1		
2		Spectral biology across scales in changing environments
3		
4		
5		
6		
7		
8		Jeannine Cavender-Bares ^{1,2*}
9		Jose Eduardo Meireles ³
10		Jesús Pinto-Ledezma ¹
11		Peter B. Reich ^{4,5}
12		Meredith C. Schuman ⁶
13		Philip A. Townsend ⁷
14		Amy Trowbridge ⁷
15		
16	*A	uthor for correspondence: Email: jcavender@fas.harvard.edu
17	1.	Department of Ecology, Evolution and Behavior, University of Minnesota
18	2.	Department of Organismic and Evolutionary Biology, Harvard University
19	3.	Department of Integrative Biology, University of Maine
20	4.	Department of Forest Resources, University of Minnesota
21	5.	Institute for Global Change Biology and School for the Environment and Sustainability,
22		University of Michigan
23	6.	Department of Geography and Department of Chemistry, University of Zurich
24	7.	Department of Forest and Wildlife Ecology, University of Wisconsin-Madison
25		

- 1 Abstract
- 2

3 Understanding ecosystem processes on our rapidly changing planet requires integration across spatial, 4 temporal and biological scales. We propose that spectral biology, using tools that enable near- to far-5 range sensing by capturing the interaction of energy with matter across domains of the electromagnetic 6 spectrum, will increasingly enable ecological insights across scales from cells to continents. Here, we 7 focus on advances using spectroscopy in the visible to short-wave infrared, chlorophyll fluorescence-8 detecting systems, and optical laser scanning (light detection and ranging, LiDAR) to introduce the topic 9 and special feature. Remote sensing using these tools, in conjunction with *in situ* measurements, can 10 powerfully capture ecological and evolutionary processes in changing environments. These tools are 11 amenable to capturing variation in life processes across biological scales that span physiological, 12 evolutionary and macroecological hierarchies. We point out key areas of spectral biology with high 13 potential to advance understanding and monitoring of ecological processes across scales—particularly at 14 large spatial extents—in the face of rapid global change. These include: the detection of plant and 15 ecosystem composition, diversity, structure and function as well as their relationships; detection of the 16 causes and consequences of environmental stress, including disease and drought, for ecosystems; and 17 detection of change through time in ecosystems over large spatial extents to discern variation in and mechanisms underlying their resistance, recovery and resilience in the face of disturbance. We discuss 18 19 opportunities for spectral biology to discover previously unseen variation and novel processes and to 20 prepare the field of ecology for novel computational tools on the horizon with vast new capabilities for 21 monitoring the ecology of our changing planet.

1 Why spectral biology has high potential to advance knowledge of ecological processes across scales 2 In this era of rapid global change, understanding how biological variation at one scale influences 3 emergent properties at other scales—including the functioning of organisms, ecosystems, and the 4 biosphere—is important to developing an integrative understanding that will allow us to actively support 5 a sustainable future for humanity. The approach we term 'spectral biology' encompasses integrative 6 measures of biological systems that harness the interaction of electromagnetic radiation in ways that are 7 scalable and support standardized, repeated measurements (Fig. 1). These spatio-temporally scalable tools 8 provide a means to measure biological variation and related emergent properties across levels of 9 organization. For example, the reflection of electromagnetic radiation by plants is influenced by their 10 phenotypic, chemical, structural, and functional properties, thereby providing a means to measure 11 biological variation and related emergent properties across levels of organization. Spectral information 12 thus provides a consistent data type to integrate aspects of the evolutionary and physiological variation 13 within and among plant species, the interactions of species within communities, and their consequences 14 for ecosystems responses to global change. The rapidly expanding use of the tools of spectral biology in 15 ecology (Fig. 2) provides the impetus to synthesize the capabilities and opportunities within the field and 16 consider the path forward. 17 The goal of this special feature is to explore how spectral biology enables integration across

17 The goal of this special feature is to explore how spectral biology enables integration across 18 spatial, temporal and biological scales to reveal novel insights in plant ecology, ecosystem dynamics and 19 global change biology. Focal areas represented in articles that are part of this special feature range from 20 quantitative genetics, phylogenetic ecology, ecophysiology, forest dynamics, global change biology, and 21 phenological variation in ecological systems, to biodiversity-ecosystem function relationships.

22

23 What is spectral biology?

We define spectral biology as the spectrally resolved observation of the interaction of electromagnetic
radiation with biological systems. We emphasize these interactions in the solar domain, specifically in the
visible-to-shortwave infrared (VSWIR, 400-2500 nm) but also include ultra-violet (UV, 100-400 nm) as

1 well as thermal emissions $(3-14 \text{ }\mu\text{m})$ and active and passive microwave (0.1-1m) domains that enable the 2 discernment of biological properties. We focus on advances made in this new discipline through studying 3 plant life using reflectance, transmittance, and absorbance spectroscopy, as well as chlorophyll 4 fluorescence emission, including solar-induced fluorescence (SIF), which is coupled to photosynthetic 5 function (Fig. 3). We also include thermal emission, which provides observations of temperature and 6 water content/flux; microwave emission, which can be used to determine soil moisture; and LiDAR (light 7 detection and ranging)-an active sensing system, which provides detailed three-dimensional structural 8 information through the measurement of distance by pulsed lasers. These tools of spectral biology can 9 help decipher the causes and consequences of biological variation across scales. Spectral variation in 10 reflected, absorbed, transmitted or re-emitted electromagnetic radiation results from the variation of 11 chemical, anatomical, morphological, and architectural plant traits, as well as variations in viewing 12 geometry due to sun position, topography or sensor position. The biological variation may originate due 13 to selection, evolutionary history, community composition, diversity, plasticity and their varied responses 14 to environmental drivers.

15 Spectral biology encompasses a continuum of close- to far-range measurements, which are often 16 described as contact (e.g., using a leaf contact probe attached to a spectrometer), proximal (such as a 17 handheld measurement above a canopy, from a tower or low-flying drones [< 100 m]), or remote (higher 18 altitude aerial to space-based or orbital). Remote and proximal sensing of spectral variation most often 19 involve measuring reflectance. Full-range surface reflectance in the solar domain is calculated as a 20 fraction of incoming (atmosphere-penetrating) solar radiation across the electromagnetic spectrum, which 21 is highest in the visible to short wave infrared (400 - 2500 nm); or as a fraction of artificial, standardized 22 light sources providing a similar source spectrum for irradiating targets at close range. Surface reflectance 23 in this range—e.g., the surface of whole ecosystems or a plant leaf, depending on the scale of 24 measurement—is detected at each wavelength (or band of multiple wavelengths) by a sensor that can be 25 placed on a range of platforms (Fig. 1). Spectral signatures can distinguish among different kinds of 26 molecules in plants (Jacquemoud and Ustin 2019), are sensitive to differences in plant traits (Fig. 3c), and

reveal variation across a range of scales from leaves of individual plants, within and among species across
the tree of life, and within and among plant communities, ecosystems, and landscapes across the global
biosphere (Gamon et al. 2020). Beyond full-range, spectrally highly resolved (often termed
'hyperspectral') reflectance data, we include in the set of tools multispectral sensors which capture
reflectance in many fewer bands that may each span a range of wavelengths of interest; fluorescence
sensors; associated technologies such as LiDAR; and new applications that emerge from the interpretation
of these signals in biological realms.

8 Chlorophyll fluorescence has long been used at the leaf level to assess photosynthetic light use 9 efficiency (Genty et al. 1989, Schreiber et al. 1994) and to scale from leaves to ecosystems (Gamon and 10 Qiu 1999, Cavender-Bares and Bazzaz 2004). Chlorophyll fluorescence associated with photosynthesis 11 can be captured proximally from UAVs, or remotely from aircraft and from space through measurements 12 of solar-induced fluorescence emission in specific wavelengths that overlap with "dark features" of the 13 Earth's incoming or reflected light spectrum (Joiner et al. 2013) (Fig. 3b). Within these wavelengths, 14 sunlight is partially absorbed by oxygen (O₂-A or O₂-B bands, centered at 760 and 687 nm, respectively). 15 Dark features can also include wavelengths where gases in the Sun's atmosphere absorb outgoing 16 radiation (Fraunhofer lines). Such absorption features where solar radiation is diminished are critical 17 because they allow distinction between the relatively weak signal emitted by plants as fluorescence and 18 the much stronger signal of solar radiation and reflectance (e.g., Köhler et al. 2018, Moya and Cerovic 19 2004, Sun et al. 2018, Mohammed et al. 2019). Like spectral data, solar-induced chlorophyll fluorescence 20 detection can involve a range of platforms from satellites (Köhler et al. 2018), aircraft (Frankenberg et al. 21 2018, Porcar-Castell et al. 2021), towers or movable carts (Kebabian et al. 1999, Flexas et al. 2000) to 22 leaf-level measurements (Magney et al. 2017) that vary in the specific detection approach and sensor 23 used.

LiDAR (Light Detection And Ranging) instruments uses pulsed laser light and detect the return time of pulses. This provides distance information and is used to generate three-dimensional point clouds with the level of detail depending on point density. The resulting three-dimensional models reveal

information that can be interpreted ecologically in terms of form and structure (e.g., Davies and Asner
2014). Long used in archeology and in the automotive industry, sensors can be hand-held, placed on
uncrewed aerial vehicles (UAVs) or on aircraft as well as on platforms orbiting the Earth (GEDI,
Dubayah et al. 2020). Many LiDAR instruments are also able to measure echo intensity, providing
additional information that can be used to classify targets (Wagner et al. 2006).

6 By harnessing these tools, spectral biology provides powerful and integrated means to capture 7 biological variation-or biodiversity-from leaves to landscapes and to determine the causal factors that 8 give rise to that variation. It is particularly powerful when spectral and remotely sensed information and 9 tools are coupled with deep biological knowledge across subdisciplines that span scales. The spectral 10 biology toolkit complements other tools, such as gas exchange systems, flux towers, and isotopic 11 measurements that can provide more precise, or different types, of information at specific biological 12 scales. The toolkit may enable biologists across disciplines to consider a greater breadth of relevant scales 13 when designing research to study focal processes, loosening constraints to focus on a specific scale 14 imposed by familiar tools and expertise.

15

16 What is the potential of spectral biology to advance ecological research?

17 Advancing our understanding of Earth's biodiversity and its response to global environmental 18 change at scales from molecules to ecosystems, revealing mechanisms that can be targeted for 19 management, is critical for societal capacity to adapt to, and mitigate, changes in biodiversity (Cavender-20 Bares et al. 2022a). Here we define the term 'biodiversity' not simply in its most common usage as 21 species diversity at a community scale but to encompass the diversity of life on Earth including variation 22 in functional and evolutionary components within and among biological scales, ranging from cells to 23 organs, to individuals to ecosystems and regions. Decades of research on species diversity at the 24 community scale and its relationship to ecosystem functions have revealed its importance for how 25 ecosystems cycle elements (Weisser et al. 2017, Schuldt et al. 2023), produce biomass (Isbell et al. 2018, 26 Huang et al. 2018), and respond to environmental change (Reich et al. 2001, Loreau and de Mazancourt

1 2013). These functions are critical to providing ecosystem services that contribute to human well-being 2 (Mori et al. 2021, O'Connor et al. 2021). Integration across biological subdisciplines is required to 3 address fundamental questions that remain poorly understood, including how biodiversity varies across 4 scales-from genes and molecules within cells and tissues, to ecosystem variation. Our capacity to 5 understand and monitor changes in these biological processes at different scales is critical to sustainably 6 managing Earth's life support systems (Gonzalez et al. 2023). However, the scientific advances required 7 to tackle this set of problems have been hindered by the fragmentation of biology into specialized sub-8 disciplines that are conceptually and methodologically divergent and do not meaningfully connect these 9 vastly different scales. The lack of a common data type to discern processes across scales has contributed 10 to these constraints.

11

12 Critical scales in biology

13 We focus on three kinds of biological hierarchies that form the basis of biological integration and scaling: 14 physiological, evolutionary, and macroecological (Fig. 4). The physiological hierarchy considers the 15 functional or metabolic units within a plant from genes and metabolites (molecular products of 16 metabolism) to organelles, cells, leaves, and other organs, to the whole plant. The evolutionary hierarchy 17 encompasses the nested and fractal organization of the tree of life from individuals nested within 18 populations, species and clades, or lineages of increasing size. Finally, the macroecological hierarchy 19 refers to the ecological processes at nested spatial and temporal scales that drive the distribution and 20 diversity of life—from density- and frequency-dependent neighborhood interactions, to sorting of species 21 across environmental gradients, and the dispersal, migration, and long-term biogeographic processes that 22 form the variation in ecosystems within and across biomes, and drive their function as well.

23

24 How spectra help integrate across scales to address complex ecological problems

25 As biological and ecological subdisciplines have become increasingly specialized, addressing complex

26 questions that span biological scales requires bridging subdisciplines. For example, resting within a single

1 subdiscipline, it is difficult to understand how climate change and landscape fragmentation influence the 2 genetic variation within species; or the complex ecological processes by which community composition 3 of ecosystems across biomes at broad geographic scales impacts changes in ecosystem function and 4 stability. Successful integration requires both conceptual and technical advances. Conceptually, we seek 5 to understand biological processes using a common data type across scales, including across evolutionary 6 hierarchies that capture the nature of phenotypic and functional variation within and among populations, 7 species and major lineages (Fig. 5a) and across temporal and spatial scales (Fig. 5b) to help elucidate how 8 processes at one scale affect processes at other scales and their combined influences on observed patterns, 9 properties, and dynamics.

10 The technological dimensions involved in generating common data types create a path forward 11 for the practical aspects of integrating across scales to address complex problems. An important point is 12 that monitoring methods should align with biological scales. For example, contact probes are appropriate 13 at the leaf scale, UAVs and low-flying piloted aircraft are often most appropriate at the community scale, 14 and satellites capture phenomena at landscape to global scales (Fig. 5b). Analysis and interpretation of 15 spectral measurements differ significantly based on measurement scale, due to the range of confounding 16 factors expressed at different scales. These factors may include atmospheric interference for high-altitude 17 and orbital imaging, or the influence of detector distance from the object of measurement (e.g., leaves or 18 canopies), or variation unrelated to biological factors due to source-sensor-object geometry. These issues 19 can be addressed through various data processing approaches (e.g., Queally et al. 2022). On the 20 conceptual side, advances emerge when we bridge subdisciplines across scales, fusing expertise from 21 different realms. For example, knowledge of genetic variation within species and how different genotypes 22 respond physiologically to environmental change emerges from the realms of quantitative genetics and 23 ecophysiology. These differences can be connected with typical functional differences among co-24 occurring species that influence their interactions, and community dynamics that influence ecosystem 25 processes spanning community and ecosystem ecology. This integration may include linkages between 26 above- and belowground processes that drive long-term responses of nutrient cycling to community

1 change, integrating soil and microbial science (Cline et al. 2018, Cavender-Bares et al. 2022b). In another 2 important example, detecting changes in biodiversity in plants at the leaf level is advanced by our 3 understanding that spectra are coupled to genetic and phylogenetic information (Cavender-Bares et al. 4 2016, Meireles et al. 2020, Stasinski et al. 2021, Griffith et al. 2023, Li et al. 2023). Recent evidence finds 5 similar relationships at canopy scales (Czyż et al. 2020, 2023, Seeley et al. 2023, Griffith et al. 2023). The 6 physiological processes and stress responses that spectra reveal also appear to scale from leaf to canopy 7 levels (Sapes et al. 2024). These findings are important for understanding physiological processes that 8 underlie disease symptoms and can help monitor and map diseases to aid management (Sapes et al. 2022, 9 Guzmán et al. 2023). Spectral biology thus facilitates scaling from individual leaves to their aggregated 10 properties at the scale of landscapes and global observations, because it provides a common measure for 11 investigating how foliar tissue and photosynthetic processes interact with the environment, biological 12 phenomena that can be examined from microscopic to ecosystem scales. Spectral information can also be 13 combined with other measures, such as gas flux rates across scales, to gain insight into how processes at 14 one scale result in emergent properties at others. All of these advances in integration emerge from 15 conceptual and technological efforts.

16

17 Avenues for major advances in spectral biology

18 We address five dimensions of ecology in which spectral information will help to bridge scales and 19 subdisciplines to address complex ecological problems that affect humanity: 1) detecting the composition, 20 structure, function, and diversity of biological components, 2) measuring the consequences of 21 composition, structure, function and diversity for functions of plants and ecosystems, and 3) measuring 22 how those consequences will vary with global environmental change, enabling us 4) to characterize and 23 quantify how those consequences play out at differing temporal and spatial scales, including detecting the 24 resistance and recovery of vegetation in response to disturbance given the ecosystem composition and 25 diversity; and 5) discovery of novel biological phenomena through detection of emergent processes and 26 patterns enabled by cross-scale observation. These dimensions build on each other (Fig. 6). The

1 characterization of composition and diversity is key to understanding how they influence ecosystem 2 function. Deciphering linkages between biodiversity and ecosystem function at large spatial extents 3 provides a baseline for understanding how ecosystems and the components within them respond to stress 4 and global change. Determining the resilience of ecosystems depends on our ability to measure and 5 understand their response to perturbations over time. The fifth dimension highlights the importance of 6 detecting phenomena we are not vet aware of and preparing for new advances in other realms. We chose 7 these dimensions to highlight the potential of spectral biology to advance understanding and monitoring 8 of ecological processes across scales—particularly at large spatial extents—in the face of rapid global 9 change. All are relevant to managing our biosphere for sustainability. We recognize that properties and 10 processes in each dimension interact with those in all others, but we view this organization as enabling us 11 to discuss and investigate key elements in an unfolding or expanding fashion (Fig. 6).

12

13 1. Composition and Diversity

Spectral biology has made considerable advances in characterizing the identity and composition of organisms,
particularly plants, and in quantifying the diversity and composition of vegetation in ecosystems. These
developments also have potential to support evaluating the many organisms that depend on plants for their life
cycles and livelihoods. We first consider these capabilities and future potential before discussing how they
may be impacted by environmental change.

19

20 Composition

One of the most powerful attributes of spectral data is its ability to discern identity and composition by coupling reflectance information across many wavelengths with pattern detection, including machine learning approaches. While spectroscopy has been widely used to identify stars and the presence of specific gases and elements in space, its application to differentiating genotypes, species and lineages of plants on Earth has more recently expanded (Asner and Martin 2016). Species and functional group identification from airborne spectra are well-established for temperate forest trees (Roberts et al. 1998,

1 Plourde et al. 2007, Williams et al. 2020, Sapes et al. 2022) and remain challenging in hyperdiverse 2 tropical systems (Baldeck et al. 2015), particularly from satellites, due to restrictions on spatial resolution 3 and signal-to-noise ratio for instruments in orbit (Papes et al. 2010). The ability to classify plant species 4 depends crucially on spatial resolution and scale (Wang and Gamon 2019). Across biological scales from 5 genotypes within species (Stasinski et al. 2021, Li et al. 2023), species within lineages and lineages 6 within larger clades, classification appears to have high accuracy at the leaf level (Meireles et al. 2020) 7 and even across canopies (Torabzadeh et al. 2019, Seeley et al. 2023, Griffith et al. 2023). Classification 8 approaches may have greater accuracy or consistency at phylogenetic scales above the level of the species 9 (Cavender-Bares et al. 2016), in other words at the scale of lineages that roughly correspond to genera or 10 subgenera. Detecting lineages rather than species may be critical in highly diverse tropical regions where 11 species-level information is often impossible to obtain on the ground.

12 Detection of ecosystem composition and identity of component lineages, species, or genotypes is 13 made challenging by shifts in spectral signatures through time (Chlus and Townsend 2022), by the 14 expression of both genetically and environmentally driven variation within taxa (Madritch et al. 2014, 15 Czyż et al. 2020), and by the many complications of different sensors and conditions across observations 16 (Li et al. 2023). The nature of these technical challenges shifts from handheld instruments to uncrewed 17 aerial vehicle (UAV) sensors to airborne sensors and the myriad satellite sensors (Schneider et al. 2017, 18 Helfenstein et al. 2022). Of the space agency-funded satellites, all have resolutions of 30 m or coarser, 19 requiring statistical approaches to discern identity at the scale of individual organisms that will be smaller 20 than the pixel size.

Using 30 m pixel satellite data (Landsat Thematic Mapper (L1TP) and Hyperion imaging
spectroscopy from NASA's EO-1 satellite, Visser et al. (2025, this feature) were able to differentiate
lianas, as a functional group, from trees. They used radiative transfer models that detect differences in leaf
angles and revealed larger apparent leaf areas and increased light scattering in the NIR and SWIR regions
in lianas, which they attributed to their less costly leaf construction compared to tree leaves.

1 Diversity

2 Various approaches have emerged for linking remotely sensed spectral diversity and *in situ* measures of 3 ecosystem diversity (Rocchini et al. 2010). Ecosystem diversity has sometimes been predicted by taking 4 advantage of identity detection using spectral libraries. For example, Williams et al. (2020) used airborne 5 spectroscopic imagery from AVIRIS NG at 1 m resolution to classify forest canopies in a young 6 experimental forest. By detecting species co-occurring within communities, they predicted forest diversity 7 with high accuracy (up to 12 species). They subsequently used remotely sensed predictions of forest 8 biomass to accurately predict tree diversity - ecosystem function relationships. Plant diversity has also 9 been directly predicted from spectra and from spectral diversity using methods that do not rely on identity 10 detection and range from simple measures of the coefficient of variation (CV) among spectra retrieved 11 from a vegetation plot to detection of spectral species (e.g., Frye et al. 2021). Wang et al. (2018) used the 12 coefficient of variation of spectra from experimental prairie systems at pixel sizes that ranged from 1 mm 13 to 1 m. Here the scale and resolution were critical, and spectral diversity was only predictive of plant 14 diversity at resolutions similar to that of whole plants, leaves, or stems. Gholizadeh et al. (2019, 2020) 15 used a similar approach in more diverse prairie systems and found that the CV of spectra predicted plant 16 diversity even at coarser resolutions up to ~ 4 m. Further studies (Schneider et al. 2017, Kamoske et al. 17 2022, Rossi et al. 2022) using additional spectral diversity metrics (e.g., convex hull volume [CHV], 18 spectral species [SS], total variance [TV]) found that accurate predictions will also depend on the metric 19 used to assess plant diversity from above. For example, some metrics are more susceptible to outliers than 20 others and thus did not capture the variability of local plant communities (Rossi et al. 2022). Despite 21 challenges, the variability of even a small number of spectral bands has enabled successful detection of 22 boreal forest diversity variation in time and space (Xi et al 2024). 23 The spectral species concept—pixels with similar signatures in the spectral space (sensu, Féret

and Asner 2014)—has gained traction as a conceptual and analytical means to predict plant species and
 communities (Féret and de Boissieu 2020, Rocchini et al. 2022). Using spectral species, Pinto-Ledezma et
 al. (in press) found consistent predictions across multiple dimensions of plant diversity across multiple

NEON sites and biomes in the United States. Guzman et al. (2025, this feature) used structural diversity
 based on UAV LiDAR measurements across the season to predict forest diversity and consequences for
 ecosystem function in an experimental forest. Forest communities that changed more in their structural
 diversity across the season also had greater ecosystem productivity.

5

6 *Connecting spectra to the tree of life*

7 Species and lineages represent points along a continuum from genetic variation among cells and 8 individuals, to quantitatively increasing genetic differentiation defining clades across the tree of life (Fig. 9 5a). In this way, genetic diversity is not distinct from species or clade diversity, but a finer point to put on 10 our understanding of biological diversity. Genetic diversity concerns differences that are passed on 11 through generations, and therefore subject to evolutionary processes, such as gene flow, selection, 12 mutation, and genetic drift. These processes can result in genotypic diversity and differentiation between 13 populations that have phenotypic consequences. Spectra are information-rich measures of the phenotypes 14 that result from the interaction between genotypes and the environment and, consequently, can be used to 15 address genetic and evolutionary questions (Babar et al. 2006, Cavender-Bares et al. 2017, Kothari and 16 Schweiger 2022). The same kinds of features that allow the separation of species and clades by their 17 spectra (Meireles et al. 2020) can also help assess within-species genetic variation, including 18 differentiation among genotypes and populations (Cavender-Bares et al. 2016). Recent work has indicated 19 that, within specific environments, genetically more diverse populations of plants are also spectrally more 20 diverse (Hernandez-Leal, in review; Li et al. 2023) and that spectra can differentiate some genotypes and 21 their F1 crosses as intermediate between signatures of the parent genotypes (Seeley et al. 2023). 22 Similarly, in naturally occurring stands of hybrid poplars, Deacon et al. (2017) showed that spectral 23 phenotypes were intermediate between the parental species. 24 Studies in this area can draw on the rich toolkit of quantitative genetics, a discipline that has 25 dissected the quantitative relationship between phenotypic and genotypic variation since before the nature

26 of genetic material was known (Falconer and Mackay 1996). More recently, as whole-genome sequencing

1 techniques became increasingly affordable and available, genome-wide association studies (GWAS) 2 became a staple of quantitative genetics (Bazakos et al. 2017). In this issue, Li and co-authors test 3 approaches to apply GWAS to spectra, as well as to spectral features related to specific traits (aspects of 4 phenotypes). They quantify narrow-sense heritability that different parts of a spectrum represent, i.e., the 5 extent to which additive genetic variation contributes to additive variation in spectra; and associate 6 specific genetic and spectral variants. Spectra have also been shown to capture genomic variation in the 7 face of biological processes that blur the lines between populations, such as gene flow, and species, such 8 as hybridization. Stasinski et al. (2021) used leaf spectra to differentiate two species of Dryas that co-9 occur and hybridize and to furthermore distinguish populations within each of those species and showed 10 that the degree of genetic ancestry of an individual plant can be predicted from spectra. 11 12 2. Linking composition and diversity to ecosystem function 13 Spectral biology further enables us to predict plant and ecosystem function—including structural, 14 chemical, photosynthetic and productivity dimensions—making possible large-scale assessments of the 15 relationships between ecosystem diversity, composition and function. Consistent, large-scale applications 16 of this potential remain untapped. 17 18 *Plant and ecosystem function* 19 The capacity of spectral information to predict a wide array of plant functional traits opens new doors for 20 mapping plant function across ecosystems (Wang et al. 2019, 2020b) and scaling up to the biosphere (Jetz 21 et al. 2016, Dechant et al. 2024). Spectral data and derived traits relate directly to photosynthesis, carbon 22 dynamics and resource allocation (Serbin et al. 2015, DuBois et al. 2018). These advances will ultimately 23 enable the inclusion of satellite-detected changes in plant function in Earth system models that predict

24 biosphere dynamics on our rapidly changing planet.

25 Pierrat et al. (2025, this feature) demonstrate the use of proximal remote sensing of solar induced
26 chlorophyll fluorescence (SIF) to discern seasonal changes in photosynthetic yields in *Pinus palustris* and

1 other evergreen needleleaf species at needle and canopy scales. This builds on long-standing efforts to use 2 SIF to measure ecosystem photosynthesis and productivity (Morales et al. 1999, Flexas and Medrano 3 2002, Freedman et al. 2002, Moya and Cerovic 2004, Sun et al. 2018) and to scale up from leaves to 4 ecosystems (Gamon and Qiu 1999, Cavender-Bares and Bazzaz 2004, Asner and Martin 2008). Detection 5 of ecosystem function has been a major global effort, with robust indices (NDVI) to detect GPP and the 6 development of Earth surface models e.g., (Sellers et al. 1996) and is at a highly advanced stage in terms 7 of predicting productivity and its change through time (Mohammed et al. 2019) in a range of diverse 8 ecosystems (Zhang et al. 2022, Dabrowska-Zielińska et al. 2022). The coupling of space-borne LiDAR 9 and satellite data is rapidly enhancing global accuracy in monitoring of global ecosystem structure and 10 function (Saarela et al. 2018, Schneider et al. 2020, Di Tommaso et al. 2021, Liu et al. 2022).

11

12 Biodiversity-Ecosystem Function relationships

13 More recent developments have involved using detection of diversity and ecosystem function to decipher 14 how dimensions of biodiversity, including spectral diversity are associated with ecosystem function 15 (Madritch et al. 2014, Schweiger et al. 2018, Williams et al. 2020). While a large body of evidence has 16 shown relationships between species diversity and ecosystem function in experimental systems for a 17 quarter of a century (e.g., Tilman 1999, Reich et al. 2001, Isbell et al. 2015, Grossman et al. 2017, Huang 18 et al. 2018), similar relationships in natural systems have been demonstrated more recently (Liang et al. 19 2016, Oehri et al. 2017, Chen et al. 2023, Liu et al. 2024) albeit with some inconsistency across scales, 20 biomes and climates (Chisholm et al. 2013, Cheng et al. 2023). Spectral biology approaches are only 21 beginning to be applied at large spatial extents to detect these relationships (Oehri et al. 2020, Schuldt et 22 al. 2023, Liu et al. 2024). Williams et al. (2025, this feature) detect the influence of forest canopy 23 composition on the transmittance of light, showing how experimental forest communities of different 24 phylogenetic lineages change the light quality and quantity that reaches the understory. Guzmán et al 25 (2025, this feature) use remotely sensed lidar across the growing season to decipher changes in forest 26 structure that are associated with critical dimensions of forest diversity and predict ecosystem biomass.

Marcilio-Silva et al. (2025, this feature) use GEDI LiDAR data from space in urban forest patches
 coupled with ground-based measurements of forest diversity and structure to map urban forests. In doing
 so, they uncover the importance of management legacies in urban forest structure. Understanding the
 linkages between plant canopies that can be spectrally observed from above and the soils processes that
 both influence and are influenced by them are critical spectral detection of belowground ecosystem
 processes (Madritch et al. 2014, 2020, Cavender-Bares et al. 2022b).

7

8 **3.** Environmental factors, stress, and global change

9 In a world exposed to increasing threats from climate change, expansion of pests and pathogens, 10 disturbance and land-use change, and increasing pollution loads in the environment, spectral biology has 11 the potential to help detect and differentiate stressors of plants at large spatial extents. Doing so across 12 scales from leaves of individual plants to tree canopies and whole landscapes will require a range of 13 methodologies that may be combined for deeper understanding of mechanisms and interactions of 14 multiple stressors. We emphasize the importance of framing spectral biology in terms of careful 15 integration of spectroscopic and remote sensing methods with stress physiology and pathology, including 16 in-depth understanding of the life-cycle and natural history of biotic stress agents and disease progression, 17 as well as the physiological responses of plants to drought, pollution and their synergies with biotically-18 induced disease. Stress leaves markers in spectral signatures of leaves, canopies and landscapes, some of 19 which can be generalized and scaled up using spectral regions that show changes in photosynthetic 20 biology, carotenoid and photoprotective pigments and changes in foliar water content across spatial 21 resolution and extent. Other stress markers are more idiosyncratic of specific stress factors and may 22 involve spatial or temporal patterns at the canopy or landscape scale that are diagnostic of a specific 23 pathogen. The degree to which more general stress signatures or system-specific responses are useful in 24 addressing questions regarding ecological processes depends on prior knowledge of organismal function 25 and species interactions as well as the scale of inquiry.

1 Using a unique open-air field experiment in Minnesota, USA, Stefanski et al (2025, this issue) 2 examined the spectral signature of experimental warming by collecting leaf spectral reflectance (400-3 2400 nm) at the peak of the growing season for three years on juveniles (two to six years old) of five tree 4 species. They found that the imprint of environmental conditions, including those associated with 5 experimental warming, experienced by plants hours to weeks prior to spectral measurements was linked 6 to spectral regions associated with stress, in particular the water absorption regions of the near-infrared 7 and shortwave infrared. In contrast, the conditions plants experienced during leaf development, again 8 including those associated with climate manipulations, left lasting imprints on the spectral profiles of 9 leaves measured much later in the growing season; those imprints were related to structural and chemical 10 leaf attributes (e.g., pigment content and associated ratios). Moreover, after accounting for species 11 differences, spectral responses to warming did not differ among species, suggesting that developing a 12 general framework for quantifying forest responses to climate change through spectral biology may be 13 feasible.

14

15

16 Signatures of stress across scales

Spectral and point cloud data are increasingly being used to detect trees that are dead or dying as a
consequence of drought, disease, and other global change-related stressors (Pontius et al. 2008, Hanavan
et al. 2015, Asner et al. 2016, 2018). Detecting mortality and discerning its causes is essential to
managing ecosystems in the face of multiple simultaneous stressors. Rapid detection of disease is critical
to management in stopping the spread of a pathogen. Less expensive containment measures can be used
when disease invasion is detected early, reducing cost.

Plants respond to environmental stress with a limited set of physiological symptoms that can
 often be detected spectrally. At the level of physiological function in leaves, for example, reduced
 photosynthetic function and water content are common responses to drought and wilting diseases as a
 consequence of damage to the photosynthetic apparatus or reduced vascular function, which limits water

1 supply for gas exchange. Changes in chlorophyll concentration and in water content in leaves are readily 2 detectable signatures of stress from leaves to canopies to landscapes (Sapes et al. 2022, 2024, Guzmán et 3 al. 2023). Increases in expression of pigments used for photoprotection may be another general stress 4 response (Savage et al. 2009, Ramirez-Valiente et al. 2015, Encinas-Valero et al. 2021, Kothari et al. 5 2021). When photosynthetic rates are slowed due to stress (e.g., drought, cold, low nutrients, disease, 6 pollution), less absorbed light can be used for photochemistry. Consequently, plants often upregulate 7 photoprotective pigments (xanthophyll-cycle carotenoid pigments) that dissipate light energy as heat to 8 prevent oxidative damage to the protein components involved in photosynthesis (Demmig-Adams and 9 Adams 2000). Increased expression of carotenoids, detected by spectral regions in the visible—including 10 indices such as the photochemical reflectance index (PRI, (Gamon et al. 1997)) and the chlorophyll 11 carotenoid index (CCI, Gamon et al. 2016)—may be fairly generalizable responses to stress that can be 12 detected across spatial resolutions and extents. At the same time, each disease or disease syndrome may 13 have a distinct temporal and spatial progression pattern, enabling early and/or rapid detection of specific 14 pathogens and differentiating them from drought.

15 Across plant taxa, environmental stress alters not only the phytochemical composition of leaves, 16 but also the structure-and ultimately function-of canopies, impacting remote sensing signals. For 17 example, drought stress causes notable physiological and chemical shifts aimed at facilitating plant 18 survival through regulating key biological processes through hormonal signaling (McDowell et al. 2022, 19 Rai et al. 2024, Sato et al. 2024). Similarly, drought has also been shown to affect leaf chemical and 20 structural attributes-including changes in amino acids and pigment composition (Demmig-Adams and 21 Adams 2006, Yang et al. 2021), leaf size and density (i.e., leaf area index), orientation, and water content. 22 The extent of these changes are highly dependent on the severity and duration of stress, resulting in high 23 temporal and spatial variation. It can be challenging to disentangle the contribution of canopy structural 24 changes and leaf-level physiological changes, particularly when the spatial resolution of the sensor is 25 course relative to the size of leaves or canopies, making this a fertile area for investigation.

1 Understanding the biology of the disease can be critical to detecting it remotely. Pests and 2 pathogens tend to be lineage-specific, often requiring biological knowledge of the host, the pathogen and 3 the biotic vector. Within the oaks, the oak wilt fungal pathogen (*Bretziella fagacearum*) is considered the 4 most deadly threat to the genus, particularly the red oak lineage (*Ouercus* section Lobatae). Its spores are 5 spread overland long distances by nitidulid sap beetles that can infect vulnerable trees if the cambium is 6 exposed from cracks or cuts (Juzwik et al. 2011). The spread to neighboring oak trees can be quite rapid 7 when roots from an infected tree graft with a neighbor, allowing the fungus to move from one tree to the 8 next (Koch et al. 2010). As trees succumb to the disease, there is a temporal progression of symptoms that 9 aid detection using time series data, as well as a characteristic spatial pattern. 10 Spectral signatures are capable of differentiating disease symptoms of the pathogen from drought 11 stress at leaf and canopy scales in both indoor (Fallon et al. 2020) and outdoor experiments (Sapes et al. 12 2024) due to differences in the spectral features that are affected and the rate of change. Heterogeneity in 13 pigment concentrations in foliage across the canopy, as a consequence of tylose formation in the xylem 14 that causes some branches to wilt, is characteristic of the disease and can be used to differentiate it from 15 drought using even inexpensive multispectral UAV sensors. At landscape scales, both spectral features

that can be characterized at the whole-plant level as well as temporal and spatial patterns can be detected
spectrally. Features in spectroscopic airborne imagery take advantage of host specificity in the disease to

18 help detect vulnerable hosts. Sapes et al. (2022) developed models to differentiate oaks from other tree

species, oak lineages vulnerable to oak wilt from less susceptible oaks and ultimately healthy from

20 diseased oaks, for accurate detection of the disease. At regional scales, land surface phenological metrics

21 used understanding of the temporal progression of disease to detect healthy, symptomatic and dead oak

trees of specific lineages using currently available satellite data (Sentinel2 and Landsat 8) in near-real

time with accuracies sufficient to aid management (Guzmán et al. 2023). Rapid and accurate detection

24 increases management options, from early options that may only involve girdling a single tree and

25 injecting herbicide, to more expensive options that involve the use of a vibratory plow and removal of

surrounding trees. If the disease is not treated early, it can spread to such extents that the cost of effective
 treatment becomes prohibitive.

3 Spectral detection of stress responses are often not diagnostic of specific diseases (Pontius and 4 Hallett 2014, Pontius et al. 2020). The extent to which particular host-disease systems are discernable and 5 whether those diagnostic responses are idiosyncratic or themselves generalizable is an open question, but 6 one where rapid progress is being made. Drought predisposes many trees to infection by pests and 7 pathogens. Most tree lineages are threatened by multiple pests and pathogens, with similarities in 8 symptoms. Deciphering the cause of decline and mortality is likely to remain complicated. Spectral 9 biology has the potential to detect ecosystem-scale stress and connect it to whole-organism understanding 10 of biotic and abiotic stress responses as a means of understanding underlying mechanisms of forest 11 decline to aid management.

12 Rapid detection of stress physiology is now possible at scales and frequencies that would be 13 infeasible from the ground. Even if the mechanism of stress is not discernable, detecting the location of 14 stress in ecosystems aids management. Forests are expressing novel phenotypes due to rapid rates of 15 change and the emergence of novel environments (Housset et al. 2018). An important question is whether 16 ecosystem-level responses to stress are generalizable or whether each specific system is distinct, requiring 17 specific local knowledge to decipher stress responses. Is there convergence in system-level responses 18 across ecosystem types and host-disease systems, from lodgepole mountain pine beetle attack to oak wilt, 19 in terms of stress physiology? Or do we need more detailed information about life histories of pests and 20 pathogens to understand how each disease is expressed? Integration among subdisciplines is critical, with 21 remote sensing of spectral information providing one tool, but only partial answers. Unique combinations 22 of stress that do not have historical analogs may produce unique signatures of stress. Given the rate of 23 global change, it is more important than ever to detect these kinds of stress responses, and it is now 24 possible to examine interacting stress factors in ways we could not before. Rapid detection of stress is 25 critical to replanting and reforesting and will advance restoration and rehabilitation efforts mandated in 26 the Global Biodiversity Framework of the Convention on Biological Diversity.

2 Genetic variation in stress response detectable from spectral phenotypes

3 Stress detection has received enormous attention in crops and with the goal of connecting spectrally 4 derived functional information to genomic mechanisms (Mohd Asaari et al. 2018, Wang et al. 2020a, 5 Calzone et al. 2021). Regulation of suites of genes in response to stress changes spectrally observable 6 phenotypes (Tirado et al. 2020). In ecology and evolution, we often need to assess the performance of 7 individual organisms or groups as indicators of their acclimation or fitness in the face of stress, but we do 8 not have complete ways to measure performance. Traditionally, we measure one or a few traits as a 9 proxy. In the worst case, one trait such as biomass accumulation is set as "equivalent" to performance, 10 which is misleading and inhibits deeper thinking about organisms as agents, and mechanisms and facets 11 of resilience. Having a more integrative measurement that lends itself to spatial and temporal scaling may 12 help us to better consider how, when, and in what ways to assess different aspects of performance, and 13 remind us that we are evaluating a multifaceted phenomenon.

14

15 4. Resistance, recovery, and resilience

16 Resistance, recovery, stability and resilience are concepts receiving increased attention in ecosystem and 17 global change ecology, in relation to both strong event-type disturbances and chronic pressures. Despite 18 inconsistent definitions (which harms progress), conceptual coherence and a variety of useful approaches 19 make this an area of current and future focus and importance (Yi and Jackson 2021, Tai et al. 2023). 20 Investigating these concepts over relevant time scales (decades to centuries) requires repeated 21 observations that are challenging to acquire with direct observations. In contrast, remotely sensed data, 22 which often is possible to acquire repeatedly over time, plays a special role in the development of both 23 resilience theory and its testing (Pontius et al. 2020, Liu et al. 2021, Yi and Jackson 2021, Tai et al. 2023). 24 Spectral biology enables us to observe ecosystems through time to test how diversity and composition 25 influence resistance, recovery, and stability (Isbell et al. 2015): processes receiving increasing attention as 26 important in a changing world (Wilcox et al. 2020, Avolio et al. 2021).

1	Frequent (e.g. every 1-2 weeks) or infrequent (e.g. seasonal to annual) satellite measurements
2	provide spectral information on ecosystems and how they change, which encompasses ecosystem
3	resistance and resilience (Fig. 6). Capturing transition states and predicting shifts in ecological function
4	under global change (Tai et al. 2023) will increasingly be critical to understanding how the Earth is
5	changing and provide important input for the sustainable management of ecosystems.
6	Diversity likely plays a key role in resilience. Linkages between diversity (e.g. species richness,
7	phylogenetic diversity, functional diversity) and stability are well established; for example, evidence is
8	increasing that forest diversity increases drought resistance in experimental systems (Blondeel et al.
9	2024). Such evidence has required long-term experiments, constraining analyses to small spatial extents, a
10	handful of biomes, and relatively few species. Time series data collected across the Earth's surface can be
11	used to feed or test models predicting relationships between diversity and function, and help decipher how
12	trends in ecosystem function are related to processes of resistance, compositional turnover, and recovery
13	after disturbance that influence resilience (Xu et al. 2022). Studies of tipping points and their signatures
14	indicate that increased variability can precede a regime shift to an alternative degraded state of an
15	ecosystem (Scheffer et al. 2001, Scheffer and Carpenter 2003, Steffen et al. 2015).
16	A mechanistic understanding of change will increase predictive capacity, even in non-linear
17	ecosystem dynamics – where detecting thresholds is critical. Changes in biome extent over time have long
18	been detected using NDVI (Simms and Ward 2013). Shifts in alpine ecotones in response to warming
19	climates have been detected in the Western US (Wei et al. 2020). Remotely sensed resilience data enabled
20	prediction of subsequent drought mortality across the continental US (Tai et al. 2023). An important
21	element is understanding the mechanisms underlying ecosystem transitions, which includes deciphering
22	causes of mortality, stress, and disturbance. High spectral resolution is important to understand
23	compositional changes and pinpoint changing physiological functions. Historically, scientists have
24	considered different stress factors in isolation. Complexities of interacting stress result in emergent
25	properties that can be detected using a holistic measurement approach such as that of spectral biology,

and untangled through a mechanistic approach by extracting specific information from spectral time
 series in combination with other data types.

3 For example, Sturm et al. (2022) used changes in the canopy Normalized Differential Water 4 Index (NDWI) from a time series of multispectral satellite imagery from Sentinel-2 to calculate the 5 resistance, resilience, and recovery of forests across Switzerland to an unusually severe drought event in 6 2018. They explained differences among forests based on landscape characteristics and forest mixing 7 ratios (e.g. proportion of needle versus broadleaf trees). Helfenstein et al. (2024) used the same approach 8 to study the relationships of resistance, resilience, and recovery with functional diversity as calculated 9 from pigments and water content in the same forests (using different images for diversity metrics versus 10 the time series calculations) and found positive relationships of functional richness with both resistance 11 and resilience to drought. These kinds of patterns can also be detected in managed, urban, or naturally 12 assembled ecosystems through spectral and LiDAR information over time that is well-connected to 13 measured biological processes on the ground (Marcilio-Silva et al. 2025, this feature). Ultimately, these 14 approaches will enable mechanistically informed monitoring of forest stress responses and resilience.

15

16 5. Discovery

17 Finally, spectral biology will advance the realm of discovery by opening our capacity to observe Earth 18 and the living world around us. What new patterns can we quantify as a consequence of the ability to 19 "see" deep patterns and mechanisms across spatial and temporal scales? The new frontiers that will 20 emerge will encompass measurements of the linkages among the full range of biological organization, 21 and evolutionary and environmental drivers of plant distributions and functions, as well as their genetic 22 structure, competitive interactions and relationships to components of ecosystems such as microbes or 23 pathogens, detected by other methods. The capability of spectral biology to detect diversity, composition 24 and function of ecosystems, and how they change in response to stress through space and time, enables 25 new pathways for discovery at vastly divergent scales.

26

1 The high dimensionality of spectra provides insurance against our ignorance

Through the linkage of spectroscopy with biology, the potential of spectral biology goes beyond what our
frameworks and methods currently allow for (Townsend et al. 2013). For example, VSWIR spectroscopy
(400 – 2500 nm) captures coherent (i.e., non-noise) information beyond the variables we currently
estimate from the imagery (Schimel et al. 2020, Cawse-Nicholson et al. 2022) or use to model leaf
reflectance via physical models (e.g., Féret and De Boissieu 2024). The high dimensionality of spectral
data can enable future discoveries unlocked by advances in machine learning models, computational
power, technological advances in associated areas, and conceptual breakthroughs (Hong et al. 2024).

9 Stronger links between genetic diversity and spectra can be forged as the cost of genomics and 10 transcriptomics come down and spectral biology becomes more democratized. Spectral biology can help 11 guide genomic and transcriptomic analyses for scientists and ecosystem managers alike by identifying 12 promising relationships for deeper investigation: it may help to more efficiently search for the proverbial 13 needle in a haystack. Specifically, advances in scalable monitoring of biological diversity enable 14 measurement prioritization. In particular, the emerging Earth observation platforms that we envision will 15 lower barriers to entry to spectral biologists and provide the foundation for more effective monitoring of 16 biological diversity with tighter links of monitoring to mechanism and response.

17 Ultimately, the ability to detect patterns at broad spatial extents through time will facilitate the 18 discovery of phenomena relevant to understanding biological processes across scales. The broad spatial 19 perspective will allow us to test whether relationships we observe at fine scales or from experimental 20 studies are generalizable at regional-to-planetary scales, and, if not, why. Thus, we expect that advances 21 in technology will be followed by increases in the spatial extent of composition, functional, and stress 22 measurements that will facilitate either verification or falsification of hypothesized mechanisms, or, 23 alternatively, reveal patterns of variation not previously characterized. Already it is clear that more 24 functional variation emerges when functional traits are spatially mapped from above than is predicted 25 from functional trait measurements on the ground, largely due to the vastly increased sample size that 26 results from using image data (Wang et al. 2020b). In order to produce comparable measurements at the pace of fieldwork, most functional ecologists adhere to specific protocols for how and when traits are measured on plants, and focus on specific seasonal and ontogenetic life stages, prioritizing certain organs over others. Remotely sensed and spectrally derived functional variation is agnostic to these protocols and can pick up otherwise hidden functional variation. The "insurance against ignorance" is that we have only scratched the surface of our understanding of the drivers of spectral variation, meaning that our archived records provide a repository of data that can be re-mined into the future as we build out our knowledge in spectral biology.

8 We will no doubt detect patterns that we could not see in other ways, and there is room for pattern 9 discovery in remote sensing across spatial and temporal scales, similar to the development of genomics. 10 Much of the focus of spectral biology to date has been on readily detected patterns, such as quantification 11 of traits that drive photosynthesis, like chlorophyll and nitrogen concentrations or leaf mass per area. 12 What is truly exciting is the potential to detect unanticipated anomalies or exceptions to expected 13 relationships—e.g., where predicted trait-trait or trait-environment relationships break down—or where 14 new relationships are observed that had not previously been identified as important. Advances in 15 modeling and computational tools may allow us to learn from the signals obtained across scales and study 16 planet Earth as a system, finally deciphering how processes at one scale influence and are influenced by 17 those at all other scales.

18 At the same time, it is important to acknowledge that many gaps remain in accurate interpretation 19 of signals, and excitement about advancing technology can result in overselling its potential. Signals can 20 only be interpreted to the extent that we can connect them to meaningful biological processes and patterns 21 that are carefully measured, understood, and verified in appropriate ways. There are technical issues with 22 signal detection from a distance based on geometry and atmospheric interference, as discussed earlier. 23 Near-surface remote sensing data are hard to acquire over time and require considerable training and 24 infrastructure investment; multiple interacting biological and environmental factors can be difficult to 25 disentangle. There is no shortcut to conducting the careful *in situ* work to decipher mechanisms

underlying biological phenomena that enables extension of our understanding across spatial and temporal
 scales.

3

4 Conclusions

5 We close by emphasizing that spectral biology has enormous potential to expand the spatial extents and 6 timeframes at which we can decipher ecological processes relevant to managing our planet. Importantly, 7 ecologists have a critical role to play in conducting the research to enable accurate biological 8 interpretation of signals, whether from spectral measurements made at fine scales, or from the sky. The 9 theoretical frameworks and extensive field, experimental, and laboratory observations and analyses that 10 underpin the inferences made from spectral data are critical to the effective use of these measurements. 11 The tools of spectral biology, which still present challenges to accurate interpretation, also provide keys 12 to understanding and monitoring vegetation on Earth from the finest scale to our entire planet in ways that 13 have not been possible before. Moreover, by linking across components of the ecosystem, such as soil 14 biota, animals and microbes, we can further disentangle trophic and other complex or non-linear 15 dynamics operating across spatial and temporal scales. Spectral biology is one framework that will help 16 us to harness the information necessary for local to global efforts to manage ecosystems and sustain a 17 habitable planet. The framework and tools will increasingly play an important role in knowing how we 18 are doing in meeting the goals and targets of the Global Biodiversity Framework (Kissling et al. 2018, 19 Skidmore et al. 2021, Cavender-Bares et al. 2022a, Gonzalez et al. 2023).

20

21 Acknowledgements

22 This work was supported by the NSF Biology Integration Institute ASCEND

23 (Advancing Spectral Biology in Changing Environments to understand Diversity; NSF-DBI-

24 2021898). The figures were partially designed by seanquinn.studio.

25

1 Literature Cited

2	Asner, G. P., P. G. Brodrick, C. B. Anderson, N. Vaughn, D. E. Knapp, and R. E. Martin. 2016.
3	Progressive forest canopy water loss during the 2012–2015 California drought. Proceedings of the
4	National Academy of Sciences 113:E249.Asner, G. P., and R. E. Martin. 2008. Spectral and
5	chemical analysis of tropical forests: Scaling from leaf to canopy levels. Remote Sensing of
6	Environment 112 :3958-3970.
7	Asner, G. P., and R. E. Martin. 2016. Spectranomics: Emerging science and conservation opportunities at
8	the interface of biodiversity and remote sensing. Global Ecology and Conservation 8:212-219.
9	Asner, G. P., R. E. Martin, L. M. Keith, W. P. Heller, M. A. Hughes, N. R. Vaughn, R. F. Hughes, and C.
10	Balzotti. 2018. A spectral mapping signature for the Rapid Ohia Death (ROD) pathogen in Hawaiian
11	forests. Remote Sensing 10:404.
12	Avolio, M. L., K. J. Komatsu, S. L. Collins, E. Grman, S. E. Koerner, A. T. Tredennick, K. R. Wilcox, S.
13	Baer, E. H. Boughton, A. J. Britton, B. Foster, L. Gough, M. Hovenden, F. Isbell, A. Jentsch, D. S.
14	Johnson, A. K. Knapp, J. Kreyling, J. A. Langley, C. Lortie, R. L. McCulley, J. R. McLaren, P. B.
15	Reich, E. W. Seabloom, M. D. Smith, K. N. Suding, K. B. Suttle, and P. M. Tognetti. 2021.
16	Determinants of community compositional change are equally affected by global change. Ecology
17	Letters 24:1892–1904.
18	Babar, M. A., M. P. Reynolds, M. van Ginkel, A. R. Klatt, W. R. Raun, and M. L. Stone. 2006. Spectral
19	reflectance to estimate genetic variation for in-season biomass, leaf chlorophyll, and canopy
20	temperature in wheat. Crop Science 46:1046–1057.
21	Baldeck, C. A., G. P. Asner, R. E. Martin, C. B. Anderson, D. E. Knapp, J. R. Kellner, and S. J. Wright.
22	2015. Operational Tree Species Mapping in a Diverse Tropical Forest with Airborne Imaging
23	Spectroscopy. PLOS ONE 10:e0118403.
24	Bazakos, C., M. Hanemian, C. Trontin, J. M. Jiménez-Gómez, and O. Loudet. 2017. New Strategies and
25	Tools in Quantitative Genetics: How to Go from the Phenotype to the Genotype. Annual Review of
26	Plant Biology 68:435–455.

1	Blondeel, H., J. Guillemot, N. Martin-StPaul, A. Druel, S. Bilodeau-Gauthier, J. Bauhus, C. Grossiord, A.
2	Hector, H. Jactel, J. Jensen, C. Messier, B. Muys, H. Serrano-León, H. Auge, N. Barsoum, E.
3	Birhane, H. Bruelheide, J. Cavender-Bares, C. Chu, J. R. Cumming, A. Damtew, N. Eisenhauer, O.
4	Ferlian, S. Fiedler, G. Ganade, D. L. Godbold, D. Gravel, J. S. Hall, D. Hölscher, K. B. Hulvey, J.
5	Koricheva, H. Kreft, C. Lapadat, J. Liang, X. Liu, C. Meredieu, S. Mereu, R. Montgomery, L.
6	Morillas, C. Nock, A. Paquette, J. D. Parker, W. C. Parker, G. B. Paterno, M. P. Perring, Q. Ponette,
7	C. Potvin, P. B. Reich, J. Rentch, B. Rewald, H. Sandén, K. Sinacore, R. J. Standish, A. Stefanski, P.
8	C. Tobin, M. van Breugel, M. V. Fagundes, M. Weih, L. J. Williams, M. Zhou, M. Scherer-
9	Lorenzen, K. Verheyen, and L. Baeten. 2024. Tree diversity reduces variability in sapling survival
10	under drought. Journal of Ecology 112:1164–1180.
11	Calzone, A., L. Cotrozzi, G. Lorenzini, C. Nali, and E. Pellegrini. 2021. Hyperspectral Detection and
12	Monitoring of Salt Stress in Pomegranate Cultivars. Agronomy 11:1038.
13	Cavender-Bares, J., and F. A. Bazzaz. 2004. From leaves to ecosystems: Using chlorophyll fluorescence
14	to assess photosynthesis and plant function in ecological studies. Pages 737-755 in Govindjee,
15	editor. Chlorophyll a Fluorescence: A Signature of Photosynthesis. Kluwer Academic Publishers,
16	Dordrecht, Netherlands.
17	Cavender-Bares, J., J. A. Gamon, S. E. Hobbie, M. D. Madritch, J. E. Meireles, A. K. Schweiger, and P.
18	A. Townsend. 2017. Harnessing plant spectra to integrate the biodiversity sciences across biological
19	and spatial scales. American Journal of Botany 104:966-969.
20	Cavender-Bares, J., J. E. Meireles, J. J. Couture, M. A. Kaproth, C. C. Kingdon, A. Singh, S. P. Serbin,
21	A. Center, E. Zuniga, G. Pilz, and P. A. Townsend. 2016. Associations of leaf spectra with genetic
22	and phylogenetic variation in oaks: prospects for remote detection of biodiversity. Remote Sensing
23	8:221.
24	Cavender-Bares, J., P. Reich, P. Townsend, A. Banerjee, E. Butler, A. Desai, A. Gevens, S. Hobbie, F.
25	Isbell, and E. Laliberté. 2021. BII-Implementation: The causes and consequences of plant

1	biodiversity across scales in a rapidly changing world. Research Ideas and Outcomes: The Open
2	Science Journal 7:e63850.
3	Cavender-Bares, J., F. Schneider, M. J. Santos, A. Armstrong, A. Carnaval, K. Dahlin, L. Fatoyinbo, G.
4	C. Hurtt, D. Schimel, P. A. Townsend, S. L. Ustin, Z. Wang, and A. M. Wilson. 2022a. Integrating
5	remote sensing with ecology and evolution to advance biodiversity conservation. Nature Ecology &
6	Evolution 6 :506-519.
7	Cavender-Bares, J., A. K. Schweiger, J. A. Gamon, H. Gholizadeh, K. Helzer, C. Lapadat, M. D.
8	Madritch, P. A. Townsend, Z. Wang, and S. E. Hobbie. 2022b. Remotely detected aboveground
9	plant function predicts belowground processes in two prairie diversity experiments. Ecological
10	Monographs 92 :e01488.
11	Cawse-Nicholson, K., A. M. Raiho, D. R. Thompson, G. C. Hulley, C. E. Miller, K. R. Miner, B. Poulter,
12	D. Schimel, F. D. Schneider, P. A. Townsend, and S. K. Zareh. 2022. Intrinsic Dimensionality as a
13	Metric for the Impact of Mission Design Parameters. Journal of Geophysical Research:
14	Biogeosciences 127:e2022JG006876.
15	Chen, X., A. R. Taylor, P. B. Reich, M. Hisano, H. Y. H. Chen, and S. X. Chang. 2023. Tree diversity
16	increases decadal forest soil carbon and nitrogen accrual. Nature 618:94-101.
17	Cheng, Z., J. Zhao, L. Ding, ZH. Shi, P. Kardol, and GL. Wu. 2023. The functioning of alpine
18	grassland ecosystems: Climate outweighs plant species richness. Journal of Ecology 111:2496-2506.
19	Chisholm, R. A., H. C. Muller-Landau, K. Abdul Rahman, D. P. Bebber, Y. Bin, S. A. Bohlman, N. A.
20	Bourg, J. Brinks, S. Bunyavejchewin, N. Butt, H. Cao, M. Cao, D. Cárdenas, LW. Chang, JM.
21	Chiang, G. Chuyong, R. Condit, H. S. Dattaraja, S. Davies, A. Duque, C. Fletcher, N. Gunatilleke, S.
22	Gunatilleke, Z. Hao, R. D. Harrison, R. Howe, CF. Hsieh, S. P. Hubbell, A. Itoh, D. Kenfack, S.
23	Kiratiprayoon, A. J. Larson, J. Lian, D. Lin, H. Liu, J. A. Lutz, K. Ma, Y. Malhi, S. McMahon, W.
24	McShea, M. Meegaskumbura, S. Mohd. Razman, M. D. Morecroft, C. J. Nytch, A. Oliveira, G. G.
25	Parker, S. Pulla, R. Punchi-Manage, H. Romero-Saltos, W. Sang, J. Schurman, SH. Su, R.
26	Sukumar, I. F. Sun, H. S. Suresh, S. Tan, D. Thomas, S. Thomas, J. Thompson, R. Valencia, A.

1	Wolf, S. Yap, W. Ye, Z. Yuan, and J. K. Zimmerman. 2013. Scale-dependent relationships between
2	tree species richness and ecosystem function in forests. Journal of Ecology 101:1214-1224.
3	Chlus, A., and P. A. Townsend. 2022. Characterizing seasonal variation in foliar biochemistry with
4	airborne imaging spectroscopy. Remote Sensing of Environment 275:113023.
5	Cline, L. C., S. E. Hobbie, M. D. Madritch, C. R. Buyarski, D. Tilman, and J. M. Cavender-Bares. 2018.
6	Resource availability underlies the plant-fungal diversity relationship in a grassland ecosystem.
7	Ecology 99:204-216
8	Czyż, E. A., C. Guillén Escribà, H. Wulf, A. Tedder, M. C. Schuman, F. D. Schneider, and M. E.
9	Schaepman. 2020. Intraspecific genetic variation of a Fagus sylvatica population in a temperate
10	forest derived from airborne imaging spectroscopy time series. Ecology and Evolution 10:7419-
11	7430.
12	Czyż, E. A., B. Schmid, A. Hueni, M. B. Eppinga, M. C. Schuman, F. D. Schneider, C. Guillén-Escribà,
13	and M. E. Schaepman. 2023. Genetic constraints on temporal variation of airborne reflectance
14	spectra and their uncertainties over a temperate forest. Remote Sensing of Environment 284:113338.
15	Dąbrowska-Zielińska, K., K. Misiura, A. Malińska, R. Gurdak, P. Grzybowski, M. Bartold, and M.
16	Kluczek. 2022. Spatiotemporal estimation of gross primary production for terrestrial wetlands using
17	satellite and field data. Remote Sensing Applications: Society and Environment 27:100786.
18	Davies, A. B., and G. P. Asner. 2014. Advances in animal ecology from 3D-LiDAR ecosystem mapping.
19	Trends in Ecology & Evolution 29:681-691.
20	Deacon, N. J., J. J. Grossman, A. K. Schweiger, I. Armour, and J. Cavender-Bares. 2017. Genetic,
21	morphological, and spectral characterization of relictual Niobrara River hybrid aspens (Populus \times
22	smithii). American Journal of Botany 104:1878-1890.
23	Dechant, B., J. Kattge, R. Pavlick, F. Schneider, F. Sabatini, A. Moreno-Martinez, E. Butler, P. van
24	Bodegom, H. Vallicrosa, T. Kattenborn, C. Boonman, N. Madani, I. Wright, N. Dong, H. Feilhauer,
25	J. Penuelas, J. Sardans, J. Aguirre-Gutierrez, P. Reich, P. Leitao, J. Cavender-Bares, I. H. Myers-
26	Smith, S. Duran, H. Croft, I. C. Prentice, A. Huth, K. Rebel, S. Zaehle, I. Simova, S. Diaz, M.

1	Reichstein, C. Schiller, H. Bruehlheide, M. Mahecha, C. Wirth, Y. Malhi, and P. Townsend. 2024.
2	Intercomparison of global foliar trait maps reveals fundamental differences and limitations of
3	upscaling approaches. Remote Sensing of Environment 311:114276
4	Demmig-Adams, B., and W. W. Adams. 2000. Photosynthesis - Harvesting sunlight safely. Nature
5	403:371-374.
6	Demmig-Adams, B., and W. W. Adams. 2006. Tansley review: Photoprotection in an ecological context:
7	the remarkable complexity of thermal energy dissipation. New Phytologist 172:11-21.
8	Di Tommaso, S., S. Wang, and D. B. Lobell. 2021. Combining GEDI and Sentinel-2 for wall-to-wall
9	mapping of tall and short crops. Environmental Research Letters 16:125002.
10	Dubayah, R., J. B. Blair, S. Goetz, L. Fatoyinbo, M. Hansen, S. Healey, M. Hofton, G. Hurtt, J. Kellner,
11	S. Luthcke, J. Armston, H. Tang, L. Duncanson, S. Hancock, P. Jantz, S. Marselis, P. L. Patterson,
12	W. Qi, and C. Silva. 2020. The Global Ecosystem Dynamics Investigation: High-resolution laser
13	ranging of the Earth's forests and topography. Science of Remote Sensing 1:100002.
14	DuBois, S., A. R. Desai, A. Singh, S. P. Serbin, M. L. Goulden, D. D. Baldocchi, S. Ma, W. C. Oechel, S.
15	Wharton, E. L. Kruger, and P. A. Townsend. 2018. Using imaging spectroscopy to detect variation
16	in terrestrial ecosystem productivity across a water-stressed landscape. Ecological Applications
17	28:1313-1324.
18	Encinas-Valero, M., R. Esteban, AM. Hereş, J. M. Becerril, J. I. García-Plazaola, U. Artexe, M. Vivas,
19	A. Solla, G. Moreno, and J. Curiel Yuste. 2022. Photoprotective compounds as early markers to
20	predict holm oak crown defoliation in declining Mediterranean savannahs. Tree Physiology 42:208-
21	224.
22	Falconer, D. S., and T. F. C. Mackay. 1996. Introduction to Quantitative Genetics. Prentice Hall, New
23	York.
24	Fallon, B., A. Yang, C. Lapadat, I. Armour, J. Juzwik, R. A. Montgomery, and J. Cavender-Bares. 2020.
25	Spectral differentiation of oak wilt from foliar fungal disease and drought is correlated with
26	physiological changes. Tree Physiology 40:377-390.

1	Féret, JB., and F. de Boissieu. 2020. biodivMapR: An r package for α - and β -diversity mapping using
2	remotely sensed images. Methods in Ecology and Evolution 11:64-70.
3	Féret, JB., and F. De Boissieu. 2024. prospect: an R package to link leaf optical properties with their
4	chemical and structural properties with the leaf modelPROSPECT. Journal of Open Source Software
5	9:6027.
6	Flexas, C. B., J. ;. Escalona, J. M. ;. Evain, S. ;. Gulías, J. ;. Moya, I. ;. Osmond, and H. Medrano. 2002.
7	Steady-state chlorophyll fluorescence (Fs) measurements as a tool to follow variations of net CO2
8	assimilation and stomatal conductance during water-stress in C3 plants. Physiologia Plantarum
9	114:231-240.
10	Flexas, J., J. M. Briantais, Z. Cerovic, H. Medrano, and I. Moya. 2000. Steady-state and maximum
11	chlorophyll fluorescence responses to water stress in grapevine leaves: A new remote sensing
12	system. Remote Sensing of Environment 73:283-297.
13	Frankenberg, C., P. Köhler, T. S. Magney, S. Geier, P. Lawson, M. Schwochert, J. McDuffie, D. T.
14	Drewry, R. Pavlick, and A. Kuhnert. 2018. The Chlorophyll Fluorescence Imaging Spectrometer
15	(CFIS), mapping far red fluorescence from aircraft. Remote Sensing of Environment 217:523–536.
16	Freedman, A., J. Cavender-Bares, P. L. Kebabian, R. Bhaskar, H. Scott, and F. A. Bazzaz. 2002. Remote
17	sensing of solar-excited plant fluorescence as a measure of photosynthetic rate. Photosynthetica
18	40:127-132.
19	Frye, H. A., M. E. Aiello-Lammens, D. Euston-Brown, C. S. Jones, H. Kilroy Mollmann, C. Merow, J. A.
20	Slingsby, H. van der Merwe, A. M. Wilson, and J. A. Silander Jr. 2021. Plant spectral diversity as a
21	surrogate for species, functional and phylogenetic diversity across a hyper-diverse biogeographic
22	region. Global Ecology and Biogeography 30:1403-1417.
23	Gamon, J. A., K. F. Huemmrich, C. Y. S. Wong, I. Ensminger, S. Garrity, D. Y. Hollinger, A. Noormets,
24	and J. Peñuelas. 2016. A remotely sensed pigment index reveals photosynthetic phenology in
25	evergreen conifers. Proceedings of the National Academy of Sciences 113:13087–13092.

1	Gamon, J. A., R. Wang, H. Gholizadeh, B. Zutta, P. A. Townsend, and J. Cavender-Bares. 2020.
2	Consideration of Scale in Remote Sensing of Biodiversity. Pages 425-447 in J. Cavender-Bares, J.
3	A. Gamon, and P. A. Townsend, editors. Remote Sensing of Plant Biodiversity. Springer
4	International Publishing, Cham.Gamon, J., and HL. Qiu. 1999. Ecological applications of remote
5	sensing at multiple scales. Marcel Dekker, Inc., New York. Pages 805-846 in F. I. Pugnaire and F.
6	Valladares, editors. Marcel Dekker, Inc., New York.
7	Gamon, J., L. Serrano, and J. Surfus. 1997. The photochemical reflectance index: an optical indicator of
8	photosynthetic radiation use efficiency across species, functional types, and nutrient levels.
9	Oecologia 112:492-501.
10	Genty, B., J. Briantais, and N. Baker. 1989. The relationship between the quantum yield of photosynthetic
11	electron transport and quenching of chlorophyll fluorescence. Biochimica Biophysica Acta 990:87-
12	92.
13	Gholizadeh, H., J. A. Gamon, C. J. Helzer, and J. Cavender-Bares. 2020. Multi-temporal assessment of
14	grassland α - and β -diversity using hyperspectral imaging. Ecological Applications 30:e02145.
15	Gholizadeh, H., J. A. Gamon, P. A. Townsend, A. I. Zygielbaum, C. J. Helzer, G. Y. Hmimina, R. Yu, R.
16	M. Moore, A. K. Schweiger, and J. Cavender-Bares. 2019. Detecting prairie biodiversity with
17	airborne remote sensing. Remote Sensing of Environment 221:38-49.
18	Gonzalez, A., P. Vihervaara, P. Balvanera, A. E. Bates, E. Bayraktarov, P. J. Bellingham, A. Bruder, J.
19	Campbell, M. D. Catchen, J. Cavender-Bares, J. Chase, N. Coops, M. J. Costello, M. Dornelas, G.
20	Dubois, E. J. Duffy, H. Eggermont, N. Fernandez, S. Ferrier, G. N. Geller, M. Gill, D. Gravel, C. A.
21	Guerra, R. Guralnick, M. Harfoot, T. Hirsch, S. Hoban, A. C. Hughes, M. E. Hunter, F. Isbell, W.
22	Jetz, N. Juergens, W. D. Kissling, C. B. Krug, Y. Le Bras, B. Leung, M. C. Londoño-Murcia, JM.
23	Lord, M. Loreau, A. Luers, K. Ma, A. J. MacDonald, M. McGeoch, K. L. Millette, Z. Molnar, A. S.
24	Mori, F. E. Muller-Karger, H. Muraoka, L. Navarro, T. Newbold, A. Niamir, D. Obura, M.
25	O'Connor, M. Paganini, H. Pereira, T. Poisot, L. J. Pollock, A. Purvis, A. Radulovici, D. Rocchini,
26	M. Schaepman, G. Schaepman-Strub, D. S. Schmeller, U. Schmiedel, F. D. Schneider, M. M.

1	Shakya, A. Skidmore, A. L. Skowno, Y. Takeuchi, MN. Tuanmu, E. Turak, W. Turner, M. C.
2	Urban, N. Urbina-Cardona, R. Valbuena, B. Van Havre, and E. Wright. 2023. A global biodiversity
3	observing system to unite monitoring and guide action. Nature Ecology & Evolution 7:1947-1952.
4	Griffith, D. M., K. B. Byrd, L. D. L. Anderegg, E. Allan, D. Gatziolis, D. Roberts, R. Yacoub, and R. R.
5	Nemani. 2023. Capturing patterns of evolutionary relatedness with reflectance spectra to model and
6	monitor biodiversity. Proceedings of the National Academy of Sciences 120:e2215533120.
7	Grossman, J. J., J. Cavender-Bares, S. E. Hobbie, P. B. Reich, and R. A. Montgomery. 2017. Species
8	richness and traits predict overyielding in stem growth in an early-successional tree diversity
9	experiment. Ecology 98:2601-2614.
10	Guzmán, J. A., J. N. Pinto-Ledezma, D. Frantz, P. A. Townsend, J. Juzwik, and J. Cavender-Bares. 2023.
11	Mapping oak wilt disease from space using land surface phenology. Remote Sensing of Environment
12	298:113794.
13	Hanavan, R. P., J. Pontius, and R. Hallett. 2015. A 10-Year Assessment of Hemlock Decline in the
14	Catskill Mountain Region of New York State Using Hyperspectral Remote Sensing Techniques.
15	Journal of Economic Entomology 108:339-349
16	Helfenstein, I. S., F. D. Schneider, M. E. Schaepman, and F. Morsdorf. 2022. Assessing biodiversity from
17	space: Impact of spatial and spectral resolution on trait-based functional diversity. Remote Sensing
18	of Environment 275:113024.
19	Helfenstein, I. S., J. T. Sturm, B. Schmid, A. Damm, M. C. Schuman, and F. Morsdorf. 2024. Satellite
20	observations reveal positive relationship between trait-based diversity and drought response in
21	temperate forests. https://doi.org/10.32942/X24619
22	Hong, D., C. Li, B. Zhang, N. Yokoya, J. A. Benediktsson, and J. Chanussot. 2024. Multimodal artificial
23	intelligence foundation models: Unleashing the power of remote sensing big data in earth
24	observation. The Innovation Geoscience 2:100055-2.

1	Housset, J. M., S. Nadeau, N. Isabel, C. Depardieu, I. Duchesne, P. Lenz, and M. P. Girardin. 2018. Tree
2	rings provide a new class of phenotypes for genetic associations that foster insights into adaptation
3	of conifers to climate change. New Phytologist 218:630-645.
4	Huang, Y., Y. Chen, N. Castro-Izaguirre, M. Baruffol, M. Brezzi, A. Lang, Y. Li, W. Härdtle, G. von
5	Oheimb, X. Yang, X. Liu, K. Pei, S. Both, B. Yang, D. Eichenberg, T. Assmann, J. Bauhus, T.
6	Behrens, F. Buscot, XY. Chen, D. Chesters, BY. Ding, W. Durka, A. Erfmeier, J. Fang, M.
7	Fischer, LD. Guo, D. Guo, J. L. M. Gutknecht, JS. He, CL. He, A. Hector, L. Hönig, RY. Hu,
8	AM. Klein, P. Kühn, Y. Liang, S. Li, S. Michalski, M. Scherer-Lorenzen, K. Schmidt, T. Scholten,
9	A. Schuldt, X. Shi, MZ. Tan, Z. Tang, S. Trogisch, Z. Wang, E. Welk, C. Wirth, T. Wubet, W.
10	Xiang, M. Yu, XD. Yu, J. Zhang, S. Zhang, N. Zhang, HZ. Zhou, CD. Zhu, L. Zhu, H.
11	Bruelheide, K. Ma, P. A. Niklaus, and B. Schmid. 2018. Impacts of species richness on productivity
12	in a large-scale subtropical forest experiment. Science 362 :80-83.
13	Isbell, F., D. Craven, J. Connolly, M. Loreau, B. Schmid, C. Beierkuhnlein, T. M. Bezemer, C. Bonin, H.
14	Bruelheide, E. de Luca, A. Ebeling, J. N. Griffin, Q. Guo, Y. Hautier, A. Hector, A. Jentsch, J.
15	Kreyling, V. Lanta, P. Manning, S. T. Meyer, A. S. Mori, S. Naeem, P. A. Niklaus, H. W. Polley, P.
16	B. Reich, C. Roscher, E. W. Seabloom, M. D. Smith, M. P. Thakur, D. Tilman, B. F. Tracy, W. H.
17	van der Putten, J. van Ruijven, A. Weigelt, W. W. Weisser, B. Wilsey, and N. Eisenhauer. 2015.
18	Biodiversity increases the resistance of ecosystem productivity to climate extremes. Nature 526:574-
19	577.
20	Jacquemoud, S., and S. Ustin. 2019. Leaf Optical Properties. Cambridge University Press, New York.
21	Jetz, W., J. Cavender-Bares, R. Pavlick, D. Schimel, F. W. Davis, G. P. Asner, R. Guralnick, J. Kattge, A.
22	M. Latimer, P. Moorcroft, M. E. Schaepman, M. P. Schildhauer, F. D. Schneider, F. Schrodt, U.
23	Stahl, and S. L. Ustin. 2016. Monitoring plant functional diversity from space. Nature Plants
24	2:16024.
25	Juzwik, J., D. N. Appel, W. L. macDonald, and S. Burks. 2011. Challenges and success in managing oak

wilt in the United States. Plant Disease 95:888-900.

1	Kamoske, A. G., K. M. Dahlin, Q. D. Read, S. Record, S. C. Stark, S. P. Serbin, and P. L. Zarnetske.
2	2022. Towards mapping biodiversity from above: Can fusing lidar and hyperspectral remote sensing
3	predict taxonomic, functional, and phylogenetic tree diversity in temperate forests? Global Ecology
4	and Biogeography 31:1440–1460.
5	Kebabian, P. L., A. F. Theisen, S. Kallelis, and A. Freedman. 1999. A passive two-band sensor of
6	sunlight-excited plant fluorescence. Review of Scientific Instruments 70:4386-4393.
7	Kissling, W. D., J. A. Ahumada, A. Bowser, M. Fernandez, N. Fernández, E. A. García, R. P. Guralnick,
8	N. J. B. Isaac, S. Kelling, W. Los, L. McRae, JB. Mihoub, M. Obst, M. Santamaria, A. K.
9	Skidmore, K. J. Williams, D. Agosti, D. Amariles, C. Arvanitidis, L. Bastin, F. De Leo, W. Egloff, J.
10	Elith, D. Hobern, D. Martin, H. M. Pereira, G. Pesole, J. Peterseil, H. Saarenmaa, D. Schigel, D. S.
11	Schmeller, N. Segata, E. Turak, P. F. Uhlir, B. Wee, and A. R. Hardisty. 2018. Building essential
12	biodiversity variables (EBVs) of species distribution and abundance at a global scale. Biological
13	Reviews 93:600-625.
14	Koch, K. A., G. L. Quiram, and R. C. Venette. 2010. A review of oak wilt management: A summary of
15	treatment options and their efficacy. Urban Forestry & Urban Greening 9:1-8.
16	Kothari, S., R. A. Montgomery, and J. Cavender-Bares. 2021. Physiological responses to light explain
17	competition and facilitation in a tree diversity experiment. Journal of Ecology 109:2000-2018.
18	Kothari, S., and A. K. Schweiger. 2022. Plant spectra as integrative measures of plant phenotypes.
19	Li, C., E. A. Czyż, R. Halitschke, I. T. Baldwin, M. E. Schaepman, and M. C. Schuman. 2023. Evaluating
20	potential of leaf reflectance spectra to monitor plant genetic variation. Plant Methods 19:108.
21	Liang, J., B. Lee, M. R. Ngugi, F. Rovero, B. Herault, B. Schmid, M. J. Schelhaas, N. Tchebakova, D.
22	Verbyla, M. Zhou, A. M. Jagodzinski, G. Alberti, H. Bruelheide, T. Jucker, T. W. Crowther, S.
23	Brandl, S. L. Lewis, P. Alvarez-Loayza, P. Schall, R. Bitariho, R. M. Vasquez, B. Jaroszewicz, A. R.
24	Marshall, N. Chamuya, G. M. Hengeveld, D. Gianelle, J. Vayreda, F. Kitahara, F. Mortier, N. L.
25	Engone-Obiang, R. Valencia, W. Marthy, H. B. Glick, H. Y. H. Chen, H. Lu, B. Sonke, C. Baraloto,
26	V. Wortel, D. E. Odeke, E. D. Schulze, F. Valladares, T. Zawi a-Nied wiecki, V. J. Neldner, R. Ba

1	azy, E. B. Searle, E. H. Martin, L. Finer, S. Wiser, X. Lei, A. D. McGuire, S. Gourlet-Fleury, D.
2	Piotto, J. Oleksyn, T. Obrien, F. Bussotti, S. Pfautsch, A. Paquette, M. Scherer-Lorenzen, O.
3	Bouriaud, J. Zhu, E. I. Parfenova, R. Tavani, J. V. Watson, H. Viana, H. Pretzsch, C. B. Barrett, P.
4	A. Niklaus, M. Fischer, D. A. Coomes, F. Bozzato, A. C. Vibrans, C. Ammer, L. Frizzera, T.
5	Sunderland, G. J. Nabuurs, C. Salas, P. L. Peri, L. V. Ferreira, C. Gonmadje, N. Picard, H. S. Kim,
6	P. B. Reich, E. Lee, and S. De-Miguel. 2016. Positive biodiversity-productivity relationship
7	predominant in global forests. Science 354:aaf8957-aaf8957.
8	Liu, M., X. Liu, L. Wu, Y. Tang, Y. Li, Y. Zhang, L. Ye, and B. Zhang. 2021. Establishing forest
9	resilience indicators in the hilly red soil region of southern China from vegetation greenness and
10	landscape metrics using dense Landsat time series. Ecological Indicators 121:106985.
11	Liu, X., Y. Su, T. Hu, Q. Yang, B. Liu, Y. Deng, H. Tang, Z. Tang, J. Fang, and Q. Guo. 2022. Neural
12	network guided interpolation for mapping canopy height of China's forests by integrating GEDI and
13	ICESat-2 data. Remote Sensing of Environment 269:112844.Loreau
14	Liu, Y., J. A. Hogan, J. W. Lichstein, R. P. Guralnick, D. E. Soltis, P. S. Soltis, and S. M. Scheiner. 2024.
15	Biodiversity and productivity in eastern US forests. Proceedings of the National Academy of
16	Sciences 121:e2314231121.
17	Loreau, M., and C. de Mazancourt. 2013. Biodiversity and ecosystem stability: a synthesis of underlying
18	mechanisms. Ecology Letters 16:106-115.
19	Madritch, M., J. Cavender-Bares, S. E. Hobbie, and P. A. Townsend. 2020. Linking Foliar Traits to
20	Belowground Processes. Pages 173-197 in J. Cavender-Bares, J. A. Gamon, and P. A. Townsend,
21	editors. Remote Sensing of Plant Biodiversity. Springer International Publishing, Cham.
22	Madritch, M. D., C. C. Kingdon, A. Singh, K. E. Mock, R. L. Lindroth, and P. A. Townsend. 2014.
23	Imaging spectroscopy links aspen genotype with below-ground processes at landscape scales.
24	Philosophical Transactions of the Royal Society B: Biological Sciences 369:20130194.

1	Magney, T. S., C. Frankenberg, J. B. Fisher, Y. Sun, G. B. North, T. S. Davis, A. Kornfeld, and K.
2	Siebke. 2017. Connecting active to passive fluorescence with photosynthesis: a method for
3	evaluating remote sensing measurements of Chl fluorescence. New Phytologist 215:1594–1608.
4	McDowell, N. G., G. Sapes, A. Pivovaroff, H. D. Adams, C. D. Allen, W. R. Anderegg, M. Arend, D. D.
5	Breshears, T. Brodribb, and B. Choat. 2022. Mechanisms of woody-plant mortality under rising
6	drought, CO ₂ and vapour pressure deficit. Nature Reviews Earth & Environment 3:294-308.
7	Meireles, J. E., J. Cavender-Bares, P. A. Townsend, S. Ustin, J. A. Gamon, A. K. Schweiger, M. E.
8	Schaepman, G. P. Asner, R. E. Martin, A. Singh, F. Schrodt, A. Chlus, and B. O'Meara. 2020. Leaf
9	reflectance spectra capture the evolutionary history of seed plants. New Phytologist 228:485-493.
10	Mohammed, G. H., R. Colombo, E. M. Middleton, U. Rascher, C. van der Tol, L. Nedbal, Y. Goulas, O.
11	Pérez-Priego, A. Damm, M. Meroni, J. Joiner, S. Cogliati, W. Verhoef, Z. Malenovský, JP.
12	Gastellu-Etchegorry, J. R. Miller, L. Guanter, J. Moreno, I. Moya, J. A. Berry, C. Frankenberg, and
13	P. J. Zarco-Tejada. 2019. Remote sensing of solar-induced chlorophyll fluorescence (SIF) in
14	vegetation: 50 years of progress. Remote Sensing of Environment 231:111177.
15	Mohd Asaari, M. S., P. Mishra, S. Mertens, S. Dhondt, D. Inzé, N. Wuyts, and P. Scheunders. 2018.
16	Close-range hyperspectral image analysis for the early detection of stress responses in individual
17	plants in a high-throughput phenotyping platform. ISPRS Journal of Photogrammetry and Remote
18	Sensing 138:121–138.
19	Morales, F., R. Belkhodja, Y. Goulas, J. Abadia, and I. Moya. 1999. Remote and near-contact chlorophyll
20	fluorescence during photosynthetic induction in iron-deficient sugar beet leaves. Remote Sensing of
21	Environment 69:170-178.
22	Mori, A. S., L. E. Dee, A. Gonzalez, H. Ohashi, J. Cowles, A. J. Wright, M. Loreau, Y. Hautier, T.
23	Newbold, and P. B. Reich. 2021. Biodiversity-productivity relationships are key to nature-based
24	climate solutions. Nature Climate Change 11:543-550.
25	Moya, I., and Z. Cerovic. 2004. Remote sensing of chlorophyll fluorescence: Instrumentation and
26	analysis. Springer, Dordrecht, The Netherlands.

1	O'Connor, M. I., A. S. Mori, A. Gonzalez, L. E. Dee, M. Loreau, M. Avolio, J. E. K. Byrnes, W. Cheung,
2	J. Cowles, A. T. Clark, Y. Hautier, A. Hector, K. Komatsu, T. Newbold, C. L. Outhwaite, P. B.
3	Reich, E. Seabloom, L. Williams, A. Wright, and F. Isbell. 2021. Grand challenges in biodiversity-
4	ecosystem functioning research in the era of science-policy platforms require explicit consideration
5	of feedbacks. Proceedings of the Royal Society B: Biological Sciences 288:20210783.
6	Oehri, J., B. Schmid, G. Schaepman-Strub, and P. A. Niklaus. 2017. Biodiversity promotes primary
7	productivity and growing season lengthening at the landscape scale. Proceedings of the National
8	Academy of Sciences 114:10160-10165.
9	Oehri, J., B. Schmid, G. Schaepman-Strub, and P. A. Niklaus. 2020. Terrestrial land-cover type richness
10	is positively linked to landscape-level functioning. Nature Communications 11:154.
11	Papeş, M., R. Tupayachi, P. Martínez, A. T. Peterson, and G. V. N. Powell. 2010. Using hyperspectral
12	satellite imagery for regional inventories: a test with tropical emergent trees in the Amazon Basin.
13	Journal of Vegetation Science 21:342-354.
14	Plourde, L. C., S. V. Ollinger, ML. Smith, and M. E. Martin. 2007. Estimating Species Abundance in a
15	Northern Temperate Forest Using Spectral Mixture Analysis. Photogrammetric Engineering &
16	Remote Sensing 73:829-840.
17	Pontius, J., and R. Hallett. 2014. Comprehensive Methods for Earlier Detection and Monitoring of Forest
18	Decline. Forest Science 60:1156-1163.
19	Pontius, J., M. Martin, L. Plourde, and R. Hallett. 2008. Ash decline assessment in emerald ash borer-
20	infested regions: A test of tree-level, hyperspectral technologies.
21	Pontius, J., P. Schaberg, and R. Hanavan. 2020. Remote sensing for early, detailed, and accurate detection
22	of forest disturbance and decline for protection of biodiversity. Springer International Publishing,
23	Cham. Remote Sensing of Environment 112:2665-2676.
24	Porcar-Castell, A., Z. Malenovský, T. Magney, S. Van Wittenberghe, B. Fernández-Marín, F. Maignan,
25	Y. Zhang, K. Maseyk, J. Atherton, L. P. Albert, T. M. Robson, F. Zhao, JI. Garcia-Plazaola, I.
26	Ensminger, P. A. Rajewicz, S. Grebe, M. Tikkanen, J. R. Kellner, J. A. Ihalainen, U. Rascher, and B.

- Logan. 2021. Chlorophyll a fluorescence illuminates a path connecting plant molecular biology to
 Earth-system science. Nature Plants 7:998-1009.
- 3 Queally, N., Z. Ye, T. Zheng, A. Chlus, F. Schneider, R. P. Pavlick, and P. A. Townsend. 2022. 4 FlexBRDF: A Flexible BRDF Correction for Grouped Processing of Airborne Imaging Spectroscopy 5 Flightlines. Journal of Geophysical Research: Biogeosciences 127:e2021JG006622. 6 Rai, G. K., D. M. Khanday, S. M. Choudhary, P. Kumar, S. Kumari, C. Martínez-Andújar, P. A. 7 Martínez-Melgarejo, P. K. Rai, and F. Pérez-Alfocea. 2024. Unlocking nature's stress buster: 8 Abscisic acid's crucial role in defending plants against abiotic stress. Plant Stress 11:100359. 9 Ramirez-Valiente, J. A., K. Koehler, and J. Cavender-Bares. 2015. Climatic origins predict variation in 10 photoprotective leaf pigments in response to drought and low temperatures in live oaks (Quercus 11 series Virentes). Tree Physiology 35:521-534. 12 Reich, P. B., J. Knops, D. Tilman, J. Craine, D. Ellsworth, M. Tjoelker, T. Lee, D. Wedin, S. Naeem, D. 13 Bahauddin, G. Hendrey, S. Jose, K. Wrage, J. Goth, and W. Bengston. 2001. Plant diversity 14 enhances ecosystem responses to elevated CO2 and nitrogen deposition. Nature 410:809-812. 15 Roberts, D. A., M. Gardner, R. Church, S. Ustin, G. Scheer, and R. O. Green. 1998. Mapping chaparral in 16 the Santa Monica Mountains using multiple endmember spectral mixture models. Remote Sensing of 17 Environment 65:267-279. 18 Rocchini, D., N. Balkenhol, G. A. Carter, G. M. Foody, T. W. Gillespie, K. S. He, S. Kark, N. Levin, K. 19 Lucas, and M. Luoto. 2010. Remotely sensed spectral heterogeneity as a proxy of species diversity: 20 recent advances and open challenges. Ecological Informatics 5:318-329. 21 Rocchini, D., M. J. Santos, S. L. Ustin, J.-B. Féret, G. P. Asner, C. Beierkuhnlein, M. Dalponte, H. 22 Feilhauer, G. M. Foody, G. N. Geller, T. W. Gillespie, K. S. He, D. Kleijn, P. J. Leitão, M. 23 Malavasi, V. Moudrý, J. Müllerová, H. Nagendra, S. Normand, C. Ricotta, M. E. Schaepman, S. 24 Schmidtlein, A. K. Skidmore, P. Šímová, M. Torresani, P. A. Townsend, W. Turner, P. Vihervaara,
- 25 M. Wegmann, and J. Lenoir. 2022. The Spectral Species Concept in Living Color. Journal of
- 26 Geophysical Research: Biogeosciences 127:e2022JG007026.
 - 40

1	Rossi, C., M. Kneubühler, M. Schütz, M. E. Schaepman, R. M. Haller, and A. C. Risch. 2022. Spatial
2	resolution, spectral metrics and biomass are key aspects in estimating plant species richness from
3	spectral diversity in species-rich grasslands. Remote Sensing in Ecology and Conservation 8:297-
4	314.
5	Saarela, S., S. Holm, S. P. Healey, HE. Andersen, H. Petersson, W. Prentius, P. L. Patterson, E. Næsset,
6	T. G. Gregoire, and G. Ståhl. 2018. Generalized hierarchical model-based estimation for
7	aboveground biomass assessment using GEDI and Landsat data. Remote Sensing 10:1832.
8	Sapes, G., C. Lapadat, A. K. Schweiger, J. Juzwik, R. Montgomery, H. Gholizadeh, P. A. Townsend, J.
9	A. Gamon, and J. Cavender-Bares. 2022. Canopy spectral reflectance detects oak wilt at the
10	landscape scale using phylogenetic discrimination. Remote Sensing of Environment 273:112961.
11	Sapes, G., L. Schroeder, A. Scott, I. Clark, J. Juzwik, R. A. Montgomery, J. A. Guzmán Q, and J.
12	Cavender-Bares. 2024. Mechanistic links between physiology and spectral reflectance enable
13	previsual detection of oak wilt and drought stress. Proceedings of the National Academy of Sciences
14	121:e2316164121.
15	Sato, H., J. Mizoi, K. Shinozaki, and K. Yamaguchi-Shinozaki. 2024. Complex plant responses to
16	drought and heat stress under climate change. The Plant Journal 117:1873-1892.
17	Savage, J., J. Cavender-Bares, and A. Verhoeven. 2009. Habitat generalists and wetland specialists in the
18	genus Salix vary in their photoprotective responses to drought. Functional Plant Biology 36:300-309.
19	Scheffer, M., S. Carpenter, J. A. Foley, C. Folke, and B. Walker. 2001. Catastrophic shifts in ecosystems.
20	Nature 413:591-596.
21	Scheffer, M., and S. R. Carpenter. 2003. Catastrophic regime shifts in ecosystems: linking theory to
22	observation. Trends in Ecology & Evolution 18:648–656.
23	Schimel, D., P. A. Townsend, and R. Pavlick. 2020. Prospects and pitfalls for spectroscopic remote
24	sensing of biodiversity at the global scale. Springer, New York.

1	Schneider, F. D., A. Ferraz, S. Hancock, L. I. Duncanson, R. O. Dubayah, R. P. Pavlick, and D. S.
2	Schimel. 2020. Towards mapping the diversity of canopy structure from space with GEDI.
3	Environmental Research Letters 15:11500
4	Schneider, F. D., F. Morsdorf, B. Schmid, O. L. Petchey, A. Hueni, D. S. Schimel, and M. E. Schaepman.
5	2017. Mapping functional diversity from remotely sensed morphological and physiological forest
6	traits. Nature Communications 8:1441.
7	Schreiber, U., W. Bilger, and C. Neubauer. 1994. Chlorophyll fluorescence as a noninvasive indicator for
8	rapid assessment of in vivo photosynthesis. Springer-Verlag, Berlin.
9	Schuldt, A., X. Liu, F. Buscot, H. Bruelheide, A. Erfmeier, JS. He, AM. Klein, K. Ma, M. Scherer-
10	Lorenzen, B. Schmid, T. Scholten, Z. Tang, S. Trogisch, C. Wirth, T. Wubet, and M. Staab. 2023.
11	Carbon-biodiversity relationships in a highly diverse subtropical forest. Global Change Biology
12	29:5321–5333.
13	Schweiger, A. K., J. Cavender-Bares, P. A. Townsend, S. E. Hobbie, M. D. Madritch, R. Wang, D.
14	Tilman, and J. A. Gamon. 2018. Plant spectral diversity integrates functional and phylogenetic
15	components of biodiversity and predicts ecosystem function. Ecology & Evolution 2:976-982.
16	Seeley, M. M., E. A. Stacy, R. E. Martin, and G. P. Asner. 2023. Foliar functional and genetic variation in
17	a keystone Hawaiian tree species estimated through spectroscopy. Oecologia 202:15-28.
18	Sellers, P., D. Randall, G. Collatz, J. Berry, C. Field, D. Dazlich, C. Zhang, G. Collelo, and L. Bounoua.
19	1996. A revised land surface parameterization (SIB2) for atmospheric GCMs. Part 1. Model
20	formulation. Journal of Climate 9:676-705.
21	Serbin, S. P., A. Singh, A. R. Desai, S. G. Dubois, A. D. Jablonski, C. C. Kingdon, E. L. Kruger, and P.
22	A. Townsend. 2015. Remotely estimating photosynthetic capacity, and its response to temperature,
23	in vegetation canopies using imaging spectroscopy. Remote Sensing of Environment 167:78-87.
24	Simms, É. L., and H. Ward. 2013. Multisensor NDVI-Based Monitoring of the Tundra-Taiga Interface
25	(Mealy Mountains, Labrador, Canada). Remote Sensing 5:1066-1090.

1	Skidmore, A. K., N. C. Coops, E. Neinavaz, A. Ali, M. E. Schaepman, M. Paganini, W. D. Kissling, P.
2	Vihervaara, R. Darvishzadeh, H. Feilhauer, M. Fernandez1, N. Fernández, N. Gorelick, I.
3	Geizendorffer, U. Heiden, M. Heurich, D. Hobern, S. Holzwarth, F. E. Muller-Karger, R. V. D.
4	Kerchove, A. Lausch, P. J. Leitãu, M. C. Lock, C. A. Mücher, B. O'Connor, D. Rocchini, W.
5	Turner, JK. Vis, T. Wang, M. Wegmann, and V. Wingate. 2021. Priority list of biodiversity metrics
6	to observe from space. Nature Ecology & Evolution 5:896–906.
7	Stasinski, L., D. M. White, P. R. Nelson, R. H. Ree, and J. E. Meireles. 2021. Reading light: leaf spectra
8	capture fine-scale diversity of closely related, hybridizing arctic shrubs. New Phytologist 232:2283-
9	2294.
10	Stefanski A, E Butler, L Williams, R Bermudez, JAQ Guzmán, A Larson, P Townsend, R Montgomery, J
11	Cavender-Bares, PB Reich. All the light we cannot see: Climate manipulations leave short and long-
12	term imprints in spectral reflectance of trees. Ecology (in press, Special Feature on Spectral
13	Ecology).
14	Steffen, W., K. Richardson, J. Rockström, S. E. Cornell, I. Fetzer, E. M. Bennett, R. Biggs, S. R.
15	Carpenter, W. de Vries, C. A. de Wit, C. Folke, D. Gerten, J. Heinke, G. M. Mace, L. M. Persson, V.
16	Ramanathan, B. Reyers, and S. Sörlin. 2015. Planetary boundaries: Guiding human development on
17	a changing planet. Science 347.
18	Sturm, J., M. J. Santos, B. Schmid, and A. Damm. 2022. Satellite data reveal differential responses of
19	Swiss forests to unprecedented 2018 drought. Global Change Biology 28:2956–2978.
20	Sun, Y., C. Frankenberg, M. Jung, J. Joiner, L. Guanter, P. Köhler, and T. Magney. 2018. Overview of
21	Solar-Induced chlorophyll Fluorescence (SIF) from the Orbiting Carbon Observatory-2: Retrieval,
22	cross-mission comparison, and global monitoring for GPP. Remote Sensing of Environment
23	209:808-823.
24	Tai, X., A. T. Trugman, and W. R. L. Anderegg. 2023. Linking remotely sensed ecosystem resilience
25	with forest mortality across the continental United States. Global Change Biology 29:1096–1105.

1	Tilman, D. 1999. The ecological consequences of changes in biodiversity: A search for general principles
2	Ecology 80:1455-1474.

- Tirado, S. B., S. S. Dennis, T. A. Enders, and N. M. Springer. 2021. Utilizing top-down hyperspectral
 imaging for monitoring genotype and growth conditions in maize. he Plant Phenome Journal
 4:e20013.
- 6 Torabzadeh, H., R. Leiterer, A. Hueni, M. E. Schaepman, and F. Morsdorf. 2019. Tree species
- classification in a temperate mixed forest using a combination of imaging spectroscopy and airborne
 laser scanning. Agricultural and Forest Meteorology 279:107744.
- 9 Townsend, P. A., S. P. Serbin, E. L. Kruger, and J. A. Gamon. 2013. Disentangling the contribution of

10 biological and physical properties of leaves and canopies in imaging spectroscopy data. Proceedings

- 11 of the National Academy of Sciences 110:E1074.
- 12 Wagner, W., A. Ullrich, V. Ducic, T. Melzer, and N. Studnicka. 2006. Gaussian decomposition and

13 calibration of a novel small-footprint full-waveform digitising airborne laser scanner. ISPRS Journal

14 of Photogrammetry and Remote Sensing 60:100-112.

Wang, L., Y. Duan, L. Zhang, J. Wang, Y. Li, and J. Jin. 2020a. LeafScope: A Portable High-Resolution
Multispectral Imager for In Vivo Imaging Soybean Leaf. Sensors 20:2194.

- Wang, R., and J. A. Gamon. 2019. Remote sensing of terrestrial plant biodiversity. Remote Sensing of
 Environment 231:111218.
- Wang, R., J. A. Gamon, J. Cavender-Bares, P. A. Townsend, and A. I. Zygielbaum. 2018. The spatial
 sensitivity of the spectral diversity-biodiversity relationship: an experimental test in a prairie
- 21 grassland. Ecological Applications 28:541-556.
- 22 Wang, Z., A. Chlus, R. Geygan, Z. Ye, T. Zheng, A. Singh, J. Couture, J. Cavender-Bares, E. Kruger, and
- P. Townsend. 2020b. Foliar functional traits from imaging spectroscopy across biomes in the eastern
- 24 North America. New Phytologist 228:494-511.

1	Wang, Z., P. A. Townsend, A. K. Schweiger, J. J. Couture, A. Singh, S. E. Hobbie, and J. Cavender-
2	Bares. 2019. Mapping foliar functional traits and their uncertainties across three years in a grassland
3	experiment. Remote Sensing of Environment 221:405-416.
4	Wei, C., D. N. Karger, and A. M. Wilson. 2020. Spatial detection of alpine treeline ecotones in the
5	Western United States. Remote Sensing of Environment 240:111672.
6	Weisser, W. W., C. Roscher, S. T. Meyer, A. Ebeling, G. Luo, E. Allan, H. Beßler, R. L. Barnard, N.
7	Buchmann, F. Buscot, C. Engels, C. Fischer, M. Fischer, A. Gessler, G. Gleixner, S. Halle, A.
8	Hildebrandt, H. Hillebrand, H. de Kroon, M. Lange, S. Leimer, X. Le Roux, A. Milcu, L. Mommer,
9	P. A. Niklaus, Y. Oelmann, R. Proulx, J. Roy, C. Scherber, M. Scherer-Lorenzen, S. Scheu, T.
10	Tscharntke, M. Wachendorf, C. Wagg, A. Weigelt, W. Wilcke, C. Wirth, ED. Schulze, B. Schmid,
11	and N. Eisenhauer. 2017. Biodiversity effects on ecosystem functioning in a 15-year grassland
12	experiment: Patterns, mechanisms, and open questions. Basic and Applied Ecology 23:1-73.
13	Wilcox, K. R., S. E. Koerner, D. L. Hoover, A. K. Borkenhagen, D. E. Burkepile, S. L. Collins, A. M.
14	Hoffman, K. P. Kirkman, A. K. Knapp, T. Strydom, D. I. Thompson, and M. D. Smith. 2020. Rapid
15	recovery of ecosystem function following extreme drought in a South African savanna grassland.
16	Ecology 101:e02983.
17	Williams, L. J., J. Cavender-Bares, P. A. Townsend, J. J. Couture, Z. Wang, A. Stefanski, C. Messier, and
18	P. B. Reich. 2020. Remote spectral detection of biodiversity effects on forest biomass. Nature
19	Ecology & Evolution 5:46-54.
20	Xi, Y., Zhang, W., Wei, F. et al. Boreal tree species diversity increases with global warming but is
21	reversed by extremes. Nat. Plants (2024). https://doi.org/10.1038/s41477-024-01794-w
22	Xu, Q., X. Yang, J. Song, J. Ru, J. Xia, S. Wang, S. Wan, and L. Jiang. 2022. Nitrogen enrichment alters
23	multiple dimensions of grassland functional stability via changing compositional stability. Ecology
24	Letters 25:2713–2725.
25	Yang, X., M. Lu, Y. Wang, Y. Wang, Z. Liu, and S. Chen. 2021. Response Mechanism of Plants to

26 Drought Stress. Horticulturae. 7:50.

1	Yi, C., and N. Jackson. 2021. A review of measuring ecosystem resilience to disturbance. Environmental
2	Research Letters 16:053008.
3	Zhang, X., K. Liu, X. Li, S. Wang, and J. Wang. 2022. Vulnerability assessment and its driving forces in
4	terms of NDVI and GPP over the Loess Plateau, China. Physics and Chemistry of the Earth, Parts
5	A/B/C 125:103106.
6	
7	
8	
9	
10	
11	

1 Figure Legends

3	Fig. 1. Capturing biological variation across spatial and temporal scales to understand ecological and evolutionary
4	processes in changing environments. Shown are biological scales of measurement (circles) from genes and leaves to
5	the biosphere and some of the tools of spectral biology that capture optical information across these scales.
6	Spectroscopy, SIF and LiDAR from satellite, aircraft, UAVs, towers and hand-held instruments showing remote,
7	proximal and in-situ sensors that capture plant foliar chemistry, structure and function, photosynthesis and
8	productivity, and vegetation height and structure. The figure emphasizes the visual to the short-wave infrared
9	(VSWIR) solar domain (400-2500 nm), but the UV (100 - 400 nm), thermal emission (3 – 14 μ m) as well as active
10	and passive microwave (0.1-1m) domains provide critical information, for example about light quality, ozone and
11	SO ₂ ; land surface temperature, water content and flux; and soil water content or atmospheric water and ozone
12	content, respectively. Towers in a fixed location close to focal observation sites can support Phenocams, continuous
13	spectroscopic measurements, terrestrial laser scanning, and other sensor types. Combined with ground-based
14	measures and understanding of biological processes, spectral biology can contribute to measuring and understanding
15	life's variation (biodiversity components at any scale), ecological and evolutionary processes and their emergent
16	properties, and how they are changing with global environmental forces.



Fig. 2. Number of publications listed within Web of Science over time from 1978 to 2024 with the
 queries 'ecology and spectroscopy or hyperpsectral' (black), 'ecology and remote sensing' (orange),
 'ecology and SIF' (green), and 'ecology and LiDAR' (brown).



1	Fig. 3. Spectral biology is defined as the interaction of electromagnetic energy, shown for (A), with
2	biological systems to reveal patterns and processes, such as (B) chlorophyll fluorescence emission
3	(middle) and (C) reflectance from plant tissues (bottom). A) Solar irradiance at the top of the atmosphere
4	(gray) and the sun's energy that penetrates the atmosphere to reach the Earth's surface (red) falls mostly
5	within the range of 250-2500 nm, spanning the ultraviolet (UV), visible range (VIS), near-infrared (NIR),
6	and two short-wave infrared regions (SWIR1, SWIR2). Plants absorb energy primarily in the red and blue
7	wavelengths for photosynthesis and re-emit a small fraction of the energy as chlorophyll fluorescence (B)
8	within the range of 625 to 800 nm, with peak emission shown at 737 (red vertical line). Solar-induced
9	fluorescence (SIF) can be differentiated from solar irradiance within features such as the O ₂ A band, where
10	oxygen absorbs (vertical blue band), providing a means to detect photosynthesis. Satellite sensors
11	designed to retrieve SIF capture emission within the range of 758–771 nm, indicated by the curly bracket,
12	taking advantage of the O ₂ A band. Different parts of the chlorophyll emission spectrum are used by
13	different sensors, depending on distance from the vegetation and depth of the atmosphere. C) Spectral
14	reflectance of fresh (green) and dried (brown) leaf tissue include features from the visible to the short-
15	wave infrared that are informative for predicting plant functional traits (e.g., leaf mass per area, LMA),
16	indicated as dotted lines. Reflectance spectra (solid curves) show the percent of incoming light reflected
17	at each wavelength within the VIS, NIR and SWIR1 and SWIR2.
18	
19	
20	
21	



1 Fig. 4. Three critical scaling hierarchies in spectral biology. Left: the physiological hierarchy encompasses how 2 functions are expressed within nested levels of organization from genes, to molecules, organelles, cells, tissues 3 (leaves) and the whole organism. Middle: the evolutionary hierarchy captures the fractal nature of the tree of life 4 based on shared ancestry, where variation among individuals is nested within populations, which are in turn nested 5 within species, and within clades of larger and larger size. Right: the macroecological hierarchy traverses the 6 ecological processes that shift with spatial and temporal scales, shown here spanning the density-dependent 7 interactions of individual trees, environmental filtering that sorts species based on niche preferences operating at the 8 scale of critical environmental gradients, dispersal processes driven by migration and propagule movement, and the 9 biogeographic and macroevolutionary processes that operate at deeper temporal and larger spatial scales. 10 Three critical hierarchies in biology: 1) the physiological hierarchy with nested biological components from DNA to 11 the whole organism, 2) the evolutionary hierarchy where variation among individuals is nested within populations, 12 which are in turn nested within species and increasingly larger clades across the tree of life, and 3) a 13 macroecological hierarchy in which ecological processes shift with spatial and temporal scale from density-14 dependent processes that involve organismal and species interactions in local environments, to environmental 15 sorting and dispersal and migration processes at landscape scales, to long-term biogeographic and evolutionary 16 processes at continental scales that extend deep in time. A typical spatial resolution (grain size) is shown below each 17 spectral image associated with the different spatial scales. This figure is adapted with permission from Cavender-18 Bares et al. (2021).





1 Fig. 5. Using a common data type (spectral reflectance) across evolutionary (A) and macroecological scaling hierarchies (B). A) Phylogenetic signal across wavelengths and phylogenetic scales from seed 2 3 plants to an adaptive radiation within a single genus (*Quercus*, the oaks) to populations within a single 4 species. Phylogenetic relationships and spectra from fresh leaves are shown for species across the seed 5 plants (bottom), for species of the oak genus Quercus (middle), and for the variation among individuals within populations of a single species (top). A filled red circle for a given wavelength indicates that close 6 7 relatives have a more similar normalized spectral reflectance value than expected at random. Data are 8 redrawn from Meireles et al. 2020 and Cavender-Bares et al. 2016. B) A range of instruments from 9 handheld devices, uncrewed aerial vehicles (UAV), aircraft and satellites capture reflectance spectra and 10 image cubes of vegetation reflectance at every biological scale. Spectral reflectance from different 11 platforms has the potential to advance ecological integration across spatial and temporal scales.







1 Fig. 6. Key realms for advancement in spectral biology. The realms are conceptual and nested. A) Plant identity, diversity, and composition as well as plant and ecosystem structure and function can be 2 3 spectrally detected in ambient steady state conditions using vegetation spectra, SIF and/or LiDAR. B) The 4 average responses of ecosystems to global change and environmental stress can also be detected 5 spectrally, across space, time or experimental treatments. C) Differences over time can further be used to 6 understand the dynamics of ecosystem responses to change, including their resistance and capacity to 7 recover from disturbance, both of which help capture the nature and underlying mechanisms of resilience 8 of ecosystems.

