

1 Experimental tests on the evolution of sex and
2 recombination and their adaptive significance

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10 **Abstract**

11 Sex and recombination generate genetic variation and facilitate adaptation by
12 reducing selective interference, but they can also disrupt allelic combinations
13 maintained by selection. We here review experimental evolution studies on the
14 adaptive significance of sex and recombination in constant environments, empha-
15 sizing insights gained from population genomic data. We discuss evidence showing
16 how meiotic segregation (sex) and crossing-over (recombination) disrupt nega-
17 tive disequilibrium between alleles within and between loci and as a consequence
18 increase the fitness variance of populations and enhance selection efficacy. While
19 sexual reproduction can facilitate adaptation when compared to asexual repro-
20 duction, the advantages of high rates of sex and recombination under facultative
21 sexual reproduction or facultative outcrossing and self-fertilization are less clear,
22 especially when overdominance and epistasis cause segregation and recombina-
23 tion loads. We further discuss the challenges of measuring interference between
24 selected alleles, particularly under polygenic adaptation and segregation of mul-
25 tiple modifiers of recombination, and propose directions for future research. Our
26 discussion underscores the nuanced role of sex and recombination in adaptation,
27 shaped by a balance between increased genetic variation and the disruption of
28 beneficial allele combinations.

29 **Keywords:** recombination, sex, crossover, segregation, meiosis, selective interference,
30 epistasis, experimental evolution, population genomics

31 Introduction

32 Sexual reproduction in eukaryotes is widely recognized for its role in generating genetic
33 variation, which fuels adaptation to novel environments by combining and shuffling
34 genotypes, despite its significant physiological and ecological costs (Weismann, 1891;
35 Fisher, 1930; Burt, 2000). With sexual reproduction, genetic variation is generated
36 during “sex” through the meiotic segregation of homologous chromosomes, as well as
37 through “recombination” and the crossing over between non-sister chromatids (Gray
38 and Cohen, 2016; Lenormand et al., 2016). Gene conversion also affects recombina-
39 tion rates, but we will ignore it here (Korunes and Noor, 2017). In prokaryotes and
40 viruses, genetic mixing is common, and may contribute to adaptation, although the
41 underlying replication processes are not necessarily tied with reproduction (Bell, 1982;
42 Maynard Smith, 1990; Redfield, 2001).

43 Understanding the evolution of recombination has been the primary focus in efforts
44 to explain the widespread occurrence of sexual reproduction (Otto and Lenormand,
45 2002; Otto, 2021). Much of the reasoning emphasizes the ability of recombination to
46 reduce selective interference by breaking negative linkage disequilibrium (LD), that
47 is, by breaking the associations between deleterious and beneficial alleles at different
48 loci (Felsenstein, 1965; Eshel et al., 1970). The disruption of negative LD increases
49 the variance in fitness within a population, thereby enhancing the efficiency of natu-
50 ral selection and the evolutionary responses of traits related to fitness (Fisher, 1930;
51 Muller, 1932, 1964; Hill and Robertson, 1966; Otto, 2021). The evolution of sex through
52 segregation is likewise important to understand adaptation and the prevalence of sex-
53 uals, as segregation can increase the fitness variance of a population, and selection
54 efficiency, through the disruption of within-locus negative associations in heterozygotes
55 (Kirkpatrick and Jenkins, 1989; Roze and Michod, 2010; Roze, 2014).

56 When explaining the evolution of sex and recombination, comparisons are often
57 made between sexually and asexually reproducing populations, which are relevant for
58 the emergence of sex and recombination when adaptation occurs from a limited supply
59 of new mutations, or to answer whether sexual populations can resist the invasion of
60 asexual mutants (Maynard Smith, 1990; Redfield, 2001). These comparative arguments
61 imply differential group selection between sexual and asexual lineages, which is not
62 necessarily the case when individuals with variable sex and recombination rates must
63 compete for similar resources in the same habitat and have the opportunity to mate
64 with each other (Burt, 2000). For instance, genetic modifiers that increase sex or
65 recombination rates might facilitate adaptation to a novel environment in the long-
66 term, due to an increase in fitness variance of the population, while not always being
67 favored on the short-term by individual selection due to the disruption of beneficial
68 allele combinations (Feldman and Liberman, 1986; Lewontin and Hubby, 1966; Eshel
69 et al., 1970; Barton, 1995; Roze, 2014).

70 Evolution experiments have been a favorite approach to test for the adaptive
71 consequences of sex and recombination (Charlesworth and Barton, 1996; Otto and
72 Lenormand, 2002; Otto and Barton, 2001; Rice, 2002). Our goal here is to highlight
73 a few of these evolution experiments, particularly those that have placed population
74 genetic observations at the center of the debate [see also Desai (2013); Sharp and Otto
75 (2016); Cvijovic et al. (2018)]. Time-series analysis of the genomic diversity found in

76 experimental populations is now possible, and several studies have attempted to mea-
77 sure the genetic basis of adaptation under different rates of sex and recombination.
78 In addition, observations from natural populations indicate that recombination rates
79 vary along chromosomes, generating recombination rate “landscapes”, which
80 themselves are heritable due to genetic differences between individuals in the distri-
81 bution of crossover position and number (Brazier and Glémin, 2022; Venu et al., 2024;
82 Johnston, 2024). How selection on genetic modifiers of crossover position and num-
83 ber relates to adaptation, particularly when there are many selected loci across the
84 genome (polygenic adaptation), is an open problem.

85 We review illustrative evolution experiments conducted in constant environments,
86 although the evolution of sex and recombination in fluctuating environments is a sig-
87 nificant topic in its own right (Becks and Agrawal, 2010; Morran et al., 2011; Gray
88 and Goddard, 2012a; Kerstes et al., 2012; Masri et al., 2013; Haafke et al., 2016;
89 Lynch et al., 2018). We begin by briefly summarizing the main theoretical predictions
90 for the evolution of sex and recombination and then discuss four related questions
91 in the context of experimental evolution: Does sexual reproduction facilitate adapta-
92 tion compared to asexual reproduction? How does adaptation depend on the realized
93 frequency of sex and recombination? Can selection explain the evolution of sex and
94 recombination? What is the evidence for selective interference in population genomic
95 data? We finish by suggesting directions for future experimental research.

96 Theoretical background

97 Sex and recombination alter genotype frequencies when there is a departure from
98 Hardy-Weiberg and linkage equilibrium within loci and between loci (Felsenstein, 1965;
99 Eshel et al., 1970; Otto, 2003). Such disequilibrium are expected to be common in
100 finite populations, as new mutations arise in phase with the genetic background in
101 which they occur (Fisher, 1930; Muller, 1932), and in a heterozygous state in diploid
102 organisms (Kirkpatrick and Jenkins, 1989). Disequilibrium can also occur due to selec-
103 tion on particular allele combinations, or because finite populations cannot contain
104 all possible alleles at the many loci across the genome (Hill and Robertson, 1966;
105 Eshel et al., 1970; Roze and Michod, 2010; Otto, 2021). Factors such as population
106 subdivision, inbreeding, or self-fertilization can generate an excess of homozygosity
107 by bringing together related genotypes, increasing effective segregation but decreas-
108 ing effective recombination (Nordborg and Donnelly, 1997; Martin et al., 2006; Roze,
109 2009; Roze and Lenormand, 2005; Teterina et al., 2023).

110 Selection can lead to within and between loci disequilibrium, particularly when
111 the fitness effects of alleles are not independent, that is, when there is dominance and
112 epistasis for fitness (Felsenstein, 1965; Eshel et al., 1970; Otto, 2003). With negative
113 epistasis for example — as when deleterious alleles at different loci act synergisti-
114 cally and beneficial alleles act antagonistically — selection leads to negative linkage
115 disequilibrium (LD) and to an excess of genotypes in the population containing dele-
116 terious and beneficial alleles. By breaking negative LD, recombination increases the
117 frequency of genotypes with multiple deleterious alleles and of genotypes with multiple
118 beneficial alleles, thereby increasing the genetic variance in fitness and the efficiency

119 of selection (Felsenstein, 1965; Eshel et al., 1970). In the short-term, however, break-
120 ing (positive or negative) LD generated by epistasis can reduce offspring fitness. A
121 balance between these short-term and long-term effects of epistasis will then dic-
122 tate the evolution of recombination (Barton, 1995). Similarly, when beneficial alleles
123 are partially-dominant, or deleterious alleles partially-recessive, selection will favor
124 negative disequilibrium and an excess of heterozygosity in the population because
125 heterozygotes are fitter on average than their parent homozygotes. Sex, by produc-
126 ing homozygotes through segregation will reduce excess of heterozygosity and thus
127 increase the fitness variance of the population but, like recombination, at the expense
128 of a fitness loss in offspring generations (Otto, 2003).

129 Besides dominance and epistasis, negative disequilibrium in a population, within
130 and between loci, can also arise from the interaction of genetic drift and selection,
131 in what is known as the Hill-Robertson effect (Hill and Robertson, 1966; Felsen-
132 stein, 1974; Barton and Otto, 2005; Roze and Barton, 2006). In any finite population,
133 stochastic fluctuations in genotype frequencies generate both positive and negative
134 disequilibrium. However, selection eliminates positive disequilibrium as the fitter geno-
135 types, which combine beneficial alleles, sweep to fixation, while the worst genotypes,
136 combining deleterious alleles, are purged from the population. Negative disequilib-
137 rium will therefore persist for longer due to selective interference among beneficial and
138 deleterious alleles. Genetic modifiers increasing sex and recombination rates can then
139 be selectively favored as they reduce this interference and allow beneficial alleles to
140 spread more rapidly (Barton and Otto, 2005; Roze and Barton, 2006).

141 In asexual populations, clones carrying different beneficial mutations also interfere
142 with each other, limiting adaptation (Fisher, 1930; Muller, 1932). Asexual populations
143 may further face the so called Muller’s ratchet as clones with less deleterious alleles
144 cannot be recreated without genetic mixing (Muller, 1964). Analogous processes to
145 those occurring in sexuals are expected to generate an excess of heterozygotes in finite
146 diploid asexual populations (Kirkpatrick and Jenkins, 1989; Roze and Michod, 2010).

147 Sexual reproduction facilitates adaptation

148 Most experimental evolution studies about the adaptive significance of sex and recom-
149 bination compared sexual with asexual populations, usually using microorganisms such
150 as the budding yeast *Saccharomyces cerevisiae* and the green alga *Chlamydomonas*
151 *reinhardtii*. [In Table 1 we highlight the evolution experiments here discussed, see
152 also Otto and Lenormand (2002); Rice (2002); Sharp and Otto (2016); Cvijovic et al.
153 (2018); Desai (2013).] A few of these studies accounted for the possibility that the
154 environment inducing sexual reproduction, typically starvation or population density,
155 influenced adaptation. This was achieved by ensuring that the asexuals underwent the
156 same environmental manipulation as sexuals without triggering sexual reproduction,
157 by maintaining a single mating type (Lachapelle and Bell, 2012) or through genetic
158 engineering of meiosis preventing segregation and recombination (Goddard et al., 2005;
159 Gray and Goddard, 2012b).

160 Many of these experiments have conclusively shown that sexual reproduction gener-
161 ally increases the fitness variance of populations and accelerates adaptation to new

162 environments, when compared to asexual reproduction. The advantage of sexual repro-
163 duction over asexual reproduction is particularly evident in harsh, novel environments,
164 but less so in benign conditions to which populations already adapted to. For instance,
165 in benign environments, one study found more extensive adaptation in sexual than in
166 asexual populations (Zeyl and Bell, 1997), another observed no differences (Gray and
167 Goddard, 2012b), and a third one reported a reduction of mean population fitness
168 with sexual reproduction (Renaut et al., 2006). These mixed results can be attributed
169 to weaker selection in benign or to the segregation of predominantly deleterious alle-
170 les once adaptation has been achieved (Hartfield et al., 2010); these explanations not
171 being mutually exclusive, as shown by a study where an increase in mutation rates
172 reduced adaptation in asexual populations under stressful conditions but not in more
173 permissive environments (Gray and Goddard, 2012b).

174 In several studies comparing sexual with asexual reproduction, experimental popu-
175 lations were maintained under predominant haploidy, with diploidy being expressed
176 only during a few sexual cycles (Colegrave, 2002; Kaltz and Bell, 2002; Lachapelle and
177 Bell, 2012; McDonald et al., 2016; Kosheleva and Desai, 2018). In these conditions,
178 the adaptive significance of sexual reproduction is due to recombination, as the effects
179 of sex in the maintenance of heterozygosity should be minimal in the near absence
180 of diploid selection. In the study of Kosheleva and Desai (2018), sexual reproduction
181 facilitated adaptation in diploid yeast populations, although the effects of segrega-
182 tion and recombination in this case remained difficult to disentangle. Segregation is
183 expected to confer an advantage to sexuals over asexuals by reducing deleterious loads
184 (Haag and Roze, 2007) or by generating more homozygotes with beneficial alleles.
185 Asexual populations on the other hand will be hindered because they maintain het-
186 erozygosity (Kirkpatrick and Jenkins, 1989). In general, comparing ploidy treatments
187 is complicated by the fact that diploid populations usually have higher mean fitness
188 and lower fitness variance than haploid populations due to the masking of recessive
189 deleterious mutations. In Kosheleva and Desai (2018) in particular, there was a rel-
190 atively smaller advantage of sex under diploidy than haploidy, which could also be
191 explained by overdominance (Lewontin and Hubby, 1966; Sellis et al., 2016).

192 In the context of asexual reproduction and the evolution of horizontal gene transfer,
193 it is worth mentioning experiments with the RNA bacteriophage $\Phi 6$, where mul-
194 tiple virions can infect the same cell (Malmberg, 1977; Poon and Chao, 2004), or
195 with *Escherichia coli*, where F-plasmid conjugation between cells is possible (Cooper,
196 2007). These experiments showed that adaptation is facilitated when there is oppor-
197 tunity for genetic mixing between virions or cells. Interestingly, in Cooper (2007),
198 a beneficial mutation spread significantly faster in conjugating *E. coli* populations
199 compared to non-conjugating populations, suggesting that selective interference can
200 explain impaired adaptation under asexuality (Desai, 2013; Cvijovic et al., 2018).

201 More comprehensive results about selective interference were found with popula-
202 tion genomic data from yeast experiments (Kao and Sherlock, 2008; Lang et al., 2013;
203 McDonald et al., 2016). In these experiments, adaptation in asexuals involved the
204 spread of clones, each carrying beneficial mutations together with neutral and deleter-
205 ious mutations. In contrast, sexual populations tended to display independent allele

206 frequency changes at many loci across the genome, suggesting that selective interfer-
207 ence is reduced when compared to asexual populations (McDonald et al., 2016). As
208 expected from theory, deleterious mutations often hitchhike with beneficial ones in
209 asexual populations while recombination in sexual populations facilitates their inde-
210 pendent evolution. Yet, how many loci can be independently selected is an unresolved
211 problem. One yeast experiment with facultative sexuals found that selection occurred
212 at fewer than ten loci across replicate populations (Burke et al., 2014), a much smaller
213 number when compared with experiments in exclusive sexuals, such as *Drosophila*
214 *simulans* (Barghi et al., 2020), and raising questions about how variable sex and recom-
215 bination rates determine the extent of selective interference and the polygenicity of
216 adaptation (see next section).

217 It is expected that sex and recombination are favored when many selected alleles
218 interfere with each other (Weissman and Barton, 2012; Weissman and Hallatschek,
219 2014). Consistent with this idea, conjugating *E. coli* and sexual yeast experimental
220 populations show stronger adaptive responses in treatments with higher mutation rates
221 compared to their asexual counterparts (Cooper, 2007; Gray and Goddard, 2012b;
222 Peabody V et al., 2017). Moreover, small population size bottlenecks, by reducing
223 overall genetic diversity, reduce the advantage of sexual reproduction vs asexual repro-
224 duction in the green alga *C. reinhardtii* (Colegrave, 2002). Smaller population sizes
225 can, however, also enhance selective interference (Otto and Barton, 2001; Iles et al.,
226 2003; Barton and Otto, 2005; Roze and Barton, 2006; Roze, 2021), as supported by
227 the benefit of genetic mixing in the RNA bacteriophage $\Phi 6$ undergoing bottlenecks
228 (Poon and Chao, 2004).

229 **Adaptation and variable sex and recombination rates**

230 The experiments outlined so far provide insights into the prevalence of sexual repro-
231 duction over asexual reproduction. However, their relevance in understanding how
232 different rates of sex and recombination influence adaptation is limited (see intro-
233 duction). Furthermore, rates of sex and recombination are generally low in microbial
234 experimental evolution. For example, budding yeast populations in the laboratory typ-
235 ically undergo one round of sexual reproduction every 25 to 120 (mitotic) generations
236 (Goddard et al., 2005; Gray and Goddard, 2012b; McDonald et al., 2016; Kosheleva
237 and Desai, 2018). With six crossovers per chromosome per meiosis (Mancera et al.,
238 2008), much less than one crossover per chromosome per generation are expected on
239 average.

240 It has been suggested that “a little sex goes a long way” (Hurst and Peck, 1996).
241 This is usually meant as: even rare segregation or crossing-over events generate novel
242 allele combinations that can be selected if adaptation depends on a small number of
243 loci (Kirkpatrick and Jenkins, 1989; Green and Noakes, 1995). As mentioned above,
244 several theoretical models suggest, however, that selective interference is stronger, and
245 the long-term advantage of sex and recombination greater, when adaptation is highly
246 polygenic and many of the selected alleles are tightly linked (Weissman and Barton,
247 2012; Weissman and Hallatschek, 2014; Hermisson and Pennings, 2005).

Organism	Ploidy	Focus of the study	Sexual reproduction	Genetic variation	Key findings	Ref.
<i>Saccharomyces cerevisiae</i>	Diploid	Adaptation sex/asex	One round of sex every 25 or 50 generations.	New mutations with or without a mutator	Use of the <i>spo11/spo13</i> genetic system enables identical environmental conditions between asexual and sexual populations. Sex has no effect in a benign environment (30°C) but facilitates adaptation to a harsh environment (37°C). Increasing mutation rates by deleting the MSH2 gene impairs adaptation of asexual populations but not sexual populations.	Goddard et al. (2005); Gray and Goddard (2012b)
	Haploid	Adaptation sex/asex	One round of sex every 90 generations	New mutations	In haploid populations, sex facilitates adaptation to a peptone dextrose medium by disrupting association between beneficial and deleterious mutations	McDonald et al. (2016)
	Haploid or diploid	daptation with varying sex and recombination	One round of sex every 40 or 120 generations.	Standing genetic variation	Sex prevents loss of fitness variance and facilitate adaptation to a peptone dextrose medium. No difference is observed between rare and frequent sex.	Kosheleva and Desai (2018)
<i>Chlamydomonas reinhardtii</i>	Haploid	Adaptation sex/asex	8 successive rounds of sexual reproduction	New mutations	Sex facilitates adaptation to a sodium bicarbonate medium, being reduced in populations having undergone a small population size bottleneck.	Colegrave (2002)
	Haploid	Initial round of sexual reproduction	Adaptation with varying sex and recombination	Standing genetic variation	Following an initial mean fitness reduction, sexual populations show higher fitness variance and faster adaptation to different media, compared to asexual populations.	Colegrave et al. (2002)
	Haploid	Adaptation with varying sex and recombination rates	1 to 3 rounds of sexual reproduction during about 150 generations	Standing genetic variation	Following an initial mean fitness reduction, sexual populations show enhanced adaptation to different media, especially with a larger number of sexual rounds.	Kaltz and Bell (2002)
	Haploid	Extinction with varying sex	1 round of facultative or environmentally-induced obligate sexual reproduction every 4–12 generations.	Varying standing genetic variation	Sexual reproduction and genetic diversity delay the extinction of population in a gradually harsher environment (salt). This effect is more pronounced at higher rate of sex (rounds of obligate vs facultative sex).	Lachapelle and Bell (2012)
<i>Drosophila melanogaster</i>	Diploid	Evolution of recombination	Obligate, outcrossing	Standing genetic variation	Increased recombination evolves in response to artificial selection, often in both directions. Increased recombination occurs in a large portion of the genome but not in all genomic intervals.	(Korol and Iliadi, 1994; Aggarwal et al., 2015)
	Diploid	Evolution of recombination	Obligate outcrossing	Standing genetic variation	Higher recombination rates evolves when artificial selection is applied to bristle number (both directions), but reduced recombination evolves when artificial stabilizing selection is applied.	(Rodel et al., 2004)
<i>Brachionus calyciflorus</i>	Diploid (female) or diploid (male)	Evolution of facultative sexual reproduction	Facultative	Standing genetic variation	Rates of sex transiently increase during adaptation to varying algal food and NaCl. Sex is associated with an increase in fitness variance and a decrease in mean fitness.	(Becks and Agrawal, 2012)
<i>Caenorhabditis elegans</i>	Diploid	Evolution of outcrossing rates	Obligate and partial, facultative outcrossing.	Mutagen-induced mutations	Outcrossing facilitates recovery from mutation loads and adaptation.	(Morran et al., 2009)
	Diploid	Evolution of a modifier of crossover position	Partial, facultative outcrossing	Standing genetic variation	The modifier <i>rec-1</i> mutant smooths the recombination landscape limiting adaptation but is indirectly favored by selection because of hitch hiking with beneficial genotypes it creates in its local genomic neighborhood.	(Paré et al., 2025)

Table 1 Illustrative experimental findings on the evolution of sex and recombination and adaptation.

248 Adaptation from standing genetic variation is typically polygenic (Hermisson and
249 Pennings, 2005; Barghi et al., 2020; Barrett and Schluter, 2008). In evolution exper-
250 iments, adaptation from standing genetic variation is usually studied by crossing
251 different wild isolates or isogenic inbred lines to create genetically diverse (mul-
252 tiparental) hybrid populations (Kaltz and Bell, 2002; Lachapelle and Bell, 2012;
253 Lachapelle and Colegrave, 2017; Kosheleva and Desai, 2018; Teotónio et al., 2017;
254 Macdonald and Long, 2007). Results from these experiments have not been conclusive.
255 In yeast populations derived from two parental isolates, sexual reproduction enhances
256 adaptation compared to asexual reproduction, but the level of adaptation is similar
257 whether sexual cycles occur every 40 or 120 generations (Kosheleva and Desai, 2018).
258 In green algae populations derived from 16 parental isolates, but not in those derived
259 from two parental isolates, frequent rounds of (environment-imposed) obligate sexual
260 reproduction delay extinction in a gradually deteriorating environment compared to
261 facultative sexual reproduction (Lachapelle and Bell, 2012). In another experiment
262 with green algae, populations derived from 12 to 15 isolates, and evolved for more
263 than 150 generations, showed that increasing the number of sexual cycles from one
264 to two improved adaptation, but further increasing to three cycles had no additional
265 effect (Kaltz and Bell, 2002). Hence, neither higher levels of standing genetic varia-
266 tion always lead to higher adaptation nor does frequent sexual reproduction always
267 facilitate adaptation.

268 The question of whether natural populations harbor enough selected alleles to
269 generate the interference explaining obligate sexual reproduction or high sex and
270 recombination rates remains unresolved (Otto, 2021). The number of independently
271 selected alleles should depend on demographic history and factors such as population
272 size and frequency of bottlenecks, population subdivision or predominant reproduc-
273 tion mode (Ellegren and Galtier, 2016). As a result, the choice of the organism can
274 influence experimental outcomes. The specific methods used to derive experimen-
275 tal populations (e.g., through funnel or round-robin designs) will also impact the
276 observations that are made. For example, populations derived from crosses between
277 distant isolates might generate standing genetic variation, but if these isolates differ
278 in their initial adaptation to the experimental environment, outbreeding depression
279 might occur, and positive disequilibrium be common, with sex and recombination in
280 these cases limiting adaptation. Some of these issues can be alleviated with domes-
281 tication by maintaining populations in the laboratory environments until they near
282 a selection-recombination equilibrium before starting with the treatments of interest
283 (Simões et al., 2010; Teotónio et al., 2017).

284 Lack of clear evidence for enhanced adaptation under frequent sexual reproduction
285 may result from a balance between higher fitness variance and reduced mean fitness
286 that is difficult to detect. Populations with standing genetic variation are expected to
287 be prone to recombination loads due to the disruption of beneficial epistatic interac-
288 tions at multiple loci that were favored by prior selection (Hansen, 2006; Neher and
289 Shraiman, 2009). Populations with standing genetic variation should also be prone
290 to segregation loads due to the disruption of heterozygotes with beneficial pseudo-
291 overdominant or truly overdominant alleles (Lewontin and Hubby, 1966; Lewontin
292 et al., 1974; Bierne et al., 2000).

293 Several experiments support the existence of segregation and recombination loads.
294 For example, recombinant lines of *Drosophila spp.* derived from wild populations
295 exhibit reduced fitness compared to lines with recombination-suppressing chromoso-
296 mal inversions or male-derived chromosomes (as males are achiasmatic), reviewed in
297 [Charlesworth and Barton \(1996\)](#). Recombination loads may also explain the fitness
298 loss observed following a round of sexual reproduction in haploid populations of the
299 green alga *C. reinhardtii* with standing genetic variation derived from wild progenitors
300 ([Colegrave et al., 2002](#); [Kaltz and Bell, 2002](#); [Renaut et al., 2006](#)). In the nematode
301 *Caenorhabditis elegans*, a modifier of crossover position that increases recombination
302 rates in chromosomal centers, which should contain epistatic loci ([Chelo and Teotónio,](#)
303 [2013](#); [Noble et al., 2017](#); [Cutter et al., 2009](#)), reduces the mean fitness of a domesti-
304 cated population with standing genetic variation ([Parée et al., 2025](#)). Reduced mean
305 fitness is also observed in sexually vs asexually produced offspring in field-derived pop-
306 ulations of the monogonont rotifer *Brachionus calyciflorus* ([Becks and Agrawal, 2011,](#)
307 [2012](#)). In this latter case, however, and because of diploidy, reduced fitness may be
308 attributed to either a recombination or a segregation load. In general, segregation and
309 recombination loads can cause an immediate fitness loss in offspring generations that
310 may be offset by enhanced selection over time ([Charlesworth and Barton, 1996](#); [Cole-](#)
311 [grave et al., 2002](#); [Kaltz and Bell, 2002](#)); it may be that there is a limit beyond which
312 sex and recombination no longer have an effect on adaptation or even become limiting
313 ([Lewontin and Hubby, 1966](#); [Neher and Shraiman, 2009](#); [Lobkovsky et al., 2016](#)), but
314 conclusive experimental evidence is needed.

315 **Facultative sex and facultative outcrossing under selection**

316 Increased rates of sexual reproduction can be selected against despite their potential
317 role in facilitating adaptation. In some evolution experiments, sexual reproduction may
318 evolve because of (inadvertent) selection on traits unrelated to meiotic segregation
319 and crossing over. This is the case of experiments using facultative sexuals, such as
320 monogonot rotifers, which show phenotype differentiation between sexual and asexual
321 individuals in egg resistance to harsh environments or between individuals in sexual
322 or ploidy identity ([Becks and Agrawal, 2012](#)). In these organisms, observed changes
323 in the rates of sexual reproduction may often result from phenotypic plasticity ([Sasso](#)
324 [et al., 2018](#); [Seudre et al., 2020](#); [Hartfield, 2016](#)). In addition, phenotypic plasticity
325 can vary with individual fitness, as in the white clover *Trifolium repens* where stress-
326 sensitive individuals tend to allocate growth resources to sexual reproduction traits
327 ([Griffiths and Bonser, 2013](#)). Plasticity to engage in sexual reproduction can itself
328 evolve. In the green alga *C. reinhardtii*, for instance, increased rates of spontaneous
329 sexual reproduction evolve when sexual offspring are artificially selected or when there
330 is a single mating type in the population ([Bell, 2005](#)).

331 In the rotifer *B. calyciflorus*, an increase in the rate of sexual reproduction was
332 observed during adaptation ([Becks and Agrawal, 2012](#)). This increase was associated
333 with higher fitness variance and a decline in the mean fitness of sexual offspring when
334 compared to asexual offspring. Because females in this species are diploid, changes in
335 offspring fitness suggest disruption of negative linkage disequilibrium ([Barton, 1995](#))
336 but also the segregation of homozygotes with deleterious recessive alleles ([Roze and](#)

337 Michod, 2010; Roze, 2014). In the experiments of Becks and Agrawal (2012), however,
338 the increase of sexual reproduction was transient. Once adaptation occurred, a reversal
339 to lower levels of sexual reproduction was observed; these results being explained
340 because of weak selection in already-adapted populations (see above) or because of
341 transgenerational environmental effects on the frequency of sexual reproduction. In
342 the sister species to *B. calyciflorus*, transgenerational effects reduce the tendency for
343 sexual reproduction in lineages undergoing more sexual rounds (Seudre et al., 2020).
344 Interestingly, frequent sexual reproduction is observed in *B. calyciflorus* populations
345 when in more complex environments and where a higher number of selected loci could
346 underlie adaptation (Luijckx et al., 2017).

347 Evidence for the evolution of traits associated with sexual reproduction also comes
348 from experiments on the evolution of facultative outcrossing and self-fertilization.
349 Outcrossing rates can change through a plastic response, such as increased outcross-
350 ing rates following a heat stress in the fava bean *Vicia faba* (Bishop et al., 2017), or
351 they can change through selection for reproductive assurance, such as the evolution of
352 self-fertilization in the absence of pollinators in the yellow monkeyflower *Mimulus gut-*
353 *tatus* (Bodbyl Roels and Kelly, 2011; Tusuubira and Kelly, 2024) or the evolution of
354 hermaphroditism in male-female populations of the nematode *C. elegans* (Theologidis
355 et al., 2014).

356 By increasing homozygosity, self-fertilization increases the fitness variance of a
357 population and facilitates the selective purging of deleterious recessive alleles. How-
358 ever, increased homozygosity also decreases effective recombination (Nordborg and
359 Donnelly, 1997), increasing selection interference and possibly reducing mean popula-
360 tion fitness in the presence of overdominant loci (Nordborg et al., 1996; Bierne et al.,
361 2000). In line with these population genetic mechanisms, the evolution of outcrossing
362 with adaptation in experiments with *C. elegans* may be driven by both the masking
363 of recessive deleterious alleles and to the expression of overdominant alleles in het-
364 erozygotes (Teotonio et al., 2012; Chelo and Teotónio, 2013; Chelo et al., 2019). Also
365 in line with these population genetic mechanisms, in the hermaphroditic freshwater
366 snail *Physa acuta*, self-fertilizing populations show fast adaptation during a limited
367 number of generations while outcrossing populations maintain a consistent response
368 during experimental evolution (Noël et al., 2017).

369 Further evidence that the evolution of outcrossing impacts adaptation comes from
370 several experiments with *C. elegans*. In *C. elegans*, hermaphrodites either self-fertilize
371 or outcross by mating with males but not with each other (Stewart and Phillips,
372 2002). In experiments depending on *de novo* mutation for evolution, outcrossing rates
373 can transiently increase upon exposure to a mutagen (Cutter, 2005; Manoel et al.,
374 2007; Morran et al., 2009). However, this increase remains modest even under envi-
375 ronmental conditions where obligate male-female populations show more extensive
376 adaptation (Morran et al., 2009), suggesting that the evolution of outcrossing rates is
377 limited because beneficial genotypes are unlikely to remain associated for long with the
378 hermaphrodite lineages that tend to outcross more often (Lande and Porcher, 2015).

379 Increased outcrossing rates have also been observed in *C. elegans* carrying nuclear
380 and mitochondrial mutations impairing mitochondrial functions (Wernick et al., 2019;

381 [Bever et al., 2022](#)). These observations are relevant to the “mitochondrial sex hypothe-
382 sis” ([Havird et al., 2015](#)), which posits that in recombining populations, high mutation
383 rates in mitochondria are maintained by selection of compensatory nuclear mutations.
384 However, deleterious mitochondrial mutations and their compensatory nuclear muta-
385 tions are beneficial epistatic combinations that can be disrupted by outcrossing and
386 the shuffling of mitochondrial and nuclear genomes ([Nguyen et al., 2023](#)).

387 Selection for recombination and polygenic adaptation

388 Early experiments on the evolution of recombination typically measured the response
389 to artificial selection in *Drosophila spp.* populations carrying large chromosomal inver-
390 sions suppressing recombination ([Charlesworth and Barton, 1996](#); [Rice, 2002](#)). These
391 studies showed that recombination increases selection responses by increasing the fit-
392 ness variance of the population. However, inversions suppress recombination over large
393 portions of the chromosomes and are possibly not representative of the modifiers of
394 recombination segregating in natural populations, such as those modifying crossover
395 position or number ([Johnston, 2024](#)).

396 Theoretical models indicate that modifiers of crossover position or number hitch-
397 hike with the allele combinations they create in their genomic neighborhood and less so
398 with more distant genotypes ([Otto and Feldman, 1997](#); [Roze, 2021](#)). To illustrate the
399 evolution of recombination in a polygenic adaptation context, we run computer sim-
400 ulations with varying distances between many selected loci along a chromosome and
401 a modifier of crossover position that uniformly changes recombination rates between
402 them ([Figure 1](#)). These simulations indicate that the distance to selected alleles deter-
403 mines the strength of indirect selection on the modifier ([Figure 1A,C,D](#)), a result that
404 was already known ([Roze, 2021](#)). Further, when the modifier changes recombination
405 rates between selected alleles non-uniformly, the direction of indirect selection will then
406 depend on whether the modifier is located near them ([Figure 1B,C,E](#)). However, while
407 selection for the modifier depends on its position, its effects on recombination among
408 all selected alleles across the chromosomes is the same independently of its position
409 and should not impact (genome-wide) polygenic adaptation. A modifier of crossover
410 positions that results in a more uniform recombination across selected alleles should
411 maximize genetic mixing and shuffling and thus reduce selective interference ([Veller
412 et al., 2019](#)).

413 For indirect selection to result in the evolution of recombination, selected loci
414 need to be in close proximity to recombination modifiers. This is plausible in many
415 experiments where increased recombination rates evolved as a correlated response to
416 artificial selection ([Korol and Iliadi, 1994](#); [Aggarwal et al., 2015](#)), as recombination
417 rates often increase across large portions of the genome, and selected traits, such as
418 stress resistance, are often highly polygenic ([Lynch and Walsh, 1998](#)). Additionally,
419 the segregation of multiple recombination modifiers in the experimental populations
420 increases the likelihood that modifiers are located near the selected alleles. The strong
421 selection pressures usually applied in these experiments [e.g., only 30 individuals out of
422 300 contributing to the next generation in [Korol and Iliadi \(1994\)](#)] may further allow
423 the modifiers to be indirectly selected with more distant selected loci ([Roze, 2021](#)).

Fig. 1 Indirect selection on modifiers of crossover position. The figure shows simulations of indirect selection on a modifier of crossover position and number along a chromosome. The ancestral population of 10^3 diploid individuals was generated with burn-in of 10^3 generations with one selected mutation ($s \sim \text{Exp}(0.005)$) introduced per generation at each of 10^4 loci across the chromosome. After burn-in, a neutral recombination modifier was introduced in 500 randomly sampled genomes. We modeled the evolution of recombination under four different scenarios (**A,B**). The black line represents the chromosome, the purple and green circle represent the deleterious and beneficial alleles, the blue circle the modifier. **A.** In scenarios a and b, individuals homozygous for the modifier allele (M) have a 3-fold chromosomal map length (R_M) increase relative to the ancestral map length (R_m ; assuming the modifier alleles are co-dominant). **B** In scenarios c and d, one of the modifier alleles increases the map length in a first interval, i , and reduces it in an adjacent interval, j , by a factor 3. **C.** The modifier allele frequency during 500 generations. Thin lines show the trajectories for 40 simulation runs per scenario, thick lines show the average of 500 simulations runs. **D.** Indirect selection coefficient (binomial generalized linear model: *modifier.freq* \sim *generation*) as a function of the genetic distance of the modifier to selected loci, for scenarios a,b, and in two additional scenarios (dots = mean; error-bars = 95% CI). Indirect selection decreases log-linearly with the genetic distance between the modifier and selected loci. **E.** Indirect selection coefficients for scenarios c,d are positive when the modifier is located in interval i , where it increases recombination, and negative when it is located in interval j , where it decreases recombination.

424 The evolution of recombination rates in response to artificial selection has been
425 quantified in many different species (Otto and Barton, 2001; Otto and Lenormand,
426 2002; Rodell et al., 2004; Aggarwal et al., 2015), including domesticated species (Ross-
427 Ibarra, 2004; Bursell et al., 2024). Increased recombination rates tend to evolve when
428 the trait is selected in a different direction (Korol and Iliadi, 1994; Rodell et al.,
429 2004; Aggarwal et al., 2015), indicating that pleiotropy (where the responding allele
430 affects both the selected trait and recombination rates), or spurious initial associa-
431 tions between the modifier and selected loci, are unlikely to explain the evolution of
432 recombination. Rather, these experiments suggest that modifiers are indirectly selected
433 because they enable more efficient selection on the targeted trait. In experiments
434 where artificial stabilizing selection was applied, reduced recombination rates evolved
435 (Rodell et al., 2004), a result explained by recombination among the fitter genotypes
436 at optimal trait value generating less fit genotypes (Charlesworth et al., 1993; Whit-
437 lock et al., 1995). Other experiments in *D. melanogaster* have shown the evolution
438 of recombination rates at particular genomic locations (Kohl and Singh, 2018; Win-
439 bush and Singh, 2021, 2022). However, in these latter experiments, the environmental
440 conditions imposed, such as temperature, may affect proper crossover formation and
441 directly affect individual fitness, explaining selection of the modifiers independently
442 of their recombination effects on selected alleles (Grushko et al., 1991; Samuk et al.,
443 2020).

444 Recently, the evolution of recombination was investigated by following the evolu-
445 tion of a known modifier of crossover position in *C. elegans* (Parée et al., 2025).
446 Mutant populations for the *rec-1* gene redistribute crossovers to chromosome centers
447 while not changing crossover number (Parée et al., 2024). Chromosome centers contain
448 less genetic diversity and less fitness variance than flanking regions, and as a conse-
449 quence, *rec-1* mutant populations showed impaired adaptation (Parée et al., 2025).
450 However, the *rec-1* mutant allele was favored by selection because it increases recom-
451 bination rates in its genomic neighborhood. Consistent with the Hill-Robertson effect

452 mentioned in the introduction, stronger selection on the recombination modifier *rec-1*
453 was observed at smaller population sizes.

454 Detecting selective interference

455 The extent of selective interference in a population will determine the benefits of
456 recombination on adaptation (Otto, 2021). Yet, detecting selective interference from
457 the genomic diversity data that is now typically collected from evolution experiments
458 is challenging. It requires identifying selected loci and estimating negative linkage dis-
459 equilibrium between them, which is not trivial when adaptation is polygenic and there
460 is an astronomical number of genotypes to measure (Macdonald and Long, 2007; Long
461 et al., 2015; Schlötterer et al., 2015). Most studies use presumably neutral markers,
462 such as single-nucleotide variants, that themselves must be in linkage with selected
463 alleles to be informative. Only rare studies have shown disruption of negative link-
464 age disequilibrium between a small number of selected alleles, e.g., McDonald et al.
465 (2016). Instead, average genome-wide estimates of marker allele frequency dynamics
466 have been used.

467 One way to estimate linkage between selected loci is by measuring the correlation
468 between marker allele frequencies at each replicate population undergoing experimen-
469 tal evolution (McDonald et al., 2016; Parée et al., 2025; Burke et al., 2014; Barghi
470 et al., 2019). For instance, in the *C. elegans rec-1* experiments mentioned above, adap-
471 tation was associated with higher correlations of allele frequency change in genomic
472 regions with reduced recombination rates (Parée et al., 2025). The temporal covari-
473 ance of allele frequency change is another metric that can be used to detect linked
474 selection (Buffalo and Coop, 2019). In this case, it is assumed that changes in the allele
475 frequencies of the (neutral) markers depend on the genetic backgrounds with selected
476 alleles. If so, then the change in allele frequency of a given marker in a period should
477 be less correlated with its frequency change at subsequent periods as recombination
478 swaps markers among genetic backgrounds (Buffalo and Coop, 2019). Although this
479 method has faced some criticism (Lynch and Ho, 2020), the expected reduction in
480 temporal covariance between distant time intervals was observed in experiments with
481 *D. melanogaster* and the marine copepod *Acartia tonsa* (Buffalo and Coop, 2020;
482 Brennan et al., 2022). In the *D. melanogaster* study, there was an excess of negative
483 temporal covariance, further indicating that marker alleles reversed their frequency
484 trajectories, and reflecting the disruption of negative linkage disequilibrium or the
485 presence of epistasis (Buffalo and Coop, 2020).

486 It has also been argued that selective interference can be measured as the non-
487 parallel genomic diversity responses among replicate populations during experimental
488 evolution (Kosheleva and Desai, 2018). Recombination should increase parallelism by
489 allowing the same alleles to be independently selected across replicates. Kosheleva and
490 Desai (2018) calculated the standard deviation of marker allele frequencies between
491 replicates across generations as the inverse of parallelism (divergence), finding that
492 it is lower in sexual than asexual yeast populations. Yet, recombination can dimin-
493 ish parallelism when calculated as the correlation of marker allele frequency change
494 among replicates [referred to as the “convergence correlation” in Buffalo and Coop
495 (2020)]. In this case, parallelism might be high in the first few generations but it should

496 quickly decay because ancestral lineages recombine and the fitness of specific genetic
497 backgrounds will then determine marker frequency trajectories.

Fig. 2 Parallelism of genomic diversity responses among replicates of experimental evolution. Similarly to [Figure 1](#), standing genetic variation of the ancestral population was generated was modeled $R=50$ cM and an average of one mutation per genome per generation, now for a total of 10^5 loci. Both neutral alleles ($s = 0$) and selected alleles [$s \sim \text{Exp}(0.005)$] were modeled with varying numbers indicated above the panels (proportion of p_{neutr} and p_{sel} in the chromosome, respectively). With a higher the number of selected alleles, the stronger interference between them. Adaptation was simulated during 500 generations with several map lengths (R ; in cM). **A.** Adaptation is faster and more extensive for larger values of R and lower number of selected alleles. **B.** Parallelism among replicates was calculated as the Pearson correlation (r_s) of selection coefficients among ten simulation runs and from the same ancestral population. Increasing R increases this correlation only when it is small and/or the number of selected alleles is high.

498 We adapted the simulation model of [Figure 1](#) to better understand this dual effect
499 of recombination on the parallelism of replicate responses during experimental evo-
500 lution ([Figure 2](#)). We simulated adaptation from standing genetic variation under
501 different recombination rates and different numbers of selected alleles. Results show
502 that the relation between recombination and parallelism among replicate simulations
503 is non-monotonic. Parallelism increases with recombination only under conditions of
504 low recombination rates and a high number of selected alleles. Conversely, with weaker
505 selective interference, increased recombination rates decrease parallelism despite facil-
506 itating adaptation. While higher parallelism with recombination may indicate strong
507 selective interference, low parallelism does not necessarily indicate its absence.

508 Future research directions

509 Evolution experiments have established that sexual reproduction facilitates adaptation
510 by breaking negative disequilibrium between selected alleles, and this may in part
511 explain the prevalence of sexual reproduction over asexual reproduction. However,
512 evidence supporting a role for high rates of sex and recombination in adaptation
513 remains limited. While adaptation is undoubtedly highly polygenic, it is still unclear
514 whether a sufficient number of alleles can be independently selected to provide an
515 advantage to high sex and recombination rates. This uncertainty is compounded by
516 the opposing effects of segregation and recombination on the maintenance of beneficial
517 epistatic or overdominant alleles ([Peters et al., 2003](#); [Sellis et al., 2016](#); [Johnson et al.,
518 2023](#)). Variable recombination rates across the genome might help maintain beneficial
519 alleles in low-recombining genomic regions, such as when populations adapt to local
520 environmental conditions ([Kenig et al., 2015](#); [Altenberg et al., 2017](#); [Wolf and Ellegren,
521 2017](#); [Venu et al., 2024](#)). In this scenario, however, it remains an open question whether
522 recombination evolves when modifiers of dispersal, or modifiers affecting dominance
523 and epistasis, also segregate within populations ([Proulx and Teotónio, 2022](#)).

524 While the consequences of high rates of sex and recombination for adaptation may
525 depend on the number of selected alleles ([Weissman and Barton, 2012](#); [Weissman and
526 Hallatschek, 2014](#)), several theoretical models suggest that modifiers of traits asso-
527 ciated with sexual reproduction are likely to evolve ([Otto, 2003](#); [Roze and Michod,](#)

528 2010; Roze, 2014). Under random mating, segregation effects should favor sexual repro-
529 duction when deleterious alleles are dominant (Roze and Michod, 2010; Glemin and
530 Ronfort, 2013). Recessive deleterious alleles, which appear to be common in natu-
531 ral populations, further tend to create segregation loads that disfavor higher rates of
532 sexual reproduction under random mating. High rates of sexual reproduction can be
533 favored when excess homozygosity occurs, as this allows heterozygotes to mask dele-
534 terious recessives or expose overdominant alleles to selection (Otto, 2003; Dolgin and
535 Otto, 2003; Bierne et al., 2000; Chelo and Teotónio, 2013; Chelo et al., 2019). Pop-
536 ulation subdivision, inbreeding among relatives, or self-fertilization – which generally
537 increase homozygosity – may play an important role in explaining the evolution of
538 sexual reproduction (Teterina et al., 2023; Roze and Lenormand, 2005; Roze, 2009).
539 Overall, many of these ideas have yet to be tested with experimental evolution.

540 The direct fitness effects of modifying crossover position and number, such as com-
541 promising DNA repair or the segregation of homologous chromosomes during meiosis,
542 could play a significant role in the evolution of sex and recombination (Kleckner et al.,
543 2004; Hunt, 2006; Otto and Payseur, 2019; Fernandes et al., 2018; Morgan et al., 2021).
544 There is strong experimental evidence, however, that indirect selection can also drive
545 the evolution of modifiers crossover position and number (Aggarwal et al., 2015; Parée
546 et al., 2025). These modifiers are expected to be favored when they generate beneficial
547 genotypes in their local genomic neighborhood, rather than by enhancing genome-
548 wide selection efficacy and facilitating polygenic adaptation (Otto and Feldman, 1997;
549 Roze, 2021). Any factor that increases linkage disequilibrium prolongs the association
550 between a recombination modifier and the genotypes it produces. This in turn alters
551 the strength of indirect selection and may even shift its direction by expanding the
552 genomic region associated with the modifier (Stetsenko and Roze, 2022). As shown
553 with evolution experiments in *C. elegans*, polygenic adaptation may often be weakly
554 related with selection of modifiers of crossover position across the genome (Parée et al.,
555 2025).

556 When recombination itself is polygenic, driven by the segregation of multiple
557 crossover modifiers (Johnston, 2024), selection conflicts may arise between them if they
558 have opposing effects on recombination among selected alleles. Our understanding of
559 how recombination evolves with multiple modifiers across the genome remains is still
560 at a very early stage. Selection on multiple modifiers across the genome might average
561 out, aligning or diverging from the effects of a given modifier on genome-wide recom-
562 bination rates. It is further unclear whether indirect selection is substantial when the
563 recombination effects of a modifier are highly localized to its genomic neighborhood,
564 e.g., (Ubeda et al., 2019). If the evolution of recombination is primarily governed by
565 modifiers of local recombination rates, rather than genome-wide, indirect selection
566 may help with the maintenance of adaptation to local environmental conditions (Wolf
567 and Ellegren, 2017; Venu et al., 2024).

568 In conclusion, advancing our understanding about the evolution of sex and recom-
569 bination, as well as their adaptive significance, will require identifying the genomic
570 location of sex and recombination rate modifiers together with the selected loci. Addi-
571 tionally, it is worth considering whether our current understanding is biased by the
572 predominant focus on microbial and metazoan experimental evolution, highlighting the

573 need for greater attention to underrepresented organisms such as plants and multicel-
574 lular fungi. These efforts will certainly pave the way for developing new experimental
575 models manipulating modifier associations with selected alleles in genomic, life-history,
576 or ecological contexts that are more representative of natural populations than those
577 studied to date.

578 **Simulation methods.** The individual-based simulations of diploid populations
579 were implemented using *SLiM* 4.0.1 (Haller and Messer, 2023), using the default
580 Wright-Fisher models for mutation, selection, and reproduction. Ancestral populations
581 with standing genetic variation were obtained by 1000 generations of reproduction
582 with recombination (map length values specified in the legend of the figures) with an
583 average of one mutation per genome (mutation rate of $u = 1/L$, where L is the number
584 of loci). Unless otherwise indicated in the figure legend, all mutations have a selection
585 coefficient sampled from an exponential distribution with an expected mean of 0.005.
586 Simulations start from an ancestral population and differ in their map length or the
587 presence and position of a modifier of recombination rates. For each ancestral popula-
588 tion generated, 10 simulations per parameter set were performed for 500 generations.
589 The mutation rate during these 500 generations was set to 0. Simulation scripts, results
590 and modeling details are available in our [GitHub repository](#) and in Dryad.

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