¹ **Understanding Host-Microbiome Evolution through the**

Lens of Evolutionary Theory:

³ **New Tricks for Old Dogs**

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¹⁴ **ABSTRACT**

All plants and animals are host to a community of microorganisms, their microbiomes, that have crucial influences on the life history and performance of their hosts. Despite the importance of such host-microbiome relationships, relatively little is known about the role microbiomes play in mediating evolution of the host as well as entire host-microbe assemblages. This knowledge gap is partly due to the lack of theoretical frameworks that generate testable predictions on the evolutionary dynamics of host-microbiome systems. In this Perspective, we argue that the foundation for such frameworks exists in evolutionary theory. We highlight four

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examples of theoretical models - niche construction, indirect genetic effects, maternal effects and multilevel selection – that capture important aspects of host-microbiome evolution. We outline how each of these frameworks can provide key insights into the involved evolutionary dynamics, while also suggesting expansions of current theory to incorporate processes unique to host-microbe assemblages, for instance focusing on nuances in microbiome transmission and ecological microbial community dynamics. Expanding evolutionary theory to accommodate host-microbiome systems is key for a more integrative understanding of evolution, which is undoubtedly impacted by the association with microorganisms across the tree of life, guiding future empirical research on the function and evolution of these omnipresent interactions.

¹⁶ **Introduction**

¹⁷ All eukaryotic life has long-standing, intimate relationships with microorganisms. These host-associated microbial

¹⁸ communities (including bacteria, archaea, viruses, protists, and fungi, together termed the microbiome) are crucial for

^{[1](#page-11-0)9} host performance, affecting host traits related to metabolism¹, pathogen resistance^{[2](#page-11-1)}, immune development^{[3](#page-11-2)}, disease^{[4](#page-11-3)},

²⁰ and behavior^{[5](#page-11-4)}, among many others. Beyond its fundamental interest, the potential applications of the microbiome ²¹ vary widely, ranging from human health^{[6](#page-11-5)}, to sustainable agriculture^{[7](#page-11-6)}, conservation biology^{[8](#page-11-7)}, and adaptation to

²² climate change^{[9](#page-11-8)}.

 Yet, despite the clear relevance of host-associated microbiomes to host performance, relatively little is known about the causes and consequences of evolution in such host-microbe associations. Proposed almost two decades ago, the hologenome theory of evolution posits that the holobiont (i.e., the host and all its associated microbes) $_{26}$ functions as a single, integrated evolutionary unit upon which selection acts^{[10](#page-11-9)}. Here, the hologenome refers to all

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²⁷ host genes together with the genes of all host-associated microbes. Since the introduction of this theory, various $_{28}$ perspectives^{[11–](#page-11-10)[19](#page-12-0)} have stimulated research on this, as it has turned out to be, controversial topic. Major challenges in ²⁹ considering hosts and their microbiomes as a single evolutionary unit are substantial variation in microbiome fidelity ³⁰ across generations (Box 1), and the multiple levels of selection and evolutionary interests possible in host-microbe 31 associations^{[19](#page-12-0)}.

 λ An in-depth evaluation of these conflicting perspectives would greatly benefit from theoretical approaches^{[20](#page-12-1)}. Theory facilitates organization of observations, identifies generalities and gaps in our understanding, predicts future events, and provides guidance on the main questions and designs of empirical studies. Theory is especially useful for understanding processes that occur at temporal or spatial scales challenging to study, such as evolution. While $_{36}$ previous studies developed theoretical models tailored to specific questions about host-microbe systems^{[21,](#page-12-2) [22](#page-12-3)}, the full breadth of well-established evolutionary theory has not been applied to understand the evolutionary dynamics and resulting consequences of host-microbe associations.

³⁹ We propose to make use of the wealth of theoretical approaches in evolutionary biology to explore and dissect the evolution of host-microbiome interactions. We highlight four existing frameworks that address key characteristics of host-microbiome evolutionary dynamics. We discuss how we may borrow useful elements from each of these frameworks, while also highlighting fundamental differences between host-microbe evolutionary dynamics and existing frameworks, pinpointing features of host-microbiome evolution that require the development of new theoretical approaches. We point to important directions for future theoretical work, while emphasizing the importance of integrating theory and empirical work.

Box 1: Microbiome Inheritance

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The fidelity of the microbiome (Figure B1) across host generations is the most critical factor that determines whether microbes share the same evolutionary interests as their hosts and thus may function as a single evolutionary unit. Several mechanisms could result in host-microbe associations exhibiting fidelity across host generations.

One process that could result in cross-generational host-microbiome fidelity is the vertical transmission of microbes from parents to offspring. Strict vertical transmission, akin to genetic inheritance, occurs through intracellular infection of germ cells, for example observed in aphid-*Buchnera*[23](#page-12-4) and in carpenter ant-*Blochmannia*[24](#page-12-5) systems. However, even in the absence of strict vertical transmission, 'intimate neighborhood transmission^{'[25](#page-12-6)} may result in the transmission from parents to offspring, for instance through the covering of eggs with microbes^{[26](#page-12-7)}, through mode of delivery in humans^{[27](#page-12-8)}, or through hosts shaping their microbial environment as a form of niche construction^{[28](#page-12-9)}. Further, vertical microbiome transmission goes beyond direct transmission from parents to offspring: living in proximity (e.g., sharing the same household with relatives) may promote microbiome fidelity^{[29](#page-12-10)}. Even in the absence of vertical transmission, host genotypes might directly influence the types of microbes that can establish in a particular host, shaping microbiome composition and increasing cross-generational fidelity^{[30](#page-12-11)}. Environmental transmission can also result in host-microbe fidelity across generations, whenever hosts faithfully acquire the same microbes from the environment every generation, as found in the Bobtail squid–*Vibrio*^{[31](#page-12-12)} and stinkbug-*Burkholderia* associations^{[32](#page-13-0)}. Whenever the environmental microbial pool responds to selection on hosts, environmental acquisition alone can lead to cross-generational microbiome fidelity, through 'collective inheritance'^{[33](#page-13-1)}.

Despite all these different biological processes that may bolster microbiome fidelity, many host-associated microbes were proposed to lack cross-generational fidelity^{[19](#page-12-0)} and the exact degree of microbiome fidelity is often unknown for most host species. One way to quantify this relationship is to estimate the microbiome 'heritability' (Figure B1); the percent of microbiome variance (e.g., variance in relative abundance of a microbial taxon across hosts in a population) attributable to host genotypic variance. Microbiome heritability has been estimated for only a limited number of plant and animal host species , suggesting low microbiome heritabilities in general, although some were on par with heritabilities of important host traits.

Figure B1: Components of microbially mediated inheritance. Heritability of host traits (left panel) measures the similarity of offspring traits to parental traits. Intergenerational microbiome fidelity (middle panel) is the similarity of microbiome compositions between host generations. Microbially mediated host trait heritability depends on microbiome fidelity, which in turn depends on the process of microbial inheritance (right panel).

Adapting Evolutionary Frameworks for Host-Microbiome Systems

⁴⁹ A fundamental question is to what extent we need to develop new theory to describe host-microbiome evolution, and where we can draw on existing frameworks. Utilizing existing frameworks has the advantage of making the $_{51}$ ideas, reasoning, and conclusions more accessible to researchers already familiar with such frameworks.

 Host-microbiome systems are shaped by an array of diverse processes across many scales of biological organization, and it is unlikely that a single existing framework will capture them all. For example, metacommunity theory has been used to describe fundamental processes that influence the assembly of microbial communities, and this approach ⁵⁵ is relatively well established^{[35,](#page-13-3) [36](#page-13-4)}. However, most such models ignore host-microbe feedbacks and host evolution^{[37](#page-13-5)}. In general, we lack an understanding of the consequences of natural selection on host-microbiome systems and the inheritance of selected (microbiome-mediated) variation in particular. In the coming sections, we explore how four evolutionary frameworks may help us understand such microbiome-mediated host adaptation over the timescale in which host microevolutionary dynamics occur (e.g., from a single to possibly thousands of host generations), each addressing different aspects of the evolution of host-microbiome associations (Figure 1): (1) Niche construction, (2) Indirect genetic effects, (3) Maternal effects, and (4) Multilevel selection.

 Frameworks 1-3 focus on microbiome-mediated host evolution, considering host-associated microbiomes essentially as a form of non-genetic inheritance (NGI). NGI involves the transmission of other factors than the DNA (e.g., epigenetic patterns, cytoplasmic transmission, nutrient provisioning, and cultural inheritance), from parents (or other conspecifics) to offspring. Depending on how these non-genetic factors covary and interact with genetic, ϵ environmental and/or stochastic factors, NGI can manifest itself as, for instance, maternal effects^{[38](#page-13-6)}, ecological ⁶⁷ inheritance^{[39](#page-13-7)} or indirect genetic effects^{[40](#page-13-8)}. There exists a large body of literature on the implications of NGI for ⁶⁸ plant and animal evolution^{[41](#page-13-9)}. Since various non-genetic inherited mechanisms share analogies with host-associated microbiomes, it provides a useful framing to think about microbiome-mediated host evolution, as we will discuss. As σ a fourth framework, we outline how we can use tools from multilevel selection and inclusive fitness theory to describe π how composites of individuals respond to selection that jointly acts on various scales of biological organization, resulting in host-microbe coevolution.

 For each framework, we briefly summarize the main concepts and discuss its merits and limitations for under-standing host-microbiome evolution. We note these frameworks are not mutually exclusive, and each has a wide

variety of perspectives through which they may be viewed.

Niche Construction

 π . The framework of niche construction is centered on the reciprocal dynamics of populations and their environment.

Niche construction considers the indirect effects of organismal activity on their own or descendants' fitness through

⁷⁹ environmental modification, and it has been considered an evolutionary process in its own right^{[42](#page-13-10)}. There are

- two interpretations of niche construction, that may both apply to host-microbiome associations (Figure 1): A)
- environmental modification by organismal activity (which may or may not have evolutionary consequences) and B)
- an evolutionary process involving feedback between environmental change and organismal evolution.

Niche Construction as Environmental Modification

- ⁸⁴ Niche construction as environmental modification by organismal activity without necessarily establishing an evolu-
- tionary process, applies to host-microbiome systems in at least two ways (Figure 1a). First, the microbiome of a
- host (such as microbiomes associated with the host's skin or gut) can be considered as a host's environment (Figure
- 1a-I). Host activity that results in microbe acquisition (such as through feeding, social behavior, or other means),
- and host immune responses that result in selection of microbes, provide mechanisms of niche construction.

 Second, the microbiome of a host's immediate surroundings (e.g., microbial communities associated with different food sources, or with surfaces the host comes into contact with) can be considered as a host's environment. Niche construction then occurs when host activity alters the environmental microbiome composition, for instance by

- shedding microbes into their surroundings at a sufficient rate (Figure 1a-II), or by other activities including host-
- ⁹³ mediated structuring of the environment (e.g., nest building)^{[43](#page-13-11)}, provision of nutrients (e.g., "priming" of soil microbes
- ⁹⁴ by plant roots)^{[44](#page-13-12)}, and any kind of "farming" activity (e.g., the cultivation of fungi by insects)^{[45](#page-13-13)} (Figure 1a-III).
- These scenarios can alter either microbiome composition or microbial activity, and, through these changes in host
- habitat, may consequently affect host fitness (Figure 1a-IV). Such modifications can include increasing nutrient
- ⁹⁷ availability or suppressing pathogens, as has been observed to occur in soil surrounding plant roots^{[46,](#page-13-14)[47](#page-13-15)}.

Niche Construction as an Evolutionary Process

 The interpretation of niche construction as an evolutionary process is more stringent, as it requires the inheritance of natural selection pressures (Figure 1b). This process, called ecological inheritance , requires that organismal activity μ_{101} shapes selection on genetic variation, and that these selection pressures are transmitted to subsequent generations^{[42](#page-13-10)}. Host-associated microbiomes can establish modes of ecological inheritance in at least two ways.

 First, by mediating host trait expression, microbiomes can facilitate the inheritance of natural selection pressures on host genetic variation associated with that trait (Figure 1b-I) To illustrate this, consider a host trait z that is 105 additively determined by a genetic component g and a microbiome component m such that $z=g+m$. This model ¹⁰⁶ of host trait architecture has previously been applied by 48 . Writing host fitness as a function of host trait, $W(z)$, 107 selection on the host trait is defined as the covariance of fitness and phenotype, $Cov(W,z)$. Then, setting $W_G(q)$ 108 as the average of $W(z) = W(g+m)$ across *m* for fixed *g* (i.e., the marginal fitness of host genetic value g across variation of host microbiomes), and $W_M(m)$ as the marginal fitness of the microbiome component *m*, selection on host trait decomposes as follows:

$$
Cov(W, z) = Cov(W_G, g) + Cov(W_M, m). \tag{1}
$$

 This decomposition illustrates that selection at the level of host trait results in indirect selection at the levels of host genotype and host microbiome, and that these selection pressures are mediated by complementary components 113 of host trait variation. In particular, because $W_G(g)$ is an average across microbiome variation, selection on host

Figure 1. Graphical illustration of the discussed frameworks that exist in evolutionary biology and that address key characteristics of host-microbiome evolution. Note that these frameworks are not mutually-exclusive.

 $_{114}$ genotype $(Cov(W_G, g))$, is a function of the distribution of host microbiome variation. Hence, in this case, the transmission of microbiome variation across host generations such that the distribution of microbiome variation in host parental generations resembles the distribution of microbiome variation in host offspring generations (see Box 1), establishes a mode of ecological inheritance.

 Second, microbiomes can modify selection pressures on host genetic variation by altering the host environment (e.g., via host shedding), such as resource availability and habitat quality, (Figure 1b-II). These modified selection pressures can have long-term evolutionary consequences on host traits, such as immunological profiles, tissue structures, and physiological processes that influence specific microbial functions^{[14](#page-11-11)}. In this case, the focal host trait need not be mediated by the host microbiome, and instead could be purely genetically mediated. For the sake of 123 clarity, in this paragraph we adopt this assumption, such that $z = g$. Summarizing the effect of the environmental 124 microbiome on host fitness as m_E , we can include it as a parameter of the fitness function: $W(g|m_E)$. Then, the 125 correlation of m_E between host generations maintained by host activity (such as shedding) results in the inheritance of selection pressures on host genetic variation, and therefore establishes a second mode of ecological inheritance.

¹²⁷ **Indirect Genetic Effects**

¹²⁸ Indirect genetic effects (IGE) are the influence of an individual's genotype on the phenotype of another (typically conspecific) individual^{[40](#page-13-8)}. Because IGEs contribute to the expression and inheritance of phenotypic variation, ¹³⁰ they have important evolutionary consequences. A major application of IGE is to understand the evolutionary $_{131}$ consequences of social interactions in social insects^{[49](#page-14-1)[–52](#page-14-2)}.

 Host-associated microbiomes establish IGE between host individuals when three conditions are met: 1) host genes determine microbiome composition, 2) microbiome composition is transmissible, and 3) microbiomes mediate a host trait. When these conditions are met, microbiome transmission (which may occur during social encounters) forms the mechanism for the genes of one host to influence the phenotype of another host (Figure 1c).

136 To illustrate this, consider microbiome composition as a host trait z_1 that is mediated by host genotype g_1 such ¹³⁷ that $z_1 = g_1 + e_1$, where e_1 is an environmental effect. Now consider another host trait that is mediated by host ¹³⁸ genes and microbiome composition

$$
z_2 = g_2 + e_2 + z_1. \tag{2}
$$

This is similar to the starting point taken by 53 to derive their model for interactions with nonreciprocal effects. ¹⁴⁰ As a consequence of microbiome transmission via social contact, we might take the same starting place and assume that each individual engages in a single interaction with another randomly chosen individual. Denote by z'_1, g'_1, e'_1 the ¹⁴² trait and trait components for the non-focal interacting individual. Suppose that the social interaction results in an ¹⁴³ exchange of microbes between the interacting partners, so that their microbiome composition traits become similar ¹⁴⁴ by some amount $0 \le \psi \le 1$. Writing z_1 as the microbiome composition of the focal individual after the interaction, ¹⁴⁵ we then have

$$
z_1 = (1 - \psi)z_1 + \psi z_1'.
$$
\n(3)

146 Assuming the second trait z_2 is expressed after the exchange of microbes has occurred, it can then be written as

$$
z_2 = g_2 + e_2 + z_1 = g_2 + e_2 + (1 - \psi)(g_1 + e_1) + \psi(g_1' + e_1').
$$
\n(4)

¹⁴⁷ The coefficient ψ , which measures microbial transmission via social contact, also quantifies an indirect genetic effect ¹⁴⁸ of the interacting partner's additive genetic value g'_1 on the expression of the focal individual's trait value z_2 .

 The IGE framework has applications for understanding the dynamics of host-associated microbiomes, particularly for systems where social transmission of microbes between unrelated individuals plays an important role. Because the IGE framework considers interactions between arbitrary individuals, and only incorporates non-random interactions mediated by trait covariances, and not necessarily relatedness^{[40](#page-13-8)}, it requires additional assumptions to apply to systems with substantial parent-offspring microbe transmission. To model this complementary scenario more directly, the related framework of maternal effects has greater utility.

Maternal Effects

 A maternal effect is the influence of a parental phenotype on an offspring phenotype, controlling for genetic variation, $_{157}$ mediated by parent-offspring interactions such as maternal care^{[54](#page-14-4)}. Host-associated microbiomes establish maternal effects between host parents and host offspring when a host trait is mediated by its microbiome, and part of the host's microbiome is inherited from direct parent-offspring transmission (see Box 1 and Figure 1d). Assume a host trait decomposes as

$$
z = g + m,\tag{5}
$$

 where *g* is the additive host genetic effect, and *m* is the additive effect of host microbiome composition. To account f_{162} for microbiome transmission directly from parents to offspring, suppose the offspring microbiome m' is given by

$$
m' = \ell m + (1 - \ell)\xi + \delta,\tag{6}
$$

¹⁶³ where ℓ is the proportion of the offspring microbiome inherited from its parent so that $(1-\ell)$ is the proportion acquired from the environment and unrelated hosts, *ξ* corresponds to the microbiome composition averaged across 165 the environment and unrelated hosts, and δ is an ontogenetic differential of the microbiome that is independent of *m*. Using this simple analytical model, we can measure the microbiome's contribution to a maternal effect by quantifying maternal effects as the partial regression coefficient of offspring trait on maternal trait, holding genetic $_{168}$ variation constant^{[38](#page-13-6)}.

 To measure the microbiome mediated maternal effect in the above model, we write the variance of the trait *z* as P, M the component of P explained by microbiome variation (i.e., the variance of m), and ω the maternal effect. 171 The maternal effect can then be expressed as

$$
\omega = \text{Cov}_{g,g'}(z, z')/P = \text{Cov}_{g,g'}(g + m, g' + m')/P = \text{Cov}(m, \ell m)/P = \ell M/P,
$$
\n(7)

 n_{172} where the subscript g, g' is a reminder that we are holding genetic variation constant in both the parent and offspring. This demonstrates that host-associated microbiomes can be modeled as maternal characters, but the application is limited to the analysis of microbiome inheritance resulting from strict parent-offspring transmission.

Multilevel Selection

 This last framework applies to microbe-host associations in which microbiome-mediated traits are heritable and subject to natural selection. Heritable variation in both host and microbe fitness is essential for the host-microbe system to be an evolutionary individual that responds to natural selection (Box 1). In other words, Lewontin's ₁₇₉ conditions^{[55](#page-14-5)} must be met. Establishing host-microbe evolutionary individuality requires the collective reproduction of hosts and microbes, either through vertical microbe transmission or through horizontal transmission mechanisms μ_{181} that link host-microbe genomes and fitnesses^{[56](#page-14-6)}. Given these restrictive conditions, most host-microbe associations are not considered evolutionary individuals (we note that terms such as demibiont have been coined to describe associations that exhibit less than perfect collective reproduction) 57 .

- ¹⁸⁴ The host-microbe systems with collective reproduction can experience natural selection as individuals, as kin, and
- ¹⁸⁵ as groups of unrelated organisms^{[58](#page-14-8)}, all explicitly captured by multilevel selection (MLS) models Multilevel selection 1
- ¹⁸⁶ (MLS1) models deal with selection among kin or relatives. For instance, populations of (nearly) clonal host-associated
- ¹⁸⁷ microbes experience inclusive fitness if their activities ensure transmission of their relatives to new host generations.
- ¹⁸⁸ Groups of individuals (e.g., hosts with all their microbes) operating as a collective are modeled by multilevel selection
- ¹⁸⁹ 2 (MLS2). Recently, multilevel selection 3 (MLS3) was proposed, merging MLS1 and MLS2 to consider the joint
- ¹⁹⁰ influence of microbe-inclusive fitness and group selection on emerging host-microbe trait evolution^{[33](#page-13-1)}. Multilevel
- ¹⁹¹ selection models have been successful in describing the major evolutionary transitions in individuality that resulted ¹⁹² in endosymbiont-derived organelles, multicellularity, and the germline^{[59–](#page-14-9)[62](#page-14-10)}.
- ¹⁹³ By testing different MLS models, we can ascertain whether individual, kin, or group selection, or a combination, ¹⁹⁴ is the predominant force in the evolution of emergent host-microbe phenotypes. As for MLS1, because individual ¹⁹⁵ microbes are subdivided into groups among hosts, their selection coefficient can be split into two components: 196 selection within-hosts and selection among-hosts^{[63](#page-14-11)}. For a population of individuals (i.e., microbes) to experience 197 selection that can be modeled with MLS1, the relatedness between individuals (r) and the indirect fitness benefit from their interaction (b) must exceed the cost to individual fitness (c), expressed as $r > c/b^{64-66}$ $r > c/b^{64-66}$ $r > c/b^{64-66}$. However, this ¹⁹⁹ equation, known as Hamilton's rule, is only valid for close relatives experiencing strong additive selection, not $_{200}$ capturing more complicated models of selection^{[67](#page-15-2)}.
- ²⁰¹ Instead, a quantitative genetic model of direct and indirect fitness effects allows for simultaneous consideration of 202 MLS1 and MLS2. Here, phenotype P of individual $i(P_i)$, interpreted as either the host or the microbe, can be 203 written as^{68} as^{68} as^{68} :

$$
P_i = A_{D,i} + E_{D,i} + \sum_{j \neq i}^{n} A_{s,j} + \sum_{j \neq j}^{n} E_{s,j},
$$
\n(8)

where $A_{D,i}$ is the direct heritable impact of individual *i* on its own phenotype, whereas $A_{S,i}$ is its indirect heritable $_{205}$ impact of other host or microbe associates in the community on the focal individual. $E_{D,i}$ is the direct environmental ²⁰⁶ impact on individual *i*, and $E_{s,j}$ is the environmental impact on the indirect effects of individual *i* on the community ²⁰⁷ associates.

208 At the population level, the selection for individual $i(C_i)$ can be expressed to depend both on focal phenotype ²⁰⁹ P_i and on all other phenotypes in the group^{[68](#page-15-3)}:

$$
C_i = P_i + g \sum_{j \neq i}^{n} P_j. \tag{9}
$$

210 Here, *g* is the degree to which group selection acts (when $q = 0$, selection acts on individuals only; when $q = 1$, selection acts on the total group-level phenotype). This approach allows us to connect levels of selection occurring simultaneously across distinct scales of biological organization. In particular, this framework can be adapted for the study of diverse microbe-host systems by considering selection on phenotypes expressed by the host, the microbes, and the joint actions of the host and their associated microbes. The associative phenotypes that feedback indirectly on these focal individuals can be experienced by the host, the microbes, or the host and their associated microbes by varying the degree of relatedness among associates when modeling the group's response to selection. More than ²¹⁷ two levels of organization can be considered by generalizing equation $(9)^{68}$ $(9)^{68}$ $(9)^{68}$ $(9)^{68}$, allowing for structured populations of host-microbe groups and within-host microbe groups. Future work is needed to simultaneously model individual-level and emergent group-level phenotypes.

Discussion

Limitations of Existing Frameworks

 Each of the discussed pre-existing frameworks is useful for understanding specific cases of host-microbiome evolution. At the same time, some fundamental properties of host-microbiome systems necessitate expanding these frameworks. Niche construction has clear applications for understanding the relationship between host and environmental microbiomes, and consequential selection of host-microbiome associations. Modification of the social environment via microbiome transmission may be considered a form of niche construction. While niche construction does not focus on nuances in microbiome transmission, the frameworks of indirect genetic effects (IGE) and maternal effects (ME) are particularly useful for understanding the evolutionary consequences of such social and parent-offspring transmission, respectively. Combining IGE and ME could account for mixed transmission modes, consisting of social and parent-offspring transmission. However, IGE and ME treat the consequences of microbiome transmission as fixed effects, limiting their ability to incorporate microbiome community dynamics, host immune response, and variation of transmission other than what is explained by trait covariances. Hence, further expansions are needed to incorporate specific biological details relevant for understanding host-microbiome dynamics.

 Multilevel selection is useful as an overarching framework for understanding selection on complex host-microbiome assemblages, but this approach has important limitations as well. The evolutionary mechanisms that enable evolutionary transitions in individuality to occur are still under debate and models are in development. For example, it is still under debate whether MLS1 and MLS2 are inequivalent because the Price equation has supported their ²³⁸ equivalency since the 1970s^{[69–](#page-15-4)[71](#page-15-5)}. Further, while some group-level phenotypes may have relevance at the individual level, not all will. For example, metabolic complementation between aphids and their Buchnera endosymbionts for amino acid synthesis is selected for in the host-microbe assemblage, but not in the individual organisms, because the individual aphid and Buchnera genomes lack genes to complete the pathway.

Opportunities for Developing Novel Frameworks

²⁴³ Beyond the four frameworks discussed here, host-microbiome systems provide a number of exciting opportunities for extending and developing theoretical approaches to describe features that are not sufficiently captured by our ²⁴⁵ focal frameworks. For example, microbiome composition varies over the course of a host's life^{[72,](#page-15-6)[73](#page-15-7)}. Here, theory ²⁴⁶ on ontogenetic changes in maternal and genetic contributions to host phenotypic variation^{[74](#page-15-8)} may provide useful ²⁴⁷ insights to microbiome changes during host development and its implications for responses to selection. For instance, ²⁴⁸ a maternal signal in microbiome composition that diminishes with host age^{[75](#page-15-9)} could be captured by a negative relationship between host age and the contribution of maternal effects. Further, host microbiome composition is shaped by fluctuating microbial abundances resulting in within-host ecological interactions, but such interactions are ignored in the non-genetic inheritance and group selection frameworks that we discussed.

 These frameworks can be extended to account for such ecological details by integrating models of microbial community dynamics into host trait architecture. These biotic interactions are even further complicated by the existence of multiple trophic levels within a microbiome community (e.g., interactions of bacteria with phages or $_{255}$ predatory bacteria)^{[76](#page-15-10)[–78](#page-15-11)}. Microbes can also show context-dependence in their contributions to host fitness, where they act as mutualists in one environment, while as pathogens in another^{[79–](#page-15-12)[82](#page-16-0)}. Theory on fluctuating selection^{[83](#page-16-1)} could be used to assess host-microbiome evolution in such a case.

258 Additionally, there are many opportunities to expand theory of multilevel selection. For example, stochastic simulation of multilevel selection processes could enable the development of new MLS models and theories to pinpoint the conditions required for cooperation among microbes and hosts to evolve. Spatial structure shapes ²⁶¹ the formation of groups that can respond to selection pressures^{[63,](#page-14-11) [70,](#page-15-13) [84,](#page-16-2) [85](#page-16-3)}, emphasizing the need to incorporate environmental parameters in MLS models. Genetic models for mapping trait selection onto the complex genetic ₂₆₃ basis for that trait^{[86](#page-16-4)} could be used to map group-level phenotypic selection onto individual genotypes. Incorporating ²⁶⁴ genetic parameters into MLS models will enable the use of genome-wide datasets. Further, the impact of host versus symbiont population size and generation time on the rates of co-evolution should be considered $33,87$ $33,87$.

 Lastly, it may be useful to consider microbiome mediated host traits as a form of phenotypic plasticity. However, the framework of phenotypic plasticity typically considers a single global environmental factor driving the plastic response of a population, and one that is not transmissible. Moreover, microbiome-mediated plasticity can act ²⁶⁹ at different levels and time scales^{[88](#page-16-6)}. For example, a new environmental challenge can be accommodated fastest by ecological changes in microbiome community composition, followed by evolutionary genetic changes in single microbial lineages. Such microbiome plasticity can further selectively favor hosts that either select the beneficial microbes from the environment or ensure their vertical transmission, as a kind of microbiome-mediated Baldwin ₂₇₃ effect^{[88](#page-16-6)}. Hence, application of the phenotypic plasticity framework to microbiome mediated traits would need to be extended to account for environmental factors taking values unique to each host individual, and possibly transmissible between hosts.

²⁷⁶ **The Need for "Empirically Friendly" Theory**

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²⁷⁷ We feel that it is important to recognize that existing theoretical frameworks were often developed with biological systems in mind other than host-microbiome systems. Not only has this resulted in frameworks that lack key aspects of the biology of host-microbiome systems (as we discuss in the previous section), but it has also limited the application of these frameworks to host-microbiome systems in the laboratory and the field. To make these frameworks maximally useful, it is important that the validity of the underlying assumptions of these frameworks is determined empirically. It is also crucial that these frameworks are constructed in a way that makes their predictions empirically testable, given the technical limitations of empirical microbiome research. For example, for many host-microbiome systems, empiricists are limited to surveying relative abundance or presence/absence of microbial taxa, and for theory to be maximally useful it must generate predictions for these microbiome attributes. Tailoring theory in this way will likely require direct collaborations between theoretical and empirical microbiome scientists, to enable an iterative refinement of theory with information from actual host-microbiome systems. This ²⁸⁸ requires suitable host systems, such as the water flea *Daphnia* (Box 2), the nematode *Caenorhabditis elegans*^{[89,](#page-16-7)[90](#page-16-8)}, the zebrafish *Danio rerio*^{[91](#page-16-9)}, or insect-*Wolbachia* associations^{[92](#page-16-10)} as examples.

Box 2: Empirical studies of the theoretical frameworks using the water flea host system as an example

The water flea *Daphnia* (Figure B2), a freshwater crustacean, is a model organism in many biological fields, such as ecotoxicology^{[93](#page-16-11)}, epidemiology^{[94](#page-16-12)}, and evo-evolutionary dynamics^{[95](#page-16-13)}, and is investigated in different settings, ranging from controlled laboratory experiments, to mesocosms, to natural field observations. *Daphnia* are also increasingly being used to study host-microbiome interactions 96 . Previous studies have revealed various aspects of the *Daphnia*-microbiome system that make this system uniquely promising to parameterize and test theoretical frameworks on the evolution of host-microbiome interactions.

First, while gut-associated microbial communities in *Daphnia* are relatively simple with a few core members, these communities are clearly distinct from their surrounding aquatic microbial communities^{[97](#page-17-1)}, suggesting a role of selective processes at play. Second, *Daphnia* microbiomes are impacted both by the environmental conditions (e.g., temperature^{[98](#page-17-2)}) and by host genotype^{[99](#page-17-3)}. Third, the microbiome of *Daphnia* is related to host fitness: associations between microbiome composition and various life history traits have been found $100, 101$ $100, 101$ $100, 101$, and microbiome-mediated plasticity may help adjust hosts to their environment^{[102](#page-17-6)}. Finally, both horizontal and vertical transmission shape $Daphnia$ microbiome composition^{[103,](#page-17-7) [104](#page-17-8)}.

Practically, *Daphnia* are easy to culture in the lab, and clonal lineages can be established from hatching sexually produced resting eggs (which remain viable in the sediment for decades). Under favorable conditions, asexual reproduction can be ensured, enabling a high amount of control on genotypic variation. Further, due to their fast life-cycle, it is straightforward to perform experimental evolution on populations that can easily consist of a few hundreds individuals. Lastly, using *Daphnia* as a host system enables causal inferences on the role of the microbiome for host fitness, both by rearing germ-free hosts^{[101](#page-17-5)} and by microbiome transplants^{[105](#page-17-9)}.

Altogether, this suggests a relatively straightforward integration of empirical data with evolutionary theory, in order to understand host-microbiome evolution. For example, *Daphnia*'s asexual reproduction facilitates the quantification of parent-offspring microbiome transmission as a maternal character, as the contribution of host genetics is known to be constant across generations. Also, *Daphnia* are primary consumers and a keystone freshwater species, and have been shown to mediate their surrounding aquatic microbial communities by grazing[28](#page-12-9). This is a clear example of niche construction as an environmental modification (Figure 1a), but may also establish a mode of ecological inheritance whenever the modified environment shapes selection on *Daphnia* genetic variation (Figure 1b).

Figure B2: Photo of the water flea *Daphnia*. Photo credit: Dr. Marjolein Bruijning.

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²⁹² **Conclusion**

 In this perspective, we presented four frameworks developed in the fields of evolutionary biology that help to generate new insights into host-microbiome evolution. In order to capture the biological diversity of such host-microbe systems and produce empirically testable predictions, these frameworks require thoughtful expansion and in some cases the development of novel theory, in close collaboration with empirical microbiome scientists. We envision that the initial result will be a mosaic of theoretical frameworks, each tuned to the set of processes considered and questions asked, with the initial goal of clarifying concepts. Such a mosaic could eventually lead to the identification of general principles underlying the interactions between microbes and their animal and plant hosts, greatly expanding our understanding of the evolutionary consequences of the host-microbe associations omnipresent across the tree of life.

References

- **1.** Cox, T. O., Lundgren, P., Nath, K. & Thaiss, C. A. Metabolic control by the microbiome. *Genome Medicine* **14**, DOI: <10.1186/s13073-022-01092-0> (2022). Publisher: Springer Science and Business Media LLC.
- **2.** Buffie, C. G. *et al.* Precision microbiome reconstitution restores bile acid mediated resistance to Clostridium difficile. *Nature* **517**, 205–208, DOI: <10.1038/nature13828> (2014). Publisher: Springer Science and Business Media LLC.
- **3.** Hooper, L. V., Littman, D. R. & Macpherson, A. J. Interactions between the microbiota and the immune system. *Science* **336**, 1268–1273, DOI: <10.1126/science.1223490> (2012). Publisher: American Association for the Advancement of Science (AAAS).
- **4.** Matsuoka, K. & Kanai, T. The gut microbiota and inflammatory bowel disease. *Semin. Immunopathol.* **37**, 47–55, DOI: <10.1007/s00281-014-0454-4> (2014). Publisher: Springer Science and Business Media LLC.
- **5.** Bercik, P. *et al.* The intestinal microbiota affect central levels of brain-derived neurotropic factor and behavior in mice. *Gastroenterology* **141**, 599–609.e3, DOI: <10.1053/j.gastro.2011.04.052> (2011). Publisher: Elsevier BV.
- **6.** Wilkinson, J. E. *et al.* A framework for microbiome science in public health. *Nat. Medicine* **27**, 766–774, DOI: <10.1038/s41591-021-01258-0> (2021). Publisher: Springer Science and Business Media LLC.
- **7.** Singh, B. K., Trivedi, P., Egidi, E., Macdonald, C. A. & Delgado-Baquerizo, M. Crop microbiome and sustainable agriculture. *Nat. Rev. Microbiol.* **18**, 601–602, DOI: <10.1038/s41579-020-00446-y> (2020). Publisher: Springer Science and Business Media LLC.
- **8.** Biggs, E., Taylor, M. W. & Middleton, D. M. R. L. Beyond the theory: From holobiont concept to microbiome engineering. *Environ. Microbiol.* **25**, 832–835, DOI: <10.1111/1462-2920.16308> (2023). Publisher: Wiley.
- **9.** Silverstein, M. R., Segrè, D. & Bhatnagar, J. M. Environmental microbiome engineering for the mitigation of climate change. *Glob. Chang. Biol.* **29**, 2050–2066, DOI: <10.1111/gcb.16609> (2023). Publisher: Wiley.
- **10.** Zilber-Rosenberg, I. & Rosenberg, E. Role of microorganisms in the evolution of animals and plants: the hologenome theory of evolution. *FEMS Microbiol. Rev.* **32**, 723–735, DOI: <10.1111/j.1574-6976.2008.00123.x> (2008). Publisher: Oxford University Press (OUP).
- **11.** Aldana, M. & Robeva, R. New challenges in systems biology: Understanding the holobiont. *Front. Physiol.* **12**, DOI: <10.3389/fphys.2021.662878> (2021). Publisher: Frontiers Media SA.
- **12.** Bordenstein, S. R. & Theis, K. R. Host biology in light of the microbiome: Ten principles of holobionts and hologenomes. *PLOS Biol.* **13**, e1002226, DOI: <10.1371/journal.pbio.1002226> (2015). Publisher: Public Library of Science (PLoS).
- **13.** Theis, K. R. *et al.* Getting the hologenome concept right: an eco-evolutionary framework for hosts and their microbiomes. *mSystems* **1**, DOI: <10.1128/msystems.00028-16> (2016). Publisher: American Society for Microbiology.
- **14.** McFall-Ngai, M. *et al.* Animals in a bacterial world, a new imperative for the life sciences. *Proc. Natl. Acad. Sci.* **110**, 3229–3236, DOI: <10.1073/pnas.1218525110> (2013). Publisher: Proceedings of the National Academy of Sciences.
- **15.** Rosenberg, E. & Zilber-Rosenberg, I. The hologenome concept of evolution after 10 years. *Microbiome* **6**, DOI: <10.1186/s40168-018-0457-9> (2018). Publisher: Springer Science and Business Media LLC.
- **16.** Morris, J. J. What is the hologenome concept of evolution? *F1000Research* **7**, 1664, DOI: [10.12688/](10.12688/f1000research.14385.1) [f1000research.14385.1](10.12688/f1000research.14385.1) (2018). Publisher: F1000 Research Ltd.
- **17.** Moran, N. A. & Sloan, D. B. The hologenome concept: Helpful or hollow? *PLOS Biol.* **13**, e1002311, DOI: <10.1371/journal.pbio.1002311> (2015). Publisher: Public Library of Science (PLoS).
- **18.** Henry, L. P., Bruijning, M., Forsberg, S. K. G. & Ayroles, J. F. The microbiome extends host evolutionary potential. *Nat. Commun.* **12**, DOI: <10.1038/s41467-021-25315-x> (2021). Publisher: Springer Science and Business Media LLC.
- **19.** Douglas, A. E. & Werren, J. H. Holes in the hologenome: Why host-microbe symbioses are not holobionts. *mBio* **7**, DOI: <10.1128/mbio.02099-15> (2016). Publisher: American Society for Microbiology.
- **20.** Egan, S., Fukatsu, T. & Francino, M. P. Opportunities and challenges to microbial symbiosis research in the microbiome era. *Front. Microbiol.* **11**, DOI: <10.3389/fmicb.2020.01150> (2020). Publisher: Frontiers Media SA.
- **21.** Zeng, Q., Sukumaran, J., Wu, S. & Rodrigo, A. Neutral models of microbiome evolution. *PLOS Comput. Biol.* **11**, e1004365, DOI: <10.1371/journal.pcbi.1004365> (2015). Publisher: Public Library of Science (PLoS).
- **22.** van Vliet, S. & Doebeli, M. The role of multilevel selection in host microbiome evolution. *Proc. Natl. Acad. Sci.* **116**, 20591–20597, DOI: <10.1073/pnas.1909790116> (2019). Publisher: Proceedings of the National Academy of Sciences.
- **23.** Koga, R., Meng, X.-Y., Tsuchida, T. & Fukatsu, T. Cellular mechanism for selective vertical transmission of an obligate insect symbiont at the bacteriocyte–embryo interface. *Proc. Natl. Acad. Sci.* **109**, DOI: <10.1073/pnas.1119212109> (2012). Publisher: Proceedings of the National Academy of Sciences.
- **24.** Sauer, C., Dudaczek, D., Hölldobler, B. & Gross, R. Tissue localization of the endosymbiotic bacterium " candidatus blochmannia floridanus" in adults and larvae of the carpenter ant camponotus floridanus. *Appl. Environ. Microbiol.* **68**, 4187–4193, DOI: <10.1128/aem.68.9.4187-4193.2002> (2002). Publisher: American Society for Microbiology.
- **25.** Roughgarden, J., Gilbert, S. F., Rosenberg, E., Zilber-Rosenberg, I. & Lloyd, E. A. Holobionts as units of selection and a model of their population dynamics and evolution. *Biol. Theory* **13**, 44–65, DOI: [10.1007/](10.1007/s13752-017-0287-1) [s13752-017-0287-1](10.1007/s13752-017-0287-1) (2017). Publisher: Springer Science and Business Media LLC.
- **26.** Fukatsu, T. & Hosokawa, T. Capsule-transmitted gut symbiotic bacterium of the japanese common plataspid stinkbug, megacopta punctatissima. *Appl. Environ. Microbiol.* **68**, 389–396, DOI: [10.1128/aem.68.1.389-396.](10.1128/aem.68.1.389-396.2002) [2002](10.1128/aem.68.1.389-396.2002) (2002). Publisher: American Society for Microbiology.
- **27.** Dominguez-Bello, M. G. *et al.* Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc. Natl. Acad. Sci.* **107**, 11971–11975, DOI: <10.1073/pnas.1002601107> (2010). Publisher: Proceedings of the National Academy of Sciences.
- **28.** Degans, H., Zöllner, E., Gucht, K., Meester, L. & Jürgens, K. Rapid Daphnia-mediated changes in microbial community structure: an experimental study. *FEMS Microbiol. Ecol.* **42**, 137–149, DOI: <10.1111/j.1574-6941.2002.tb01003.x> (2002). Publisher: Oxford University Press (OUP).
- **29.** Tung, J. *et al.* Social networks predict gut microbiome composition in wild baboons. *eLife* **4**, DOI: [10.7554/](10.7554/elife.05224) ³⁷⁵ [elife.05224](10.7554/elife.05224) (2015). Publisher: eLife Sciences Publications, Ltd.
- **30.** Weinstein, S. B. *et al.* Microbiome stability and structure is governed by host phylogeny over diet and geography in woodrats (Neotoma spp.). *Proc. Natl. Acad. Sci.* **118**, DOI: <10.1073/pnas.2108787118> (2021). Publisher: Proceedings of the National Academy of Sciences.
- **31.** Nyholm, S. V. & McFall-Ngai, M. The winnowing: establishing the squid–vibrio symbiosis. *Nat. Rev. Microbiol.* **2**, 632–642, DOI: <10.1038/nrmicro957> (2004). Publisher: Springer Science and Business Media LLC.
- **32.** Kikuchi, Y., Hosokawa, T. & Fukatsu, T. Insect-microbe mutualism without vertical transmission: a stinkbug acquires a beneficial gut symbiont from the environment every generation. *Appl. Environ. Microbiol.* **73**, 4308–4316, DOI: <10.1128/aem.00067-07> (2007). Publisher: American Society for Microbiology.
- **33.** Roughgarden, J. Holobiont evolution: Population theory for the hologenome. *The Am. Nat.* **201**, 763–778, DOI: <10.1086/723782> (2023). Publisher: University of Chicago Press.
- **34.** Morris, A. H. & Bohannan, B. J. M. Estimates of microbiome heritability across hosts. *Nat. Microbiol.* DOI: <10.1038/s41564-024-01865-w> (2024). Publisher: Springer Science and Business Media LLC.
- **35.** Leibold, M. A. *et al.* The metacommunity concept: a framework for multi-scale community ecology. *Ecol. Lett.* **7**, 601–613, DOI: <10.1111/j.1461-0248.2004.00608.x> (2004). Publisher: Wiley.
- **36.** Costello, E. K., Stagaman, K., Dethlefsen, L., Bohannan, B. J. M. & Relman, D. A. The application of ecological theory toward an understanding of the human microbiome. *Science* **336**, 1255–1262, DOI: <10.1126/science.1224203> (2012). Publisher: American Association for the Advancement of Science (AAAS).
- **37.** Miller, E. T., Svanbäck, R. & Bohannan, B. J. Microbiomes as metacommunities: Understanding host-associated microbes through metacommunity ecology. *Trends Ecol. & Evol.* **33**, 926–935, DOI: <10.1016/j.tree.2018.09.002> (2018). Publisher: Elsevier BV.
- **38.** Kirkpatrick, M. & Lande, R. The evolution of maternal characters. *Evol. international journal organic evolution* **43**, 485–503, DOI: <10.1111/j.1558-5646.1989.tb04247.x> (1989). Publisher: Wiley.
- **39.** Odling-Smee, F., Lala, K. & Feldman, M. *Niche construction: The neglected process in evolution*. Monographs in population biology (Princeton University Press, 2013). Tex.lccn: 2002031747.
- **40.** Wolf, J. B., Brodie III, E. D. & Moore, A. J. Interacting phenotypes and the evolutionary process. II. Selection resulting from social interactions. *The Am. Nat.* **153**, 254–266, DOI: <10.1086/303168> (1999). Publisher: University of Chicago Press.
- **41.** Bonduriansky, R. & Day, T. Nongenetic inheritance and its evolutionary implications. *Annu. Rev. Ecol. Evol. Syst.* **40**, 103–125, DOI: <10.1146/annurev.ecolsys.39.110707.173441> (2009). Publisher: Annual Reviews.
- **42.** Odling-Smee, J., Erwin, D. H., Palkovacs, E. P., Feldman, M. W. & Laland, K. N. Niche construction theory: a practical guide for ecologists. *The Q. Rev. Biol.* **88**, 3–28, DOI: <10.1086/669266> (2013). Publisher: University of Chicago Press.
- **43.** Campos-Cerda, F. & Bohannan, B. J. The nidobiome: a framework for understanding microbiome assembly in neonates. *Trends Ecol. & Evol.* **35**, 573–582, DOI: <10.1016/j.tree.2020.03.007> (2020). Publisher: Elsevier BV.
- **44.** Zhu, B. *et al.* Rhizosphere priming effects on soil carbon and nitrogen mineralization. *Soil Biol. Biochem.* **76**, 183–192, DOI: <10.1016/j.soilbio.2014.04.033> (2014). Publisher: Elsevier BV.
- **45.** Mueller, U. G. & Gerardo, N. Fungus-farming insects: Multiple origins and diverse evolutionary histories. *Proc. Natl. Acad. Sci.* **99**, 15247–15249, DOI: <10.1073/pnas.242594799> (2002). Publisher: Proceedings of the National Academy of Sciences.
- **46.** Finzi, A. C. *et al.* Rhizosphere processes are quantitatively important components of terrestrial carbon and nutrient cycles. *Glob. Chang. Biol.* **21**, 2082–2094, DOI: <10.1111/gcb.12816> (2015). Publisher: Wiley.
- **47.** Mendes, R. *et al.* Deciphering the rhizosphere microbiome for disease-suppressive bacteria. *Science* **332**, 1097–1100, DOI: <10.1126/science.1203980> (2011). Publisher: American Association for the Advancement of Science (AAAS).
- **48.** Bruijning, M., Henry, L. P., Forsberg, S. K. G., Metcalf, C. J. E. & Ayroles, J. F. Natural selection for imprecise vertical transmission in host–microbiota systems. *Nat. Ecol. & Evol.* **6**, 77–87, DOI: <10.1038/s41559-021-01593-y> (2021). Publisher: Springer Science and Business Media LLC.
- **49.** Linksvayer, T. A. Direct, maternal, and sibsocial genetic effects on individual and colony traits in an ant. *Evol. international journal organic evolution* **60**, 2552, DOI: <10.1554/06-011.1> (2006). Publisher: Wiley.
- **50.** Sokolowski, M. B. Honey bee colony aggression and indirect genetic effects. *Proc. Natl. Acad. Sci.* **117**, 18148–18150, DOI: <10.1073/pnas.2012366117> (2020). Publisher: Proceedings of the National Academy of Sciences.
- **51.** Piekarski, P. K., Valdés-Rodríguez, S. & Kronauer, D. J. C. Conditional indirect genetic effects of caregivers on brood in the clonal raider ant. *Behav. Ecol.* **34**, 642–652, DOI: <10.1093/beheco/arad033> (2023). Publisher: Oxford University Press (OUP).
- **52.** Carter, M. J., Wilson, A. J., Moore, A. J. & Royle, N. J. The role of indirect genetic effects in the evolution of interacting reproductive behaviors in the burying beetle,Nicrophorus vespilloides. *Ecol. Evol.* **9**, 998–1009, DOI: <10.1002/ece3.4731> (2019). Publisher: Wiley.
- **53.** Moore, A. J., Brodie, E. D. & Wolf, J. B. Interacting phenotypes and the evolutionary process: I. Direct and indirect genetic effects of social interactions. *Evol. international journal organic evolution* **51**, 1352–1362, DOI: <10.1111/j.1558-5646.1997.tb01458.x> (1997). Publisher: Wiley.
- **54.** Mousseau, T. A. & Fox, C. W. *Maternal effects as adaptations* (Oxford University Press, 1998).
- **55.** Lewontin, R. C. The units of selection. *Annu. Rev. Ecol. Syst.* **1**, 1–18, DOI: [10.1146/annurev.es.01.110170.](10.1146/annurev.es.01.110170.000245) [000245](10.1146/annurev.es.01.110170.000245) (1970). Publisher: Annual Reviews.
- **56.** Stencel, A. & Wloch-Salamon, D. M. Some theoretical insights into the hologenome theory of evolution and the role of microbes in speciation. *Theory Biosci.* **137**, 197–206, DOI: <10.1007/s12064-018-0268-3> (2018). Publisher: Springer Science and Business Media LLC.
- **57.** Suárez, J. & Stencel, A. A part-dependent account of biological individuality: why holobionts are individuals and ecosystems simultaneously. *Biol. Rev.* **95**, 1308–1324, DOI: <10.1111/brv.12610> (2020). Publisher: Wiley.
- **58.** Provorov, N. A. & Vorobyov, N. I. Macroevolution of symbiosis as self-organization of superspecies system controlled by natural selection. *Biol. Bull. Rev.* **3**, 274–285, DOI: <10.1134/s2079086413040063> (2013). Publisher: Pleiades Publishing Ltd.
- **59.** Zachar, I. & Boza, G. Endosymbiosis before eukaryotes: mitochondrial establishment in protoeukaryotes. *Cell. Mol. Life Sci.* **77**, 3503–3523, DOI: <10.1007/s00018-020-03462-6> (2020). Publisher: Springer Science and Business Media LLC.
- **60.** Stencel, A. & Suárez, J. Do somatic cells really sacrifice themselves? Why an appeal to coercion may be a helpful strategy in explaining the evolution of multicellularity. *Biol. Theory* **16**, 102–113, DOI: [10.1007/](10.1007/s13752-021-00376-9) [s13752-021-00376-9](10.1007/s13752-021-00376-9) (2021). Publisher: Springer Science and Business Media LLC.
- **61.** Niklas, K. J. The evolutionary-developmental origins of multicellularity. *Am. J. Bot.* **101**, 6–25, DOI: <10.3732/ajb.1300314> (2014). Publisher: Wiley.
- **62.** Brutovský, B. Scales of cancer evolution: Selfish genome or cooperating cells? *Cancers* **14**, 3253, DOI: <10.3390/cancers14133253> (2022). Publisher: MDPI AG.
- **63.** Hermsen, R. Emergent multilevel selection in a simple spatial model of the evolution of altruism. *PLOS Comput. Biol.* **18**, e1010612, DOI: <10.1371/journal.pcbi.1010612> (2022). Publisher: Public Library of Science (PLoS).
- **64.** Hamilton, W. The genetical evolution of social behaviour. I. *J. Theor. Biol.* **7**, 1–16, DOI: [10.1016/0022-5193\(64\)](10.1016/0022-5193(64)90038-4) [90038-4](10.1016/0022-5193(64)90038-4) (1964). Publisher: Elsevier BV.
- **65.** Yokoyama, S. & Felsenstein, J. A model of kin selection for an altruistic trait considered as a quantitative character. *Proc. Natl. Acad. Sci.* **75**, 420–422, DOI: <10.1073/pnas.75.1.420> (1978). Publisher: Proceedings of the National Academy of Sciences.
- **66.** GARDNER, A., WEST, S. A. & WILD, G. The genetical theory of kin selection. *J. Evol. Biol.* **24**, 1020–1043, DOI: <10.1111/j.1420-9101.2011.02236.x> (2011). Publisher: Oxford University Press (OUP).
- **67.** van Veelen, M. The group selection–inclusive fitness equivalence claim: not true and not relevant. *Evol. Hum. Sci.* **2**, DOI: <10.1017/ehs.2020.9> (2020). Publisher: Cambridge University Press (CUP).
- **68.** Bijma, P., Muir, W. M. & Van Arendonk, J. A. M. Multilevel selection 1: Quantitative genetics of inheritance and response to selection. *Genetics* **175**, 277–288, DOI: <10.1534/genetics.106.062711> (2007). Publisher: Oxford University Press (OUP).
- **69.** van Veelen, M., García, J., Sabelis, M. W. & Egas, M. Group selection and inclusive fitness are not equivalent; the Price equation vs. models and statistics. *J. Theor. Biol.* **299**, 64–80, DOI: <10.1016/j.jtbi.2011.07.025> (2012). Publisher: Elsevier BV.
- **70.** Simon, B., Fletcher, J. A. & Doebeli, M. Towards a general theory of group selection. *Evol. international journal organic evolution* **67**, 1561–1572, DOI: <10.1111/j.1558-5646.2012.01835.x> (2012). Publisher: Wiley.
- **71.** Lehtonen, J. Multilevel selection in kin selection language. *Trends Ecol. & Evol.* **31**, 752–762, DOI: [10.1016/j.](10.1016/j.tree.2016.07.006) [tree.2016.07.006](10.1016/j.tree.2016.07.006) (2016). Publisher: Elsevier BV.
- **72.** Moore, R. E. & Townsend, S. D. Temporal development of the infant gut microbiome. *Open Biol.* **9**, 190128, DOI: <10.1098/rsob.190128> (2019). Publisher: The Royal Society.
- **73.** Zimmermann, J. *et al.* Gut-associated functions are favored during microbiome assembly across a major part of C. elegans life. *mBio* **15**, DOI: <10.1128/mbio.00012-24> (2024). Publisher: American Society for Microbiology.
- **74.** Wilson, A. J. & Réale, D. Ontogeny of additive and maternal genetic effects: Lessons from domestic mammals. *The Am. Nat.* **167**, E23–E38, DOI: <10.1086/498138> (2006). Publisher: University of Chicago Press.
- **75.** Ferretti, P. *et al.* Mother-to-infant microbial transmission from different body sites shapes the developing infant gut microbiome. *Cell Host & Microbe* **24**, 133–145.e5, DOI: <10.1016/j.chom.2018.06.005> (2018). Publisher: Elsevier BV.
- **76.** Kirsch, J. M. *et al.* Bacteriophage-bacteria interactions in the gut: From invertebrates to mammals. *Annu. Rev. Virol.* **8**, 95–113, DOI: <10.1146/annurev-virology-091919-101238> (2021). Publisher: Annual Reviews.
- **77.** Shkoporov, A. N., Turkington, C. J. & Hill, C. Mutualistic interplay between bacteriophages and bacteria in the human gut. *Nat. Rev. Microbiol.* **20**, 737–749, DOI: <10.1038/s41579-022-00755-4> (2022). Publisher: Springer Science and Business Media LLC.
- **78.** Johnke, J., Fraune, S., Bosch, T. C. G., Hentschel, U. & Schulenburg, H. Bdellovibrio and like organisms are predictors of microbiome diversity in distinct host groups. *Microb. Ecol.* **79**, 252–257, DOI: [10.1007/](10.1007/s00248-019-01395-7) $\frac{\text{600248-019-01395-7 (2019)}}{1}$
- **79.** VORBURGER, C. & GOUSKOV, A. Only helpful when required: a longevity cost of harbouring defensive symbionts. *J. Evol. Biol.* **24**, 1611–1617, DOI: <10.1111/j.1420-9101.2011.02292.x> (2011). Publisher: Oxford University Press (OUP).
- **80.** Drew, G. C., Stevens, E. J. & King, K. C. Microbial evolution and transitions along the parasite–mutualist continuum. *Nat. Rev. Microbiol.* **19**, 623–638, DOI: <10.1038/s41579-021-00550-7> (2021). Publisher: Springer Science and Business Media LLC.
- **81.** Couret, J., Huynh-Griffin, L., Antolic-Soban, I., Acevedo-Gonzalez, T. S. & Gerardo, N. M. Even obligate symbioses show signs of ecological contingency: Impacts of symbiosis for an invasive stinkbug are mediated by host plant context. *Ecol. Evol.* **9**, 9087–9099, DOI: <10.1002/ece3.5454> (2019). Publisher: Wiley.
- **82.** Griem-Krey, H., Petersen, C., Hamerich, I. K. & Schulenburg, H. The intricate triangular interaction between protective microbe, pathogen and host determines fitness of the metaorganism. *Proc. Royal Soc. B: Biol. Sci.* **290**, DOI: <10.1098/rspb.2023.2193> (2023).
- **83.** Gillespie, J. H. Natural selection with varying selection coefficients – a haploid model. *Genet. Res.* **21**, 115–120, DOI: <10.1017/s001667230001329x> (1973). Publisher: Hindawi Limited.
- **84.** Henriques, G. J. B., van Vliet, S. & Doebeli, M. Multilevel selection favors fragmentation modes that maintain cooperative interactions in multispecies communities. *PLOS Comput. Biol.* **17**, e1008896, DOI: <10.1371/journal.pcbi.1008896> (2021). Publisher: Public Library of Science (PLoS).
- **85.** Doekes, H. M. & Hermsen, R. Multiscale selection in spatially structured populations. *Proc. Royal Soc. B: Biol. Sci.* **291**, DOI: <10.1098/rspb.2023.2559> (2024). Publisher: The Royal Society.
- **86.** Kimura, M. & Crow, J. F. The number of alleles that can be maintained in a finite population. *Genetics* **49**, 725–738, DOI: <10.1093/genetics/49.4.725> (1964).
- **87.** Lynch, M. Mutation pressure, drift, and the pace of molecular coevolution. *Proc. Natl. Acad. Sci.* **120**, DOI: <10.1073/pnas.2306741120> (2023). Publisher: Proceedings of the National Academy of Sciences.
- **88.** Kolodny, O. & Schulenburg, H. Microbiome-mediated plasticity directs host evolution along several distinct time scales. *Philos. Transactions Royal Soc. B: Biol. Sci.* **375**, 20190589, DOI: <10.1098/rstb.2019.0589> (2020).
- **89.** Dirksen, P. *et al.* Cembio - thecaenorhabditis elegansmicrobiome resource. *G3 Genes|Genomes|Genetics* **10**, $3025-3039$, DOI: $10.1534/g3.120.401309$ (2020).
- **90.** Petersen, C. *et al.* Host and microbiome jointly contribute to environmental adaptation. *The ISME J.* **17**, 1953–1965, DOI: <10.1038/s41396-023-01507-9> (2023).
- **91.** Stagaman, K., Sharpton, T. J. & Guillemin, K. Zebrafish microbiome studies make waves. *Lab Animal* **49**, 201–207, DOI: <10.1038/s41684-020-0573-6> (2020).
- **92.** Mirchandani, C. *et al.* Mixed Wolbachia infections resolve rapidly during in vitro evolution. *PLOS Pathog.* **20**, e1012149, DOI: <10.1371/journal.ppat.1012149> (2024). Publisher: Public Library of Science (PLoS).
- **93.** Tkaczyk, A., Bownik, A., Dudka, J., Kowal, K. & Ślaska, B. Daphnia magna model in the toxicity assessment of pharmaceuticals: A review. *Sci. The Total. Environ.* **763**, 143038, DOI: <10.1016/j.scitotenv.2020.143038> (2021). Publisher: Elsevier BV.
- **94.** Ebert, D. *Ecology, epidemiology, and evolution of parasitism in Daphnia* (National Library of Medicine, 2005).
- **95.** Ebert, D. Daphnia as a versatile model system in ecology and evolution. *EvoDevo* **13**, DOI: [10.1186/](10.1186/s13227-022-00199-0) [s13227-022-00199-0](10.1186/s13227-022-00199-0) (2022). Publisher: Springer Science and Business Media LLC.
- **96.** Akbar, S. *et al.* Understanding host-microbiome-environment interactions: Insights from Daphnia as a model organism. *Sci. The Total. Environ.* **808**, 152093, DOI: <10.1016/j.scitotenv.2021.152093> (2022). Publisher: Elsevier BV.
- **97.** Freese, H. M. & Schink, B. Composition and stability of the microbial community inside the digestive tract of the aquatic crustacean daphnia magna. *Microb. Ecol.* **62**, DOI: <10.1007/s00248-011-9886-8> (2011). Publisher: Springer Science and Business Media LLC.
- **98.** Frankel-Bricker, J., Song, M. J., Benner, M. J. & Schaack, S. Variation in the microbiota associated with daphnia magna across genotypes, populations, and temperature. *Microb. Ecol.* **79**, 731–742, DOI: <10.1007/s00248-019-01412-9> (2019). Publisher: Springer Science and Business Media LLC.

 99. Sison-Mangus, M. P., Metzger, C. M. J. A. & Ebert, D. Host genotype-specific microbiota do not influence the susceptibility of D. magna to a bacterial pathogen. *Sci. Reports* **8**, DOI: <10.1038/s41598-018-27681-x> (2018). Publisher: Springer Science and Business Media LLC.

- **100.** Gurung, A., Mukherjee, S., Declercq, M., Souffreau, C. & De Meester, L. Strain-dependent and host genotype–dependent priority effects in gut microbiome assembly affect host fitness in ¡scp¿Daphnia¡/scp¿. *Limnol. Oceanogr.* **69**, 1782–1796, DOI: <10.1002/lno.12614> (2024). Publisher: Wiley.
- **101.** Sison-Mangus, M. P., Mushegian, A. A. & Ebert, D. Water fleas require microbiota for survival, growth and reproduction. *The ISME J.* **9**, 59–67, DOI: <10.1038/ismej.2014.116> (2014). Publisher: Oxford University Press (OUP).
- **102.** Macke, E., Callens, M., De Meester, L. & Decaestecker, E. Host-genotype dependent gut microbiota drives zooplankton tolerance to toxic cyanobacteria. *Nat. Commun.* **8**, DOI: <10.1038/s41467-017-01714-x> (2017). Publisher: Springer Science and Business Media LLC.
- **103.** Callens, M., Watanabe, H., Kato, Y., Miura, J. & Decaestecker, E. Microbiota inoculum composition affects holobiont assembly and host growth in Daphnia. *Microbiome* **6**, DOI: <10.1186/s40168-018-0444-1> (2018). Publisher: Springer Science and Business Media LLC.
- **104.** Mushegian, A. A., Walser, J., Sullam, K. E. & Ebert, D. The microbiota of diapause: How host–microbe associations are formed after dormancy in an aquatic crustacean. *J. Animal Ecol.* **87**, 400–413, DOI: <10.1111/1365-2656.12709> (2017). Publisher: Wiley.

 105. Houwenhuyse, S., Stoks, R., Mukherjee, S. & Decaestecker, E. Locally adapted gut microbiomes mediate host stress tolerance. *The ISME J.* **15**, 2401–2414, DOI: <10.1038/s41396-021-00940-y> (2021). Publisher: Oxford University Press (OUP).

Acknowledgements

 This paper was initiated at the 2023 Symbiosis Theory Workshop. We are grateful to the Gordon and Betty Moore Foundation for their support of this workshop, and to the workshop participants for their stimulating discussion. MB is supported by NWO Veni grant VI.Veni.222.344. BW and BJMB are supported by grant no.10001 from the Gordon and Betty Moore Foundation. BW is further funded through KiTE (i.e., Kiel Training for Excellence) which is part of the European Union's Horizon Europe research and innovation programme under the Marie Sklodowska-Curie grant agreement number No 101081480. BJMB is grateful for the support of the NIH (P01GM125576). BJMB and HS are grateful for financial support from the German Science Foundation within the collaborative research center CRC 1182 on the Origin and Function of Metaorganisms (Mercator fellowship to BJMB; projects A1.1 and A4.3 to HS). HS further thanks the Max-Planck Society for a Max-Planck fellowship.. SLR thanks the NIH (R00GM135583) for financial support.