

1 Land use gradients drive spatial variation in 2 Lassa fever host communities in the 3 Eastern Province of Sierra Leone.

4 Author details.

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31 Conflict of interest

32 All authors report no financial conflicts of interest.

33 Authors' contributions

34 DS, RG, EF-C, DW-J, RK and KEJ conceived the ideas and designed the methodology. DS,
35 UB, DiS, JL, JK, MJ, MD, JoL and RA collected the data. DS and RG analysed the data. DS,
36 RG and KEJ interpreted the analysis. DS, UB, EF-C, DW-J, RK and KEJ provided supervision
37 and resources to conduct the study. DS led the writing of the manuscript. RG, DW-J, RK
38 and KEJ contributed critically to the drafts. All authors gave final approval for publication.

39 Statement on inclusion

40 This study brings together authors from several countries, including scientists based in the
41 country where the study was carried out. All authors were engaged early on with the
42 research and study design to ensure that the diverse sets of perspectives they represent
43 was considered. Literature published by scientists from the study country and wider region
44 was cited. Study protocols were discussed with local scientists for appropriateness of
45 design. Consultations were held with community leaders of the study villages and the
46 wider region before finalising the study design and enrolment of sites into the study.

47 Data availability

48 Data on detected rodent individuals are available on PHAROS
49 (<https://pharos.viralemergence.org/projects/?prj=prjyg91YQvrnk>).

50 All R scripts and data to reproduce the analysis are available on GitHub
51 (<https://github.com/DidDrog11/land-use-lassa-hosts>).

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53

54 Abstract

- 55 1. The natal multimammate mouse (*Mastomys natalensis*) is the primary reservoir
56 host of *Lassa marmarenavirus* (LASV), a zoonotic pathogen causing Lassa fever
57 that is endemic to West Africa. The occurrence and abundance of this species is
58 regulated by the human environment and biotic interactions with other small-
59 mammal species, but these ecological drivers remain poorly understood in the
60 regions where Lassa fever outbreaks are observed.
- 61 2. We developed a Bayesian multi-species occupancy model incorporating
62 incomplete detection to assess habitat use from data obtained as part of a multi-
63 year small-mammal trapping study (43,226 trap nights across four village sites in
64 Sierra Leone, 2020-2023). We investigated the effects of land use gradients and
65 small-mammal community dynamics on the spatial distribution of *M. natalensis*.
- 66 3. *Mastomys natalensis* occupancy increased along a gradient from forest to
67 agriculture to village habitats but was reduced in peri-urban settings compared to
68 rural settings. Invasive rodent species influenced this pattern, with *Mus musculus*
69 presence associated with reduced *M. natalensis* occupancy in peri-urban settings.
70 We did not observe a similar effect when considering the co-occurrence of invasive
71 *Rattus rattus* with *M. natalensis* in rural settings.
- 72 4. These findings suggest that land use and species interactions drive spatial
73 heterogeneity in *M. natalensis* populations, potentially explaining reduced Lassa
74 fever incidence in urban areas. The results highlight the importance of considering

75 community dynamics when predicting the risk of outbreaks of endemic zoonoses
76 and the need to widen the context of studies of LASV transmission beyond the
77 primary reservoir host species.

78 5. To better assess public health risk and improve allocation of limited resources, we
79 recommend more precise characterisation of small-mammal communities in LASV
80 endemic regions, particularly in areas undergoing rapid land use change which may
81 alter community level small-mammal biodiversity.

82 Introduction

83 Global biodiversity is in decline, with biodiversity loss directly influencing zoonotic disease
84 risk (Sala et al. 2000; Mantyka-Pringle et al. 2015; IPBES 2020). Land use change,
85 particularly the conversion of natural habitats into agricultural or urban landscapes, is a
86 key driver of biodiversity loss, reducing mammalian species diversity across several
87 dimensions (Newbold et al. 2015). These include taxonomic diversity (the number and
88 relative abundance of taxa), functional biodiversity (an increase in the abundance of
89 generalist, synanthropic animals) and interaction diversity (the biotic interactions among
90 species) (Naeem, Duffy, and Zavaleta 2012; Glidden et al. 2021). These declines in
91 biodiversity, particularly in rodent-associated disease systems, may exacerbate zoonotic
92 disease risks by promoting the proliferation of generalist, synanthropic rodents that thrive
93 in human-modified landscapes, where they host zoonotic pathogens (Young et al. 2014;
94 Gibb et al. 2020; Ecke et al. 2022). This shift may therefore not only increase the

95 prevalence of zoonotic pathogens but also human exposure in complex socio-ecological
96 systems (Gibb et al. 2025).

97 The role of host community diversity in mediating zoonotic outbreaks is nuanced, with
98 changes in species composition and pathogen prevalence often interacting with
99 anthropogenic stressors to create context-specific outcomes (Gibb et al. 2020; Keesing
100 and Ostfeld 2021; Carlson et al. 2025). A deeper mechanistic understanding of these
101 processes, including community structure, biotic interactions, and responses to
102 anthropogenic land use change, is critical for leveraging biodiversity knowledge to predict
103 and mitigate zoonotic risks (Salkeld, Padgett, and Jones 2013; Glidden et al. 2021; Carlson
104 et al. 2025). These approaches have only been taken for a few rodent associated zoonoses
105 (Keesing and Ostfeld 2024).

106 Rodents are an important mammalian host taxa for zoonotic diseases (Han et al. 2015;
107 Mendoza et al. 2019). Rodent-associated zoonoses, such as Lyme disease caused by
108 *Borrelia burgdorferi sensu lato*, have been shown to involve a complex interplay of
109 community structure, biotic interactions, and the effect of land use change (Ostfeld and
110 Holt 2004). For instance, land use change can increase reservoir abundance and
111 subsequently zoonotic risk, though responses vary across systems and settings,
112 highlighting the idiosyncratic or system specific nature of these dynamics (Young et al.
113 2017; Mendoza et al. 2019; Pei et al. 2024).

114 Lassa fever, caused by *Lassa marmarenavirus* (LASV), is an important rodent-associated
115 zoonosis endemic to West Africa, reported from Nigeria, Guinea, Sierra Leone, Liberia,

116 Mali, Benin, Ghana and Togo and causes an estimated 900,000 annual infections with
117 substantial morbidity and mortality (World Health Organisation 2022; Basinski et al. 2021).
118 Risk of infection in human populations is spatially heterogeneous with patchy distributions
119 of reported cases across the endemic region (Gibb et al. 2017; Agbonlahor et al. 2021;
120 Grant et al. 2023). While epidemiological factors such as acquired immunity and
121 asymptomatic infections may contribute, reservoir host ecology and distribution are likely
122 critical drivers of this variation (McCormick et al. 1987).

123 *Mastomys natalensis*, considered the main reservoir host, is a synanthropic rodent species
124 found throughout sub-Saharan Africa (IUCN 2016). It is abundant in and around areas of
125 human-dominated landscapes where it is considered a pest species (Leirs, Verhagen, and
126 Verheyen 1993). Population dynamics in *M. natalensis* are influenced by resource
127 availability and seasonal rainfall pulses, which drive fluctuations in abundance that
128 coincide with increased human Lassa fever outbreaks (Leirs et al. 1996; Leirs et al. 1997;
129 Redding et al. 2021). However, most intensive ecological studies of *M. natalensis* have
130 been conducted outside the Lassa fever endemic region, in regions with differing agro-
131 ecologies, potentially limiting their applicability to West Africa (Leirs et al. 1996). The
132 effects of land use change on rodent communities in the endemic region have only been
133 explored across a limited geographic area, with most studies focusing solely on the role of
134 the rodent reservoir (Fichet-Calvet et al. 2005, 2014; Fichet-Calvet et al. 2009; Olayemi et
135 al. 2018; Arruda et al. 2021). The limited number of studies that do consider the wider
136 rodent community typically focus on abundance (measured through summary proxies

137 such as trap-success) rather than explicitly investigating community dynamics (Fichet-
138 Calvet et al. 2009; Eskew et al. 2024).

139 There is a pressing need for systematic small-mammal community studies in West Africa
140 to ensure generalisability of these findings to Lassa fever, and to disentangle the
141 respective roles of abiotic factors, such as land use type, and biotic interactions within
142 small-mammal communities in shaping *M. natalensis* distributions and pathogen
143 exposure risks. *Mastomys natalensis* occurs within species rich settings in West Africa,
144 biotic interactions between this species and other native and invasive rodents within
145 communities and how these may regulate *M. natalensis* occurrence are not well described
146 (Fichet-Calvet et al. 2009; Garba et al. 2014; Cuyppers et al. 2017; Hima et al. 2019). Past
147 studies have indicated that competitive interactions with *R. rattus* and *M. musculus* can
148 alter native rodent communities, and might therefore reshape *M. natalensis* local
149 distributions with subsequent effects on LASV transmission and Lassa fever hazard
150 (Dalecky et al. 2015; Lippens et al. 2017; Eskew et al. 2024). Sierra Leone is associated
151 with frequent outbreaks of Lassa fever in human populations, with evidence of outbreaks
152 beyond the traditionally accepted endemic region with limited evidence for how
153 community dynamics (i.e., occurrence of invasive rodent species) might influence these
154 (Monath et al. 1974; Keenlyside et al. 1983; McCormick et al. 1987; Bonner et al. 2007;
155 Bonwitt et al. 2017; Bangura et al. 2021; Grant et al. 2023; Eskew et al. 2024).

156 Given these gaps in understanding, we sought to explore how small-mammal communities
157 are structured across land use gradients in Eastern Sierra Leone and how these dynamics
158 influence the distribution of *M. natalensis*. To achieve this we conducted repeated,

159 systematic, rodent trapping along a land use gradient in Eastern Sierra Leone to investigate
160 how land use influences small-mammal communities and the distribution of *M.*
161 *natalensis*. We hypothesised that: (1) small-mammal community diversity varies with land
162 use, with lower diversity in more anthropogenic habitats; (2) *M. natalensis* occupancy is
163 positively associated with anthropogenic land use but negatively influenced by
164 competition with sympatric species; and (3) species interactions within small-mammal
165 communities regulate the local distribution of *M. natalensis*, thereby shaping LASV
166 outbreak risk. Sierra Leone, a hotspot for Lassa fever, provides a unique opportunity to
167 explore these dynamics in a region where rodent ecology studies have largely focused on
168 *M. natalensis*, often neglecting the broader small-mammal community. We expect these
169 analyses to further understanding of small-mammal community structures that may
170 explain observed patterns of LASV outbreaks within this context and the wider endemic
171 region.

172 Methods

173 Small-mammal sampling

174 We conducted small-mammal trapping surveys from October 2020 to April 2023 at four
175 village study sites (Baiana, Lalehun, Lambayama, and Seilama) located in the Lassa fever
176 endemic zone of Sierra Leone's Eastern Province (Figure 1A). Site selection was informed
177 by discussions with the Kenema Government Hospital Lassa Fever team and guided by
178 remote imaging data to ensure representation of prior disease outbreak areas. Trapping

179 grids were established along a gradient of anthropogenic disturbance, encompassing
180 forest, agricultural land (both fallow and active), and village areas (inside and outside
181 permanent structures). Each village study site was assigned one forest grid, three to four
182 agricultural grids, and two village grids, except for Lambayama, which lacked forested land
183 (Supplementary Figure 1 A-D). Trapping survey sessions within each village occurred four
184 times annually with two trapping surveys in each of the rainy and dry seasons (May to
185 November and December to April, respectively), producing a total of 10 trapping sessions
186 over the study period (Figure 1B).

187 We selected village study sites and trapping grids within the village study sites to be
188 representative of land use in the Eastern Province of Sierra Leone and based on
189 accessibility to the sites during all seasons and acceptability of the study protocol to the
190 village study site communities (Supplementary Information 1). Trapping grids consisted of
191 49 Sherman traps ($7.62 \times 8.89 \times 22.86$ cm; H.B. Sherman Traps, Tallahassee, USA),
192 arranged in a 7×7 layout with 7-metre spacing between traps, adjusted to the local terrain
193 (median grid area = $3,882 \text{ m}^2$). For grids within permanent structures, the grid arrangement
194 was modified: individual buildings were semi-randomly selected using a projected village
195 grid, and four traps were deployed per structure. The location of each individual trap within
196 trapping grids was geolocated for subsequent data processing. Traps were baited with a
197 locally produced mixture of oats, palm oil and dried fish. Each morning the traps were
198 checked and closed for the day prior to re-baiting during the evening. Each trapping survey
199 session consisted of four consecutive trap-nights (TN) at each trapping grid within the
200 village study site.

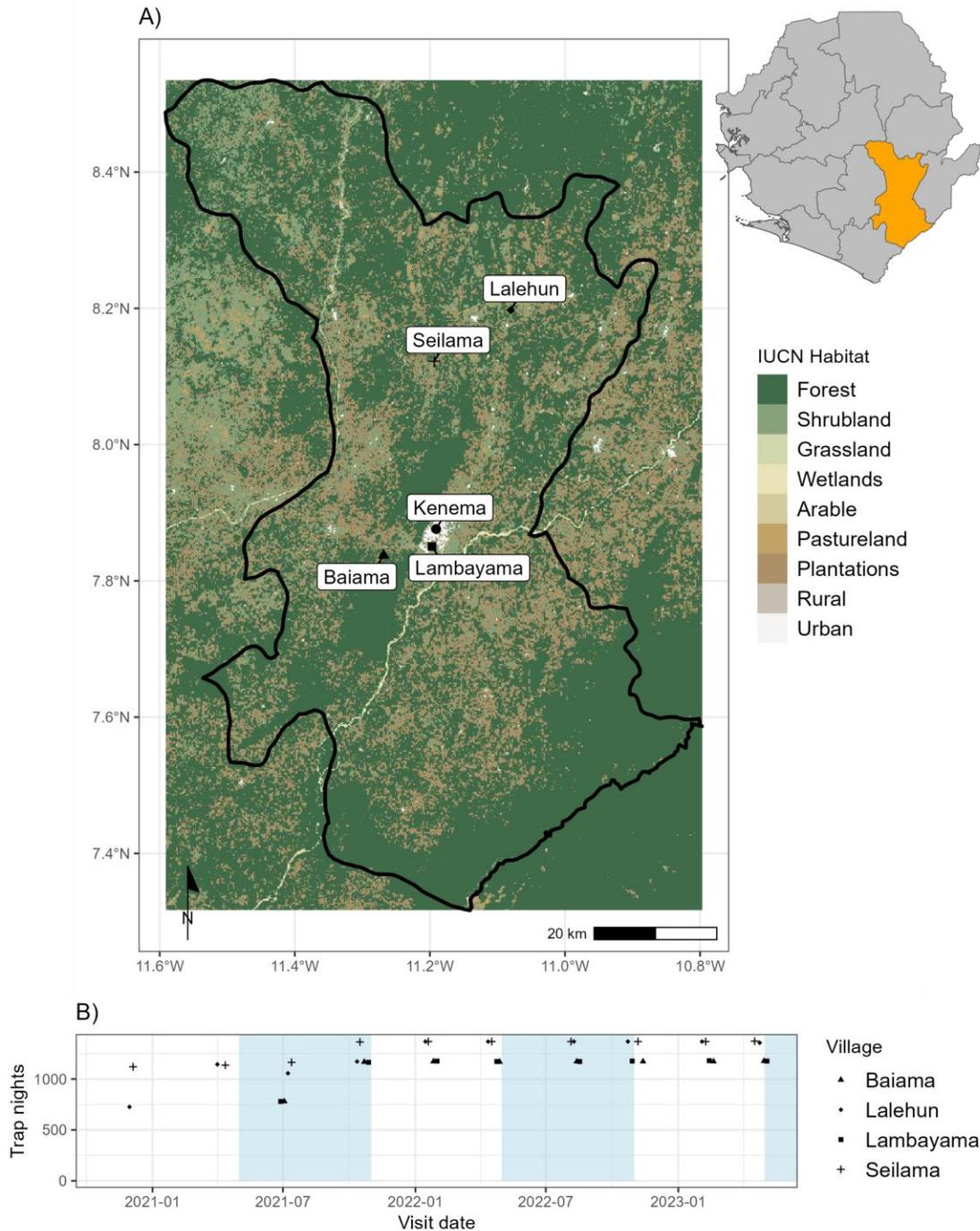
201 We standardised individual trap locations to 49 m² trapping grid cells for analysis. First, a
202 convex hull encompassing all trapping sessions for a grid was generated. A regular 49 m²
203 grid was overlaid on this polygon, and traps were assigned to cells based on their spatial
204 coordinates. This process resulted in 2,068 unique grid cells across all sites
205 (Supplementary Figure 2). Geospatial processing was performed using the sf package R
206 (version 4.1.2) (Pebesma 2018; R Core Team 2021). The four consecutive trap-nights
207 obtained from each trap during a single survey are pooled as a single replicate (i.e.,
208 trapping session) for the subsequent statistical analysis.

209 All small mammals were handled by trained researchers wearing appropriate personal
210 protective equipment. Animals were sedated using halothane and euthanized according to
211 established protocols (Fichet-Calvet 2014). Morphological measurements and samples of
212 blood and tissue were collected. The study was approved by the Clinical Research Ethical
213 Review Board and Animal Welfare Ethical Review Board of the Royal Veterinary College, UK
214 (URN: 2019 1949-3), and the ethical review board of Njala University, Sierra Leone, and
215 adhered to national and institutional ethical guidelines. Sex was determined based on
216 external and internal genitalia. Age classification was informed by reproductive status
217 (e.g., perforate vs. imperforate vagina, scarring from prior pregnancies, current pregnancy
218 status, descended testes, and seminal vesicle development) and lens weight from dried
219 eye samples. Photographs of the dorsal and ventral aspects of each rodent were taken for
220 morphological documentation. All carcasses were incinerated to mitigate pathogen
221 transmission risks.

222 Species classification

223 Species identification was performed in the field based on external morphological
224 characteristics, including body length, tail length, ear length, and pelage colouration,
225 following the taxonomic keys of Kingdon and Happold (Kingdon and Happold 2013) and
226 Monadjem *et al.* (Monadjem *et al.* 2015) (Supplementary Information 2). Field identification
227 was supplemented by molecular methods to confirm species identity for individuals
228 identified as *Mastomys sp.*, *Mus sp.*, *Rattus sp.* and *Crocidura sp.* alongside a random
229 subset of remaining individuals (50% of remaining samples).

230 Samples were stored at -20°C until processing and genomic DNA was extracted using
231 QIAGEN DNAeasy kits as per the manufacturers instructions (QIAGEN 2023)
232 (Supplementary Information 1). DNA extracts were amplified via polymerase chain
233 reaction (PCR) using *cytochrome B* primers, and amplification success was confirmed via
234 gel electrophoresis (Bangura *et al.* 2021). Full details of the PCR and sequencing protocols,
235 including thermocycler settings, are provided in Supplementary Information 1. Successful
236 PCR products were purified and submitted for Sanger sequencing (performed by Eurofins
237 Genomics). Attribution of sequences to rodent species was performed using the BLAST
238 programme, comparing obtained sequences to *cytochrome B* records in the NCBI
239 database (accessed 2023-06-30) (Altschul *et al.* 1990).



240

241 **Figure 1:** Village site locations and dates of rodent trapping in Sierra Leone. A) Location of
 242 village study sites (coloured labels), in the Eastern Province of Sierra Leone, Kenema, the
 243 largest city of the province is shown with a white label. The inset map shows the location of
 244 Sierra Leone in West Africa. B) Number of trap nights obtained from each study village, blue
 245 shaded regions represent the rainy season in Sierra Leone. The background raster image of
 246 Eastern Province, Sierra Leone was adapted from data presented in Jung et al. 2020.
 247 Shapefiles of Sierra Leone and administrative districts were obtained from GADM, 2022.

248 Description of small-mammal detection and species community

249 structure

250 Adequacy of sampling effort was assessed using species accumulation curves produced
251 for each village study site and each land use type within a village study site, generated with
252 the `specaccum` function from the `vegan` package (Oksanen et al. 2022). The rarefaction
253 method was applied to evaluate whether sampling was sufficient to detect the expected
254 rodent species within these categories. Curves were visually inspected, and most
255 demonstrated a plateau, suggesting adequate sampling effort (Supplementary Figure 3).

256 Detection/non-detection histories for each grid cell and small-mammal species were
257 constructed, assigning “1” when a species was detected and “0” otherwise. Species
258 communities were described at multiple spatial scales: (i) all species identified across all
259 village sites and land use types, (ii) all species identified within a village study site, and (iii)
260 all species identified within a single land use type within a single village study site. Species
261 richness and Shannon diversity were calculated at each of these spatial scales (Shannon
262 1948).

263 Estimating the effect of land use on species occurrence and richness

264 To adjust for differential probabilities of detection that may be driven by environmental
265 conditions and trapping effort during the trapping study and between species, we use a
266 Bayesian spatial latent factor multi-species occupancy model that incorporates residual
267 species correlations, imperfect detection and spatial autocorrelation. Variable selection

268 was informed by a pre-specified conceptual model (Supplementary Figure 4). Models were
 269 defined using the `sFMsPGOcc` function in the `spOccupancy` package in the R statistical
 270 computing language (Doser et al. 2022). This approach defines the true presence or
 271 absence (z) of a species (i), at grid cell (j) as arising from a Bernoulli process (Equation 1).
 272 Where ψ_j is the probability of occurrence of a species at a grid cell. This is modelled using
 273 a logit link where β_i are the species-specific regression coefficients of the site-specific
 274 covariates (\mathbf{x}_j^T) and a latent process $w_{i,j}^*$. This latent process incorporates residual species
 275 correlations through a small number of latent spatial factors and latent variables
 276 representing unmeasured grid cell covariates (Equation 2). Latent spatial factors account
 277 for spatial autocorrelation using a Nearest Neighbour Gaussian Process.

$$278 \quad z_{i,j} \sim \text{Bernoulli}(\psi_{i,j}) \quad (\text{Equation 1})$$

$$279 \quad \text{logit}(\psi_{i,j}) = \mathbf{x}_j^T \boldsymbol{\beta}_i + w_{i,j}^* \quad (\text{Equation 2})$$

280 The species-specific regression coefficients (β_i) are specified as random effects arising
 281 from a common community level distribution (Equation 3). Where $\boldsymbol{\mu}_\beta$ represents the
 282 community level mean effect for each occurrence covariate effect and \mathbf{T}_β is a diagonal
 283 matrix representing the variability of these among the species in the community.

$$284 \quad \boldsymbol{\beta}_i \sim \text{Normal}(\boldsymbol{\mu}_\beta, \mathbf{T}_\beta) \quad (\text{Equation 3})$$

285 The detection component estimates the unobserved $z_{i,j}$. Here, $y_{i,j,k}$ is the observed
 286 detection or non-detection of a species i , at site j , during replicate k (Equation 4). This is
 287 approached as arising from a Bernoulli process conditional on the true latent occurrence

288 process $p_{i,j,k}$. The probability of a species being detected at a grid cell, during a replicate
289 (given it is present at grid cell j), is a function of grid cell and replicate specific covariates \mathbf{v}
290 and a set of species-specific regression coefficients α_i (Equation 6).

$$291 \quad y_{i,j,k} \sim \text{Bernoulli}(p_{i,j,k} z_{i,j}) \quad (\text{Equation 4})$$

$$292 \quad \text{logit}(p_{i,j,k}) = \mathbf{v}_{i,j,k}^T \alpha_i \quad (\text{Equation 5})$$

293 Similarly to Equation 3, these coefficients are specified as random effects arising from a
294 common community level distribution, where μ_α represents the community level mean
295 effect for each detection covariate effect and \mathbf{T}_α is a diagonal matrix representing the
296 variability of these among species in the community (Equation 6).

$$297 \quad \alpha_i \sim \text{Normal}(\mu_\alpha, \mathbf{T}_\alpha) \quad (\text{Equation 6})$$

298 Minimally informative priors were specified for community and species level coefficients
299 (α and β , a normal prior of mean = 0, variance = 2.72) and for community level occurrence
300 and detection variance parameters (\mathbf{T}_α and \mathbf{T}_β , 0.1 for the scale and shape parameters of
301 the inverse Gamma prior).

302 We included covariates in the model based on a pre-specified conceptual model and after
303 assessing for co-linearity (defined as strong correlation >0.8) among variables. Continuous
304 variables were standardised by scaling values between 0-1. The fully specified model is
305 defined in Equation 7 and 8 and using a single latent spatial factor.

$$306 \quad \text{Probability of occurrence} \sim \text{Land use type} + \text{Village} + \text{scale}(\text{Distance to permanent structure}) + \text{scale}(\text{Elevation})$$

$$307 \quad (\text{Equation 7})$$

308 Probability of detection \sim scale(Monthly precipitation) + Moon fraction + scale(Number of trap nights)

309 (Equation 8)

310 Model checks, including mixing patterns of the MCMC sampler and posterior predictive
311 checks, were performed as an assessment of goodness of fit. Bayesian p-values were
312 calculated for both community-level and species-level fits, with values between 0.1 and
313 0.9 suggesting an adequate fit. The Widely Applicable Information Criterion (WAIC) was
314 used to guide final model selection (Watanabe 2010). Using this model, we estimate
315 occupancy probability for each species in different land use types. Only estimates for
316 species with at least 25 detections are included to avoid inference from limited data
317 (resulting in 7 species being included in this analysis).

318 We drew posterior samples from the most parsimonious Bayesian occupancy model
319 incorporating spatial autocorrelation to estimate the probability of occurrence of a species
320 within a trapping grid cell (Supplementary Table 1). The most parsimonious model included
321 variables for land use type and study village (Equation 9) to model the probability of
322 occurrence. Equation 9 was used for the probability of detection component with a single
323 latent spatial factor used.

324 Probability of occurrence \sim Land use type + Village

325 (Equation 9)

326 Co-occurrence of *Mastomys natalensis* with sympatric species

327 To investigate the potential for competitive exclusion of *Mastomys natalensis*, the reservoir
328 host of LASV, by other sympatric species, we examined correlations in the predicted

329 probabilities of occupancy between species pairs. These probabilities were derived for
330 each grid cell from the spatial multi-species occupancy model. Analyses were stratified by
331 land use type, and the Spearman rank correlation coefficient (ρ) was calculated for each
332 species pair. A two-sided test for statistical significance was performed, with the null
333 hypothesis being no correlation in the probabilities of occupancy for species pairs.

334 To minimise inference from sparse data, this analysis was restricted to species pairs that
335 were detected in the observed data within the specific land use type being examined.
336 Additionally, given the multiple statistical tests performed, we adopted a conservative
337 threshold for statistical significance ($p \leq 0.0005$) to reduce the likelihood of reporting false-
338 positive associations (Benjamin et al. 2018).

339 Statistically significant correlations were interpreted as indicating that one species was
340 either more likely (positive correlation) or less likely (negative correlation) to occupy a grid
341 cell within a given land use type when the other species was present. However, this
342 analysis does not allow for inferences regarding the causal mechanism or directionality of
343 any observed relationships (e.g., whether one species excludes or is excluded by another).

344 Results

345 Small-mammal detection and species community structure

346 Over the study period 684 individual small mammals were detected from 43,266 trap-
347 nights across the four village study sites, yielding an overall trap success (TS) of 1.6%. The

348 agricultural areas had the highest species richness and Shannon diversity values, while TS
349 was greatest in village settings, including both inside and outside permanent structures
350 (Table 1). Among the study sites, Seilama had the highest overall TS, species richness, and
351 Shannon diversity. Notably, Seilama also exhibited the greatest TS within agricultural
352 areas, unlike the other three sites. Species richness in Seilama was twice that of
353 Lambayama, the peri-urban village study site, which also had the lowest species richness
354 and Shannon diversity. In Lambayama, most rodents were detected within built-up village
355 areas, reflecting its proximity to Kenema city.

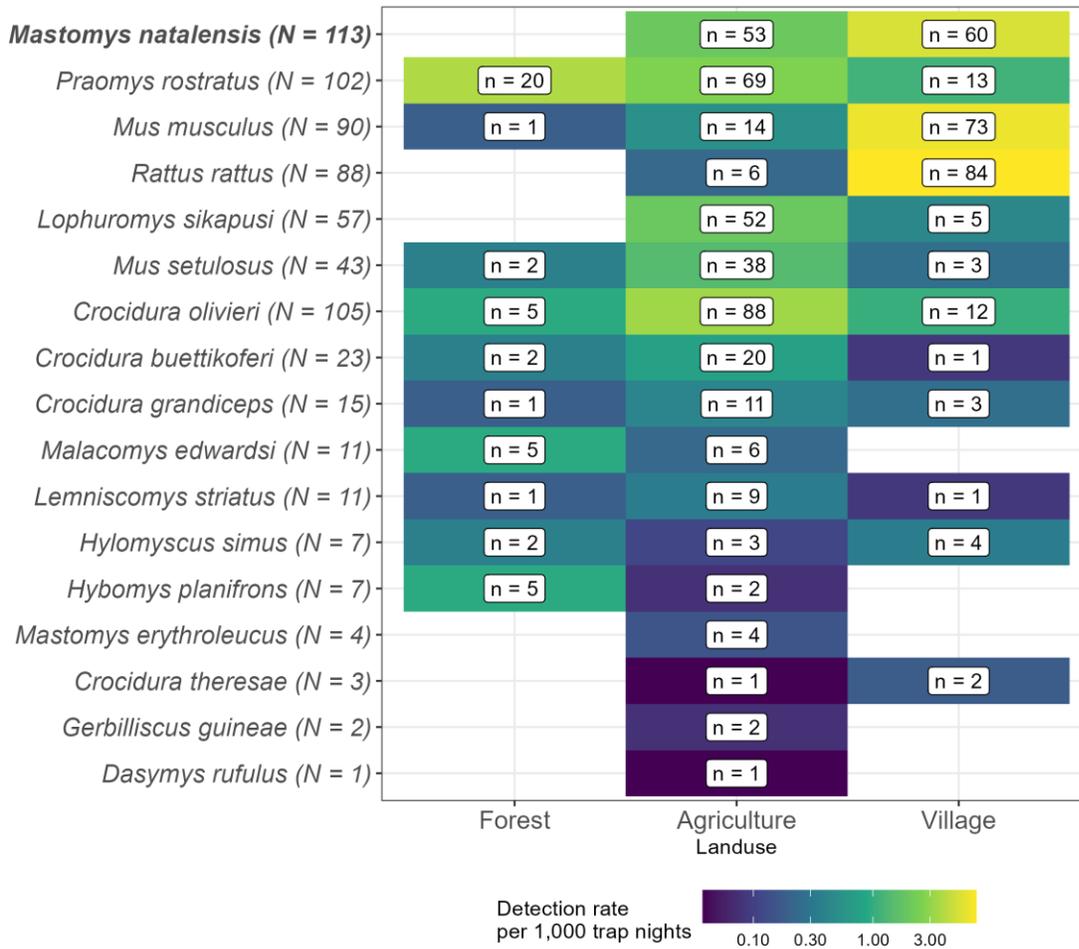
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Table 1. The number of trapped individuals (N), the number of trap nights (TN), trap-success (TS %), species richness and Shannon diversity by village and land use type.

Village	Land use	N	TN (TS %)	Species richness	Shannon diversity
All villages					
	Village	261	11516 (2.3%)	12	1.67
	Agriculture	379	26400 (1.4%)	17	2.20
	Forest	44	5350 (0.8%)	10	1.78
Baiaama					
	Village	73	2716 (2.7%)	8	1.11
	Agriculture	45	4696 (1%)	9	1.96
	Forest	3	1568 (0.2%)	2	0.64
	Total	121	8980 (1.3%)	12	1.73
Lalehun					
	Village	54	2824 (1.9%)	9	1.65
	Agriculture	98	7608 (1.3%)	13	2.18
	Forest	5	1862 (0.3%)	3	1.05
	Total	157	12294 (1.3%)	13	2.21
Lambayama					
	Village	93	2736 (3.4%)	4	0.42
	Agriculture	50	6260 (0.8%)	6	1.19
	Total	143	8996 (1.6%)	6	1.03
Seilama					
	Village	41	3240 (1.3%)	8	1.54
	Agriculture	186	7836 (2.4%)	13	1.97
	Forest	36	1920 (1.9%)	8	1.51
	Total	263	12996 (2%)	14	2.07

359 The most commonly detected rodent species across all land use types was *M. natalensis*
360 (N = 113, 16.5%), followed by *Praomys rostratus* (N = 102, 14.9%), *M. musculus* (N = 90,
361 13.2%), *R. rattus* (N = 88, 12.9%) and *Lophuromys sikapusi* (N = 57, 8.3%) and *Mus*
362 *setulosus* (N = 43, 6.3%). Among non-rodent species, the insectivorous shrew *Crocidura*
363 *olivieri* was the most frequently detected (N = 105, 15.4%). *Mastomys natalensis* and *R.*
364 *rattus* were found in all village study sites, although *M. natalensis* was absent in forest
365 settings (Figure 2). The invasive rodent *M. musculus* was detected only in Lambayama, the
366 peri-urban site.

367 Detection rates (number of individuals per 1,000 trap-nights) varied significantly by
368 species, land use type, and village study site. The highest detection rate was observed for
369 *M. musculus* in the Lambayama village site, while *M. natalensis* and *R. rattus* showed high
370 detection rates across multiple sites in village land use types. In contrast, *P. rostratus* had
371 the highest detection rates in forest and agricultural areas.



372

373 **Figure 2.** Detection rate per 1,000 trap nights of small mammal species across different
 374 land use types (grouped across village study sites). The total number of detections in this
 375 study is shown next to the species binomial name (N). The number of detections of each
 376 species in each land use is shown in the label (n). The colour of the tile corresponds to the
 377 detection rate per 1,000 trap nights.

378 Seasonal variation in species detection was observed but did not show consistent trends

379 across all species. While *M. musculus* had higher detection rates during the rainy season,

380 species such as *L. sikapusi* and *Mus setulosus* were more frequently detected in the dry

381 season. For most other species, detection rates were similar across seasons

382 (Supplementary Figure 5A). When stratified by land use type, some variations were evident.

383 For instance, *M. natalensis* was more frequently detected in village settings than in

384 agricultural areas during the rainy season, but detection rates were similar between the
385 two land use types in the dry season. *Praomys rostratus* showed higher detection rates in
386 forests during the dry season compared with the rainy season. No other species showed
387 notable seasonal variations when stratified by land use type (Supplementary Figure 5B).

388 Estimating the effect of land use on species occurrence and richness

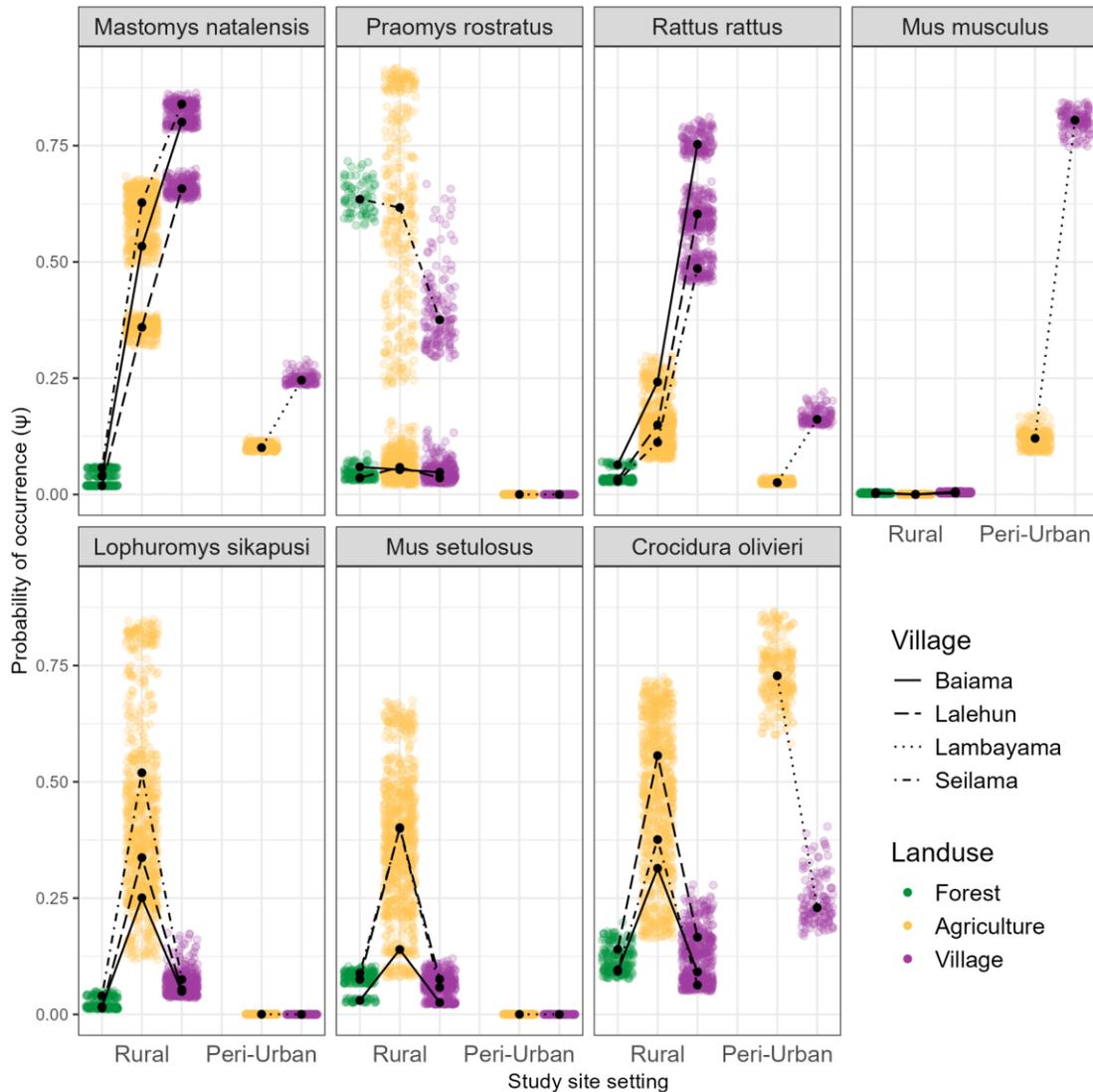
389 We found three patterns of probability of occurrence (ψ) within a trapping grid cell for the
390 seven included species (Figure 3 and, marginal effects of the detection parameters are
391 shown in Supplementary Figures 6-8). First, *M. natalensis*, *R. rattus* and *M. musculus* had
392 greatest probabilities of occurrence in villages with lower occurrence probabilities in
393 agricultural and forest areas. *Mastomys natalensis* differed from the two commensal,
394 invasive species (*R. rattus* and *M. musculus*) as their probability of occurrence in
395 agricultural settings, while lower than village settings, remained generally high. Second, *P.*
396 *rostratus* had high probability of occurrence in forests with lower probabilities in
397 agricultural areas and villages. Finally, *C. olivieri*, *L. sikapusi* and *M. setulosus* had their
398 highest probabilities of occurrence in agricultural areas with lower probabilities of
399 occurrence in forests and villages. No species showed high probability of occurrence
400 across all land use types, consistent with species being adapted to distinct ecological
401 niches.

402 The probability of occurrence within a trapping grid cell, within the same land use type
403 showed wide variability for some species. For example, the narrow range of probabilities
404 for *M. natalensis* in forests (0-13%) is suggestive that the probability of this species

405 occurring within forests is low. This compares to the wide variability for *P. rostratus* in
406 agricultural areas (0-90%), this suggests that additional environmental factors beyond land
407 use type are affecting the probability occurrence.

408 To further explore this, we stratified village study sites by human population density into
409 rural and peri-urban sites (rural \leq 500 individuals per 1km²) (Figure 3). The probability of
410 occurrence of *M. natalensis* was importantly different between these settings, with high
411 probability of occurrence in both agricultural and village settings in rural areas but
412 substantially lower probability in peri-urban study sites. The same pattern was observed
413 for *R. rattus*. For the rodent species predicted to have a lower probability of occurrence in
414 village settings (*P. rostratus*, *L. sikapusi* and *M. setulosus*) probabilities of occurrence were
415 greater in all land use types in rural areas compared to peri-urban areas. Shrew species
416 had similar probabilities of occurrence in rural and peri-urban areas. Human population
417 density itself or other environmental factors strongly associated with human population
418 density may therefore be importantly contributing to small-mammal species occurrence.

419 In contrast to species found throughout our study area, *M. musculus* was predicted to
420 have a low probability of occurrence in all land use types in rural areas, with high
421 probabilities of occurrence only for village settings in peri-urban areas. The occurrence
422 probabilities for the three commensal species (*M. natalensis*, *R. rattus* and *M. musculus*)
423 suggest that competition may be a factor in reducing the occurrence probabilities of *M.*
424 *natalensis* and *R. rattus* in the presence of *M. musculus* as in its absence these two
425 species have high occurrence probabilities in villages.



426

427 **Figure 3.** Probability of species occurrence (ψ) across a land use gradient, stratified by
 428 rural and peri-urban village study sites, for the seven rodent and shrew species. Each
 429 coloured point represents the median predicted probability of occurrence for a species,
 430 obtained from the posterior distribution at each trapping grid cell. Colours correspond to
 431 different land use types. Predictions are based on 2,068 trapping grid cells. Black points
 432 indicate the median probability of occurrence within each land use type, grouped by
 433 village, with lines connecting estimates within villages for different land use types. Lines
 434 are shown only to link the occupancy estimates for each land use type within a given
 435 village.

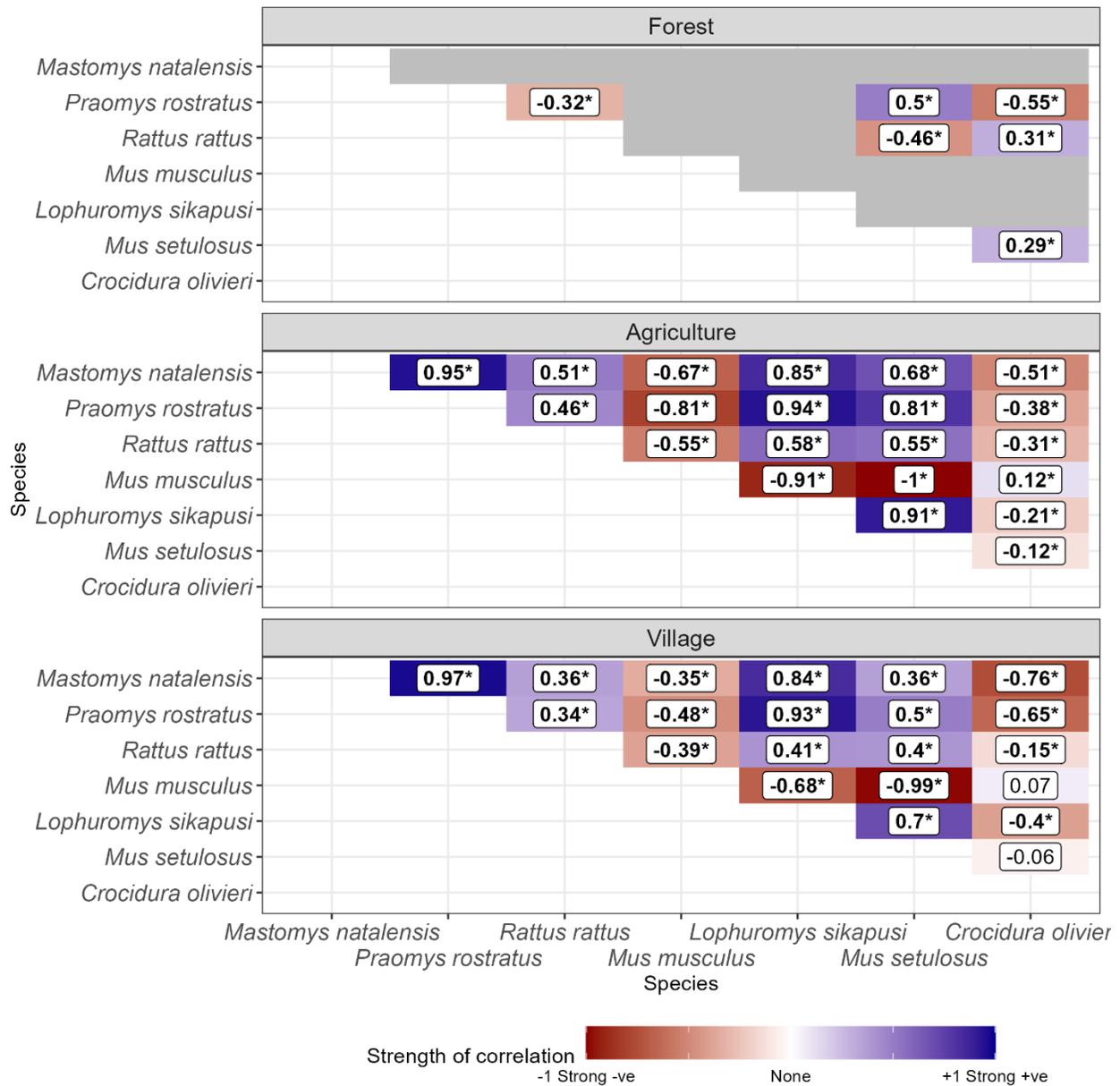
436 These patterns of species occurrence, particularly in relation to human population density
 437 and land use type, suggest that urbanisation and land use change could substantially

438 influence the distribution of small-mammal species, with potential implications for
439 zoonotic disease dynamics and biodiversity conservation.

440 Co-occurrence of species within land use types

441 The tests for species correlations reveal patterns consistent with our original hypothesis
442 that the local spatial distribution of *M. natalensis* is regulated by biotic interactions with
443 co-occurring species (Figure 4). While this analytical approach cannot demonstrate
444 causality between co-occurrence and the probability of occurrence, it is possible that
445 unmeasured factors are influencing the observed associations. In land use types where
446 both *M. natalensis* and *M. musculus* co-occurred, the presence of one species was
447 associated with a reduced probability of occurrence for the other. Statistically significant
448 negative correlations were observed in agricultural (Spearman's $\rho = -0.67, p < 0.0001$) and
449 village ($\rho = -0.35, p < 0.0001$) settings. In contrast, a positive correlation was found
450 between the occurrences of *M. natalensis* and the other commensal, invasive rodent *R.*
451 *rattus*, with significant positive correlations in both agricultural ($\rho = 0.51, p < 0.0001$) and
452 village ($\rho = 0.36, p < 0.0001$) settings.

453 Within villages, high probabilities of *M. musculus* presence were associated with lower
454 probabilities for all other rodent species. This pattern was not replicated for *M. natalensis*
455 and *R. rattus*, which were positively correlated with the co-occurrence of native rodent
456 species, including *P. rostratus*, *L. sikapusi*, and *M. setulosus*. Across all land use types, the
457 presence of the shrew species, *C. olivieri* was negatively correlated with the presence of
458 rodent species.



459

460 **Figure 4.** Spearman's rank correlations for the modelled site-level probability of
 461 occurrence of species pairs in different land use types. Positive values (blue shades)
 462 represent positive correlation coefficients between the occurrence of two species.
 463 Negative values (red shades) represent negative correlation between the occurrence of
 464 two species. Numbers in bold typeface and indicated with an asterisk (*) are statistically
 465 significant at a level of $p \leq 0.0005$. Grey tiles are used where no detections of the species
 466 pair in the land use type were observed and therefore excluded from analysis.

467 Discussion

468 This study provides valuable insights into the dynamics of small-mammal communities in
469 a Lassa fever endemic region of Eastern Sierra Leone, focusing on how land-use types
470 influence species richness, diversity, and the risk of viral spillover. Our results indicate that
471 species richness and diversity were highest in agricultural settings, followed by villages,
472 with lower richness in forests. Agricultural habitats support both synanthropic and non-
473 synanthropic species, increasing the potential for viral sharing.

474 The highest probability of occupancy for the LASV reservoir host, *M. natalensis*, was
475 observed in villages and agricultural settings, while it was likely absent from forests. This
476 pattern mirrors that of the invasive commensal species *M. musculus* and *R. rattus*,
477 suggesting that human-modified landscapes may be key drivers of rodent distribution.
478 Stratification by human population density revealed that *M. natalensis* occupancy was
479 lower in peri-urban settings, where *M. musculus* dominates. Importantly, we identified a
480 negative correlation in the probability of co-occurrence between *M. natalensis* and *M.*
481 *musculus* within villages, which could have important implications for understanding
482 observed Lassa fever outbreaks.

483 Our study suggests that agricultural settings, with their higher species richness, may be
484 hotspots for LASV transmission among rodents. The diverse rodent communities found in
485 these areas could facilitate viral persistence and reintroduction, particularly among
486 isolated populations of *M. natalensis*. These findings underscore the importance of

487 considering both species diversity and land use when assessing the risk of zoonotic
488 spillover.

489 Small-mammal communities are associated with land use type

490 Small-mammal species richness was greatest in agricultural settings. In these settings
491 both synanthropic and non-synanthropic species were found. The higher species richness
492 in agricultural environments provides more opportunities for cross-species transmission
493 of LASV within diverse small-mammal communities.

494 Evidence suggests that the wider species community may play a role in LASV transmission.

495 Current or prior infection with LASV (through detection of virus or antibodies), has been
496 identified in 11 additional small-mammal species to *M. natalensis*. Whether these
497 infections are incidental or represent competent chains of viral transmission remains
498 unclear (Monath et al. 1974; Demby et al. 2001; Fichet-Calvet et al. 2014; Olayemi et al.

499 2016; Simons et al. 2023). It is possible that viral sharing within small-mammal
500 communities is greatest in the more species rich agricultural settings, allowing

501 introduction or re-introduction of LASV into isolated commensal species populations
502 following local extinction of virus (Bordes, Blasdell, and Morand 2015). This may be

503 particularly important for maintaining viral persistence through time in spatially isolated *M.*

504 *natalensis* populations, where rapid depletion of susceptible individuals is expected in
505 well-mixed populations (Goyens et al. 2013). Our findings, along with previous studies,

506 show that *M. natalensis* is absent from forested regions and populations are spatially

507 isolated, further supporting the hypothesis of limited geographic dispersal of this species

508 (Leirs, Verheyen, and Verhagen 1996; Denys et al. 2005; Mariën et al. 2018). However, the
509 role of the wider rodent community in facilitating LASV transmission between isolated *M.*
510 *natalensis* populations warrants further investigation.

511 Although previous studies from Guinea and Sierra Leone have reported seasonal
512 fluctuations in *M. natalensis* prevalence, our findings of similar or increased prevalence
513 during the dry season suggest that localized factors such as agricultural practices or food
514 storage, may influence rodent behaviour in this region somewhat differently than
515 previously documented (Fichet-Calvet et al. 2007; Bangura et al. 2021). Alternatively,
516 increased trap-shyness during periods of increased abundance may mask replication of
517 previous findings. Further studies on small-mammal communities, incorporating local
518 human community behaviour and practices, conducted over longer time periods and
519 across different geographic regions, would be valuable in identifying how seasonal habitat
520 preferences of these rodents vary (Kelly et al. 2013; Leach et al. 2017). These findings
521 underscore the importance of considering local human activities, such as food storage
522 and agricultural practices (e.g., one vs. two annual planting and harvesting cycles), when
523 modelling seasonal patterns in rodent behaviour. Such practices may alter rodent
524 abundance and movement in ways that differ from areas where *M. natalensis* displays
525 more predictable seasonal patterns due to other environmental or ecological factors.

526 The prevalence of *M. natalensis* within households has been suggested as a key driver of
527 Lassa fever outbreaks, likely due to increased human-rodent contact in household settings
528 (Bonwitt et al. 2017; Mariën et al. 2020). Therefore, the movement of *M. natalensis* from
529 species-rich agricultural areas to households may play a critical role in the transmission

530 dynamics of LASV. Understanding these movement patterns and their interactions with
531 local human activities will be essential for developing targeted interventions to reduce the
532 risk of Lassa fever outbreaks.

533 Evidence for biotic interactions shaping patterns of small-mammal 534 species diversity

535 The segregation of small-mammal species into distinct ecological niches, categorised by
536 human-dominated (village and agricultural) or non-human-dominated (forest) land use
537 types, underscores the significant role of biotic factors in species distribution. Our findings
538 show a high predicted occupancy of both *M. natalensis* and *R. rattus* in human-dominated
539 landscapes, with a positive correlation in their co-occurrence, aligning with a similar study
540 conducted in another region of Sierra Leone (Bangura et al. 2021). In contrast, a study
541 combining data from Guinea and Sierra Leone reported a decreased abundance
542 (measured as catch per trap) of *M. natalensis* in the presence of *R. rattus* (Eskew et al.
543 2024). This suggests that *M. natalensis* may be displaced by *R. rattus* at a lower rate
544 compared to *M. musculus*, but the extent of this displacement likely depends on which
545 invasive species has already established dominance in a given area. Further evidence from
546 Senegal suggests that *M. musculus* and *R. rattus* may invade along different routes
547 (Dalecky et al. 2015). Therefore, the findings from this study and the Eskew study likely
548 reflect a more intricate process of invasion and species displacement, which is contingent
549 on the timing and dominance of either *M. musculus* or *R. rattus* within a landscape.

550 In village environments, the probability of *M. musculus* occurrence was negatively
551 correlated with the co-occurrence of both *M. natalensis* and *R. rattus*. Interestingly, *M.*
552 *musculus* was absent from rural villages where *R. rattus* and *M. natalensis* had high
553 probabilities of occurrence. This pattern extended across all other rodent species in village
554 environments, suggesting that *M. musculus* may be currently unable to establish
555 populations in areas already dominated by *R. rattus* and *M. natalensis*. While it is not
556 possible from this study to determine whether *M. musculus* is actively expanding into rural
557 settings, the potential implications for small-mammal community dynamics and LASV
558 transmission remain a critical area for future research.

559 To better understand the causal processes behind changes in small-mammal community
560 structures, especially in response to the expansion of invasive species, long-term
561 monitoring of rodent populations is necessary. Similar efforts conducted in Senegal would
562 be valuable for tracking the progression of invasive species and their effects on local
563 biodiversity and disease transmission risks (Dalecky et al. 2015).

564 Benefits and challenges of systematic small-mammal community 565 sampling

566 Systematic investigation of small-mammal communities entails a greater sampling effort
567 than targeted sampling of specific species in selected habitats. In this study, the overall TS
568 was relatively low compared to other studies focusing on synanthropic rodent species
569 (Olayemi et al. 2018; Bangura et al. 2021; Happi et al. 2022). Our TS rate of 3.3% within
570 villages aligns with the 3% reported in Bo, Sierra Leone, but is notably lower than the 17%

571 and 14% reported in Nigeria and Guinea, respectively (Fichet-Calvet et al. 2007; Happi et
572 al. 2022). This discrepancy may arise from differences in trapping methodologies,
573 including the size of traps, trap locations, or the behaviour of target species. For example,
574 *M. natalensis* captures in its Western range (Nigeria, Guinea, Sierra Leone) were less
575 frequent than those observed in Tanzania, where TS rates of around 24% were reported in
576 agricultural settings (Mulungu et al. 2013). One potential explanation is that the food
577 availability in each setting might affect the species' behaviour, with environmental food
578 sources influencing trap-shyness (Taylor, Hammond, and Quy 1974; Stryjek, Kalinowski,
579 and Parsons 2019).

580 Despite the higher sampling effort required, systematic small-mammal community
581 sampling offers distinct advantages over targeted species sampling, particularly in
582 addressing biases related to species representation and pathogen detection. By adopting
583 a broader, more inclusive approach, this method reduces the risk of overlooking less
584 common species that may act as pathogen hosts but would otherwise be
585 underrepresented in more focused sampling. Furthermore, this comprehensive approach
586 provides greater insight into biotic interactions between species. While targeted sampling
587 may focus on a single species or a small subset of species, a more systematic approach
588 enables the detection of interactions among different rodent species, which may be
589 crucial for understanding pathogen dynamics. For instance, our sampling design allowed
590 us to observe interactions between synanthropic species like *M. musculus* and native
591 species, potentially influencing pathogen transmission patterns. Wider sampling can

592 reveal such interactions that might otherwise remain hidden, offering a more holistic view
593 of the ecological factors contributing to pathogen spread and spillover risk.

594 Comparison between studies using different sampling techniques and study designs
595 presents several challenges. Previous studies on rodent communities in the Lassa fever
596 endemic region have used TS rates as an indirect measure of rodent abundance in the
597 absence of capture-mark-recapture studies (Fichet-Calvet et al. 2009; Olayemi et al. 2018;
598 Bangura et al. 2021). Our analysis, incorporating a model of imperfect detection, suggests
599 estimating abundance from TS may not be applicable across different land use types and
600 species (Supplementary Figure 8). For example, we observed that the probability of
601 detecting *M. musculus* and *R. rattus* was higher than for native species when a consistent
602 amount of trapping effort was applied. This finding aligns with previous research showing
603 that detection rates, when used as a proxy for relative abundance, may be unreliable,
604 especially when combining live-trapping and camera-trapping methods (Parsons, Clark,
605 and Kays 2022). Several factors can contribute to variability in detection, including trap-
606 shyness (neophobia) in non-synanthropic species, local resource availability, and trap
607 placement (Stryjek, Kalinowski, and Parsons 2019). Understanding how these factors
608 affect detection rates is essential for improving our ability to assess small-mammal
609 populations accurately. Therefore, improving harmonisation of rodent sampling designs —
610 particularly those incorporating systematic small-mammal community sampling — could
611 enhance direct comparisons of species communities and pathogen prevalence across the
612 Lassa fever endemic region (Simons et al. 2023). More broadly, this could lead to a deeper

613 understanding of how different land use types and ecological factors influence rodent
614 populations and their role in pathogen transmission.

615 There are several limitations to the current study. One limitation is the relatively short
616 duration of our animal sampling (less than three years). Given that rodent populations can
617 fluctuate significantly over longer time periods, it is possible that important variations in
618 abundance were missed during our study. As a result, the probability of occurrence for
619 certain species may have been underestimated, particularly for species that were at low
620 abundance during our survey period. Extending the sampling period would help to better
621 capture temporal changes in species occurrence and abundance. Additionally, land use in
622 Sierra Leone, particularly in agricultural areas, follows multi-year cycles of cultivation,
623 fallow, and reconversion to cultivated land. To more comprehensively understand the
624 effects of land use change on rodent communities, it would be useful to focus on a single
625 location that spans the transition from forested land to agricultural, fallow, and degraded
626 forest land use. Such a study would provide insights into how these different land
627 management practices influence rodent community dynamics and pathogen risk.
628 Furthermore, unobserved characteristics of the villages in our study may have contributed
629 to variations in the composition of rodent communities, as suggested by the wide posterior
630 distributions for some species. Expanding our study to include a greater number of villages
631 would help increase the generalisability of our findings and improve the robustness of
632 conclusions drawn for the wider region.

633 Implications for understanding the risk of LASV spillover

634 The lower occurrence of *M. natalensis* in agricultural and forest land use is consistent with
635 growing evidence of LASV prevalence heterogeneity across the endemic region (Mariën et
636 al. 2020). In some village communities, no current evidence of LASV transmission has
637 been detected within rodent populations, despite prior human cases or serological
638 evidence of outbreaks. This suggests that pathogen prevalence in rodents may vary
639 significantly over time and space (Bangura et al. 2021; McCormick et al. 1987; Leski et al.
640 2015). As discussed earlier, LASV transmission among the rodent community may be
641 short-lived, with rapid local extinction of the virus (Goyens et al. 2013). These dynamics
642 raise the possibility that non-*M. natalensis* species may play a crucial role in transferring
643 the virus between communities of *M. natalensis* that reside in villages separated by forest
644 or other land use types that are not colonised by *M. natalensis*, leading to pathogen re-
645 introduction. Species found in forested settings, such as *P. rostratus*, *M. setulosus*,
646 *Malacomys edwardsi*, and *L. striatus*, have shown evidence of prior LASV exposure, as
647 indicated by antibodies (Monath et al. 1974; Demby et al. 2001; Fichet-Calvet et al. 2014;
648 Olayemi et al. 2016; Simons et al. 2023). To better understand the temporal and spatial
649 variability in LASV prevalence, it is imperative to conduct small-mammal sampling and
650 movement ecology studies across the broader land use gradient.

651 Current disease models of LASV risk largely fail to incorporate the role of multiple rodent
652 species and the potential biotic interactions between them (Basinski et al. 2021; Mylne et
653 al. 2015; Ologasa et al. 2014; Redding et al. 2016; Fichet-Calvet and Rogers 2009; Klitting

654 et al. 2022). Our findings, which highlight interactions between *M. natalensis* and primarily
655 *M. musculus*, suggest that Lassa fever risk could potentially be reduced in areas where *M.*
656 *musculus* is present. This raises the need for further research into the competence of *M.*
657 *musculus* as a host for LASV, as previous serological evidence has shown exposure to the
658 virus in this species (Demby et al. 2001). If *M. musculus* is not a competent host for LASV,
659 this could help explain why Lassa fever is more commonly reported in rural areas of the
660 endemic region, rather than urban centers, where *M. musculus* may have displaced more
661 competent viral hosts. To test this hypothesis, further research systematically sampling
662 along the urban-rural gradient is essential. Such work could significantly impact future
663 projections of Lassa fever risk, especially given the rapid population growth and
664 urbanisation occurring across West Africa. The expansion of *M. musculus* in these settings
665 may moderate the risk of increased Lassa fever outbreaks, making it crucial to better
666 understand the implications of urbanisation on zoonotic outbreak risks.

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