

1 **Selection on *Picea mariana* shifts with climate and evolutionary response to**
2 **climate change is largely unconstrained by phenotypic integration**

3
4 **Authors**

5 Sabina Henry¹, Julie Messier¹

6 **Affiliations**

7 1 Department of Biology, University of Waterloo, Waterloo, Ontario, Canada

8
9 **Corresponding author:**

10 Email address: sihenry@uwaterloo.ca

11
12 **ABSTRACT**

13 The local persistence of long-lived organisms is at risk as climate change drives a rapid shift in
14 selection regimes world-wide. Although adaptive evolution is one of the main mechanisms by
15 which populations persist in changing environments, we have little information regarding how
16 selection regimes will shift in response to continued climate change, nor on the potential for trees
17 to evolve adaptively under novel selection pressures. To address these gaps, here we assessed the
18 changes in selection in three sites along a spatial climate gradient which mimics expected
19 temporal changes in climate and determined whether trait covariance might accelerate or impede
20 the rate of adaptive evolution of seven *Picea mariana* populations in the warmer and drier
21 environment. In three common garden sites established 50 years ago, we measured an array of
22 traits which represent water use, response to temperature, structural investment, and metabolic
23 efficiency. Our findings reveal that all 10 traits measured in this study were under selection in at
24 least one site. We also find different traits are under selection in each site, with four instances
25 where the shift in selection gradient is consistent with shifts in climate: water use efficiency
26 (WUE); needle carbon to nitrogen ratio (CN); the interaction between WUE and CN; and the
27 interaction between CN and huber value. In the warm and dry site, traits under selection were
28 largely uncorrelated, with only four of the 49 trait combinations under selection exhibiting intra
29 population trait covariances. The shifts in selection gradient suggest that climate change may
30 select for needles with higher WUE, higher structural carbon and higher hydraulic supply to the
31 needles. The few trait-trait correlations indicate that phenotypic integration should neither
32 impede nor facilitate adaptive evolution, leaving *P. mariana* provenances with the evolutionary
33 flexibility to respond to climate change regardless of the direction to selection.

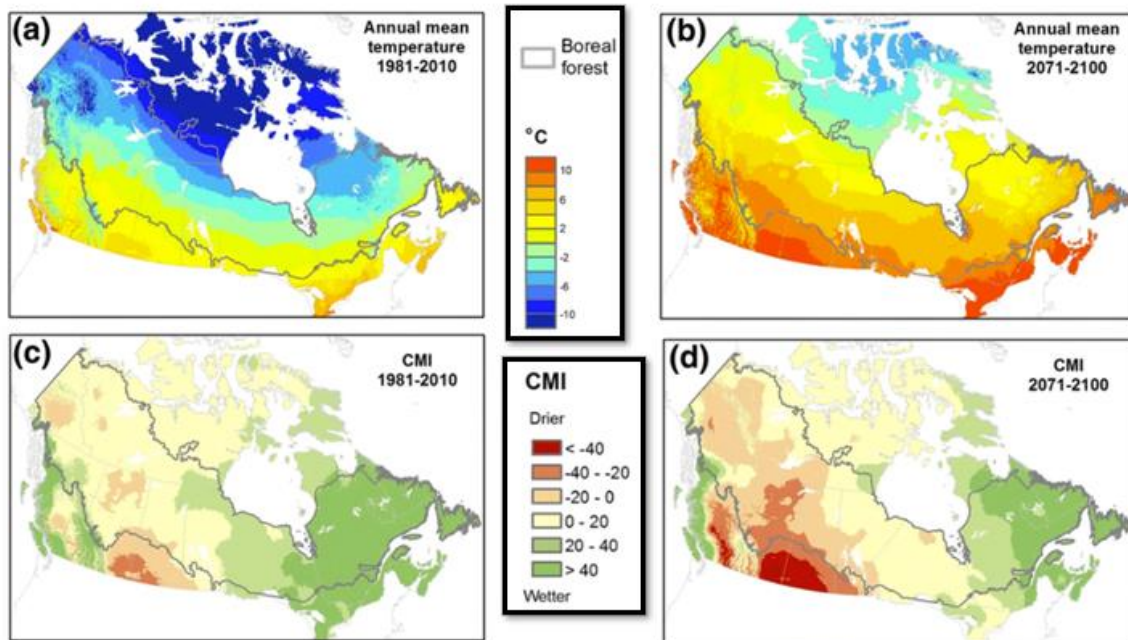
34 INTRODUCTION

35 As climates shift under global climate change, the growth, productivity and persistence of
36 tree species globally are at risk (Allen *et al.*, 2010; Choat *et al.*, 2012). This is especially true for
37 the boreal forest which is experiencing climate change at a rapid rate compared to regions at
38 lower latitudes (Aubin *et al.*, 2018; Chagnon *et al.*, 2022). These rapid shifts in environmental
39 conditions expose local populations to new selection pressures with little time for evolution by
40 natural selection to occur, leaving long-lived sessile organisms at particular risk of becoming
41 mismatched with new environmental conditions (Barrett & Schluter, 2008; de Lafontaine *et al.*,
42 2018). Studies worldwide have reported widespread forest tree mortality driven by this
43 mismatch, leaving adaptation and migration as possible outcomes which may preserve
44 population persistence (Shaw *et al.*, 2005; McDowell *et al.*, 2020; Münchinger *et al.*, 2023).
45 However, migration rates are likely too slow to match the pace of climate change (Malcolm *et al.*
46 *et al.*, 2002; Iverson *et al.*, 2004; Brecka *et al.*, 2018). As a result, research has focused on
47 identifying adaptations which would allow for the persistence of populations in the face of
48 climate change. Adaptations are phenotypic characteristics associated with higher fitness within
49 a population in a given environment. These can occur through adaptive plasticity, whereby an
50 individual's plastic response increases fitness or through adaptive evolution whereby traits which
51 confer high fitness are preferentially passed on from one generation to the next. Although both
52 adaptive plasticity and adaptive evolution can contribute to population persistence most work to
53 date focuses on adaptive plasticity and we know little about the potential for adaptive evolution
54 in trees (henceforth, adaptive potential). Indeed, in recent years many studies have aimed to
55 assess adaptive capacity in the face of climate change by solely quantifying adaptive plasticity of
56 phenotypic traits using locally adapted populations from warm and dry environments (Andrés-
57 Hernández *et al.*, 2023; Blasini *et al.*, 2021; Challis *et al.*, 2022; Robert *et al.*, 2024). However,

58 recent meta-analyses on adaptive capacity have highlighted the need to also examine the role of
59 adaptive evolution to provide a more comprehensive assessment of population adaptive capacity
60 in the face of climate change (Lindner *et al.*, 2010; Royer-Tardif *et al.*, 2021). As a result,
61 research on the adaptive potential of important tree species is required to determine whether
62 populations will be able to persist in their current environments. However, a missing piece of the
63 puzzle remains; in order to assess the adaptive potential of a population we need to understand
64 how selection on populations might shift with climate change. Once we understand how
65 selection is likely to shift, we can then identify populations which may have a higher adaptive
66 potential than others in the face of climate change. Here I first ask how selection on *Picea*
67 *mariana* shifts with climate in the Canadian boreal forest and then ask whether different *P.*
68 *mariana* provenances have different adaptive potentials to warmer and drier environments.

69 Over the next 100 years, the climate of the boreal forest is expected to get warmer and drier
70 (Figure 1). Current emission trends suggest that continued increase in greenhouse gas emission
71 over time is likely. Trends modelled using this climate scenario (CanESM2 RCP 8.5 model)
72 show warming temperatures across the boreal forest, with most regions experiencing a ~ 4 °C
73 increase in mean annual temperature (Boucher *et al.*, 2020). In addition to warming, water
74 availability in environments is predicted to decrease. Water availability in environments depends
75 on the interplay between evaporation, due to temperature and irradiation, and levels of
76 precipitation. Water availability is reflected by the climate moisture index (CMI), which is the
77 difference between annual precipitation and potential evapotranspiration. While a decrease in
78 CMI is predicted across most of the boreal forest (with the exception of the most eastern
79 regions), in the western regions CMI is expected to reach negative values, which indicate that a

80 closed canopy forest would not be maintained in this region. Yet exactly how these shifts in
81 environmental conditions will affect boreal tree species remains unclear.



82

83 **Figure 1.** Projections of annual mean temperature (°C) and Climate Moisture Index (CMI) for
84 the 1981 – 2010 and 2071 – 2100 periods based on the CanESM2 RCP 8.5 model. Grey line
85 represents the range of the Canadian boreal forest. Figure adapted from Boucher *et al.* (2020).

86 In the boreal forest the response of *P. mariana* to warming has not been uniform, with
87 warming expected to increase the growth rates of trees in the northern boreal forest where water
88 is not limiting, and southern regions expected to face increased drought stress and thus lower
89 growth rates and higher mortality (Sniderhan *et al.*, 2021). Indeed, many studies have found
90 productivity to increase in response to warming (Gamache & Payette, 2004) and many others
91 have found it to decrease (Hogg *et al.*, 2005; Girardin *et al.*, 2016; Robert *et al.*, 2024). An
92 advance of the timing of spring since 1950 has been documented for most of North America
93 (Cayan *et al.*, 2001; Sniderhan *et al.*, 2021) however for southern latitudes, productivity is more

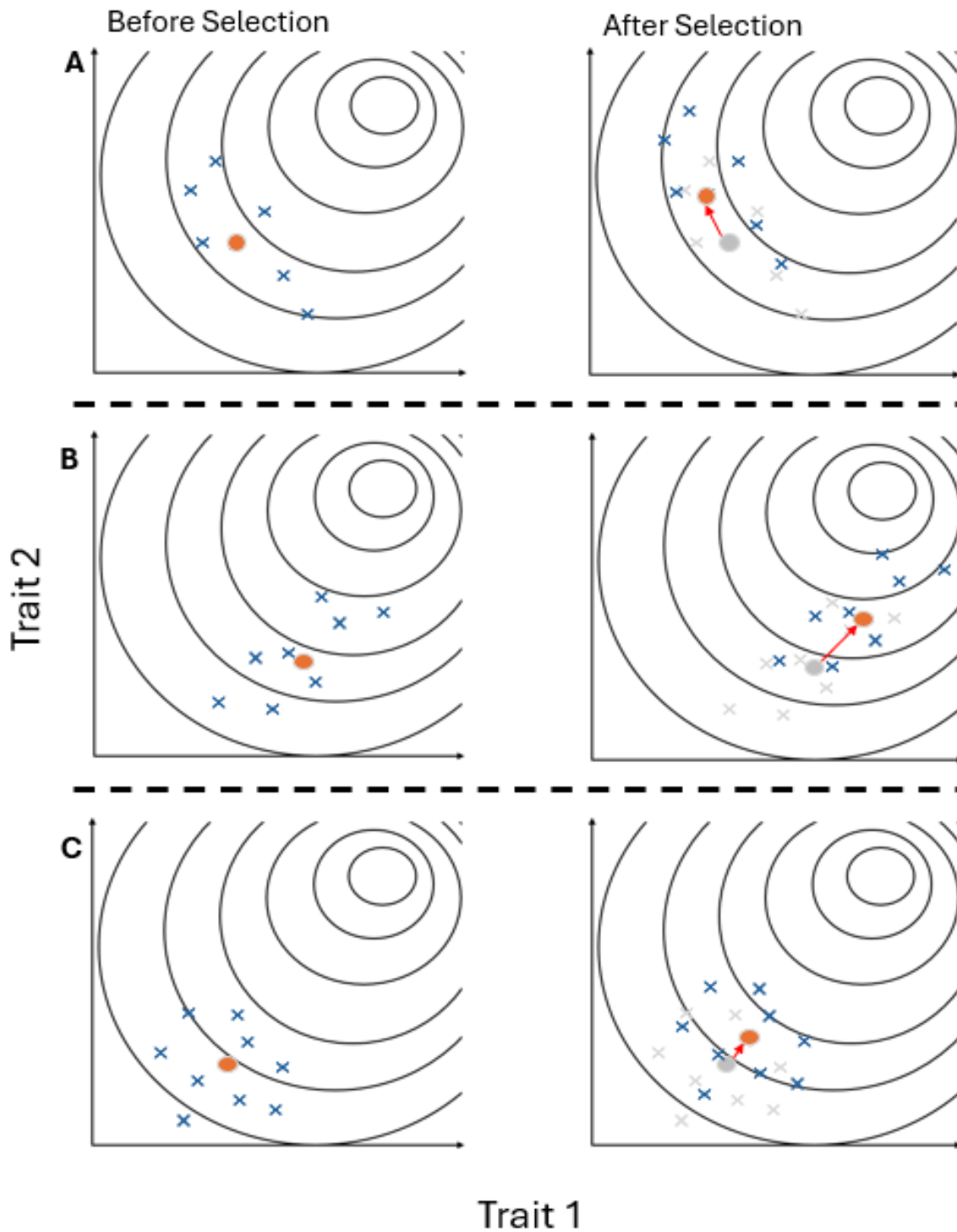
94 water limited (Chagnon *et al.*, 2022). Changes in temperature can change the length of the
95 growing season, an important determinant of forest productivity in temperate and boreal forests
96 (Kimball *et al.*, 2004). Water limitation in the southern regions of the boreal forest can be further
97 exacerbated by increases in temperature increasing water loss from the environment which
98 negatively impacts growth and increases mortality from the leaves in dryer sites, unless stomata
99 close (Shaw *et al.*, 2005; Peng *et al.*, 2011; Choat *et al.*, 2018; Chagnon *et al.*, 2022; Münchinger
100 *et al.*, 2023). Research on the impact of climate change on *P. mariana* focuses mainly on
101 changes in growth. Looking at growth integrates the response of the entire phenotype and is thus
102 informative of the overall impacts of climate change on individual- and population-level
103 performance, but we do not know which traits are driving this response. Here, I address this by
104 measuring which traits are under selection in a natural population and are thus driving this
105 change in growth response.

106 I expect selection from warmer and dryer climates to act on traits related to water use,
107 photosynthesis, structural support and temperature regulation (Aubin *et al.*, 2016; Boisvert-
108 Marsh *et al.*, 2020; Sniderhan *et al.*, 2021). Water is required in the photosynthetic pathway and
109 is the main chemical component of live plant cells. It is also required to maintain turgor and
110 therefore support the softer tissues, as well as to transport the nutrients across the plant tissues.
111 Low water availability in environments has been shown to decrease forest productivity, as such,
112 trees have shown adaptations to cope with this environmental stress. Therefore, in environments
113 with low water availability traits representing water conservation strategies are likely to confer
114 higher performance and be the targets of selection. Trees must balance resource allocation
115 between growth, reproduction, and structural support. A portion of the tree's energy is invested
116 in building strong wood and bark to support height and canopy spread. Temperature limited trees

117 exposed to warming temperatures may develop weaker wood. This is because elevated
118 temperatures accelerate growth and as a consequence reduces investment in structural support
119 leading to lower wood and branch density. This makes them more vulnerable to break under
120 mechanical stress, such as during storms (Ahrens *et al.*, 2020). Temperature (heat) controls the
121 rate of plant metabolism, which in turn determines the rate of photosynthesis. Most biological
122 metabolic activity takes place within the range of 0–50° C (Hopkins & Hüner, 2004). The
123 optimal tree productivity coincides with 15–25 °C, which is the optimal range of photosynthesis
124 (Hopkins & Hüner, 2004). Temperatures become lethal over 44°C (Schulze *et al.*, 2002).
125 However, it is important to note that plants have been shown to thermoregulate, whether it is to
126 maintain stable tissue temperatures or to produce heat to attract pollinators, there is evidence that
127 plant temperatures are not just passively tracking ambient temperatures. The “leaf homeothermy
128 hypothesis” states that specific suites of leaf traits have evolved to buffer variation in
129 temperature and maintain leaf temperatures within a narrower range of variation around the
130 metabolic optima for photosynthesis (Michaletz *et al.*, 2015). In theory, the warmer an
131 environment is, the more cooling the leaves will need to maintain this metabolic optimum. As
132 such, in the range of temperatures that decrease plant metabolic activity, I expect enhanced leaf
133 cooling to confer higher performance and thus be a target of selection in such an environment.

134 In addition to the adaptive value of individual traits, phenotypic integration may affect
135 population response to selection. Specifically, when the phenotype is highly integrated, (i.e. trait
136 covariance is strong) then the response to selection of one trait will be affected by selection on
137 the other trait. A population’s response to climate change can either facilitate or impede the
138 adaptive evolution of the population depending on the magnitude and direction of trait
139 covariance. Adaptive evolution is constrained when trait covariances are strong such that the

140 response to selection is determined largely by the direction of maximum trait covariation rather
141 than the direction of selection (Figure 2A) (Björklund, 1996; Schluter, 1996). When integration
142 is strong and the direction of maximum trait covariation conflicts with the direction of selection,
143 then the response to selection is weak. However, when phenotypic integration is strong and
144 selection is aligned with the direction of maximum trait covariation, then the response to
145 selection can be greater than in a population with no integration present (Figure 2B) (Björklund,
146 1996). Last, when there is no covariance between traits under selection, the trait values of the
147 populations can evolve independently (Figure 2C). Provided that adequate heritable trait
148 variation is present in the population, this can benefit populations as their response to climate
149 change can proceed unconstrained by trait covariation (Via & Lande, 1985).



150

151 **Figure 2.** Hypothetical fitness landscapes for trait covariance that impedes (A) facilitates (B) or
 152 leaves response to selection unconstrained (C). Contour lines represent the topography of the
 153 fitness landscape with the top right corner representing the adaptive peak.

154 *P. mariana* is well adapted to tolerate cold environments, it is one of the species which marks
155 the northern tree line of Canada and is particularly well adapted to waterlogged, poorly drained
156 and rich organic soils (Burns, 1990), which contrast with the warming and drying climate.
157 However, this generalist species grows on a wide range of soil conditions and is present along
158 the entire range of the boreal forest (Burns, 1990). It is thus unclear how this cold- and wet-
159 adapted species will persist in warmer and drier conditions that are expected to accompany
160 climate change (Robert *et al.*, 2024).

161 Here I ask, how does selection on *P. mariana* change along a spatial climate gradient, and do
162 different *P. mariana* populations have trait covariances that accelerate or impede response to
163 selection in a warm and dry environment? In each of three common garden sites which span a
164 spatial temperature and water availability gradient, I studied seven provenances from across *P.*
165 *mariana*'s range. I first assess for each site the strength and direction of selection on a set of
166 traits associated with plant response to temperature and drought and then I assess which of the
167 seven provenances, if any, have higher potential for adaptive evolution in the warmest and driest
168 site.

169

170 **METHODS**

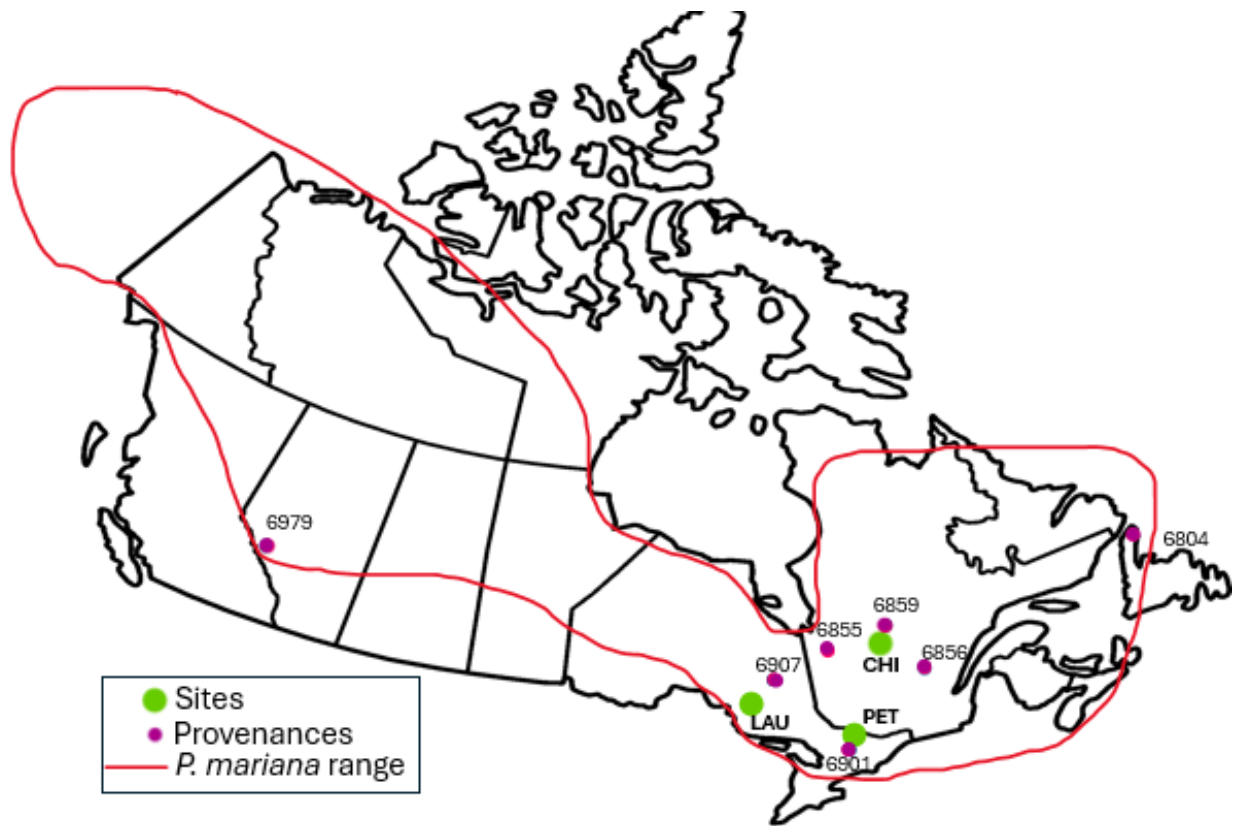
171 **STUDY SITES**

172 This research makes use of three sites from a range wide *P. mariana* provenance trial
173 started in 1967 (Morgenstern and Kokocinski, 1976). Sites included in my research were selected
174 based on the following criteria: sites containing the same planted provenances, sites including
175 provenances that span as much of *P. mariana*'s range as possible, and sites which fall along a
176 spatial temperature and water availability gradient.

177 The three sites selected are located near Petawawa, Ontario (PET); Chapleau, Ontario
178 (LAU); and Chibougamau, Quebec (CHI). Five provenances are common to all three sites and
179 two provenances, (6856 and 6979) are common to PET and CHI only. The provenances included
180 in this research span the southern latitudes of the boreal forest, from Newfoundland to Alberta
181 (Figure 3). The sites selected fall along a spatial temperature and water availability gradient
182 (Figure 4). Site climates were defined with BioSIM data using the yearly averages from 1980-
183 2020 (Fortin, 2022).

184 The warmest and driest of the three sites, PET, is characterized by a mean annual
185 temperature (MAT) of 4.7 °C and a climate moisture index (CMI) of -1 (Table 1). The
186 intermediate site, LAU, has moderate MAT (1.5 °C) and CMI (9). The coldest and dampest site,
187 CHI, is characterized by a MAT of -0.4 °C and a CMI of 27. In addition to temperature and water
188 availability, sites were characterized by soil water holding capacity (WHC), total annual
189 precipitation (TAP), spacing between trees, and survival of provenances (Table 1).

190 Each provenance trial was set up with at least three blocks. Within a block, 16 trees per
191 provenance were planted in a 4x4 grid forming small plots with consistent spacing between the
192 trees (Morgenstern and Kokocinski, 1976) (Figure 5). Spacing between trees differed among
193 sites (Table 1).



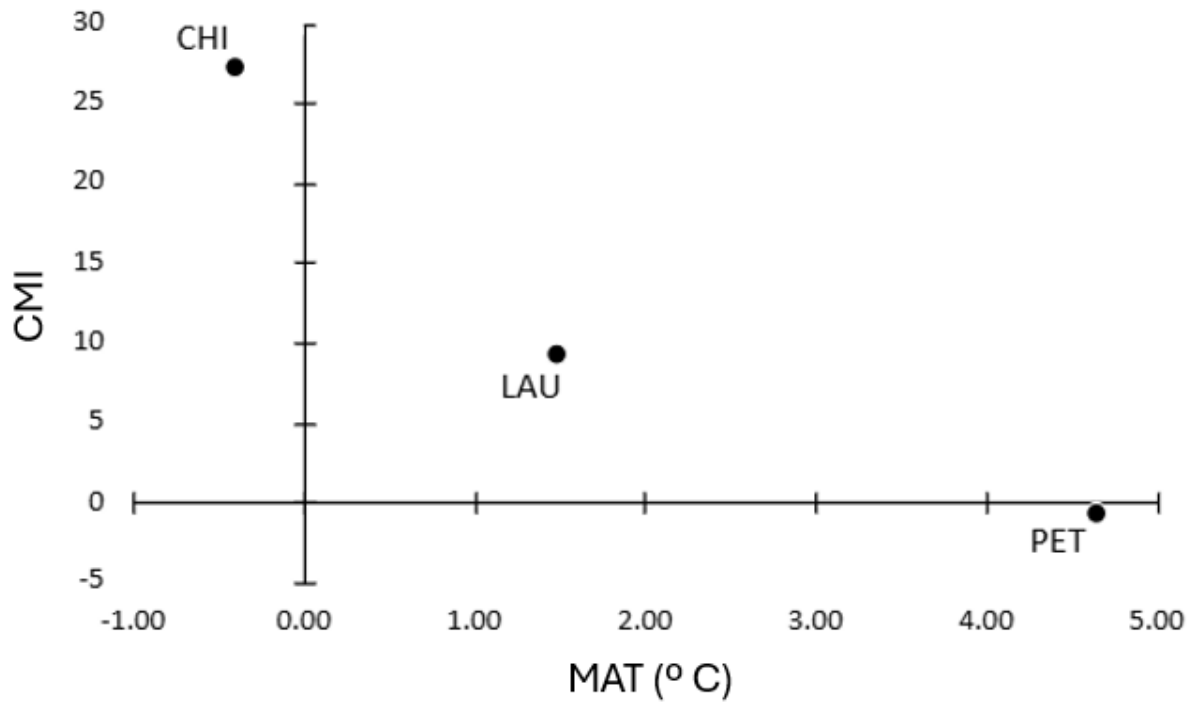
194

195 **Figure 3.** Location of each site (green) and provenance (purple) used in this study. Approximate
 196 range of the boreal forest shown in red.

197

198

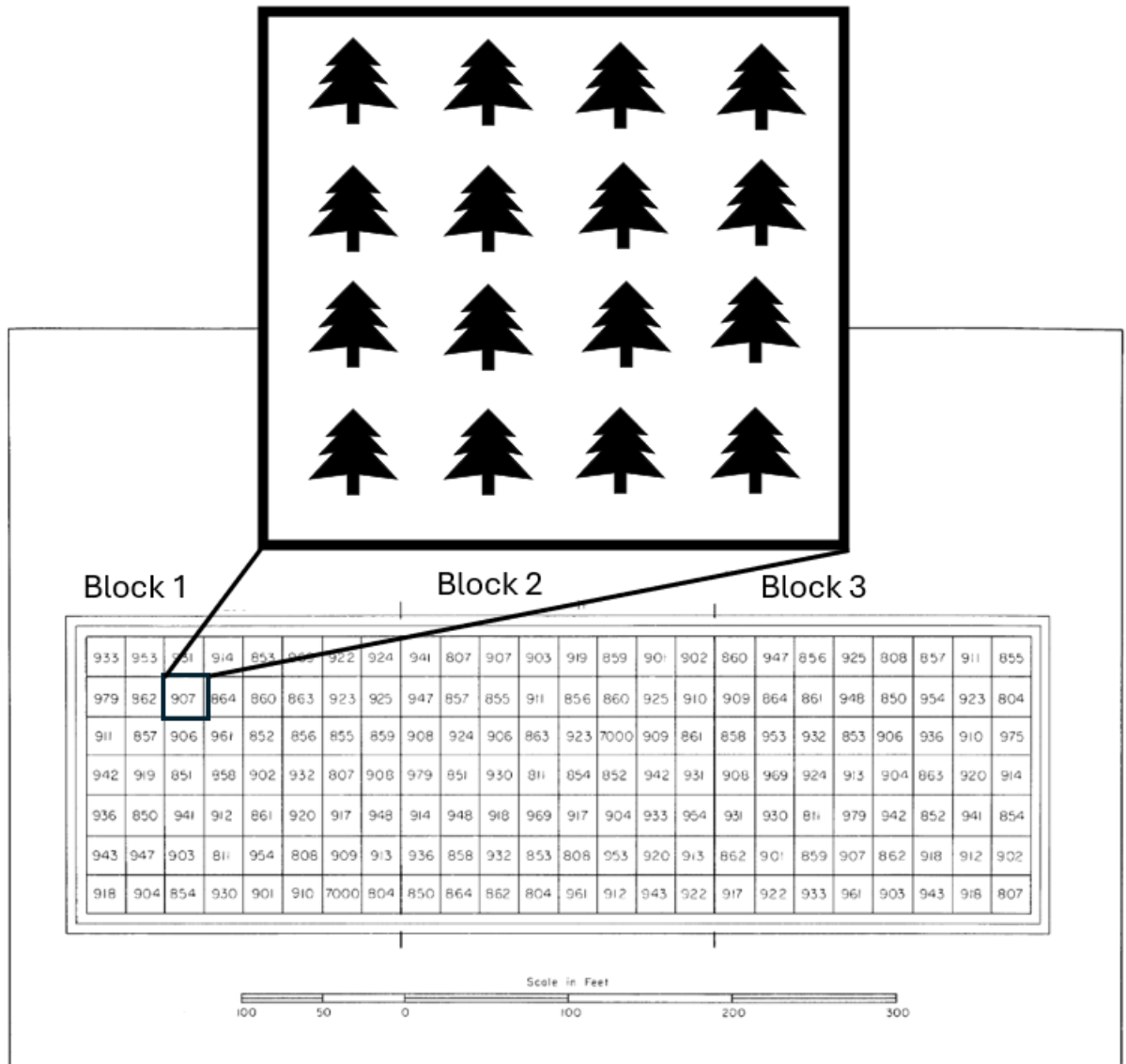
199



200

201 **Figure 4.** Visualization of the temperature (MAT) and water availability (CMI) between the sites
202 used in this study.

203



204

205 **Figure 5.** Representation of provenance trial experimental set up (Morgenstern & Kokocinski,
 206 1976). Numbers represent provenance IDs. Each provenance is planted once per block in a 4x4
 207 grid of 16 trees.

208

209

210

211 **Table 1.** Table of site characteristics for PET, LAU and CHI.

Characteristic	PET	LAU	CHI
Mean Annual Temperature (°C)	4.7	1.5	-0.4
Climate Moisture Index	-0.9	9.0	27.2
Total Annual Precipitation (mm)	841.6	842.3	975.5
Water Holding Capacity (%)	77.7	78.1	62.4
Spacing between trees (m)	1.8 x 1.8	1.8 x 1.8	2.4 x 3
Survival	42 % (n = 336)	65 % (n = 240)	78 % (n = 672)

212

213 **GROWTH DATA**

214 Height data was collected by Natural Resources Canada in 2022 for PET, 2023 for LAU and
 215 2016 for CHI. The height of each individual tree was measured using a secant scale clinometer
 216 (Suunto, PM-5), commonly used in forestry to determine stand height. Relative growth rate
 217 (RGR; m/yr) over an individual's lifetime was calculated as the height of the tree divided by the
 218 age at the time of measurement.

219 **TRAIT DATA**

220 Individual trees were sampled from at least three blocks at each site. Branch samples were
 221 collected from PET in July 2022 and May 2023 (n = 143), from LAU in July 2023 (n = 156) and
 222 from CHI in July 2022 (n = 132). Since I only considered fully mature needles and some samples
 223 were not yet mature at the time of collection, needles that emerged during the year of collection
 224 (yr 0) were not considered. For tree branch sampling, it is common practice to sample the most

225 sun-exposed branches of a tree in order to standardize the light environment however, the height
226 and geometry of many trees prevented the collection of branches from the top of the crown.
227 Therefore, I standardized for branch position in the crown. Each sampling morning, branches
228 containing at least four years of complete growth were cut from the bottom of the top third of
229 foliar branches using a 13.7 m telescopic pole pruner (HV-245 Tel-O-Pole). To keep the
230 branches hydrated, the cut ends were placed in florist tubes filled with water. The branches were
231 then placed in plastic bags with a damp paper towel, and stored in a cooler with ice until time-
232 sensitive traits were measured indoors in the afternoon (Garnier *et al.*, 2001).

233 I measured 10 traits associated with tree response to temperature, water availability, or
234 both. Three traits, Huber value (HV), water use efficiency (WUE) and needle carbon to nitrogen
235 ratio (CN) were measured at the branch level. The other seven traits were measured for each of
236 the four most recent years of mature growth. Branch level trait values were calculated from year-
237 level trait values by taking an abundance-weighted average. The relative abundance was
238 calculated from dry biomass, with needle biomass for needle traits and twig biomass for twig
239 traits. For example, the branch-level mean needle length (NL) was weighted by the total dry
240 needle biomass for each year of growth and twig specific density (TSD) was weighted by the
241 total dry twig biomass for each year of growth. Traits were classified by association with
242 structure, photosynthesis, or water use in plants (Table 2).

243 To characterize investment in structural support at the needle and branch levels, leaf dry matter
244 content (LDMC; g/g) and twig specific density (TSD; g/cm³) were measured (Poorter *et al.*,
245 2010; Bartlett *et al.*, 2012). The LDMC of 12 needles from each growth year was measured as
246 needle dry mass divided by fresh mass, measured using analytical balances (10⁻⁴ g AG104 Metler
247 Toledo from Switzerland, and 10⁻⁶ g XSR205 Metler Toledo, as needed). Needles were dried

248 using a forced air oven at 60 °C to a constant weight or for a minimum of 72 hours. To measure
249 TSD, after removing the needles from the branch, the volume was measured for three fresh twigs
250 per growth year. Since the twigs varied in size within a growth year, the three twigs were
251 randomly sampled from a subset excluding the largest and smallest sizes. Twig fresh volume was
252 measured using the water immersion method (Sukul *et.al.*, 1993). After fresh measurements, the
253 twigs were dried at 60 °C to constant weight for at least 72 hours and dry weight for three twigs
254 was measured using analytical balances (10⁻⁴ g AG104 Metler Toledo from Switzerland, and 10⁻⁶
255 g XSR205 Metler Toledo, as needed). TSD was calculated for each of the three branches as twig
256 dry mass (g) divided by twig fresh volume (cm³) and averaged. Additionally, total dry twig
257 biomass was measured for each growth year and used for abundance-weighted trait averaging.

258 Huber value (HV; mm/g), the ratio of sap-wood diameter to needle weight, was measured to
259 characterize water supply per water demand at the branch level (Carter & White, 2009). Two
260 perpendicular diameter measurements were taken at the base of the fourth oldest year of
261 complete growth using electronic calipers (±0.01 cm, Series 500 ABSOLUTE Digimatic
262 Caliper) and averaged. All needles were removed from the branch and dried at 60 °C to constant
263 weight. The total mass of needles for each branch was measured with the Mettler Toledo AG104
264 Analytical Balance. A high value for HV indicates a liberal water supply per needle weight
265 whereas a low value for HV reflects conservative water supply per needle weight. HV was
266 calculated as branch base diameter (mm) divided by the dry mass of the needles supplied by that
267 branch (g).

268 To characterize water use and regulation at the needle level, water use efficiency (WUE; ‰) and
269 stomatal density (SD; count/cm) were measured. To measure WUE, I used carbon isotope ratio
270 ($\delta^{13}\text{C}$), which is commonly used to estimate intrinsic water use efficiency (Ma *et al.*, 2023).

271 During photosynthesis, $^{12}\text{CO}_2$ is preferentially fixed over $^{13}\text{CO}_2$ for various reasons, the main one
272 being that the enzyme RuBisCO discriminates against $^{13}\text{CO}_2$ and only binds it when $^{12}\text{CO}_2$ is
273 limited, which occurs when stomata are closed (Farquhar *et al.*, 1982; Lambers *et al.*, 2008). The
274 ratio of ^{13}C to ^{12}C present in plant tissue, therefore reflects the amount of time leaves spent with
275 their stomata closed, and $\delta^{13}\text{C}$ gives the ^{13}C isotope composition relative to the primary reference
276 scale of Vienna Pee Dee Belemnite. Needles from all growth years were combined and ground
277 into a fine powder using a ball mill grinder to ensure a homogenous mixture. Analysis of carbon
278 isotopes was performed at the Environmental Isotope Laboratory at the University of Waterloo.
279 The analysis for ^{13}C isotope measurements was done by combustion conversion of sample
280 material to gas through a 4010 Elemental Analyzer (Costech Instruments, Italy) coupled to a
281 Delta Plus XL (Thermo-Finnigan, Germany) continuous flow isotope ratio mass spectrometer
282 (CFIRMS). To determine the homogeneity of the ground needle samples, duplicate
283 measurements were performed for 34 samples, spaced at regular intervals throughout the
284 measurement process. To measure SD images of needle abaxial surface were taken using Leica
285 EZ4 W stereo microscope at 35X magnification and the microscope imaging software LAS X for
286 Life Sciences (Leica Microsystems, 2021). Since stomata were not clearly visible over the full
287 length of the needles, the number of stomata along the maximum length possible was recorded
288 along with stomata count using WinSeedle Software (Regent Instruments, 2020).

289 Needle cooling (NC; °C), the needle temperature relative to ambient temperature, was measured
290 to assess needle thermal regulatory ability. Under the ‘limited leaf homeothermy’ hypothesis
291 leaves buffer environmental variation by controlling internal leaf temperature to maintain the
292 optimal temperature range for the photosynthesis reaction (Michaletz *et al.*, 2015). These
293 measurements were taken while the branches were secured in a florist tub full of water to

294 maintain needle transpiration. Needle temperature was measured indoors, on-site, a few hours
295 after sample collection, using a thermal camera, TCAM-300 (Infrared Camera INC, 2021). To
296 validate the thermal camera's measurements, a high accuracy temperature sensor (TS, ± 0.1 °C,
297 TMP117 High-Accuracy, Low-Power, Digital Temperature Sensor) was included in each image.
298 The sensor plate of the TS was placed inside each thermal image and its temperature was
299 recorded when each thermal picture was taken. The difference between the temperature recorded
300 on the TS and the temperature of the sensor plate on the thermal camera was used as an
301 adjustment for all temperature readings in each thermal image. Needle cooling was measured for
302 each of the four most recent complete years of growth and was calculated as needle temperature
303 minus ambient temperature.

304 To characterize photosynthetic ability of the needles, the concentration of chlorophyll
305 (CHL; mg/m²) was measured using a chlorophyll content meter: CCM-300 (Opti-Sciences). The
306 CCM-300 uses the fluorescence ratio technique to measure total chlorophyll content (Gitelson *et*
307 *al.*, 1999). A few hours after sample collection, CHL was measured for healthy needles from
308 each of the four most recent years of mature growth.

309 Leaf mass per area (LMA; mg/cm³) represents the leaf level cost of light interception and in
310 global interspecific comparisons is an important indicator of plant carbon-use strategies
311 (Gutschick & Wiegand, 1988; Grime, 2001; Westoby *et al.*, 2002). To measure needle fresh area
312 (cm³), 12 needles from each of the four most recent years of complete growth were haphazardly
313 selected and scanned using the STD4800 Scanner for WinRhizo and WinSeedle and area was
314 measured using WinSeedle software (Regent Instruments, 2020). Needles were dried in a VWR
315 Forced Air Incubator at 60 °C to a constant weight for a minimum of 72 hours and needle dry
316 weight was measured using analytical balances (10⁻⁴ g AG104 Metler Toledo from Switzerland,

317 and 10^{-6} g XSR205 Metler Toledo, as needed). LMA was calculated for each growth year as the
318 weight of the 12 needles over their area.

319 Needle length (NL; mm) is an indicator of the size of the needle and affects a plant's light
320 capture, water balance, and thermal stability (Perez-Harguindeguy *et al.*, 2013). To measure NL
321 needles were scanned using the STD4800 Scanner for WinRhizo and WinSeedle and the length
322 of the needle was measured using WinSeedle software (Regent Instruments, 2020). NL was
323 measured for 12 needles from each of the four most recent full years of growth and the average
324 length of the needles was calculated for each growth year.

325

326

327

328

329

330

331

332

333

334

335

336

337 **Table 2.** Traits measured, abbreviations, units, and associated physiological function(s).

Trait	Unit	Function
Leaf dry matter content (LDMC)	g g^{-1}	Structural investment
Twig specific density (TSD)	g cm^{-3}	Structural investment
Huber value (HV)	$\text{mm}^2 \text{g}^{-1}$	Water use and transport
Water use efficiency (WUE)	%	Water use and transport
Stomata density (SD)	count cm^{-1}	Water use and transport
Needle cooling ability (NC)	$^{\circ}\text{C}$	Photosynthesis
Chlorophyll concentration (CHL)	mg m^{-2}	Photosynthesis
Needle carbon to nitrogen ratio (CN)	g g^{-1}	Photosynthesis
Leaf mass per area (LMA)	mg cm^{-3}	Structure + Photosynthesis
Needle length (NL)	mm	Structure + Photosynthesis + Water use and transport

338

339 **STATISTICAL ANALYSIS**

340 All statistical analyses were conducted in R version 4.4.0 (R Core Team, 2024). Two percent of
 341 all data was missing due to lost samples or measurement errors. In PET, 3% of the data was
 342 missing, in LAU, <1% of the data was missing and in Chibougamau, 3% of the data was
 343 missing. Missing values were imputed for each site using linear predictive models. Traits as
 344 predictors which yielded models with the highest predictive power for missing values were

345 identified using the “dredge()” function from the “MuMIn” package. Data were only imputed
346 when the prediction model R^2 value was above 0.6. Otherwise, the missing values were left as
347 NA. After imputation, there was less than 0.01% data missing from each site for sample sizes of
348 $n = 142$ trees in PET, $n = 151$ trees in LAU, and $n = 132$ trees in CHI.

349 To identify significant differences in survival among sites and among provenances within sites
350 one-way ANOVAs were used. Due to missing provenances in LAU (6956 and 6979) a two-way
351 ANOVA to accommodate both Site and Provenance as factors was not possible. Principal
352 component analyses (PCAs) were conducted on trait data within each site using the “PCA()”
353 function of the “FactoMineR” package to assess trait covariance strength and structure.

354 Standardized selection gradients were measured for each site by regressing relativized RGR onto
355 all 10 standardized traits and their two-way interactions using multiple linear regression models
356 using the function “lme()” from the “nlme” package (Connor 1988). Traits were standardized by
357 taking the z-scores for all trait values within each site. Additionally, RGR was relativized by
358 dividing RGR values within a site by the mean RGR value for that site. In this method, the
359 partial regression coefficients represent the strength and direction of selection, referred to as
360 selection gradients (β) (Lande and Arnold, 1983). To control for the effect of different lineages,
361 provenance was included as a random factor in each model.

362 Differences in selection between the two most extreme sites, PET and CHI, were assessed
363 using t-tests to determine statistical significance using the “tsum.test()” function from the
364 “BDSA” library. Additionally, selection gradients that changed directionally with the climate
365 gradient were interpreted as being driven by the climate gradient.

366 In order to test whether trait integration was aligned with the direction of selection,
367 significant selection gradients in PET were used to create performance surfaces (trait1 x trait2 x
368 RGR) using the “geom_contour()” function in “ggplot” library. The performance surface is a
369 topographic map of selection and allows us to visualize the peaks and valleys of performance in
370 relation to traits. Smooth contour lines were generated using the “interp()” function from the
371 “akima” library. This helped to remove noise from the data so trends in the performance surface
372 could be better identified. For each provenance, significant correlations between the residuals of
373 the traits were determined using the Pearson Correlation Coefficient from the “cor.test()”
374 function from the “stats” library. The residuals of the traits from the regression models were
375 collected using the “residuals()” function from the “stats” library. Significant correlations
376 between phenotypic traits were later assessed to determine if integration might facilitate a
377 provenance’s response to selection.

378

379

380

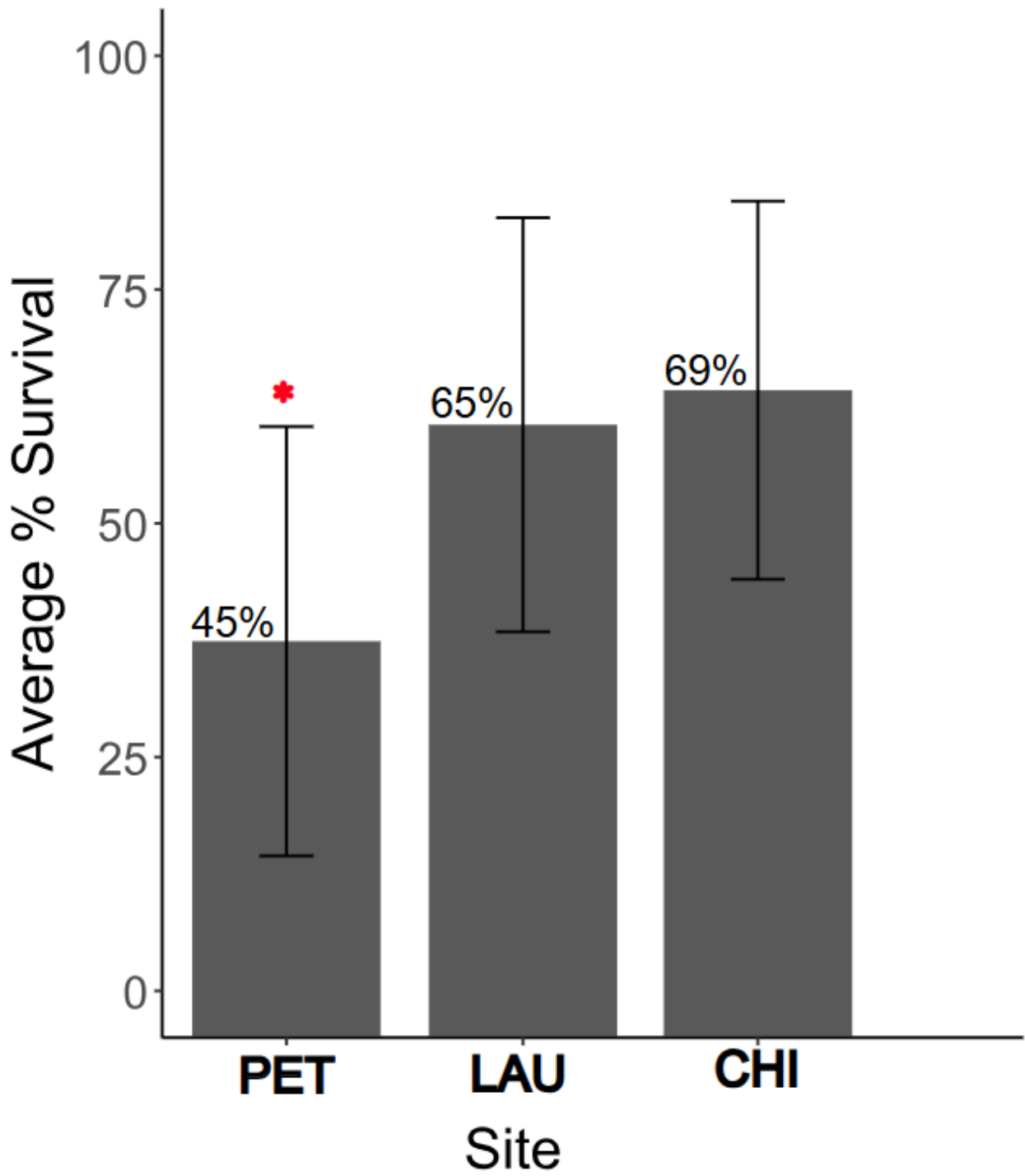
381

382 **RESULTS**

383 **SURVIVAL**

384 Average survival was significantly lower in Petawawa (42%) than in the other two sites
385 (LAU: 65%, CHI: 69%) (Figure 6). In terms of differences in provenance survival, in Petawawa
386 there was no significant difference among the survival of provenances (One-way ANOVA,
387 $F_{(6,14)} = 1.113$, $p = 0.403$). However, in Chapleau and Chibougamau the survival of some

388 provenances differed. In Chapleau, it was significantly lower in provenance 6907 (39.58%) than
389 in provenance 6855 and 6859 (83.33% and 79.17% respectively; One-way ANOVA, $F_{(4,10)} =$
390 3.491, $p = 0.0495$). In Chibougamau provenance 6979 had significantly lower average survival
391 (45.00%) than provenances 6855, 6856 and 6907 (83.75%, 77.50%, and 77.50% respectively)
392 (One-way ANOVA, $F_{(6,28)} = 2.492$, $p = 0.0466$).

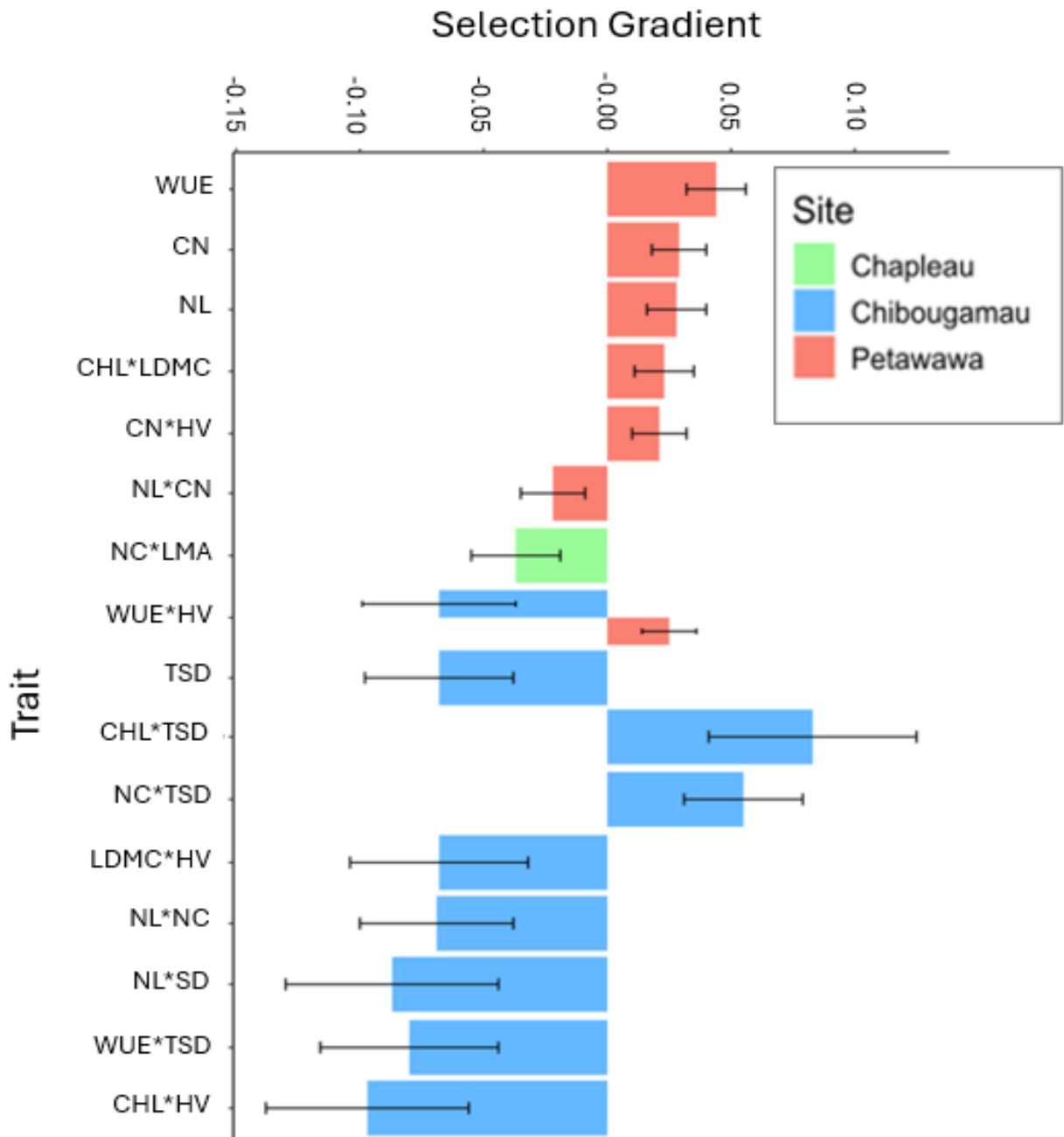


395 **Figure 6.** Average percent survival of individual trees in each of the three study sites. Red star

396 represents significantly different site (PET: n = 336, LAU: n = 240, CHI: n = 672).

397 **TRAIT SELECTION ALONG THE CLIMATE GRADIENT**

398 All 10 traits measured experienced significant selection in at least one site, either alone or in
399 interaction with another trait, hereafter referred to a trait interaction and represented by
400 trait1*trait2 (Figure 7). Many traits were under selection, with a total of 17 significant selection
401 gradients across all 3 sites. Across sites, most of the selection gradients (β) were for trait
402 interactions (12 out of 17). In Petawawa, most β were positive (6 out of 7) and in Chibougamau
403 most β were negative (7 out of 9). Here a positive trait interaction β indicates that individuals
404 with high performance are associated with high values of both traits. For example, in PET high
405 CHL is associated with high performance only in individuals that also have high LDMC. A
406 negative trait interaction β indicates that individuals with high performance are associated with a
407 high value in the first trait combined with low values in the second trait, and vice-versa (low
408 value in the first trait combined with a high trait value for the second trait). For example, in PET
409 individuals with high performance either had high NL and low CN values, or low NL and high
410 CN values. The traits and trait interactions under selection differed among sites. In Petawawa,
411 three traits were under significant or marginally significant selection: WUE ($\beta = 0.044$, $p < 0.01$)
412 CN ($\beta = 0.029$, $p = 0.10$) and NL ($\beta = 0.028$, $p = 0.17$). Four trait interactions were under
413 significant or marginally significant selection: WUE*HV ($\beta = 0.025$, $p = 0.01$) CHL*LDMC (β
414 = 0.023, $p = 0.06$), CN*HV ($\beta = 0.021$, $p = 0.06$) and NL*CN ($\beta = -0.022$, $p = 0.01$) (Figure 7).
415 In Chapleau selection only acted on LC*LMA ($\beta = -0.037$, $p = 0.04$) (Figure 7). In CHI only one
416 trait was significantly under selection TSD ($\beta = -0.068$, $p = 0.03$) and 8 trait interactions were
417 under significant or marginally significant selection: CHL*HV ($\beta = -0.097$, $p = 0.02$), CHL*TSD
418 ($\beta = 0.083$, $p = 0.05$), NL*LC ($\beta = -0.069$, $p = 0.03$), NL*SD ($\beta = -0.087$, $p = 0.05$), LC*TSD (β
419 = 0.055, $p = 0.03$), WUE*HV ($\beta = -0.068$, $p = 0.03$) and WUE*TSD ($\beta = -0.080$, $p = 0.03$) and
420 LDMC*HV ($\beta = -0.068$, $p = 0.07$).



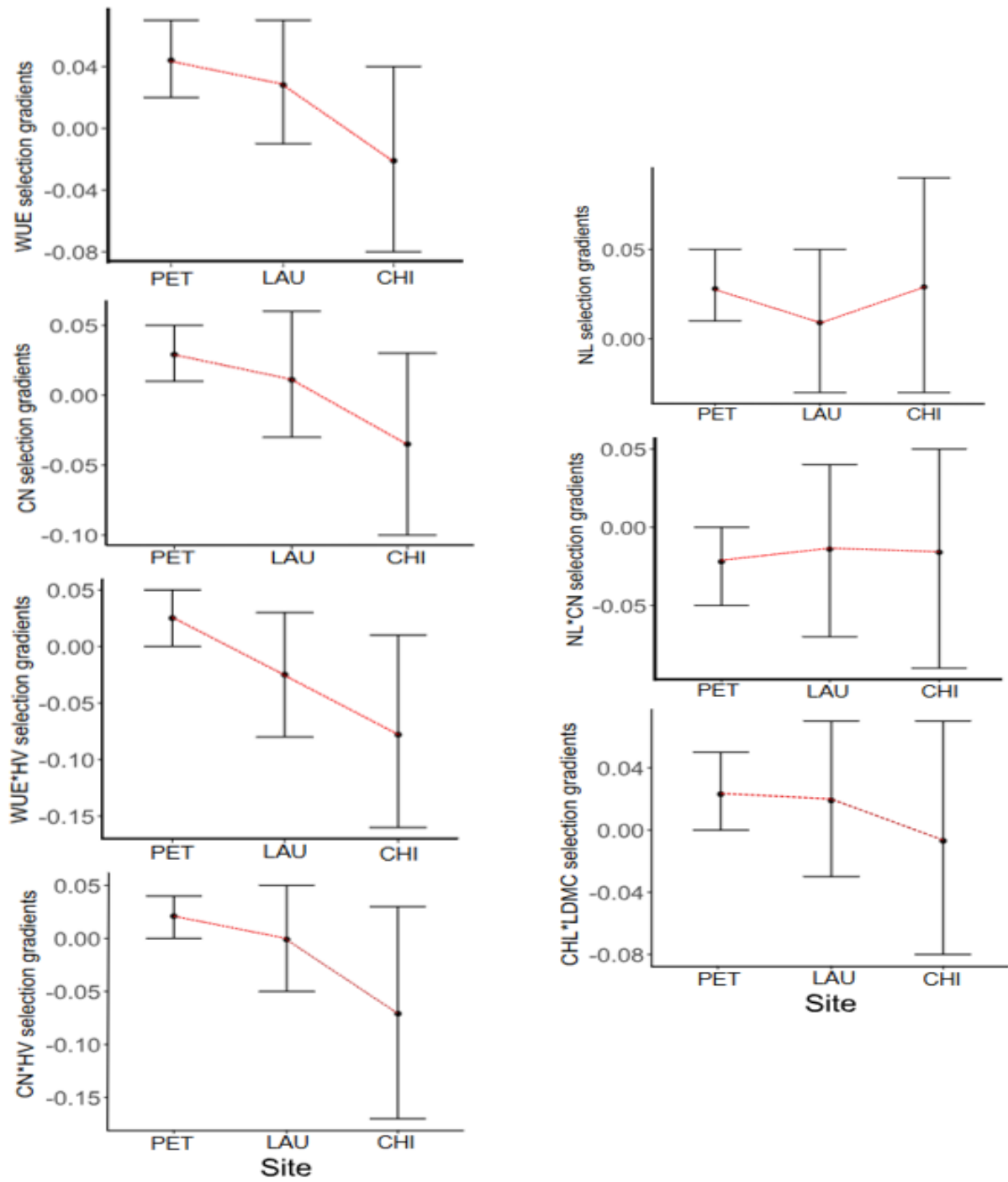
421

422 **Figure 7.** Significant and marginally significant selection gradients (β) for each of the three sites.

423 Error bars represent standard error.

424 **DIFFERENCES IN SELECTION ASSOCIATED WITH CLIMATE**

425 To identify differences associated with climate, I identified the β changing directionally with
426 the climate gradient among the three sites. Here I make the assumption that changes that occur
427 directionally along our climate gradient are likely due to climate. While I acknowledge that there
428 are many factors that vary among natural environments, directional changes along our gradient
429 are most likely due to differences in climate, because the other factors known to differ among the
430 sites do not differ directionally (Table 1). Selection gradients in Petawawa that varied
431 directionally with climate are WUE, CN, WUE*HV, and CN*HV (Figure 8). The selection
432 gradients for WUE and WUE*HV significantly differed between PET and CHI (two-sample t-
433 test, $p = 0.046$; two-sample t-test, $p = 0.025$, respectively; Figure 8). The selection gradients for
434 CN and CN*HV were marginally different between PET and CHI (two-sample t-test, $p = 0.060$,
435 $p = 0.092$; Figure 8)



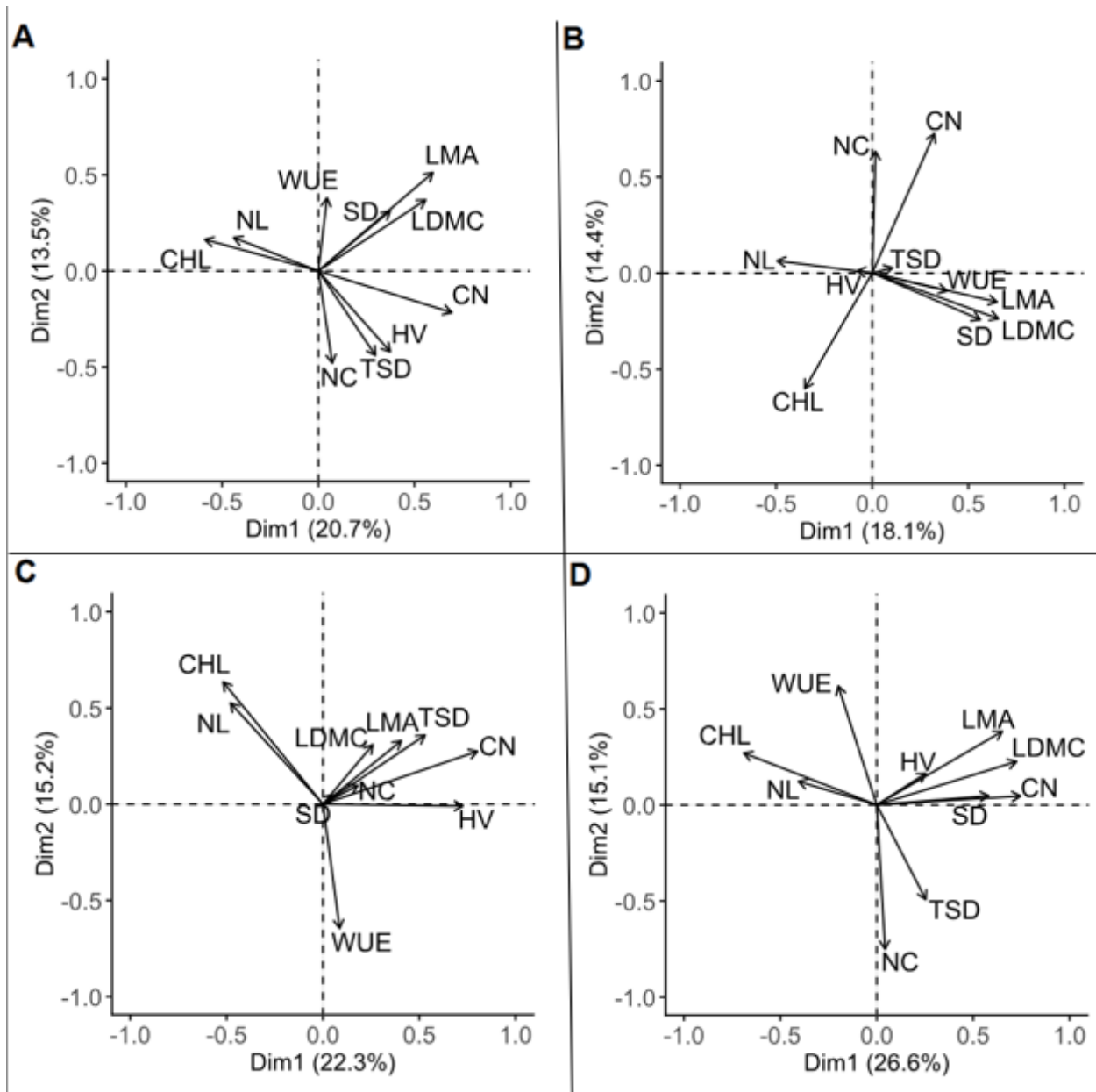
436

437 **Figure 8.** 95% Confidence intervals of β s in each site for β s under significant selection in

438 Petawawa.

439 **PHENOTYPIC INTEGRATION IN *P. MARIANA* PROVENANCES**

440 In general, traits showed weak covariation. For the experiment wide PCA on trait values,
441 together the first two principal components accounted for only 34.1% of total trait variation
442 (Figure 9). The site-specific PCAs for PET and LAU are different from each other and from the
443 experiment wide PCA. The amount of variance explained by the first two axes of the site-
444 specific PCA remained low (PET: 32.5%; LAU 37.5%; CHI: 41.6%) (see Supplementary
445 Information Appendix S1 Table S3 for loadings).



446

447 **Figure 9.** PCA of trait values among all the sites as well as for individual sites. Tell us what the

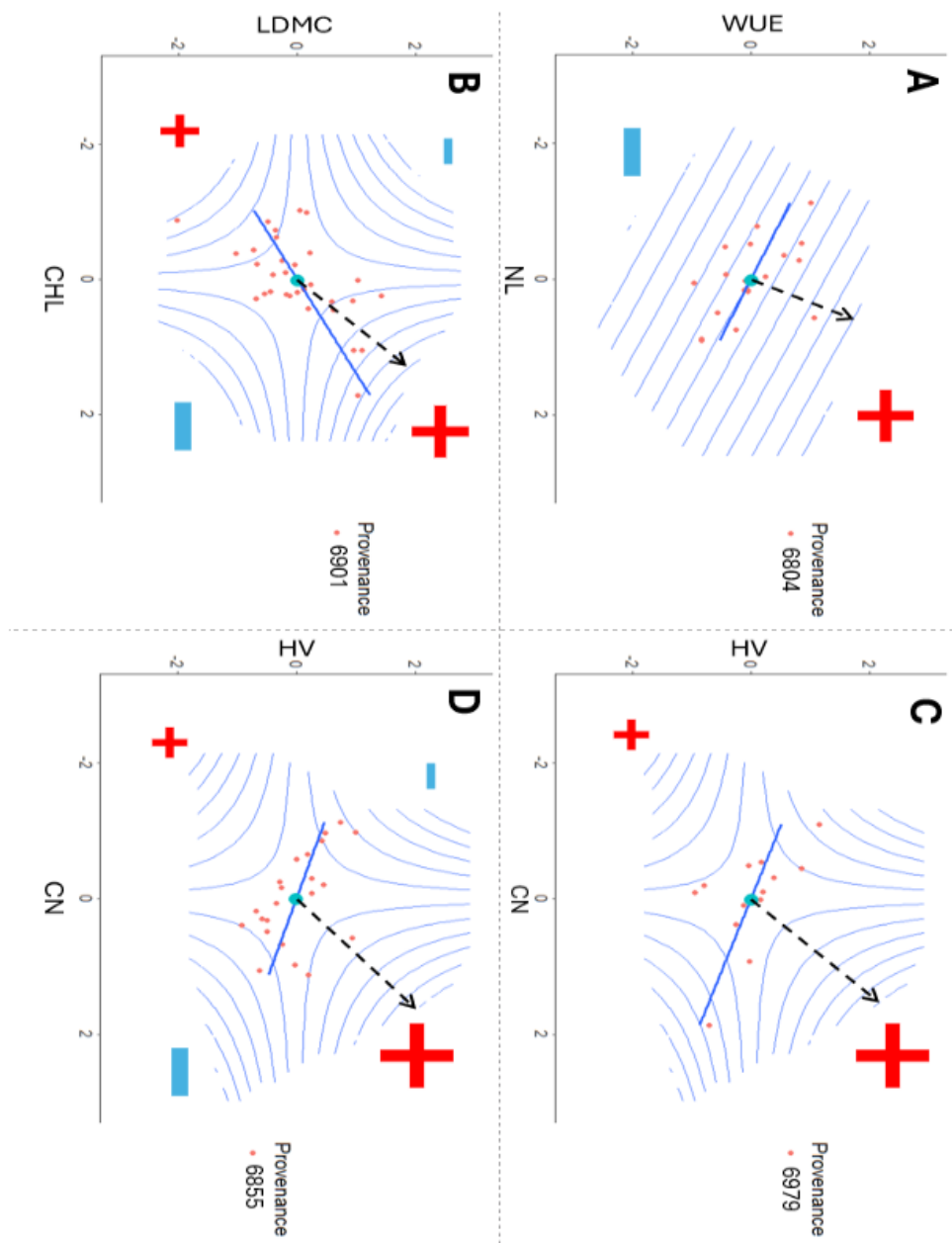
448 circles mean. PCAs of trait values among all sites (A), in PET (B), in LAU (C) and in CHI (D).

449

450 Most traits under selection in PET are not correlated, except for three trait pairs with

451 significant covariances in one or two provenances: WUE and NL in 6804, CN and HV in 6979

452 and 6855. The covariance between WUE and NL in provenance 6804 is largely perpendicular to
453 the direction of selection (Pearson's correlation, $t = -0.194$, $df = 20$, $p = 0.021$, $r = -0.53$, Figure
454 10A). Similarly, the covariances between needle CN and HV for provenances 6855 and 6979 are
455 largely perpendicular to the direction of selection (Pearson's correlation, $t = -2.75$, $df = 20$, $p =$
456 0.012 , $r = -0.52$, Figure 10C & D) (Pearson's correlation, $t = -2.30$, $df = 11$, $p = 0.042$, $r = -0.57$).



458 **Figure 10.** Significant intra-provenance trait covariances in traits under climate-linked selection.

459 **DISCUSSION**

460 The differences in climate among the sites in this study were large enough to adequately
461 detect differences in selection. Further, the directional change in selection along the climate
462 gradient are consistent with the expectation that selection at each site is partly driven by climate
463 (Linhart & Grant, 1996). The lower survival of *P. mariana* individuals in the warmer and drier
464 site (PET), suggests that the climate at his site is stressful for the trees and leads to stronger
465 selection than in cooler climates. Additionally, the traits under selection in each site showed
466 significant differences between selection gradients in the warmest site (PET) and the coldest site
467 (CHI). These changes in selection on traits in these two sites confirm that the climate gradient
468 used in this research is broad enough to register the influence of climate on patterns of selection.

469 *P. mariana* experienced lower survival in the hottest and driest site, indicating that the fitness
470 of these trees is likely lower in a warm and dry environment. The low survival in a warm and dry
471 environment provides insight into the strength of selection on the full phenotype before selection
472 was measured for individual traits. In principle, within a generation, the adaptive plasticity of a
473 population could help to maintain survival and thus fitness in the face of climate change
474 (Bradshaw, 1965; C. D. Schlichting, 1986; Sultan, 1987; Van Tienderen, 1991; C. Schlichting,
475 1998; Kingsolver *et al.*, 2001). Although the provenances did exhibit trait plasticity (Figure S1),
476 the low survival rate in Petawawa indicates that the adaptive plasticity of the *P. mariana*
477 provenances to hot and dry environments is insufficient to maintain survival at levels similar to
478 those in cooler climates. If findings from this spatial climate gradient apply to the temporal
479 effects of climate change on *P. mariana* in general, this will likely result in a lower abundance of
480 *P. mariana* individuals as climates increase in temperature and decrease in water availability.

481 First, the lower survival in the warmer and drier site resulted in weaker selection gradients in
482 Petawawa than at the other sites. Weak selection gradients despite strong selection can occur
483 because of low variation in trait values (resulting in survivor bias) (Mitchell-Olds & Shaw, 1987;
484 Wade & Kalisz, 1990). In other words, mortality of individuals led to an underestimate of the
485 selection gradients. Further, since biomass is often allocated in priority to the most limiting
486 resource a common response to water limitation is a shift in biomass allocation to belowground
487 growth (Poorter *et al.*, 2012). If belowground allocation increases plastically with water
488 limitation, then height-based RGR would become an increasingly poor indicator of performance
489 in increasingly water-limited sites. Thus, the weaker selection gradients in PET could be an
490 artifact of aboveground relative growth rate becoming an increasingly imperfect indicator of
491 performance at the drier site.

492 The magnitude of selection in this study is smaller than the values from the body of
493 literature. Selection gradients for natural populations typically range from negative one to one,
494 with a mean of 0.22 and median of 0.16 (Kingsolver *et al.*, 2001), whereas the largest selection
495 gradient in this study is 0.07 in CHI and the smallest is 0.021 in PET. To contrast this,
496 experimental studies have been shown to detect stronger selection than observational studies
497 since they can impose more extreme environmental conditions than are naturally present (Caruso
498 *et al.*, 2017). As such the pressures faced in the natural environment may be less severe resulting
499 in lower selection pressures. However, selection in the present study was lower than what is
500 typically observed in natural environments. One possible explanation is that all the provenances
501 used in this study originated from the southern latitudes of *P. mariana*'s range, where
502 environmental conditions may not differ drastically from those experienced in previous
503 generations. As a result, the fitness of the present individuals may have been somewhat impacted

504 but not to a large extent, leading to relatively moderate fitness levels for surviving individuals.
505 This aligns with findings that lower fitness tends to result in larger selection gradients (Caruso et
506 al., 2017). Therefore, the low selection gradients in this study indicate that the fitness of
507 surviving individuals is high compared to other selection studies in the literature. Again, this is
508 likely due to the mortality in the experiments, removing low performing individuals before this
509 study took place.

510 Response to selection for the *P. mariana* provenances under study is largely
511 unconstrained by phenotypic covariance. Phenotypic integration is unlikely to hinder or facilitate
512 response to selection of *P. mariana* in warmer climates. Not only is the overall strength of trait
513 integration weak, as shown by the PCAs performed within site, but very few of the provenances
514 showed intra-provenance covariation for the four climate-linked selection gradients. A corollary
515 of the lack of trait integration in all provenances is that their integration does not differ, and none
516 of the studied provenances show higher adaptive potential compared to others. The three
517 instances where trait covariation was present in fact showed integration that would slow down
518 evolutionary response to selection. However, given the rarity of these instances, overall, I do not
519 expect integration to play a meaningful role in black spruce response to selection from warming
520 climates. Over the long term given unknown future environmental change, low integration is
521 beneficial as it provides more flexibility for the phenotype to respond to selection.

522 **REFERENCES**

523 Ahrens, C. W., Andrew, M. E., Mazanec, R. A., Ruthrof, K. X., Challis, A., Hardy, G., Byrne,
524 M., Tissue, D. T., & Rymer, P. D. (2020). Plant functional traits differ in adaptability and
525 are predicted to be differentially affected by climate change. *Ecology and Evolution*,
526 *10*(1), 232–248. <https://doi.org/10.1002/ece3.5890>

527 Allen, C. D., Macalady, A. K., Chenchouni, H., Bachelet, D., McDowell, N., Vennetier, M.,
528 Kitzberger, T., Rigling, A., Breshears, D. D., Hogg, E. H. (Ted), Gonzalez, P., Fensham,
529 R., Zhang, Z., Castro, J., Demidova, N., Lim, J.-H., Allard, G., Running, S. W., Semerci,
530 A., & Cobb, N. (2010). A global overview of drought and heat-induced tree mortality
531 reveals emerging climate change risks for forests. *Forest Ecology and Management*,
532 259(4), 660–684. <https://doi.org/10.1016/j.foreco.2009.09.001>

533 Andrés-Hernández, A. R., Luna-Vega, I., & Rodríguez-Ramírez, E. C. (2023). Functional traits
534 and adaptive capacity of cloud forest *Ternstroemia* species in response to climatic
535 variation. *Flora*, 307, 152383. <https://doi.org/10.1016/j.flora.2023.152383>

536 Aubin, I., Boisvert-Marsh, L., Kebli, H., McKenney, D., Pedlar, J., Lawrence, K., Hogg, E. H.,
537 Boulanger, Y., Gauthier, S., & Ste-Marie, C. (2018). Tree vulnerability to climate
538 change: Improving exposure-based assessments using traits as indicators of sensitivity.
539 *Ecosphere*, 9(2), e02108. <https://doi.org/10.1002/ecs2.2108>

540 Aubin, I., Munson, A. D., Cardou, F., Burton, P. J., Isabel, N., Pedlar, J. H., Paquette, A., Taylor,
541 A. R., Delagrange, S., Kebli, H., Messier, C., Shipley, B., Valladares, F., Kattge, J.,
542 Boisvert-Marsh, L., & McKenney, D. (2016). Traits to stay, traits to move: A review of
543 functional traits to assess sensitivity and adaptive capacity of temperate and boreal trees
544 to climate change. *Environmental Reviews*, 24(2), 164–186. [https://doi.org/10.1139/er-](https://doi.org/10.1139/er-2015-0072)
545 2015-0072

546 Barrett, R. D. H., & Schluter, D. (2008). Adaptation from standing genetic variation. *Trends in*
547 *Ecology & Evolution*, 23(1), 38–44. <https://doi.org/10.1016/j.tree.2007.09.008>

548 Bartlett, M. K., Scoffoni, C., Ardy, R., Zhang, Y., Sun, S., Cao, K., & Sack, L. (2012). Rapid
549 determination of comparative drought tolerance traits: Using an osmometer to predict

550 turgor loss point. *Methods in Ecology and Evolution*, 3(5), 880–888.
551 <https://doi.org/10.1111/j.2041-210X.2012.00230.x>

552 Björklund, M. (1996). The importance of evolutionary constraints in ecological time scales.
553 *Evolutionary Ecology*, 10(4), 423–431. <https://doi.org/10.1007/BF01237727>

554 Blasini, D., Koepke, D., Grady, K., Allan, G., Gehring, C., Whitham, T., Cushman, S., &
555 Hultine, K. (2021). Adaptive trait syndromes along multiple economic spectra define cold
556 and warm adapted ecotypes in a widely distributed foundation tree species. *Journal of*
557 *Ecology*, 109(3), 1298–1318. <https://doi.org/10.1111/1365-2745.13557>

558 Boisvert-Marsh, L., Royer-Tardif, S., Nolet, P., Doyon, F., & Aubin, I. (2020). Using a Trait-
559 Based Approach to Compare Tree Species Sensitivity to Climate Change Stressors in
560 Eastern Canada and Inform Adaptation Practices. *Forests*, 11(9), Article 9.
561 <https://doi.org/10.3390/f11090989>

562 Boucher, D., Gauthier, S., Thiffault, N., Marchand, W., Girardin, M., & Urli, M. (2020). How
563 climate change might affect tree regeneration following fire at northern latitudes: A
564 review. *New Forests*, 51(4), 543–571. <https://doi.org/10.1007/s11056-019-09745-6>

565 Bradshaw, A. D. (1965). Evolutionary Significance of Phenotypic Plasticity in Plants. In E. W.
566 Caspari & J. M. Thoday (Eds.), *Advances in Genetics* (Vol. 13, pp. 115–155). Academic
567 Press. [https://doi.org/10.1016/S0065-2660\(08\)60048-6](https://doi.org/10.1016/S0065-2660(08)60048-6)

568 Brecka, A., Shahi, C., & Chen, H. (2018). Climate change impacts on boreal forest timber
569 supply. *Forest Policy and Economics*, 92, 11–21.
570 <https://doi.org/10.1016/j.forpol.2018.03.010>

571 Burns, R. M. (1990). *Silvics of North America*. U.S. Department of Agriculture, Forest Service.

572 Carter, J. L., & White, D. A. (2009). Plasticity in the Huber value contributes to homeostasis in
573 leaf water relations of a mallee Eucalypt with variation to groundwater depth. *Tree*
574 *Physiology*, 29(11), 1407–1418. <https://doi.org/10.1093/treephys/tpp076>

575 Caruso, C. M., Martin, R. A., Sletvold, N., Morrissey, M. B., Wade, M. J., Augustine, K. E.,
576 Carlson, S. M., MacColl, A. D. C., Siepielski, A. M., & Kingsolver, J. G. (2017). What
577 Are the Environmental Determinants of Phenotypic Selection? A Meta-analysis of
578 Experimental Studies. *The American Naturalist*, 190(3), 363–376.
579 <https://doi.org/10.1086/692760>

580 Cayan, D. R., Kammerdiener, S. A., Dettinger, M. D., Caprio, J. M., & Peterson, D. H. (2001).
581 Changes in the Onset of Spring in the Western United States. *Bulletin of the American*
582 *Meteorological Society*, 82(3), 399–415.

583 Chagnon, C., Wotherspoon, A. R., & Achim, A. (2022). Deciphering the black spruce response
584 to climate variation across eastern Canada using a meta-analysis approach. *Forest*
585 *Ecology and Management*, 520, 120375. <https://doi.org/10.1016/j.foreco.2022.120375>

586 Challis, A., Blackman, C., Ahrens, C., Medlyn, B., Rymer, P., & Tissue, D. (2022). Adaptive
587 plasticity in plant traits increases time to hydraulic failure under drought in a foundation
588 tree. *Tree Physiology*, 42(4), 708–721. <https://doi.org/10.1093/treephys/tpab096>

589 Choat, B., Brodribb, T. J., Brodersen, C. R., Duursma, R. A., López, R., & Medlyn, B. E. (2018).
590 Triggers of tree mortality under drought. *Nature*, 558(7711), 531–539.
591 <https://doi.org/10.1038/s41586-018-0240-x>

592 Choat, B., Jansen, S., Brodribb, T. J., Cochard, H., Delzon, S., Bhaskar, R., Bucci, S. J., Feild, T.
593 S., Gleason, S. M., Hacke, U. G., Jacobsen, A. L., Lens, F., Maherali, H., Martínez-
594 Vilalta, J., Mayr, S., Mencuccini, M., Mitchell, P. J., Nardini, A., Pittermann, J., ...

595 Zanne, A. E. (2012). Global convergence in the vulnerability of forests to drought.
596 *Nature*, 491(7426), Article 7426. <https://doi.org/10.1038/nature11688>

597 de Lafontaine, G., Napier, J. D., Petit, R. J., & Hu, F. S. (2018). Invoking adaptation to decipher
598 the genetic legacy of past climate change. *Ecology*, 99(7), 1530–1546.

599 Farquhar, G., O’Leary, M. H. O., & Berry, J. (1982). On the relationship between carbon isotope
600 discrimination and the intercellular carbon dioxide concentration in leaves. *Australian*
601 *Journal of Plant Physiology*, 9, 121–137.

602 Gamache, I., & Payette, S. (2004). Height growth response of tree line black spruce to recent
603 climate warming across the forest-tundra of eastern Canada. *Journal of Ecology*, 92(5),
604 835–845. <https://doi.org/10.1111/j.0022-0477.2004.00913.x>

605 Garnier, E., Shipley, B., Roumet, C., & Laurent, G. (2001). A standardized protocol for the
606 determination of specific leaf area and leaf dry matter content. *Functional Ecology*,
607 15(5), 688–695. <https://doi.org/10.1046/j.0269-8463.2001.00563.x>

608 Girardin, M. P., Hogg, E. H., Bernier, P. Y., Kurz, W. A., Guo, X. J., & Cyr, G. (2016). Negative
609 impacts of high temperatures on growth of black spruce forests intensify with the
610 anticipated climate warming. *Global Change Biology*, 22(2), 627–643.
611 <https://doi.org/10.1111/gcb.13072>

612 Grime, J. (2001). Plant Strategies, Vegetation Processes, and Ecosystem Properties. In *Biological*
613 *Conservation—Biol Conserv* (Vol. 107). [https://doi.org/10.1016/S0006-3207\(02\)00055-1](https://doi.org/10.1016/S0006-3207(02)00055-1)

614 Gutschick, V. P., & Wiegand, F. W. (1988). Optimizing the Canopy Photosynthetic Rate by
615 Patterns of Investment in Specific Leaf Mass. *The American Naturalist*, 132(1), 67–86.
616 <https://doi.org/10.1086/284838>

617 Hogg, E. (Ted), Brandt, J. P., & Kochtubajda, B. (2005). Factors affecting interannual variation
618 in growth of western Canadian aspen forests during 1951-2000. *Canadian Journal of*
619 *Forest Research*, 35(3), 610–622. <https://doi.org/10.1139/x04-211>

620 Hopkins, W.G. and Hüner, N.P. (2004) *Introduction to Plant Physiology. 3rd Edition, John*
621 *Wiley & Sons, Inc, Hoboken. - References—Scientific Research Publishing.* (n.d.).
622 Retrieved September 17, 2024, from
623 <https://www.scirp.org/reference/referencespapers?referenceid=1590176>

624 Iverson, L. R., Schwartz, M. W., & Prasad, A. M. (2004). How Fast and Far Might Tree Species
625 Migrate in the Eastern United States Due to Climate Change? *Global Ecology and*
626 *Biogeography*, 13(3), 209–219.

627 Kaulesar Sukul, D. M., den Hoed, P. T., Johannes, E. J., van Dolder, R., & Benda, E. (1993).
628 Direct and indirect methods for the quantification of leg volume: Comparison between
629 water displacement volumetry, the disk model method and the frustum sign model
630 method, using the correlation coefficient and the limits of agreement. *Journal of*
631 *Biomedical Engineering*, 15(6), 477–480. [https://doi.org/10.1016/0141-5425\(93\)90062-4](https://doi.org/10.1016/0141-5425(93)90062-4)

632 Kimball, J. S., McDonald, K. C., Running, S. W., & Frohling, S. E. (2004). Satellite radar remote
633 sensing of seasonal growing seasons for boreal and subalpine evergreen forests. *Remote*
634 *Sensing of Environment*, 90(2), 243–258. <https://doi.org/10.1016/j.rse.2004.01.002>

635 Kingsolver, J. G., Hoekstra, H. E., Hoekstra, J. M., Berrigan, D., Vignieri, S. N., Hill, C. E.,
636 Hoang, A., Gibert, P., Beerli, P., & Travis, E. J. (2001). The Strength of Phenotypic
637 Selection in Natural Populations. *The American Naturalist*, 157(3), 245–261.
638 <https://doi.org/10.1086/319193>

639 Lambers, H., Raven, J. A., Shaver, G. R., & Smith, S. E. (2008). Plant nutrient-acquisition
640 strategies change with soil age. *Trends in Ecology & Evolution*, 23(2), 95–103.
641 <https://doi.org/10.1016/j.tree.2007.10.008>

642 Lindner, M., Maroschek, M., Netherer, S., Kremer, A., Barbati, A., Garcia-Gonzalo, J., Seidl, R.,
643 Delzon, S., Corona, P., Kolström, M., Lexer, M. J., & Marchetti, M. (2010). Climate
644 change impacts, adaptive capacity, and vulnerability of European forest ecosystems.
645 *Forest Ecology and Management*, 259(4), 698–709.
646 <https://doi.org/10.1016/j.foreco.2009.09.023>

647 Linhart, Y. B., & Grant, M. C. (1996). Evolutionary significance of local genetic differentiation
648 in plants. *Annual Review of Ecology, Evolution, and Systematics*, 27(Volume 27, 1996),
649 237–277. <https://doi.org/10.1146/annurev.ecolsys.27.1.237>

650 Malcolm, J. R., Markham, A., Neilson, R. P., & Garaci, M. (2002). Estimated migration rates
651 under scenarios of global climate change. *Journal of Biogeography*, 29(7), 835–849.
652 <https://doi.org/10.1046/j.1365-2699.2002.00702.x>

653 McDowell, N. G., Allen, C. D., Anderson-Teixeira, K., Aukema, B. H., Bond-Lamberty, B.,
654 Chini, L., Clark, J. S., Dietze, M., Grossiord, C., Hanbury-Brown, A., Hurtt, G. C.,
655 Jackson, R. B., Johnson, D. J., Kueppers, L., Lichstein, J. W., Ogle, K., Poulter, B., Pugh,
656 T. A. M., Seidl, R., ... Xu, C. (2020). Pervasive shifts in forest dynamics in a changing
657 world. *Science*, 368(6494), eaaz9463. <https://doi.org/10.1126/science.aaz9463>

658 Michaletz, S. T., Weiser, M. D., Zhou, J., Kaspari, M., Helliker, B. R., & Enquist, B. J. (2015).
659 Plant Thermoregulation: Energetics, Trait–Environment Interactions, and Carbon
660 Economics. *Trends in Ecology & Evolution*, 30(12), 714–724.
661 <https://doi.org/10.1016/j.tree.2015.09.006>

662 Mitchell-Olds, T., & Shaw, R. G. (1987). Regression Analysis of Natural Selection: Statistical
663 Inference and Biological Interpretation. *Evolution*, 41(6), 1149–1161.
664 <https://doi.org/10.1111/j.1558-5646.1987.tb02457.x>

665 Morgenstern, E. K., & Kokocinski, G. H. (1976). *Range-wide, cooperative black spruce*
666 *provenance study, Morgenstern and Kokocinski, 1976* (Special Joint Report 2).
667 Environment Canada.

668 Münchinger, I. K., Hajek, P., Akdogan, B., Caicoya, A. T., & Kunert, N. (2023). Leaf thermal
669 tolerance and sensitivity of temperate tree species are correlated with leaf physiological
670 and functional drought resistance traits. *Journal of Forestry Research*, 34(1), 63–76.
671 <https://doi.org/10.1007/s11676-022-01594-y>

672 Peng, C., Ma, Z., Lei, X., Zhu, Q., Chen, H., Wang, W., Liu, S., Li, W., Fang, X., & Zhou, X.
673 (2011). A drought-induced pervasive increase in tree mortality across Canada’s boreal
674 forests. *Nature Climate Change*, 1(9), 467–471. <https://doi.org/10.1038/nclimate1293>

675 Perez-Harguindeguy, N., Díaz, S., Garnier, E., Lavorel, S., Poorter, H., Jaureguiberry, P., Bret-
676 Harte, M. S., Cornwell, W. K., Craine, J. M., Gurvich, D. E., Urcelay, C., Veneklaas, E.
677 J., Reich, P. B., Poorter, L., Wright, I. J., Ray, P., Enrico, L., Pausas, J. G., de Vos, A. C.,
678 ... Cornelissen, J. H. C. (2013). New handbook for standardised measurement of plant
679 functional traits worldwide. <https://doi.org/10.1071/BT12225>

680 Poorter, H., Niklas, K. J., Reich, P. B., Oleksyn, J., Poot, P., & Mommer, L. (2012). Biomass
681 allocation to leaves, stems and roots: Meta-analyses of interspecific variation and
682 environmental control. *New Phytologist*, 193(1), 30–50. [https://doi.org/10.1111/j.1469-](https://doi.org/10.1111/j.1469-8137.2011.03952.x)
683 [8137.2011.03952.x](https://doi.org/10.1111/j.1469-8137.2011.03952.x)

684 Poorter, L., McDonald, I., Alarcón, A., Fichtler, E., Licona, J.-C., Peña-Claros, M., Sterck, F.,
685 Villegas, Z., & Sass-Klaassen, U. (2010). The importance of wood traits and hydraulic
686 conductance for the performance and life history strategies of 42 rainforest tree species.
687 *New Phytologist*, 185(2), 481–492. <https://doi.org/10.1111/j.1469-8137.2009.03092.x>

688 Robert, E., Lenz, P., Bergeron, Y., de Lafontaine, G., Bouriaud, O., Isabel, N., & Girardin, M.
689 (2024). Future carbon sequestration potential in a widespread transcontinental boreal tree
690 species: Standing genetic variation matters! *Global Change Biology*, 30(6).
691 <https://doi.org/10.1111/gcb.17347>

692 Royer-Tardif, S., Boisvert-Marsh, L., Godbout, J., Isabel, N., & Aubin, I. (2021). Finding
693 common ground: Toward comparable indicators of adaptive capacity of tree species to a
694 changing climate. *Ecology and Evolution*, 11(19), 13081–13100.
695 <https://doi.org/10.1002/ece3.8024>

696 Schlichting, C. D. (1986). The evolution of phenotypic plasticity in plants. *Annual Review of*
697 *Ecology, Evolution, and Systematics*, 17(Volume 17.), 667–693.
698 <https://doi.org/10.1146/annurev.es.17.110186.003315>

699 Schlichting, C. (with Pigliucci, M.). (1998). Phenotypic evolution: A reaction norm perspective.
700 Sinauer.

701 Schluter, D. (1996). Adaptive Radiation Along Genetic Lines of Least Resistance. *Evolution*,
702 50(5), 1766–1774. <https://doi.org/10.2307/2410734>

703 Shaw, J. D., Steed, B. E., & DeBlander, L. T. (2005). Forest Inventory and Analysis (FIA)
704 Annual Inventory Answers the Question: What Is Happening to Pinyon-Juniper
705 Woodlands? *Journal of Forestry*, 103(6), 280–285. <https://doi.org/10.1093/jof/103.6.280>

706 Sniderhan, A. E., Mamet, S. D., & Baltzer, J. L. (2021). Non-uniform growth dynamics of a
707 dominant boreal tree species (*Picea mariana*) in the face of rapid climate change.
708 *Canadian Journal of Forest Research*, 51(4), 565–572. [https://doi.org/10.1139/cjfr-2020-](https://doi.org/10.1139/cjfr-2020-0188)
709 0188

710 Sultan, S. E. (1987). Evolutionary Implications of Phenotypic Plasticity in Plants. In M. K.
711 Hecht, B. Wallace, & G. T. Prance (Eds.), *Evolutionary Biology: Volume 21* (pp. 127–
712 178). Springer US. https://doi.org/10.1007/978-1-4615-6986-2_7

713 Van Tienderen, P. H. (1991). Evolution of Generalists and Specialist in Spatially Heterogeneous
714 Environments. *Evolution*, 45(6), 1317–1331. <https://doi.org/10.2307/2409882>

715 Via, S., & Lande, R. (1985). Genotype-environment interaction and the evolution of phenotypic
716 plasticity. *Evolution*, 39(3), 505–522. [https://doi.org/10.1111/j.1558-](https://doi.org/10.1111/j.1558-5646.1985.tb00391.x)
717 5646.1985.tb00391.x

718 Wade, M. J., & Kalisz, S. (1990). The Causes of Natural Selection. *Evolution*, 44(8), 1947–1955.
719 <https://doi.org/10.1111/j.1558-5646.1990.tb04301.x>

720 Westoby, M., Falster, D. S., Moles, A. T., Vesk, P. A., & Wright, I. J. (2002). Plant Ecological
721 Strategies: Some Leading Dimensions of Variation Between Species. *Annual Review of*
722 *Ecology, Evolution, and Systematics*, 33(Volume 33, 2002), 125–159.
723 <https://doi.org/10.1146/annurev.ecolsys.33.010802.150452>