

1 **Cryptic ontogenetic changes in the ventral coloration of a colour**
2 **polymorphic wall lizard (*Podarcis muralis*)**

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33 **Abstract (284 words)**

34 Many animals undergo irreversible ontogenetic colour changes (OCCs), yet these changes are often overlooked
35 despite their potential ethological relevance. The problem is compounded when OCCs involve wavelengths
36 invisible to humans. Wall lizards can perceive ultraviolet (UV) light, and their conspicuous ventral and
37 ventrolateral coloration—including UV-reflecting patches—likely serves social communication. Here, we
38 describe OCCs in the ventral (throat and belly) and ventrolateral (outer ventral scales, OVS) coloration of
39 juvenile common wall lizards (*Podarcis muralis*) as perceived by conspecifics. We measured reflectance in
40 newborn and yearling lizards raised under semi-natural conditions and used visual modelling to estimate
41 chromatic distances within individuals and across life stages (i.e. newborns, yearlings, and adults). Newborns
42 typically exhibit UV-enhanced white (UV⁺white) on their ventral surfaces (throat, belly, and OVS), a colour
43 that is likely discriminable to conspecifics from the most frequent adult colours in the throat (i.e. orange, yellow,
44 and UV-reduced white; UV⁻white) and OVS (i.e. UV-blue). The prevalence of UV⁺white decreases with age,
45 with the decline being less pronounced in female bellies. OCCs to UV-blue in the OVS are more apparent in
46 males than in females and appear delayed relative to changes in the throat and belly. While throat colours in
47 yearlings are indistinguishable to conspecifics from adult throat colours, yearling UV-blue patches remain
48 chromatically distinct from those of adults. This delay may reflect variations in the mechanisms of colour
49 production or distinct selective pressures acting on these patches. Overall, our results show that OCCs in *P.*
50 *muralis* fulfil a key requirement for social signals by being perceptible to conspecifics. This supports the
51 hypothesis that OCCs may play a role mediating interactions between juveniles and adults, as well as delaying
52 the onset of colours involved in social communication.

53

54 **Keywords:** Ontogenetic colour changes; Colour polymorphism; Ultraviolet light perception; Sensory ecology;
55 Visual modelling; Social communication; Wall lizards; *Podarcis muralis*

56

57 Introduction

58 Development exposes juvenile animals to varying selective forces arising from changes in size, vulnerability,
59 diet, habitat, or social environment, which are sometimes associated with changes in body coloration.
60 Ontogenetic colour change (OCCs) is defined as an irreversible colour change that is part of the normal
61 development of individuals in a species (Buckman 1985; Booth 1990). External conditions may affect the timing
62 of the change and the resulting colour expression (Frédérich et al. 2010; Stückler et al. 2022). OCCs can be so
63 dramatic that the juvenile and corresponding adult forms were sometimes mistakenly labelled as different
64 species (Smale and Kok 1983). Despite their potential ethological and selective relevance, research on OCCs
65 has often lagged behind the study of adult coloration due to the logistical difficulties it poses, to the point that
66 we sometimes ignore whether certain species undergo OCCs at all. The risk of inadvertently disregarding OCCs
67 increases when colour changes take place out of the spectrum of light visible to humans (Rivas and Burghardt
68 2002; Caves et al. 2019). Despite its widespread occurrence and notable biological importance, the ultraviolet
69 (UV) component in animal coloration has long gone unnoticed by scientists. However, its potential as a private
70 communication channel hidden from animals lacking UV vision makes it particularly worthy of attention
71 (Bradbury and Vehrencamp 2011; Cronin and Bok 2016).

72 Studies on lizard coloration have been crucial for advancing our understanding of a great range of
73 evolutionary processes, from sexual selection and animal communication to the formation of new species
74 (Olsson et al. 2013). Species showing heritable colour polymorphism (i.e. the coexistence of two or more
75 alternative colour morphs of a species, with the rarer being too frequent to be solely the result of recurrent
76 mutation (White and Kemp 2016) have been particularly popular in evolutionary biology and ethology (Majerus
77 2008; Svensson 2017; Endler et al. 2023). Ventral colour polymorphism has evolved in at least seven families
78 of lizards, providing a unique opportunity to study the evolutionary processes responsible of maintaining
79 intraspecific diversity (Stuart-Fox et al. 2020). Unfortunately, OCCs has rarely been examined in colour
80 polymorphic lizards, and colour variation has often been described from the human perspective (which in some
81 cases may have led to biologically irrelevant morph categorization; but see Rankin et al. 2016; Pérez i de Lanuza
82 et al. 2018). Indeed, the historical neglect of UV colours in lizards has been corrected only in recent decades,
83 revealing a great array of UV-coloured patches which may play a role as chromatic signals (e.g. involved in
84 deimatic displays; Abramjan et al. 2015; Badiane et al. 2018); or male-male competition; Stapley and Whiting
85 2006; Whiting et al. 2006), but the relevance of UV in OCCs has not yet been examined (e.g. Yang et al. 2023).

86 Wall lizards (genus *Podarcis*, Lacertidae), comprising ca. 26 species with circum-Mediterranean
87 distribution (Speybroeck et al. 2016; O'Shea 2021; Yang et al. 2021), have recently attracted attention as a
88 suitable group in which to study colour (Pérez i de Lanuza et al. 2013, 2014; Andrade et al. 2019; Names et al.
89 2019; Miñano et al. 2021; Sacchi et al. 2021; de la Cruz et al. 2023; Abalos et al. 2024; Escoriza 2024). The
90 dorsal coloration of *Podarcis* lizards is typically cryptic, consisting of dark melanin-based patches forming a
91 reticulate pattern or bands over an olive background coloration (Salvador 2014; Ortega et al. 2019). In contrast,
92 most species exhibit conspicuous ventral and ventrolateral coloration, often combining polymorphic throat and
93 belly coloration, melanin-based patches, and conspicuous UV-reflecting patches in the shoulder region and/or
94 the flanks (Arnold et al. 2007; Huyghe et al. 2007; Runemark et al. 2010; Marshall and Stevens 2014; Badiane
95 and Font 2021; Brock et al. 2022b; de la Cruz et al. 2023; Solan et al. 2023). One such species (which is also the

96 most widely distributed) is the European common wall lizard, *Podarcis muralis*. In common with many diurnal
97 lizards, *P. muralis* has a sophisticated colour vision system with four different types of cones that are sensitive
98 to light in the wavelengths between 320 and 700 nm (Pérez i de Lanuza and Font 2014a; Martin et al. 2015).
99 Newborn *P. muralis* show a creamy white ventral coloration, but adults of both sexes may show up to 5
100 alternative ventral colour morphs to the human observer: three uniform (pure) morphs (orange, white, and
101 yellow), and 2 mixed-morph mosaics combining orange and white or yellow and orange (Sacchi et al. 2007;
102 Calsbeek et al. 2010; Pérez i de Lanuza et al. 2019). However, spectrophotometry and UV photography have
103 revealed the existence of two types of whites differing in their UV reflectance (UV⁺white and UV⁻white) in the
104 ventral coloration of both male and female *Podarcis* lizards, which may increase the number of discrete colours
105 conforming lacertid colour polymorphism (Abalos et al. 2016; Pérez i de Lanuza & Font, 2024, *in press*). In
106 addition, adults of both sexes show UV-blue patches in their outer ventral scales (OVS), but these are typically
107 larger, more abundant, and more UV-biased and conspicuous in males (Pérez i de Lanuza and Font 2015).

108 Because of their ventral and ventrolateral location (allowing lizards to control their visibility through
109 posturing) and conspicuousness, ventral polymorphic colours and UV-blue patches have been often thought
110 to play a role in intraspecific communication (Pérez i de Lanuza et al. 2013, 2017; Abalos et al. 2016a, 2020;
111 Names et al. 2019; Scali et al. 2019). The spectral properties of UV-blue patches in males are tuned to lacertid
112 vision (Pérez i de Lanuza and Font 2014a; Martin et al. 2015), correlate with bite force and body condition
113 (Pérez i de Lanuza et al. 2014), and behavioural evidence suggest that males making the OVS colour pattern
114 visible during male-male confrontations have higher probability of prevailing over their rivals (Abalos et al.
115 2024). Research on *Podarcis* ventral colour polymorphism has largely focused on testing for the existence of
116 alternative phenotypic optima, often in the form of alternative reproductive strategies involving differential
117 sociosexual behaviour or breeding investment (Huyghe et al. 2007; Sacchi et al. 2009; Calsbeek et al. 2010;
118 Galeotti et al. 2013; Mangiacotti et al. 2019; Brock et al. 2022a). Although the evidence in this regard is, at best,
119 inconclusive (Sacchi et al. 2009; Stuart-Fox et al. 2020; Abalos et al. 2020, 2021), there are other lines of evidence
120 suggesting a social role for alternative ventral colours. The average orange, yellow, and UV-white colours are
121 discriminable to conspecifics (Pérez i de Lanuza et al. 2018), a crucial prerequisite for alternative colours to
122 function as social signals. In *P. muralis*, ventral colour may be playing a role in the formation of lasting male-
123 female associations, with homomorphic pairs being more common than heteromorphic pairs in natural
124 populations (Pérez i de Lanuza et al. 2013a, 2016b; but see Abalos et al. 2020; Aguilar et al. 2022a). The positive
125 correlation observed between increased morph diversity and male-biased sex ratios across natural populations
126 in the eastern Pyrenees further indicates that variation in ventral coloration among sympatric lizards may play
127 a role in sexual selection under conditions of heightened male-male competition (Pérez i de Lanuza et al. 2017;
128 Aguilar et al. 2024). Frequency-dependent effects of ventral colour on social interactions—such as a rare morph
129 advantage or providing additional cues for social recognition—could potentially explain this finding but remain
130 unexplored (Sheehan and Tibbetts 2009; Wellenreuther et al. 2014; Sheehan and Bergman 2016; Sheehan et al.
131 2017).

132 Despite the considerable attention given to the potential signalling role of these colour patches,
133 research into their development (ontogeny) remains limited. For instance, newborn ventral coloration (which
134 is perceived as identical to the adult white morph by human observers) has never been objectively characterized,
135 and white morph adult lizards have been assumed to retain the juvenile coloration during growth (Pérez i de

136 [Lanuza et al. 2013](#)). Here, we set out to describe ontogenetic changes in the ventral coloration of juvenile *P.*
137 *muralis* lizards both objectively (i.e. quantifying changes in reflectance) and as perceived by conspecifics (i.e.
138 using visual models to simulate lacertid colour vision). To do so, we raised juveniles coming from two different
139 breeding experiments under semi-natural conditions ([Abalos et al. 2020, 2021](#)), taking spectrophotometric
140 measurements 3-4 weeks after hatching and 9-10 months later. We then used visual modelling to estimate
141 chromatic distances between paired spectra from the same individual. Lastly, we assessed chromatic differences
142 in ventral coloration between newborns, yearlings, and adult lizards by including ventral reflectance data from
143 adult individuals sampled in the same localities where the parents of the juvenile lizards were captured.

144

145 **Materials and methods**

146 In 2018 and 2019 we conducted mesocosm experiments with *P. muralis* at the Station d'Écologie Théorique
147 et Expérimentale (Moulis, France). Lizards participating in these experiments (135 males and 225 females) were
148 captured in 14 different localities across the Cerdanya plateau, in eastern Pyrenees ([Abalos et al. 2020, 2021,](#)
149 [2024](#)). Lizards mated under semi-natural conditions, after which females were housed individually until
150 oviposition and their clutches were incubated (**Fig. S1**). In September, 3-4 weeks after hatching, we released
151 417 juveniles resulting from these experiments (2018: $N = 45$, 2019: $N = 372$) into 44 plastic tubs (170 cm
152 diameter, 60 cm high; **Fig. S1**), that were kept outdoors, under natural temperature and illumination conditions,
153 in groups of 10-12 individuals ([Abalos et al. 2021](#)). Prior to release into the tanks, we permanently marked each
154 newborn on the ventral scales using a disposable medical cautery unit ([Ekner et al. 2011](#)). We took
155 spectrophotometric measurements of the juveniles at two ontogenetic stages: as newborns (3-4 weeks after
156 hatching) and as yearlings (10.5 months after hatching). Since newborn lizards show uniform coloration across
157 their ventral and ventrolateral surface, we measured all newborn lizards in the throat ($N = 417$) and only a
158 subset also in the belly ($N = 43$). Dorsal reflectance was measured in eight newborn lizards. Approximately ten
159 months after hatching, we re-captured surviving yearlings in the tanks (June 2019: $N = 16$; July 2020: $N = 50$)
160 and obtained spectrophotometric measurements of the throat, belly, and dorsum ($N = 66$), as well as the UV-
161 blue patches, if present ($N = 36$). We also measured mass (± 0.01 g) and SVL (± 1 mm) in all newborn and
162 yearling lizards using a ruler and a digital pocket scale.

163 Reflectance was measured using a USB 2000 portable diode-array spectrometer equipped with a
164 QP200-2-UV/VIS-BX reading-illumination probe and a PX-2 Xenon strobe light for full spectrum illumination
165 (Ocean Optics Inc., Dunedin, FL, USA; see details in [Font et al. 2009](#)). Small colour patches (such as UV-blue
166 patches in juvenile lizards) may result in chimeric spectra when the cone of light projected by the
167 spectrophotometer probe exceeds the diameter of the measured patch ([Badiane et al. 2017](#)). To avoid this
168 problem, we attached an entomological pin with the nylon head downward to the side of the probe—ensuring
169 a consistent 3 mm distance from the target surface—and limited measurements to colour patches with a
170 diameter of at least 2 mm ([Badiane et al. 2017](#)). For analyses, we restricted the reflectance spectra to the 300-
171 700 nm range to encompass the visual sensitivity of lacertids ([Pérez i de Lanuza and Font 2014b; Martín et al.](#)
172 [2015](#)). To control for noisy variation in luminance, spectra were normalized by subtracting the minimum value
173 at all wavelengths. Spectral data were analysed in R v.4.0.3 (R Development Core Team 2017) using the package
174 *pavo 2* ([Maia et al. 2019](#)).

175 We assigned specific colours (i.e. UV⁺white, UV-white, yellow, orange) to each juvenile colour patch
176 based on the chromatic differences observed among adult throat and belly colours in the reflectance spectra
177 used in Pérez i de Lanuza and Font (2015) (Fig. 1). These spectra correspond to adult males and females
178 collected across various locations in the Cerdanya plateau, the same area where the parent lizards from the
179 present experiment were captured. Notably, in these populations the orange and yellow colours seem to cover
180 the entire ventral surface in males but are restricted to the throat in females (Pérez i de Lanuza et al. 2013a,
181 2017; Abalos et al. 2016, 2020). To determine objective thresholds for morph categorization, for each adult and
182 juvenile spectrum we extracted five standard variables (Table 1): luminance, UV chroma (C_{UV}), and two
183 variables describing hue ($\lambda \beta_{max}$, and λR_{mid}) (Endler 1990; Kemp et al. 2015; Maia et al. 2019). We calculated
184 luminance (Q_t) as the sum of the reflectance across the visible range of lizards (i.e., $R_{300-700}$) and UV chroma
185 (C_{UV}) as the relative reflectance in the UV waveband (i.e. summing the reflectance in the 300-400 range and
186 dividing it by total reflectance; $R_{300-400}/R_{300-700}$). In adults, the wavelength corresponding to the maximum
187 positive slope between 325 and 560 nm ($\lambda \beta_{max_{325-560}}$) shows a gap between 440 and 500 nm that separates
188 most lizards assigned to the orange and yellow colours from lizards assigned to the white morph (Fig. 1).
189 Hence, juvenile spectra showing values of $\beta_{max_{325-560}}$ equal or below 500 nm were classified as white, while
190 lizards showing higher values were classified as either yellow or orange. We then distinguished between yellow
191 and orange juveniles based on the wavelength at which reflectance is halfway between its maximum and its
192 minimum within the 400-650 nm waveband ($\lambda R_{mid_{400-650}}$) (Fig. 1). Specifically, juveniles were classified as
193 yellow if showing values of $\lambda R_{mid_{400-650}}$ equal or lower than 550 nm, and orange if showing higher values.
194 Following Pérez i de Lanuza & Font (2024, *in press*), we classified juvenile spectra as UV⁺white if showing values
195 of $\lambda R_{mid_{300-450}}$ equal or lower than 365 nm, and UV-white if showing higher values (Fig. 1). To validate these
196 thresholds for morph categorization, we reclassified a subset of 460 throat spectra from adult males, previously
197 assigned to the white, orange, or yellow morphs by visual inspection. The high agreement between
198 classifications confirms the reliability of the criteria (Cohen's $kappa \pm CI_{95} = 0.914 \pm 0.031$).

199 We built visual models using the Vorobyev and Osorio receptor noise model (Vorobyev and Osorio
200 1998) implemented in *pavo 2* to assess colour discrimination in terms of chromatic (dS) and achromatic (dL)
201 distances, to test whether ontogenetic changes were large enough to be perceived by conspecifics (Maia et al.
202 2019). Specifically, we quantified within-individual chromatic and achromatic distances in throat and UV-blue
203 coloration based on conspecific cone sensitivities and relative frequencies (Martin et al. 2015; Pérez i de Lanuza
204 et al. 2018). We also estimated the colour distances between the different throat colours shown by yearling
205 lizards. To quantify OCCs based on lacertid colour vision, we estimated the colour distances between the
206 average ventral colour in newborn lizards ($N = 417$) and each of the colours found in full-blown adults (OVS
207 and throat spectra from Pérez i de Lanuza and Font, 2015, the latter re-classified as either UV⁺white, UV-white,
208 yellow, or orange according to the criteria specified above). We explored the similarity between yearling and
209 adult throat colours when viewed by conspecifics by estimating colour distances among all possible
210 combinations. Lastly, we examined OCCs in the OVS of yearling lizards by estimating within-sex colour
211 distances between UV-reflecting patches in yearling lizards and full-blown UV patches from adult lizards.

212 We used the cone sensitivities (UVS:SWS:MSW:LSW, 367:456:497:562 nm) and cone ratios (1:1:1:4)
213 published for *P. muralis* in Martin et al., (2015). In the absence of behavioural estimates of the Weber fraction
214 for colour discrimination in lizards, we set its value to 0.05 (Siddiqi et al. 2004; Pérez i de Lanuza and Font

215 2015; Pérez i de Lanuza et al. 2018), and a standard daylight “D65” irradiance spectrum, as implemented in
216 *pavo*. Colour distances between pairs of colors were measured in units of just noticeable differences (JND),
217 where one JND is assumed to be the threshold of discrimination between two colors under good illumination
218 conditions (Vorobyev et al. 1998). However, as JND values between one and three could mean that two colors
219 are barely discriminated, we also evaluated our results using a more conservative discrimination threshold of 3
220 JNDs (Siddiqi et al. 2004; Santiago et al. 2020). We then used distance-based MANOVAs or PERMANOVAs
221 to test for discriminability between ventral colorations with respect to ontogeny (Anderson 2014). To do so,
222 we first ran a MANOVA procedure on the chromatic and achromatic contrasts using the *adonis2* function from
223 the *pairwiseAdonis* R package (Martinez Arbizu 2017). For comparisons between more than two groups, we ran
224 a PERMANOVA using the *pairwise.adonis2* function in *pairwiseAdonis*, a modified version of the *adonis* function
225 from the *vegan* R package (Oksanen et al. 2016). When comparing reflectance spectra from the same individual
226 taken at different points in time, we accommodated the repeated measures nature of the data in the MANOVA
227 by blocking contrasts at the within-individual level. Finally, as significance thresholds do not necessarily match
228 the theoretical perceptual threshold of one JND above which colours can be said to be distinguishable, we used
229 a bootstrap procedure to generate confidence intervals for the mean colour distance between the different
230 colours. We used the *bootcoldist* function from the *pavo* package on the visual model described above with 999
231 replicates and a 0.95 level for confidence intervals.
232

233 Results

234 Juvenile lizards (yearlings) gained an average of 2.16 ± 0.19 g (mean \pm CI₉₅) in body mass and grew by an
235 average of 22.33 ± 1.21 mm of SVL in the period examined (ca. 10 months). Males grew slightly faster than
236 females (m-f: 2.33 ± 2.34 mm, $t = -1.96$, $p = 0.054$) and increased their mass significantly more (m-f: $0.55 \pm$
237 0.36 g, $t = -3.07$, $p = 0.003$). We found strong evidence that *P. muralis* ventral coloration undergoes ontogenetic
238 changes (Table 1, Fig. 2) that are large enough to be sensed by conspecifics (Tables 2, 3 and S2). Most
239 newborn lizards showed white throats (97%), with three quarters (74%) of them being classified as UV+white
240 according to their spectral properties. Yellow (39%), orange (17%) and UV white (30%) throats predominated
241 in yearling lizards, with only a subset (8%) of re-sampled individuals showing UV+white throats (Fig. 3).
242 UV+white coloration is particularly frequent in the bellies of adult and yearling females, even when their throats
243 are UV-white (Fig. 4). Sex differences in the prevalence and spectral properties of the UV-blue patches are
244 already noticeable in yearlings (Table 1): UV-blue patches were present in 54% of the yearling lizards (29% of
245 the females, 80% of the males). Dorsal coloration showed little differences between newborn, yearling and
246 adult lizards (Table S1, Fig. S2). We did not quantify melanin-based patches, which were present in the ventral
247 surface of many resampled yearlings but largely absent in the throat and belly of newborn lizards.

248 Within-individual contrasts (i.e. MANOVAs) found significant chromatic and achromatic differences
249 between newborn and yearling throat colours for every yearling colour except UV+white (Table 2). Chromatic
250 distances between newborn-yearling paired spectra averaged 8.68 ± 1.55 JND, and were significantly greater
251 than the theoretical threshold of 3 JND for orange, yellow, and UV white, but not for UV+white (Fig. 3).
252 Within-individual achromatic distances were shorter than the theoretical threshold of 3 JNDs for every yearling
253 throat colour (Table 2).

254 Between-individual contrasts (PERMANOVA) found significant chromatic differences among all
255 pairwise combinations of yearling throat colours (**Table 2**). In contrast, achromatic differences were non-
256 significant for all contrasts except UV-white–orange and yellow–orange (**Table 2**). Bootstrapped chromatic
257 distances were larger than the threshold of 3 JNDs for all combinations, with Orange and UV+white being the
258 most different pair and Yellow–UV white the least different (**Fig. S3**).

259 Spectra from the UV-blue patches of yearlings showed a trough shape (with two peaks at the extremes
260 of the spectral range), which differed from the single peak around longer wavelengths found in newborn lizards
261 (**Fig. 5**). Paired spectra contrasting newborn ventral coloration and yearling UV-blue patches showed
262 significant differences according to the MANOVAs (**Table 2**). Chromatic and achromatic distances were large
263 enough to allow for discrimination, and more pronounced for males than females (**Table 2; Fig. 5**).

264 All contrasts between the average newborn colour ($N = 417$) and adult throat colours were statistically
265 significant according to the PERMANOVA except the chromatic contrast against UV+white adult throats
266 (**Table 3**). Bootstrapped chromatic and achromatic distances followed this same pattern, with the contrasts
267 against orange adult throats showing the largest distances, and the chromatic contrast against UV+white adult
268 throats being the only one below the theoretical threshold of 3 JNDs (**Fig. 6**). The distance-based
269 PERMANOVA fitted on chromatic contrasts between yearlings and adult throat colours yielded significant
270 results for all contrasts between lizards of different colours (**Tables 3 and S2**). Chromatic contrasts between
271 same-coloured lizards were non-significant except for yellow yearlings and adults. According to bootstrapped
272 chromatic distances, all contrasts between same-coloured yearlings and adults result shorter distances than the
273 theoretical threshold of 3 JNDs (**Fig 6 and S5**).

274 Adult UV-blue patches were found to differ chromatically and achromatically from the average
275 newborn coloration (**Table 3**), with contrasts in males resulting in larger bootstrapped distances than contrasts
276 in females (**Fig. 6**). Within-sex contrasts between the UV-blue coloration found in yearling and adult lizards
277 showed significant results only for chromatic differences (**Table 3**). Bootstrapped chromatic distances were
278 similar for both sexes and approximately half of those estimated between newborns and adults. Achromatic
279 distances were shorter than the theoretical threshold of 3 JNDs for both males and females (**Fig. 6**).

280

281 Discussion

282 We monitored colour expression in newborn (3–4 weeks old) and yearling (9–10 months old) *P. muralis* lizards
283 raised under semi-natural conditions, providing the first account of OCCs in their ventral and ventrolateral
284 coloration. Objective colour characterization revealed that newborn lizards exhibit enhanced UV reflectance
285 compared to yearlings, which begin to show incipient morph expression (**Figs. 2, 3**). Our results support the
286 distinction of two types of white ventral coloration in *P. muralis*: UV+white, typically found in newborns and
287 some adult female bellies, and UV-white, present in the throats of adults and the bellies of adult males ([Abalos
288 et al. 2016b](#); [Pérez i de Lanuza and Font 2025, in press](#)). Within-individual chromatic distances suggest that
289 OCCs to orange, yellow, and UV-white are perceptible to conspecifics. Among yearlings, throat colours are
290 theoretically distinguishable, with orange and UV+white being the most dissimilar and yellow and UV-white the

291 most similar. Contrasts with adults align with our repeated measures results, showing a similar rank order in
292 divergence from average newborn coloration among throat colours (orange > yellow > UV-white > UV+white).
293 Additionally, adult-yearling comparisons indicate that by 9–10 months after hatching, 92% of yearlings exhibit
294 throat coloration that is indistinguishable to conspecifics from adult colours.

295 These findings have significant implications for understanding the ontogeny of ventral colour morphs
296 in *P. muralis* and other lacertids. White morph lizards have often been assumed to retain their juvenile coloration
297 into adulthood (unlike orange, yellow, or mosaic lizards; e.g., [Pérez i de Lanuza et al. 2013a](#)), but our results
298 suggest that newborn lizards express a distinct UV+white coloration that conspecifics likely perceive as
299 chromatically different from adult throat colours. Similar UV+white coloration is found in the bellies (but not
300 throats) of many females in the eastern Pyrenees, suggesting that females in these populations may retain this
301 juvenile trait into adulthood (**Figs. 2, 4**, and Fig. 2 in [Pérez i de Lanuza and Font 2015](#)). We have previously
302 highlighted the need to establish reliable criteria for adult colour morph classification ([Pérez i de Lanuza et al.](#)
303 [2013a](#); [Abalos et al. 2020](#)). Based on our findings, we propose that decreased UV reflectance could be used to
304 identify adult morph expression in white subadult lizards in future studies of *P. muralis* ventral colour variation.

305 Within-individual contrasts suggest that OCCs to UV-blue in the OVS are perceptible to conspecifics,
306 and more apparent in males than in females (**Fig. 5**). However, nearly a year after hatching, yearling UV-blue
307 patches remain chromatically distinguishable from adult colours, unlike the throat and belly coloration. This
308 finding highlights an ontogenetic delay in the development of UV-blue patches, potentially reflecting
309 differences in the underlying mechanisms of colour production ([Pérez i de Lanuza et al. 2014](#); [Andrade et al.](#)
310 [2019](#)) and/or distinct selective pressures acting on these colour patches ([Hebets and Papaj 2005](#); [Cuthill et al.](#)
311 [2017](#); [Tibbetts et al. 2017](#)). Notably, UV-reflecting patches in the OVS undergo the most dramatic OCCs
312 possible, shifting their reflectance from the extreme long-wavelength end to the minimum of the lacertid visual
313 range in the UV wavelength band ([Pérez i de Lanuza et al. 2014](#); [Martin et al. 2015](#); [Fleishman and Font 2019](#)).
314 This causes reflectance in the UV-blue patches of yearlings to adopt a characteristic trough shape (i.e. two peaks
315 separated by a depression at intermediate wavelengths) which may explain the large chromatic distances found
316 in this study. This result suggests that the ontogeny of UV-blue patches involves an increase in UV reflectance
317 paired with a decrease in long-wavelength reflectance, instead of a progressive displacement towards shorter
318 wavelengths of a single reflectance peak. The intermediate stages of this process would result in the trough-
319 shaped spectra we observe in yearling lizards and adult females (**Fig. 6**).

320 Here, we did not test for possible adaptive explanations for the ventral and ventrolateral OCCs
321 described in *P. muralis*, which could represent a nonfunctional by-product of chromatophore maturation
322 ([Bagnara et al. 2007](#); [Umbers 2013](#)). However, our findings demonstrate that OCCs fulfil a key requirement for
323 social signals by being perceptible to conspecifics. Studies on OCCs in lizards have mostly focused on exploring
324 the function of bright tail colorations as a decoy, acting in combination with striped dorsal patterns and
325 autotomy to deflect predator attacks towards the expendable tail ([Castilla et al. 1999](#); [Hawlena et al. 2006](#);
326 [Hawlena 2009](#); [Watson et al. 2012](#); [Ortega et al. 2014](#); [Fresnillo et al. 2015b, a, 2016](#); [Murali et al. 2018](#)). In
327 contrast, a role in predator avoidance seems unlikely for ventral and ventrolateral OCCs. These colorations are
328 rarely visible to avian predators, which typically attack from above ([Marshall and Stevens 2014](#); [Marshall et al.](#)
329 [2015, 2016](#)), and are only occasionally visible to terrestrial predators as lizards are most commonly observed in

330 postures where these scales remain largely hidden (Pérez i de Lanuza et al. 2016a). However, lizards shift their
331 postures with increasing body temperature or during social interactions, making their ventral and ventrolateral
332 scales visible to observers positioned parallel to and level with the lizard's flank (Noble and Bradley 1933;
333 Kitzler 1941; Font and Carazo 2010; Pérez i de Lanuza et al. 2016a; Abalos et al. 2024). Considering the
334 ontogenetic decline in UV⁺white coloration and its distinct appearance to lizards compared to the colours
335 predominantly found in adults, we suggest that OCCs affecting ventral and ventrolateral scales could influence
336 social interactions by revealing juvenile status and sexual immaturity (Booth 1990). Juvenile-specific coloration
337 may reduce aggression from adult conspecifics, as observed in several vertebrate species (Hill 1989; Bergman
338 and Sheehan 2013; Ochi and Awata 2016; Hendershott et al. 2019), including lizards (Clark and Hall 1970;
339 Hawlena et al. 2006; Fresnillo et al. 2015a). In wild populations of *P. muralis*, agonistic confrontations between
340 adult males can escalate into physical fights, but juvenile or even subadult males (i.e. satellites) are often allowed
341 to set their home-range within the boundaries of an adult male's territory (Waltz 1982; Barbault and Mou 1986;
342 Edsman 1990, 2001; Brown et al. 1995; Font et al. 2012; Abalos et al. 2020). OCCs in ventral and ventrolateral
343 coloration could mediate this tolerance by signalling the subordinate status and low reproductive threat posed
344 by younger males, reducing the likelihood of aggression by resident males. Future studies should explore the
345 role of ventral and ventrolateral OCCs in mediating male-male competition and territorial dynamics. The
346 ontogenetic changes described here could also play a role in the context of mate choice. Evidence from lab
347 experiments and field studies suggest that, as in many other lizards (Tokarz 1995; Cuadrado 2006; Uller and
348 Olsson 2008; Vicente and Halloy 2016), pre-copulatory mate choice in wall lizards is largely under male control,
349 with males selecting larger conspecific females based on either chemical or visual stimuli (Edsman 1990, 2001;
350 Font and Desfilis 2002; Barbosa et al. 2006; Carazo et al. 2011; Font et al. 2012; Sacchi et al. 2015). Although
351 rudimentary male traits in females are often assumed to represent a by-product of their function in males, OCCs
352 in female ventral and ventrolateral scales may allow males to avoid courting immature females (mate
353 recognition) (Edward and Chapman 2011; Swierk and Langkilde 2013). At the same time, immature females
354 may benefit from being recognized as such if excessive male attention is costly to female fitness (Eberhard and
355 Cordero 2003; Arnqvist and Rowe 2005; Le Galliard et al. 2008). An objection to this hypothesis, however, lies
356 in explaining why body size alone might not be sufficient for receivers to identify young lizards. While body
357 size is an obvious correlate of age in most reptiles, it may not provide enough information for conspecifics to
358 distinguish between those that are sexually immature and those that are fully grown but still sexually inactive
359 (Booth 1990; Roucort Cezário et al. 2022).

360 Variation in the ventral and ventrolateral colour pattern of wall lizards could also be playing an
361 overlooked role in sexual selection by providing visual cues for social recognition (Tibbetts 2004; Tibbetts et
362 al. 2008; Sheehan and Bergman 2016). Many territorial lizards mitigate the costs of defence by showing
363 attenuated aggression toward known neighbours while remaining aggressive toward unfamiliar intruders (Qualls
364 and Jaeger 1991; Husak and Fox 2003; Osborne 2005; Baird 2013; Whiting and Miles 2019). In wall lizards,
365 males discriminate familiar rivals based on scent marks, allocating aggression based on perceived threat (Carazo
366 et al. 2008; Font et al. 2012). Given the sophistication of their visual system, it is plausible that wall lizards also
367 use visual cues for social recognition (Font et al. 2012; Pérez i de Lanuza and Font 2014b; Fleishman and Font
368 2019). In other taxa, visual cues have been shown to play a crucial role alongside chemical signals in individual
369 recognition (Tibbetts 2002; Sheehan and Tibbetts 2010), and in lizards like the tawny dragon (*Ctenophorus*
370 *decrevii*), throat coloration aids rival recognition (Osborne et al. 2012). While studies of social recognition in

371 intersexual interactions in lizards are limited (e.g. Font and Desfilis 2002; Leu et al. 2015; Bordogna et al. 2016),
372 evidence suggests that OCCs could influence mate choice. In *P. muralis*, ventral coloration is linked to male-
373 female pair bond formation (Pérez i de Lanuza et al. 2013, 2016b; Aguilar et al. 2022a), potentially acting as
374 mate-guarding or deterring harassment (in Den Bosch and Zandee 2001; Zaldívar-Rae and Drummond 2007;
375 Olsson et al. 2019; Abalos et al. 2020). Subtle chromatic and achromatic variation within the orange, white, and
376 yellow alternative colours, along with between-individual differences in patterning—such as melanin-based
377 patches or mosaic coloration—may hold relevance for social recognition (Martin et al. 2015; Pérez i de Lanuza
378 et al. 2018; Aguilar et al. 2022b). At the cohort level, ontogenetic transitions from uniform UV⁺white coloration
379 to distinct ventral and ventrolateral colour patterns enhance phenotypic divergence among maturing lizards, as
380 expected if showing a distinctive appearance becomes increasingly advantageous with age (Sheehan and
381 Tibbetts 2009; Sheehan and Bergman 2016; Gokcekus et al. 2021). Future studies could examine whether *P.*
382 *muralis* OCCs may influence social interactions by delaying the onset of sexually-selected colour signals, as well
383 as providing visual cues for individual recognition.

384 In conclusion, here we show that *P. muralis* ventral coloration undergoes OCCs perceptible to
385 conspecifics, though not always to humans, providing valuable insights into the ontogeny of lacertid coloration.
386 Future research should focus on detailed descriptions of OCCs, their cellular mechanisms, and their integration
387 with melanin-based patches (Sheehan et al. 2017; Pérez-Rodríguez et al. 2017; Zhang et al. 2023). Altogether,
388 our results underscore the importance of considering receiver perspectives in ethological studies of animal
389 coloration (Endler et al. 2023).

390

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407 **Author contributions**

408 Conceptualization and experimental design: J.A., G.P.L., F.A. and E.F.; Investigation: J.A., A.B.; Statistical
409 analysis: J.A.; Visualization: J.A.; Writing-original draft: J.A.; Writing-review & editing: all authors.

410 **Conflict of interests**

411 The authors declare no conflict of interest.

412 **Data availability**

413 The dataset used and analysed in this study will be made available upon acceptance.

414 **References**

415 Abalos J, i de Lanuza GP, Bartolomé A, et al (2021) Viability, behavior, and colour expression in the
416 offspring of matings between common wall lizard *Podarcis muralis* colour morphs. *Curr Zool* 1–15.
417 <https://doi.org/10.1093/cz/zoab039>

418 Abalos J, Pérez i de Lanuza G, Bartolomé A, et al (2024) Behavioral threat and appeasement signals take
419 precedence over static colors in lizard contests. *Behav Ecol* 35:.
420 <https://doi.org/10.1093/beheco/arae045>

421 Abalos J, Pérez i de Lanuza G, Bartolomé A, et al (2020) No evidence for differential sociosexual behavior
422 and space use in the color morphs of the European common wall lizard (*Podarcis muralis*). *Ecol Evol*
423 00:1–20. <https://doi.org/10.1002/ece3.6659>

424 Abalos J, Pérez i de Lanuza G, Carazo P, Font E (2016a) The role of male coloration in the outcome of
425 staged contests in the European common wall lizard (*Podarcis muralis*). *Behaviour* 153:607–631.
426 <https://doi.org/10.1163/1568539X-00003366>

427 Abalos J, Pérez i de Lanuza G, Reguera S, et al (2016b) Whiter than white: dimorphism in the white ventral
428 coloration of *Podarcis muralis*. XIV Iber Congr Herpetol Poster.
429 https://doi.org/https://www.lacerta.de/AF/Bibliografie/BIB_10924.pdf

430 Abramjan A, Bauerová A, Somerová B, Frynta D (2015) Why is the tongue of blue-tongued skinks blue?
431 Reflectance of lingual surface and its consequences for visual perception by conspecifics and predators.
432 *Sci Nat* 102:.. <https://doi.org/10.1007/s00114-015-1293-4>

433 Aguilar P, Andrade P, Afonso S, et al (2022a) No genomic repercussions of assortative pairing in a colour
434 polymorphic lizard. *J Evol Biol* 35:648–656. <https://doi.org/10.1111/jeb.13990>

435 Aguilar P, Andrade P, Pérez I De Lanuza G (2022b) Epistatic interactions between pterin and carotenoid
436 genes modulate intra-morph color variation in a lizard. *Integr Zool* 17:44–53.
437 <https://doi.org/10.1111/1749-4877.12570>

438 Aguilar P, Pérez i de Lanuza G, Carneiro M, et al (2024) The role of historical biogeography in shaping colour

- 439 morph diversity in the common wall lizard. *Mol Ecol* 33:e17338. <https://doi.org/10.1111/MEC.17338>
- 440 Anderson MJ (2014) *Permutational Multivariate Analysis of Variance (PERMANOVA)*. Wiley StatsRef Stat
441 Ref Online 1–15. <https://doi.org/10.1002/9781118445112.stat07841>
- 442 Andrade P, Pinho C, de Lanuza GP i., et al (2019) Regulatory changes in pterin and carotenoid genes underlie
443 balanced color polymorphisms in the wall lizard. *Proc Natl Acad Sci U S A* 116:5633–5642.
444 <https://doi.org/10.1073/pnas.1820320116>
- 445 Arnold EN, Arribas O, Carranza S (2007) Zootaxa, Systematics of the Palaearctic and Oriental lizard tribe
446 Lacertini (Squamata: Lacertidae: Lacertinae), with descriptions of eight new genera. *Zootaxa* 1430:1–86.
447 <https://doi.org/10.11646/zootaxa.1430.1.1>
- 448 Arnqvist G, Rowe L (2005) *Sexual conflict*. Princeton university press
- 449 Badiane A, Carazo P, Price-Rees SJ, et al (2018) Why blue tongue? A potential UV-based deimatic display in a
450 lizard. *Behav Ecol Sociobiol* 72:1–11. <https://doi.org/10.1007/s00265-018-2512-8>
- 451 Badiane A, Font E (2021) Information content of ultraviolet - reflecting colour patches and visual perception
452 of body coloration in the Tyrrhenian wall lizard *Podarcis tiliguerta*. *Behav Ecol Sociobiol* 1–15.
453 <https://doi.org/10.1007/s00265-021-03023-2>
- 454 Badiane A, Pérez i de Lanuza G, García-Custodio M del C, et al (2017) Colour patch size and measurement
455 error using reflectance spectrophotometry. *Methods Ecol Evol* 8:1585–1593.
456 <https://doi.org/10.1111/2041-210X.12801>
- 457 Bagnara JT, Fernandez PJ, Fujii R (2007) On the blue coloration of vertebrates. *Pigment Cell Res* 20:14–26.
458 <https://doi.org/10.1111/j.1600-0749.2006.00360.x>
- 459 Baird TA (2013) Lizards and other reptiles as model systems for the study of contest behaviour. In: Hardy
460 ICW, Briffa M (eds) *Animal Contests*. Cambridge University Press, Cambridge, pp 258–286
- 461 Barbault R, Mou YP (1986) A Population Analysis of the Common Wall Lizard *Podarcis muralis* in
462 Southwestern France. *Stud Herpetol Rocek Z* (ed) 513–518
- 463 Barbosa D, Font E, Desfilis E, Carretero MA (2006) Chemically mediated species recognition in closely
464 related *Podarcis* wall lizards. *J Chem Ecol* 32:1587–1598. <https://doi.org/10.1007/s10886-006-9072-5>
- 465 Bergman TJ, Sheehan MJ (2013) Social knowledge and signals in primates. *Am J Primatol* 75:683–694.
466 <https://doi.org/10.1002/ajp.22103>
- 467 Booth CL (1990) Evolutionary significance of ontogenetic colour change in animals. *Biol J Linn Soc* 40:125–
468 163. <https://doi.org/10.1111/j.1095-8312.1990.tb01973.x>
- 469 Bordogna G, Cunningham G, Fitzpatrick LJ, et al (2016) An experimental test of relatedness-based mate

- 470 discrimination in a social lizard. *Behav Ecol Sociobiol* 70:2139–2147
- 471 Bradbury JW, Vehrencamp SL (2011) *Principles of Animal Communication*, 2nd edn. Sinauer Associates,
472 Sunderland (MA)
- 473 Brock KM, Chelini MC, Ayton C, et al (2022a) Colour morph predicts social behaviour and contest outcomes
474 in a polymorphic lizard (*Podarcis erhardii*). *Anim Behav* 191:91–103.
475 <https://doi.org/10.1016/j.anbehav.2022.06.017>
- 476 Brock KM, McTavish EJ, Edwards DL (2022b) Color Polymorphism is a Driver of Diversification in the
477 Lizard Family Lacertidae. *Syst Biol* 71:24–39. <https://doi.org/10.1093/sysbio/syab046>
- 478 Brown RM, Gist DH, Taylor DH (1995) Home range ecology of an introduced population of the European
479 wall lizard *Podarcis muralis* (Lacertilia; Lacertidae) in Cincinnati, Ohio. *Am Midl Nat* 344–359
- 480 Buckman D (1985) Color change in insects. In: Bagnara J, Klaus SN, Paul E, Schartl M (eds) *Pigment Cell*.
481 University of Tokyo Press, Tokyo, pp 209–217
- 482 Calsbeek B, Hasselquist D, Clobert J (2010) Multivariate phenotypes and the potential for alternative
483 phenotypic optima in wall lizard (*Podarcis muralis*) ventral colour morphs. *J Evol Biol* 23:1138–1147.
484 <https://doi.org/10.1111/j.1420-9101.2010.01978.x>
- 485 Carazo P, Font E, Desfilis E (2008) Beyond “nasty neighbours” and “dear enemies”? Individual recognition
486 by scent marks in a lizard (*Podarcis hispanica*). *Anim Behav* 76:1953–1963.
487 <https://doi.org/10.1016/j.anbehav.2008.08.018>
- 488 Carazo P, Font E, Desfilis E (2011) The role of scent marks in female choice of territories and refuges in a
489 lizard (*Podarcis hispanica*). *J Comp Psychol* 125:362–365. <https://doi.org/10.1037/a0023404>
- 490 Castilla AM, Gosá A, Galán P, Pérez-Mellado V (1999) Green Tails in Lizards of the Genus *Podarcis* : Do
491 They Influence the Intensity of Predation ? *Herpe* 55:530–537.
492 <https://doi.org/https://www.jstor.org/stable/3893319>
- 493 Caves EM, Nowicki S, Johnsen S (2019) Von Uexküll revisited: Addressing human biases in the study of
494 animal perception. *Integr Comp Biol* 59:1451–1462. <https://doi.org/10.1093/icb/icz073>
- 495 Clark DR, Hall RJ (1970) Function of the blue tail coloration of the five-lined skink *Eumeces fasciatus*.
496 *Herpetologica* 26:271–274
- 497 Cronin TW, Bok MJ (2016) Photoreception and vision in the ultraviolet. *J Exp Biol* 219:2790–2801.
498 <https://doi.org/10.1242/jeb.128769>
- 499 Cuadrado M (2006) Mate guarding and social mating system in male common chameleons (*Chamaeleo*
500 *chamaeleon*). *J Zool* 255:425–435. <https://doi.org/10.1017/S0952836901001510>

- 501 Cuthill IC, Allen WL, Arbuckle K, et al (2017) The biology of color. *Science* (80-) 357:.
502 <https://doi.org/10.1126/science.aan0221>
- 503 de la Cruz F, Pérez i de Lanuza G, Font E (2023) Signalling on islands : the case of Lilford ' s wall lizard (
504 *Podarcis lilfordi gigliolii*) from Dragonera. *Biol J Linn Soc* 138:1–20.
505 <https://doi.org/https://doi.org/10.1093/biolinnean/blac152>
- 506 Eberhard WG, Cordero C (2003) Sexual conflict and female choice. *Trends Ecol Evol* 18:438–439.
507 [https://doi.org/10.1016/S0169-5347\(03\)00180-0](https://doi.org/10.1016/S0169-5347(03)00180-0)
- 508 Edsman L (2001) Female mate choice of male characteristics and resources in the wall lizard. In: Vicente L,
509 Crespo EG (eds) *Mediterranean basin lizards: a biological approach*. Instituto de Conservação da
510 Natureza, Lisboa, pp 133–134
- 511 Edsman L (1990) Territoriality and competition in wall lizards. Dept. of Zool., Univ. of Stockholm
- 512 Edward DA, Chapman T (2011) The evolution and significance of male mate choice. *Trends Ecol Evol*
513 26:647–654
- 514 Ekner A, Sajkowska Z, Dudek K, Tryjanowski P (2011) Medical cautery units as a permanent and non-
515 invasive method of marking lizards. *Acta Herpetol* 6:229–236.
516 https://doi.org/10.13128/Acta_Herpetol-9346
- 517 Endler JA (1990) On the measurement and classification of colour in studies of animal colour patterns. *Biol J*
518 *Linn Soc* 41:315–352. <https://doi.org/10.1111/j.1095-8312.1990.tb00839.x>
- 519 Endler JA, Kemp DJ, Reznick DN, et al (2023) How to generate and test hypotheses about colour : insights
520 from half a century of guppy research
- 521 Escoriza D (2024) Environmental colour pattern variation in Mediterranean *Podarcis*. 1–8
- 522 Fleishman LJ, Font E (2019) Sensory Processing in Relation to Signaling Behavior
- 523 Font E, Barbosa D, Sampedro C, Carazo P (2012) Social behavior, chemical communication, and adult
524 neurogenesis: Studies of scent mark function in *Podarcis* wall lizards. *Gen Comp Endocrinol* 177:9–17.
525 <https://doi.org/10.1016/j.ygcen.2012.02.015>
- 526 Font E, Carazo P (2010) Animals in translation: Why there is meaning (but probably no message) in animal
527 communication. *Anim Behav* 80:e1–e6. <https://doi.org/10.1016/j.anbehav.2010.05.015>
- 528 Font E, Desfilis E (2002) Chemosensory recognition of familiar and unfamiliar conspecifics by juveniles of
529 the Iberian wall lizard *Podarcis hispanica*. *Ethology* 108:319–330. <https://doi.org/10.1046/j.1439-0310.2002.00782.x>
- 530
- 531 Font E, Pérez i de Lanuza G, Sampedro C (2009) Ultraviolet reflectance and cryptic sexual dichromatism in

- 532 the ocellated lizard, *Lacerta (Timon) lepida* (Squamata: Lacertidae). *Biol J Linn Soc* 97:766–780.
533 <https://doi.org/10.1111/j.1095-8312.2009.01251.x>
- 534 Frédéric B, Mills SC, Denoël M, et al (2010) Colour differentiation in a coral reef fish throughout ontogeny:
535 habitat background and flexibility. *Aquat Biol* 9:271–277
- 536 Fresnillo B, Belliure J, Cuervo JJ (2015a) Red coloration in juvenile spiny-footed lizards, *Acanthodactylus*
537 *erythrurus*, reduces adult aggression. *Anim Behav* 102:59–67.
538 <https://doi.org/10.1016/j.anbehav.2015.01.004>
- 539 Fresnillo B, Belliure J, Cuervo JJ (2016) Ontogenetic shifts in risk behaviours are related to body size and
540 coloration in spiny-footed lizards. *Anim Behav* 119:165–172.
541 <https://doi.org/10.1016/J.ANBEHAV.2016.07.009>
- 542 Fresnillo B, Belliure J, Cuervo JJ (2015b) Red tails are effective decoys for avian predators. *Evol Ecol* 29:123–
543 135. <https://doi.org/10.1007/S10682-014-9739-2/TABLES/2>
- 544 Galeotti P, Sacchi R, Pellitteri-Rosa D, et al (2013) Colour polymorphism and alternative breeding
545 strategies: effects of parent's colour morph on fitness traits in the common wall lizard. *Evol Biol*
546 40:385–394. <https://doi.org/10.1007/s11692-012-9222-3>
- 547 Gokcekus S, Firth JA, Regan C, Sheldon BC (2021) Recognising the key role of individual recognition in
548 social networks. *Trends Ecol Evol* 36:1024–1035. <https://doi.org/10.1016/j.tree.2021.06.009>
- 549 Hawlena D (2009) Colorful tails fade when lizards adopt less risky behaviors. *Behav Ecol Sociobiol* 64:205–
550 213. <https://doi.org/10.1007/s00265-009-0837-z>
- 551 Hawlena D, Bochnik R, Abramsky Z, Bouskila A (2006) Blue tail and striped body: why do lizards change
552 their infant costume when growing up? *Behav Ecol* 17:889–896.
553 <https://doi.org/10.1093/BEHECO/ARL023>
- 554 Hebets EA, Papaj DR (2005) Complex signal function: Developing a framework of testable hypotheses.
555 *Behav Ecol Sociobiol* 57:197–214. <https://doi.org/10.1007/s00265-004-0865-7>
- 556 Hendershott R, Hu G, Groves C, Behie A (2019) Natal-to-juvenile pelage change in free-living François'
557 (*Trachypithecus francoisi*) and Cat Ba langurs (*T. poliocephalus*). *Vietnamese J Primatol* 3:55–69
- 558 Hill GE (1989) Late spring arrival and dull nuptial plumage: aggression avoidance by yearling males? *Anim*
559 *Behav* 37:665–673. [https://doi.org/10.1016/0003-3472\(89\)90045-6](https://doi.org/10.1016/0003-3472(89)90045-6)
- 560 Husak JF, Fox SF (2003) Adult male collared lizards, *Crotaphytus collaris*, increase aggression towards displaced
561 neighbours. *Anim Behav* 65:391–396. <https://doi.org/10.1006/anbe.2003.2058>
- 562 Huyghe K, Vanhooydonck B, Herrel A, et al (2007) Morphology, performance, behavior and ecology of three
563 color morphs in males of the lizard *Podarcis melisellensis*. *Integr Comp Biol* 47:211–220.

564 <https://doi.org/10.1093/icb/icm043>

565 in Den Bosch HAJ, Zandee M (2001) Courtship behaviour in lacertid lizards: phylogenetic interpretations of
566 the *Lacerta kulzeri* complex (Reptilia: Lacertidae). Netherlands J Zool 51:263–284

567 Kemp DJ, Herberstein ME, Fleishman LJ, et al (2015) An integrative framework for the appraisal of
568 coloration in nature. Am Nat 185:705–724. <https://doi.org/10.1086/681021>

569 Kitzler G (1941) Die Paarungsbiologie einiger Eidechsen. Z Tierpsychol 353–402

570 Le Galliard J-F, Cote J, Fitze PS (2008) Lifetime and intergenerational fitness consequences of harmful male
571 interactions for female lizards. Ecology 89:56–64

572 Leu ST, Burzacott D, Whiting MJ, Bull CM (2015) Mate Familiarity Affects Pairing Behaviour in a Long-
573 Term Monogamous Lizard: Evidence from Detailed Bio-Logging and a 31-Year Field Study. Ethology
574 121:760–768. <https://doi.org/10.1111/eth.12390>

575 Maia R, Gruson H, Endler JA, White TE (2019) pavo 2: New tools for the spectral and spatial analysis of
576 colour in r. Methods Ecol Evol 10:1097–1107. <https://doi.org/10.1111/2041-210X.13174>

577 Majerus MEN (2008) Industrial Melanism in the Peppered Moth, *Biston betularia*: An Excellent Teaching
578 Example of Darwinian Evolution in Action. Evol Educ Outreach 2:63–74.
579 <https://doi.org/10.1007/s12052-008-0107-y>

580 Mangiacotti M, Pezzi S, Fumagalli M, et al (2019) Seasonal Variations in Femoral Gland Secretions Reveals
581 some Unexpected Correlations Between Protein and Lipid Components in a Lacertid Lizard. J Chem
582 Ecol 45:673–683. <https://doi.org/10.1007/s10886-019-01092-2>

583 Marshall KLA, Philpot KE, Damas-Moreira I, Stevens M (2015) Intraspecific colour variation among lizards
584 in distinct island environments enhances local camouflage. PLoS One 10:1–20.
585 <https://doi.org/10.1371/journal.pone.0135241>

586 Marshall KLA, Philpot KE, Stevens M (2016) Microhabitat choice in island lizards enhances camouflage
587 against avian predators. Sci Rep 6:1–10. <https://doi.org/10.1038/srep19815>

588 Marshall KLA, Stevens M (2014) Wall lizards display conspicuous signals to conspecifics and reduce
589 detection by avian predators. Behav Ecol 25:1325–1337. <https://doi.org/10.1093/beheco/aru126>

590 Martin M, Le Galliard JF, Meylan S, Loew ER (2015) The importance of ultraviolet and near-infrared
591 sensitivity for visual discrimination in two species of lacertid lizards. J Exp Biol 218:458–465.
592 <https://doi.org/10.1242/jeb.115923>

593 Martinez Arbizu P (2017) pairwiseAdonis: Pairwise multilevel comparison using adonis. R Packag version 1:

594 Miñano MR, While GM, Yang W, et al (2021) Climate shapes the geographic distribution and introgressive

- 595 spread of color ornamentation in common wall lizards. *Am Nat.* <https://doi.org/10.1086/715186>
- 596 Murali G, Merilaita S, Kodandaramaiah U (2018) Grab my tail: evolution of dazzle stripes and colourful tails
597 in lizards. *J Evol Biol* 31:1675–1688
- 598 Names G, Martin M, Badiane A, Le Galliard JF (2019) The relative importance of body size and UV
599 coloration in influencing male-male competition in a lacertid lizard. *Behav Ecol Sociobiol* 73:.
600 <https://doi.org/10.1007/s00265-019-2710-z>
- 601 Noble GK, Bradley HT (1933) The mating behavior of lizards; its bearing on the theory of sexual selection.
602 *Ann N Y Acad Sci* 35:25–100. <https://doi.org/10.1111/j.1749-6632.1933.tb55365.x>
- 603 O'Shea M (2021) *Lizards of the World: A Guide to Every Family*. Princeton University Press
- 604 Ochi H, Awata S (2016) Resembling the Juvenile Colour of Host Cichlid Facilitates Access of the Guest
605 Cichlid to Host Territory Author (s): Haruki Ochi and Satoshi Awata Published by : Brill Stable URL :
606 <http://www.jstor.org/stable/40296094> REFERENCES Linked references are av. 146:741–756
- 607 Oksanen AJ, Blanchet FG, Friendly M, et al (2016) Vegan: community ecology package. R Packag version
608 2.0:321–326
- 609 Olsson M, Schwartz TS, Wapstra E, Shine R (2019) How accurately do behavioural observations predict
610 reproductive success in free-ranging lizards? *Biol Lett* 15:.
<https://doi.org/10.1098/rsbl.2019.0030>
- 611 Olsson M, Stuart-Fox D, Ballen C (2013) Genetics and evolution of colour patterns in reptiles. *Semin Cell*
612 *Dev Biol* 24:529–541. <https://doi.org/10.1016/j.semcdb.2013.04.001>
- 613 Ortega J, López P, Martín J (2014) Conspicuous blue tails, dorsal pattern morphs and escape behaviour in
614 hatchling Iberian wall lizards (*Podarcis hispanicus*). *Biol J Linn Soc* 113:1094–1106
- 615 Ortega J, Martín J, Crochet PA, et al (2019) Seasonal and interpopulational phenotypic variation in
616 morphology and sexual signals of *Podarcis liolepis* lizards. *PLoS One* 14:1–25.
617 <https://doi.org/10.1371/journal.pone.0211686>
- 618 Osborne L (2005) Rival recognition in the territorial tawny dragon (*Ctenophorus decresii*). *Acta Ethol* 8:45–
619 50. <https://doi.org/10.1007/s10211-005-0108-6>
- 620 Osborne L, Umbers KDL, Backwell PRY, Keogh JS (2012) Male tawny dragons use throat patterns to
621 recognize rivals. *Naturwissenschaften* 99:869–872. <https://doi.org/10.1007/s00114-012-0968-3>
- 622 Pérez-Rodríguez L, Jovani R, Stevens M (2017) Shape matters: Animal colour patterns as signals of individual
623 quality. *Proc. R. Soc. B Biol. Sci.* 284
- 624 Pérez i de Lanuza G, Abalos J, Bartolomé A, Font E (2018) Through the eye of a lizard: hue discrimination in
625 a lizard with ventral polymorphic coloration. *J Exp Biol* 221;jeb.169565.

- 626 <https://doi.org/10.1242/jeb.169565>
- 627 Pérez i de Lanuza G, Bellati A, Pellitteri-Rosa D, et al (2019) Colour variation between different lineages of a
628 colour polymorphic lizard. *J Zool* 308:175–187. <https://doi.org/10.1111/jzo.12658>
- 629 Pérez i de Lanuza G, Carazo P, Font E (2014) Colours of quality: Structural (but not pigment) coloration
630 informs about male quality in a polychromatic lizard. *Anim Behav* 90:73–81.
631 <https://doi.org/10.1016/j.anbehav.2014.01.017>
- 632 Pérez i de Lanuza G, Carretero MA, Font E (2017) Intensity of male-male competition predicts morph
633 diversity in a color polymorphic lizard. *Evolution (N Y)* 71:1832–1840.
634 <https://doi.org/10.1111/evo.13256>
- 635 Pérez i de Lanuza G, Carretero MÁ, Font E (2016a) Thermal dependence of signalling: do polymorphic wall
636 lizards compensate for morph-specific differences in conspicuousness? *Behav Ecol Sociobiol* 70:1151–
637 1159. <https://doi.org/10.1007/s00265-016-2123-1>
- 638 Pérez i de Lanuza G, Font E (2014a) Ultraviolet vision in lacertid lizards: evidence from retinal structure, eye
639 transmittance, SWS1 visual pigment genes and behaviour. *J Exp Biol* 217:2899–909.
640 <https://doi.org/10.1242/jeb.104281>
- 641 Pérez i de Lanuza G, Font E (2015) Differences in conspicuousness between alternative color morphs in a
642 polychromatic lizard. *Behav Ecol* 26:1432–1446. <https://doi.org/10.1093/beheco/arv075>
- 643 Pérez i de Lanuza G, Font E (2014b) Ultraviolet vision in lacertid lizards: evidence from retinal structure, eye
644 transmittance, SWS1 visual pigment genes and behaviour. *J Exp Biol* 217:2899–909.
645 <https://doi.org/10.1242/jeb.104281>
- 646 Pérez i de Lanuza G, Font E, Carazo P (2013) Color-assortative mating in a color-polymorphic lacertid lizard.
647 *Behav Ecol* 24:273–279. <https://doi.org/10.1093/beheco/ars164>
- 648 Pérez i de Lanuza G, Font E, Carretero MÁ (2016b) Colour assortative pairing in a colour polymorphic lizard
649 is independent of population morph diversity. *Sci Nat* 103:82. <https://doi.org/10.1007/s00114-016-1407-7>
650
- 651 Qualls CP, Jaeger RG (1991) Dear enemy recognition in *Anolis carolinensis*. *J Herpetol* 25:361.
652 <https://doi.org/10.2307/1564599>
- 653 R Development Core Team (2017) R: A Language and Environment for Statistical Computing
- 654 Rankin KJ, McLean CA, Kemp DJ, Stuart-Fox D (2016) The genetic basis of discrete and quantitative colour
655 variation in the polymorphic lizard, *Ctenophorus decresii*. *BMC Evol Biol* 16:179.
656 <https://doi.org/10.1186/s12862-016-0757-2>
- 657 Rivas JA, Burghardt GM (2002) Crotalomorphism: A metaphor to understand anthropomorphism by omission.

- 658 In: Bekoff M, Colin A, Burghardt GM (eds) The cognitive animal: empirical and theoretical
659 perspectives on animal cognition. MIT Press, Cambridge, pp 9–17
- 660 Roucourt Cezário R, Moreira Therézio E, Marletta A, et al (2022) Ontogenetic colour change of a sexual
661 ornament in males of a damselfly: female mimicry, crypsis or both? Sci Nat 109:2.
662 <https://doi.org/10.1007/s00114-021-01775-5>
- 663 Runemark A, Hansson B, Pafilis P, et al (2010) Island biology and morphological divergence of the Skyros
664 wall lizard *Podaris gaigeae*: a combined role for local selection and genetic drift on color morph
665 frequency divergence? BMC Evol Biol 10:269. <https://doi.org/10.1186/1471-2148-10-269>
- 666 Sacchi R, Coladonato AJ, Battaiola M, et al (2021) Subjective resource value affects aggressive behavior
667 independently of resource-holding-potential and color morphs in male common wall lizard. J Ethol
668 39:179–189. <https://doi.org/10.1007/s10164-021-00690-6>
- 669 Sacchi R, Ghitti M, Scali S, et al (2015) Common wall lizard females (*Podaris muralis*) do not actively choose
670 males based on their colour morph. Ethology 121:1145–1153. <https://doi.org/10.1111/eth.12431>
- 671 Sacchi R, Pupin F, Gentili A, et al (2009) Male-male combats in a polymorphic lizard: Residency and size, but
672 not color, affect fighting rules and contest outcome. Aggress Behav 35:274–283.
673 <https://doi.org/10.1002/ab.20305>
- 674 Sacchi R, Scali S, Pupin F, et al (2007) Microgeographic variation of colour morph frequency and biometry of
675 common wall lizards. 273:389–396. <https://doi.org/10.1111/j.1469-7998.2007.00342.x>
- 676 Salvador A (2014) Reptiles, 2ª edición revisada y aumentada. Fauna Ibérica 10:
- 677 Santiago C, Green NF, Hamilton N, et al (2020) Does conspicuousness scale linearly with colour distance? A
678 test using reef fish: Detectability of suprathreshold colours. Proc R Soc B Biol Sci 287:.
679 <https://doi.org/10.1098/rspb.2020.1456>
- 680 Scali S, Sacchi R, Falaschi M, et al (2019) Mirrored images but not silicone models trigger aggressive
681 responses in male common wall lizards. Acta Herpetol 14:35–41.
682 https://doi.org/10.13128/Acta_Herpetol-24651
- 683 Sheehan MJ, Bergman TJ (2016) Is there an evolutionary trade-off between quality signaling and social
684 recognition? Behav Ecol 27:2–13. <https://doi.org/10.1093/beheco/arv109>
- 685 Sheehan MJ, Choo J, Tibbetts EA (2017) Heritable variation in colour patterns mediating individual
686 recognition. R Soc Open Sci 4: <https://doi.org/10.1098/rsos.161008>
- 687 Sheehan MJ, Tibbetts EA (2010) Selection for individual recognition and the evolution of polymorphic
688 identity signals in polistes paper wasps. J Evol Biol 23:570–577. <https://doi.org/10.1111/j.1420-9101.2009.01923.x>
689

- 690 Sheehan MJ, Tibbetts EA (2009) Evolution of identity signals: Frequency-dependent benefits of distinctive
691 phenotypes used for social recognition. *Evolution* (N Y) 63:3106–3113.
692 <https://doi.org/10.1111/j.1558-5646.2009.00833.x>
- 693 Siddiqi A, Cronin TW, Loew ER, et al (2004) Interspecific and intraspecific views of color signals in the
694 strawberry poison frog *Dendrobates pumilio*. *J Exp Biol* 207:2471–2485.
695 <https://doi.org/10.1242/jeb.01047>
- 696 Smale MJ, Kok HM (1983) The occurrence and feeding of *Pomatomus saltarix* (el) and *Lichia amia* (leervis)
697 juveniles in two Cape south coast estuaries. *South African J Zool* 18:337–342.
698 <https://doi.org/10.1080/02541858.1983.11447836>
- 699 Solan T De, Geniez P, Solan T De, et al (2023) Colour polymorphism and conspicuousness do not increase
700 speciation rates in Lacertids. *Peer Community J* 3:e111. <https://doi.org/10.24072/pcjournal.345>
- 701 Speybroeck J, Beukema W, Bok B, et al (2016) Field guide to the amphibians and reptiles of Britain and
702 Europe. Bloomsbury Publishing Plc, London
- 703 Stapley J, Whiting MJ (2006) Ultraviolet signals fighting ability in a lizard. *Biol Lett* 2:169–172.
704 <https://doi.org/10.1098/rsbl.2005.0419>
- 705 Stuart-Fox D, Aulsebrook A, Rankin KJ, et al (2020) Convergence and divergence in lizard colour
706 polymorphisms. *Biol Rev* 000–000:1–21. <https://doi.org/10.1111/brv.12656>
- 707 Stückler S, Cloer S, Hödl W, Preininger D (2022) Carotenoid intake during early life mediates ontogenetic
708 colour shifts and dynamic colour change during adulthood. *Anim Behav* 187:121–135
- 709 Svensson EI (2017) Back to basics: using colour polymorphisms to study evolutionary processes. *Mol Ecol*
710 26:2204–2211. <https://doi.org/10.1111/mec.14025>
- 711 Swierk L, Langkilde T (2013) Bearded ladies: Females suffer fitness consequences when bearing male traits.
712 *Biol Lett* 9:. <https://doi.org/10.1098/rsbl.2013.0644>
- 713 Tibbetts EA (2004) Complex social behaviour can select for variability in visual features: A case study in
714 *Polistes* wasps. *Proc R Soc B Biol Sci* 271:1955–1960. <https://doi.org/10.1098/rspb.2004.2784>
- 715 Tibbetts EA (2002) Visual signals of individual identity in the wasp *Polistes fuscatus*. *Proc R Soc B Biol Sci*
716 269:1423–1428. <https://doi.org/10.1098/rspb.2002.2031>
- 717 Tibbetts EA, Mullen SP, Dale J (2017) Signal function drives phenotypic and genetic diversity: The effects of
718 signalling individual identity, quality or behavioural strategy. *Philos Trans R Soc B Biol Sci*
719 372:20160347. <https://doi.org/10.1098/rstb.2016.0347>
- 720 Tibbetts EA, Sheehan MJ, Dale J (2008) A testable definition of individual recognition. *Trends Ecol Evol*
721 23:356. <https://doi.org/10.1016/j.tree.2008.03.007>

- 722 Tokarz RR (1995) Mate choice in lizards: a review. *Herpetol Monogr* 9:17–40.
723 <https://doi.org/10.2307/1466994>
- 724 Uller T, Olsson M (2008) Multiple paternity in reptiles: Patterns and processes. *Mol Ecol* 17:2566–2580.
725 <https://doi.org/10.1111/j.1365-294X.2008.03772.x>
- 726 Umbers KDL (2013) On the perception, production and function of blue colouration in animals. *J Zool*
727 289:229–242. <https://doi.org/10.1111/jzo.12001>
- 728 Vicente NS, Halloy M (2016) Chemical recognition of conspecifics in a neotropical lizard, *Liolaemus pacha*
729 (Iguania: Liolaemidae): relation to visual displays, season and sex. *J Ethol* 34:329–335
- 730 Vorobyev M, Osorio D (1998) Receptor noise as a determinant of colour thresholds. *Proc R Soc London Ser*
731 *B Biol Sci* 265:351–358. <https://doi.org/10.1098/rspb.1998.0302>
- 732 Vorobyev M, Osorio D, Bennett ATD, et al (1998) Tetrachromacy, oil droplets and bird plumage colours. *J*
733 *Comp Physiol - A Sensory, Neural, Behav Physiol* 183:621–633.
734 <https://doi.org/10.1007/s003590050286>
- 735 Waltz EC (1982) Alternative mating tactics and the law of diminishing returns: The satellite threshold model.
736 *Behav Ecol Sociobiol* 10:75–83. <https://doi.org/10.1007/BF00300166>
- 737 Watson CM, Roelke CE, Pasichnyk PN, Cox CL (2012) The fitness consequences of the autotomous blue tail
738 in lizards : an empirical test of predator response using clay models. 115:339–344
- 739 Wellenreuther M, Svensson EI, Hansson B (2014) Sexual selection and genetic colour polymorphisms in
740 animals. *Mol Ecol* 23:5398–5414. <https://doi.org/10.1111/mec.12935>
- 741 White TE, Kemp DJ (2016) Colour polymorphism. *Curr Biol* 26:R517–R518.
742 <https://doi.org/10.1016/j.cub.2016.03.017>
- 743 Whiting MJ, Miles DB (2019) Behavioral ecology of aggressive behavior in lizards. In: Bels VL, Russel AP
744 (eds) *Behavior of Lizards*. CRC Press, Boca Ratón, FL, pp 289-319.
- 745 Whiting MJ, Stuart-Fox DM, O'Connor D, et al (2006) Ultraviolet signals ultra-aggression in a lizard. *Anim*
746 *Behav* 72:353–363. <https://doi.org/10.1016/j.anbehav.2005.10.018>
- 747 Yang C, Chen S, Wang J (2023) Ontogenetic color change in the tail of blue-tailed skinks (*Plestodion*
748 *elegans*). *Ecol Evol* 13:e10152
- 749 Yang W, Feiner N, Pinho C, et al (2021) Extensive introgression and mosaic genomes of Mediterranean
750 endemic lizards. *Nat Commun* 2021 12:1–8. <https://doi.org/10.1038/s41467-021-22949-9>
- 751 Zaldívar-Rae J, Drummond H (2007) Female accompaniment by male whiptail lizards: Is it mate guarding?
752 *Behaviour* 144:1383–1402. <https://doi.org/10.1163/156853907782418187>

753 Zhang G, Yallapragada VJ, Halperin T, et al (2023) Lizards exploit the changing optics of developing
754 chromatophore cells to switch defensive colors during ontogeny. Proc Natl Acad Sci 120:e2215193120
755

756 **Tables**

757

758 **Table 1.** Spectral variables (mean \pm CI₉₅) describing reflectance curves in the ventral coloration of newborn,
 759 yearling, and adult lizards. Spectra have been normalized by subtracting the minimum value at all wavelengths.

760 Qt = luminance ($R_{300-700}$), C_{UV} = UV chroma ($R_{300-400}/R_{300-700}$), $\lambda \beta_{\max_{325-560}}$ = wavelength at which reflectance
 761 shows the maximum positive slope in the 325-560 nm waveband, $\lambda R_{\text{mid}_{300-450}}$ and $\lambda R_{\text{mid}_{400-650}}$ = wavelength
 762 at which reflectance is halfway between its minimum and its maximum for the wavebands 300-450 nm and
 763 400-650 nm respectively. Spectra from adult lizards correspond to [Pérez i de Lanuza and Font \(2015\)](#).

764

| Colour patch | Age | N | Qt | C_{UV} | $\lambda \beta_{\max_{325-560}}$ (nm) | $\lambda R_{\text{mid}_{300-450}}$ (nm) | $\lambda R_{\text{mid}_{400-650}}$ (nm) |
|-------------------|----------------------|-----|---------------------|---------------|---------------------------------------|-----------------------------------------|-----------------------------------------|
| Conflated throats | Newborns (all) | 417 | 8303.9 \pm 133.0 | .12 \pm .00 | 345.1 \pm 4.3 | 345.16 \pm 2.0 | 513.4 \pm 2.4 |
| | Newborns (resampled) | 46 | 8384.1 \pm 357.6 | .13 \pm .01 | 329.8 \pm 0.1 | 334.5 \pm 2.7 | 520.0 \pm 5.5 |
| UV+white throat | Yearlings | 5 | 8303.9 \pm 133.0 | .12 \pm .03 | 328.8 \pm 1.6 | 329.4 \pm 5.4 | 536.2 \pm 27.5 |
| | Adults | 26 | 7306.0 \pm 598.5 | .13 \pm .01 | 330.2 \pm 0.2 | 336.8 \pm 5.1 | 563.3 \pm 10.2 |
| UV-white throat | Newborns (resampled) | 11 | 8091.9 \pm 909.3 | .08 \pm .01 | 388.2 \pm 13.0 | 386.1 \pm 4.1 | 483.1 \pm 11.9 |
| | Yearlings | 20 | 6142.5 \pm 547.3 | .05 \pm .01 | 402.0 \pm 3.4 | 399.6 \pm 5.3 | 511.6 \pm 14.8 |
| | Adults | 159 | 5544.1 \pm 173.5 | .04 \pm .00 | 410.9 \pm 2.4 | 407.5 \pm 1.9 | 505.9 \pm 5.3 |
| Yellow throat | Newborns (resampled) | 5 | 8241.2 \pm 2896.2 | .09 \pm .01 | 554.2 \pm 7.3 | 355.8 \pm 27.3 | 534.4 \pm 11.2 |
| | Yearlings | 26 | 5754.7 \pm 571.4 | .05 \pm .01 | 512.2 \pm 4.3 | 376.7 \pm 11.5 | 527.4 \pm 4.2 |
| | Adults | 150 | 4764.8 \pm 155.9 | .04 \pm .00 | 513.3 \pm 1.5 | 384.4 \pm 4.7 | 527.8 \pm 1.4 |
| Orange throat | Yearlings | 11 | 4848.9 \pm 1060.3 | .05 \pm .02 | 537.3 \pm 15.1 | 361.4 \pm 26.8 | 560.7 \pm 4.7 |
| | Adults | 125 | 3443.3 \pm 206.5 | .03 \pm .00 | 554.7 \pm 1.4 | 363.9 \pm 6.1 | 566.0 \pm 1.1 |
| OVS-UV females | Yearlings | 10 | 4534.9 \pm 997.2 | .18 \pm .05 | 329.5 \pm .5 | 327.1 \pm 11.0 | 544.5 \pm 30.4 |
| | Adults | 37 | 6294.7 \pm 420.6 | .28 \pm .02 | 330.2 \pm 0.2 | 342.0 \pm 12.8 | 498.2 \pm 24.9 |
| OVS-UV males | Yearlings | 26 | 4710.1 \pm 311.13 | .24 \pm .02 | 336.5 \pm 14.1 | 336.7 \pm 15.7 | 511.3 \pm 23.9 |
| | Adults | 510 | 6620.9 \pm 119.5 | .36 \pm .00 | 330.2 \pm .1 | 367.7 \pm 4.7 | 438.9 \pm 1.7 |

765

766

767 **Table 2.** Pairwise comparisons between the ventral coloration of lizards measured as newborns and re-sampled
768 as yearlings, per colour patch. Within-individual contrasts were calculated using a distance-based MANOVA
769 on the chromatic and achromatic distances obtained by modelling the vision of conspecifics for the ventral
770 coloration of each individual at birth and when re-captured one year after. Between-individual contrasts were
771 calculated using a distance-based PERMANOVA on the chromatic and achromatic distances between the
772 throat spectra of yearlings assigned into each of the four different morphs. Significant contrasts are indicated
773 in bold ($p < 0.05$). F and R^2 represent pseudo F -statistics and effect size estimate, respectively. Adj. P represents
774 adjusted P values (Bonferroni correction).
775

| Colour patch | | Within-individual contrasts | | | | | |
|-------------------------------|-----|-----------------------------|-------|-------------------|---------------------|-------|-------------------|
| | | Chromatic contrast | | | Achromatic contrast | | |
| Yearling colour | N | F | R^2 | P | F | R^2 | P |
| UV ⁺ white throats | 5 | 3.00 | 0.27 | 0.125 | 0.49 | 0.06 | 0.813 |
| UV ⁻ white throats | 20 | 34.07 | 0.47 | < 0.001 | 27.30 | 0.42 | < 0.001 |
| Yellow throats | 26 | 42.54 | 0.46 | < 0.001 | 20.92 | 0.29 | < 0.001 |
| Orange throats | 11 | 12.31 | 0.38 | 0.002 | 14.82 | 0.43 | < 0.001 |
| OVS females | 10 | 9.31 | 0.34 | 0.010 | 37.23 | 0.67 | 0.002 |
| OVS males | 26 | 71.96 | 0.59 | < 0.001 | 185.89 | 0.79 | < 0.001 |

| Colour patch | | Between-individual contrasts | | | | | |
|-----------------------|-----------------------|------------------------------|-------|--------------|---------------------|-------|--------------|
| | | Chromatic contrast | | | Achromatic contrast | | |
| | | F | R^2 | adj. P | F | R^2 | adj. P |
| UV ⁺ white | Orange | 3.36 | 0.19 | 0.039 | 2.88 | 0.17 | 0.091 |
| UV ⁻ white | Orange | 9.59 | 0.25 | 0.003 | 5.27 | 0.15 | 0.010 |
| UV ⁺ white | Yellow | 12.41 | 0.30 | 0.001 | 1.29 | 0.04 | 0.276 |
| UV ⁺ white | UV ⁻ white | 10.86 | 0.32 | 0.001 | 1.64 | 0.07 | 0.179 |
| Yellow | Orange | 9.44 | 0.21 | 0.003 | 4.31 | 0.11 | 0.016 |
| UV ⁻ white | Yellow | 4.17 | 0.09 | 0.020 | 0.32 | 0.01 | 0.734 |

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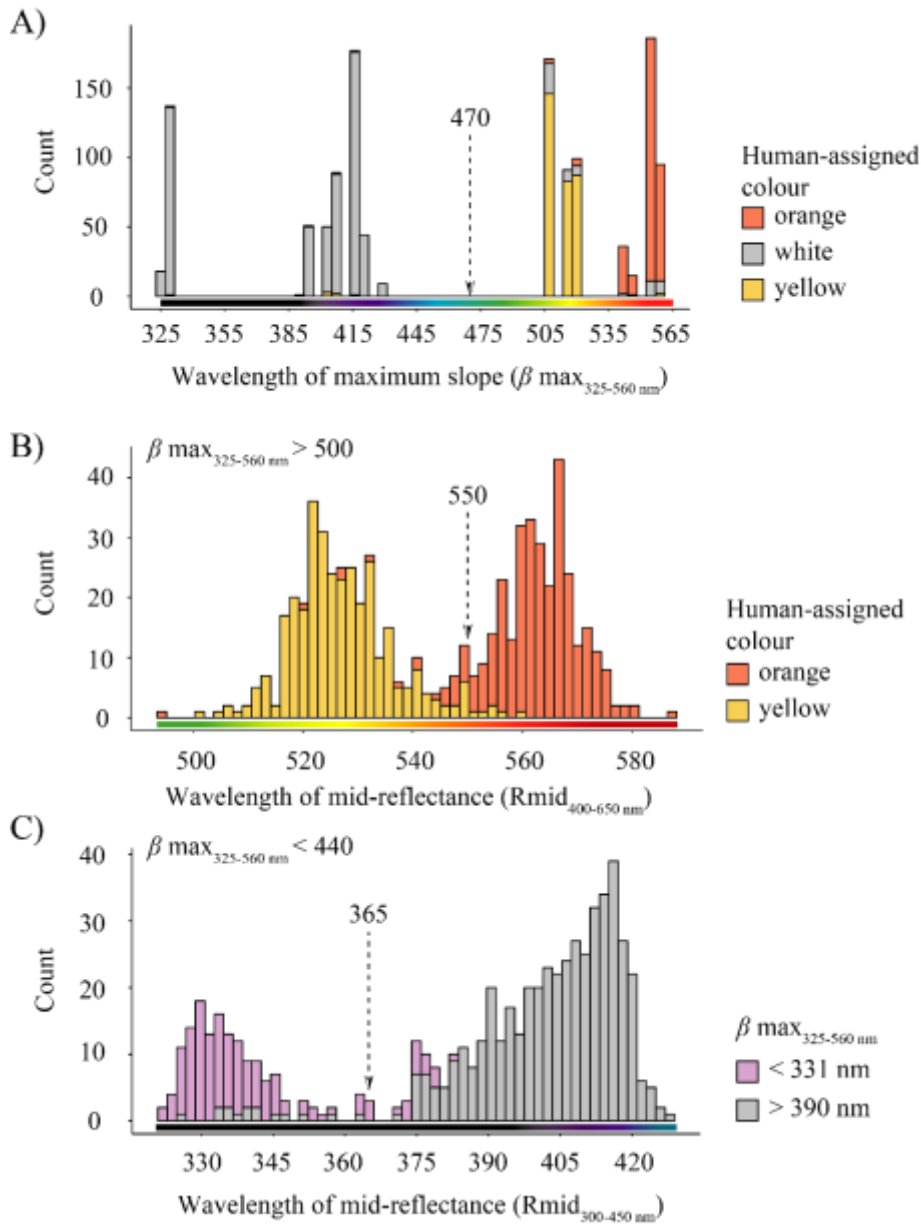
779 **Table 3.** Pairwise comparison contrasting the ventral coloration of adults with newborns, and adults with
780 yearlings, per colour patch. In the former, every adult colour was contrasted against the average newborn ventral
781 colour. In the latter, contrasts were estimated only between same-coloured yearling and adult lizards. Contrasts
782 were calculated using a distance-based PERMANOVA on the chromatic and achromatic distances obtained by
783 modelling the vision of conspecifics for the ventral coloration of newborn, yearling, and adult *P. muralis* lizards.
784 Significant contrasts ($p < 0.05$) are indicated in bold. *F* and *R*² represent pseudo *F*-statistics and effect size
785 estimate, respectively. Adj. *P* represents adjusted *P* values (Bonferroni correction).
786

| Colour patch | Newborn average - Adult colours | | | | | |
|-----------------|---------------------------------|-----------------------|---------------|---------------------|-----------------------|---------------|
| | Chromatic contrast | | | Achromatic contrast | | |
| | <i>F</i> | <i>R</i> ² | adj. <i>P</i> | <i>F</i> | <i>R</i> ² | adj. <i>P</i> |
| UV+white throat | -2.5 | -0.01 | 1.00 | 416.78 | 0.49 | 0.001 |
| UV-white throat | 705.5 | 0.55 | 0.001 | 2314.7 | 0.80 | 0.001 |
| Orange throat | 1406.8 | 0.72 | 0.001 | 1947.9 | 0.78 | 0.001 |
| Yellow throat | 1194.8 | 0.68 | 0.001 | 2270.9 | 0.80 | 0.001 |
| OVS males | 7252.7 | 0.89 | 0.001 | 5038.6 | 0.84 | 0.001 |
| OVS females | 556.2 | 0.56 | 0.001 | 623.85 | 0.58 | 0.001 |

| Colour patch | Yearlings - Adults (equivalent colours) | | | | | |
|-----------------|-----------------------------------------|-----------------------|---------------|---------------------|-----------------------|---------------|
| | Chromatic contrast | | | Achromatic contrast | | |
| | <i>F</i> | <i>R</i> ² | adj. <i>P</i> | <i>F</i> | <i>R</i> ² | adj. <i>P</i> |
| UV+white throat | 1.29 | 0.04 | 0.255 | 0.77 | 0.03 | 0.393 |
| UV-white throat | 2.91 | 0.02 | 0.069 | 3.66 | 0.02 | 0.033 |
| Yellow throat | 9.42 | 0.05 | 0.002 | 9.73 | 0.05 | 0.001 |
| Orange throat | 1.72 | 0.01 | 0.178 | 7.41 | 0.05 | 0.003 |
| OVS females | 33.52 | 0.43 | 0.001 | 1.73 | 0.04 | 0.189 |
| OVS males | 194.25 | 0.27 | 0.001 | 0.34 | 0.00 | 0.673 |

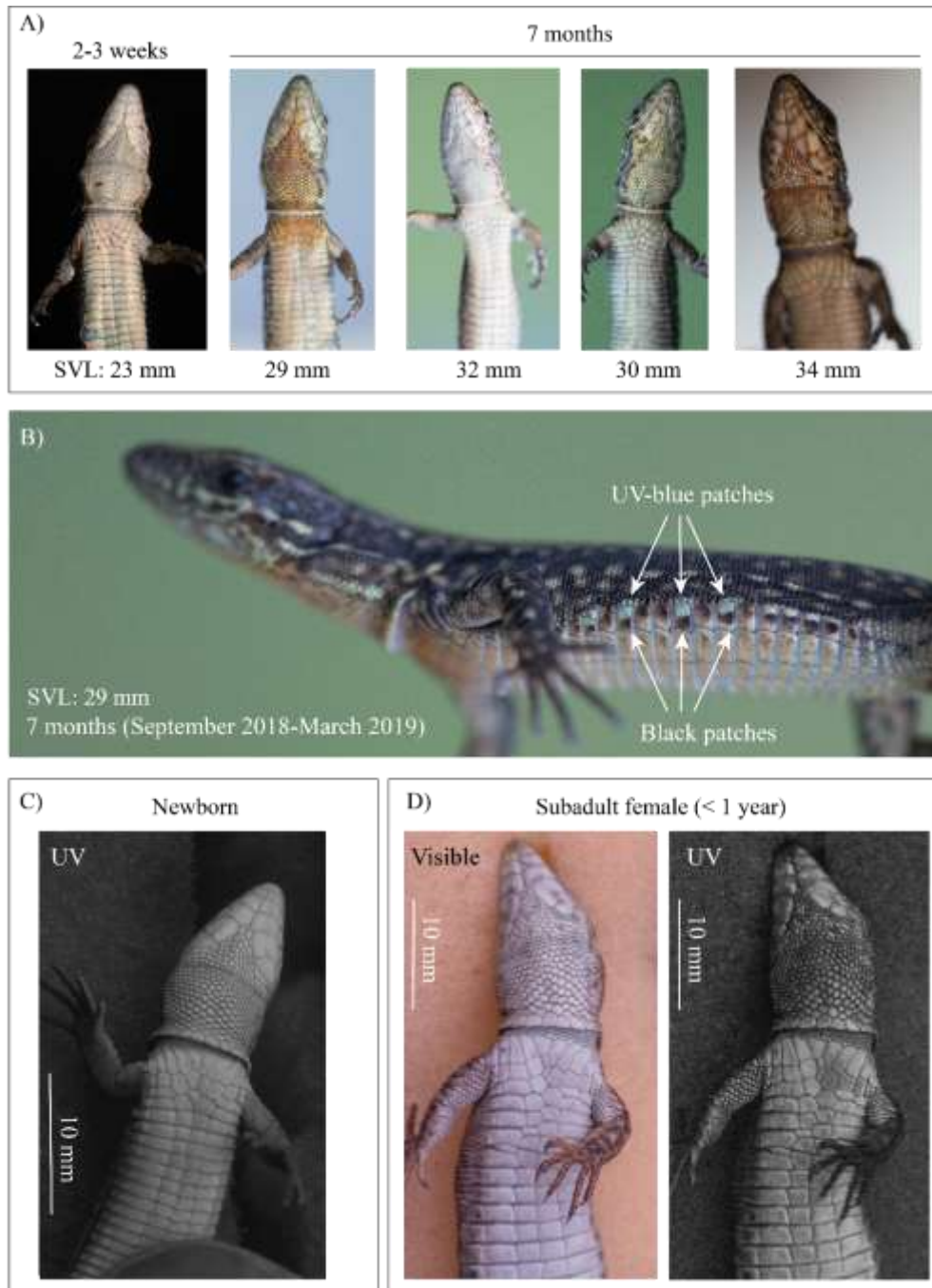
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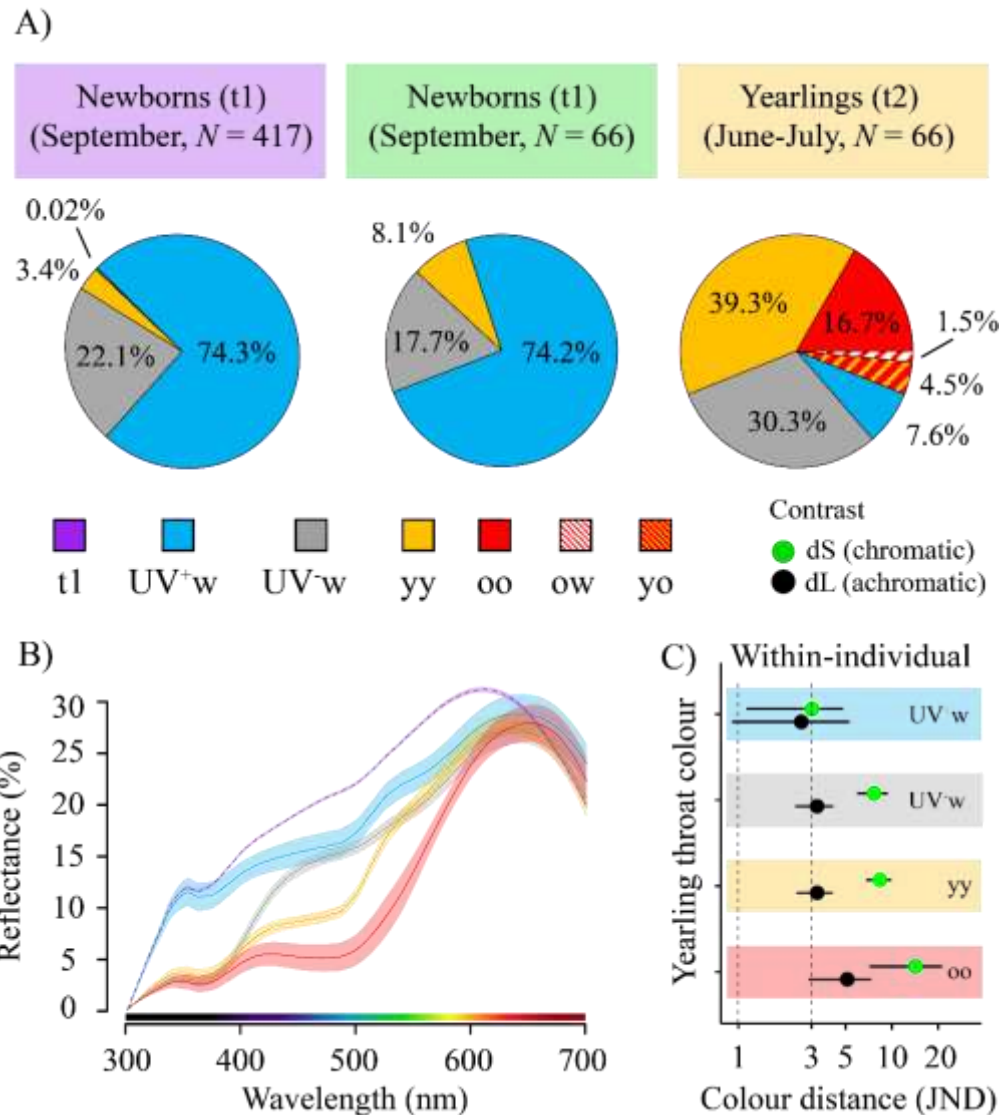
792

793 **Figure 1.** Categorization of *P. muralis* ventral colours according to their spectral properties in a sample of 1293
794 throat and belly spectra collected in adult males and females and assigned to a colour morph by human visual
795 inspection. A) Stacked bar plot showing the discontinuous distribution of $\lambda \beta \max_{325-560 \text{ nm}}$ (i.e. wavelength of
796 maximum slope between 325 and 560 nm). A gap between 440 and 500 nm separates most lizards assigned to the
797 orange and yellow colours from lizards assigned to the white morph. B) Stacked bar plot showing the
798 bimodal distribution of $\lambda \text{ Rmid}_{400-650 \text{ nm}}$ (i.e. wavelength at which reflectance is halfway between its minimum and
799 its maximum in the 400-650 nm waveband). We set a threshold at 550 nm to separate orange and yellow lizards,
800 which largely agrees with morph categorization by visual inspection. C) Stacked bar plot showing the bimodal
801 distribution of $\lambda \text{ Rmid}_{300-450 \text{ nm}}$. Filled bars show the high agreement between a threshold to distinguish between
802 UV⁺white and UV⁻white based on $\lambda \beta \max_{325-560 \text{ nm}}$ and one based on $\lambda \text{ Rmid}_{300-450 \text{ nm}}$. For consistency with Pérez i
803 de Lanuza and Font (2024, *in press*), we set a threshold at $\lambda \text{ Rmid}_{300-450 \text{ nm}} = 365 \text{ nm}$ to separate UV⁺white from
804 UV⁻white lizards.



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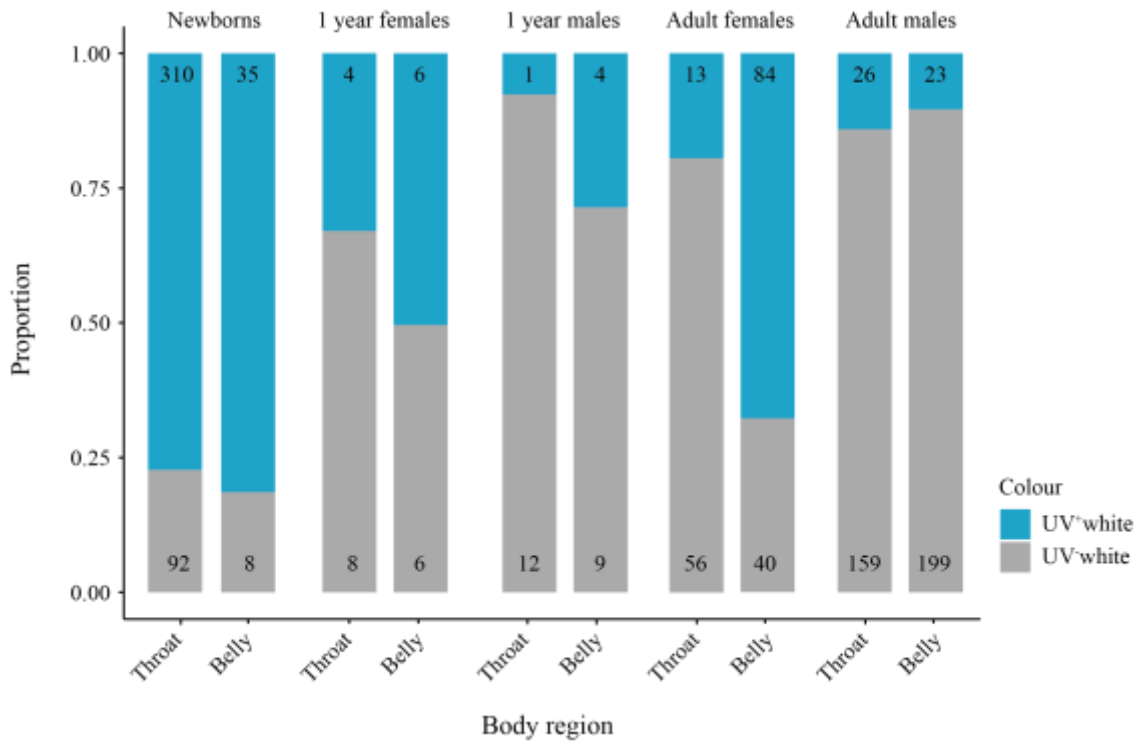
806 **Figure 2.** Ontogeny of ventral coloration in *P. muralis*. A) Representative photographs of ventral coloration in
 807 a newborn lizard 2-3 weeks after hatching (left), and in four yearlings re-captured in March 2019 (7 months
 808 after hatching). SVL stands for snout to vent length in mm. B) Young male of *P. muralis* showing incipient UV-
 809 blue and melanin-based black patches in the outer ventral scales (OVS). C) UV photograph of a newborn *P.*
 810 *muralis* lizard showing UV+white colour in its ventral surface. D) Paired UV and visible photographs of the
 811 same subadult female (<1 year) showing UV-white in the throat and UV+white in the belly. The difference is
 812 only apparent in the UV image. Photographs in C and D were obtained with a full-spectrum camera and two
 813 filters, each transmitting light either in the visible (400-700 nm) or the near-UV (320-380 nm) range. Brighter
 814 areas in the UV image have higher UV reflectance (i.e. newborn ventral surface and female belly).
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817 **Figure 3.** Ontogenetic changes in *P. muralis* throat coloration. A) Pie charts represent the proportions of
 818 juveniles showing different throat colours (UV⁺w = UV⁺white, UV⁻w= UV⁻white, oo = orange, yy = yellow,
 819 ow = orange-white, yo = yellow-orange) in September (t1; 3-4 weeks after hatching), and June-July (t2; one
 820 year after hatching). The three pie charts correspond to the full dataset of 417 newborns measured at t1 (purple),
 821 the subset of 66 surviving juveniles measured as newborns at t1 (green), and these same juveniles measured as
 822 yearlings in t2 (yellow). B) Reflectance spectra from the 62 surviving yearlings showing pure colour morphs
 823 (solid lines and shaded area represent means \pm SEM). The dashed purple line represents the average throat
 824 spectra of 417 newborn lizards (similar to that of the 62 surviving yearlings when first measured, see central pie
 825 chart). Spectra have been normalized by subtracting the minimum reflectance of each spectrum at all
 826 wavelengths. C) Mean and CI₉₅ of the chromatic and achromatic distances (dS and dL respectively) between
 827 paired measurements of throat reflectance (i.e. within-individual newborn-yearling contrasts), separated by
 828 yearling throat colour. JND stands for “Just Noticeable Differences”. Two dashed lines at 1 and 3 JNDs
 829 represent two theoretical discriminability thresholds. If the confidence interval of a point includes a
 830 discriminability threshold, the two colours are not distinguishable at this threshold according to receiver cone
 831 sensitivities and relative abundance.

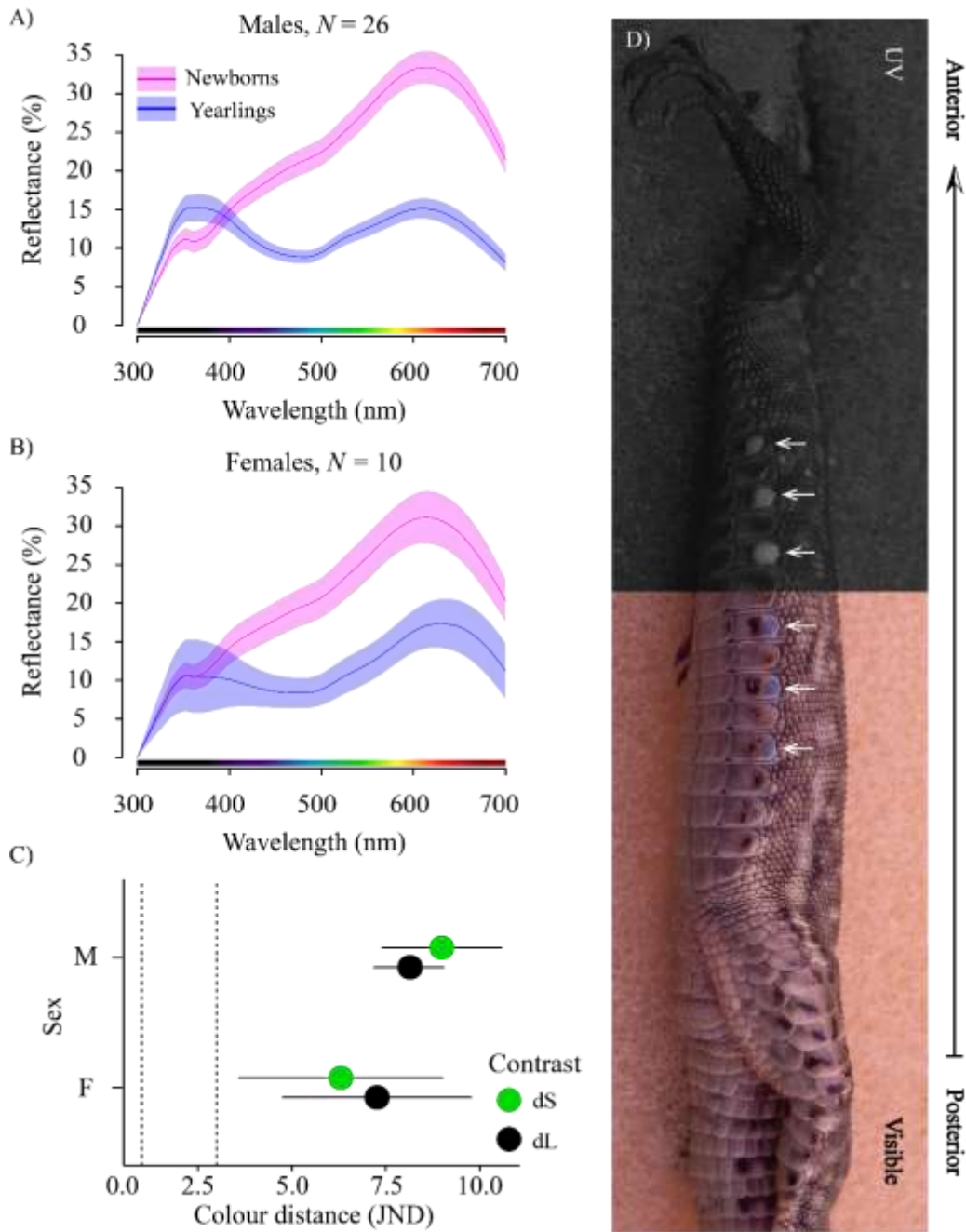
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835 **Figure 4.** Stacked bar plot showing the proportion of UV+white and UV-white colorations in the throats and
 836 bellies of lizards classified according to the spectral properties of their reflectance curves (i.e. $\lambda \beta_{\max_{325-560}} \leq$
 837 500): UV+white if showing a $\lambda R_{\text{mid}_{300-450}} \leq 365$ nm, UV-white if $\lambda R_{\text{mid}_{300-450}} > 365$ nm. Numbers inside filled
 838 bars indicate sample size. UV+white coloration is particularly frequent among newborn lizards, and in the belly
 839 of yearling and adult females.

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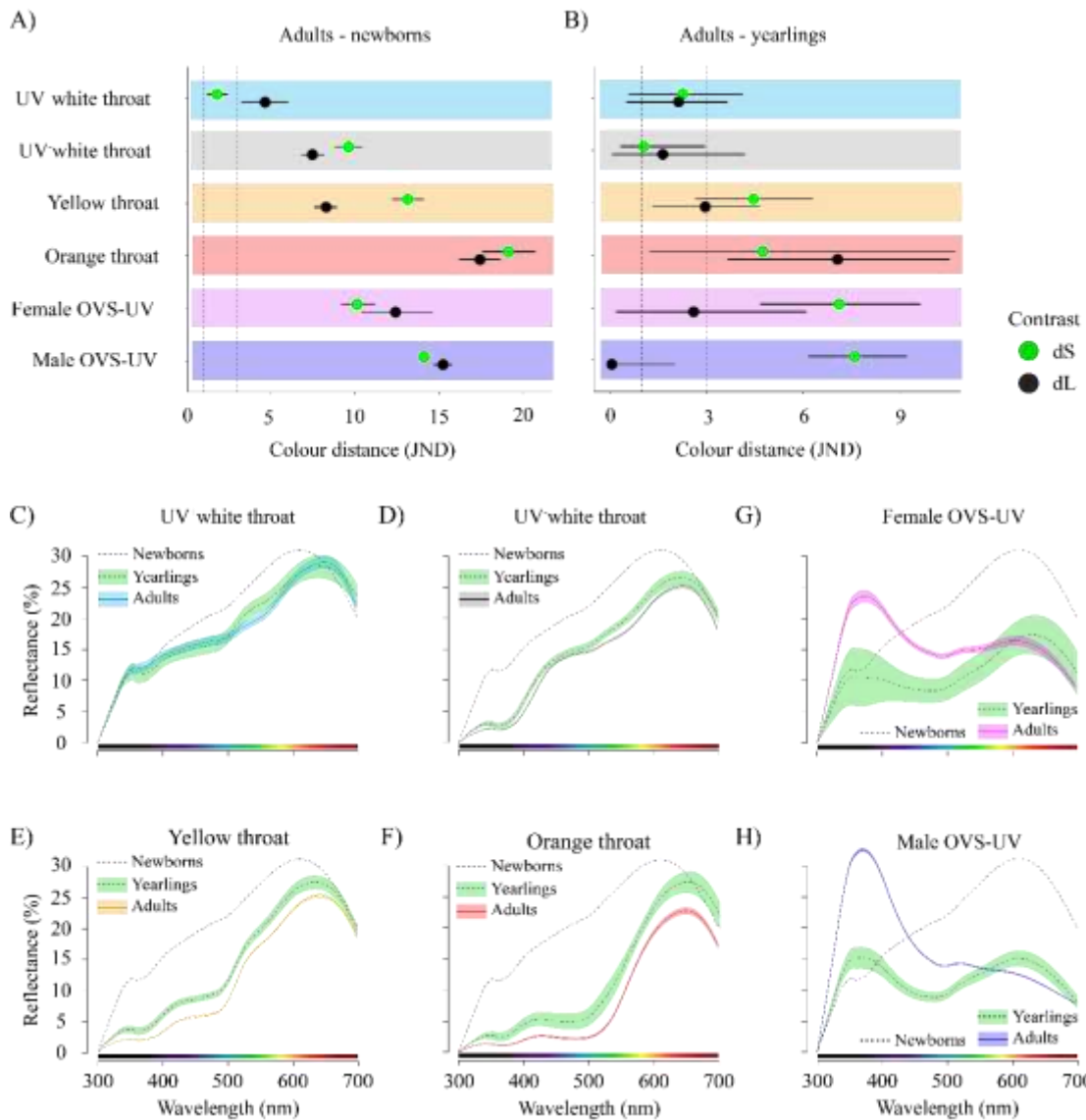
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Figure 5. Ontogenetic change in *P. muralis* outer ventral scales (OVS) coloration. A) Reflectance spectra from the 36 lizards showing UV-blue patches as yearlings in their OVS (solid lines and shaded area represent means \pm SEM). B) Mean and CI₉₅ of the chromatic (dS) and achromatic (dL) distances between paired measurements of OVS reflectance (newborns-yearlings) in males and females. Two dashed lines at 1 and 3 JNDs represent two theoretical discriminability thresholds. If the confidence interval of a point includes a discriminability threshold, the two colours are not distinguishable at this threshold according to receiver cone sensitivities and relative abundance. D) Composite UV + visible image of a subadult (<1 year) *P. muralis* male showing incipient UV-blue coloration in the OVS.



853

854 **Figure 6.** Comparison between the ventral coloration of newborn and yearling lizards with adult *P. muralis* from eastern
 855 Pyrenees. A) Mean and CI₉₅ of the chromatic (dS) and achromatic (dL) distances between the average ventral coloration of
 856 newborn and adult lizards, by adult colour. B) Mean and CI₉₅ of the chromatic and achromatic distances between equivalent
 857 colour patches in yearling and adult lizards (see Fig. S5 for all possible pairwise contrasts between throat colours). Two
 858 dashed lines at 1 and 3 JNDs represent two theoretical discriminability thresholds. If the confidence interval of a point
 859 includes a discriminability threshold, the two colours are not distinguishable at this threshold according to receiver cone
 860 sensitivities and relative abundance. C-F) Throat reflectance spectra from yearlings and adults of each colour morph. G-H)
 861 Reflectance spectra of the UV-blue patches in the OVS of yearling and adult lizards, separated by sex. Solid lines and shaded
 862 area represent mean \pm SEM. Dashed black line represents the average ventral reflectance of newborn lizards.

863

864

865 **Appendix S1: Supplementary material of “Cryptic ontogenetic changes in the**
 866 **ventral coloration of a colour polymorphic wall lizard (*Podarcis muralis*)”**

867 **Tables**

868

869 **Table S1.** Spectral variables (mean \pm CI₉₅) describing reflectance curves in the dorsal coloration of 8 newborn,
 870 66 yearlings, and 50 adult lizards of each sex. Spectra have been normalized by subtracting the minimum
 871 reflectance of each spectrum at all wavelengths. *N* = sample size, *Qt* = luminance (*R*₃₀₀₋₇₀₀), λ max = wavelength
 872 corresponding to peak reflectance in the 300-700 nm waveband, Chroma = Reflectance over the λ max \pm 50
 873 nm range, divided by total luminance (*Qt*).

| Colour patch | Age | <i>N</i> | <i>Qt</i> | λ max (nm) | Chroma |
|--------------|---------------|----------|--------------------|--------------------|-----------------|
| Dorsum | Newborns | 8 | 1315.7 \pm 155.8 | 640.3 \pm 9.5 | 0.53 \pm 0.03 |
| | Yearlings | 66 | 1011.7 \pm 74.6 | 642.0 \pm 4.2 | 0.55 \pm 0.01 |
| | Adult males | 50 | 1562.5 \pm 183.0 | 631.5 \pm 4.7 | 0.50 \pm 0.01 |
| | Adult females | 50 | 1237.4 \pm 132.1 | 642.8 \pm 5.1 | 0.50 \pm 0.01 |

874

875 **Table S2.** Pairwise comparisons contrasting the throat colours of adult and yearling *P. muralis* lizards. Contrasts
876 were calculated using a distance-based PERMANOVA on the chromatic and achromatic distances obtained by
877 modelling the vision of conspecifics for the ventral coloration of yearling and adult lizards. Significant contrasts
878 ($p < 0.05$) are indicated in bold. F and R^2 represent pseudo F -statistics and effect size estimate, respectively.
879 Adj. P represents adjusted P values (Bonferroni correction).

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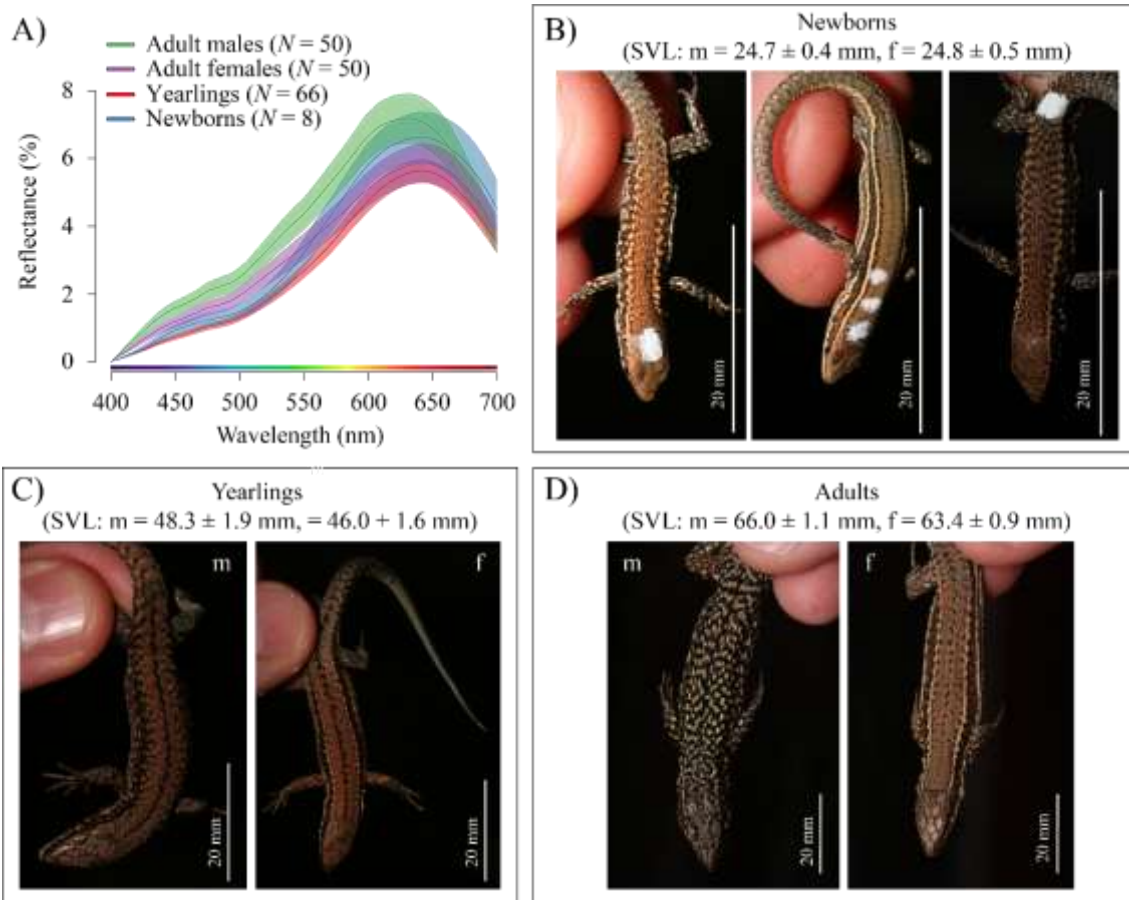
| Yearling colour | Adult colour | Chromatic contrast dS | | | Achromatic contrast dL | | |
|-----------------|--------------|-----------------------|-------|--------------|------------------------|-------|--------------|
| | | F | R^2 | adj. P | F | R^2 | adj. P |
| UV+white | UV+white | 1.29 | 0.04 | 0.255 | 0.77 | 0.03 | 0.393 |
| | UV-white | 14.98 | 0.08 | 0.001 | 5.16 | 0.03 | 0.010 |
| | Yellow | 15.58 | 0.09 | 0.001 | 8.28 | 0.05 | 0.002 |
| | Orange | 8.28 | 0.06 | 0.005 | 9.71 | 0.07 | 0.003 |
| UV-white | UV+white | 77.92 | 0.64 | 0.001 | 0.54 | 0.01 | 0.548 |
| | UV-white | 2.91 | 0.02 | 0.066 | 3.66 | 0.02 | 0.033 |
| | Yellow | 19.86 | 0.11 | 0.001 | 8.52 | 0.05 | 0.001 |
| | Orange | 32.76 | 0.19 | 0.001 | 32.65 | 0.19 | 0.001 |
| Yellow | UV+white | 99.66 | 0.67 | 0.001 | 0.40 | 0.01 | 0.651 |
| | UV-white | 14.50 | 0.07 | 0.001 | 4.17 | 0.02 | 0.028 |
| | Yellow | 9.42 | 0.05 | 0.002 | 9.73 | 0.05 | 0.001 |
| | Orange | 33.09 | 0.18 | 0.001 | 38.01 | 0.20 | 0.001 |
| Orange | UV+white | 19.19 | 0.35 | 0.001 | 6.07 | 0.15 | 0.006 |
| | UV-white | 30.60 | 0.15 | 0.001 | 3.94 | 0.02 | 0.028 |
| | Yellow | 9.64 | 0.06 | 0.001 | 3.61 | 0.02 | 0.042 |
| | Orange | 1.72 | 0.01 | 0.178 | 7.41 | 0.05 | 0.003 |

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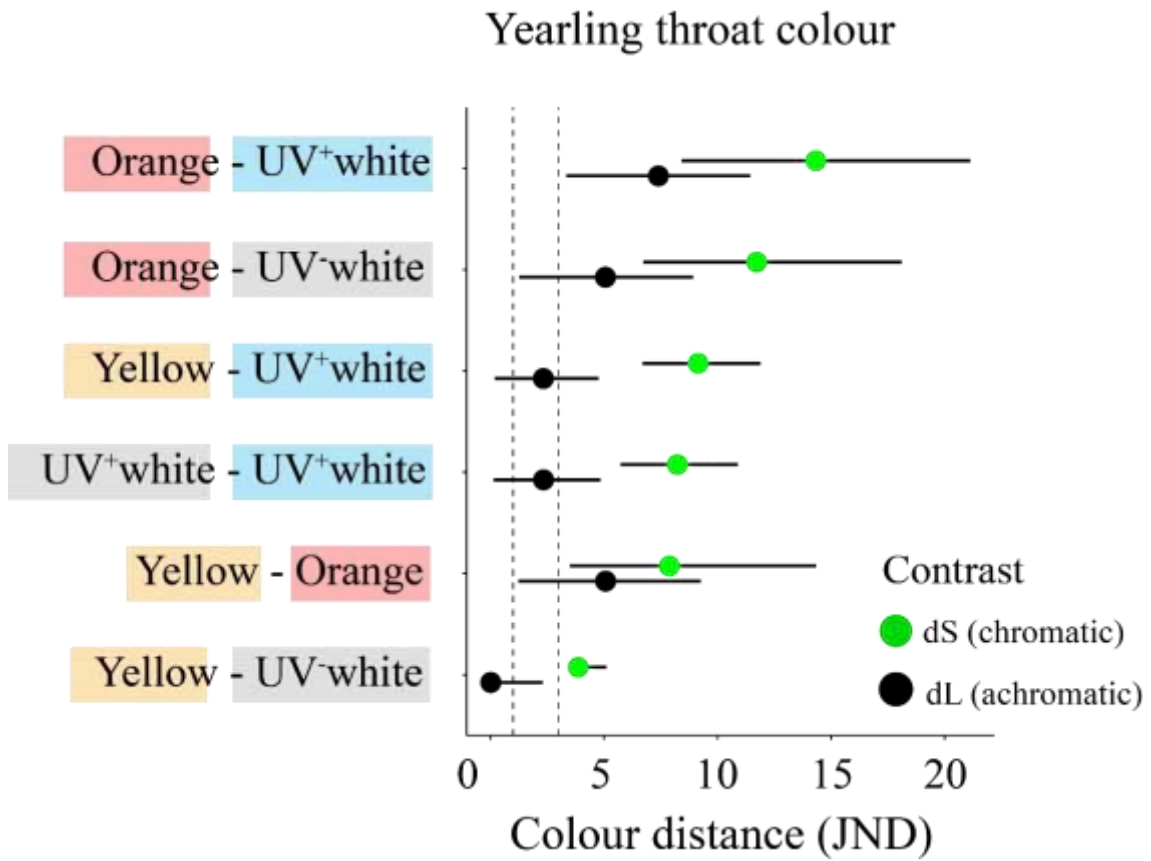
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885 **Figure S1.** Breeding and housing of wall lizard juveniles. A) Newborn lizard hatching from the egg. Clutches
886 were incubated in plastic cups filled with moist coco husk (1:2 coco:water by weight) and covered with a
887 perforated lid at a constant temperature of 28°C. B) Outdoor plastic tubs (170cm diameter, 60cm high) used to
888 house juveniles under natural conditions at the Station d'Ecologie Théorique et Expérimentale (SETE, Moulis,
889 France). We covered each tank with a net, which excluded avian predators while allowing local invertebrates to
890 get inside the tanks and serve as primary food source. To increase habitat complexity, we complemented the
891 naturally occurring vegetation inside the tanks by providing a perforated brick to serve as shelter, two wood
892 logs, and a water dish to collect rainwater. We maintained the tanks by regularly trimming weeds and
893 supplementing the lizards' diet with *Tenebrio molitor* larvae and *Acheta domestica* adults. Unfortunately, the
894 COVID-19 lockdown caused a two-month interruption in tank maintenance, enabling some juveniles to escape
895 through the overgrown vegetation. More details in [Abalos et al., \(2021\)](#).



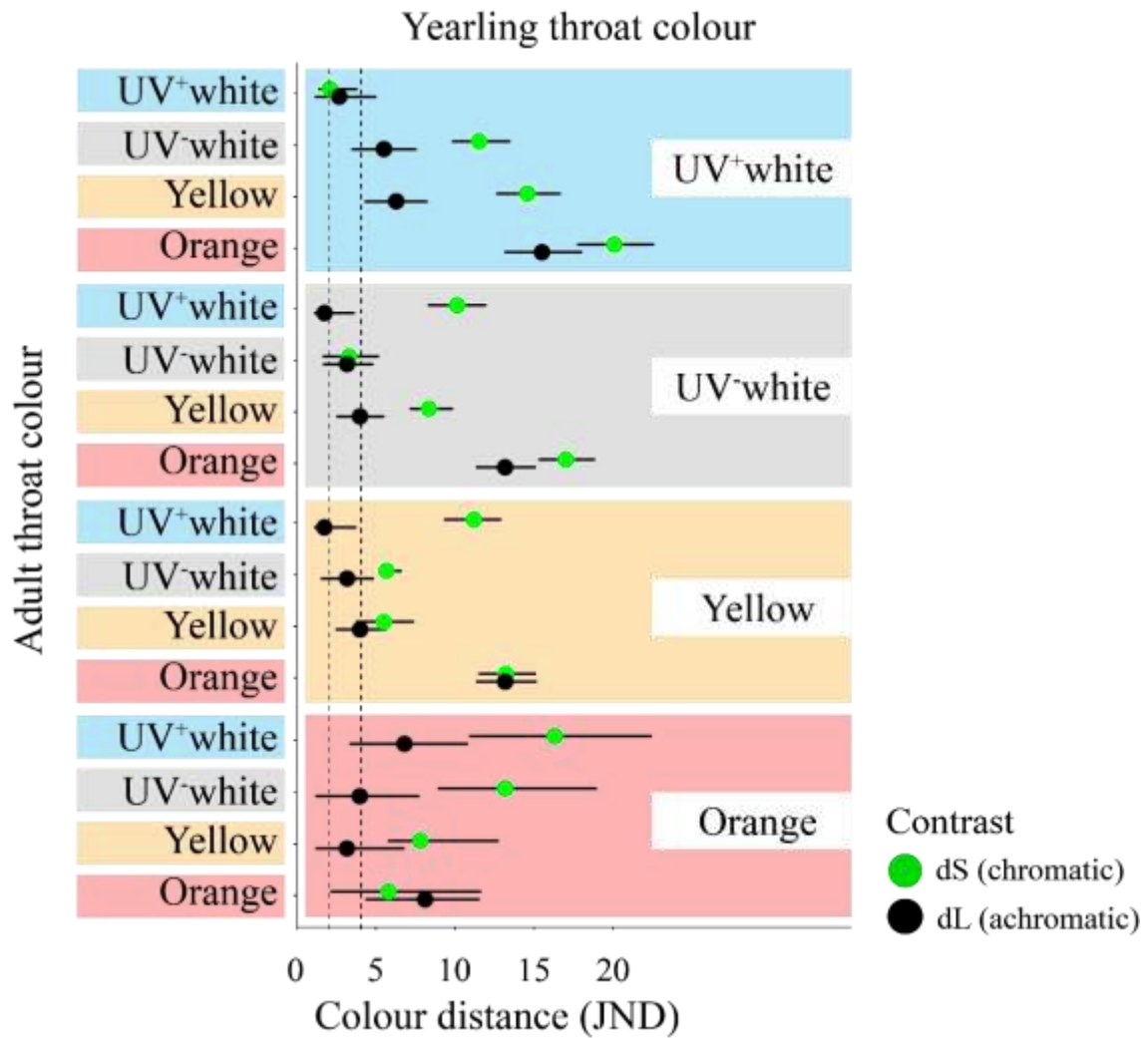
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898 **Figure S2.** Ontogenetic variation in the dorsal coloration of *Podarcis muralis*. A) Reflectance spectra measured
 899 in eight newborn lizards, 66 yearlings, and 50 adults of each sex (solid lines and shaded area represent means \pm
 900 CI_{95}). Spectra correspond to dorsal skin not covered by melanin patches and have been normalized by
 901 subtracting the minimum reflectance of each spectrum at all wavelengths. B) Photographs of three
 902 representative newborn lizards obtained 3-4 weeks after hatching. C) Photographs of two representative
 903 yearling lizards (one male and one female) obtained 10 months after hatching. D) Photographs of a male and
 904 female *P. muralis* captured in the same localities where the parent generation of the juveniles in B and C where
 905 sampled (i.e. along the Cerdanya plateau in Pyrénées-Orientales, France).



907

908 **Figure S3.** Mean and CI₉₅ of the chromatic and achromatic distances between the four different colours found
 909 in the throat of yearling lizards. Two dashed lines at 1 and 3 JNDs represent two theoretical discriminability
 910 thresholds. If the confidence interval of a point includes a discriminability threshold, the two colours are not
 911 distinguishable at this threshold according to receiver cone sensitivities and relative abundance.



912

913 **Figure S4.** Mean and CI_{95} of the chromatic and achromatic distances between the colours found in the throat
 914 of yearling and adult lizards. Two dashed lines at 1 and 3 JNDs represent two theoretical discriminability
 915 thresholds. If the confidence interval of a point includes a discriminability threshold, the two colours are not
 916 distinguishable at this threshold according to receiver cone sensitivities and relative abundance.

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918 **References**

- 919 Abalos, J., i de Lanuza, G.P., Bartolomé, A., Aubret, F., Uller, T. & Font, E. (2021). Viability, behavior, and
920 colour expression in the offspring of matings between common wall lizard *Podarcis muralis* colour
921 morphs. *Curr. Zool.* 1–15.
- 922 Abalos, J., Pérez i de Lanuza, G., Bartolomé, A., Liehrmann, O., Laakkonen, H., Aubret, F., Uller, T., Carazo,
923 P., Font, E., Lanuza, G.P. i de, Bartolomé, A., Liehrmann, O., Laakkonen, H., Aubret, F., Uller, T.,
924 Carazo, P., Font, E., Pérez i de Lanuza, G., Bartolomé, A., Liehrmann, O., Laakkonen, H., Aubret, F.,
925 Uller, T., Carazo, P. & Font, E. (2020). No evidence for differential sociosexual behavior and space use
926 in the color morphs of the European common wall lizard (*Podarcis muralis*). *Ecol. Evol.* **00**, 1–20.
- 927
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