

1 **Spider colour polymorphism is shaped by precipitation, not**
2 **ambient temperature**

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6

7 **ABSTRACT**

8 Colour polymorphism, the presence of multiple colour variants within a population, is a
9 common example of intraspecific phenotypic variation and has served as a model for
10 studying drivers of diversity. Climatic factors can influence the distribution and abundance of
11 colour variants, yet research often focuses on lineages where sexual selection covaries with
12 the climate-colouration associations. Research has also focussed disproportionately on
13 vertebrates and a few insects, neglecting other taxa where polymorphism is widespread, like
14 arachnids. Here, we investigated climatic factors influencing colour variation in a widely
15 distributed colour polymorphic spider using a combination of controlled experiments, field
16 measurements and tests for macroecological associations. We showed in the lab and field
17 that dark colouration is unlikely to provide biologically relevant thermal benefits; yet different
18 colour morphs occupy distinct climatic niches across Australia. Precipitation, rather than
19 temperature, appears to be the strongest driver of colour morph frequencies: there were
20 higher frequencies of dark individuals in regions with higher precipitation while orange
21 individuals were more common in drier areas. We discuss potential mechanisms related to
22 background matching or pathogen protection in humid environments. Our findings highlight
23 the role of climatic factors in shaping colour variation, and demonstrate the value of
24 combining macroecological, field, and laboratory approaches.

25

26 **KEYWORDS**

27 Colour variation, Gloger's rule, humidity, geographic variation, thermal melanism hypothesis

28

29 INTRODUCTION

30 Intraspecific variation has extensive evolutionary and ecological implications and is the raw
31 material for adaptation (Des Roches et al., 2018). For instance, populations with greater
32 variation generally have increased resilience to perturbations and enhanced ability to adapt
33 to rapidly changing conditions (González-Suárez & Revilla, 2013; Moran et al., 2016; Palacio
34 & Clark, 2023). Identifying the drivers that produce and maintain variation is essential to
35 understand eco-evolutionary processes, including patterns of species diversity, resilience,
36 and responses to environmental change (Bolnick et al., 2011). Environmental factors can
37 shape variation within and between populations; but these are commonly studied in isolation.
38 For example, many studies have examined large-scale correlations between phenotypic
39 traits and climate variables among populations. Conversely, many studies consider adaptive
40 variation within populations (Mclean & Stuart-Fox, 2014; Svensson, 2017). However, to
41 understand how environmental factors shape intraspecific variation, it is important to
42 integrate both macroecological patterns and within-population dynamics.

43

44 Species with discrete within-population phenotypic variation (i.e., polymorphism) provide an
45 ideal model to understand how environment factors shape intraspecific variation (Ford, 1945;
46 Gray & McKinnon, 2007; Jamie & Meier, 2020). Specifically, environmental factors can
47 determine the presence or frequency of discrete phenotypes within and between populations
48 (Endler, 1977; Kassen, 2002; Dreiss et al., 2012). For instance, in some colour polymorphic
49 species, morph frequencies — and even the presence of polymorphism — can vary strongly
50 across environmental gradients (Mullen & Hoekstra, 2008; Wittkopp et al., 2011; Roulin et
51 al., 2011; Dreiss et al., 2012a; Pérez i de Lanuza et al., 2018). In the black sparrowhawk,
52 *Accipiter melanoleucus*, for example, the frequency of dark and white colour morphs varies
53 clinally with light environment due to differential crypsis (Tate et al., 2016), while in oldfield
54 mice, pigmentation varies along a vegetation gradient (Mullen & Hoekstra, 2008). It is no
55 surprise that these and many other examples involve colour polymorphic species because

56 colour is easily scored and can be functionally associated with different environmental
57 variables.

58

59 Two of the most important climatic gradients associated with colour variation are
60 temperature and humidity. Correlations between temperature or humidity have been
61 observed in a wide range of taxa and are known as Bogert's rule (or the thermal melanism
62 hypothesis) and Gloger's rule respectively (Goldenberg et al., 2022). According to the
63 thermal melanism hypothesis (Clusella Trullas et al., 2007; Clusella-Trullas et al., 2008)
64 darker individuals should be more abundant in environments with low ambient temperatures,
65 given the thermoregulatory benefits of dark colours, especially for ectotherms (Watt, 1968;
66 Meredith & Sarna, 2006; Castella et al., 2013). Even though this pattern has been observed
67 in multiple species (Clusella-Trullas et al., 2008; Broennimann et al., 2014; Azócar et al.,
68 2016; Bishop et al., 2016; Pinkert et al., 2017; Heidrich et al., 2018; Dalrymple et al., 2018;
69 Hantak et al., 2022), there are cases where dark colouration is associated with other functions
70 or climatic factors (Roulin, 2016; Smith et al., 2016; Delhey et al., 2019; Lopez et al., 2021;
71 Pepi et al., 2022; Passarotto et al., 2022; Hastings et al., 2023). Additionally, in many cases,
72 we ignore how ambient temperature is mechanistically associated with the presence of dark
73 colouration and whether the degree of colour variation in the wild can lead to biologically
74 relevant differences in temperature regulation across morphs. In other words, large-scale
75 correlations are often not supported by experimental evidence.

76

77 Similarly to temperature, humidity shapes the geographic distribution of colouration in many
78 taxa (Friedman & Remeš, 2017; Delhey, 2019; Cerezer et al., 2020; Lopez et al., 2021;
79 Marcondes et al., 2021; Kang et al., 2021). Darker individuals appear to be more abundant
80 in places with higher humidity or precipitation than in drier places (Delhey, 2019). This
81 pattern does not have a clear causal explanation; it is possible that it emerges as a

82 consequence of improved background matching (Stevens & Merilaita, 2008; Cheng et al.,
83 2018), pathogen protection (Mackintosh, 2001; Nosanchuk & Casadevall, 2003) or a
84 consequence of pleiotropic effects (Ducrest et al., 2008). In addition to the lack of clarity on
85 the mechanisms behind Gloger's rule, there are some taxa, such as amphibians and
86 arachnids, that are notably underrepresented in the available empirical evidence supporting
87 these macroecological patterns (reviewed in Delhey, 2019).

88

89 Most of our understanding of how climatic variables shape colour variation comes from
90 vertebrate taxa where sexual selection can strongly affect the presence and maintenance of
91 colour polymorphism (Roulin & Bize, 2007; Wellenreuther et al., 2014). This means that the
92 interpretation of the association between climate and coloration needs to account for the
93 confounding effect of sexual selection (Lancaster et al., 2017; Pérez i de Lanuza et al.,
94 2018). Web-building spiders provide an opportunity to explore climatic drivers of phenotypic
95 diversity interpedently of sexual selection. They are a diverse group (> 6,000 species in the
96 families Theridiidae and Araneidae alone) with at least 140 colour polymorphic species
97 (Salgado-Roa et al., 2024), yet they have limited visual capabilities. Most web-building
98 spiders examined to date have only one photoreceptor (Yamashita, 1985; Tiedemann et al.,
99 1986), suggesting they may lack colour vision. This visual limitation implies that colour
100 signals might not play a significant role in sexual selection for these species. As a result,
101 their coloration may be shaped entirely by environmental factors rather than intraspecific
102 interactions.

103

104 Here, we combined controlled experiments, field measurements and macroecological
105 analyses to examine the climatic drivers that influence the composition and abundance of
106 different colour variants across multiple populations in a widespread colour polymorphic
107 spider (*Austracantha minax*) with three different morphs (Figure 1). We first explored the

108 thermal benefits of dark colouration by comparing heating rates of light and dark morphs
109 under a solar simulator in the laboratory and comparing body temperatures of the two
110 morphs in the field. At a broader scale, we investigated which climatic variables could
111 explain colour variation in the Christmas spider within and across populations. We compared
112 the climatic niche of the colour morphs using over 1254 field observations and public records
113 from databases and museum specimens. Considering that the frequency of the morphs can
114 change throughout the species' distribution, we also quantified morph frequency across 49
115 populations in Australia. Our study reconciles experiments conducted under controlled
116 conditions with field observations and macroecological associations to understand the
117 drivers of phenotypic diversity at the intraspecific level in an unexplored group.

118

119 MATERIALS AND METHODS

120 Study species

121 *Austracantha minax*, the Christmas spider, is a colour polymorphic species widely distributed
122 in Australia. Females exhibit two main colour morphs: one is completely dark on both its
123 dorsal and ventral sides; and the other presents black, white, and yellow on the dorsal side,
124 and a mosaic yellow pattern ventrally (Waldock, 1991). A third colour morph has been
125 described for this species; however the reports are limited to only a few places in Western
126 Australia (Hogg, 1914; Waldock, 1991). This colour variant appears to be orange and white
127 on its dorsal side and completely orange on its ventral side (Waldock, 1991). Based on
128 previous museum and field observations, as well as early studies (Waldock, 1991; Elgar &
129 Bathgate, 1996; Mascord, 1996; Lloyd & Elgar, 1997), females of the Christmas spider tend
130 to be bigger, more abundant, live longer, and exhibit clearer phenotypic variation than males.
131 These observations prompted us to explore the effect of climatic variables on the colour
132 polymorphism of *A. minax*, focusing exclusively on females.

133

134 Heating rates experiment

135 Correlations between colour variation and climate can be driven by several factors, one of
136 the most important being thermal needs. To investigate potential functional explanations for
137 ecogeographical patterns, we quantified heating rates in the two most common morphs —
138 the black morph and the black-and-white morph — under controlled conditions. The
139 equipment and protocol for this experiment are based on and fully described in Wang et al.
140 (2021) and Ospina-Rozo et al. (2022). For this experiment, we used a solar simulator
141 (Sciencetech UHE-NS) emitting an energy intensity of 1,000 W/m², equivalent to the solar
142 radiation typical at an equatorial site during a clear day around solar noon. The light source
143 mimics the power and irradiance spectrum of the sun, spanning 300 nm to 1700 nm
144 (AM1.5G). To measure heating rates, we enclosed the samples within a sealed glass

145 thermal chamber, illuminated from above by the light source (Supplementary figure 1). This
146 chamber was linked to a cooling system (thermofisher accel 250 lc) to minimise conduction
147 or convection effects during experimentation by maintaining air temperature at $20 \pm 0.5^{\circ}\text{C}$
148 (Supplementary figure 1).

149

150 To measure the change in spider temperatures, we sacrificed the spiders immediately before
151 the experiment using a CO_2 chamber, then inserted a thermocouple (approximately 5 mm
152 TP-K01, K type, Center Technology Corp., Taiwan) into the spinnerets of the spider. Another
153 thermocouple was positioned at the same level as the sample to monitor the air temperature
154 inside the chamber. Both thermocouples were connected to a thermometer (Center 521,
155 Center Technology Corp., Taiwan), recording temperatures every 10 seconds for 5 minutes.

156

157 We measured a total of 20 black individuals and 20 black-and-white individuals on both their
158 ventral and dorsal sides. To compare heating rates between the colour morphs on each side,
159 we employed two methods. One method involved calculating the total temperature
160 differences between the initial and final measurements after 5 minutes (ΔT), while the other
161 measured the maximum change in temperature in 20-second intervals ($\text{maxHR} = \Delta T/\text{sec}$).
162 Additionally, considering that larger objects are expected to heat up more slowly, we
163 recorded the weight of each sample before the start of each trial. To test for differences in
164 heating rates between morphs, we built linear models per side (dorsal/ventral) in R v4.2.0 (R
165 Core Team, 2021) using either ΔT or maxHR as response variables and colour morph (black
166 or black-and-white) and weight as predictors. The assumptions of the linear model were
167 verified using diagnostic plots.

168

169 ***Field thermal measurements***

170 To explore if our measurements of heating rates under controlled conditions were reflected in
171 the field and biologically relevant, we compared the body temperature of the black and
172 black-and-white individuals in the field. We measured the body temperature of 15 black
173 females and 15 black-and-white females in a population near Ballarat, Victoria, Australia
174 (latitude: -37.8215, longitude:143.7585). To ensure we measured all individuals under the
175 most similar conditions possible, we conducted measurements on a cloudless day at noon
176 along a transect of less than 50 meters.

177

178 We measured the body temperature of the individuals on their dorsal and ventral side using
179 an infrared thermal camera (FLIR t540). Raw images were processed with the manufacturer
180 software (FLIR Tools v6.4) to extract average temperatures per side. Simultaneously, we
181 recorded wind speed with an Anemometer (BTMETER BT-100), air temperature with a
182 thermometer (Center 521, Center Technology Corp., Taiwan) and the luminosity level
183 (measured in lux) using a digital illuminance device (Model: LX1330B-Dr. meter; spectral
184 sensitivity ranging from 400 to 700 nm). These devices were positioned parallel to the
185 individuals. We also measured the height of the web of every spider.

186

187 We analysed each body side (dorsal and ventral) independently by building linear models in
188 R v4.2.0 (R Core Team, 2021). The models included body temperature as response variable
189 and colour morph (black or black-and-white), ambient temperature, wind speed, luminosity,
190 and web height as predictors. We checked for collinearity between the predictors using the
191 function *check_collinearity* from the R package *performance v0.10* (Lüdecke et al., 2021)
192 and calculating their Pearson's correlation. Diagnostic plots were used to confirm the
193 assumptions of the linear model.

194

195 ***Geographical records data collection***

196 To test whether and which climatic variables can explain colour variation in *A. minax*, we
197 obtained geographical records from three different sources. First, we visited four of the main
198 arachnological collections in Australia: Western Australian Museum, Queensland Museum,
199 Australian Museum, and Museums Victoria, and extracted coordinates for a total of 357
200 specimen records. Second, we collated all the research-grade records of the species on
201 iNaturalist on the 11th of November 2023. All these records had images associated and
202 consensus about their identification. We removed records with poor-quality pictures and a
203 coordinate uncertainty > 1 km. This resulted in a total of 1829 records with photographs from
204 iNaturalist. Finally, we also added 47 new geographical records from our own surveys of
205 locations across southeast and southwest Australia that had suitable environments for the
206 spider. Our final dataset consisted of 1,254 records, 1097 records for the black-and-white
207 colour morph, 123 records for the black morph, and 34 records of the orange morph (Figure
208 1).

209

210 ***Climatic niche variation among colour morphs***

211 We tested if the three colour morphs of the Christmas spider differ in their realised climatic
212 niche (Peterson et al., 2011). To extract climatic data that represents the conditions in which
213 adults are found in the field, we focused on the months when the species shows the highest
214 activity levels. Based on all the records with associated collection dates, we defined the
215 months of peak activity by excluding those months that had fewer than 5% of the reports.
216 We found that 95% of the records were from October to May, so we used those months to
217 extract climatic data.

218

219 Using the months of activity we downloaded climatic data from CHELSA v2.1 (Karger et al.,
220 2017). This resource offers monthly climatic information from 1979 to 2019, derived by
221 downscaling ERA-Interim climatic reanalysis to a resolution of 1 km. We selected this global

222 dataset over others because its spatial resolution is higher (e.g. Era5 has a spatial resolution
223 of approximately 31 km). From this dataset, we downloaded six climatic variables that we
224 hypothesised could be linked to morph variation: mean daily maximum air temperature
225 (extracted automatically at 2 meters), mean daily minimum air temperature (extracted
226 automatically at 2 meters), precipitation, surface downwelling shortwave flux in air (i.e. solar
227 radiation), potential evapotranspiration, and climatic moisture index. To account for temporal
228 variation or biases due to extreme conditions, we generated raster layers for Australia for
229 each variable by averaging the values for the months of peak activity of the species (October
230 to May) over the last decade of available data (2009-2019). The values for each record were
231 extracted using the function *extract* from the R package *terra* v1.7 (Hijmans, 2023).

232

233 Using the climatic data, we implemented univariate and multivariate comparisons between
234 the colour morphs of the Christmas spider. For the univariate comparisons, we computed the
235 density profiles per colour morph of every climatic variable and Tukey honest significant
236 difference (Tukey HSD) tests to determine which of the colour morphs were significantly
237 different from one another across the six climatic variables. For the multivariate analysis, we
238 implemented a COUE (Centroid shift, Overlap, Unfilling and Expansion) scheme (Guisan et
239 al., 2014). This framework is commonly used to test differences in the climatic niche of
240 different populations of the same species (Collart et al., 2021; Farquhar et al., 2022; Y. Wang
241 et al., 2023); for example, to compare invasive and native ranges of invasive species (Bates
242 & Bertelsmeier, 2021).

243

244 For the COUE scheme, we first defined the geographical range of each colour morph by
245 adding a buffer of 100 km radius to every geographical record of each morph (Following,
246 Pittiglio et al., 2018; Farquhar et al., 2022; Whitford et al., 2024). Using the geographical
247 range of each colour morph, we defined the spatial extent of their environmental background

248 by intersecting the range of each morph with a layer of the terrestrial ecoregions of the
249 world, which were obtained from Dinerstein et al. (2017; <https://ecoregions.appspot.com/>).
250 The black-and-white morph occupied a greater number of ecoregions (34 ecoregions),
251 followed by the black morph (21 ecoregions) and orange morph (17 ecoregions). Then, we
252 randomly sampled 10,000 background points within all the ecoregions occupied by each
253 colour morph (30,000 background points). These points represent the environmental
254 background, which is the available environment that might be suitable for each colour
255 morph, but that is not in our sampling (Barbet-Massin et al., 2012). These steps were done
256 using functions from the R package *sf* (Pebesma, 2018). We repeated this analysis adding a
257 buffer of 50 km radius to every geographical location, instead of 100 km, obtaining
258 equivalent qualitative results.

259

260 After sampling the background random points, we extracted the climatic values for each of
261 these background records from the CHELSA v2.1 layers, and combined them with the
262 climatic values of the geographical records of the colour morphs (i.e. presence records). We
263 summarized climatic variation from all (presence and background) records in two principal
264 components (PCs) which explained 95.4% of the climatic variation and represented the
265 environmental space of the whole species (i.e including both observations and possible
266 suitable areas for all the morphs; Supplementary figure 2). This information was used as an
267 input for the R package *ecospat* (Di Cola et al., 2017), where we implemented the COUE
268 scheme to explore the similarity of the climatic niche of the colour morphs. First, we created
269 grids of 100 x 100 cells per colour morph that represented the occurrence density of each
270 morph over the environmental space of the whole species (summarized in the two principal
271 components). Then, using the grid topology of all the colour morphs, we compared their
272 climatic niche estimating the D index (Schoener, 1968) and the similarity statistic I (Warren
273 et al., 2008) between all the colour morphs with the R package *ecospat* (Di Cola et al.,

274 2017). The values of D and I range between 0 and 1, where 0 indicates no niche overlap
275 between the niche occupied by colour morphs, and 1 represents complete overlap.

276

277 To evaluate the statistical significance of D and I , we performed tests of niche equivalency
278 and similarity using 10,000 permutations (pseudo-replicates) to create a null distribution of
279 similarity values based on the background environment of each morph. This allowed us to
280 compare the observed values of D and I against a null distribution to implement a one-tailed
281 hypothesis test. We tested this for niche conservatism (niche overlap is more
282 equivalent/similar than random expectations) and for niche divergence (niche overlap is less
283 equivalent/similar than random expectations).

284

285 ***Climatic factors shaping colour morph frequency***

286 The analysis of geographical distribution of the colour morphs described above provides a
287 broad scale comparison between the environments where morphs are found, but multiple
288 morphs can be found in some locations. To gain insights into the drivers of abundance for
289 each colour morph, we measured their frequency in the field. We sampled 49 locations
290 encompassing 36 degrees of longitude and 6 degrees of latitude in south-eastern and south-
291 western Australia (Figure 2). Taking advantage of the semi-social behaviour of the species,
292 we searched for aggregations of individuals of *A. minax* at these locations. When these
293 aggregations were found, we conducted an exhaustive search aiming for a minimum of 50
294 females within a radius of 1km. We categorised the individuals found at each location as
295 black, black-and-white, or orange (Figure 1). In total, we counted 15,214 spiders, with a
296 mean of 310.49 spiders per location, a minimum of 85 individuals and a maximum of 817.

297

298 We used the colour morph frequency information from the field to test whether the variation
299 in morph frequency was associated with climatic factors. The climatic information in this case
300 was obtained from the Australian Gridded Climate Data / Australian Water Availability Project
301 dataset (AGCD/AWAP; Jones et al., 2009), rather than from CHELSA v2.1. We decided to
302 use this dataset instead of the global dataset (CHELSA v2.1), because AGCD/AWAP offers
303 higher regional specificity and temporal specificity (daily data per hour), which is ideal for this
304 finer-scale analysis. We were unable to use this dataset for the continental climatic analyses
305 (climatic niche variation among colour morphs) because extracting data from this dataset for
306 such a high number of records (~32,000) was extremely computationally intensive. In the
307 supplementary material we show that both CHELSA and AWAP datasets are highly
308 correlated and lead to similar conclusions.

309

310 From the AGCD/AWAP, we downloaded the daily values of the following five climatic
311 variables: vapor pressure, minimum temperature, maximum temperature, precipitation, and
312 solar radiation. We also accounted for temporal bias by calculating the average of each
313 location for the months of peak activity of the species (October to May) over the last decade
314 of available data (2011-2021). Before the analyses, we assessed potential collinearity
315 between the climatic variables by exploring their Pearson correlation (Supplementary figure
316 3). Following that, we built a dissimilarity matrix with the absolute values of the correlation (1-
317 correlation values) to perform a hierarchical clustering analysis with the function *hclust* in R
318 v4.2.0 (R Core Team, 2021). We considered a pair of climatic variables as independent if
319 they had a pairwise distance > 0.5 (Supplementary figure 4).

320

321 We constructed multiple matrix regression (MMR) models using non-collinear variables with
322 the R package *ecodist* v2.1 (Goslee & Urban, 2007). The predictors comprised distance
323 matrices for each climatic variable, calculated as the Euclidean distance between pairs of

324 locations, along with a matrix of geographic distances between sampling locations. The latter
325 calculation was performed using the R package *geosphere* v1.5 (Hijmans, 2016). The
326 response variable was the Euclidean distance in colour morph frequencies between pairs of
327 locations (e.g., between locations A and B), calculated as follows:

$$328 \quad dist(A, B) = \sum_{i=1}^n \sqrt{(fA_i - fB_i)^2}$$

329 Here, fA_i and fB_i represent the relative frequency of morph i in locations A and B,
330 respectively. Predictors were log-transformed if their distribution significantly deviated from
331 normality. All matrices were scaled and centered before analysis to ensure comparable
332 linear coefficients (Wang, 2013).

333

334 We conducted two MMR models considering the three colour variants of the Christmas
335 spider to avoid including highly correlated predictors in the same model. We found that
336 vapour pressure was highly correlated with minimum temperature, and maximum
337 temperature correlated with solar radiation, while precipitation was not strongly correlated
338 with the other variables. Hence, the first model included precipitation, minimum temperature,
339 maximum temperature, and geographical distance as predictors. For the second model, we
340 used precipitation, vapor pressure, solar radiation, and geographical distance as predictors.
341 If either of these models contained a statistically significant predictor, we evaluated the same
342 model using pair-wise comparisons between morphs (instead of the three morphs at once)
343 to examine if certain variables strongly influenced frequency changes in specific morphs.
344 Therefore, in this case the estimation of the Euclidean distance in colour morph frequencies
345 was done the same way as explained earlier but excluding one morph from the calculation.

346

347 Given that measures of distance can reduce the statistical power to test associations
348 between variables (Legendre & Fortin, 2010), we also built linear models for each colour

349 morph using morph frequency as the response and the climatic variables that had a
350 significant effect in the MMR models as predictors. We applied the logit transformation to the
351 relative frequency of the colour morph in each location as the response variable. We
352 examined whether the model assumptions were met by plotting the normality of residuals
353 and assessing heteroscedasticity. The same methodology described in this section was
354 repeated using the CHELSA v2.1 dataset to validate the results across climatic datasets.

355

356 **RESULTS**

357 **Thermal differences between black and black-and-white morph**

358 We found that under controlled conditions, the black morph heated faster than the black-
359 and-white morph on both the dorsal ($F_{1,97} = 6.44$, P-value = 0.01, $R^2 = 0.18$) and ventral
360 sides ($F_3 = 7.24$, P-value = 0.01, $R^2 = 0.1$). The same pattern was supported by the two
361 methods of calculating heating rate (Supplementary Table 1-2 and Figure 3). The difference
362 in heating rate was small, however, varying in less than 1 degree after 5 minutes of solar
363 exposure on both ventral and dorsal sides. Our field measurements revealed no temperature
364 difference between the black and black-and-white morphs on either the dorsal (Figure 3) or
365 ventral sides (Figure 3). Ambient temperature, wind speed and light intensity appear to be
366 the most important factors explaining the variation in body temperature in our field data
367 (Supplementary Table 3).

368

369 ***Climatic niche variation among colour morphs***

370 In the different sources of geographical records that we accessed (public records, museum
371 collections, and field observations), the black-and-white morph was the most prevalent
372 colour morph (87.5%), followed by the black morph (9.8%). The orange morph was rare, with
373 a prevalence of only 2.7% (Figure 1).

374

375 Visual examination of the climatic niche of all colour morphs in a density space per climatic
376 variable, along with the pairwise significance test (Tukey HSD), suggests that the black and
377 orange morphs exhibit differences across all variables except for mean minimum
378 temperature (Figure 4, Supplementary Table 4, Supplementary Figures 5 & 6). We also
379 observed a similar pattern when comparing the black-and-white with the orange morph;
380 however, they showed similar values of mean maximum temperature. (Supplementary Table
381 4). Notably, the black and black-and-white morphs differ in potential evapotranspiration,
382 precipitation, solar radiation and air temperature, but not in climatic moisture index
383 (Supplementary Table 4).

384

385 We found that the climatic niche of the orange morph was the most dissimilar, presenting low
386 to medium overlap values when compared with the black ($D=0.25$, $I=0.45$) and black-and-
387 white ($D=0.21$, $I=0.39$) morphs (Supplementary Table 5). In contrast, the climatic niches of
388 the black and the black-and-white morphs were more similar, with high to medium overlap
389 values ($D=0.64$, $I=0.82$). These results are consistent with a visual inspection of the density
390 of occurrences in the environmental space (reduced multivariate dimensions to two principal
391 components) where the black-and-white morph has a higher overlap with the black morph
392 than with the orange morph (Figure 2). When comparing the environmental densities of the
393 black and orange morphs, the segregation is even more clear (Figure 2); the black morph is
394 more frequent in areas with higher precipitation and climatic moisture index, and the orange
395 morph has a higher density of points in areas with more solar radiation and higher
396 temperature (Figure 2).

397

398 Our test for niche conservatism and niche divergence supports the idea that the black-and-
399 white and black morph share more similar niches than expected by chance ($D=0.64$, $I=0.82$,

400 P-value < 0.001; Supplementary Table 5). Conversely, we found that the orange morph's
401 climatic niche is less equivalent than expected by chance when compared to the black
402 morph (D=0.25, I=0.45, P-value= 0.003; Supplementary Table 5) and the black-and-white
403 morph (D=0.21, I=0.39, P-value= 0.003; Supplementary Table 5).

404

405 ***Climatic factors shaping colour morph frequency***

406 In the field, we observed that the black-and-white morph was the most abundant colour
407 variant in most locations, exhibiting a mean relative frequency of 88% (min= 43.5% to max=
408 100%). The black morph ranked as the second most abundant morph in our sampling,
409 showing a mean relative frequency of 11.8% (0.3% to 48.8%). Lastly, the orange morph
410 appeared as the least frequent, with a mean relative frequency of 9% (0.18% to 56%).
411 (Figure 2).

412

413 Our MMR models indicate that precipitation is the strongest predictor of morph frequency
414 change across our sampling, and there was little effect of geographical distance and
415 temperature (Figure 4; supplementary table 6). Pairwise models suggest that the effect of
416 precipitation is reduced ($r_{\text{partial}}=0.087$, P-value=0.051) when only comparing black and black-
417 and-white morphs. However, the effect of precipitation is stronger when comparing only the
418 orange morph and black morph ($r_{\text{partial}}=0.37$, P-value<0.001; supplementary table 7) or the
419 orange morph and the black-and-white morph ($r_{\text{partial}}=0.16$, P-value<0.001; supplementary
420 table 7). Similar results were obtained with the CHELSA dataset (supplementary table 7).

421

422 Given that precipitation had the strongest effect in the analysis above, we explored the effect
423 of precipitation on the frequency of each colour variant using linear models. Precipitation did
424 not predict the frequency of the black-and-white morph ($F_{5,49}=2.12$, P-value = 0.15, $R^2=0.02$;

425 Figure 2), but had a significant positive effect on the frequency of the black morph
426 ($F_{17,15}=14.77$, P-value < 0.001, $R^2=0.30$), which was more abundant in places with higher
427 precipitation (Figure 2). Conversely, the orange morph seems to follow the opposite pattern,
428 with higher relative frequency in places with lower precipitation (Figure 2), but there is only
429 marginal statistical support, possibly due to lower sample size ($F_{6,44}=4.64$, P-value = 0.06,
430 $R^2=0.31$, morph present in 13/49 populations). We obtained similar results with CHELSA
431 dataset for all the colour morphs, except the orange morph ($F_{0,27}=0.11$, P-value = 0.74,
432 $R^2=0.01$).

433

434

435

436 **Discussion**

437 Integrating macroecological patterns, within-population trends, experiments under controlled
438 conditions and field observations, we revealed the climatic drivers shaping intraspecific
439 colour variation of a widespread colour-polymorphic spider. We found little evidence for
440 thermal benefits of dark colouration, or that temperature drives variation in morph frequency
441 or presence. Instead, differences in the realised climatic niche of colour morphs, and their
442 frequency and presence, were strongly linked to precipitation. Specifically, there is a higher
443 frequency of dark individuals in wetter areas and a higher frequency of orange individuals in
444 drier areas. Our results uncover the crucial role of environmental factors in shaping the
445 presence and frequency of discrete phenotypes within and between populations.

446

447 Under controlled laboratory conditions the black morph had a slightly higher heating rate
448 compared to the black-and-white morph. However, we detected no difference in body
449 temperature of the two colour morphs under typical field conditions (average air temperature
450 30.32°C). Although our field data represent a single snapshot of environmental conditions,
451 our results are consistent with previous evidence that discounted, under field or laboratory
452 conditions, any thermal differences between bright and dark colour variants in other
453 arthropods (Umbers et al., 2013). The small differences in heating rate under controlled
454 conditions may only correspond to differences in heating rate or body temperature in
455 variable field conditions under very limited circumstances. For example, differences in
456 radiative heat gain are expected to be highest in cool, sunny conditions (Kingsolver &
457 Wiernasz, 1991; Zverev et al., 2018; Britton & Davidowitz, 2023). Colour may also have
458 minimal effect on body temperature for very small animals because their body temperature is
459 coupled to convection rather than radiation (Parry, 1951; Stevenson, 1985; Shine & Kearney,
460 2001; Umbers et al., 2013). In such cases, other factors such as convective and evaporative
461 heat loss can have a greater impact on body temperature than body colouration (Turner &
462 Lombard, 1990; Umbers et al., 2013). This is supported by our field measurements of body

463 temperature, where variation is mostly explained by ambient temperature and wind speed
464 (Supplementary Table 3).

465

466 The lack of effect of temperature in our field and laboratory approaches aligns well with our
467 realised climatic niche comparison, which shows that colour morphs differ in almost all
468 climatic variables except for minimum temperature. The black and black-and-white morphs
469 are the most similar in their climatic niche, while the orange and black morphs are the most
470 climatically distinct replacing each other throughout our sampling areas (9 of 38 populations
471 with black individuals also have orange individuals). Our colour frequency measurements
472 across multiple populations also indicated that precipitation is the most important climatic
473 factor affecting the distribution of colour morphs in the Christmas spider. Specifically, higher
474 precipitation favours a greater abundance of black individuals while limiting the presence of
475 the orange morph in the same areas. Higher abundance of black individuals in wetter areas
476 has been reported in other lineages (Friedman & Remeš, 2017; Delhey, 2019; Cerezer et al.,
477 2020; Lopez et al., 2021; Marcondes et al., 2021; Kang et al., 2021). This pattern may arise
478 as a consequence of the role of melanin in antimicrobial protection (Mackintosh, 2001;
479 Wilson et al., 2001; Nosanchuk & Casadevall, 2003), assuming higher pathogen density in
480 high-humidity environments. However, in spiders black and browns are usually produced by
481 ommochrome pigments (a combination of ommatins and ommins) (Seligy, 1972); in fact,
482 melanin was only recently reported in spiders for the first time (Hsiung et al., 2015).

483

484 An alternative explanation for the frequency variation of the black and orange morphs of the
485 Christmas spider across a precipitation gradient is that could arise, at least partly, as a
486 consequence of local adaptation associated with background matching (Stevens & Merilaita,
487 2008; Cheng et al., 2018). Background matching in web-building spiders is crucial to reduce
488 predation (Pekár, 2014; Robledo-Ospina & Rao, 2022) and increase prey capture (Nakata &

489 Shigemiya, 2015; Nakata, 2021) because they are restricted to their webs. A post-hoc
490 exploration of the association between soil brightness and colour variation suggests that the
491 black morph is more prevalent in forested locations with darker soils, while the orange morph
492 is more abundant in brighter soils characteristic of the most arid regions of Australia
493 (Supplementary figure 7). Available data on background colour are very coarse, which
494 hinders a formal analysis, and there are inescapable links between soil colour and climatic
495 variables, but background matching could be behind the association between precipitation
496 and coloration in this system. In fact, background matching to reduce predation or enhance
497 prey capture can promote colour variation both within and between populations across
498 multiple lineages and types of environments (Bond, 2007; Tate et al., 2016; Matthews et al.,
499 2018; Nosil et al., 2018; Fark et al., 2022; Villoutreix et al., 2023). However, this hypothesis
500 needs to be formally tested by collecting more detailed data on background colours in the
501 field and quantifying background matching. We also note that the sources of selection could
502 be different between the alternative morphs: pathogen protection could drive prevalence of
503 black individuals and camouflage could favour orange individuals in each environment.

504

505 Our data also showed that the black-and-white morph is the most successful colour variant.
506 This morph was the most abundant in all our sources of collection records, exhibited the
507 highest frequency in nearly all sampled locations (minimum 43.5%), and was the only colour
508 variant found in isolation. This geographical pattern, coupled with the spatial replacement of
509 black and orange colour variants correlated with precipitation, suggests that colour
510 polymorphism in this species may be maintained through the interplay of multiple selective
511 processes. Colour polymorphism can arise and be maintained when diverse selective
512 pressures act in opposing directions, favouring different colour variants (Endler, 1983;
513 Nokelainen et al., 2011; Poloni et al., 2024). This phenomenon has been documented in
514 various arthropods, where positive frequency-dependent selection favouring one morph is
515 counterbalanced by negative frequency-dependent selection favouring another phenotype

516 (Takahashi et al., 2011; Chouteau et al., 2017; Poloni et al., 2024). Another possibility is that
517 the coexistence of multiple colour morphs is a consequence of highly heterogeneous
518 environments that impose different forms of spatially and temporally varying selection at a
519 small geographical scale (Nosil et al., 2024). To test these hypotheses, further research is
520 needed to identify the functions of body coloration in *Austracantha minax*, given that we
521 currently do not know whether colour in this species acts as a warning signal or improves
522 prey capture.

523

524 Taken together, our work reveals a strong link between precipitation and intraspecific colour
525 variation in the Christmas spider and rules out any important effect of ambient temperature
526 driving colour variation in this species. This level of understanding was possible only through
527 the combination of broad scale, field, and laboratory approaches, providing important
528 evidence for the role of the environment in driving the evolution of colour morphs in spiders.
529 The patterns described here should be explored in depth to elucidate the mechanisms
530 maintaining both within and between colour variation in this species. Given the lack of
531 studies on most invertebrate taxa, it is also important to test the prevalence of the links found
532 here in other polymorphic species, especially in those species with dark or melanic morphs.

533

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542 **Conflict of Interest**

543 We declare that there are no competing interests in the publication of this work.

544 **Author contributions**

545 F.C.S.-R conceptualisation, funding acquisition, methodology, investigation, data curation,
546 formal analysis, visualization, writing—original draft; D.S.-F. conceptualisation, funding
547 acquisition, supervision, writing—review and editing; I.M. conceptualisation, funding
548 acquisition, supervision, writing—review and editing.

549 **Data availability**

550 All the scripts and information about the data are available here:

551 https://github.com/fcsalgado/christmas_spider_climate

552 **References**

- 553 Azócar, D. L. M., Bonino, M. F., Perotti, M. G., Schulte, J. A., Abdala, C. S., & Cruz, F. B.
554 (2016). Effect of body mass and melanism on heat balance in *Liolaemus* lizards of the
555 goetschi clade. *Journal of Experimental Biology*, 219(8), 1162–1171.
556 <https://doi.org/10.1242/jeb.129007>
- 557 Barbet-Massin, M., Jiguet, F., Albert, C. H., & Thuiller, W. (2012). Selecting pseudo-
558 absences for species distribution models: how, where and how many? *Methods in*
559 *Ecology and Evolution*, 3(2), 327–338. [https://doi.org/https://doi.org/10.1111/j.2041-](https://doi.org/https://doi.org/10.1111/j.2041-210X.2011.00172.x)
560 [210X.2011.00172.x](https://doi.org/https://doi.org/10.1111/j.2041-210X.2011.00172.x)
- 561 Bates, O. K., & Bertelsmeier, C. (2021). Climatic niche shifts in introduced species. *Current*
562 *Biology*, 31(19), R1252–R1266. <https://doi.org/https://doi.org/10.1016/j.cub.2021.08.035>
- 563 Bishop, T. R., Robertson, M. P., Gibb, H., van Rensburg, B. J., Braschler, B., Chown, S. L.,
564 Foord, S. H., Munyai, T. C., Okey, I., Tshivhandekano, P. G., Werenkraut, V., & Parr, C.
565 L. (2016). Ant assemblages have darker and larger members in cold environments.
566 *Global Ecology and Biogeography*, 25(12), 1489–1499.
567 <https://doi.org/10.1111/geb.12516>
- 568 Bolnick, D. I., Amarasekare, P., Araújo, M. S., Bürger, R., Levine, J. M., Novak, M., Rudolf,
569 V. H. W., Schreiber, S. J., Urban, M. C., & Vasseur, D. A. (2011). Why intraspecific trait
570 variation matters in community ecology. *Trends in Ecology & Evolution*, 26(4), 183–192.
571 <https://doi.org/https://doi.org/10.1016/j.tree.2011.01.009>
- 572 Bond, A. B. (2007). The Evolution of Color Polymorphism: Crypticity, Searching Images, and
573 Apostatic Selection. *Annual Review of Ecology, Evolution, and Systematics*, 38(1),
574 489–514. <https://doi.org/10.1146/annurev.ecolsys.38.091206.095728>
- 575 Britton, S., & Davidowitz, G. (2023). The adaptive role of melanin plasticity in thermally
576 variable environments. *Journal of Evolutionary Biology*, 36(12), 1811–1821.

577 <https://doi.org/10.1111/jeb.14243>

578 Broennimann, O., Ursenbacher, S., Meyer, A., Golay, P., Monney, J. C., Schmocker, H.,
579 Guisan, A., & Dubey, S. (2014). Influence of climate on the presence of colour
580 polymorphism in two montane reptile species. *Biology Letters*, *10*(11).
581 <https://doi.org/10.1098/rsbl.2014.0638>

582 Castella, B., Golay, J., Monney, J. C., Golay, P., Mebert, K., & Dubey, S. (2013). Melanism,
583 body condition and elevational distribution in the asp viper. *Journal of Zoology*, *290*(4),
584 273–280. <https://doi.org/10.1111/jzo.12037>

585 Cerezer, F. O., Ribeiro, J. R. I., Graipel, M., & Cáceres, N. C. (2020). The dark side of
586 coloration: Ecogeographical evidence supports Gloger’s rule in American marsupials.
587 *Evolution*, *74*(9), 2046–2058. <https://doi.org/10.1111/evo.13989>

588 Cheng, W., Xing, S., Chen, Y., Lin, R., Bonebrake, T. C., & Nakamura, A. (2018). Dark
589 butterflies camouflaged from predation in dark tropical forest understories. *Ecological*
590 *Entomology*, *43*(3), 304–309. <https://doi.org/10.1111/een.12499>

591 Chouteau, M., Llaurens, V., Piron-Prunier, F., & Joron, M. (2017). Polymorphism at a
592 mimicry supergene maintained by opposing frequency-dependent selection pressures.
593 *Proceedings of the National Academy of Sciences*, *114*(31), 8325 LP – 8329.
594 <https://doi.org/10.1073/pnas.1702482114>

595 Clusella-Trullas, S., Terblanche, J. S., Blackburn, T. M., & Chown, S. L. (2008). Testing the
596 thermal melanism hypothesis: A macrophysiological approach. *Functional Ecology*,
597 *22*(2), 232–238. <https://doi.org/10.1111/j.1365-2435.2007.01377.x>

598 Clusella Trullas, S., van Wyk, J. H., & Spotila, J. R. (2007). Thermal melanism in ectotherms.
599 *Journal of Thermal Biology*, *32*(5), 235–245.
600 <https://doi.org/10.1016/j.jtherbio.2007.01.013>

601 Collart, F., Hedenäs, L., Broennimann, O., Guisan, A., & Vanderpoorten, A. (2021).

602 Intraspecific differentiation: Implications for niche and distribution modelling. *Journal of*
603 *Biogeography*, 48(2), 415–426. <https://doi.org/https://doi.org/10.1111/jbi.14009>

604 Dalrymple, R. L., Flores-Moreno, H., Kemp, D. J., White, T. E., Laffan, S. W., Hemmings, F.
605 A., Hitchcock, T. D., & Moles, A. T. (2018). Abiotic and biotic predictors of
606 macroecological patterns in bird and butterfly coloration. *Ecological Monographs*, 88(2),
607 204–224. <https://doi.org/https://doi.org/10.1002/ecm.1287>

608 Delhey, K. (2019). A review of Gloger’s rule, an ecogeographical rule of colour: definitions,
609 interpretations and evidence. *Biological Reviews*, 94(4), 1294–1316.
610 <https://doi.org/10.1111/brv.12503>

611 Delhey, K., Dale, J., Valcu, M., & Kempenaers, B. (2019). Reconciling ecogeographical
612 rules: rainfall and temperature predict global colour variation in the largest bird
613 radiation. *Ecology Letters*, 22(4), 726–736. <https://doi.org/10.1111/ele.13233>

614 Des Roches, S., Post, D. M., Turley, N. E., Bailey, J. K., Hendry, A. P., Kinnison, M. T.,
615 Schweitzer, J. A., & Palkovacs, E. P. (2018). The ecological importance of intraspecific
616 variation. *Nature Ecology & Evolution*, 2(1), 57–64. [https://doi.org/10.1038/s41559-017-](https://doi.org/10.1038/s41559-017-0402-5)
617 0402-5

618 Di Cola, V., Broennimann, O., Petitpierre, B., Breiner, F. T., D’Amen, M., Randin, C., Engler,
619 R., Pottier, J., Pio, D., Dubuis, A., Pellissier, L., Mateo, R. G., Hordijk, W., Salamin, N.,
620 & Guisan, A. (2017). ecospat: an R package to support spatial analyses and modeling
621 of species niches and distributions. *Ecography*, 40(6), 774–787.
622 <https://doi.org/10.1111/ecog.02671>

623 Dinerstein, E., Olson, D., Joshi, A., Vynne, C., Burgess, N. D., Wikramanayake, E., Hahn,
624 N., Palminteri, S., Hedao, P., Noss, R., Hansen, M., Locke, H., Ellis, E. C., Jones, B.,
625 Barber, C. V., Hayes, R., Kormos, C., Martin, V., Crist, E., ... Saleem, M. (2017). An
626 Ecoregion-Based Approach to Protecting Half the Terrestrial Realm. *BioScience*, 67(6),
627 534–545. <https://doi.org/10.1093/biosci/bix014>

628 Dreiss, A. N., Antoniazza, S., Burri, R., Fumagalli, L., Sonnay, C., Frey, C., Goudet, J., &
629 Roulin, A. (2012a). Local adaptation and matching habitat choice in female barn owls
630 with respect to melanic coloration. *Journal of Evolutionary Biology*, 25(1), 103–114.
631 <https://doi.org/10.1111/j.1420-9101.2011.02407.x>

632 Dreiss, A. N., Antoniazza, S., Burri, R., Fumagalli, L., Sonnay, C., Frey, C., Goudet, J., &
633 Roulin, A. (2012b). Local adaptation and matching habitat choice in female barn owls
634 with respect to melanic coloration. *Journal of Evolutionary Biology*, 25(1), 103–114.
635 <https://doi.org/10.1111/j.1420-9101.2011.02407.x>

636 Ducrest, A.-L., Keller, L., & Roulin, A. (2008). Pleiotropy in the melanocortin system,
637 coloration and behavioural syndromes. *Trends in Ecology & Evolution*, 23(9), 502–510.
638 <https://doi.org/https://doi.org/10.1016/j.tree.2008.06.001>

639 Elgar, M. A., & Bathgate, R. (1996). Female Receptivity and Male Mate-Guarding in the
640 Jewel Spider *Gasteracantha minax* Thorell (Araneidae). In *Journal of Insect Behavior*
641 (Vol. 9, Issue 5).

642 Endler, J. A. (1977). *Geographic Variation, Speciation and Clines*. Princeton University
643 Press. <https://doi.org/10.2307/j.ctvx5wbdg>

644 Endler, J. A. (1983). Natural and sexual selection on color patterns in poeciliid fishes.
645 *Environmental Biology of Fishes*, 9(2), 173–190. <https://doi.org/10.1007/BF00690861>

646 Fark, S. N., Gerber, S., Alonzo, S. H., Kindsvater, H. K., Meier, J. I., & Seehausen, O.
647 (2022). Multispecies colour polymorphisms associated with contrasting microhabitats in
648 two Mediterranean wrasse radiations. *Journal of Evolutionary Biology*, 35(4), 633–647.
649 <https://doi.org/10.1111/jeb.13999>

650 Farquhar, J. E., Pili, A., & Russell, W. (2022). Using crowdsourced photographic records to
651 explore geographical variation in colour polymorphism. *Journal of Biogeography*,
652 August, 1–13. <https://doi.org/10.1111/jbi.14500>

653 Ford, E. B. (1945). Polymorphism. *Biological Reviews*, 20(2), 73–88.
654 <https://doi.org/https://doi.org/10.1111/j.1469-185X.1945.tb00315.x>

655 Friedman, N. R., & Remeš, V. (2017). Ecogeographical gradients in plumage coloration
656 among Australasian songbird clades. *Global Ecology and Biogeography*, 26(3), 261–
657 274. <https://doi.org/https://doi.org/10.1111/geb.12522>

658 Goldenberg, J., Bisschop, K., D’Alba, L., & Shawkey, M. D. (2022). The link between body
659 size, colouration and thermoregulation and their integration into ecogeographical rules:
660 a critical appraisal in light of climate change. *Oikos*, 2022(6), e09152.
661 <https://doi.org/https://doi.org/10.1111/oik.09152>

662 González-Suárez, M., & Revilla, E. (2013). Variability in life-history and ecological traits is a
663 buffer against extinction in mammals. *Ecology Letters*, 16(2), 242–251.
664 <https://doi.org/https://doi.org/10.1111/ele.12035>

665 Goslee, S. C., & Urban, D. L. (2007). The ecodist package for dissimilarity-based analysis of
666 ecological data. *Journal of Statistical Software*, 22(7), 1–19.
667 <https://doi.org/10.18637/jss.v022.i07>

668 Gray, S. M., & McKinnon, J. S. (2007). Linking color polymorphism maintenance and
669 speciation. *Trends in Ecology and Evolution*, 22(2), 71–79.
670 <https://doi.org/10.1016/j.tree.2006.10.005>

671 Guisan, A., Petitpierre, B., Broennimann, O., Daehler, C., & Kueffer, C. (2014). Unifying
672 niche shift studies: Insights from biological invasions. *Trends in Ecology and Evolution*,
673 29(5), 260–269. <https://doi.org/10.1016/j.tree.2014.02.009>

674 Hantak, M. M., Guralnick, R. P., Cameron, A. C., Griffing, A. H., Harrington, S. M., Weinell,
675 J. L., & Paluh, D. J. (2022). Colour scales with climate in North American ratsnakes: A
676 test of the thermal melanism hypothesis using community science images. *Biology*
677 *Letters*, 18(12). <https://doi.org/10.1098/rsbl.2022.0403>

678 Hastings, B. T., Melnyk, A., Ghyabi, M., White, E., Barroso, F. M., Carretero, M. A., Lattanzi,
679 D., Claude, J., & Chiari, Y. (2023). Melanistic coloration does not influence
680 thermoregulation in the crepuscular gecko *Eublepharis macularius*. *Biology Open*,
681 12(10), bio060114. <https://doi.org/10.1242/bio.060114>

682 Heidrich, L., Friess, N., Fiedler, K., Brändle, M., Hausmann, A., Brandl, R., & Zeuss, D.
683 (2018). The dark side of Lepidoptera: Colour lightness of geometrid moths decreases
684 with increasing latitude. *Global Ecology and Biogeography*, 27(4), 407–416.
685 <https://doi.org/10.1111/geb.12703>

686 Hijmans, R. J. (2016). *geosphere: Spherical Trigonometry*.

687 Hijmans, R. J. (2023). *terra: Spatial Data Analysis*. <https://cran.r-project.org/package=terra>

688 Hogg, H. R. (1914). Spiders from the Montebello Islands. *Proceedings of the Zoological*
689 *Society of London*, 84(1), 69–92.

690 Hsiung, B.-K., Blackledge, T. A., & Shawkey, M. D. (2015). Spiders do have melanin after
691 all. *Journal of Experimental Biology*, 218(22), 3632–3635.
692 <https://doi.org/10.1242/jeb.128801>

693 Jamie, G. A., & Meier, J. I. (2020). The Persistence of Polymorphisms across Species
694 Radiations. *Trends in Ecology & Evolution*, 1–14.
695 <https://doi.org/10.1016/j.tree.2020.04.007>

696 Jones, D. A., Wang, W., & Fawcett, R. (2009). High-quality spatial climate data-sets for
697 Australia. *Australian Meteorological and Oceanographic Journal*, 58(4), 233.

698 Kang, C., Im, S., Lee, W. Y., Choi, Y., Stuart-Fox, D., & Huertas, B. (2021). Climate predicts
699 both visible and near-infrared reflectance in butterflies. *Ecology Letters*, 24(9), 1869–
700 1879. <https://doi.org/https://doi.org/10.1111/ele.13821>

701 Karger, D. N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza, R. W.,
702 Zimmermann, N. E., Linder, H. P., & Kessler, M. (2017). Climatologies at high

703 resolution for the earth's land surface areas. *Scientific Data*, 4(1), 170122.
704 <https://doi.org/10.1038/sdata.2017.122>

705 Kassen, R. (2002). The experimental evolution of specialists, generalists, and the
706 maintenance of diversity. *Journal of Evolutionary Biology*, 15(2), 173–190.
707 <https://doi.org/https://doi.org/10.1046/j.1420-9101.2002.00377.x>

708 Kingsolver, J. G., & Wiernasz, D. C. (1991). Seasonal Polyphenism in Wing-Melanin Pattern
709 and Thermoregulatory Adaptation in Pieris Butterflies. *The American Naturalist*, 137(6),
710 816–830. <https://doi.org/10.1086/285195>

711 Lancaster, L. T., Dudaniec, R. Y., Hansson, B., & Svensson, E. I. (2017). Do group
712 dynamics affect colour morph clines during a range shift? *Journal of Evolutionary*
713 *Biology*, 30(4), 728–737. <https://doi.org/10.1111/jeb.13037>

714 Legendre, P., & Fortin, M.-J. (2010). Comparison of the Mantel test and alternative
715 approaches for detecting complex multivariate relationships in the spatial analysis of
716 genetic data. *Molecular Ecology Resources*, 10(5), 831–844.
717 <https://doi.org/10.1111/j.1755-0998.2010.02866.x>

718 Lloyd, N. J., & Elgar, M. A. (1997). Costs and benefits of facultative aggregating behaviour in
719 the orb-spinning spider *Gasteracantha minax* Thorell (Araneae: Araneidae). In
720 *Australian Journal of Ecology* (Vol. 22).

721 Lopez, V. M., Azevedo Tosta, T. A., da Silva, G. G., Bartholomay, P. R., Williams, K. A., &
722 Ferreira, G.-F. (2021). Color lightness of velvet ants (Hymenoptera: Mutillidae) follows
723 an environmental gradient. *Journal of Thermal Biology*, 100(February), 103030.
724 <https://doi.org/10.1016/j.jtherbio.2021.103030>

725 Lüdecke, D., Ben-Shachar, M. S., Patil, I., Waggoner, P., & Makowski, D. (2021).
726 performance: An R Package for Assessment, Comparison and Testing of Statistical
727 Models. *Journal of Open Source Software*, 6(60), 3139.

728 <https://doi.org/10.21105/joss.03139>

729 Mackintosh, J. A. (2001). The Antimicrobial Properties of Melanocytes, Melanosomes and
730 Melanin and the Evolution of Black Skin. *Journal of Theoretical Biology*, 211(2), 101–
731 113. <https://doi.org/10.1006/jtbi.2001.2331>

732 Marcondes, R. S., Nations, J. A., Seeholzer, G. F., & Brumfield, R. T. (2021). Rethinking
733 Gloger's Rule: Climate, Light Environments, and Color in a Large Family of Tropical
734 Birds (Furnariidae). *The American Naturalist*, 197(5), 592–606.
735 <https://doi.org/10.1086/713386>

736 Mascord, R. E. (1996). THE MATING BEHAVIOUR OF GASTERACANTHA MINAX
737 THORELL, 1859 (ARANEIDA: ARGIOPIDAE). *The Journal of the Entomological*
738 *Society of Australia*, 3.

739 Matthews, G., Goulet, C. T., Delhey, K., Atkins, Z. S., While, G. M., Gardner, M. G., &
740 Chapple, D. G. (2018). Avian predation intensity as a driver of clinal variation in colour
741 morph frequency. *Journal of Animal Ecology*, 87(6), 1667–1684.
742 <https://doi.org/10.1111/1365-2656.12894>

743 Meredith, P., & Sarna, T. (2006). The physical and chemical properties of eumelanin.
744 *Pigment Cell Research*, 19(6), 572–594. [https://doi.org/10.1111/j.1600-](https://doi.org/10.1111/j.1600-0749.2006.00345.x)
745 [0749.2006.00345.x](https://doi.org/10.1111/j.1600-0749.2006.00345.x)

746 Moran, E. V, Hartig, F., & Bell, D. M. (2016). Intraspecific trait variation across scales:
747 implications for understanding global change responses. *Global Change Biology*, 22(1),
748 137–150. <https://doi.org/10.1111/gcb.13000>

749 Mullen, L. M., & Hoekstra, H. E. (2008). NATURAL SELECTION ALONG AN
750 ENVIRONMENTAL GRADIENT: A CLASSIC CLINE IN MOUSE PIGMENTATION.
751 *Evolution*, 62(7), 1555–1570. <https://doi.org/10.1111/j.1558-5646.2008.00425.x>

752 Nakata, K. (2021). Relationship between body colour and microhabitat breadth in an orb-

753 web spider. *Biological Journal of the Linnean Society*, 134(3), 679–684.
754 <https://doi.org/10.1093/biolinnean/blab118>

755 Nakata, K., & Shigemiyama, Y. (2015). Body-colour variation in an orb-web spider and its effect
756 on predation success. *Biological Journal of the Linnean Society*, 116(4), 954–963.
757 <https://doi.org/10.1111/bij.12640>

758 Nokelainen, O., Hegna, R. H., Reudler, J. H., Lindstedt, C., & Mappes, J. (2011). Trade-off
759 between warning signal efficacy and mating success in the wood tiger moth.
760 *Proceedings of the Royal Society B: Biological Sciences*, 279(1727), 257–265.
761 <https://doi.org/10.1098/rspb.2011.0880>

762 Nosanchuk, J. D., & Casadevall, A. (2003). The contribution of melanin to microbial
763 pathogenesis. *Cellular Microbiology*, 5(4), 203–223.
764 <https://doi.org/https://doi.org/10.1046/j.1462-5814.2003.00268.x>

765 Nosil, P., De Carvalho, C. F., Villoutreix, R., Zamorano, L. S., Sinclair-Waters, M., Planidin,
766 N. P., Parchman, T. L., Feder, J., & Gompert, Z. (2024). Evolution repeats itself in
767 replicate long-term studies in the wild. *Science Advances*, 10(21), 34–36.
768 <https://doi.org/10.1126/sciadv.adl3149>

769 Nosil, P., Villoutreix, R., De Carvalho, C. F., Farkas, T. E., Soria-Carrasco, V., Feder, J. L.,
770 Crespi, B. J., & Gompert, Z. (2018). Natural selection and the predictability of evolution
771 in timema stick insects. *Science*, 359(6377), 765–770.
772 <https://doi.org/10.1126/science.aap9125>

773 Ospina-Rozo, L., Subbiah, J., Seago, A., & Stuart-Fox, D. (2022). Pretty Cool Beetles: Can
774 Manipulation of Visible and Near-Infrared Sunlight Prevent Overheating? *Integrative*
775 *Organismal Biology*, 4(1), obac036. <https://doi.org/10.1093/iob/obac036>

776 Palacio, R. D., & Clark, J. S. (2023). Incorporating intraspecific variation into species
777 responses reveals both their resilience and vulnerability to future climate change.

778 *Ecography*, 2023(11), e06769. <https://doi.org/https://doi.org/10.1111/ecog.06769>

779 Parry, D. A. (1951). Factors Determining the Temperature of Terrestrial Arthropods in
780 Sunlight. *Journal of Experimental Biology*, 28(4), 445–462.
781 <https://doi.org/10.1242/jeb.28.4.445>

782 Passarotto, A., Rodríguez-Caballero, E., Cruz-Miralles, Á., & Avilés, J. M. (2022).
783 Ecogeographical patterns in owl plumage colouration: Climate and vegetation cover
784 predict global colour variation. *Global Ecology and Biogeography*, 31(3), 515–530.
785 <https://doi.org/https://doi.org/10.1111/geb.13444>

786 Pebesma, E. (2018). Simple Features for R: Standardized Support for Spatial Vector Data.
787 *The R Journal*, 10(1), 439–446. <https://doi.org/10.32614/RJ-2018-009>

788 Pekár, S. (2014). Comparative analysis of passive defences in spiders (Araneae). *Journal of*
789 *Animal Ecology*, 83(4), 779–790. <https://doi.org/10.1111/1365-2656.12177>

790 Pepi, A., Hemstrom, W., Dang, A., Beck, C., & Beyerle, F. (2022). Comparing the roles of
791 climate, predation and phylogeography in driving wing colour variation in Ranchman's
792 tiger moth (*Arctia virginalis*). *Biological Journal of the Linnean Society*, 204–215.
793 <https://doi.org/10.1093/biolinnean/blac138>

794 Pérez i de Lanuza, G., Sillero, N., & Carretero, M. Á. (2018). Climate suggests environment-
795 dependent selection on lizard colour morphs. *Journal of Biogeography*, 45(12), 2791–
796 2802. <https://doi.org/10.1111/jbi.13455>

797 Peterson, A. T., Soberon, J., Pearson, R. G., Anderson, R. P., Martinez-Meyer, E.,
798 Nakamura, M., & Araujo, M. B. (2011). Ecological niches and geographic distributions.
799 In *Princeton University Press*. <https://doi.org/10.5860/CHOICE.49-6266>

800 Pinkert, S., Brandl, R., & Zeuss, D. (2017). Colour lightness of dragonfly assemblages
801 across North America and Europe. *Ecography*, 40(9), 1110–1117.
802 <https://doi.org/10.1111/ecog.02578>

803 Pittiglio, C., Khomenko, S., & Beltran-Alcrudo, D. (2018). Wild boar mapping using
804 population-density statistics: From polygons to high resolution raster maps. *PLOS ONE*,
805 13(5), e0193295. <https://doi.org/10.1371/journal.pone.0193295>

806 Poloni, R., Dhennin, M., Mappes, J., Joron, M., & Nokelainen, O. (2024). Exploring
807 polymorphism in a palatable prey: predation risk and frequency dependence in relation
808 to distinct levels of conspicuousness. *Evolution Letters*, 8(3), 406–415.
809 <https://doi.org/10.1093/evlett/grad071>

810 R Core Team. (2021). *R: A Language and Environment for Statistical Computing*.
811 <https://www.r-project.org/>

812 Robledo-Ospina, L. E., & Rao, D. (2022). Dangerous visions: a review of visual antipredator
813 strategies in spiders. *Evolutionary Ecology*, 36(2), 163–180.
814 <https://doi.org/10.1007/s10682-022-10156-x>

815 Roulin, A. (2016). Condition-dependence, pleiotropy and the handicap principle of sexual
816 selection in melanin-based colouration. *Biological Reviews*, 91(2), 328–348.
817 <https://doi.org/https://doi.org/10.1111/brv.12171>

818 Roulin, A., & Bize, P. (2007). Sexual selection in genetic colour-polymorphic species: a
819 review of experimental studies and perspectives. *Journal of Ethology*, 25(2), 99–105.
820 <https://doi.org/10.1007/s10164-006-0006-z>

821 ROULIN, A., BURRI, R., & ANTONIAZZA, S. (2011). Owl melanin-based plumage redness
822 is more frequent near than away from the equator: implications on the effect of climate
823 change on biodiversity. *Biological Journal of the Linnean Society*, 102(3), 573–582.
824 <https://doi.org/10.1111/j.1095-8312.2010.01614.x>

825 Salgado-Roa, F. C., Stuart-Fox, D., White, T. E., & Medina, I. (2024). Colour polymorphism
826 is prevalent on islands but shows no association with range size in web-building
827 spiders. *Journal of Evolutionary Biology*, voae118. <https://doi.org/10.1093/jeb/voae118>

828 Schoener, T. W. (1968). The Anolis Lizards of Bimini: Resource Partitioning in a Complex
829 Fauna. *Ecology*, 49(4), 704–726. <https://doi.org/10.2307/1935534>

830 Seligy, V. L. (1972). Ommochrome pigments of spiders. *Comparative Biochemistry and*
831 *Physiology Part A: Physiology*, 42(3), 699–709.
832 [https://doi.org/https://doi.org/10.1016/0300-9629\(72\)90448-3](https://doi.org/https://doi.org/10.1016/0300-9629(72)90448-3)

833 Shine, R., & Kearney, M. R. (2001). Field studies of reptile thermoregulation: how well do
834 physical models predict operative temperatures? *Functional Ecology*, 15(2), 282–288.
835 <https://doi.org/https://doi.org/10.1046/j.1365-2435.2001.00510.x>

836 Smith, K. R., Cadena, V., Endler, J. A., Kearney, M. R., Porter, W. P., & Stuart-Fox, D.
837 (2016). Color Change for Thermoregulation versus Camouflage in Free-Ranging
838 Lizards. *The American Naturalist*, 188(6), 668–678. <https://doi.org/10.1086/688765>

839 Stevens, M., & Merilaita, S. (2008). Animal camouflage: current issues and new
840 perspectives. *Philosophical Transactions of the Royal Society B: Biological Sciences*,
841 364(1516), 423–427. <https://doi.org/10.1098/rstb.2008.0217>

842 Stevenson, R. D. (1985). The Relative Importance of Behavioral and Physiological
843 Adjustments Controlling Body Temperature in Terrestrial Ectotherms. *The American*
844 *Naturalist*, 126(3), 362–386. <https://doi.org/10.1086/284423>

845 Takahashi, Y., Morita, S., Yoshimura, J., & Watanabe, M. (2011). A geographic cline
846 induced by negative frequency-dependent selection. *BMC Evolutionary Biology*, 11(1),
847 256. <https://doi.org/10.1186/1471-2148-11-256>

848 Tate, G. J., Bishop, J. M., & Amar, A. (2016). Differential foraging success across a light
849 level spectrum explains the maintenance and spatial structure of colour morphs in a
850 polymorphic bird. *Ecology Letters*, 19(6), 679–686. <https://doi.org/10.1111/ele.12606>

851 Turner, J. S., & Lombard, A. T. (1990). Body color and body temperature in white and black
852 Namib desert beetles. *Journal of Arid Environments*, 19(3), 303–315.

853 [https://doi.org/https://doi.org/10.1016/S0140-1963\(18\)30795-X](https://doi.org/https://doi.org/10.1016/S0140-1963(18)30795-X)

854 Umbers, K. D. L., Herberstein, M. E., & Madin, J. S. (2013). Colour in insect
855 thermoregulation: Empirical and theoretical tests in the colour-changing grasshopper,
856 *Kosciuscola tristis*. *Journal of Insect Physiology*, *59*(1), 81–90.
857 <https://doi.org/10.1016/j.jinsphys.2012.10.016>

858 Villoutreix, R., de Carvalho, C. F., Feder, J. L., Gompert, Z., & Nosil, P. (2023). Disruptive
859 selection and the evolution of discrete color morphs in *Timema* stick insects. *Science*
860 *Advances*, *9*(13), eabm8157. <https://doi.org/10.1126/sciadv.abm8157>

861 Waldock, J. M. (1991). The colour-forms of the Christmas spider *Gasteracantha minax* in
862 south-western Australia. *The Western Australian Naturalist*, *18*, 207–215.

863 Wang, I. J. (2013). Examining the full effects of landscape heterogeneity on spatial genetic
864 variation: A multiple matrix regression approach for quantifying geographic and
865 ecological isolation. *Evolution*, *67*(12), 3403–3411. <https://doi.org/10.1111/evo.12134>

866 Wang, L.-Y., Franklin, A. M., Black, J. R., & Stuart-Fox, D. (2021). Heating rates are more
867 strongly influenced by near-infrared than visible reflectance in beetles. *Journal of*
868 *Experimental Biology*, *224*(19), jeb242898. <https://doi.org/10.1242/jeb.242898>

869 Wang, Y., Pineda-Munoz, S., & McGuire, J. L. (2023). Plants maintain climate fidelity in the
870 face of dynamic climate change. *Proceedings of the National Academy of Sciences*,
871 *120*(7), e2201946119. <https://doi.org/10.1073/pnas.2201946119>

872 Warren, D. L., Glor, R. E., & Turelli, M. (2008). Environmental niche equivalency versus
873 conservatism: quantitative approaches to niche evolution. *Evolution*, *62*(11), 2868–
874 2883. <https://doi.org/10.1111/j.1558-5646.2008.00482.x>

875 Watt, W. B. (1968). Adaptive Significance of Pigment Polymorphisms in *Colias* Butterflies. I.
876 Variation of Melanin Pigment in Relation to Thermoregulation. *Evolution*, *22*(3), 437–
877 458. <https://doi.org/10.2307/2406873>

878 Wellenreuther, M., Svensson, E. I., & Hansson, B. (2014). Sexual selection and genetic
879 colour polymorphisms in animals. *Molecular Ecology*, 23(22), 5398–5414.
880 <https://doi.org/https://doi.org/10.1111/mec.12935>

881 Whitford, A. M., Shipley, B. R., & McGuire, J. L. (2024). The influence of the number and
882 distribution of background points in presence-background species distribution models.
883 *Ecological Modelling*, 488, 110604.
884 <https://doi.org/https://doi.org/10.1016/j.ecolmodel.2023.110604>

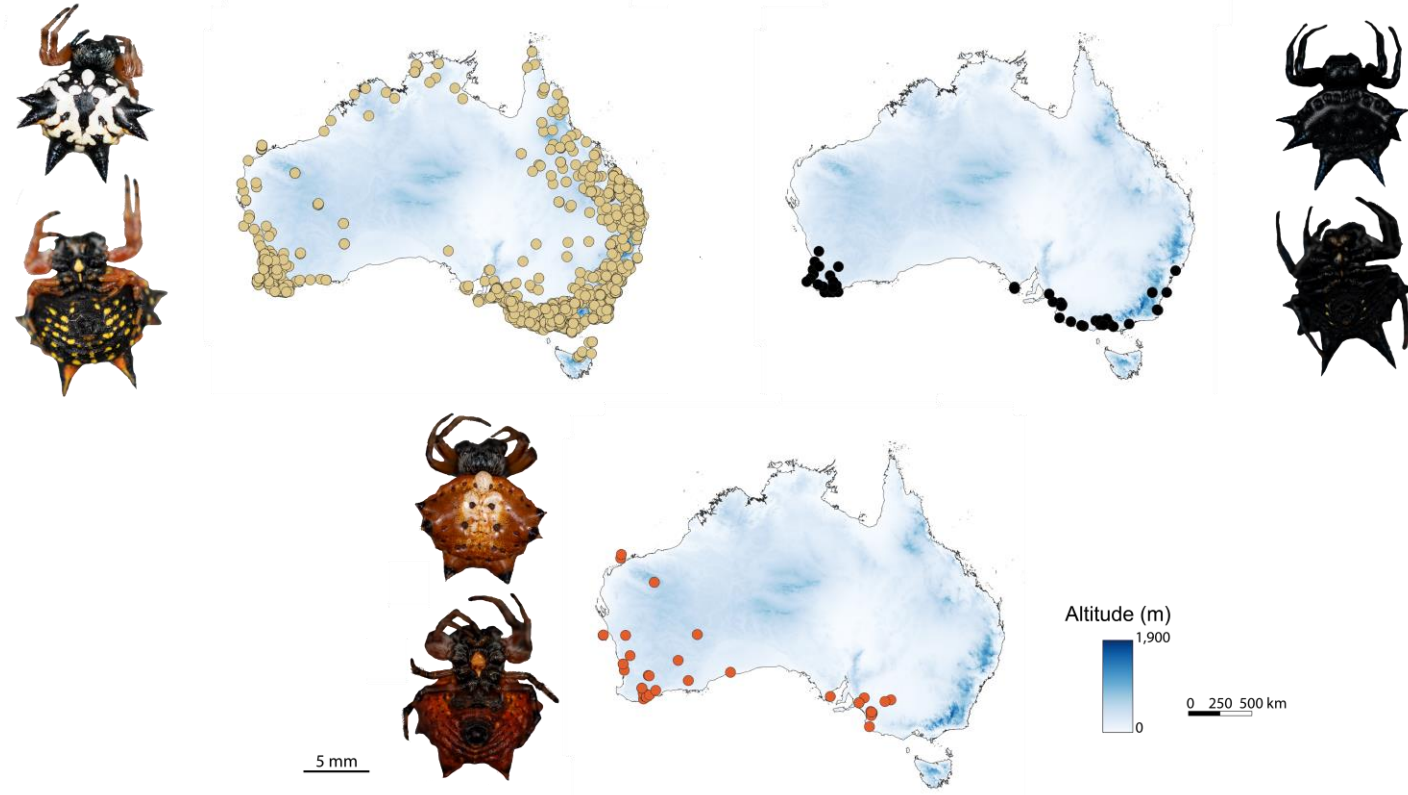
885 Wilson, K., Cotter, S. C., Reeson, A. F., & Pell, J. K. (2001). Melanism and disease
886 resistance in insects. *Ecology Letters*, 4(6), 637–649.
887 <https://doi.org/https://doi.org/10.1046/j.1461-0248.2001.00279.x>

888 Wittkopp, P. J., Smith-Winberry, G., Arnold, L. L., Thompson, E. M., Cooley, A. M., Yuan, D.
889 C., Song, Q., & McAllister, B. F. (2011). Local adaptation for body color in *Drosophila*
890 *americana*. *Heredity*, 106(4), 592–602. <https://doi.org/10.1038/hdy.2010.90>

891 Zverev, V., Kozlov, M. V, Forsman, A., & Zvereva, E. L. (2018). Ambient temperatures
892 differently influence colour morphs of the leaf beetle *Chrysomela lapponica*: Roles of
893 thermal melanism and developmental plasticity. *Journal of Thermal Biology*, 74, 100–
894 109. <https://doi.org/https://doi.org/10.1016/j.jtherbio.2018.03.019>

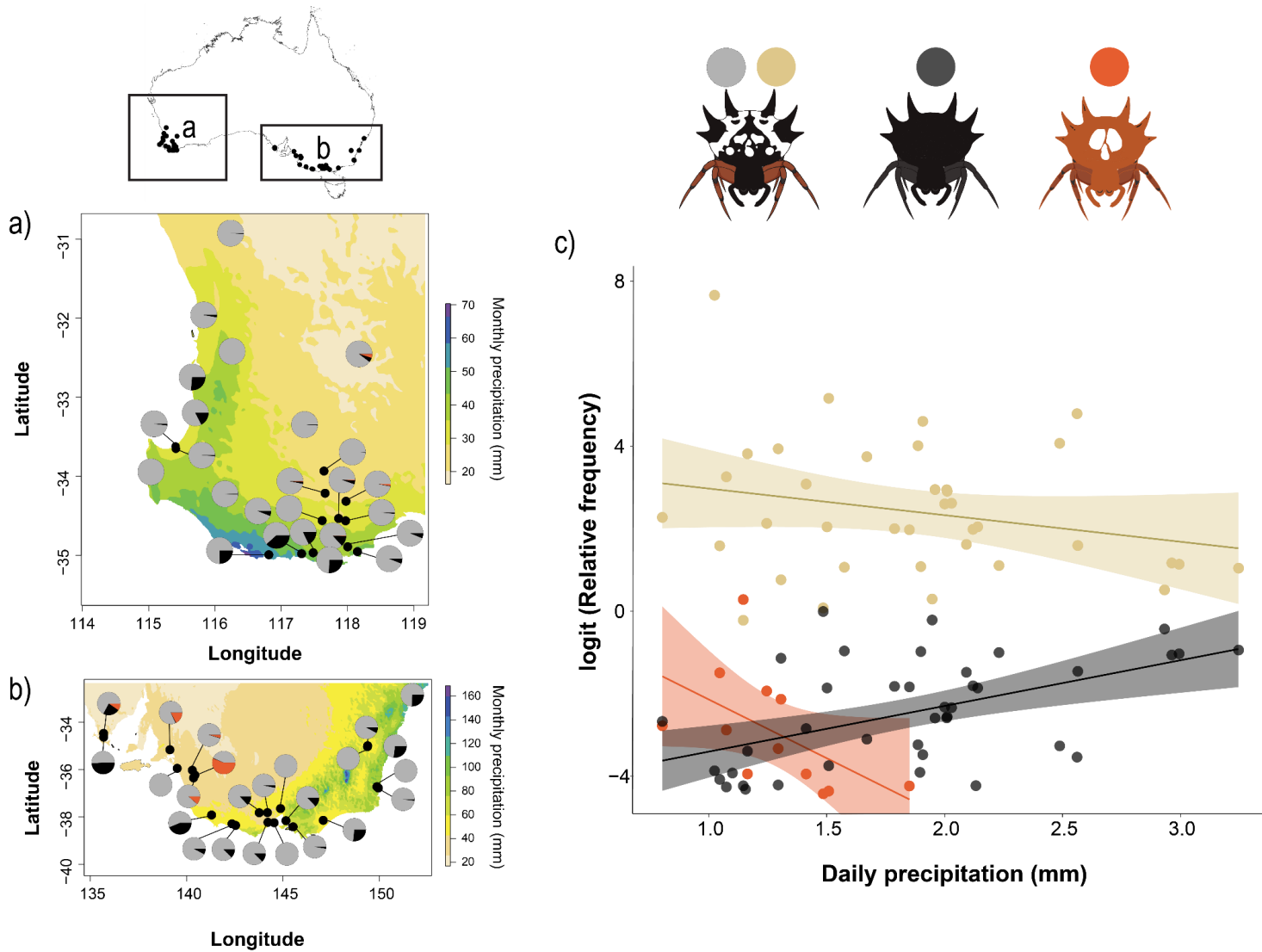
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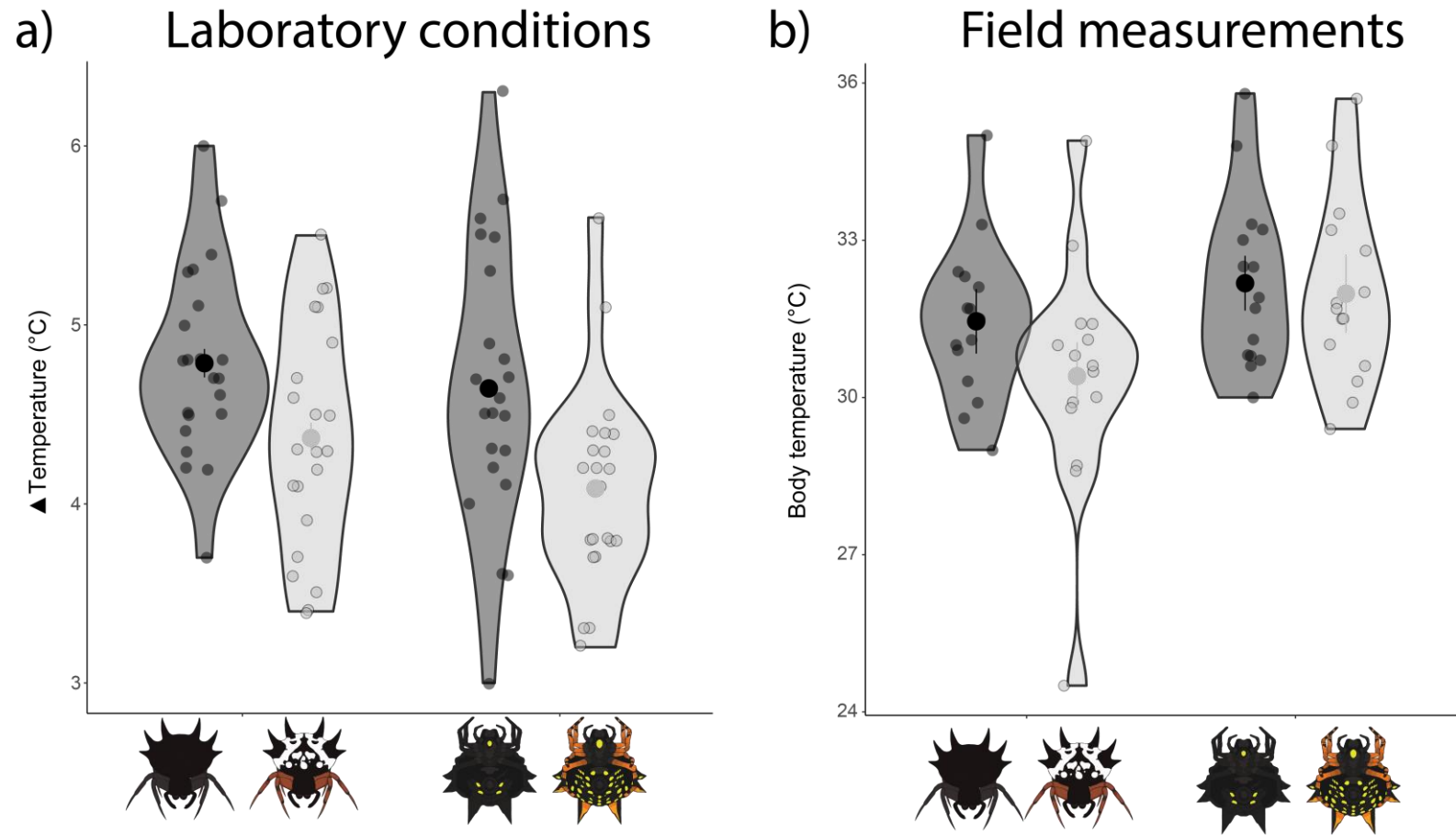
898 **Figure 1.** *Austracantha minax* colour variation and morph distribution. Each map represents the geographical records collated for each female
 899 colour variant plotted on an altitude background. The top-left panel corresponds to the black-and-white morph (1097 records), the top-right to
 900 the black morph (123 records), and the bottom panel to the orange morph (34 records).



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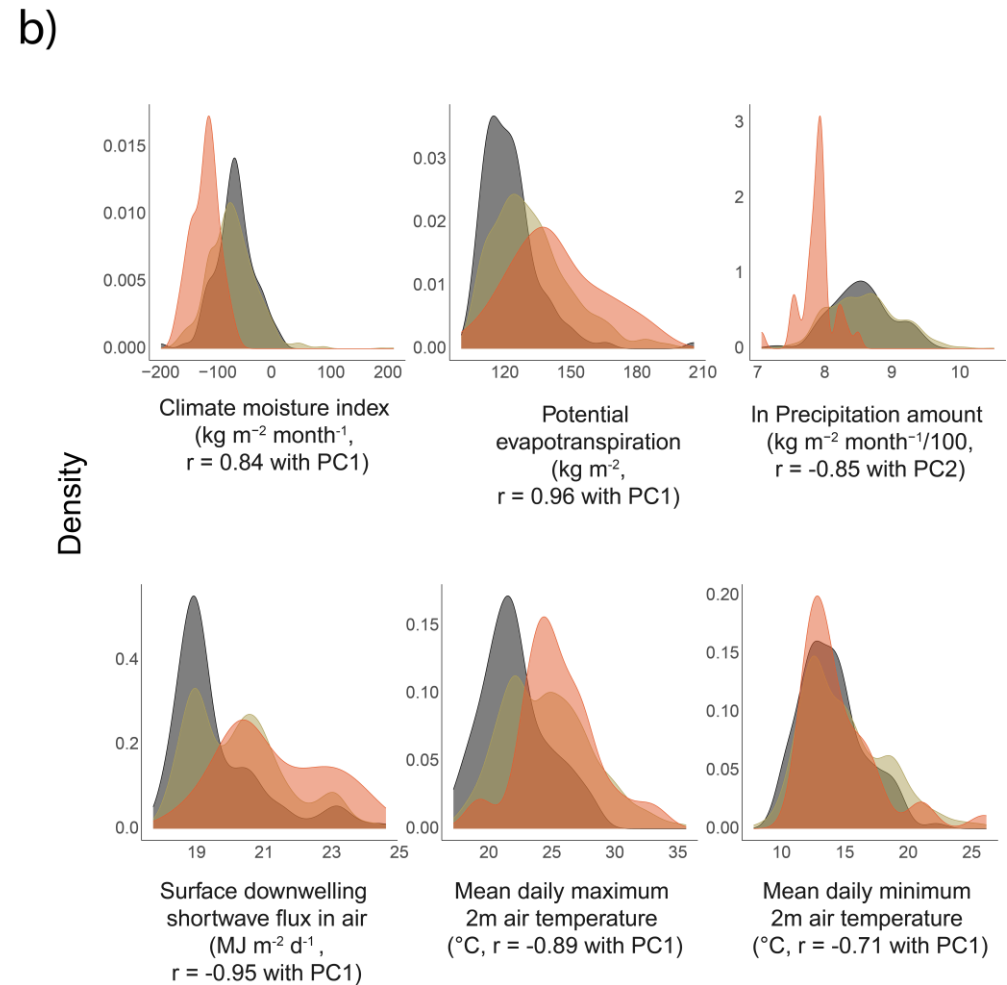
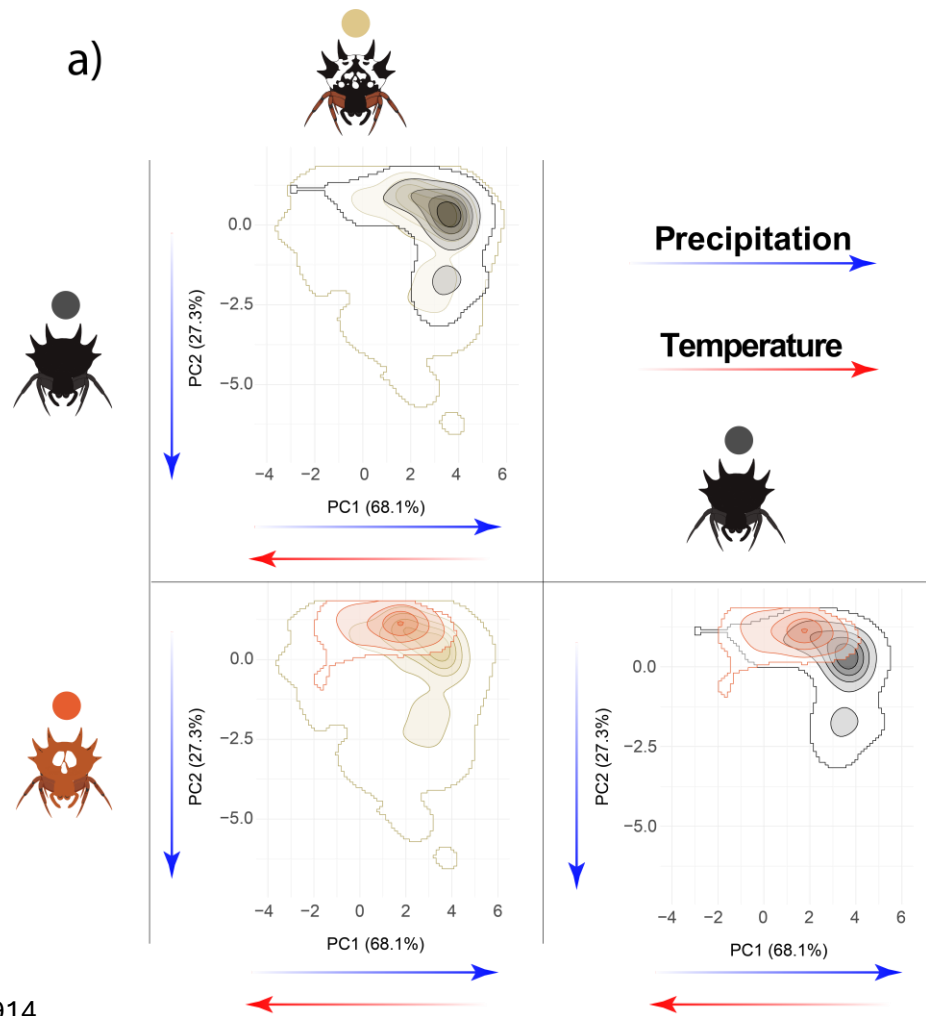
902 **Figure 2.** Variation in relative frequency for each colour morph and its association with
 903 precipitation. The maps show the relative frequency of the black-and-white (grey colour),
 904 black, and orange morphs across 49 locations in a) south-western and b) south-eastern
 905 Australia. c) Linear model association between the relative frequency of each colour morph
 906 and daily precipitation. Line represents estimate from the model and shaded area represents
 907 the confidence interval of the regression.

908



910

911 **Figure 3.** Differences in temperature between the black-and-white and black colour morphs on their ventral and dorsal sides. a) Heating rates
 912 experiment under controlled laboratory conditions. b) Field temperature measurements. The dots represent measured values per individual per
 913 morph, while the big circles and bars represent the mean \pm standard error of the predicted values from the linear models.



914

915 **Figure 4.** Realized climatic niche of the colour morphs of the Christmas spider. A) Multivariate pairwise comparison of the climatic environment of the
 916 black-and-white (yellow), black (black), and orange (orange) morphs. Darker colours represent a higher density of records. Blue arrows represent the

917 direction of increase of precipitation, climatic moisture index and potential evapotranspiration values in both PC1 and PC2. Red arrows symbolize the
918 increase of solar radiation and temperature values on PC1. B) Density distribution of the records of each climatic variable and their correlation with
919 the principal components in plot A. All density plots per variable are available in supplementary material (Supplementary Figures 5 & 6).

920