- 1 **Title:**
- 2 New technology for an ancient fish: A lamprey life cycle modeling tool with an R Shiny
- 3 application
- 4

5 **Authors:**

- 6 Dylan G. E. Gomes^{1*}, Joseph R. Benjamin², Benjamin J. Clemens³, Ralph Lampman⁴, Jason B. Dunham⁵ 7
- 8

9¹ United States Geological Survey, Forest and Rangeland Ecosystem Science Center, Seattle, 10 WA

- ² United States Geological Survey, Forest and Rangeland Ecosystem Science Center, Boise, ID
- ³ Oregon Department of Fish and Wildlife, Corvallis Research Lab, Corvallis, OR
- 13 ⁴ Yakama Nation Fisheries Resource Management Program, Toppenish, WA
- ⁵ United States Geological Survey, Forest and Rangeland Ecosystem Science Center, Corvallis,
- 15 OR
- 16
- 17 *** Correspondence: dgomes@usgs.gov**

Abstract:

 Lampreys (Petromyzontiformes) are an ancient group of fishes with complex life histories. We created a life cycle model that includes an R Shiny interactive web application interface to simulate abundance by life stage. This will allow scientists and managers to connect available demographic information in a framework that can be applied to questions regarding lamprey biology and conservation. We used Pacific lamprey (*Entosphenus tridentatus*) as a case study to highlight the utility of this model. We applied a global sensitivity analysis to explore the importance of individual life stage parameters to overall population size, and to better understand the implications of existing gaps in knowledge. We also provided example analyses of selected management scenarios (dam passage, fish translocations, and hatchery additions) influencing Pacific lamprey in fresh water. These applications illustrate how the model can be applied to inform conservation efforts. This tool will provide new capabilities for users to explore their own questions about lamprey biology and conservation. Simulations can hone hypotheses and predictions, which can then be empirically tested in the real world.

Introduction

 Lampreys are a widespread group of ecologically and culturally important fishes, with many species of conservation concern (Close, Fitzpatrick & Li, 2002; Docker, Hume & Clemens, 2015; Potter et al., 2015; Docker, 2019; Clemens et al., 2021). Lampreys represent an 'ancient' lineage, branching from the ancestors of modern jawed vertebrates between 450 and 550 million years ago (Donoghue, Forey & Aldridge, 2000; Shimeld & Donoghue, 2012; Miyashita et al., 2019), with body forms that have remained mostly unchanged for at least 360 million years (Gess, Coates & Rubidge, 2006; Janvier, 2006). Given their unique evolutionary history, potential losses of lampreys represent a threat to the overall loss of the evolutionary diversity of fishes (IUCN, 2012; Isaac & Pearse, 2018; Gumbs et al., 2023). However, the formal status of many species of lamprey suffers from limited data availability (Clemens et al., 2021). One major difficulty in studying lampreys is the complexity and cryptic nature of their life cycles (Figure 1). For example, lampreys are photophobic and their larval life stage reside within the substrate for years (Dawson et al., 2015). Additionally, lampreys undergo substantial transformations between larval and adult stages, which can include anadromous or adfluvial migration behaviors in large bodies of water, such as oceans and lakes (Potter et al., 2015; Mateus et al., 2021). These life stages are difficult to study due to the sparseness of animals in such pelagic environments and the consequent high cost of data collection. Furthermore, within a single lamprey species, considerable variation in environmental conditions is experienced (Potter et al., 2015; Mateus et al., 2021), necessitating a flexible tool to explore the understudied aspects of lamprey biology. Few reliable methods exist for studying lampreys throughout their life cycle (Moser, Butzerin & Dey, 2007; Bonar, Hubert & Willis, 2009; Clemens et al., 2022).

Methods

 The model framework is built on the basic life cycle of parasitic, anadromous lampreys and simulates abundance across six serial life stages: spawning adults, eggs, larvae, transformers, juveniles, and returning adults, which spawn in freshwater (Clemens, 2019; Figure 1). Fish move sequentially through each life stage and age class (Figure 1) and are subjected to survival processes including density-dependent survival (Figure 2). For simplicity, fish growth, among other physiological processes, is not currently considered. To add demographic stochasticity, and to explore parameter uncertainty, processes include draws from probability distributions (see below). Process-based models, such as this one, often consist of repeated simulation runs with stochastic sampling from probability distributions or multiple variants of a base model (i.e., Monte Carlo models), which allows parameter uncertainty to compound in model outputs and to propagate over time and/or space for the population or ecosystem of interest.

 We used Pacific lamprey as a model species for the design and implementation of this life cycle model. The Pacific lamprey is an anadromous species of fish inhabiting a widespread area across the Northern Pacific Ocean and associated tributaries (Potter et al., 2015; Clemens et al., 2019;

Mateus et al., 2021). They are semelparous (die after spawning), construct redds (nests) in stream

Life Cycle Model Formulation

Spawners to eggs

125 Survival is represented by the transition of individuals from one life stage to the next. The 126 transition between the spawner and egg life stages includes fecundity in addition to egg survival. 127 The number of eggs, n_{ege} , for each individual, *i*, is drawn from a negative binomial distribution 128 with mean $λ_{\text{fecund}}$ and standard deviation $σ_{\text{fecund}}$:

129
$$
n_{\text{egg}_i} \sim \text{negative binomial}(\lambda_{\text{fecund}}, k_{\text{fecund}}),
$$
 (1)

130 where λ_{fecund} equals the mean fecundity and the negative binomial shape parameter, k_{fecund} , is

131 related to the standard deviation via moment matching (Hobbs & Hooten, 2015):

$$
k_{\text{fecund}} = \frac{\lambda^2_{\text{fecund}}}{\sigma^2_{\text{fecund}}}.
$$
 (2)

133 Total eggs, n_{egg} , are summed across all spawners, where the number of spawners is the product 134 of the total number of adults and the female sex ratio:

$$
n_{\text{egg}} = \sum_{i=1}^{n_{\text{spawners}}} n_{\text{egg}_i}, \tag{3}
$$

136

137
$$
n_{\text{spawners}} = n_{\text{adults}} \cdot [9: \sigma]. \tag{4}
$$

138

139 *Eggs to larvae*

140 The Beverton-Holt function was used to account for density dependence in the freshwater

141 survival of eggs (Moussalli & Hilborn, 1986). The calculated survival estimates, *π*egg, are then

- used as probabilities in a binomial distribution to incorporate demographic stochasticity to the
- simulated number of individuals at the subsequent age-0 larval life stage:

$$
\pi_{\rm egg} = \frac{1}{\left(\frac{1}{S_{\rm egg}} + \frac{n_{\rm egg}}{K_{\rm egg}}\right)},\tag{5}
$$

$$
n_{\text{larvae}_0} \sim \text{binomial}(n_{\text{egg}}, \pi_{\text{egg}}), \tag{6}
$$

148 where n_{larvae_0} is the number of age-0 larvae; π_{egg} is the Beverton-Holt estimated, density-149 dependent, egg to larva survival probability; S_{egg} is the baseline egg to larva survival, without 150 density dependence (defined by the user); n_{ege} is the number of eggs present, and K_{ege} is the carrying capacity of eggs in the system or spawning capacity (in # eggs; defined by the user).

Hatchery additions

 Hatchery releases occur during this step, before fish experience density dependence in the wild (see next step). The user can select any number of hatchery releases consisting of any number of age-1 to age-10 larvae and transformers. For simplicity, the model treats hatchery fish as externally-sourced (i.e., does not account for in-system hatchery production logistics such as removing fish for brood), and fish are added directly to the existing population. That is, the model assumes no differences between hatchery released fish and naturally occurring fish.

161 *Annual larval survival*

- 162 Larval survival occurs through density-independent (see below) and density-dependent
- 163 processes. Density-dependent survival is calculated across all age-1+ larvae, regardless of size,
- 164 through modifications of mortality $(M,$ optional) and carrying capacity, K , during two sequential
- 165 periods, summer and winter.
- 166 First, we sum larvae across all age-1+ larvae available at the beginning of the year (in the

$$
167 spring):
$$

168
$$
n_{\text{spring.larvae}} = \sum_{l=1}^{10} n_{\text{larvae}_{l}}.
$$
 (7)

169 We can specify any summer mortality, M , here, or we can set to 0 to simplify:

$$
S_{\text{summer}} = 1 - M_{\text{summer}}.\tag{8}
$$

171 The Beverton-Holt function is again used for including carrying capacity, K :

$$
\pi_{\text{summer}} = \frac{1}{\left(\frac{1}{S_{\text{summer}}} + \frac{n_{\text{spring}.\text{larvae}}}{K_{\text{ summer}}}\right)},\tag{9}
$$

173 where, like in eq. 5, S_{summer} is the baseline summer larval survival, without density dependence 174 (defined by the user via summer mortality in eq. 8). Larval lamprey surviving the summer are 175 then drawn from a binomial distribution, using π_{summer} as the survival probability:

$$
n_{\text{fall}.\text{larvae}} \sim \text{binomial}(n_{\text{spring}.\text{larvae}}, \pi_{\text{summer}}) \tag{7}
$$

178 Winter conditions are explored analogously to summer conditions. Similar to summer

- 179 conditions, winter conditions can be explored by setting the winter rearing capacity, $K_{\text{winter.larvae}}$
- 180 and winter mortality, M_{winter} . Again, this affects all age-1+ larvae together, regardless of size.
- 181 The equations for winter conditions are the same as for summer conditions:

$$
S_{\text{winter}} = 1 - M_{\text{winter}} \tag{8}
$$

183
$$
\pi_{\text{winter}} = \frac{1}{\left(\frac{1}{S_{\text{winter}}} + \frac{n_{\text{fall.larvae}}}{K_{\text{winter}}}\right)}
$$
(9)

184
$$
n_{\text{spring}+1.\text{larvae}} \sim \text{binomial}(n_{\text{fall}.\text{larvae}}, \pi_{\text{winter}})
$$
 (10)

185 This process is used to calculate the density-dependent survival, π_{dd} , which is applied to each 186 age class, along with an age-class specific density-independent survival, π_{di_l} , which is specified 187 by the user.

$$
\pi_{dd} = \frac{n_{\text{spring}+1.\text{larvae}}}{n_{\text{spring}.\text{larvae}}} \tag{11}
$$

189 Surviving larvae in each age-class, $n_{\text{survived.larvae}}$, is drawn from the pool of available larvae in 190 that age-class, n_{larvae_l} :

191
$$
n_{\text{survived.larvae}_l} \sim \text{binomial}(n_{\text{larvae}_l}, \pi_{dd} \cdot \pi_{di_l}).
$$
 (15)

192

193 *Larval age classes and transformation*

Any surviving age-2 to age-10 larvae in the model can transform with probability δ_l for age class *l.* Individuals are stochastically drawn from a binomial distribution:

$$
n_{\text{transform}_l} \sim \text{binomial}(n_{\text{survived.larvae}_l}, \delta_l) \tag{12}
$$

 The total number of transformers is the sum of individuals that transformed from each age class, l :

204
$$
n_{\text{transformers}} = \sum_{l=2}^{10} n_{\text{transform}_{l}},
$$
 (13)

205 once larval fish reach 10 years of age, they transform $(\delta_{10} = 1)$.

 Larvae that do not enter the intermediate transformer life stage will enter the next age class with 208 probability $(1 - \delta_l)$, where $(1 - \delta_l)$ is the probability of not transforming in a given year. The next age class of larvae could then be, again, drawn from a binomial distribution:

$$
n_{\text{larvae}_{l+1}} \sim \text{binomial}(n_{\text{survived.larvae}_{l}}, 1 - \delta_{l}) \tag{18}
$$

 However, fish that have already transformed are removed from the pool of larvae that can move to the next age class. Thus, to ensure that larval lamprey are not being created nor destroyed (via

- 213 stochastic resampling of the same individuals), we instead subtract the number of transformed
- 214 fish from the number of surviving larvae (within that age class):

$$
n_{\text{larvae}_{l+1}} = n_{\text{survived.larvae}_{l}} - n_{\text{transform}_{l}} \tag{19}
$$

216

217 *Downstream barriers and juveniles at sea*

 Before juvenile lamprey reach the ocean, they encounter any downstream barriers. The user can specify any number of barriers and the probability of passage for each barrier (or average proportion of fish that pass those barriers). The resulting number of juveniles that continue 221 towards the marine environment, $n_{\text{juveniles}_M}$, (soon to become ocean age-0 juveniles once they survive ocean entry) is estimated as:

$$
n_{\text{juveniles}_M} \sim \text{binomial}(n_{\text{transformers}}, \theta_{\text{DS}}) \tag{14}
$$

224 where θ_{DS} is the total downstream probability of riverine juveniles, $n_{\text{iupeniles}}$, passing all 225 barriers. The total downstream probability of passing barriers equals the product of all the 226 probabilities of passing each successive downstream barrier, θ_{D_d} , for d downstream barriers:

$$
\theta_{\rm DS} = \prod_{k=1}^d \theta_{D_d}.
$$
\n(15)

228 If there are no barriers, $d = 0$, $\theta_{DS} = 1$, and $n_{\text{juveniles}_M} = n_{\text{transformers}}$. However, if the 229 translocation option is selected, translocated juveniles $(n_{juveniles_T})$ are removed from the fish 230 experiencing barrier passage, and instead are moved below the barriers, with the user-defined 231 juvenile translocation survival probability θ_{IT} :

232
$$
n_{\text{juveniles}_P} \sim \text{binomial}(n_{\text{juveniles}} - n_{\text{juveniles}_T}, \theta_{\text{DS}}),
$$
 (22)

233
$$
n_{\text{juveniles}_{ST}} \sim \text{binomial}(n_{\text{juveniles}_{T}}, \theta_{\text{JT}}).
$$
 (23)

234 Fish that experienced and passed barriers $(n_{juveniles_P})$ are then summed with surviving

translocated fish $(n_{\text{inveniles}})$, to calculate the number of juveniles entering the ocean, in lieu of 236 equation 20:

$$
n_{\text{juveniles}_M} = n_{\text{juveniles}_P} + n_{\text{juveniles}_{ST}} \tag{24}
$$

238

239 Juveniles then may experience mortality as they meet marine predators in the ocean (e.g., the 240 river plume). We again use moment matching (Hobbs $\&$ Hooten, 2015), to include our moments 241 (mean, μ and standard deviation σ of ocean entry survival, $\pi_{\text{entry. ocean}}$) in the beta distribution 242 parameters, α and β :

$$
\alpha = \left(\frac{\mu \cdot (1 - \mu)}{\sigma_{\text{EO}}^2} - 1\right) \cdot \mu,\tag{25}
$$

244
$$
\beta = \left(\frac{\mu \cdot (1 - \mu)}{\sigma_{\text{EO}}^2} - 1\right) \cdot (1 - \mu), \tag{26}
$$

$$
\pi_{\text{entry. ocean}} \sim \text{beta}(\alpha, \beta). \tag{27}
$$

246 We can then draw our resulting juvenile fish (in the $0th$ juvenile age-class) from the number of

247 juveniles entering the marine environment with probability $\pi_{\text{entry.ocean}}$.

248
$$
n_{\text{juvenile}_0} \sim \text{binomial}(n_{\text{juveniles}_M}, \pi_{\text{entry. ocean}}).
$$
 (28)

249

250 *Juveniles at sea to adults in river*

251 Once juveniles are at sea, each age class, *j*, can transition to an adult returner, heading upstream 252 with an age-class specific probability ζ_j , and have a river entry survival probability π_{entry} . 253 Both ζ_i and $\pi_{\text{entry river}}$ are ultimately defined by the user, however $\pi_{\text{entry river}}$ is specified with a

254 mean, μ, and standard deviation, σ , for river entry survival as in equations 25-27:

$$
n_{\text{return}_j} \sim \text{binomial}\left(n_{\text{juvenile}_j}, \zeta_j \cdot \pi_{\text{entry river}}\right),\tag{29}
$$

256 where n_{return_j} is the number of juveniles in class, *j*, that will return to a riverine system. The

257 sum across all age classes is the total in-river adults (n_{adults_R}) :

$$
n_{\text{adults}_{\text{R}}} = \sum_{j=1}^{10} n_{\text{return}_j} \tag{30}
$$

259 Juveniles that remain at sea advance to the next juvenile age-class with a probability equal to 260 yearly ocean survival (which is parameterized with different user-defined inputs, but relies on the 261 same equations as the ocean entry survival above):

262
$$
n_{juvenile_{j+1}} \sim binomial(n_{juvenile_j} - n_{return_j}, \pi_{yearly. ocean}).
$$
 (31)

263 That is, $\pi_{\text{vearly. ocean}}$ is specified with a mean, μ , and standard deviation, σ , for yearly ocean 264 survival as in equations 25-27. If fish spend 10 years as juveniles, they automatically swim 265 upriver if they do not happen to die during the process.

267 *Upstream barriers and translocation*

268 Adults swimming upstream may encounter upstream barriers, which impede the probability of 269 fish passage. The resulting number of adult fish of age-class 0 in the river, n_{adult_0} , is estimated 270 as:

271

$$
n_{\text{adult}_0} \sim \text{binomial}(n_{\text{adults}_R}, \theta_{\text{US}}) \tag{32}
$$

273 where θ_{US} is the total upstream probability of each adult, n_{adults_R} , passing all barriers. The total 274 upstream passability, θ_{US} , equals the product of all the probabilities of passing each successive 275 barrier, θ_{U_u} , for *u* upstream barriers:

$$
\theta_{\text{US}} = \prod_{k=1}^{u} \theta_{U_u} \tag{33}
$$

277 However, in the event that the translocation option is selected, translocated adults (n_{adult_T}) are 278 removed from the fish experiencing barrier passage, and instead are dropped off above the 279 barriers, with the user-defined adult translocation survival probability θ_{AT} :

280
$$
n_{\text{adult}_{P}} \sim \text{binomial}(n_{\text{adult}_{R}} - n_{\text{adult}_{T}}, \theta_{\text{US}}), \tag{34}
$$

$$
n_{\text{adult}_{ST}} \sim \text{binomial}(n_{\text{adult}_T}, \theta_{\text{AT}}). \tag{35}
$$

282 Fish that experienced and passed barriers (n_{adult_P}) are then summed with surviving translocated 283 fish (n_{adulter}) :

$$
n_{\text{adult}_0} = n_{\text{adult}_P} + n_{\text{adult}_{ST}} \tag{36}
$$

Adults to spawners

- 287 Some adults may spawn immediately (adult age-0, n_{adult_0}), while others will hold in the river
- without feeding and spawn within the next two years (adult ages 1 and 2). Adult age-classes, *a*,
- 289 spawn in a given year with a probability η_a , where age-2 adults all spawn $(\eta_2=1)$:

$$
n_{\text{spawn}_a} \sim \text{binomial}(n_{\text{adult}_a}, \eta_a). \tag{37}
$$

The sum across all age classes is the total spawners for that year:

$$
n_{\text{spawners}} = \sum_{a=0}^{2} n_{\text{spawn}_a} \tag{38}
$$

 Spawners now contribute to the next generation via equations 1 and 3 and the life cycle continues to the next year.

Default parameter values

 Pacific lamprey occupy a broad geographic range (Potter et al., 2015). Thus, many of their life cycle parameters are likely to vary across their range. Here, we provide the model structure and some rationale for default lamprey life cycle parameters. That is, if the user runs the model code in R, or in the Shiny app, without specifying parameters, the defaults will be automatically used. However, it is up to the user to specify the parameters that most closely represents their system and lamprey species of interest. Sometimes information is limited, and one will have to make assumptions about how to use information from other populations or species. Models are

Spawning and fecundity

 An initial number of spawners is a necessary simulation input. After the first year the internal population dynamics determine the spawner numbers in subsequent years. An arbitrary default of 800 spawners was selected. Longer burn-in time (also known as warm-up or simulation years that are thrown away [Hoad, Robinson & Davies, 2010]) will ensure that this initial number of spawners does not affect the simulation since internal dynamics will dominate after a few years (Figure 3). The number of females is determined each year by the female sex ratio, which is defaulted to 0.5 (or 50% females). The fecundity of each female is drawn from a distribution (eq. 322 1) with a mean, λ_{ferund} , and standard deviation, σ_{ferund} , defaulted to 127,000 and 33,500 eggs, respectively (rounded to the nearest 500 eggs [Clemens et al., 2013]).

Egg survival

Larval survival

349 Total summer and winter larval carrying capacity, K_{summer} and K_{winter} , can be input directly by the user (arbitrary default is 1.3 million larvae during each time period), or it can be calculated from the total river area and mean larval density measured at relevant sites. The default mean 352 density (individuals/ $m²$) for larvae is 0.13, which is taken as an average across 14 tributaries from the Willamette River Basin (Appendix 1 in Schultz et al., 2014) and 10 tributaries of the Columbia River Basin (Table 4 in Harris & Jolley, 2017). If larval carrying capacity is defined, instead of larval density, the default of 1.3 million larvae is identical to using the default density 356 of 0.13 larvae/m² with 10 km² of available rearing habitat. Survival probability can be selected by age class (age-0 – age-10; default) or with the same probability for each age class (less realistic but requires fewer decisions). Default larval survival by age-class values were taken from Schultz et al., 2014 (Table 5-1; Chen/Watanabe estimates without age-0, since age-0 survival is input directly here). Only values for age-1 to age-7 were provided by Schultz et al., 2014. We estimated age-0 survival to be 33% because a 0.33 age-0 362 survival makes cumulative survival across age- $0 - \text{age-7}$ to be 22.5%, which is in-line with previous estimates of 19-26% for entire larval life cycle survival (Schultz et al., 2014). To estimate age-8 through age-10 survival, we fit a sigmoid curve to age-1 to age-7 data (from Schultz et al., 2014) with R package `nls` to predict the survival values, which were rounded to the nearest 2 decimal places (see Table 1; Gomes, 2024).

Larval transformation by age-class

 Transformation probability can be selected by age class (age-0 – age-10; default) or with the same probability for each age class (less realistic but requires fewer decisions). Transformation probabilities were modified until matched empirical data for the Yakima River Basin (age-2: 1%, age-3: 9%, age-4: 62%, and age-5: 28%, R. Lampman, *unpublished*). This particular set of values (see Table 1) only matches the Yakima River empirical transformer composition data given the other default model parameters. Thus, if users change life cycle parameters, such as egg or larval survival, these transformation probabilities will need to be modified again to match the desired transformer composition. These particular values also do not allow larvae to reach age-6, as they will automatically transform at age-5, assuming they survive. This is realistic for some streams, but not others. These values should be adjusted to fit the age-class distribution for the stream of interest. That is, the user can adjust transformation probabilities such that transformer age class composition matches that of the focal system.

Entering the marine environment

383 We used a value of 0.9 (90%) for the default downstream passage success per dam, θ_{D_d} . This value is estimated at Lower Granite Dam (Deng et al., 2023). To our knowledge, data do not exist specifically for ocean entry survival (Laurie Weitkamp, *personal communication*). We assume that marine predators in the plume are opportunistically consuming available prey. Thus, as a rough approximation, we have taken ocean entry survival as the mean (across years) value of Columbia River plume survival for Chinook salmon (*Oncorhynchus tshawytscha*) smolts 389 (survival \pm SD: 0.46 \pm 0.09 from Table 1 in Brosnan et al., 2014; Gomes, 2024). Annual survival for juveniles in the ocean (referred to here as yearly ocean survival) is assumed to be 391 substantially higher, and a best guess of 0.7 ± 0.1 (mean \pm SD) was used.

Re-entering freshwater and spawning

The probability that each juvenile age class $(0 - 10)$ heads upstream, ζ_j , is set to 0, 0.02, 0.05,

0.1, 0.15, 0.45, 1, 1, 1, 1, 1 as a default. These values are based on age-class distributions for

each life stage of Snake River Pacific lamprey (Hess et al., 2022). Figure 5 in Hess et al., (2022)

suggests that most fish spend 5 or 6 years in the ocean; thus, we have selected nearly half (0.45)

398 of 5th ocean year juveniles and all of the remaining $(\zeta_6 = 1)$ 6th ocean year juveniles to swim

upriver. With this parameterization fish never spend more than 6 years in the ocean (Hess et al.,

400 2022), but this can be extended by the user $(\zeta_6 < 1)$.

 Once lamprey enter the mouth of the river, they may experience increased levels of predation or mortality at these physical bottlenecks (Roffe & Mate, 1984), which is controlled in this model via a river entry survival. Given that predation on lamprey re-entering riverine systems (such as by pinnipeds) can be equal to, or higher than, predation on salmonids (Roffe & Mate, 1984; Riemer, Wright & Brown, 2011), we have used information on river mouth survival of adult salmonids as a stand-in for the more data-limited lamprey populations. Thus, default survival for 407 Pacific lamprey entering the river mouth was taken as 0.67 ± 0.09 (mean \pm SD; from Figure 5; Wargo Rub et al., 2019; Gomes, 2024).

For fish encountering barriers, we used a value of 0.6 (60%) for the default upstream passage

410 success, θ_{U_u} (the mean of all lower and upper extreme values estimated at Bonneville [38-47%],

The Dalles [50-82%], and McNary [65-75%] dams [Moser et al., 2002; Keefer et al., 2013]).

Any number of available adult fish can be translocated above all barriers (Hess et al., 2022), but

 the user must specify the survival probability of translocation. We use a default value of 0.99 for adult translocation survival (Aaron Jackson, *personal communication*).

 Adults can wait in the river for up to two years prior to spawning. As a default probability of spawning for entry-year adults (adult age-class 0) and adults that have waited one year (adult age-class 1), we have used 0.05 and 0.7, respectively, to reflect that most Pacific lamprey appear to wait at least one year, while we assume all adult age-class 2 individuals spawn after two years $(\eta_2=1)$.

Global sensitivity analysis

 Sensitivity analyses allow for an exploration of model sensitivities to particular parameters. As opposed to altering one parameter at a time, and keeping all others at fixed values, global sensitivity analyses (GSA) allow an assessment of parameter influence on overall model dynamics while considering all possible interactions among parameters (Saltelli et al., 2008; Iooss & Lemaître, 2015). A GSA is advantageous to fixing all non-modified parameter values to mean values and assuming that they are correct, because parameter space, and thus parameter influence, is more thoroughly explored. To more efficiently sample parameter space, we used Latin hypercube sampling (LHS). LHS generates near-random sampling of parameter values from a multidimensional distribution where the sampling space is broken up into strata or breaks (Loh, 1996; Helton & Davis, 2003). Each break in the sampling space is randomly sampled only once for each parameter such that the range of parameter values is efficiently sampled across all parameters simultaneously. We used the R (R Core Team, 2017) package `lhs` (Carnell, 2022) to

- set up two global sensitivity analysis where parameter values were sampled with a Latin hypercube with 100 breaks.
- For the first GSA, we sampled parameters more broadly. For example, values that are constrained between 0 and 1 (e.g., survival, transformation probabilities, etc.) are mostly not empirically resolved for Pacific lamprey and, thus, were sampled across the entire range of 439 possible values $(0 - 1)$ to fully explore potential influence of the parameter space. Fecundity is hypothetically only constrained on the lower end to 0, whereas the upper end could be infinite. Since this was not feasible, fecundity was explored with a normal distribution with the mean and standard deviation as defined in *Default values* (see Tables 1, 2). Carrying capacities vary by location and spatial extent; thus, they were explored across a large range of values with uniform distributions:

445
$$
K_{\text{egg}} \sim \text{uniform}(1 \times 10^5, 1 \times 10^{12})
$$
 (39)

$$
K_{\text{summer}} \sim \text{uniform}(1 \times 10^4, 1 \times 10^8) \tag{40}
$$

$$
K_{\text{winter}} \sim \text{uniform}(1 \times 10^4, 1 \times 10^8) \tag{41}
$$

 Since the GSA relies on LHS, the deterministic option (function argument) was turned on in the `MODEL.R` code, and, thus, standard deviation values are not necessary to systematically sample (i.e., the LHS handles the variation in inputs already).

For each of the 100 GSA runs (100 LHS breaks), we ran the model for 50 years after 50

additional burn-in years (Figure 3). Since stochastic sampling was turned off, each run reached

nearly constant stable states after a conservative 50-year burn-in. The resulting number of

- spawners in the last year of the simulation was used as the response variable in a random forest
- model to assess parameter influence.

 constrained to more reasonable values (see Table 2). For example, instead of sampling uniformly between 0 and 1 for the sex ratio, this parameter-restricted GSA explored sex ratios drawn from a beta distribution with mean of 0.5 (50% males and females) and a standard deviation of 0.05. That is,

$$
[9: \sigma] \sim \text{beta}(\alpha, \beta), \tag{42}
$$

470 where α and β are moment matched to μ and σ via equations 25 and 26 (Hobbs & Hooten, 2015). All of the remaining parameters were similarly drawn from a beta distribution with their respective mean and standard deviations. All parameters retained their mean values from the *Default parameters* section (Table 1) except where probabilities were too close to 0 or 1 for proper sampling (Table 2). In these cases, values of 0.05 and 0.9, respectively, were used. Resulting beta distributions were visualized to ensure that values of 0 and 1 were well-represented. Most parameters were sampled with a conservative standard deviation equal to 0.1,

 to ensure that a broad (yet more restricted relative to the first GSA) parameter space was sampled (see Table 2).

Management scenario case study examples

 To demonstrate potential management applications, we chose five simple scenarios to explore (Table 3). Each scenario consists of 100 stochastic replications of a 100-year simulation (50-year run time + 50-year burn-in period). The first scenario is considered baseline conditions, where only default parameter values are input. The second scenario adds one upstream barrier with the default 60% passage probability. In this scenario, the number of adults immigrating to freshwater are reduced by 60%. The third scenario is the same as the second but includes the translocation of up to 100 adults for each year, within each simulation. If there are fewer adults available in the model in any given year, then all adults are translocated above the barrier for that year. The fourth scenario is also the same as the second but includes yearly hatchery releases of 1000 fish at the 'transformer' stage, which is the short stage in between larvae and juveniles, before the fish have made it to the ocean (Figures 1, 2). The fifth scenario is a combination of scenarios 3 and 4, which includes one upstream barrier with the default 60% passage probability, the translocation of up to 100 adults for each year, and yearly hatchery releases of 1000 fish at the 'transformer' stage. Results are visualized as the median value of the 100 replications for each of the 5 scenarios for each year (see Figure 6 and function `PlotTSComparison` in Gomes, 2024).

Results

 Simulations with default parameter values, during 100-year simulations, were stable and did not lead to population extinctions or explosions. The initial number of spawners did not affect the final number of spawners after a sufficient burn-in period (Figure 3). The burn-in period smoothed the initial startup population dynamics and leads to more consistent results across minor changes in initial conditions.

Global sensitivity analysis

 The global sensitivity analysis (GSA) with the broad parameter sampling scheme suggested that the most sensitive parameters in the model were age-0 larval survival, juvenile ocean age-7 swim upriver probability, and age-1 and age-5 larval survival (Figure 4). When more realistic parameter values were explored in a second GSA, the most sensitive parameters in the model were egg survival, age-7 larval transformation probability, yearly ocean survival, and age-7 larval survival (Figure 5). In both GSA random forest analyses, 17 of 44 parameters had positive effects (i.e., they were considered influential) on predicting (measured as % mean squared error) the number of spawners at the end of a 100-year simulation. Many of these influential parameters were the same across the two analyses (yearly ocean survival, fecundity, river entry survival, summer and winter larval carrying capacity [K], age-6 larval survival, age-1 larval transformation probability), or very nearly the same (multiple ages of larval survival and transformation probabilities, and juvenile ocean age-5, -6, and -7 probabilities of swimming upriver), yet their order of importance differed between the two analyses.

Management scenario sensitivity analysis

Model limitations

 Process-based models are useful tools in exploring connected systems, complex ecological phenomena, and whole-ecosystem management (Urban, 2005; Cuddington et al., 2013; Geary et al., 2020). Yet, all models are necessarily simplified approximations of reality. Structurally, there are a few model limitations. The current model assumes a closed population. That is, lampreys do not emigrate from nor immigrate to other populations. Similarly, adult translocations in the model do not consider that in the real world, depending on the collection and release points, lamprey may be diverted from one tributary or population to another. While the addition of a meta-population module can be added to future versions of the LLCM, this simplifying assumption may or may not be an important omission, depending on the ecology and population dynamics of the particular lamprey species or population in question. Lampreys do not appear to home to natal streams. If watersheds are not population sources nor sinks at the scale of interest, the modeled population dynamics likely will yield similar results, despite ignoring emigration and immigration. This limitation is also likely to vary spatially, considering larger areas will likely lead to neutral net emigration and immigration into and out of the model domain (e.g., modeling the global population versus local populations).

 For the sake of simplicity and computational efficiency, we ignored growth and biomass in the LLCM, and instead focused on tracking individual lamprey numbers. This has several important consequences worth considering. Fecundity is a function of lamprey size (Docker & Beamish, 1991; Gambicki & Steinhart, 2017), yet since size is not considered in the model, all individual fecundity values are drawn from the same distribution (regardless of pre-spawn instream holding time, which may or may not affect fecundity in reality). LLCM users can currently explore the

 We were able to find default parameters from Pacific lamprey-specific work for many, but not all parameters. Life cycle parameters in Table 1 came from different populations and, sometimes, different species. While these default values are a useful starting place, some parameter values will likely vary for different populations and species of lamprey. Future LLCM users should attempt to parameterize important differences for populations of interest. Notably for Pacific lamprey, we were unable to find larval freshwater survival and transformation probability values, juvenile survival values upon entering and exiting the ocean, and yearly ocean survival values.

 This is not surprising given the difficulty of marking or tracking wild lamprey in either the sediment or in the open ocean. Interestingly, these parameters appeared to be important in the global sensitivity analyses (see below).

Global sensitivity analysis

 We employed two strategies for exploring parameter influence. In the first GSA we allowed a broad range of possibilities for parameter values. Some parameter values are highly uncertain, so exploring the entire range of possibilities can be a more conservative approach. However, this means that we explored potentially unrealistic values (e.g., survival parameters close to 0 or 1). Thus, in the second GSA, constraining values to a more realistic range will shift the 'importance' weighting to other parameters since the influence of some parameters will become more restricted (e.g., no longer uniform distribution from 0 to 1). Thus, we are more inclined to trust the results of the second GSA, while recognizing that the assigned uncertainty distributions we used to explore parameter space might not fully capture the realm of possibilities across all populations or species of lamprey. This analysis highlights the importance of continued monitoring to better estimate the life cycle parameter values and variability for populations of interest, since these inputs can change the relative importance of each parameter in the life cycle model. Our hope is that future model users continue to explore these parameters and their influence on population dynamics.

 In the GSA with the broadest explored parameter space, survival values for juveniles entering the ocean and the river, and yearly ocean survival values were all influential, whereas ocean entry survival was not influential in the more restricted parameter space analysis. All three of these

 parameters are not well-resolved in the empirical lamprey literature, such that we used Chinook salmon values for two of them and based the third on assumptions. While these are difficult values to estimate, the GSA suggests that they would be useful in being able to predict spawning lamprey numbers, especially yearly ocean survival, which came out in the top 5 most influential parameters in both GSAs.

 Early survival values were highly influential in each GSA. Age-0 larval survival was the most influential parameter in the broad GSA (Figure 4), while egg survival was the most influential in the restricted parameter GSA (Figure 5). Both age-0 larval survival and egg survival default values are based here on some empirical literature and assumptions. Other larval survival (and transformation probability) parameters were also influential in both analyses, although the two GSAs differed in which larval age-classes had the most influential effects on predicting spawners. Larval survival and transformation probabilities were adjusted to match an age-class distribution of transformers from a single stream, but otherwise were not constrained by real data. More information on all of the above parameters would help to resolve a Pacific lamprey-specific life cycle model.

 Fecundity was one of the few parameters that was consistent across GSAs (Table 2), as it is relatively well-known and, thus, constrained. This parameter was influential in both GSAs, however, the magnitude of importance changed substantially between analyses (Figure 4, 5). These analyses suggest that better constraining all parameters can alter, and potentially help elucidate, which parameters are influential. These simulations highlight the value of fine-tuning life cycle model parameters using field and laboratory research. Future uses of the LLCM can include more systematically altering the parameter mean values and variation to understand the effect that fine-tuning any particular parameter has on parameter influence in the model.

Management scenario sensitivity analysis

 Our management scenarios are a few simple examples of what the LLCM can explore. We focused on two management strategies that have been highlighted as potentially useful for the conservation and restoration of Pacific lamprey (CRITFC, 2011; CRITFC et al., 2018; Hess et al., 2022). In Figure 6 the population of spawners stabilizes at various values given the management scenarios employed and the default values used for all other parameters. It is important to note that absolute numbers of spawners are not meaningful outside the context of the particular values selected for the model life cycle parameters (including arbitrary carrying capacity values that do not represent any particular stream). Additionally, these absolute numbers are only possibly meaningful if we can validate these numbers with empirical data for particular populations. Instead, we are focusing on the relative values between scenarios to explore management alternatives.

 In these case studies, we demonstrated that the barrier addition, fish translocation, and hatchery release functions are operational in the LLCM. The barrier addition reduced the spawning population relative to baseline conditions, which was expected. Each of the adult translocation and the hatchery release of transformers options appeared to increase the spawning population to some degree, although either strategy in isolation was not enough to overcome the deficit from the addition of one upstream barrier. These two strategies in this simplistic simulation appeared to be equal in efficacy, although note an order of magnitude difference in the number of transformers released from hatcheries (1000) relative to the maximum number of translocated

Conclusions and future directions

 Management actions, climate, and life cycle parameters can interact in unexpected ways, which can be explored with our life cycle model. Management actions can be financially costly, which typically results in only a limited number of methods being available to any one conservation group or effort. Understanding the expected efficacy of such alternative actions, before taking such actions, can be greatly beneficial to species conservation. Future studies using this model could explore tradeoffs between barriers (including multiple barriers in both the upstream and downstream directions), fish translocation, and hatchery releases amidst a changing climate. Users of the model can adjust summer and winter mortality and carrying capacities to explore what future climate scenarios might bring to lamprey populations. In addition to conservation efforts, this model could prove useful in assisting the control of the invasive sea lampreys in the Laurentian Great Lakes. We hope that this modeling framework is useful in aiding hypothesis

- generation and directing future research efforts, with the ultimate goal of improving lamprey conservation and management.
-

Acknowledgments

- We thank Burke Strobel, Mariah Mayfield, Tom Stahl and the USGS Viz Lab for helpful
- feedback on the design and layout of the Shiny application. This manuscript benefitted from a
- review by Bob Rose, Megan Sabal, and Nicholas Schloesser. Monica R. Blanchard provided the
- beautiful illustration used in Figure 1. Thomas Hobbs and Mevin Hooten provided a Bayesian
- Modeling course to DGEG (NSF DEB 2042028), which was helpful to understand and document
- probability density/mass functions, moment matching, and math notation. Funding was
- recommended by Pacific Lamprey Conservation Initiative and provided by the Bonneville Power
- Administration. Any use of trade, firm, or product names is for descriptive purposes only and
- does not imply endorsement by the US government.
-
-

References

- Bonar SA, Hubert WA, Willis DW. 2009. Standard methods for sampling North American freshwater fishes.
- Brosnan IG, Welch DW, Rechisky EL, Porter AD. 2014. Evaluating the influence of
- environmental factors on yearling Chinook salmon survival in the Columbia River plume (USA).
- *Marine Ecology Progress Series* 496:181–196.
- Carnell R. 2022. lhs: Latin Hypercube Samples.
- Chang W, Cheng J, Allaire JJ, Sievert C, Schloerke B, Xie Y, Allen J, McPherson J, Dipert A,
- Borges B. 2023. shiny: Web Application Framework for R.

- Clemens BJ. 2019. A call for standard terminology for lamprey life stages. *Fisheries* 44:243– 245.
- Clemens BJ, Arakawa H, Baker C, Coghlan S, Kucheryavyy A, Lampman R, Lança MJ, Mateus
- CS, Miller A, Nazari H. 2021. Management of anadromous lampreys: Common threats, different
- approaches. *Journal of Great Lakes Research* 47:S129–S146.
- Clemens BJ, Binder TR, Docker MF, Moser ML, Sower SA. 2010. Similarities, differences, and unknowns in biology and management of three parasitic lampreys of North America. *Fisheries* 35:580–594.
- Clemens BJ, Harris JE, Starcevich SJ, Evans TM, Skalicky JJ, Neave F, Lampman RT. 2022.
- Sampling Methods and Survey Designs for Larval Lampreys. *North American Journal of*
- *Fisheries Management* 42:455–474. DOI: 10.1002/nafm.10762.
- Clemens BJ, Van De Wetering S, Sower SA, Schreck CB. 2013. Maturation characteristics and
- life-history strategies of the Pacific lamprey, *Entosphenus tridentatus*. *Canadian Journal of*
- *Zoology* 91:775–788. DOI: 10.1139/cjz-2013-0114.
- Clemens BJ, Weitkamp L, Siwicke K, Wade J, Harris J, Hess J, Porter L, Parker K, Sutton T,
- Orlov AM. 2019. Marine biology of the Pacific lamprey Entosphenus tridentatus. *Reviews in*
- *Fish Biology and Fisheries* 29:767–788.
- Close DA, Fitzpatrick MS, Li HW. 2002. The ecological and cultural importance of a species at risk of extinction, Pacific lamprey. *Fisheries* 27:19–25.
- CRITFC. 2011. Tribal Pacific Lamprey Restoration Plan.
- CRITFC, Yakama Nation, Confederated Tribes of the Umatilla Indian Reservation, Nez Perce
- Tribe. 2018. Master Plan: Pacific Lamprey Artificial Propagation, Translocation, Restoration, and Research.
- Cuddington K, Fortin M-J, Gerber LR, Hastings A, Liebhold A, O'Connor M, Ray C. 2013.
- Process‐based models are required to manage ecological systems in a changing world. *Ecosphere* 4:1–12. DOI: 10.1890/ES12-00178.1.
- Dawson HA, Quintella BR, Almeida PR, Treble AJ, Jolley JC. 2015. The Ecology of Larval and
- Metamorphosing Lampreys. In: Docker MF ed. *Lampreys: Biology, Conservation and Control*.
- Dordrecht: Springer Netherlands, 75–137. DOI: 10.1007/978-94-017-9306-3_3.
- Deng ZD, Deters KA, Martinez JJ, Harnish RA, Mueller RP, Titzler PS, Fu T, Li H, Wu B.
- 2023. *Juvenile Pacific Lamprey Passage Behavior and Survival at Lower Granite Dam, 2022.* Pacific Northwest National Laboratory.
-
- Docker MF (ed.). 2019. *Lampreys: Biology, Conservation and Control: Volume 2*. Dordrecht:
- Springer Netherlands. DOI: 10.1007/978-94-024-1684-8.
- Docker MF, Beamish FWH. 1991. Growth, fecundity, and egg size of least brook lamprey,
- Lampetra aepyptera. *Environmental Biology of Fishes* 31:219–227.

- Docker MF, Hume JB, Clemens BJ. 2015. Introduction: a surfeit of lampreys. *Lampreys:*
- *Biology, Conservation and Control: Volume 1*:1–34.
- Donoghue PC, Forey PL, Aldridge RJ. 2000. Conodont affinity and chordate phylogeny.
- *Biological Reviews* 75:191–251.
- Gambicki S, Steinhart GB. 2017. Changes in sea lamprey size and fecundity through time in the Great Lakes. *Journal of Great Lakes Research* 43:209–214.
- Geary WL, Bode M, Doherty TS, Fulton EA, Nimmo DG, Tulloch AI, Tulloch VJ, Ritchie EG.
- 2020. A guide to ecosystem models and their environmental applications. *Nature Ecology & Evolution* 4:1459–1471.
- Gess RW, Coates MI, Rubidge BS. 2006. A lamprey from the Devonian period of South Africa. *Nature* 443:981–984.
- Gomes DGE. 2024. Lamprey life cycle model: U.S. Geological Survey Software Release. DOI: https://doi.org/10.5066/F7CV4H1T.
- Goodman DH, Reid SB. 2022. Rapid development of larval Pacific lamprey Entosphenus
- tridentatus in southern populations provides adaptive benefits for uncertain flow regimes.
- *Environmental Biology of Fishes* 105:403–411.
- Gumbs R, Gray CL, Böhm M, Burfield IJ, Couchman OR, Faith DP, Forest F, Hoffmann M,
- Isaac NJ, Jetz W. 2023. The EDGE2 protocol: Advancing the prioritisation of Evolutionarily
- Distinct and Globally Endangered species for practical conservation action. *PLoS Biology*
- 21:e3001991.
- Hardisty MW. 1961. The growth of larval lampreys. *The Journal of Animal Ecology*:357–371.
- Hardisty MW. 2006. *Lampreys: life without jaws*. Forrest Text.
- Harris JE, Jolley JC. 2017. Estimation of occupancy, density, and abundance of larval lampreys
- in tributary river mouths upstream of dams on the Columbia River, Washington and Oregon.
- *Canadian Journal of Fisheries and Aquatic Sciences* 74:843–852. DOI: 10.1139/cjfas-2016-
- 0212.
- Helton JC, Davis FJ. 2003. Latin hypercube sampling and the propagation of uncertainty in analyses of complex systems. *Reliability Engineering & System Safety* 81:23–69.
- Hemming V, Camaclang AE, Adams MS, Burgman M, Carbeck K, Carwardine J, Chadès I,
- Chalifour L, Converse SJ, Davidson LNK, Garrard GE, Finn R, Fleri JR, Huard J, Mayfield HJ,
- Madden EM, Naujokaitis‐Lewis I, Possingham HP, Rumpff L, Runge MC, Stewart D, Tulloch
- VJD, Walshe T, Martin TG. 2022. An introduction to decision science for conservation.
- *Conservation Biology* 36:e13868. DOI: 10.1111/cobi.13868.
- Hess JE, Delomas TA, Jackson AD, Kosinski MJ, Moser ML, Porter LL, Silver G, Sween T,
- Weitkamp LA, Narum SR. 2022. Pacific Lamprey Translocations to the Snake River Boost

- Abundance of All Life Stages. *Transactions of the American Fisheries Society* 151:263–296. DOI: 10.1002/tafs.10359.
- Hoad K, Robinson S, Davies R. 2010. Automating warm-up length estimation. *Journal of the Operational Research Society* 61:1389–1403. DOI: 10.1057/jors.2009.87.
- Hobbs NT, Hooten MB. 2015. *Bayesian models: a statistical primer for ecologists*. Princeton University Press.
- Houston KA, Kelso JR. 1991. Relation of sea lamprey size and sex ratio to salmonid availability in three Great Lakes. *Journal of Great Lakes Research* 17:270–280.
- Hume JB, Bennis S, Bruning T, Docker MF, Good S, Lampman R, Rinchard J, Searcy T, Wilkie
- MP, Johnson NS. 2024. Evaluation of Larval Sea Lamprey Petromyzon marinus Growth in the
- Laboratory: Influence of Temperature and Diet. *Aquaculture Research* 2024:1–11. DOI:
- 10.1155/2024/5547340.
- Iooss B, Lemaître P. 2015. A Review on Global Sensitivity Analysis Methods. In: Dellino G,
- Meloni C eds. *Uncertainty Management in Simulation-Optimization of Complex Systems*.
- Operations Research/Computer Science Interfaces Series. Boston, MA: Springer US, 101–122.
- 787 DOI: 10.1007/978-1-4899-7547-8_5.
- Isaac NJB, Pearse WD. 2018. The Use of EDGE (Evolutionary Distinct Globally Endangered)
- and EDGE-Like Metrics to Evaluate Taxa for Conservation. In: Scherson RA, Faith DP eds.
- *Phylogenetic Diversity*. Cham: Springer International Publishing, 27–39. DOI: 10.1007/978-3-
- 319-93145-6_2.
- IUCN. 2012. Resolutions and Recommendations. *World Conservation Congress.* International Union for Conservation of Nature.
- Janvier P. 2006. Modern look for ancient lamprey. *Nature* 443:921–923.
- Johnson WJ. 1982. Body lengths, body weights and fecundity of sea lampreys (Petromyzon
- marinus) from Green Bay, Lake Michigan [Wisconsin]. *Transactions of the Wisconsin Academy of Sciences, Arts and Letters* 70.
- Jolley JC, Uh CT, Silver GS, Whitesel TA. 2015. Feeding and Growth of Larval Pacific
- Lamprey Reared in Captivity. *North American Journal of Aquaculture* 77:449–459. DOI:
- 10.1080/15222055.2015.1044630.
- Keefer ML, Boggs CT, Peery CA, Caudill CC. 2013. Factors affecting dam passage and
- upstream distribution of adult Pacific lamprey in the interior Columbia River basin. *Ecology of Freshwater Fish* 22:1–10. DOI: 10.1111/j.1600-0633.2012.00586.x.
- Lampman RT, Maine AN, Moser ML, Arakawa H, Neave FB. 2021. Lamprey aquaculture
- successes and failures: A path to production for control and conservation. *Journal of Great Lakes*
- *Research* 47:S201–S215.

- Lampman R, Moser ML, Jackson A, Rose R, Gannam A, Barron J. 2016. Developing techniques
- for artificial propagation and early rearing of Pacific lamprey (Entosphenus tridentatus) for
- species recovery. *Jawless fishes of the world* 2:160.
- Liaw A, Wiener M. 2002. Classification and Regression by randomForest. *R News* 2:18–22.
- Loh W-L. 1996. On Latin hypercube sampling. *The annals of statistics* 24:2058–2080.
- Manion PJ, Hanson LH. 1980. Spawning Behavior and Fecundity of Lampreys from the Upper
- Three Great Lakes. *Canadian Journal of Fisheries and Aquatic Sciences* 37:1635–1640. DOI:
- 10.1139/f80-211.
- Mateus CS, Docker MF, Evanno G, Hess JE, Hume JB, Oliveira IC, Souissi A, Sutton TM.
- 2021. Population structure in anadromous lampreys: Patterns and processes. *Journal of Great Lakes Research* 47:S38–S58.
- Mayfield MP, Schultz LD, Wyss LA, Clemens BJ, Schreck CB. 2014. Spawning Patterns of
- Pacific Lamprey in Tributaries to the Willamette River, Oregon. *Transactions of the American*
- *Fisheries Society* 143:1544–1554. DOI: 10.1080/00028487.2014.949013.
- McGree M, Whitesel TA, Stone J. 2008. Larval Metamorphosis of Individual Pacific Lampreys
- Reared in Captivity. *Transactions of the American Fisheries Society* 137:1866–1878. DOI:
- 10.1577/T07-206.1.
- Meeuwig MH, Bayer JM, Seelye JG. 2005. Effects of Temperature on Survival and
- Development of Early Life Stage Pacific and Western Brook Lampreys. *Transactions of the*
- *American Fisheries Society* 134:19–27. DOI: 10.1577/FT03-206.1.
- Miyashita T, Coates MI, Farrar R, Larson P, Manning PL, Wogelius RA, Edwards NP, Anné J,
- 828 Bergmann U, Palmer AR, Currie PJ. 2019. Hagfish from the Cretaceous Tethys Sea and a
- reconciliation of the morphological–molecular conflict in early vertebrate phylogeny.
- *Proceedings of the National Academy of Sciences* 116:2146–2151. DOI:
- 10.1073/pnas.1814794116.
- Moser ML, Butzerin JM, Dey DB. 2007. Capture and collection of lampreys: the state of the
- science. *Reviews in Fish Biology and Fisheries* 17:45–56. DOI: 10.1007/s11160-006-9037-3.
- Moser ML, Ocker PA, Stuehrenberg LC, Bjornn TC. 2002. Passage Efficiency of Adult Pacific
- Lampreys at Hydropower Dams on the Lower Columbia River, USA. *Transactions of the*
- *American Fisheries Society* 131:956–965. DOI: 10.1577/1548-8659.
- Moussalli E, Hilborn R. 1986. Optimal Stock Size and Harvest Rate in Multistage Life History
- Models. *Canadian Journal of Fisheries and Aquatic Sciences* 43:135–141. DOI: 10.1139/f86-
- 014.
- Murdoch SP, Docker MF, Beamish FWH. 1992. Effect of density and individual variation on
- growth of sea lamprey (*Petromyzon marinus*) larvae in the laboratory. *Canadian Journal of*
- *Zoology* 70:184–188. DOI: 10.1139/z92-027.

- Potter IC, Gill HS, Renaud CB, Haoucher D. 2015. The Taxonomy, Phylogeny, and Distribution
- of Lampreys. In: Docker MF ed. *Lampreys: Biology, Conservation and Control*. Dordrecht:
- Springer Netherlands, 35–73. DOI: 10.1007/978-94-017-9306-3_2.
- Purvis HA. 1980. Effects of Temperature on Metamorphosis and the Age and Length at
- Metamorphosis in Sea Lamprey (*Petromyzon marinus*) in the Great Lakes. *Canadian Journal of Fisheries and Aquatic Sciences* 37:1827–1834. DOI: 10.1139/f80-225.
- R Core Team. 2017. *R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing*.
- Riemer SD, Wright BE, Brown RF. 2011. Food habits of Steller sea lions (Eumetopias jubatus) off Oregon and northern California, 1986–2007.
- Rodríguez‐Muñcoz R, Nicieza AG, Braña F. 2001. Effects of temperature on developmental
- performance, survival and growth of sea lamprey embryos. *Journal of Fish Biology* 58:475–486.
- 855 DOI: 10.1111/j.1095-8649.2001.tb02266.x.
- Rodriguez-Munoz R, Nicieza AG, Brana F. 2003. Density-dependent growth of sea lamprey larvae: evidence for chemical interference. *Functional Ecology*:403–408.
- Roffe TJ, Mate BR. 1984. Abundances and feeding habits of pinnipeds in the Rogue River, Oregon. *The Journal of Wildlife Management*:1262–1274.
- Saltelli A, Ratto M, Andres T, Campolongo F, Cariboni J, Gatelli D, Saisana M, Tarantola S.
- 2008. *Global sensitivity analysis: the primer*. John Wiley & Sons.
- Schultz LD, Mayfield MP, Wyss LA, Sheoships GT, Clemens BJ, Chasco B, Schreck CB. 2014.
- The distribution and relative abundance of spawning and larval Pacific Lamprey in the
- Willamette River basin. *Final Report to the Columbia River Inter-Tribal Fish Commission,*
- *Portland, Oregon*.
- Shimeld SM, Donoghue PC. 2012. Evolutionary crossroads in developmental biology:
- cyclostomes (lamprey and hagfish). *Development* 139:2091–2099.
- Starcevich SJ, Gunckel SL, Jacobs SE. 2014. Movements, habitat use, and population characteristics of adult Pacific lamprey in a coastal river. *Environmental biology of fishes* 97:939–953.
- Swink WD. 1993. Effect of Water Temperature on Sea Lamprey Growth and Lake Trout
- Survival. *Transactions of the American Fisheries Society* 122:1161–1166. DOI: 10.1577/1548-
- 8659(1993)122<1161:EOWTOS>2.3.CO;2.
- Torgersen CE, Close DA. 2004. Influence of habitat heterogeneity on the distribution of larval
- Pacific lamprey (*Lampetra tridentata*) at two spatial scales. *Freshwater Biology* 49:614–630.
- 876 DOI: 10.1111/j.1365-2427.2004.01215.x.
- Urban DL. 2005. Modeling ecological processes across scales. *Ecology* 86:1996–2006. DOI: 10.1890/04-0918.

- Wargo Rub AM, Som NA, Henderson MJ, Sandford BP, Van Doornik DM, Teel DJ, Tennis MJ,
- Langness OP, van der Leeuw BK, Huff DD. 2019. Changes in adult Chinook salmon
- (*Oncorhynchus tshawytscha*) survival within the lower Columbia River amid increasing pinniped
- abundance. *Canadian Journal of Fisheries and Aquatic Sciences* 76:1862–1873.
- Weitkamp LA, Tuttle V, Ward EJ, Kamikawa D, Billings A, Buchanan J, Hess JE. 2023. Pacific
- Lamprey and Western River Lamprey marine ecology: Insight from new ocean collections.
- *North American Journal of Fisheries Management* 43:1492–1510. DOI: 10.1002/nafm.10936.
- Whitlock SL, Schultz LD, Schreck CB, Hess JE. 2017. Using genetic pedigree reconstruction to
- estimate effective spawner abundance from redd surveys: an example involving Pacific lamprey
- (*Entosphenus tridentatus*). *Canadian Journal of Fisheries and Aquatic Sciences* 74:1646–1653.
- DOI: 10.1139/cjfas-2016-0154.
- Zerrenner A. 2004. Effect of Density and Age on Larval Sea Lamprey Growth and Survival in
- Three Lake Champlain Streams. *Journal of Freshwater Ecology* 19:515–519. DOI:
- 10.1080/02705060.2004.9664927.

894 Table 1: User-defined parameters and default values with references. All references are for Pacific lamprey (*Entosphenus tridentatus*),

895 unless otherwise stated in the column, "Reference River Basin."

896

898 Table 2: Distributions and hyperparameters for global sensitivity analyses (GSA). The broad GSA explored a larger parameter space,

899 relying on a uniform distribution across the entire range of possible values $(0 - 1$ in most cases). The restricted GSA explored a more

900 constrained, and realistic, parameter space.

Table 3: Case study management scenario examples.

- Figure 1: (A) Artistic diagram of the Pacific lamprey anadromous life cycle. Artwork by Monica R. Blanchard. (B) Images depicting
- Pacific lamprey life stages with details on notable traits at various stages of development. Images by Ralph Lampman.

This information product has been peer reviewed and approved for publication as a preprint by the U.S. Geological Survey.

908 Figure 2: Conceptual diagram of the Lamprey Life Cycle Model (LLCM). Grey boxes indicate 909 specific life-stage and age-class combinations. Black arrows indicate processes that occur within 910 a given year (e.g., reproduction, transformation, etc.), while red arrows indicate processes that 911 occur between years (e.g., aging). Purple symbols indicate life cycle parameters (see Table 1). 912 Black icons indicate potential management scenarios to explore (1 = summer [drought] mortality 913 and carrying capacity, $2 =$ winter [flood/scouring] mortality and carrying capacity, $3 =$ hatchery 914 additions, $4 =$ downstream barriers, $5 =$ downstream [transformer] translocation, $6 =$ upstream 915 barriers, $7 =$ upstream [adult] translocation).

916

 Figure 3: Initialization of the model with 5 different starting values for initial spawners leads to similar results after a conservative burn-in period of 50 years. Colors indicate mean values for 100 stochastic runs of each initial starting value for spawners, while grey lines represent variation (uncertainty) in stochastic simulation runs. It is important to note that the absolute number of spawners is not meaningful outside the context of the particular values selected for model life cycle parameters (including arbitrary carrying capacity values that do not represent any particular stream).

 Figure 4: Broad global sensitivity analysis (GSA) with parameter space sampled across a full range of possible values (see Table 2). Importance of life cycle parameters in predicting the number of spawners (% change in mean squared error, MSE). We used a global sensitivity analysis (GSA) and a Latin hypercube to systematically vary all life cycle model parameters simultaneously across a broad range of possible values. We then ran the deterministic versions of the life cycle model for 100 years with 100 different parameter sets. With the resulting model output for the number of spawners, 100 random forests, each with 501 trees, were used to regress the number of spawners across the 100 life cycle parameter sets for the 44 life cycle variables (only those with positive % change in MSE are shown on the y-axis here, others are omitted for brevity). Dots indicate mean parameter importance and error bars indicate 95% confidence

- intervals (as 1.96 x SE). Positive x-axis values indicate an improvement in model skill (when y-
- axis parameter is not randomly permuted) and negative values indicate a reduction in model skill
- 938 (relative to random permutations). $K =$ carrying capacity; $SUR =$ swim upriver probability for
- each juvenile ocean age-class.

 Figure 5: Restricted global sensitivity analysis (GSA) with parameter space sampled across more reasonable values (see Table 2). Importance of life cycle parameters in predicting the number of spawners (% change in mean squared error, MSE). We used a global sensitivity analysis (GSA) and a Latin hypercube to systematically vary all life cycle model parameters across a range of reasonable values. We then ran the deterministic versions of the life cycle model for 100 years with 100 different parameter sets. With the resulting model output for the number of spawners, 100 random forests, each with 501 trees, were used to regress the number of spawners across the 100 life cycle parameter sets for the 44 life cycle variables (only those with positive % change in MSE are shown on the y-axis here, others are omitted for brevity). Dots indicate mean parameter importance and error bars indicate 95% confidence intervals (as 1.96 x SE). Positive x-axis

- values indicate an improvement in model skill (when y-axis parameter is not randomly
- permuted) and negative values indicate a reduction in model skill (relative to random
- 954 permutations). $K =$ carrying capacity; $SUR =$ swim upriver probability for each juvenile ocean
- age-class.

+ Barrier.US.Trans + Barriers.US.Hatch + Barriers.US.Trans.Hatch Scenario Barrier.US **Baseline**

 Figure 6: Hypothetical management scenarios as examples of model utility. Scenario 1 (pink): "Baseline" (default parameters, including no barriers). Scenario 2 (red): "Barrier.US" addition of upstream barrier with 60% passability. Scenario 3 (olive): "Barrier.US.Trans" same as Scenario $2 +$ translocating 100 adults (when available) above the barriers. Scenario 4 (green): "Barriers.US.Hatch" same as Scenario 2 + 1000 hatchery transformers released into river. Scenario 5 (blue): "Barriers.US.Trans.Hatch" combination of Scenario 3 and 4; that is, upstream 964 barrier with 60% passability + translocating 100 adults (if available) above the barriers $+1000$ hatchery transformers released into river. It is important to note that the absolute number of spawners is not meaningful outside the context of the particular values selected for model life cycle parameters (including arbitrary carrying capacity values that do not represent any particular stream). Instead, we are focusing on the relative values between scenarios to explore management alternatives.