

# The Development and Evolution of Arthropod Tagmata

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## **Abstract**

The segmented body plan is a hallmark of the arthropod body plan. Morphological segments are formed during embryogenesis, through a complex procedure involving the activation of a series of gene regulatory networks. The segments of the arthropod body are organized into functional units known as tagmata, and these tagmata are different among the arthropod classes (e.g. head, thorax and abdomen in insects). Based on embryological work on segment generation in a number of arthropod species, coupled with a survey of classical descriptions of arthropod development, I suggest a new framework for the evolution of arthropod tagmata. The ancestral condition involves three developmental tagmata: The pre-gnathal segments, a tagma that is formed within a pre-existing developmental field and a tagma that is formed through the activity of a segment-addition zone, that may be embryonic or post-embryonic. These embryonic tagmata may fuse post-embryonically to generate more complex tagmata. This framework is consistent with the evolution of tagmosis seen in the early arthropod fossil record. It also calls for a re-thinking of the decades-old division of arthropod development into short-germ vs. long-germ development, and a re-thinking of questions of segment identity determination, and the role of Hox genes in tagma differentiation.

Keywords: Arthropods, tagma, segment-identity, Hox genes, body-plan

26

27 **Overview**

28 The arthropod body plan is characterized by repeated morphological units, or segments,  
29 along the antero-posterior axis. These segments are usually organized into higher level units,  
30 each composed of several segments, known as tagmata (tagma in singular). The segments of  
31 each tagma normally have shared functions, and roughly similar size and shape, and they are  
32 often differentiated from the segments of the adjoining tagmata by a distinct morphological  
33 boundary. While the nature and composition of the tagmata vary among arthropod classes, they  
34 are more or less conserved at the level of the class, and are often used as defining characters of  
35 the class. The morphological literature has given a lot of attention to the defining features of  
36 different tagmata, and to the variability in their composition, but there has been almost no recent  
37 discussion of the evolutionary history of the tagmata or of their development. I present a  
38 synthesis of what is known about tagmosis (the arrangement of segments into tagmata) from  
39 comparative morphology, from developmental biology and from the fossil record. I use this  
40 synthesis to suggest a novel model for the evolution of arthropod tagmata.

41 The details of arthropod tagmosis have been extensively reviewed by Fusco and Minelli  
42 [1], and will not be repeated here. I will give a brief overview of the typical tagmosis pattern in  
43 the main arthropod lineages, emphasizing the most likely plesiomorphic condition for each  
44 lineage.

45 *Chelicerata*

46 The basic chelicerate body plan is composed of two tagmata, usually known as the  
47 prosoma (or cephalothorax) and opisthosoma (or abdomen). This is most clearly seen in the  
48 terrestrial arachnids, exemplified by spiders. The prosoma includes four pairs of walking  
49 appendages and anterior to them two pairs of feeding or sensory appendages, the pedipalps and  
50 the chelicerae, The anteriormost segment of the prosoma carries the eyes and the labrum. The  
51 opisthosoma normally does not carry walking appendages, but may carry breathing appendages,  
52 or specialized structures such as spinnerets.

53 There are numerous minor variations on this theme, and these have been reviewed  
54 extensively by Dunlop and Lamsdell [2]. In pycnogonids, the number of walking appendages can

55 vary, and the opisthosoma is rudimentary or nonexistent [3]. Some arachnids have a subdivided  
56 opisthosoma (e.g. scorpions), whereas in others there is no clear border between the prosoma and  
57 opisthosoma (e.g. acarids, opiliones). Nonetheless, it is generally accepted that the two-tagma  
58 structure is ancestral for extant chelicerates, as are four walking appendages and two sensory /  
59 feeding appendages in the prosoma [2].

#### 60 *Myriapoda*

61 The myriapod body plan consists of a head and a mostly homonomous trunk. The head is  
62 composed of six segments: three pre-gnathal segments and three gnathal segments. This is the  
63 composition of the head in all members of the Mandibulata (Myriapoda + Pancrustacea). The  
64 pre-gnathal segments include the three parts of the brain and carry eyes on the first segment and  
65 a pair of antennae on the third segment. The gnathal segments carry feeding appendages [4].

66 The trunk is composed of all of the remaining segments, most of which carry a pair of  
67 walking appendages. The myriapod trunk may display a number of lineage specific novelties,  
68 such as a mid-body transition zone in many centipedes, or a mismatch between dorsal and  
69 ventral segmentation in millipedes. There may be specialized segments, such as the centipede  
70 venom-claw or the posteriormost segment, or gonopods used for sperm transfer in millipedes. In  
71 some cases, there is some degree of regionalization within the trunk [4]. Nonetheless, none of  
72 these specializations alter the basic tagmosis of the myriapod body plan, which is always  
73 composed of a head and a trunk.

#### 74 *Pancrustacea*

75 The non-hexapod pancrustaceans display a bewildering array of tagmosis patterns [5]. In  
76 all cases, the anteriormost tagma is a six-segment mandibulate head, although the head is not  
77 always separated by a clear morphological boundary from the tagma behind it. Indeed, the head  
78 is often covered by a head shield or carapace that also covers segments posterior to the head. The  
79 region covered by the carapace is sometime known as a cephalothorax, although this is a very  
80 different structure from the one known by the same name in chelicerates.

81 The body of malacostracan crustaceans is usually divided into two regions, known as a  
82 pereon and pleon. The pereon carries walking appendages, as well as maxillipedes, which are  
83 modified appendages used in feeding. The pleon usually does not have walking appendages, but

84 often has other modified appendages. In brachiurans, as well as in other “crab-like” crustaceans,  
85 most or all of the pleon is reduced and folded ventrally under the pereon [5, 6].

### 86 *Hexapoda*

87 Insects and their close relatives in Hexapoda have the most consistent pattern of tagmosis  
88 among arthropods. Their body is composed of three sharply defined tagmata: a six-segment  
89 mandibulate head, a three-segment thorax with walking appendages, and an abdomen with  
90 between 9 and 11 segments or as few as 6 in Collembola, that normally do not carry appendages  
91 [7].

92 There are minor variations on this structure. E.g., in some hymenopterans, the border  
93 between the thorax and the abdomen shifts. The head and thorax are sometimes fused, as in some  
94 beetles. In some holometabolous insects there is a reduction in the number of adult abdominal  
95 segments. Nonetheless, there is little doubt that the ancestral pattern of tagmosis is the common  
96 three-tagma structure.

97

### 98 **The fossil record**

99 The earliest arthropod stem group members, the paraphyletic assemblage known as  
100 lobopodians, show no tagmosis, and are thus similar to the extant arthropod sister groups,  
101 onychophorans and tardigrades [8]. The head is composed of a single segment, with no  
102 additional segments attached to it [9]. While there may be some regionalization and  
103 specialization of specific segments along the trunk, these are not grouped into functional regions  
104 and do not display morphological borders, and should thus not be considered as tagmata.

105 The gilled lobopodians (such as *Kerygmachela* and *Pambdelurion*), as well as diverse  
106 organisms as *Opabinia* and the radiodonts, also have a single-segment head, and no obvious  
107 tagmata. The single head carries the eyes and usually a single pair of large raptorial or sensory  
108 appendages [10, 11]. Members of these group display varying degrees of regionalization. The  
109 size of the segments and appendages varies along the body axis, but without a sharp  
110 discontinuity. The neck region in radiodonts is a distinct group of 4-5 segments, but it is not  
111 delimited by a morphological boundary. The tail fan is a distinct structure in some gilled  
112 lobopodians and radiodonts, and it may be composed of a number of segments [12, 13]. All this

113 considered, while members of the gilled lobopodian / radiodont grade start to show increasing  
114 levels of regionalization, there are no obviously differentiated tagmata.

115 Deuteropoda is the clade that includes both crown-group arthropods and the upper stem  
116 group. Members of Deuteropoda have three pre-gnathal segments [14]. In the upper stem group,  
117 these three segments comprise the entire head (but see O'Flynn, Liu [15] for a different  
118 viewpoint). These head segments often bear specialized appendages (sometimes called “great  
119 appendages”), while in some cases, there are 1-3 specialized appendages just posterior to the  
120 head [16-19]. It is possible to describe these animals as having a distinct head and trunk, as in  
121 myriapods, although the head is composed of only three segments. It is not clear whether the  
122 specialized post-cephalic appendages should be considered part of the head or of the trunk, as  
123 there is no sharp morphological boundary. These appendages may be the precursors of the  
124 gnathal segments in mandibulate arthropods.

125 Tagmosis in trilobites and its development has been studied fairly extensively, due to the  
126 excellent fossil record of these animals, which allows the reconstruction of complete post-  
127 embryonic developmental series [20, 21]. The trilobite body is divided into three tagmata: the  
128 cephalon (or head), thorax and pygidium. The posterior border of the cephalon is clear and  
129 stable, and is established in the earliest recoverable stages, which have a head only, or a head  
130 with a single additional segment. However, the thorax and pygidium are dynamic, with new  
131 segments being added to the pygidium in successive molts, and then “released” to the thorax, to  
132 maintain a more-or-less stable number of pygidial segments, and a growing number of thoracic  
133 segments [22]. The thorax-pygidium border is thus a dynamic border, and the tagmata  
134 themselves vary in number of segments and in the border between them throughout ontogeny. In  
135 this sense, they are unlike the tagmata in all extant arthropods and in most fossil arthropods.

136 Stem-group chelicerates all have a distinction between a prosoma and an opisthosoma, but  
137 the number of segments in each of these tagmata and the identity of the appendages carried on  
138 these segments does not always conform to what is found in extant chelicerates. The diversity  
139 and evolution of chelicerate tagmata has been reviewed in detail by Dunlop and Lamsdell [2].  
140 Stem-group mandibulates have a typical head tagma, and variable posterior tagmata. Their  
141 diversity has been reviewed by Waloszek and Maas [23]. A more recent alternative framework  
142 for the mandibulate stem group is outlined by Izquierdo-López and Caron [24].

143            **Developmental tagmosis**

144            The basis for tagma differentiation is laid down during embryonic development. While this  
145 statement sounds intuitively almost obvious, the embryonic basis of tagmosis has hardly been  
146 studied, and has only been looked at explicitly in a handful of species. There is however enough  
147 implicit information in both classical and modern descriptions of embryonic development to be  
148 able to draw some general conclusions.

149            The connection between regionalization of embryonic segments and adult tagmata is not  
150 direct, and there is often a mismatch between the two. The most obvious example is in the pre-  
151 gnathal segments (PGS). There is an ongoing debate about the evolutionary origin of these units  
152 [25, 26], with Lev and Chipman [27] arguing that they should not even be considered segments.  
153 However, regardless of these differing opinions, there is no doubt that the three anterior segments  
154 in all arthropods develop differently and stand on their own as a distinct embryonic unit, despite  
155 being incorporated into the adult head (in mandibulates) or prosoma (in chelicerates).

156            In mandibulates, the three posterior segments of the head, the gnathal segments, are  
157 indistinguishable from the segments immediately posterior to them during early stages of  
158 development. In most insects, the gnathal and thoracic segments (abbreviated as gnatho-thoracic  
159 segments hereafter) form a distinct series of six segments that form together and are of similar  
160 size and shape, until the differentiation of the appendages (mouthparts or walking legs) [28-31].  
161 In myriapods, the gnathal segments are identical to trunk segments in the early germband before  
162 appendage differentiation [32, 33]. In chelicerates, prosomal segments and opisthosomal  
163 segments have different morphologies [34-36], although this is probably mostly due to the  
164 presence of walking limbs on prosomal segments, which affect the segments' morphology.

165            In many cases, there are also differences in the way different segments are formed. In what  
166 follows, I review what is known about these differences in species where all segments are  
167 formed during embryogenesis (epimorphic development). Species where some of the segments  
168 are formed post-embryonically (anamorphic development) are discussed in the following section.

169            *Insects*

170            The similarity between gnathal and thoracic segments is not limited to their morphology in  
171 the germband stage. In many cases, they also develop through a similar developmental program.

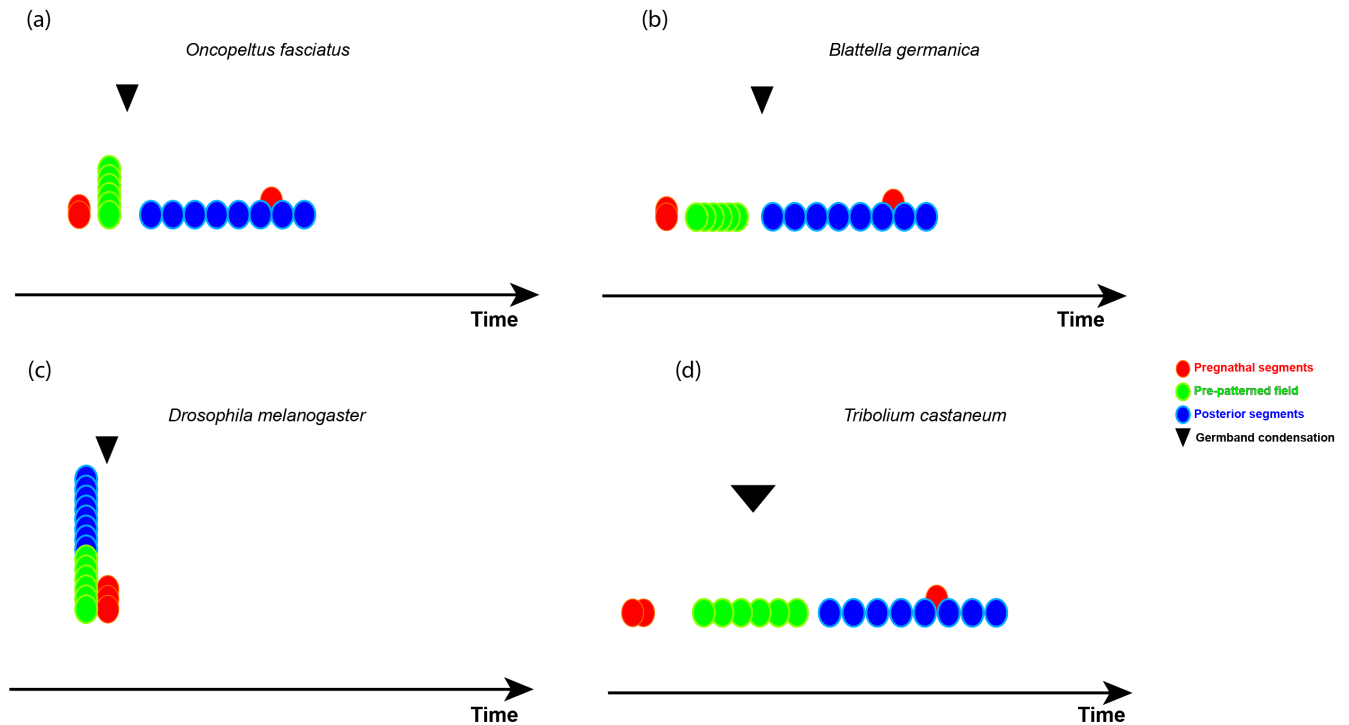
172 This similarity has been best demonstrated in the milkweed bug *Oncopeltus fasciatus* (Fig. 1a).  
173 In this species (and in other hemipterans), early development can be divided into a blastoderm  
174 stage and a germband stage [37-39]. In the blastoderm stage, the embryo is composed of a single  
175 layer of cells covering an ovoid yolk mass. The cells of the blastoderm then undergo a process of  
176 invagination, to form a germband that is embedded inside the yolk. The segmentation process  
177 begins already in the blastoderm stage, with orthologs of the *Drosophila* segmentation cascade  
178 genes being expressed in sequence to form segmental stripes of segment polarity genes such as  
179 *engrailed* and *wingless* [40]. During this early phase of segmentation, the gnatho-thoracic  
180 segments are patterned at the molecular level, and the borders between them are established by  
181 the interactions of the segment-polarity network. There is no morphological evidence of  
182 segmental borders during the blastoderm stage.

183 During the invagination process, the embryonic tissues condense to give the germband, and  
184 shortly afterwards, the gnatho-thoracic segments can be seen morphologically [41]. About half-  
185 way through the invagination process, a specialized embryonic zone forms at the posterior of the  
186 germband, and it is from this posterior zone, known as the segment addition zone (SAZ) or  
187 growth-zone, that the abdominal segments start appearing sequentially [42].

188 The question arises whether this distinction between gnatho-thoracic and abdominal  
189 segmentation is unique to hemipterans, or whether a similar distinction can be seen in the  
190 development of other insects. An analysis of an additional species of hemimetabolous insects,  
191 together with a survey of reported segmentation patterns in other insects, suggests that this may  
192 be a general pattern – at least in hemimetabolous insects – although the details may vary. In the  
193 German cockroach *Blattella germanica* (Fig. 1b), the gnathal and thoracic segments are  
194 patterned sequentially at the level of segment-polarity genes, but the pair rule gene *even-skipped*  
195 is expressed more or less simultaneously in all gnatho-thoracic segments, and fades sequentially,  
196 with *hedgehog* expression coming up in its place. Only after all of the gnatho-thoracic *hedgehog*  
197 expression stripes come up, does the SAZ form and abdominal segments appear sequentially,  
198 driven by a cyclic expression of *even-skipped* [31].

199 In the cricket *Gryllus bimaculatus* gnatho-thoracic stripes of *hedgehog* expression come up  
200 rapidly and sequentially. There is then a gap of a few hours before the SAZ forms and abdominal  
201 segments arise sequentially [43]. This pattern is similar to that seen in *B. germanica*.

202 A closer analysis of the pattern in these three hemimetabolous insects shows that the main  
 203 difference between the formation of gnatho-thoracic and abdominal segments is not whether they  
 204 are formed simultaneously or sequentially. The main difference is in the embryonic environment  
 205 in which they are formed. Gnatho-thoracic segments are patterned within a pre-existing  
 206 embryonic field or anlage, whereas abdominal segments are formed from a posterior domain  
 207 where axial elongation is taking place – the SAZ.



208

209

210 Figure 1: Schematic representation of the different modes of segment generation in four insect  
 211 species. (a) In the milkweed bug *Oncopeltus fasciatus*, two of the pre-gnathal segments (red) and the  
 212 gnatho-thoracic segments (green) are patterned nearly simultaneously in the blastoderm. Germband  
 213 condensation (marked with a black arrowhead) occurs through a process of invagination. The abdominal  
 214 segments (blue) are patterned sequentially in the germband. The third pre-gnathal segment, the intercalary  
 215 segment, is patterned during abdominal segmentation, as in many insects. (b) In the German cockroach  
 216 *Blattella germanica*, there is no sharp distinction between a blastoderm and germband stage. However,  
 217 the gnatho-thoracic segments are patterned rapidly and sequentially in a pre-patterned field, which then  
 218 condenses to form the germ-band, where abdominal segmentation takes place. (c) In the well-studied  
 219 fruit-fly, *Drosophila melanogaster*, all segments are patterned simultaneously in the blastoderm, followed



220 rapidly by the condensation of the germ-band. (d) In the flour beetle *Tribolium castaneum*, all segments  
221 are patterned sequentially. However, there is a difference in rate between the gnatho-thoracic segments  
222 and the abdominal segments, and the segment-addition zone is only active during abdominal  
223 segmentation. Germ-band condensation occurs simultaneously with gnatho-thoracic segmentation  
224 (marked with a broad black arrowhead).

225

226

227 Detailed descriptions of segmentation in additional hemimetabolous insects are rare, but a  
228 survey of the literature suggests a similar pattern in many cases (see Supplementary table 1 for  
229 examples). Anterior segments form within a pre-existing field, without addition of new tissue,  
230 and posterior segments form from a SAZ. The border between the two processes is not always  
231 evident from the existing description, but the data are consistent with the border being the  
232 thoracic-abdominal border. Even in short-germ insects such as the grasshopper *Schistocerca*, the  
233 early embryonic field encompasses only the gnathal segments and expands through tissue  
234 condensation to include the thoracic segments [28]. Only after this process is done are abdominal  
235 segments added sequentially.

236 Since hemimetabolous insects form a paraphyletic group, basal to the more widely studied  
237 Holometabola, we can assume that the pattern described above is the ancestral segmentation  
238 mode. This pattern was thus presumably lost in Holometabola, otherwise it would have been  
239 discovered before, given the extensive body of work on holometabolan development (e.g.  
240 *Drosophila melanogaster*, Fig. 1c). But is this really the case? In the well-studied red flour beetle  
241 *Tribolium castaneum* (Fig. 1d), a study of the dynamics of segment generation showed that the  
242 thoracic segments form at a uniform rate, there is then a slowing down of the process, followed  
243 by a dramatic increase in segmentation rate during abdominal segmentation [44]. Indeed, looking  
244 at the germ-band of *T. castaneum*, the SAZ is only evident during abdominal segmentation, with  
245 gnatho-thoracic segments being patterned in a rapidly condensing – but not extending –  
246 embryonic field.

247 In the parasitic jewel wasp *Nasonia vitripennis*, where there is no obvious SAZ, there is a  
248 transition in segment generation between the six anterior segments (the gnatho-thoracic  
249 segments) and the posterior abdominal segments [45]. The first three pair-rule stripes

250 (corresponding to the six anterior segments) arise simultaneously and the following stripes arise  
251 in what has been called “progressive segmentation” – e.g. sequential segmentation without a  
252 SAZ [46].

253 Even in *Drosophila melanogaster*, the paradigm for simultaneous, long-germ segmentation,  
254 there are two patterning centers: an anterior patterning center responsible for gnathal and  
255 thoracic segments, and a posterior center responsible for abdominal segments [47-49]. Even  
256 when all obvious evidence of a difference in segmentation mode between segments has been lost,  
257 there is a vestige hinting at an ancestral distinction between two groups of segments.

### 258 *Crustaceans*

259 The diverse morphologies and tagmosis modes seen in crustaceans are reflected in diverse  
260 modes of segmentation. There is not a lot of information about embryonic segmentation in  
261 different crustaceans. Most crustaceans for which we have any information display indirect  
262 development, hatching as a larva with only three visible segments – two pairs of antennal  
263 segments and the mandibular segment. This type of larva is known as a nauplius [50], and is  
264 discussed in the next section on post-embryonic segmentation.

265 Species with direct development are found throughout crustacean diversity. In many cases,  
266 even when all segments are formed during embryogenesis, there is an early developmental stage  
267 where the three naupliar segments develop and start differentiating, before the appearance of any  
268 other segments. This stage is called the “egg nauplius”, and is usually understood to be  
269 homologous to the recently hatched nauplius of indirect developing species [51, 52]. An example  
270 of such development can be seen in the branchiopod *Daphnia magna* [53, 54]. In this species, the  
271 naupliar segments appear very early in development, followed by all posterior segments.  
272 However, the posterior segments do not follow a strict anterior-posterior sequence. The two  
273 maxillary segments, which form part of the head, appear at a slight delay relative to the thoracic  
274 segments, suggesting the possibility of a somewhat different mechanism underlying their  
275 formation, relative to the sequential, SAZ-based formation of thoracic segments.

276 The crustacean group in which embryonic development has been best studied is  
277 Malacostraca. Most malacostracan crustaceans form their segments through posterior stem cells  
278 known as ectoteloblasts [55, 56]. In this mode of segmentation, every division of the  
279 ectoteloblasts generates a daughter cell that will be the precursor to cells of a single parasegment.

280 Segmentation via ectoteloblasts has been studied in the isopod *Porcellio scaber* (among others)  
281 [57-59]. The anterior segments, including the pregnathal segments and the gnathal segments, are  
282 formed through cell rearrangements prior to the activity of the ectoteloblasts. All segments from  
283 the first thoracic segment onwards are formed sequentially via divisions of the ectoteloblast row.  
284 It is worth noting that in post-hatching morphology, the cephalothorax, which is the anteriormost  
285 tagma, includes the pregnathal and gnathal segments, in addition to the first thoracic segment.

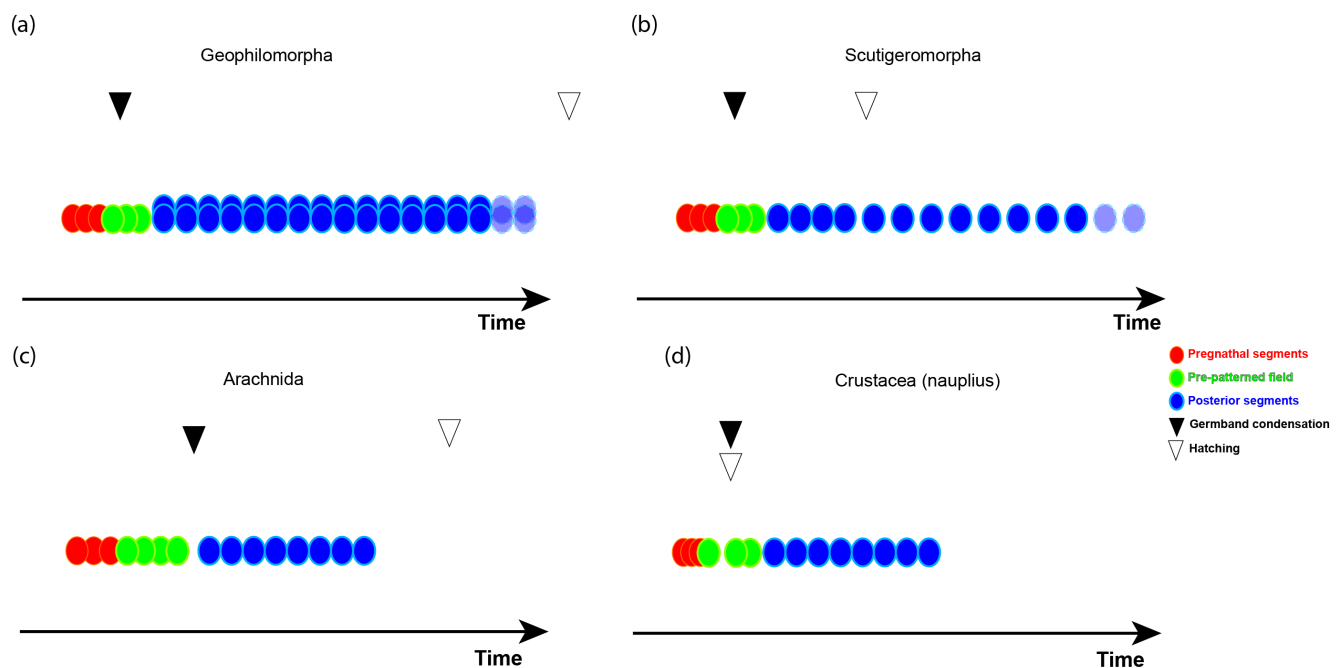
286 Studying the neural development of two crayfish species (members of Malacostraca),  
287 Vilpoux, Sandeman [60] showed that the central nervous system in the naupliar region (pre-  
288 gnathal + mandibular neuromeres) develops almost simultaneously. After its formation there is a  
289 short lag, after which the posterior neuromeres develop sequentially.

290 The best studied model for crustacean embryonic segmentation is the amphipod *Parhyale*  
291 *hawaiensis*. However, similar to *D. melanogaster*, and in common with all amphipods, *P.*  
292 *hawaiensis* displays a simultaneous mode of segment generation, which is almost certainly  
293 derived [61, 62]. In all amphipods, ectodermal cells assemble into a grid, without going through  
294 the sequential ectoteloblast divisions typical of other malacostracans. The pregnathal and  
295 mandibular segments (the naupliar segments) display a slightly different mode of assembly,  
296 whereas there is no noticeable difference among the way all other segments are formed.

### 297 *Myriapods*

298 In the centipede *Strigamia maritima* (Fig. 2a), a “head bulge” appears from the germ disc  
299 before segmentation begins, and the pre-gnathal segments form within the bulge [33]. The  
300 gnathal segments, the segment bearing the venom-claw and the first leg-bearing segment appear  
301 nearly simultaneously [32, 33, 63]. All posterior segments form sequentially, initially with a two-  
302 segment periodicity of a segmentation clock [64]. There is no apparent difference in the  
303 segmentation of the gnathal and trunk segments. Towards the end of the segmentation process,  
304 there is a shift from a pattern where two segments are patterned from each stripe of *even-skipped*  
305 expression to a pattern, to one where each *even-skipped* stripe generates a single segment [65].

306 The only diplopod species where segmentation has been studied is the pill millipede  
 307 *Glomeris marginata* [66]. Segmentation in this species is not very different from that of the  
 308 centipede *S. maritima*. Segmentation is sequential throughout, with the first few segments  
 309 appearing more or less simultaneously, with no obvious difference between gnathal segments and  
 310 trunk segments. Typically for diplopods, there is a mismatch between dorsal and ventral  
 311 segmental patterning, with each dorsal tergite corresponding to two ventral leg-bearing segments.  
 312 This mismatch is only seen in the trunk segments, and not in the head segments, where dorsal  
 313 and ventral segments are aligned. The anterior-most trunk segment also shows no dorsal-ventral  
 314 mismatch [66].



315  
 316 Figure 2: Schematic representation of the different modes of segmentation in four arthropod  
 317 species. (a) In geophilomorph centipedes (such as *Strigamia maritima*), the germ-band condenses (black  
 318 arrowhead) during the segmentation of the pre-gnathal segments (red) and the gnathal segments (green),  
 319 without the activity of a segment addition zone. Trunk segments (blue) are formed mostly two at a time  
 320 (transparent fill and dotted lines indicate that not all segments are portrayed). Segmentation ends well  
 321 before hatching (white arrowhead). (b) In scutigermomorph centipedes, anterior segmentation is probably  
 322 similar to that of geophilomorph centipedes, although there is very little data). Four trunk segments form  
 323 during embryogenesis, with successive segments added post hatching. (c) In arachnids, including spiders  
 324 and others, the pregnathal segments and the limb-bearing segments are patterned rapidly within the early

325 embryonic disk. The disk then condenses to give the germband, and opisthosomal segments are patterned  
326 sequentially from a segment-addition zone. (d) in crustaceans that have a nauplius stage, the pre-gnathal  
327 segments and the mandibular segments are patterned embryonically. The germ-band condenses and the  
328 nauplius hatches, with additional segments added sequentially. In crustaceans without a nauplius stage  
329 (not shown) the main difference is that hatching is heterochronically shifted to a after the end of  
330 segmentation.

331

332

333 *Chelicerates*

334 In spider segmentation, the prosomal segments are patterned rapidly within the initial  
335 germ-disc, without any extension of the disc (Fig. 2c). After prosomal segments are formed, the  
336 SAZ begins to function, generating single segments sequentially through a cyclic process [35,  
337 67-69]. This pattern is seen both in the wandering spider *Cupiennius salei* and in the house  
338 spider *Parasteatoda tepidariorum*.

339 A similar pattern is seen in other arachnids where segmentation has been studied, although  
340 the details are usually not as clear. Both in the whip scorpion *Phrynus marginemacultus* [70] and  
341 in the harvestman *Phalangium opilio* [71], the prosomal segments are formed rapidly within the  
342 germ disc, while the opisthosomal segments are formed sequentially from a segment addition  
343 zone.

344 **Post-embryonic segmentation**

345 There are two main modes of post-embryonic (or anamorphic) segmentation found within  
346 arthropods. The first is addition of segments within a nauplius / post-nauplius larva (Fig. 2d). In  
347 this mode, segment addition takes place in a free-swimming larva that hatches with a minimal  
348 number of segments. Segment addition is not normally accompanied by molting. Although there  
349 are molts during the segment addition process, they are few relative to the number of segments  
350 added. The process occurs continuously and at a relatively constant rate [72]. This mode of post-  
351 embryonic segmentation is found in many crustacean taxa, and may be ancestral for  
352 Pancrustacea.

353           The second mode is the addition of segments in sequential molts, in an animal that hatches  
354 with functional walking limbs. This is the case in a number of myriapod taxa, and it is also found  
355 to a limited extent (addition of a single post-embryonic segment) in some mites and in oniscidian  
356 isopods.

357           Post-naupliar segment addition has been studied in only a handful of crustacean species  
358 [72-74]. In all cases, the two antennal segments and the mandibular segment form rapidly in  
359 embryogenesis. The development of the ocular segment, which lies anterior to the antennal  
360 segments, is usually not described explicitly, but it is probably determined early in larval  
361 development. The post-mandibular segments arise sequentially, and there is no apparent  
362 difference in their mode or dynamics of development. In the fairy shrimp *Thamnocephalus*  
363 *platyurus*, the thoracic and abdominal segments are different in shape and size, but arise  
364 similarly [72].

365           Centipedes of the orders Scutigermorpha (Fig. 2b) and Lithobiomorpha hatch from the  
366 egg with four or six to eight leg bearing segments, respectively. Additional segments are added  
367 one or two at a time in successive molts, up to a total of 15 leg bearing segments [4]. Most  
368 millipedes also hatch with seven leg-bearing segments and add segments in successive molts, in  
369 some cases indefinitely [4].

370           Development of the pycnogonids (sea spiders) is very diverse, but the most common mode  
371 of development involves the hatching of a pronymphon larva that includes the pre-gnathal  
372 segments and an additional one or two segments, similar to the situation in crustaceans with a  
373 nauplius. Additional segments are added in successive molts [75].

374           The arthropod fossil record indicates that post-embryonic segment addition was much  
375 more phylogenetically diverse in the past. A number of fossil taxa have a rich enough fossil  
376 record that allows reconstruction of post-embryonic developmental series. This has already been  
377 discussed above for trilobites, and it is important to point out here that the earliest developmental  
378 stage found for trilobites is a head larva, with no externally obvious segmentation. It is not clear  
379 how many segments these larvae contain, but based on exceptionally preserved fossils of adult  
380 trilobite heads, they probably contain five segments, presumably including the pre-gnathal  
381 segments and two additional cephalic appendages [76].

382 “Orsten” type fossilization is an exceptional mode of preservation wherein small  
383 individuals are fossilized in exquisite detail through phosphatic replacement of organic tissues.  
384 Late Cambrian Orsten fossils provide a wealth of information about larval development in  
385 crustaceans and their relatives. These fossils indicate that the nauplius is a very early invention  
386 within the crustacean lineage, and that sequential post-naupliar segment addition was established  
387 and common by the late Cambrian [77, 78].

388 Developmental series for a number of stem and putative crown group arthropods have been  
389 published in the past decade. These include the radiodont *Stanleycaris* [79, 80], the upper stem  
390 species *Isoxys* [81] and *Chandianella* [82], and the putative crown arthropods *Leancoilia* [83,  
391 84] and *Fuxianhuia* [85]. All of these cases indicate that post-embryonic segment addition was  
392 found throughout arthropod phylogeny in the early stages of their evolution. However, none of  
393 these developmental series are as complete as that of trilobites, so we don’t know the minimal  
394 number of segments with which these animals hatched from the egg.

#### 395 **A model for the evolution of arthropod tagmosis**

396 The many different patterns of tagmosis found in extant arthropods emerged from an  
397 unknown ancestral pattern. I suggest that ancestral tagmosis was originally a developmental  
398 phenomenon, with the segments of diverse regions of the animal patterned differently. Over  
399 evolutionary time, the segments that were patterned using different developmental mechanisms  
400 evolved differential morphologies and different functional roles. The original differences in  
401 development are preserved to varying extents in extant arthropods. In many cases, the borders  
402 between regions that are patterned differently are preserved as tagma borders. In some cases, the  
403 mode of development has changed so the developmental borders are not immediately obvious. In  
404 many cases, additional tagmata evolved as sub-divisions of the original ones. In other cases,  
405 there has been fusion within and between adult tagmata, masking the original borders, although  
406 these can still be seen in development.

407 In the common ancestor of all arthropods, segments were patterned via three mechanisms:  
408 1) The anterior three segments (the pre-gnathal segments) were patterned separately and not  
409 always in sequence with other segments, via a developmental mechanism that did not involve  
410 pair-rule gene homologs, and probably including a mechanism of “stripe-splitting”. 2) A number  
411 of trunk segments, probably numbering on the order of 5-10 segments, were patterned within a

412 pre-existing embryonic field, either by subdivision of the field (simultaneous segmentation) or  
413 through a sequential process that did not involve the addition of new embryonic tissue  
414 (progressive segmentation). 3) Posterior segments were patterned sequentially, through the  
415 activity of a segment addition zone, which involved axis extension via a combination of the  
416 generation of new tissue through cell division and of convergent extension movements of the  
417 SAZ. Vestiges of these three segmentation mechanisms can be found in almost all extant  
418 arthropods.

419 I raise the intriguing possibility that originally, the distinction between the second  
420 segmentation mechanism (segmentation within a pre-existing developmental field) and the third  
421 mechanism (segmentation from a SAZ), was a distinction between embryonic and post-  
422 embryonic segmentation. In Chipman [86], I made the implicit assumption that the common  
423 ancestor of all arthropods was a direct developer forming all segments during embryogenesis,  
424 and that indirect development evolved convergently in different lineages. This was based mostly  
425 on lack of evidence for indirect development in the fossil record. Subsequently, Wolfe [87]  
426 argued that metamorphosis was ancestral, based on phylogenetic considerations. With the new  
427 data available, I now suggest a revised and intermediate solution. The last arthropod common  
428 ancestor was hemianamorphic, i.e. some segments were formed in embryogenesis and some  
429 post-embryonically. The anamorphically forming segments were generated via a SAZ in a  
430 gradual process, with new segments probably appearing during the molting process. Indeed, the  
431 SAZ may have evolved as a developmental mechanism for post-embryonic segment generation.  
432 Since the closest sister groups to arthropods (onychophorans and tardigrades) both generate all  
433 segments during embryonic development, we cannot say for certain when post-embryonic  
434 segmentation, and by extension the SAZ, evolved. However, the fact that we find post-  
435 embryonic segment addition in a radiodont provides a possible phylogenetic bracketing [80].

436 The extent of the segments formed in a pre-existing field (for brevity, these segments will  
437 be referred to as PEF segments hereafter) varies in different arthropods. This field still exists as a  
438 clear and distinct field in arachnids and in insects. In insects it comprises 6 segments (gnathal +  
439 thoracic). In arachnids it comprises 4 segments (walking limbs). In crustaceans with a nauplius  
440 larva it may have been lost, with only one post-PGS segment (the mandibular segment) patterned  
441 not via the SAZ. Nonetheless, in some crustaceans, the first few post-naupliar segments appear  
442 simultaneously, which may be a vestige of this mechanism. In myriapods, the pre-existing field



443 is not as distinct, but the dynamics of segmentation suggest that the SAZ only starts generating  
444 segments from first or second trunk segment, indicating that at least the embryonic gnathal  
445 segments are within this field.

446 Most of the diversity of developmental tagmosis modes in different arthropods can be seen  
447 as stemming from variation in three parameters: 1) The number of segments patterned under  
448 each of the mechanisms, 2) The point in development at which the germband forms and  
449 gastrulation takes place, 3) the stage of development at which the embryo hatches. The latter two  
450 are based on heterochronic shifts, which can occur fairly rapidly and easily from an evolutionary  
451 point of view. The former tends to be conserved at high taxonomic levels, although departures  
452 from the taxon-typical mode can be found. The diversity of arthropods is such that there are  
453 many examples of unusual development and tagmosis. The strangeness of these examples should  
454 not distract us from identifying the deeply rooted ancestral pattern.

#### 455 **Implications of the model**

##### 456 *Short germ vs. long germ development*

457 For nearly a century, the literature on insect and arthropod development had made the  
458 distinction between short-germ development and long-germ development [88-90]. The difference  
459 lies in the extent of the embryo that is patterned before gastrulation, or germ-band condensation.  
460 In long-germ development, the entire embryonic axis is patterned prior to gastrulation and in  
461 short-germ development, only a small part of the axis is (usually, only the head). Intermediate-  
462 germ is used for cases where the head + thorax are patterned prior to gastrulation. Short and  
463 long-germ are often also used to make a distinction between simultaneous and sequential  
464 segmentation, although this was not the original meaning of the terms.

465 The proposed model for the evolution and development of tagmata requires a reframing of  
466 long and short-germ development in the context of the different embryonic tagmata. Two  
467 separate modes of segmentation exist in almost all arthropods, except for in the most extreme  
468 cases of long-germ, simultaneous segmentation such as *Drosophila*. The main difference  
469 between the modes is in the heterochronic shift of gastrulation and germ-band condensation  
470 relative to the segmentation process.

##### 471 *The mandibulate head*

472 The head of mandibulate arthropods (myriapods, crustaceans and insects) incorporates  
473 segments from two embryonic tagmata: the PGS and the PEF segments [91, 92]. The fossil  
474 record shows that the ancestral head in crown group arthropods was composed of only the PGS  
475 [26]. The evolution of the mandibulate head initially involved the fusion of the entire PEF tagma  
476 with the PGS to give rise to the six-segment head. This is the case in extant myriapods and may  
477 be the ancestral mode in crustaceans (but see discussion of head larvae below).

#### 478 *The insect thorax*

479 Insects are the largest class within arthropods and the most conservative in terms of their  
480 overall body-plan (despite being extremely variable in modifications of this body plan). Perhaps  
481 one of the sources of the insects' success is the tightly integrated thorax, which specializes in  
482 locomotory function, and has no direct equivalent in any other arthropods class. The uniqueness  
483 of the insect thorax extends to its developmental origin. There is no other case where the PEF  
484 tagma subdivides into two functional adult tagmata. In myriapods and in non-hexapod  
485 crustaceans, the PEF is small – probably only three segments – and is entirely incorporated into  
486 the mandibulate head. In insects, this tagma encompasses six segments, half of which are  
487 incorporated into the head and half of which form the thorax. This realization puts the evolution  
488 of the insect body plan in a new light and suggests additional avenues for evo-devo research,  
489 while also predicting potential transitional body plans that might be found in the fossil record.

#### 490 *The issue of head larvae*

491 The term “head larva” is often used to describe larval forms that comprise only 3-4  
492 segments, essentially the pre-gnathal segments and possibly an additional gnathal segment. The  
493 best-known head larvae are the nauplius larvae common in many crustaceans (and possibly  
494 plesiomorphic for Pancrustacea), but head larvae are also found in some pycnogonids and in  
495 horseshoe crabs. Because of their highly derived post-embryonic development mode, it is  
496 difficult to incorporate these forms into the current model. However, looking at the cases where  
497 there is an embryonic nauplius (egg-nauplius) helps clarify the situation. As mentioned above,  
498 many crustaceans exhibit a slightly different mode of segmentation in the anterior few  
499 gnathal/trunk segments, suggesting that there is an anterior tagma that is distinct from the SAZ-  
500 driven sequentially segmenting posterior tagma. The evolution of head larvae involved an  
501 extreme heterochronic shift in the time of hatching relative to segment formation. The

502 postponement of segmentation to the post-embryonic period obscures the different segmentation  
503 modes, and they may even be lost in some lineages.

#### 504 *Homology of tagmata*

505 The view articulated by Fusco and Minelli [1] is that tagmata cannot be homologized  
506 across arthropod classes. I argue that if all PEF tagmata are derived from an ancestral embryonic  
507 tagma that was defined embryonically in a pre-existing field, they should be seen as homologous.  
508 Similarly, all SAZ-derived tagmata are homologous (it is already generally accepted that the PGS  
509 are homologous across arthropods). However, although similar terms are used in different  
510 lineages (cephalothorax, trunk), there is no way to unambiguously homologize specific segments  
511 or structures within a given tagma across arthropod phylogeny. When comparing between  
512 species with the same number of segments in a specific tagma, it may be possible to equate a  
513 segment in a given position with a segment in the same position in a different species, but this  
514 should be done with caution and with consideration of the evolutionary history of the species  
515 being compared.

#### 516 *The role of Hox genes in segment identity*

517 The prevailing paradigm is that Hox genes are the earliest determinants of segment identity  
518 in arthropods. Specific Hox genes confer tagma fate (e.g. *Antennapedia* is responsible for  
519 thoracic fate). Data on the development of numerous arthropods reviewed above indicates that  
520 the difference between the thorax and abdomen in insects or between the prosoma and  
521 opisthosoma in arachnids is determined very early in development, before the segments are  
522 visible morphologically, even before the segmentation cascade is completed, and indeed, before  
523 Hox gene expression begins in the relevant regions.

524 There is no doubt that Hox genes have a role in specific segment identity and this is  
525 supported by an enormous body of evidence, not only from *Drosophila melanogaster*, but from  
526 many other species [93]. Nonetheless, we must reconsider their role in determining higher-level  
527 identity, i.e. the distinction between segments belonging to different tagmata, based on the  
528 observation that these differences are based on very early embryonic differences.

529 I suggest a two-phase model for segment identity determination. The first phase defines  
530 tagma identity based on embryonic differences in the segmentation process of segments in the

531 different tagmata. The genes responsible for these differences are currently unknown. The second  
532 phase defines the morphological differences between the segments and is responsible for the  
533 development of segment and tagma specific modifications. This phase is largely driven by Hox  
534 genes. The level of integration and mutual regulation and feedback between these two processes  
535 is likewise unknown and can form the basis for a fruitful research agenda.

## 536 **Conclusions**

537 The evolution and development of tagmosis have been discussed intermittently in the  
538 arthropod literature for decades, but there has never been an attempt to offer a synthetic model  
539 for tagmosis. Most of the data presented in this paper have been known for many years. Crucial  
540 pieces of the puzzle were provided by the more recent study of segmentation in hemimetabolous  
541 insects within the context of tagma identity. I have attempted to provide a model of tagmosis that  
542 takes into account morphology, embryonic and post-embryonic segmentation, developmental  
543 dynamics, phylogeny, and data from the fossil record. I have attempted to identify the core  
544 conserved elements of the developmental basis of tagmosis, while disregarding the many  
545 lineage-specific variations in tagmosis that have mostly led to confusion in the field. The  
546 synthetic model I present provides a relatively simple explanation for the observed patterns of  
547 extant and fossil arthropod body plans, but also opens a slew of new questions that can – and  
548 should – be studied experimentally in a range of arthropod species.

549

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Supplementary table 1

Non-exhaustive representative list of papers describing embryonic development of different insect orders, where there is a clear difference between gnatho-thoracic and abdominal segmentation. This difference is usually not pointed out explicitly in the text, but can be seen in the figures.

Species	Taxon	Description	Ref.
<i>Macrocentrotus cingulum</i>	Hymenoptera	Described as a short-germ reversal from a long-germ ancestor. Gnathal segments form pre-gastrulation. Thoracic segments probably arise without axial growth. SAZ active after at least 6 segments express <i>engrailed</i> .	(1)
<i>Cloeon dipterum</i>	Ephemeroptera	The embryonic anlage extends slightly before the morphological appearance of the gnathal and pro-leg (thoracic) segments. Pro-leg anlage are clearly evident before abdominal segmentation begins from the SAZ.	(2, 3)
<i>Galloisiana yuasai</i>	Grylloblattidea	The transition between thoracic and abdominal segmentation is not clear in the description, however, when the first thoracic segments form there is no SAZ and the youngest segments are not at the posterior end of the embryos, while the abdominal segments clearly form from the SAZ.	(4)
<i>Eucorydia masumatsui</i>	Blattodea	An unsegmented germ-band forms within the egg reaching the posterior pole. After its formation, it rapidly segments to give morphologically evident gnathal and thoracic segments, without axial elongation. Abdominal segments form as the germ-band elongates.	(5)
<i>Euphaea yayeyamana</i>	Odonata	Morphologically visible segments differentiate within an extended early germ-band, encompassing the gnathal and thoracic segments. A large SAZ appears after these have differentiated and abdominal segments develop from it sequentially.	(6)
<i>Zorotypus caudelli</i>	Zoroptera	The germ-band elongates and extends to fill the entire egg axis. The SAZ appears at the posterior shortly after gnathal and thoracic segments are evident within the germ-band.	(7)
<i>Metallyticus splendidus</i>	Mantodea	The germ-band is extremely condensed in the posterior of the egg. The transition between thoracic and abdominal segmentation is not clear. The SAZ is very broad and distinct and is not seen during gnathal and thoracic segmentation but only during abdominal segmentation.	(8)
<i>Diplatys flavicola</i>	Dermaptera	The germ-band elongates without obvious segmentation and with no evident SAZ. Morphological segmentation in the gnatho-thoracic region is evident only after abdominal segmentation begins, but covers the region that had elongated initially. Posterior abdominal segments clearly form from the SAZ.	(9)

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