpetological Journal*
993 **26**, 207–211 (2016).
- 994 244. Komaki, S., Lau, Q. & Igawa, T. Living in a Japanese onsen: Field observations and
995 physiological measurements of hot spring amphibian tadpoles, *Buergeria japonica*. *Amphibia*
996 *Reptilia* **37**, 311–314 (2016).
- 997 245. Krakauer, T. Tolerance limits of the toad, *Bufo marinus*, in South Florida. *Comparative*
998 *Biochemistry And Physiology* **33**, 15–26 (1970).
- 999 246. Kurabayashi, A. *et al.* Improved transport of the model amphibian, *Xenopus tropicalis*, and its
1000 viable temperature for transport. *Current Herpetology* **33**, 75–87 (2014).
- 1001 247. Lange, L. Influences environnementales précoces et plasticité phénotypique : étude d'un modèle
1002 amphibien avec soins parentaux prénataux, l'Alyte accoucheur. (Université de La Rochelle, La
1003 Rochelle, France, 2020).
- 1004 248. Lange, Z. THERMAL QUALITY EXPLAINS SHIFT IN HABITAT ASSOCIATION FROM
1005 FOREST TO CLEARINGS FOR TERRESTRIAL-BREEDING FROGS ALONG AN
1006 ELEVATION GRADIENT IN COLOMBIA. (John Carroll University, University Heights, Ohio,
1007 USA, 2019).

- 1008 249. Lau, E. T. C., Leung, K. M. Y. & Karraker, N. E. Native amphibian larvae exhibit higher upper
1009 thermal limits but lower performance than their introduced predator *Gambusia affinis*. *Journal*
1010 *of Thermal Biology* **81**, 154–161 (2019).
- 1011 250. Layne, J. R. Freeze tolerance and cryoprotectant mobilization in the gray treefrog (*Hyla*
1012 *versicolor*). *J Exp Zool* **283**, 221–225 (1999).
- 1013 251. Layne, J. R. & Claussen, D. L. Seasonal variation in the thermal acclimation of critical thermal
1014 maxima (CTMax) and minima (CTMin) in the salamander *Eurycea bislineata*. *Journal of*
1015 *Thermal Biology* **7**, 29–33 (1982).
- 1016 252. Layne, J. R. & Claussen, D. L. The time courses of CTMax and CTMin acclimation in the
1017 salamander *Desmognathus fuscus*. *Journal of Thermal Biology* **7**, 139–141 (1982).
- 1018 253. Layne, J. R. & Claussen, D. L. Time courses of thermal acclimation for critical thermal minima
1019 in the salamanders *Desmognathus quadramaculatus*, *Desmognathus monticola*, *Desmognathus*
1020 *ochrophaeus*, and *Plethodon jordani*. *Comparative Biochemistry and Physiology Part A:*
1021 *Physiology* **87**, 895–898 (1987).
- 1022 254. Layne, J. R. Seasonal variation in the cryobiology of *Rana sylvatica* from Pennsylvania. *Journal*
1023 *of Thermal Biology* **20**, 349–353 (1995).
- 1024 255. Layne, J. R. & Romano, M. A. Critical Thermal Minima of *Hyla chrysoscelis*, *H. cinerea*, *H.*
1025 *gratiosa* and Natural Hybrids (*H. cinerea* × *H. gratiosa*). *Herpetologica* **41**, 216–221 (1985).
- 1026 256. Lee, P.-T. Acidic effect on tadpoles living in container habitats. (Tunghai University, Taichung,
1027 Taiwan, 2019).
- 1028 257. Leger, J. P. & Mathieson, W. B. Effects of Bombesin on Behavioral Thermoregulation in the
1029 Bullfrog. *Brain Behavior and Evolution* **50**, 304–312 (2008).
- 1030 258. Li, Z. *et al.* Landscape Connectivity Limits the Predicted Impact of Fungal Pathogen Invasion.
1031 *Journal of Fungi* **6**, 205 (2020).
- 1032 259. Lillywhite, H. B. Temperature selection by the bullfrog, *Rana catesbeiana*. *Comparative*
1033 *Biochemistry and Physiology Part A: Physiology* **40**, 213–227 (1971).
- 1034 260. Litmer, A. R. & Murray, C. M. Critical Thermal Capacities of *Hyla chrysoscelis* in Relation to
1035 Season. *Journal of Herpetology* **54**, 413–417 (2020).
- 1036 261. Llewellyn, D., Brown, G. P., Thompson, M. B. & Shine, R. Behavioral Responses to Immune-
1037 System Activation in an Anuran (the Cane Toad, *Bufo marinus*): Field and Laboratory Studies.
1038 *Physiological and Biochemical Zoology* **84**, 77–86 (2011).
- 1039 262. Longhini, L. S., De Almeida Prado, C. P., Bícego, K. C., Zena, L. A. & Gargaglioni, L. H.
1040 Measuring cardiorespiratory variables on small tadpoles using a non-invasive methodology.
1041 *Revista Cubana de Investigaciones Biomedicas* **38**, (2019).
- 1042 263. López Rosero, A. C. Ontogenetic variation of thermal tolerance in two anuran species of
1043 Ecuador: *Gastrotheca pseustes* (Hemiphractidae) and *Smilisca phaeota* (Hylidae) and their
1044 relative vulnerability to environmental temperature change. (Pontificia Universidad Católica Del
1045 Ecuador, Quito, Ecuador, 2015).
- 1046 264. Lotshaw, D. P. Temperature adaptation and effects of thermal acclimation in *Rana sylvatica* and
1047 *Rana catesbeiana*. *Comparative Biochemistry and Physiology -- Part A: Physiology* **56**, 287–294
1048 (1977).
- 1049 265. Lu, H.-L., Wu, Q., Geng, J. & Dang, W. Swimming performance and thermal resistance of
1050 juvenile and adult newts acclimated to different temperatures. *Acta Herpetologica* **11**, 189–195
1051 (2016).
- 1052 266. Lu, H. L., Geng, J., Xu, W., Ping, J. & Zhang, Y. P. Physiological response and changes in
1053 swimming performance after thermal acclimation in juvenile Chinese fire-belly newts, *Cynops*
1054 *orientalis*. *Acta Ecologica Sinica* **37**, 1603–1610 (2017).
- 1055 267. Lutterschmidt, W. I. & Hutchison, V. H. The critical thermal maximum: Data to support the
1056 onset of spasms as the definitive end point. *Canadian Journal of Zoology* **75**, 1553–1560
1057 (1997).
- 1058 268. Madalozzo, B. Variação latitudinal nos limites de tolerância e plasticidade térmica em anfíbios
1059 em um cenário de mudanças climáticas: efeito dos micro-habitats, sazonalidade e filogenia.
1060 (Universidade Federal de Santa Maria, Santa Maria, Brazil, 2018).
- 1061 269. Mahoney, J. J. & Hutchison, V. H. Photoperiod acclimation and 24-hour variations in the critical
1062 thermal maxima of a tropical and a temperate frog. *Oecologia* **2**, 143–161 (1969).

- 1063 270. Malvin, G. M. & Wood, S. C. Behavioral thermoregulation of the toad, *Bufo marinus*: Effects of
1064 air humidity. *Journal of Experimental Zoology* **258**, 322–326 (1991).
- 1065 271. Maness, J. D. & Hutchison, V. H. Acute adjustment of thermal tolerance in vertebrate
1066 ectotherms following exposure to critical thermal maxima. *Journal of Thermal Biology* **5**, 225–
1067 233 (1980).
- 1068 272. Manis, M. L. & Claussen, D. L. Environmental and genetic influences on the thermal
1069 physiology of *Rana sylvatica*. *Journal of Thermal Biology* **11**, 31–36 (1986).
- 1070 273. Markle, T. M. & Kozak, K. H. Low acclimation capacity of narrow-ranging thermal specialists
1071 exposes susceptibility to global climate change. *Ecology and Evolution* **8**, 4644–4656 (2018).
- 1072 274. Markle, T. Ecology and Evolution of Geographic Range Size Variation in North American
1073 Plethodontid Salamanders: Perspectives from Thermal Physiology. (University of Minnesota
1074 Twin Cities, Minneapolis, Minnesota, USA, 2015).
- 1075 275. Marshall, E. & Grigg, G. C. Acclimation of CTM, LD50, and Rapid Loss of Acclimation of
1076 Thermal Preferendum in Tadpoles of *Limnodynastes peronii* (Anura, Myobatrachidae). *The
1077 Australian Zoologist* **20**, 447–456 (1980).
- 1078 276. Mathias, J. H. The Comparative Ecologies of Two Species of Amphibia (*B. bufo* and *B.
1079 calamita*) on the Ainsdale Sand Dunes National Nature Reserve. (The University of Manchester,
1080 United Kingdom, 1971).
- 1081 277. McCann, S., Greenlees, M. J., Newell, D. & Shine, R. Rapid acclimation to cold allows the cane
1082 toad to invade montane areas within its Australian range. *Functional Ecology* **28**, 1166–1174
1083 (2014).
- 1084 278. McCann, S. M., Kosmala, G. K., Greenlees, M. J. & Shine, R. Physiological plasticity in a
1085 successful invader: rapid acclimation to cold occurs only in cool-climate populations of cane
1086 toads (*Rhinella marina*). *Conservation Physiology* **6**, cox072 (2018).
- 1087 279. McManus, J. J. & Nellis, D. W. The critical thermal maximum of the marine toad, *Bufo
1088 marinus*. *Caribbean Journal of Science* **15**, 67–70 (1975).
- 1089 280. Menke, M. E. & Claussen, D. L. Thermal acclimation and hardening in tadpoles of the bullfrog,
1090 *Rana catesbeiana*. *Journal of Thermal Biology* **7**, 215–219 (1982).
- 1091 281. Merino-Viteri, A. R. The vulnerability of microhylid frogs, *Cophixalus* spp., to climate change
1092 in the Australian Wet Tropics. (James Cook University, Townsville, Australia, 2018).
- 1093 282. Messerman, A. F. Tales of an ‘Invisible’ Life Stage: Survival and Physiology Among Terrestrial
1094 Juvenile Ambystomatid Salamanders. (University of Missouri, Columbia, Missouri, USA,
1095 2019).
- 1096 283. Meza-Parral, Y., García-Robledo, C., Pineda, E., Escobar, F. & Donnelly, M. A. Standardized
1097 ethograms and a device for assessing amphibian thermal responses in a warming world. *Journal
1098 of Thermal Biology* **89**, (2020).
- 1099 284. Miller, K. & Packard, G. C. Critical thermal maximum: Ecotypic variation between montane
1100 and piedmont chorus frogs (*Pseudacris triseriata*, Hylidae). *Experientia* **30**, 355–356 (1974).
- 1101 285. Miller, K. & Packard, G. C. An Altitudinal Cline in Critical Thermal Maxima of Chorus Frogs
1102 (*Pseudacris triseriata*). *The American Naturalist* **111**, 267–277 (1977).
- 1103 286. Mittan, C. S. & Zamudio, K. R. Rapid adaptation to cold in the invasive cane toad *Rhinella
1104 marina*. *Conservation Physiology* **7**, coy075 (2019).
- 1105 287. Moretti, E. H., Ortega Chinchilla, J. E., Marques, F. S., Fernandes, P. A. C. M. & Gomes, F. R.
1106 Behavioral fever decreases metabolic response to lipopolysaccharide in yellow Cururu toads
1107 (*Rhinella icterica*). *Physiology & Behavior* **191**, 73–81 (2018).
- 1108 288. Mueller, C. A., Bucsky, J., Korito, L. & Manzanares, S. Immediate and persistent effects of
1109 temperature on oxygen consumption and thermal tolerance in embryos and larvae of the baja
1110 California chorus frog, *pseudacris hypochondriaca*. *Frontiers in Physiology* **10**, (2019).
- 1111 289. Mullens, D. P. & Hutchison, V. H. Diel, seasonal, postprandial and food-deprived
1112 thermoregulatory behaviour in tropical toads (*Bufo marinus*). *Journal of Thermal Biology* **17**,
1113 63–67 (1992).
- 1114 290. Navas, C. A., Antoniazzi, M. M., Carvalho, J. E., Suzuki, H. & Jared, C. Physiological basis for
1115 diurnal activity in dispersing juvenile *Bufo granulosus* in the Caatinga, a Brazilian semi-arid
1116 environment. *Comparative Biochemistry and Physiology - A Molecular and Integrative
1117 Physiology* **147**, 647–657 (2007).

- 1118 291. Navas, C. A., Úbeda, C. A., Logares, R. & Jara, F. G. Thermal Tolerances in Tadpoles of Three
1119 Species of Patagonian Anurans. *South American Journal of Herpetology* **5**, 89–96 (2010).
- 1120 292. Nietfeldt, J. W., Jones, S. M., Droege, D. L. & Ballinger, R. E. Rate of thermal acclimation of
1121 larval *Ambystoma tigrinum*. *Journal of Herpetology* **14**, 209–211 (1980).
- 1122 293. Nol, Rosemarie & Ultsch, G. R. The Roles of Temperature and Dissolved Oxygen in
1123 Microhabitat Selection by the Tadpoles of a Frog (*Rana pipiens*) and a Toad (*Bufo terrestris*).
1124 *Copeia* **1981**, 645–652 (1981).
- 1125 294. Novarro, A. J. Thermal Physiology in a Widespread Lungless Salamander. (University of
1126 Maryland, College Park, Maryland, USA, 2018).
- 1127 295. Nowakowski, A. J. *et al.* Thermal biology mediates responses of amphibians and reptiles to
1128 habitat modification. *Ecology Letters* **21**, 345–355 (2018).
- 1129 296. Nowakowski, A. J. *et al.* Tropical amphibians in shifting thermal landscapes under land-use and
1130 climate change. *Conservation Biology* **31**, 96–105 (2017).
- 1131 297. O'Connor, M. P. & Tracy, C. R. Thermoregulation by Juvenile Toads of *Bufo woodhousei* in the
1132 Field and in the Laboratory. *Copeia* **1992**, 865–876 (1992).
- 1133 298. Orille, A. C., McWhinnie, R. B., Brady, S. P. & Raffel, T. R. Positive Effects of Acclimation
1134 Temperature on the Critical Thermal Maxima of *Ambystoma mexicanum* and *Xenopus laevis*.
1135 *Journal of Herpetology* **54**, 289–292 (2020).
- 1136 299. Oyamaguchi, H. M. *et al.* Thermal sensitivity of a Neotropical amphibian (*Engystomops*
1137 *pustulosus*) and its vulnerability to climate change. *Biotropica* **50**, 326–337 (2018).
- 1138 300. Paez Vacas, M. I. Mechanisms of population divergence along elevational gradients in the
1139 tropics. (Colorado State University, Fort Collins, Colorado, USA, 2016).
- 1140 301. Paulson, B. K. & Hutchison, V. H. Blood changes in *Bufo cognatus* following acute heat stress.
1141 *Comparative Biochemistry and Physiology -- Part A: Physiology* **87**, 461–466 (1987).
- 1142 302. Paulson, B. & Hutchison, V. Origin of the stimulus for muscular spasms at the critical thermal
1143 maximum in anurans. *Copeia* 810–813 (1987).
- 1144 303. Percino-Daniel, R. *et al.* Environmental heterogeneity shapes physiological traits in tropical
1145 direct-developing frogs. *Ecology and Evolution* (2021).
- 1146 304. Perotti, M. G., Bonino, M. F., Ferraro, D. & Cruz, F. B. How sensitive are temperate tadpoles to
1147 climate change? The use of thermal physiology and niche model tools to assess vulnerability.
1148 *Zoology* **127**, 95–105 (2018).
- 1149 305. Piasečná, K., Pončová, A., Tejedo, M. & Gvoždík, L. Thermoregulatory strategies in an aquatic
1150 ectotherm from thermally-constrained habitats: An evaluation of current approaches. *Journal of*
1151 *Thermal Biology* **52**, 97–107 (2015).
- 1152 306. Pintanel, P., Tejedo, M., Almeida-Reinoso, F., Merino-Viteri, A. & Gutiérrez-Pesquera, L. M.
1153 Critical thermal limits do not vary between wild-caught and captive-bred tadpoles of *agalychnis*
1154 *spurrelli* (Anura: Hylidae). *Diversity* **12**, (2020).
- 1155 307. Pintanel, P., Tejedo, M., Ron, S. R., Llorente, G. A. & Merino-Viteri, A. Elevational and
1156 microclimatic drivers of thermal tolerance in Andean *Pristimantis* frogs. *Journal of*
1157 *Biogeography* **46**, 1664–1675 (2019).
- 1158 308. Pintanel, P. Thermal adaptation of amphibians in tropical mountains. Consequences of global
1159 warming. (Universitat de Barcelona, Barcelona, Spain, 2018).
- 1160 309. Pintanel, P., Tejedo, M., Salinas-Ivanenko, S., Jervis, P. & Merino-Viteri, A. Predators like it hot:
1161 Thermal mismatch in a predator-prey system across an elevational tropical gradient. *The Journal*
1162 *of animal ecology* (2021).
- 1163 310. Pough, F. H. Natural daily temperature acclimation of eastern red efts, *Notophthalmus v.*
1164 *viridescens* (rafinesque) (amphibia: caudata). *Comparative Biochemistry and Physiology -- Part*
1165 *A: Physiology* **47**, 71–78 (1974).
- 1166 311. Pough, F. H., Stewart, M. M. & Thomas, R. G. Physiological basis of habitat partitioning in
1167 Jamaican *Eleutherodactylus*. *Oecologia* **27**, 285–293 (1977).
- 1168 312. Quiroga, L. B., Sanabria, E. A., Fornés, M. W., Bustos, D. A. & Tejedo, M. Sublethal
1169 concentrations of chlorpyrifos induce changes in the thermal sensitivity and tolerance of anuran
1170 tadpoles in the toad *Rhinella arenarum*? *Chemosphere* **219**, 671–677 (2019).

- 1171 313. Rausch, C. M., Starkweather, P. L. & van Breukelen, F. One year in the life of *Bufo punctatus*:
1172 annual patterns of body temperature in a free-ranging desert anuran. *Naturwissenschaften* **95**,
1173 531–535 (2008).
- 1174 314. Rausch, C. The thermal ecology of the Red-spotted toad, *Bufo punctatus*, across life history.
1175 (University of Nevada, Las Vegas, Nevada, USA, 2007).
- 1176 315. Reichenbach, N. & Brophy, T. R. Natural history of the peaks of otter salamander (*Plethodon*
1177 *Hubrichti*) along an Elevational gradient. *Herpetological Bulletin* 7–15 (2017).
- 1178 316. Reider, K. E., Larson, D. J., Barnes, B. M. & Donnelly, M. A. Thermal adaptations to extreme
1179 freeze–thaw cycles in the high tropical Andes. *Biotropica* **53**, 296–306 (2021).
- 1180 317. Richter-Boix, A. *et al.* Local divergence of thermal reaction norms among amphibian
1181 populations is affected by pond temperature variation. *Evolution* **69**, 2210–2226 (2015).
- 1182 318. Riquelme, N. A., Díaz-Páez, H. & Ortiz, J. C. Thermal tolerance in the Andean toad *Rhinella*
1183 *spinulosa* (Anura: Bufonidae) at three sites located along a latitudinal gradient in Chile. *Journal*
1184 *of Thermal Biology* **60**, 237–245 (2016).
- 1185 319. Ritchart, J. P. & Hutchison, V. H. The effects of ATP and cAMP on the thermal tolerance of the
1186 mudpuppy, *Necturus maculosus*. *Journal of Thermal Biology* **11**, 47–51 (1986).
- 1187 320. Rivera-Burgos, A. C. Habitat Suitability for Eleutherodactylus Frogs in Puerto Rico: Indexing
1188 Occupancy, Abundance and Reproduction to Climatic and Habitat Characteristics. (North
1189 Carolina State University, Raleigh, North Carolina, USA, 2019).
- 1190 321. Rivera-Ordonez, J. M., Justin Nowakowski, A., Manansala, A., Thompson, M. E. & Todd, B. D.
1191 Thermal niche variation among individuals of the poison frog, *Oophaga pumilio*, in forest and
1192 converted habitats. *Biotropica* **51**, 747–756 (2019).
- 1193 322. Rocha, P. L. & Branco, L. G. S. Physiological significance of behavioral hypothermia in
1194 hypoglycemic frogs (*Rana catesbeiana*). *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* **119**, 957–961 (1998).
- 1195 323. Rodríguez, C. Y., Bustos, D. A. & Sanabria, E. A. Adaptation of the Andean Toad *Rhinella*
1196 *spinulosa* (Anura: Bufonidae) at Low Temperatures: The Role of Glucose as Cryoprotectant.
1197 *Physiological and Biochemical Zoology* **92**, 473–480 (2019).
- 1198 324. Romero Barreto, P. Requerimientos fisiológicos y microambientales de dos especies de anfibios
1199 (Scinax ruber e Hyloxalus yasuni) del bosque tropical de Yasuní y sus implicaciones
1200 (Pontificia Universidad Católica Del Ecuador, Quito, Ecuador, 2013).
- 1201 325. Ruiz-Aravena, M. *et al.* Impact of global warming at the range margins: Phenotypic plasticity
1202 and behavioral thermoregulation will buffer an endemic amphibian. *Ecology and Evolution* **4**,
1203 4467–4475 (2014).
- 1204 326. Ruthsatz, K. *et al.* Thyroid hormone levels and temperature during development alter thermal
1205 tolerance and energetics of *Xenopus laevis* larvae. *Conservation Physiology* **6**, (2018).
- 1206 327. Ruthsatz, K. *et al.* Post-metamorphic carry-over effects of altered thyroid hormone level and
1207 developmental temperature: physiological plasticity and body condition at two life stages in
1208 *Rana temporaria*. *Journal of Comparative Physiology B: Biochemical, Systemic, and*
1209 *Environmental Physiology* **190**, 297–315 (2020).
- 1210 328. Rutledge, P. S., Spotila, J. R. & Easton, D. P. Heat hardening in response to two types of heat
1211 shock in the lungless salamanders *Eurycea bislineata* and *Desmognathus ochrophaeus*. *Journal*
1212 *of Thermal Biology* **12**, 235–241 (1987).
- 1213 329. Sanabria, E. *et al.* Effect of salinity on locomotor performance and thermal extremes of
1214 metamorphic Andean Toads (*Rhinella spinulosa*) from Monte Desert, Argentina. *Journal of*
1215 *Thermal Biology* **74**, 195–200 (2018).
- 1216 330. Sanabria, E. A., González, E., Quiroga, L. B. & Tejedo, M. Vulnerability to warming in a desert
1217 amphibian tadpole community: the role of interpopulational variation. *Journal of Zoology* **313**,
1218 283–296 (2021).
- 1219 331. Sanabria, E. A. & Quiroga, L. B. Change in the thermal biology of tadpoles of *Odontophrynus*
1220 *occidentalis* from the Monte desert, Argentina: Responses to photoperiod. *Journal of Thermal*
1221 *Biology* **36**, 288–291 (2011).
- 1222 332. Sanabria, E. A., Quiroga, L. B., González, E., Moreno, D. & Cataldo, A. Thermal parameters
1223 and locomotor performance in juvenile of *Pleurodema nebulosum* (Anura: Leptodactylidae)
1224 from the Monte Desert. *Journal of Thermal Biology* **38**, 390–395 (2013).

- 1226 333. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal changes in the thermal tolerances of
1227 the toad *Rhinella arenarum* (Bufonidae) in the Monte Desert of Argentina. *Journal of Thermal*
1228 *Biology* **37**, 409–412 (2012).
- 1229 334. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal Changes in the thermal tolerances of
1230 *odontophryne occidentalis* (BERG, 1896) (Anura: Cycloramphidae). *Belgian Journal of*
1231 *Zoology* **143**, 23–29 (2013).
- 1232 335. Sanabria, E. A. *et al.* Thermal ecology of the post-metamorphic Andean toad (*Rhinella*
1233 *spinulosa*) at elevation in the monte desert, Argentina. *Journal of Thermal Biology* **52**, 52–57
1234 (2015).
- 1235 336. Sanabria, E. A., Vaira, M., Quiroga, L. B., Akmentins, M. S. & Pereyra, L. C. Variation of
1236 thermal parameters in two different color morphs of a diurnal poison toad, *Melanophryniscus*
1237 *ruberiventralis* (Anura: Bufonidae). *Journal of Thermal Biology* **41**, 1–5 (2014).
- 1238 337. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal changes in the thermoregulatory
1239 strategies of *Rhinella arenarum* in the Monte desert, Argentina. *Journal of Thermal Biology* **36**,
1240 23–28 (2011).
- 1241 338. Sanabria, E. A., Vergara, S. C., Rodríguez, C. Y. & Quiroga, L. B. Thermophilic response post
1242 feeding in *Pleurodema nebulosum* (Anura: Leptodactylidae) from Monte desert, Argentina.
1243 *Journal of Thermal Biology* **90**, 102605 (2020).
- 1244 339. Sanabria, E. A. & Quiroga, L. B. Thermal parameters changes in males of *Rhinella arenarum*
1245 (Anura: Bufonidae) related to reproductive periods. *Revista de Biología Tropical* **59**, 347–353
1246 (2011).
- 1247 340. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Variation in the Thermal Parameters of
1248 *Odontophryne occidentalis* in the Monte Desert, Argentina: Response to the Environmental
1249 Constraints. *Journal of Experimental Zoology Part A: Ecological Genetics and Physiology* **317**,
1250 185–193 (2012).
- 1251 341. Sauer, E. L. *et al.* Variation in individual temperature preferences, not behavioural fever, affects
1252 susceptibility to chytridiomycosis in amphibians. *Proceedings of the Royal Society B: Biological*
1253 *Sciences* **285**, 20181111 (2018).
- 1254 342. Sauer, E. L., Trejo, N., Hoverman, J. T. & Rohr, J. R. Behavioural fever reduces ranaviral
1255 infection in toads. *Functional Ecology* **33**, 2172–2179 (2019).
- 1256 343. Sauer, E. L. Behavioral Thermoregulation and Thermal Mismatches Influence Disease
1257 Dynamics in Amphibians. (University of South Florida, Tampa, Florida, USA, 2018).
- 1258 344. Scheffers, B. R. *et al.* Thermal buffering of microhabitats is a critical factor mediating warming
1259 vulnerability of frogs in the Philippine biodiversity hotspot. *Biotropica* **45**, 628–635 (2013).
- 1260 345. Scheffers, B. R., Edwards, D. P., Diesmos, A., Williams, S. E. & Evans, T. A. Microhabitats
1261 reduce animal's exposure to climate extremes. *Global Change Biology* **20**, 495–503 (2014).
- 1262 346. Schmid, W. D. Survival of frogs in low temperature. *Science* **215**, 697–698 (1982).
- 1263 347. Schmid, W. D. High Temperature Tolerances of *Bufo hemiophrys* and *Bufo cognatus*. *Ecology*
1264 **46**, 559–560 (1965).
- 1265 348. Seal, er, J. A. & West, B. W. Critical Thermal Maxima of Some Arkansas Salamanders in
1266 Relation to Thermal Acclimation. *Herpetologica* **25**, 122–124 (1969).
- 1267 349. Seibel, R. V. Variables Affecting the Critical Thermal Maximum of the Leopard Frog, *Rana*
1268 *pipiens Schreber*. *Herpetologica* **26**, 208–213 (1970).
- 1269 350. Sherman, E. Ontogenetic change in thermal tolerance of the toad *Bufo woodhousii fowleri*.
1270 *Comparative Biochemistry and Physiology -- Part A: Physiology* **65**, 227–230 (1980).
- 1271 351. Sherman, E. Thermal biology of newts (*Notophthalmus viridescens*) chronically infected with a
1272 naturally occurring pathogen. *Journal of Thermal Biology* **33**, 27–31 (2008).
- 1273 352. Sherman, E., Baldwin, L., Fern, ez, G. & Deurell, E. Fever and thermal tolerance in the toad
1274 *Bufo marinus*. *Journal of Thermal Biology* **16**, 297–301 (1991).
- 1275 353. Sherman, E. & Levitis, D. Heat hardening as a function of developmental stage in larval and
1276 juvenile *Bufo americanus* and *Xenopus laevis*. *Journal of Thermal Biology* **28**, 373–380 (2003).
- 1277 354. Shi, L., Zhao, L., Ma, X. & Ma, X. Selected body temperature and thermal tolerance of tadpoles
1278 of two frog species (*Fejervarya limnocharis* and *Microhyla ornata*) acclimated under different
1279 thermal conditions. *Acta Ecologica Sinica* **32**, 0465–0471 (2012).

- 1280 355. Siddons, S. R. & Searle, C. L. Exposure to a fungal pathogen increases the critical thermal
1281 minimum of two frog species. *Ecology and Evolution* **11**, 9589–9598 (2021).
- 1282 356. Sievert, L. M. Thermoregulatory behaviour in the toads *Bufo marinus* and *Bufo cognatus*.
1283 *Journal of Thermal Biology* **16**, 309–312 (1991).
- 1284 357. Sievert, L. M. & Andreadis, P. T. Differing Diel Patterns of Temperature Selection in Two
1285 Sympatric *Desmognathus*. *Copeia* **2002**, 62–66 (2002).
- 1286 358. Simon, M. N., Ribeiro, P. L. & Navas, C. A. Upper thermal tolerance plasticity in tropical
1287 amphibian species from contrasting habitats: Implications for warming impact prediction.
1288 *Journal of Thermal Biology* **48**, 36–44 (2015).
- 1289 359. Simon, M. Plasticidade fenotípica em relação à temperatura de larvas de *Rhinella* (Anura:
1290 Bufonidae) da caatinga e da floresta Atlântica. (Universidade de São Paulo, São Paulo, Brazil,
1291 2010).
- 1292 360. Skelly, D. K. & Freidenburg, L. K. Effects of beaver on the thermal biology of an amphibian.
1293 *Ecology Letters* **3**, 483–486 (2000).
- 1294 361. Smolinský, R. & Gvoždík, L. The ontogenetic shift in thermoregulatory behaviour of newt
1295 larvae: testing the ‘enemy-free temperatures’ hypothesis. *Journal of Zoology* **279**, 180–186
1296 (2009).
- 1297 362. Smolinský, R. & Gvoždík, L. Interactive influence of biotic and abiotic cues on the plasticity of
1298 preferred body temperatures in a predator–prey system. *Oecologia* **170**, 47–55 (2012).
- 1299 363. Sos, T. Thermoconformity even in hot small temporary water bodies: a case study in yellow-
1300 bellied toad (*Bombina v. variegata*). *Herpetologica Romanica* **1**, 1–11 (2007).
- 1301 364. Spotila, J. R. Role of Temperature and Water in the Ecology of Lungless Salamanders.
1302 *Ecological Monographs* **42**, 95–125 (1972).
- 1303 365. Strickland, J. C., Pinheiro, A. P., Cecala, K. K. & Dorcas, M. E. Relationship between
1304 Behavioral Thermoregulation and Physiological Function in Larval Stream Salamanders.
1305 *Journal of Herpetology* **50**, 239–244 (2016).
- 1306 366. Swanson, D. L., Graves, B. M. & Koster, K. L. Freezing tolerance/intolerance and
1307 cryoprotectant synthesis in terrestrially overwintering anurans in the Great Plains, USA. *J Comp
1308 Physiol B* **166**, 110–119 (1996).
- 1309 367. Tattersall, G. J. & Boutilier, R. G. Balancing Hypoxia and Hypothermia in Cold-Submerged
1310 Frogs. *Journal of Experimental Biology* **200**, 1031–1038 (1997).
- 1311 368. Tattersall, G. J., Tyson, T. M., Lenchyshyn, J. R. & Carbone, R. L. Temperature Preference
1312 During Forelimb Regeneration in the Red-Spotted Newt *otophthalmus Viridescens*. *Journal of
1313 Experimental Zoology Part A: Ecological Genetics and Physiology* **317**, 248–258 (2012).
- 1314 369. Toufarová, E. & Gvoždík, L. Do female newts modify thermoregulatory behavior to manipulate
1315 egg size? *Journal of Thermal Biology* **57**, 72–77 (2016).
- 1316 370. Tracy, C. R., Christian, K. A., O'Connor, M. P. & Tracy, C. R. Behavioral Thermoregulation by
1317 *Bufo americanus*: The Importance of the Hydric Environment. *Herpetologica* **49**, 375–382
1318 (1993).
- 1319 371. Tracy, C. R., Christian, K. A., Betts, G. & Tracy, C. R. Body temperature and resistance to
1320 evaporative water loss in tropical Australian frogs. *Comparative Biochemistry and Physiology -
1321 A Molecular and Integrative Physiology* **150**, 102–108 (2008).
- 1322 372. Tracy, C. R. & Christian, K. A. Preferred Temperature Correlates with Evaporative Water Loss
1323 in Hylid Frogs from Northern Australia. *Physiological and Biochemical Zoology* **78**, 839–846
1324 (2005).
- 1325 373. Trochet, A. *et al.* Variation of preferred body temperatures along an altitudinal gradient: A multi-
1326 species study. *Journal of Thermal Biology* **77**, 38–44 (2018).
- 1327 374. Turriago, J. L., Parra, C. A. & Bernal, M. H. Upper thermal tolerance in anuran embryos and
1328 tadpoles at constant and variable peak temperatures. *Canadian Journal of Zoology* **93**, 267–272
1329 (2015).
- 1330 375. Vidal, M. A., Novoa-Muñoz, F., Werner, E., Torres, C. & Nova, R. Modeling warming predicts a
1331 physiological threshold for the extinction of the living fossil frog *Calyptocephalella gayi*.
1332 *Journal of Thermal Biology* **69**, 110–117 (2017).

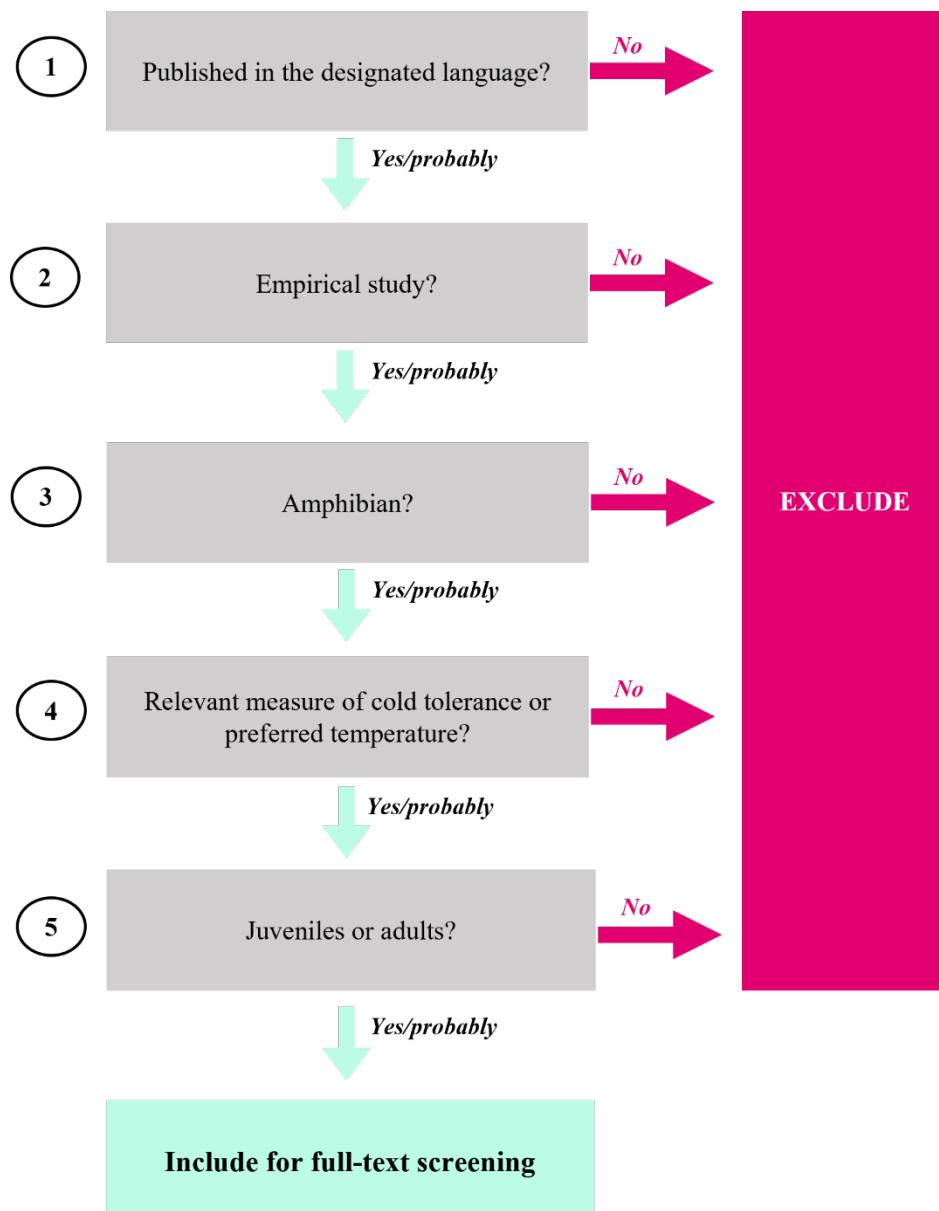
- 1333 376. Viertel, B. The reaction of suspension feeding anuran larvae to temperature, hyperbaric and
1334 hyperoxic waters. *Zoologische Jahrbücher. Abteilung für Systematik, Ökologie und Geographie*
1335 der Tiere
- 1336 120, 1–11 (1993).
- 1336 377. Vimercati, G. Exploring the invasion of the guttural toad Sclerophrys gutturalis in Cape Town
1337 through a multidisciplinary approach. (Stellenbosch University, Stellenbosch, South Africa,
1338 2017).
- 1339 378. Vinšálková, T. & Gvoždík, L. Mismatch between temperature preferences and morphology in F1
1340 hybrid newts (*Triturus carnifex* × *T. dobrogicus*). *Journal of Thermal Biology* **32**, 433–439
1341 (2007).
- 1342 379. Vo, P. & Gridi-Papp, M. Low temperature tolerance, cold hardening and acclimation in tadpoles
1343 of the neotropical túngara frog (*Engystomops pustulosus*). *Journal of Thermal Biology* **66**, 49–
1344 55 (2017).
- 1345 380. Voituron, Y., Paaschburg, L., Holmstrup, M., Barré, H. & Ramløv, H. Survival and metabolism
1346 of *Rana arvalis* during freezing. *J Comp Physiol B* **179**, 223–230 (2009).
- 1347 381. von May, R. *et al.* Divergence of thermal physiological traits in terrestrial breeding frogs along a
1348 tropical elevational gradient. *Ecology and Evolution* **7**, 3257–3267 (2017).
- 1349 382. von May, R. *et al.* Thermal physiological traits in tropical lowland amphibians: Vulnerability to
1350 climate warming and cooling. *PLOS ONE* **14**, (2019).
- 1351 383. Wagener, C., Kruger, N. & Measey, J. Progeny of *Xenopus laevis* from altitudinal extremes
1352 display adaptive physiological performance. *Journal of Experimental Biology* **224**, (2021).
- 1353 384. Wang, H. & Wang, L. Thermal adaptation of the common giant toad (*Bufo gargarizans*) at
1354 different earlier developmental stages. *Journal of Agricultural University of Hebei* **31**, 79–83
1355 (2008).
- 1356 385. Wang, L. The effects of constant and variable thermal acclimation on thermal tolerance of the
1357 common giant toad tadpoles (*Bufo gargarizans*). *Acta Ecological Sinica* **34**, 1030–1034 (2014).
- 1358 386. Wang, L.-Z. & Li, X.-C. Effect of Temperature on Incubation and Thermal Tolerance of the
1359 Chinese Forest Frog. *Chinese Journal of Zoology* (2007).
- 1360 387. Wang, L. & Li, X.-C. Effects of constant thermal acclimation on thermal tolerance of the
1361 Chinese forest frog (*Rana chensineniss*). *Acta Hydrobiologica Sinica* **31**, 748–750 (2007).
- 1362 388. Wang, L.-Z., Li, X.-C. & Sun, T. Preferred temperature, avoidance temperature and lethal
1363 temperature of tadpoles of the common giant toad (*Bufo gargarizans*) and the Chinese forest
1364 frog (*Rana chensinensis*). *Chinese Journal of Zoology* **40**, 23–27 (2005).
- 1365 389. Warburg, M. R. On the water economy of israel amphibians: The anurans. *Comparative
1366 Biochemistry and Physiology -- Part A: Physiology* **40**, 911–924 (1971).
- 1367 390. Warburg, M. R. The water economy of israel amphibians: The urodeles *Triturus vittatus*
1368 (Jenyns) and *Salamandra salamandra* (L.). *Comparative Biochemistry and Physiology -- Part A:
1369 Physiology* **40**, 1055–1056, IN11, 1057–1063 (1971).
- 1370 391. Whitehead, P. J., Puckridge, J. T., Leigh, C. M. & Seymour, R. S. Effect of Temperature on
1371 Jump Performance of the Frog *Limnodynastes tasmaniensis*. *Physiological Zoology* **62**, 937–949
1372 (1989).
- 1373 392. Willhite, C. & Cupp, P. V. Daily rhythms of thermal tolerance in *Rana clamitans* (Anura:
1374 Ranidae) tadpoles. *Comparative Biochemistry and Physiology -- Part A: Physiology* **72**, 255–
1375 257 (1982).
- 1376 393. Williams, A. A. & Wygoda, M. L. Dehydration stimulates behavioral hypothermia in the gulf
1377 coast toad, *Bufo valliceps*. *Journal of thermal biology* **18**, 223–227 (1993).
- 1378 394. Winterová, B. & Gvoždík, L. Influence of interspecific competitors on behavioral
1379 thermoregulation: developmental or acute plasticity? *Behav Ecol Sociobiol* **72**, 169 (2018).
- 1380 395. Witters, L. R. & Sievert, L. Feeding causes thermophilic in the woodhouse's toad (*Bufo
1381 woodhousii*). *Journal of Thermal Biology* **26**, 205–208 (2001).
- 1382 396. Wollmuth, L. P., Crawshaw, L. I., Forbes, R. B. & Grahn, D. A. Temperature Selection during
1383 Development in a Montane Anuran Species, *Rana cascadae*. *Physiological Zoology* **60**, 472–480
1384 (1987).
- 1385 397. Wu, C.-S. & Kam, Y.-C. Thermal tolerance and thermoregulation by Taiwanese rhacophorid
1386 tadpoles (*Buergeria japonica*) living in geothermal hot springs and streams. *Herpetologica* **61**,
1387 35–46 (2005).

- 1388 398. Wu, Q.-H. & Hsieh, C.-H. *Thermal Tolerance and Population Genetics of Hynobius Fuca*. 64
1389 (2016).
- 1390 399. Wu, Q.-X. *Study on the Temperature Preference and Temperature Acclimation Ability of*
1391 *Tadpoles in Different Microhabitats*. (2021).
- 1392 400. Xu, X. The effect of temperature on body temperature and thermoregulation in different
1393 geographic populations of *Rana dybowskii*. (Harbin Normal University, Harbin, China, 2017).
- 1394 401. Yandún Vela, M. C. Capacidad de aclimatación en renacuajos de dos especies de anuros:
1395 *Rhinella marina* (Bufonidae) y *Gastrotheca riobambae* (Hemiphractidae) y su vulnerabilidad al
1396 cambio climático. (Pontificia Universidad Católica Del Ecuador, Quito, Ecuador, 2017).
- 1397 402. Young, V. K. H. & Gifford, M. E. Limited capacity for acclimation of thermal physiology in a
1398 salamander, *Desmognathus brimleyorum*. *Journal of Comparative Physiology B: Biochemical,*
1399 *Systemic, and Environmental Physiology* **183**, 409–418 (2013).
- 1400 403. Yu, Z., Dickstein, R., Magee, W. E. & Spotila, J. R. Heat shock response in the salamanders
1401 *Plethodon jordani* and *Plethodon cinereus*. *Journal of Thermal Biology* **23**, 259–265 (1998).
- 1402 404. Zheng, R.-Q. & Liu, C.-T. Giant spiny-frog (*Paa spinosa*) from different populations differ in
1403 thermal preference but not in thermal tolerance. *Aquatic Ecology* **44**, 723–729 (2010).
- 1404 405. Zweifel, R. G. Studies on the Critical Thermal Maxima of Salamanders. *Ecology* **38**, 64–69
1405 (1957).
- 1406 406. Pintanel, P. *et al.* Elevational and local climate variability predicts thermal breadth of mountain
1407 tropical tadpoles. *Ecography* **2022**, e05906 (2022).
- 1408 407. Gutiérrez-Pesquera, L. M. *et al.* Phenology and plasticity can prevent adaptive clines in thermal
1409 tolerance across temperate mountains: The importance of the elevation-time axis. *Ecology and*
1410 *Evolution* **12**, e9349 (2022).
- 1411 408. Pottier, P. *et al.* New horizons for comparative studies and meta-analyses. *Trends in Ecology &*
1412 *Evolution* **39**, 435–445 (2024).
- 1413 409. Jenkins, C. N., Pimm, S. L. & Joppa, L. N. Global patterns of terrestrial vertebrate diversity and
1414 conservation. *Proceedings of the National Academy of Sciences* **110**, E2602–E2610 (2013).
- 1415 410. Asubiaro, T. V. & Onaolapo, S. A comparative study of the coverage of African journals in Web
1416 of Science, Scopus, and CrossRef. *Journal of the Association for Information Science and*
1417 *Technology* **74**, 745–758 (2023).
- 1418 411. Bonnet, X., Shine, R. & Lourdais, O. Taxonomic chauvinism. *Trends in Ecology & Evolution*
1419 **17**, 1–3 (2002).
- 1420 412. Giustini, D. & Boulos, M. N. K. Google Scholar is not enough to be used alone for systematic
1421 reviews. *Online Journal of Public Health Informatics* **5**, e61265 (2013).
- 1422 413. Haddaway, N. R., Collins, A. M., Coughlin, D. & Kirk, S. The Role of Google Scholar in
1423 Evidence Reviews and Its Applicability to Grey Literature Searching. *PLOS ONE* **10**, e0138237
1424 (2015).
- 1425 414. Amano, T. *et al.* Tapping into non-English-language science for the conservation of global
1426 biodiversity. *PLOS Biology* **19**, e3001296 (2021).
- 1427 415. Noble, D. W. A. *et al.* Meta-analytic approaches and effect sizes to account for ‘nuisance
1428 heterogeneity’ in comparative physiology. *Journal of Experimental Biology* **225**, jeb243225
1429 (2022).
- 1430

Supplementary Information

TABLE OF CONTENTS

5	Figure S1	2
6	Figure S2	3
7	Table S1	4
8	Table S2	8
9	Table S3	14
10	Table S4	15
11	Table S5	16



18

19 **Figure S1: Decision tree used to screen titles, abstracts, and keywords.** Additional details can be
20 found in Table S3.

21

22

23

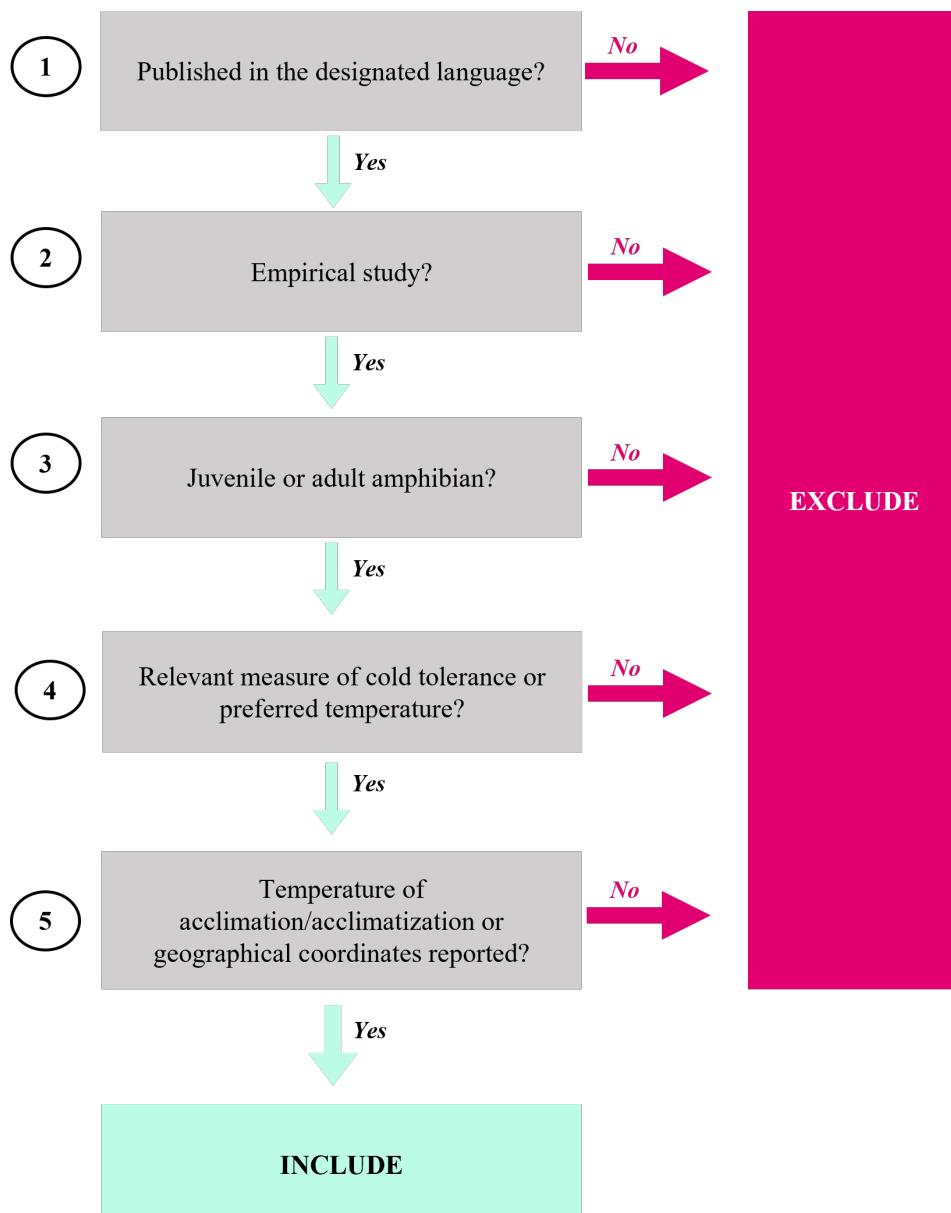
24

25

26

27

28



29

30 **Figure S2: Decision tree used to assess full articles for eligibility.** Additional details can be found
31 in Table S4.

32

33

34

35

36

37

38

39

40

41

42 **Table S1: Search strings used for the different databases.**

Database	Search strings
Scopus	TITLE-ABS-KEY("temperature*" OR "thermal" OR "cold*" OR "cool*") AND TITLE-ABS-KEY("cold tolerance*" OR "tolerance* to cold" OR "thermal min*" OR "CTmin" OR "CT min" OR "chill coma" OR "cold stress tolerance*" OR "tolerance to cold stress" OR "cold stupor" OR "cold resistance" OR "resistance to cold stress" OR "supercooling point" OR "SCP" OR "crystal* temperature*" OR "cold hardiness" OR "freez* tolerance" OR "tolerance to freezing" OR "preferred temperature*" OR "preferred body temperature*" OR "temperature preference*" OR "selected temperature*" OR "selected body temperature*" OR "thermal prefer*" OR "temperature* prefer*" OR "temperature* select*" OR "thermal selection") AND TITLE-ABS-KEY("amphibia*" OR "frog*" OR "toad*" OR "salamand*" OR "newt" OR "newts" OR "tadpole*" OR "metamorph" OR "metamorphs" OR "caecili*" OR "rhinatrema*" OR "ichthyophi*" OR "scolecomorph*" OR "chikil*" OR "herpelidae" OR "typhlonect*" OR "indotyphlid*" OR "dermophi*" OR "siphonop*" OR "caudata" OR "urodela" OR "cryptobranch*" OR "hynobiid*" OR "sirenidae" OR "ambystoma*" OR "dicamptodon*" OR "proteidae" OR "rhyacotriton*" OR "amphium*" OR "plethodon*" OR "anura*" OR "ascaph*" OR "leiopelma*" OR "bombina*" OR "alyt*" OR "rhinophryn*" OR "pipidae" OR "xenopus" OR "scaphiop*" OR "pelodyt*" OR "megophry*" OR "pelobat*" OR "heleophryn*" OR "calyptocephalell*" OR "myobatrach*" OR "rhinoderma*" OR "alsod*" OR "hylod*" OR "batrachyl*" OR "cycloramph*" OR "telmatob*" OR "ceratophry*" OR "hemiphract*" OR "hyla*" OR "hylidae" OR "bufo*" OR "leptodactyl*" OR "odontophryn*" OR "allophryn*" OR "centrolen*" OR "dendrobat*" OR "ceuthomanti*" OR "eleutherodactyl*" OR "brachycephalidae" OR "craugastor*" OR "strabomantidae" OR "pristimantis" OR "nasikabatrach*" OR "soogloss*" OR "microhyl*" OR "arthroleptid*" OR "hyperol*" OR "brevicipitidae" OR "hemisus" OR "odontobatrach*" OR "phrynobatrach*" OR "ptychaden*" OR "conraua" OR "petropedet*" OR "pyxicephal*" OR "micrixalus" OR "nyctibatrach*" OR "ranixalidae" OR "ceratobatrach*" OR "dicroglossidae" OR "rana" OR "ranidae" OR "rhacophor*" OR "mantellidae") AND (EXCLUDE(PUBYEAR , 2022))
Web of Science (core collection)	TS=("temperature*" OR "thermal" OR "cold*" OR "cool*") AND TS=("cold tolerance*" OR "tolerance* to cold" OR "thermal min*" OR "CTmin" OR "CT min" OR "chill coma" OR "cold stress tolerance*" OR "tolerance to cold stress" OR "cold stupor" OR "cold resistance" OR "resistance to cold stress" OR "supercooling point" OR "SCP" OR "crystal* temperature*" OR "cold hardiness" OR "freez* tolerance" OR "tolerance to freezing" OR "preferred temperature*" OR "preferred body temperature*" OR "temperature preference*" OR "selected temperature*" OR "selected body temperature*" OR "thermal prefer*" OR "temperature* prefer*" OR "temperature* select*" OR "thermal selection") AND TS=("amphibia*" OR "frog*" OR "toad*" OR "salamand*" OR "newt" OR "newts" OR "tadpole*" OR "metamorph" OR "metamorphs" OR "caecili*" OR "rhinatrema*" OR "ichthyophi*" OR "scolecomorph*" OR "chikil*" OR "herpelidae" OR "typhlonect*" OR "indotyphlid*" OR "dermophi*" OR "siphonop*" OR "caudata" OR "urodela" OR "cryptobranch*" OR "hynobiid*" OR "sirenidae" OR "ambystoma*" OR "dicamptodon*" OR "proteidae" OR "rhyacotriton*" OR "amphium*" OR "plethodon*" OR "anura*" OR "ascaph*" OR "leiopelma*" OR "bombina*" OR "alyt*" OR "rhinophryn*" OR "pipidae" OR "xenopus" OR "scaphiop*" OR "pelodyt*" OR "megophry*" OR "pelobat*" OR "heleophryn*" OR "calyptocephalell*" OR "myobatrach*" OR "rhinoderma*" OR "alsod*" OR "hylod*" OR "batrachyl*" OR "cycloramph*" OR "telmatob*" OR "ceratophry*" OR "hemiphract*" OR "hyla*" OR "hylidae" OR "bufo*" OR "leptodactyl*" OR "odontophryn*" OR "allophryn*" OR "centrolen*" OR

	"dendrobat*" OR "ceuthomanti*" OR "eleutherodactyl*" OR "brachycephalidae" OR "craugastor*" OR "strabomantidae" OR "pristimantis" OR "nasikabatrach*" OR "soogloss*" OR "microhyl*" OR "arthroleptid*" OR "hyperol*" OR "brevicipitidae" OR "hemisus" OR "odontobatrach*" OR "phrynobatrach*" OR "ptychaden*" OR "conraua" OR "petropedet*" OR "pyxicephal*" OR "micrixalus" OR "nyctibatrach*" OR "ranixalidae" OR "ceratobatrach*" OR "dicroglossidae" OR "rana" OR "ranidae" OR "rhacophor*" OR "mantellidae") NOT PY=(2022)
Lens	("temperature*" OR "thermal" OR "cold*" OR "cool*") AND ("cold tolerance*" OR "tolerance* to cold" OR "thermal min*" OR "CTmin" OR "CT min" OR "chill coma" OR "cold stress tolerance*" OR "tolerance to cold stress" OR "cold stupor" OR "cold resistance" OR "resistance to cold stress" OR "supercooling point" OR "SCP" OR "crystal* temperature*" OR "cold hardiness" OR "freez* tolerance" OR "tolerance to freezing" OR "preferred temperature*" OR "preferred body temperature*" OR "temperature preference*" OR "selected temperature*" OR "selected body temperature*" OR "thermal prefer*" OR "temperature* prefer*" OR "temperature* select*" OR "thermal selection") AND ("amphibia*" OR "frog*" OR "toad*" OR "salamand*" OR "newt" OR "newts" OR "tadpole*" OR "metamorph" OR "metamorphs" OR "caecili*" OR "rhinatrema*" OR "ichthyophi*" OR "scolecomorph*" OR "chikil*" OR "herpelidae" OR "typhonect*" OR "indotyphlid*" OR "dermophi*" OR "siphonop*" OR "caudata" OR "urodela" OR "cryptobranch*" OR "hynobiid*" OR "sirenidae" OR "ambystoma*" OR "dicamptodon*" OR "proteidae" OR "rhyacotriton*" OR "amphium*" OR "plethodon*" OR "anura*" OR "ascaph*" OR "leiopelma*" OR "bombina*" OR "alyt*" OR "rhinophryn*" OR "pipidae" OR "xenopus" OR "scaphiop*" OR "pelodyt*" OR "megophry*" OR "pelobat*" OR "heleophryn*" OR "calyptocephalell*" OR "myobatrach*" OR "rhinoderma*" OR "alsod*" OR "hylod*" OR "batrachyl*" OR "cycloramph*" OR "telmatob*" OR "ceratophry*" OR "hemiphract*" OR "hyla*" OR "hylidae" OR "bufo*" OR "leptodactyl*" OR "odontophrynn*" OR "allophrynn*" OR "centrolen*" OR "dendrobat*" OR "ceuthomanti*" OR "eleutherodactyl*" OR "brachycephalidae" OR "craugastor*" OR "strabomantidae" OR "pristimantis" OR "nasikabatrach*" OR "soogloss*" OR "microhyl*" OR "arthroleptid*" OR "hyperol*" OR "brevicipitidae" OR "hemisus" OR "odontobatrach*" OR "phrynobatrach*" OR "ptychaden*" OR "conraua" OR "petropedet*" OR "pyxicephal*" OR "micrixalus" OR "nyctibatrach*" OR "ranixalidae" OR "ceratobatrach*" OR "dicroglossidae" OR "rana" OR "ranidae" OR "rhacophor*" OR "mantellidae") Year Published = (1900 - 2021) Field of Study = (excl Botany , excl Physics , excl Geology , excl Materials science , excl Context (language use) , excl Geochemistry , excl Internal medicine , excl Mutant , excl Arabidopsis , excl Condensed matter physics , excl Computer science , excl Biophysics , excl Biotechnology , excl Computational biology , excl Horticulture , excl Aquaporin , excl Crop , excl Mineralogy , excl Nanotechnology , excl Arabidopsis thaliana , excl Chemical physics)
Proquest (Dissertation and Theses)	(noft(cold tolerance*) OR noft(CTmin*) OR noft(preferred temperature*) OR noft(selected temperature*)) AND (noft(amphibia*) OR noft(frog*) OR noft(toad*) OR noft(anura*) OR noft(tadpole*) OR noft(salamand*) OR noft(newts))
Google Scholar (French)	("température préférée" OR "température sélectionnée" OR "température choisie" OR "préférences thermiques") AND (amphibiens OR grenouille OR crapaud OR salamandres OR triton OR têtards OR Amphibia OR Caudata OR Anura OR batracien OR anoure) CTmin AND ("amphibiens" OR grenouille OR crapaud OR "salamandres" OR triton OR têtards OR batracien OR anoure) (Tpref OR Tsel) AND ("amphibiens" OR grenouille OR crapaud OR "salamandres" OR triton OR têtards OR batracien OR anoure)

Google Scholar (Japanese)	(好適温度 OR 選択温度 OR 温度嗜好性) AND (両生類 OR カエル OR ヒキガエル OR サンショウウオ OR イモリ OR おたまじゃくし OR “Amphibia” OR “Caudata” OR “Anura”) CTmin AND (両生類 OR カエル OR ヒキガエル OR サンショウウオ OR イモリ OR オタマジャクシ) (Tpref OR Tsel) AND (両生類 OR カエル OR ヒキガエル OR サンショウウオ OR イモリ OR オタマジャクシ)
Google Scholar (Portuguese)	(“temperatura preferida” OR “temperatura selecionada” OR “preferência termal”) AND (anfíbio OR “rã” OR sapos OR salamandra OR tritão OR girino OR Amphibia OR Caudata OR Anura OR anuros) CTmin AND (anfíbio OR “rã” OR sapos OR salamandra OR tritão OR girino) (Tpref OR Tsel) AND (anfíbio OR “rã” OR sapos OR salamandra OR tritão OR girino)
Google Scholar (simplified Chinese)	(合适温度 OR 选温度 OR 耐热程度) AND (两栖动物 OR 青蛙 OR 蛤蟆 OR 蟆螈 OR 蝌蚪 OR 小鲵 OR 大鲵 OR Amphibia OR Caudata OR Anura) CTmin AND (两栖动物 OR 青蛙 OR 蛤蟆 OR 蟆螈 OR 蝌蚪 OR 小鲵 OR 大鲵) (Tpref OR Tsel) AND (两栖动物 OR 青蛙 OR 蛤蟆 OR 蟆螈 OR 蝌蚪 OR 小鲵 OR 大鲵)
Google Scholar (traditional Chinese)	(偏好溫度 OR 溫度選擇OR 熱偏好) AND (兩棲類 OR 蛙青蛙 OR 蟾蜍蛤蟆 OR 蟆螈 OR 蝌蚪 OR 小鯢山椒魚 OR 鯢大鯢娃娃魚OR Amphibia OR Caudata OR Anura) CTmin AND (兩棲類 OR 蛙青蛙 OR 蟾蜍蛤蟆 OR 蟆螈 OR 蝌蚪 OR 小鯢山椒魚 OR 鯢大鯢娃娃魚) (Tpref OR Tsel) AND (兩棲類 OR 蛙青蛙 OR 蟾蜍蛤蟆 OR 蟆螈 OR 蝌蚪 OR 小鯢山椒魚 OR 鯢大鯢娃娃魚)
Google Scholar (Spanish)	(“temperatura preferida” OR “temperatura seleccionada” OR “preferencias térmicas”) AND (anfibio OR rana OR sapo OR salamandra OR triton OR renacuajo OR Amphibia OR Caudata OR Anura OR anuros) CTmin AND (anfibio OR rana OR sapo OR salamandra OR triton OR renacuajo or Anuros) (Tpref OR Tsel) AND (anfibio OR rana OR sapo OR salamandra OR triton OR renacuajo or Anuros)

45 **Table S2: Metadata.**

Data	Description
unique_ID	Unique identifier for each row in the data.
name	Name of the researcher who performed the data extraction.
ref	Abbreviated reference for the study.
title	Title of the paper or thesis.
pub_year	Publication year of the paper or thesis.
thesis_chapter	If the study is a thesis, the chapter the data is taken from (e.g., 2).
chapter_title	The title of the thesis chapter the data is taken from.
peer-reviewed	Whether the study was peer-reviewed or not (i.e., thesis). Factor with two levels: “peer-reviewed”, “not_peer-reviewed”.
doi	DOI of the paper.
language	Language of the paper (main text). Factor with seven levels: “English”, “traditional Chinese”, “simplified Chinese”, “French”, “Japanese”, “Portuguese”, “Spanish”.
population_ID	Unique identifier for each population. Note that populations were considered individuals of the same species taken from different geographical locations. For studies without geographical coordinates, populations were assigned based on descriptions made by the authors (e.g., “Northern population” vs. “Southern population”).
cohort_ID	Unique identifier for each cohort. By “cohort”, we refer to independent groups of animals. In some cases, traits were measured multiple times on the same cohort of animals (e.g., using different endpoints, or at different life stages). As such, the same cohort_ID was assigned to repeated measures. Note that cohort_ID was assigned at the trait-level; as it was not always possible to assign whether multiple traits (e.g., CTmin and CTmax) were measured with the same, or independent groups of animals.
notes_ID	General notes related to population_ID and cohort_ID.
order	Species order, according to Jetz and Pyron (2018).
family	Species family, according to Jetz and Pyron (2018).
species	Species name, according to Jetz and Pyron (2018).
strain	The strain, variety, subspecies, or morph of the species, as reported in the study.
IUCN_status	International Union for the Conservation of Nature (IUCN) threat status. Factor with 7 levels: “DD”, “LC”, “NT”, “VU”, “EN”, “CR” and “EX”, for “data-deficient”, “least-concern”, “near threatened”,

“vulnerable”, “endangered”, “critically endangered” and “extinct”, respectively.

origin	Origin of studied animals. Factor with four levels: recently collected from the wild (i.e., “wild”), eggs laid in the laboratory (i.e., “lab”), animals provided from a supplier (i.e., “supplier”) or “unclear”. For studies collecting eggs from the wild and testing the same generation of animals, animals were considered as “wild”.
n_generations_lab	Number of generations spent in the laboratory, if reported in the study.
latitude	Latitude from which animals were collected (decimal degrees). Latitudes presented in degrees/minutes/seconds were converted to decimal degrees. When geographical coordinates were not presented, the coordinates were estimated using Google Maps.
longitude	Longitude from which animals were collected (decimal degrees). Longitudes presented in degrees/minutes/seconds were converted to decimal degrees. When geographical coordinates were not presented, the coordinates were estimated using Google Maps.
elevation	Elevation from which animals were collected (meters above sea level), as reported in the study. When not reported, elevation was estimated using latitude and longitude and freemaptools.com.
date_sampling	Date at which the animals were sampled (format YEAR/MONTH/DAY, e.g., “2020/07/26”).
month_sampling	Month from which the animals were collected.
year_sampling	Year from which the animals were collected.
start_range_sampling_dates	The beginning of the range of dates over which animals were collected. Indicated are both the month and the year of collection (e.g., “January_2015”).
end_range_sampling_dates	The end of the range of dates over which animals were collected. Indicated are both the month and the year of collection (e.g., “September_2015”).
notes_sampling	General notes regarding the sampling of the animals.
ambient_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean ambient temperature (°C) in the month of collection, if reported in the study. If animals were collected over a range of months, the mean temperature across this sampling period was reported.
substrate_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean temperature of the substrate (°C) in the month of capture. If animals were collected over a range of months, the mean temperature across this sampling period was reported.

water_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean water temperature (°C) in the month of collection. If animals were collected over a range of months, the mean temperature across this sampling period was reported.
field_body_temp	For animals recently sampled from the wild (eggs not laid in the laboratory), the mean body temperature (°C) measured in the field when animals were collected. If animals were collected over a range of months, the mean temperature across this sampling period was reported.
notes_env_temp	General notes regarding the sampling of animals in the field.
acclimated	Whether the animals were maintained in the laboratory for >12h or tested shortly after collection. Factor with two levels: “acclimated” or “field-fresh”.
incubation_temp	For animals born in the laboratory, the mean temperature (°C) at which the embryos were incubated.
sd_incubation_temp	Variability (standard deviation) in incubation_temp (°C).
life_stage_acclimated	For acclimated animals, the life stage acclimated prior to the upper thermal limit assessment. Factor with five levels: “embryos_and_larvae”, “larvae”, “juveniles”, “metamorphs” or “adults”. Larval stages of salamanders and tadpoles were referred to as “larvae”. Animals between Gosner stages 42 and 45 were considered “metamorphs”, while those between Gosner stage 45 and sexual maturity were considered “juveniles”.
gosner_acclimated	For acclimated animals, the Gosner stage when the acclimation started, if reported in the study.
acclimation_temp	For acclimated animals, the mean temperature of acclimation (°C). Note that “acclimation” refers to a prolonged (>12h) exposure to a new temperature. Therefore, cold/heat shocks or housing conditions just prior to assessing thermal tolerance or preference (e.g., 2 hours at 25°C) were not considered as “acclimation” conditions. If animals were exposed to multiple acclimation conditions (e.g., 15°C for 1 month, and then re-acclimated to 25°C for 7 days), we took the latest acclimation condition as the “acclimation_temp”.
sd_acclimation_temp	Variability (standard deviation) in acclimation_temp (°C).
acclimation_time	The duration of acclimation (days).
notes_acclimation	General notes regarding the laboratory acclimation of animals.
life_stage_tested	The life stage tested for thermal tolerance or preference. Factor with four levels: “larvae”, “metamorphs”, “juveniles” or “adults”. Larval stages of salamanders and tadpoles were referred to as “larvae”. Animals between Gosner stages 42 and 45 were considered

	"metamorphs", while those between Gosner stage 45 and sexual maturity were considered "juveniles".
gosner_tested	Gosner stage when the animals were assessed for thermal tolerance or preference.
SVL	Mean snout-vent length of the animals (mm) when assessed for thermal tolerance or preference, if reported in the study. Note that SVL data was often taken from Rohr et al. (2018).
body_mass	Mean body mass of the animals (g) when assessed for thermal tolerance or preference.
age_tested	The age (days-post-hatching) at which the animals were tested for thermal tolerance or preference.
sex	The sex of the animals. Factor with four levels: "male", "female", "mixed", "unknown". The "mixed" category was used when authors clearly stipulate that they mixed males and females.
metric	The metric used to assess thermal tolerance (CTmax, LT50_hot, CTmin, LT50_cold) or preference (Tpref). Factor with two levels: "CTmax", "LT50_hot", "CTmin", "LT50_cold", "Tpref".
endpoint	The endpoint that was used for assessing thermal tolerance (loss or righting response, loss of equilibrium, onset of spasms, no response to prodding, supercooling point, death). Factor with seven levels: "LRR", "LOE", "OS", "prodding", "SCP", "death", "other". If "other", details are reported in "notes_test" (see below).
medium_test_temp	Whether the temperature measured during the test was the ambient, the water, the substrate, or the body temperature. Factor with three levels: "ambient", "substrate", "water", "body".
start_temp	If the metric was CTmax, the starting temperature used in the upper thermal limit assay (°C).
ramping	If the metric was CTmax, the ramping (heating) rate applied to the animals (°C/min).
set_time	If the metric was LT50, the time the animals spent at the test temperature (the time after which the animals the survival was assessed, in hours). If the authors report e.g., 96h-LT50, then set_time would be 96.
n_test_temp	If the metric was LT50, the number of temperatures tested to assess upper thermal limits. E.g., if authors measured survival at 36, 38, 39, and 41°C, n_test_temp = 4.
n_replicates_per_temp	If the metric was LT50, the number of replicates used at each test temperatures. E.g., if authors used 5 test temperatures and measured the survival of three independent cohorts of animals at each test temperature, then n_replicates_per_temp = 3.

n_animals_per_replicate	If the metric was LT50, the number of animals in each replicate.
duration_measurement	If the metric was Tpref, the duration of the assay to measure thermal preference (hours).
rate_measurement	If the metric was Tpref, the rate at which body temperature was measured (measurements/hour).
gradient_type	If the metric was Tpref, the type of thermal gradient used. Factor with two levels: “linear”, “shuttlebox”.
gradient_low_temp	Lowest temperature in the thermal gradient (°C).
gradient_high_temp	Highest temperature in the thermal gradient (°C).
notes_test	General notes regarding the thermal tolerance or preference assays.
humidity	Humidity at which animals were acclimated or tested (% relative humidity). If the humidity during the acclimation and the test were different, priority was given to the conditions of the test.
oxygen	Oxygen at which animals were acclimated or tested (mg.L ⁻¹ dissolved oxygen). If the oxygen concentration during the acclimation and the test were different, priority was given to the conditions of the test.
salinity	Salinity at which animals were acclimated or tested (parts per thousands). If the salinity during the acclimation and the test were different, priority was given to the conditions of the test.
pH	pH at which animals were acclimated. If the pH during the acclimation and the test were different, priority was given to the conditions of the test.
photoperiod	Photoperiod at which animals were acclimated (number of hours of light per day).
chemical	If any, which chemical (e.g., pollutant, toxin) was added to the animals’ environment. If animals were in a control group (i.e., only supplemented with a solvent), “control” was indicated.
hormone	If any, which hormone (e.g., corticosterone, thyroid hormone) was added to the animals’ environment. If animals were in a control group (i.e., only supplemented with a solvent), “control” was indicated.
concentration_chemical_hormone	If any, the concentration of the hormones or chemicals used. If animals were in a control group, “0” was indicated.
unit_chemical_hormone	The unit used to quantify the chemical or hormonal concentration administered (e.g., g/L, ng/g of sediment).
infected	Whether the animals were infected with a pathogen. Indicate “infected” if the animals were infected with a pathogen. Otherwise, leave the field blank.
pathogen	If the animals were infected with a pathogen, the name of the pathogen (e.g., <i>Batrachochytrium dendrobatidis</i>).

notes_supplements	General notes regarding the addition of chemicals, hormones, or pathogens.
data_source	Where the upper thermal limit data is reported (main text, table, figure, published data).
data_url	If the data was published in a repository, the url link to the repository containing the data.
flag	Whether the study has procedural concerns (with details).
mean_trait	Mean thermal tolerance or preference (°C).
error_trait	Standard deviation or standard error of mean_trait (see error_type)
n_trait	Sample size of mean_trait. When the metric was LT50, the sample size was taken as the number of test temperatures (“n_test_temp”) * the number of replicates per test temperature (n_replicates_per_temp).
error_type	Whether the error is presented as standard deviations (i.e., “sd”) or standard errors (i.e., “se”).
notes_trait	General notes about thermal tolerance or preference estimates

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63 **TABLE S3: Inclusion criteria used to screen abstracts, titles and keywords.** Numbers match those
64 used in in Figure S1 (decision tree).

Description	
1	Studies not published in French, Japanese, Portuguese, simplified Chinese, traditional Chinese or Spanish were excluded.
2	We only included empirical studies presenting original data. Therefore, we excluded reviews, syntheses, simulations, theoretical studies, and conference abstracts, unless supplemented with original data.
3	“Amphibians” refer to frogs, toads, salamanders, newts, and caecilians. We only included studies on whole organisms.
Desired measures of cold tolerance included the:	
i)	critical thermal minimum (CTmin), where animals are subject to incremental decreases in temperature until an endpoint (e.g. loss of righting response, onset of spasms, supercooling point, crystallisation) is reached;
ii)	the temperature lethal for 50% of the animals (LT50; sometimes referred to as the “incipient lethal temperature”), where survival is recorded after animals are abruptly transferred to a set of cold temperatures for a given period of time (e.g. 24 hours) and LT50 is interpolated from the survival curve; and
iii)	the death time or chill coma time where animals are abruptly transferred to cold temperatures and the time needed for animals to reach an endpoint (e.g. immobilisation, death) is recorded as the response. With the latter measure, the thermal tolerance limit can be inferred from the relationship between chill coma time and the temperature of the knockdown assay. Therefore, cold knockdown times must have been measured at >2 temperatures (e.g. chill coma times at 10, 15, and 17°C).
4	We exclude alternative measures of cold tolerance which cannot be converted to the temperature scale (e.g. chill coma recovery time) or CTmin extrapolated from physiological performance curves (e.g. critical temperature for ATPase activity).
Desired measures of preferred temperature were:	
i)	where animals are placed in a temperature gradient and the body temperature of the animals is measured at regular intervals, or inferred from photography. The mode or median body temperatures animal select is usually defined as the preferred (or selected) body temperature.
ii)	where animals are placed in an experimental set up with levers that trigger the warming or cooling of the surface or experimental chamber (shuttlebox). Similarly to above, the body temperatures of animals is tracked, and the mode or median body temperature animals experienced is usually defined as the preferred (or selected) body temperature.
5	We focused our search on juveniles (i.e., tadpole, metamorph, froglet) or adults. Hence, we excluded studies only measuring the cold tolerance of embryos.

65

66

67

68

69

70 **TABLE S4: Inclusion criteria used to assess full articles for eligibility.** Numbers match those used
71 in Figure S2 (decision tree).

Description	
1	Studies not published in French, Japanese, Portuguese, simplified Chinese, traditional Chinese or Spanish were excluded.
2	We only included empirical studies presenting original data. Therefore, we excluded reviews, syntheses, simulations, theoretical studies, and conference abstracts, unless supplemented with original data.
3	"Amphibians" refer to frogs, toads, salamanders, newts and caecilians. We only included studies on juveniles (i.e. tadpole, metamorph, froglet) or adults. Hence, we exclude studies only measuring the cold tolerance of embryos.
	Desired measures of cold tolerance included the:
4	iv) critical thermal minimum (CTmin), where animals are subject to incremental decreases in temperature until an endpoint (e.g. loss of righting response, onset of spasms, supercooling point, crystallisation) is reached; v) the temperature lethal for 50% of the animals (LT50; sometimes referred to as the "incipient lethal temperature"), where survival is recorded after animals are abruptly transferred to a set of cold temperatures for a given period of time (e.g. 24 hours) and LT50 is interpolated from the survival curve; and vi) the death time or chill coma time where animals are abruptly transferred to cold temperatures and the time needed for animals to reach an endpoint (e.g. immobilisation, death) is recorded as the response. With the latter measure, the thermal tolerance limit can be inferred from the relationship between chill coma time and the temperature of the knockdown assay. Therefore, cold knockdown times must have been measured at >2 temperatures (e.g. chill coma times at 10, 15, and 17°C).
	We exclude alternative measures of cold tolerance which cannot be converted to the temperature scale (e.g. chill coma recovery time) or CTmin extrapolated from physiological performance curves (e.g. critical temperature for ATPase activity).
	Desired measures of preferred temperature were:
	iii) where animals are placed in a temperature gradient and the body temperature of the animals is measured at regular intervals, or inferred from photography. The mode or median body temperatures animal select is usually defined as the preferred (or selected) body temperature. iv) where animals are placed in an experimental set up with levers that trigger the warming or cooling of the surface or experimental chamber (shuttlebox). Similarly to above, the body temperatures of animals is tracked, and the mode or median body temperature animals experienced is usually defined as the preferred (or selected) body temperature.
5	To be included, the study must have reported the temperature at which animals were maintained in the laboratory (i.e. temperature of acclimation), the temperature of the environment from which animals were captured (i.e. temperature of acclimatization), or the geographical coordinates and dates of capture.

72

73

74

75
76
77
78
79
80
81
82

83 **TABLE S5: Summary of procedural concerns found in some studies.** Note that estimates having
84 procedural concerns were excluded during the data curation (see main text).

Procedural concerns	Number of estimates concerned
Data from a single individual	70
Uncommon or inconsistent methodology	55
Unclear/uncommon acclimation conditions	53
Animals were starved prior to testing	34
Animals underwent surgery or amputation	16
Highly uncertain estimates	14
Animals were dehydrated prior to testing	14
Animals were exposed to hypoxic or hypercapnic conditions	13
Animals were exposed to high levels to UV radiation	5
Animals were perfused with pH solution	3
Statistical dispersion and sample sizes not reported	3
Animals were exposed to predators	2

85
86
87
88
89