

# The Architecture of Theory and Data in Microbiome Design: towards an S-matrix for microbiomes

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Designing microbiomes for applications in health, bioengineering, and sustainability is intrinsically linked to a fundamental theoretical understanding of the rules governing microbial community assembly. Microbial ecologists have used a range of mathematical models to understand, predict, and control microbiomes, ranging from mechanistic models, putting microbial populations and their interactions as the focus, to purely statistical approaches, searching for patterns in empirical and experimental data. We review the success and limitations of these modeling approaches when designing novel microbiomes, especially when guided by (inevitably) incomplete experimental data. Although successful at predicting generic patterns of community assembly, mechanistic and phenomenological models tend to fall short of the precision needed to design and implement specific functionality in a microbiome. We argue that to effectively design microbiomes with optimal functions in diverse environments, ecologists should combine data-driven techniques with mechanistic models—a middle, third way for using theory to inform design.

## Introduction

Microbiomes are complex systems, comprising multiple taxa interacting with their biotic and abiotic environment, and can be found from inhospitable cold, heat, and acidity (1), to the familiar host-associated environments of humans, plants, and animals (2). Across these environments, microbiomes modulate a broad range of functions, from biogeochemical cycles to host health (3–6). This breadth of possible microbiome function has highlighted the potential for us to engineer microbiomes to produce novel, beneficial outcomes, or even just to keep pace with the demands of a changing global environment (7). Humans as engineers of microbiomes have a long history: with records going as far back as 10,000 years ago, human populations have been developing fermented food and drink, including wine, beer, and dairy products (8), extending more recently to produce biofuel, enhance treatment of infections and inflammatory conditions, increase crop yield, and waste-water treatment (9–12). Microbial communities can exhibit emergent functions that no one microbe can perform, and there is also evidence for greater stability and resilience of diverse consortia (13, 14). While we can quantify communities in unprecedented detail (15–18), the question is whether we can go beyond merely documenting communities, and harness these data to inform design and engineering of functionally-relevant microbiomes.

A brute-force search for novel or optimized functions is unlikely to be efficient—but theoretical frameworks have often provided guiding principles in ecology and beyond, suggesting that the right theory might allow us to characterize the outcome of microbiome assembly, and thus predict function. This interplay between prediction and theoretical modeling mirrors a related goal in a seemingly distant area of science: calculating the scattering- or S-matrix in theoretical physics. The S-matrix provides a framework for mapping initial particles and momenta to predict late-time outcomes following interactions among those incoming particles. Although the component parts differ, this resembles the goal of mapping taxonomic and nutrient initial conditions to late-time community function, at least in cases where there is a well-defined steady-state, and where nutrient conditions themselves are not changing dramatically over time. Moreover, physicists calculating the S-matrix have wrestled with questions similar to those tackled by microbiome scientists. Specifically, a key theme has been the balance between deriving the S-matrix from specific, mechanistic assumptions in a given field theory, versus identifying general principles underlying its structure that abstract away the details of these processes (19).

We will review several microbiome modeling approaches below, that in part reflect this analogy with calculating the S-matrix. We propose that the difficulty in matching predictive power with available taxonomic, genomic, and metabolomic data will likely require a re-thinking of modeling in ecology and the adoption of data-driven design and control alongside traditional approaches. This may reveal principles of microbiome design that are independent of any specific theoretical model.

## Mechanistic and Phenomenological Models

Models assuming pairwise interactions among microbial taxa (20–22) are characterized by a community matrix,  $A = \{a_{ij}\}$ , where the strength and the sign of the coefficient  $a_{ij}$  denotes the effect that species  $j$  has on the growth of species  $i$ . The generalized Lotka-Volterra (gLV) model is the standard way to model pairwise interactions, and is able to accommodate many kinds of observed behaviors: amensalism, commensalism, exploitation, competition, and mutualism; this flexibility as much as anything else has positioned gLV as a foundational model in studying microbial communities. This approach can be extended to include higher-order interactions (23–26) or to include the role of space (27). In particular, inspired by Robert May’s use of random interaction matrices to probe the diversity-stability relationships in ecology, and using tools from physics of disordered systems, random gLV models have been explored extensively to inform equilibria, community coexistence, stability, and feasibility outcomes (21, 22, 28).

The gLV model has been used to make predictions for statistical patterns in microbiomes, as well as coarse-grained properties like the dependence of community stability on community diversity. But the number of parameters required to model a specific community grows quadratically with the size of the pool, and so a major challenge is posed by the number of experiments needed to infer these parameters accurately. When considering in-vivo communities like the human gut microbiome, the

required data suffers from non-uniformity of sampling times, and experimental noise, although studies have incorporated various algorithms to address these issues (29–31). Many studies have applied gLV models to infer parameters for in-vitro and in-vivo communities, and for explanation and prediction of community function (17, 22, 23, 31–34). Though when community function is something that depends in a non-trivial way on species composition (for example, the production of a metabolite), additional hypotheses must then be generated and tested for the relationship between composition and function, and this mapping might well be idiosyncratic to a given community and context (17, 35–37). In summary, the strength of the gLV model lies in the potential for explaining generic, statistical assembly patterns of microbiomes, and in making specific predictions, but only when the diversity of a community is relatively small and the relationship between late-time taxonomic composition and function can be independently estimated.

One of the main drawbacks of the gLV modelling framework is that it does not account for chemical mediators and resources and metabolites—some of the key mechanisms underlying species interactions. As a result, gLV parameters inferred from a system with a given resource and nutrient condition are unlikely to extend to arbitrarily-different resource environments. Second, the addition of new taxa may change the effective strength of existing pairwise interactions through modification of resources available to other species (38, 39). This makes pairwise models like the gLV unsuitable for design of consortia whenever there are strong effects of this type. This issue has been addressed using adaptations of the classic MacArthur’s consumer-resource model (40) that include explicit dynamics for both abiotic resources and species dynamics (41, 42). More generally, chemically-mediated interactions via toxins and anti-microbial production (43) can also be included. Consumer-resource models also hold a lot of promise in incorporating the various aspects of microbial metabolism. Constraints on and flexibility of microbial metabolism leads to diverse phenotypes and behaviors of consumer-resource dynamics (44–51)

These additional mechanisms also generally increase the number of parameters, which makes accurate parameter inference challenging. This issue is compounded by the fact that it is generally harder to measure and interpret resource concentrations. These twin difficulties in fitting consumer-resource models mean that they have so far been used less often than gLV models for parameter inference from experiments. More often these models have also been used to explain statistical patterns in microbial communities, including stability-diversity relationships (41, 42, 52, 53), and qualitative patterns in large, varied data such as the Human Microbiome and the Earth Microbiome Projects (53, 54). But as estimating metabolite concentrations becomes more tractable, we expect a continuing move towards fitting consumer-resource model parameters using combined resource and taxonomic data. Gowda et al (55) used a regression model to map gene content to consumer-resource model parameters and then used the consumer-resource model to predict metabolite dynamics in consortia, while Ho et al (18) used a coarse-grained consumer-resource model to characterize communities assembled from a 15-species pool. This work highlights how conservative metabolomics and spent-media experiments pave the way for using coarse-grained CR models to make predictions for community function.

In addition to the generalized Lotka-Volterra model and consumer-resource dynamics, more detailed metabolic models also provide a way of understanding, designing, and controlling microbiomes. Consumer-resource models themselves can (at least qualitatively) capture several more complex relationships relating to the uptake and processing of resources. But sequencing across a broad taxonomic range means that we are able to map the precise metabolic pathways of many individual taxa (56–58). Genome-scale metabolic models for individual taxa can be integrated to build models that capture metabolic functions in whole communities and also summarize interactions between species (59–61). However, most of these consortia comprise only a few strains, bringing scalability into question. While both consumer-resource and genome-scale models speak to the use of resources, they lie at either end of a mechanistic spectrum. It could be that intermediate, coarse-grained descriptions of internal metabolism might provide a bridge, trading off data overhead (the potentially prohibitive requirements for appropriate training data) and mechanistic accuracy. Metabolically informed consumer-resource models can give insights into small consortia, correctly capturing dynamics and interaction mechanisms (62–66).

In summary, every mechanistic model we know of has its drawbacks, centering around difficulties in accurately formulating and fitting these models given limited data. The full spectrum of mechanisms which underpin microbiome stability also includes spatial structure and dynamics resulting from spatial heterogeneity, adding further complexity to the application of these models. Any more exhaustive range of mechanistic models would have many of the same (or exacerbated) issues of data overhead and model specification.

## **Data-driven modelling and model-free approaches**

Mechanistic models serve as obvious starting points to build an architecture for microbiomes, but have clear limitations. Integration of multiple approaches may provide a key step needed for effective design and control of microbiomes. For example, Brunner and Chia (67) combine pairwise genome-scale modeling with generalized Lotka-Volterra equations in the

context of engraftment of probiotics in the human gut microbiome. The effectiveness of this particular approach rests on two inputs: using previously-published GEMs or genomes to build a qualitative, backbone network of possible microbe-microbe interactions, and then inferring quantitative gLV parameters on this interaction network.

Moving beyond this kind of gluing together different types of mechanistic approaches, there has also been a trend to employ statistical and machine learning methods for predicting outcomes for a specific microbiome composition, without any reference to the specific mechanisms underlying those outcomes. We think of these as data-driven, or model-free approaches, meaning that there is no specific model choice invoked to structure the predictions being made. On the other hand, there clearly are choices being made in order to build these statistical predictions, even if these choices are not biologically-motivated, and this has led to a range of distinct statistical methods being used to make predictions for microbiomes.

For example, Baranwal et al (24) have shown that a variant of recurrent neural networks (RNN-LSTM) trained on temporal data can outperform a generalized Lotka-Volterra model, especially when tested on data from a simulated model with higher-order interactions. That Lotka-Volterra models cannot recapitulate higher order interactions might not be surprising, but the success of the neural network approach was not guaranteed either, and yet made it possible to predict multiple community functions, primarily concentrations of short chain fatty acids, along with the abundances of species in a 25-species dataset, generated in vitro. In the related context of predicting colonization success of a given strain, mechanism-agnostic approaches like logistic regression, random forest regression, and compositional neural nets have also been used to make successful predictions (68). It has been proposed that more generally we may be able to guide design of communities and understand co-variation in community composition using generative models (69), and a range of methods have been employed to predict (70–73) and understand interactions amongst microbes (74).

Despite this promising array of methods, the single most highlighted downside to data-driven methods may be that our level of understanding is limited, and biological interpretability is compromised. That is, we might be able to predict outcomes successfully within a given parameter range, but we do not know *why* we are predicting these well, or where this prediction might begin to break down. An important feature of these approaches has been the degree to which predictions are constrained in order to bake in biologically sensible outcomes (75). These include obvious constraints, like ensuring that species abundances are non-negative and species which were absent in an assemblage are not predicted to be present in the steady-state outcome. Similarly, when the target variable for prediction is a metabolite, the target value can also be constrained by known biochemical limits and stoichiometry. For interpretability, LIME (76) and SHAP (77) have been used to partially address this issue by building post-hoc interaction networks, after the machine learning has been carried out. This approach can thus provide some insight into biological mechanisms (78). Even so, these models may need to be tailored for the specific datasets they were built for, and thus require overhauling when translating to other data, undermining their generality.

## The Middle Ground

Recent work has made progress using statistical approaches which straddle the divide between mechanistic models and statistical learning. The central object of interest addressed so far in these approaches is the ecological landscape. Inspired by landscapes in evolutionary biology which map genotypes to phenotypes (79, 80), these ecological landscapes map initial community composition to steady-state abundances and functions. Community landscapes have been defined and studied in multiple recent papers, showing that it may be possible to efficiently navigate this landscape to design microbial communities (81–83).

What exactly defines the ‘middle-ground’-ness of this kind of landscape thinking? First, the object of interest is fundamentally biological, given that the effect of individual taxa on other taxon abundances, and/or community function, naturally emerges from the landscape framework. Thus, a certain kind of biological interaction is inherently part of this description. Further, multiple studies demonstrate that across a variety of environments, both abundances and functions at steady-state can be predicted from species presence/absence data (84–87), and these predictions do not rest on the assumption of any particular mechanistic framework.

The success of these approaches show that microbial landscapes are highly constrained, with only a few strong interactions. This signal of sparsity in the landscape interaction space, akin to sparse higher-order epistasis in protein and genomics datasets (88), can be leveraged to infer entire landscapes from limited experimental data using regression techniques with appropriate weightings (86). We think that these signatures of sparsity may be used to a) aid design of experiments, b) engineer desirable microbial communities, c) inform and refine mechanism and interaction based models like CR and gLV, and d) define the role of environmental heterogeneity when probing the same sets of species across habitats. Further development of these approaches may generalize the input-output map to incorporate environmental conditions and nutrient availability, alongside

taxonomic composition, as part of the model input, to predict community properties as an output. The landscape might then incorporate regimes where nutrients determine function independent of taxonomy (44), regimes where taxonomy is key (86), and regimes where both act in concert to determine function.

On the other hand, several challenges remain for the landscape framework and the middle-ground. The most obvious is that the landscape has so far been treated as a map from initial composition to a steady-state—a description that inspired our S-matrix analogy in the introduction above. But several studies show that steady-states may not be guaranteed, even in the very long-term (90), or typical for microbial communities (91, 92). Thus whether this approach can be extended to incorporate those situations will be key for its generality. Another complication is that even if environmental conditions and nutrient availability are successfully incorporated into the landscape approach, these too may be highly variable over time. The success of middle-ground approaches may lie in thinking of them as limiting cases. Or, it could be that the landscape thinking explored in recent papers turns out to be just one corner of the middle-ground, and that there may be many other approaches that combine interpretability alongside statistical tractability.

## Discussion

Approaches to the control and design of microbiomes with optimal functions must wrestle with the question of what is important: no single model can capture and integrate the complexity posed by all the fluctuating biotic and abiotic components of an ecosystem. Although phenomenological and mechanistic models like the generalized Lotka-Volterra and the consumer-resource models can be, and have been used to design and test functionally beneficial communities, their success in making accurate predictions is compromised by either model limitations or the number of parameters to fit. This means that models like generalized Lotka-Volterra, consumer-resource models, and even metabolic models mostly enjoy success when employed to understand and explain statistical patterns of communities, rather than precise predictions of function—at least in cases where these communities comprise large numbers of diverse taxa. On the other hand, approaches which rely on machine learning methods tend to sideline mechanisms and hence may not be straightforwardly translated to datasets beyond those they were developed for. In between these two sets of approaches, methods which rely on finding low-dimensional representations of microbial community-function landscapes may prove to be useful and worthy of further exploration.

We think that some recent successes lay out a potential roadmap, integrating traditional modeling and statistical learning to understand, predict, and design microbiomes. The key here may be to find the right unit of analysis. Coarse-grained taxonomic units, or functional groups, will likely play a role in making microbiome analysis tractable. Although this idea has been around for a while in ecology, it may need to be revisited and re-hauled in the light of importance of microbial metabolism and consumer-resource dynamics. These suggest that we may need to go beyond just finding the right taxonomic units: we will likely need to identify coarse-grained *modules* of consumers and resources (18), encoding the key processes, not just the key players in a microbial community. In short, we need to identify functional groups in natural communities using methods that go beyond taxonomy and phylogeny (93).

The end-point of this back-and-forth might resemble progress in other fields. The particle physics S-matrix was at one point proposed to be uniquely determined by fundamental principles (94). Later, it was thought of as predictable, but only given knowledge of the specific combinations of symmetries and particles in nature, and modeled using the language of field theory (95). While now the S-matrix is sometimes thought of again as an object of study in its own right, with certain fundamental features that are independent of any specific, mechanistic models (19). The map from input nutrients and taxa to late-time community function might turn out to be the S-matrix of microbiome ecology. Understanding this input-output map may similarly benefit from back-and-forth between mechanistic models and the identification of fundamental, inevitable structures. Simplifying features in the composition-function landscape certainly suggest that there may be robust rules that govern this microbial S-matrix. But it could also be that we will only be able to leverage those rules for microbiome design if we formulate microbiome composition and dynamics in the right language, and using the right variables.

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1. Wen-Sheng Shu and Li-Nan Huang. Microbial diversity in extreme environments. *Nature Reviews Microbiology*, 20(4):219–235, 2022.
2. Courtney J Robinson, Brendan JM Bohannan, and Vincent B Young. From structure to function: the ecology of host-associated microbial communities. *Microbiology and Molecular Biology Reviews*, 74(3):453–476, 2010.
3. Paul G Falkowski, Tom Fenchel, and Edward F Delong. The microbial engines that drive earth's biogeochemical cycles. *science*, 320(5879):1034–1039, 2008.
4. Siyu Chen, Shuyan Luo, and Chao Yan. Gut microbiota implications for health and welfare in farm animals: A review. *Animals*, 12(1):93, 2021.
5. Hirokazu Toju, Kabir G Peay, Masato Yamamichi, Kazuhiko Narisawa, Kei Hiruma, Ken Naito, Shinji Fukuda, Masayuki Ushio, Shinji Nakaoka, Yusuke Onoda, et al. Core microbiomes for sustainable agroecosystems. *Nature plants*, 4(5):247–257, 2018.
6. Kaijian Hou, Zhuo-Xun Wu, Xuan-Yu Chen, Jing-Quan Wang, Dongya Zhang, Chuanxing Xiao, Dan Zhu, Jagadish B Koya, Liuya Wei, Jilin Li, et al. Microbiota in health and diseases. *Signal transduction and targeted therapy*, 7(1):1–28, 2022.
7. Ricardo Cavicchioli, William J Ripple, Kenneth N Timmis, Farooq Azam, Lars R Bakken, Matthew Baylis, Michael J Behrenfeld, Antje Boetius, Philip W Boyd, Aimée T Classen, et al. Scientists' warning to humanity: microorganisms and climate change. *Nature Reviews Microbiology*, 17(9):569–586, 2019.
8. Jean-Luc LeGras, Didier Merdinoglu, Jean-Marie Cornuet, and Francis Karst. Bread, beer and wine: *Saccharomyces cerevisiae* diversity reflects human history. *Molecular ecology*, 16(10):2091–2102, 2007.

9. Thomas Louie, Yoav Golan, Sahil Khanna, Dmitri Bobilev, Nathalie Eperding, Candida Fratazzi, Meg Carini, Rajita Menon, Mary Ruisi, Jason M Norman, et al. Ve303, a defined bacterial consortium, for prevention of recurrent clostridioides difficile infection: a randomized clinical trial. *Jama*, 329(16):1356–1366, 2023.
10. Posy E Busby, Chinmay Soman, Maggie R Wagner, Maren L Friesen, James Kremer, Alison Bennett, Mustafa Morsy, Jonathan A Eisen, Jan E Leach, and Jeffery L Dangl. Research priorities for harnessing plant microbiomes in sustainable agriculture. *PLoS biology*, 15(3):e2001793, 2017.
11. Yuancai Chen, Che-Jen Lin, Gavin Jones, Shiyu Fu, and Huaiyu Zhan. Enhancing biodegradation of wastewater by microbial consortia with fractional factorial design. *Journal of hazardous materials*, 171(1-3):948–953, 2009.
12. Sudarshan Paramsothy, Michael A Kamm, Nadeem O Kaakoush, Alissa J Walsh, Johan van den Bogaerde, Douglas Samuel, Rupert WL Leong, Susan Connor, Watson Ng, Ramesh Paramsothy, et al. Multidonor intensive faecal microbiota transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *The Lancet*, 389(10075):1218–1228, 2017.
13. Jasmine Shong, Manuel Rafael Jimenez Diaz, and Cynthia H Collins. Towards synthetic microbial consortia for bioprocessing. *Current Opinion in Biotechnology*, 23(5):798–802, 2012.
14. Wael Sabra, David Dietz, Donna Tjahjarsari, and An-Ping Zeng. Biosystems analysis and engineering of microbial consortia for industrial biotechnology. *Engineering in Life Sciences*, 10(5):407–421, 2010.
15. Curtis Huttenhower, Dirk Gevers, Rob Knight, Sahar Abubucker, Jonathan H. Badger, Asif T. Chinwalla, Heather H. Creasy, Ashlee M. Earl, Michael G. FitzGerald, Robert S. Fulton, Michelle G. Giglio, Kymberlie Hallsworth-Pepin, Elizabeth A. Lobos, Ramana Madupu, Vincent Magrini, John C. Martin, Makedonka Mitreva, Donna M. Muzny, Erica J. Sodergren, James Versalovic, Aye M. Wollam, Kim C. Worley, Jennifer R. Wortman, Sarah K. Young, Qiangdong Zeng, Kjersti M. Aagaard, Olukemi O. Abolude, Emma Allen-Vercoe, Eric J. Alm, Lucia Alvarado, Gary L. Andersen, Scott Anderson, Elizabeth Appelbaum, Harindra M. Arachchi, Gary Armitage, Cesar A. Arze, Tulin Ayvaz, Carl C. Baker, Lisa Beggs, Tsegahiwot Belachew, Veena Bhonagiri, Monika Bihan, Martin J. Blaser, Toby Bloom, Vivien Bonazzi, J. Paul Brooks, Gregory A. Buck, Christian J. Buhay, Dana A. Busam, Joseph L. Campbell, Shane R. Canon, Brandi L. Cantarel, Patrick S. G. Chain, I-Min A. Chen, Lei Chen, Shailla Chhibba, Ken Chu, Dawn M. Ciulla, Jose C. Clemente, Sandra W. Clifton, Sean Conlan, Jonathan Crabtree, Mary A. Cutting, Noam J. Davidovics, Catherine C. Davis, Todd Z. DeSantis, Carolyn Deal, Kimberley D. Delehaunty, Floyd E. Dewhurst, Elena Deych, Yan Ding, David J. Dooling, Shannon P. Dugan, Wm Michael Dunne, A. Scott Durkin, Robert C. Edgar, Rachel L. Erlich, Candace N. Farmer, Ruth M. Farrell, Karoline Faust, Michael Feldgarden, Victor M. Felix, Sheila Fisher, Anthony A. Fodor, Larry J. Forney, Leslie Foster, Valentina Di Francesco, Jonathan Friedman, Dennis C. Friedrich, Catrina C. Fronick, Lucinda L. Fulton, Hongyu Gao, Nathalia Garcia, Georgia Giannoukos, Christina Giblin, Maria Y. Giovanni, Jonathan M. Goldberg, Johannes Goll, Antonio Gonzalez, Allison Griggs, Sharvari Gujja, Susan Kinder Haake, Brian J. Haas, Holli A. Hamilton, Emily L. Harris, Theresa A. Hepburn, Brandi Herter, Diane E. Hoffmann, Michael E. Holder, Clinton Howarth, Katherine H. Huang, Susan M. Huse, Jacques Izard, Janet K. Jansson, Huaiyang Jiang, Catherine Jordan, Vandita Joshi, James A. Katanick, Wendy A. Keitel, Scott T. Kelley, Cristyn Kells, Nicholas B. King, Dan Knights, Heidi H. Kong, Omry Koren, Sergey Koren, Karthik C. Kota, Christie L. Kovar, Nikos C. Kyrpides, Patricio S. La Rosa, Sandra L. Lee, Katherine P. Lemon, Niall Lennon, Cecil M. Lewis, Lora Lewis, Ruth E. Ley, Kelvin Li, Konstantinos Liolios, Bo Liu, Yue Liu, Chien-Chi Lo, Catherine A. Lozupone, R. Dwayne Lunsford, Tessa Madden, Anup A. Mahurkar, Peter J. Mannon, Elaine R. Mardis, Victor M. Markowitz, Konstantinos Mavromatis, Jamison M. McCorison, Daniel McDonald, Jean McEwen, Amy L. McGuire, Pamela McInnes, Teena Mehta, Kathie A. Mihindukulasuriya, Jason R. Miller, Patrick J. Minx, Irene Newsham, Chad Nusbaum, Michelle O’Laughlin, Joshua Orvis, Ioanna Pagani, Krishna Palaniappan, Shital M. Patel, Matthew Pearson, Jane Stevenson, Mircea Podar, Craig Pohl, Katherine S. Pollard, Mihai Pop, Margaret E. Priest, Lita M. Proctor, Xiang Qin, Jeroen Raes, Jacques Ravel, Jeffrey G. Reid, Mina Rho, Rosamond Rhodes, Kevin P. Riehl, Maria C. Rivera, Beltran Rodriguez-Mueller, Yu-Hui Rogers, Matthew C. Ross, Carsten Russ, Ravi K. Sanka, Pamela Sankar, J. Fah Sathirapongsasuti, Jeffery A. Schloss, Patrick D. Schloss, Thomas M. Schmidt, Matthew Scholz, Lynn Schriml, Alyxandra M. Schubert, Nicola Segata, Julia A. Segre, William D. Shannon, Richard R. Sharp, Thomas J. Sharpton, Narmada Shenoy, Nihar U. Sheth, Gina A. Simone, Indresh Singh, Christopher S. Smillie, Jack D. Sobel, Daniel D. Sommer, Paul Spicer, Granger G. Sutton, Sean M. Sykes, Diana G. Tabbaa, Mathangi Thiagarajan, Chad M. Tomlinson, Manolito Torralba, Todd J. Treangen, Rebecca M. Truty, Tatiana A. Vishnivetskaya, Jason Walker, Lu Wang, Zhengyuan Wang, Doyle V. Ward, Wesley Warren, Mark A. Watson, Christopher Wellington, Kris A. Wetterstrand, James R. White, Katarzyna Wilczek-Boney, Yuanqing Wu, Kristine M. Wylie, Todd Wylie, Chandri Yandava, Liang Ye, Yuzhen Ye, Shibu Yooseph, Bonnie P. Youmans, Lan Zhang, Yanjiao Zhou, Yiming Zhou, Laurie Zoloth, Jeremy D. Zucker, Bruce W. Birren, Richard A. Gibbs, Sarah K. Highlander, Barbara A. Methé, Karen E. Nelson, Joseph F. Petrosino, George M. Weinstock, Richard K. Wilson, Owen White, and The Human Microbiome Project Consortium. Structure, function and diversity of the healthy human microbiome. *Nature*, 486(7402):207–214, June 2012. ISSN 1476-4687. doi: 10.1038/nature11234. Publisher: Nature Publishing Group.
16. Luke R Thompson, Jon G Sanders, Daniel McDonald, Amnon Amir, Joshua Ladau, Kenneth J Locey, Robert J Prill, Anupriya Tripathi, Sean M Gibbons, Gail Ackermann, et al. A communal catalogue reveals earth’s multiscale microbial diversity. *Nature*, 551(7681):457–463, 2017.
17. Ryan L. Clark, Bryce M. Connors, David M. Stevenson, Susan E. Hromada, Joshua J. Hamilton, Daniel Amador-Noguez, and Ophelia S. Venturelli. Design of synthetic human gut microbiome assembly and butyrate production. *Nat Commun*, 12(1):3254, December 2021. ISSN 2041-1723. doi: 10.1038/s41467-021-22938-y.
18. Po-Yi Ho, Taylor H Nguyen, Juan M Sanchez, Brian C DeFelicce, and Kerwyn Casey Huang. Resource competition predicts assembly of gut bacterial communities in vitro. *Nature Microbiology*, pages 1–13, 2024.
19. Sebastian Mizera. Physics of the analytic s-matrix. *Physics Reports*, 1047:1–92, 2024.
20. Wenping Cui, Robert Marsland III, and Pankaj Mehta. Les houches lectures on community ecology: From niche theory to statistical mechanics. *ArXiv*, 2024.
21. Matthieu Barbier, Jean-François Arnoldi, Guy Bunin, and Michel Loreau. Generic assembly patterns in complex ecological communities. *Proceedings of the National Academy of Sciences*, 115(9): 2156–2161, 2018.
22. Jiliang Hu, Daniel R Amor, Matthieu Barbier, Guy Bunin, and Jeff Gore. Emergent phases of ecological diversity and dynamics mapped in microcosms. 2022.
23. Aamir Faisal Ansari, Yugandhar BS Reddy, Janhavi Raut, and Narendra M Dixit. An efficient and scalable top-down method for predicting structures of microbial communities. *Nature Computational Science*, 1(9):619–628, 2021.
24. Mayank Baranwal, Ryan L Clark, Jaron Thompson, Zeyu Sun, Alfred O Hero, and Ophelia S Venturelli. Recurrent neural networks enable design of multifunctional synthetic human gut microbiome dynamics. *eLife*, 11:e73870, June 2022. ISSN 2050-084X. doi: 10.7554/eLife.73870.
25. Theo Gibbs, Gabriel Gellner, Simon A. Levin, Kevin S. McCann, Alan Hastings, and Jonathan M. Levine. Can higher-order interactions resolve the species coexistence paradox?, June 2023. Pages: 2023.06.19.545649 Section: New Results.
26. Theo Gibbs, Simon A. Levin, and Jonathan M. Levine. Coexistence in diverse communities with higher-order interactions. *Proceedings of the National Academy of Sciences*, 119(43):e2205063119, October 2022. doi: 10.1073/pnas.2205063119. Publisher: Proceedings of the National Academy of Sciences.
27. Michael T Pearce, Atish Agarwala, and Daniel S Fisher. Stabilization of extensive fine-scale diversity by ecologically driven spatiotemporal chaos. *Proceedings of the National Academy of Sciences*, 117(25):14572–14583, 2020.
28. Guy Bunin. Ecological communities with lotka-volterra dynamics. *Physical Review E*, 95(4):042414, 2017.
29. Vanni Bucchi, Belinda Tzen, Ning Li, Matt Simmons, Takeshi Tanoue, Elijah Bogart, Luxue Deng, Vladimir Yeliseyev, Mary L Delaney, Qing Liu, et al. Mdsine: Microbial dynamical systems inference engine for microbiome time-series analyses. *Genome biology*, 17(1):121, 2016.
30. Travis E. Gibson, Younhun Kim, Sawal Acharya, David E. Kaplan, Nicholas DiBenedetto, Richard Lavin, Bonnie Berger, Jessica R. Allegretti, Lynn Bry, and Georg K. Gerber. Microbial dynamics inference at ecosystem-scale, June 2023. Pages: 2021.12.14.469105 Section: New Results.
31. Charles K. Fisher and Pankaj Mehta. Identifying Keystone Species in the Human Gut Microbiome from Metagenomic Timeseries Using Sparse Linear Regression. *PLoS ONE*, 9(7):e102451, July 2014. ISSN 1932-6203. doi: 10.1371/journal.pone.0102451.
32. Ophelia S Venturelli, Alex V Carr, Garth Fisher, Ryan H Hsu, Rebecca Lau, Benjamin P Bowen, Susan Hromada, Trent Northen, and Adam P Arkin. Deciphering microbial interactions in synthetic human gut microbiome communities. *Molecular systems biology*, 14(6):e8157, 2018.
33. Sudeep Ghimire, Chayan Roy, Supapit Wongkuna, Linto Antony, Abhijit Maji, Mitchel Chan Keena, Andrew Foley, and Joy Scaria. Identification of clostridioides difficile-inhibiting gut commensals using culturomics, phenotyping, and combinatorial community assembly. *Msystems*, 5(1):10–1128, 2020.
34. Richard R Stein, Vanni Bucchi, Nora C Toussaint, Charlie G Buffie, Gunnar Rättsch, Eric G Pamer, Chris Sander, and João B Xavier. Ecological modeling from time-series inference: insight into dynamics and stability of intestinal microbiota. *PLoS computational biology*, 9(12):e1003388, 2013.
35. Alicia Sanchez-Gorostiaga, Djordje Bajić, Melisa L Osborne, Juan F Poyatos, and Alvaro Sanchez. High-order interactions distort the functional landscape of microbial consortia. *PLoS Biology*, 17(12):e3000550, 2019.
36. Glen D’Souza, Julia Schwartzman, Johannes Keegstra, Jeremy E Schreier, Michael Daniels, Otto X Cordero, Roman Stocker, and Martin Ackermann. Interspecies interactions determine growth dynamics of biopolymer-degrading populations in microbial communities. *Proceedings of the National Academy of Sciences*, 120(44):e2305198120, 2023.
37. Zongting Cai, Esther Karunakaran, and Jagroop Pandhal. Bottom-up construction and screening of algae-bacteria consortia for pollutant biodegradation. *Frontiers in Microbiology*, 15:1349016, 2024.
38. Babak Momeni, Li Xie, and Wenying Shou. Lotka-volterra pairwise modeling fails to capture diverse pairwise microbial interactions. *Elife*, 6:e25051, 2017.
39. James D Brunner and Nicholas Chia. Metabolite-mediated modelling of microbial community dynamics captures emergent behaviour more effectively than species–species modelling. *Journal of the Royal Society Interface*, 16(159):20190423, 2019.
40. Robert MacArthur. Species packing and competitive equilibrium for many species. *Theoretical Population Biology*, 1(1):1–11, 1970.
41. Robert Marsland III, Wenping Cui, Joshua Goldford, Alvaro Sanchez, Kiryll Korolev, and Pankaj Mehta. Available energy fluxes drive a transition in the diversity, stability, and functional structure of microbial communities. *PLoS computational biology*, 15(2):e1006793, 2019.
42. Joshua E. Goldford, Nanxi Lu, Djordje Bajić, Sylvie Estrela, Mikhail Tikhonov, Alicia Sanchez-Gorostiaga, Daniel Segrè, Pankaj Mehta, and Alvaro Sanchez. Emergent simplicity in microbial community assembly. *Science*, 361(6401):469–474, August 2018. ISSN 0036-8075, 1095-9203. doi: 10.1126/science.aat1168.
43. Lori Niehaus, Ian Boland, Minghao Liu, Kevin Chen, David Fu, Catherine Henckel, Kaitlin Chung, Suyen Espinoza Miranda, Samantha Dyckman, Matthew Crum, et al. Microbial coexistence through chemical-mediated interactions. *Nature communications*, 10(1):2052, 2019.
44. Ashish B George, Tong Wang, and Sergei Maslov. Functional convergence in slow-growing microbial communities arises from thermodynamic constraints. *The ISME Journal*, 17(9):1482–1494,

2023.

45. Zihan Wang, Akshit Goyal, Veronika Dubinkina, Ashish B George, Tong Wang, Yulia Fridman, and Sergei Maslov. Complementary resource preferences spontaneously emerge in diauxic microbial communities. *Nature communications*, 12(1):6661, 2021.
46. Akshit Goyal, Avi I Flamholz, Alexander P Petroff, and Arvind Murugan. Closed ecosystems extract energy through self-organized nutrient cycles. *Proceedings of the National Academy of Sciences*, 120(52):e2309387120, 2023.
47. Zihan Wang, Yu Fu, Akshit Goyal, and Sergei Maslov. Emergent ecological advantage of sequential metabolic strategies in complex microbial communities. *bioRxiv*, pages 2024–06, 2024.
48. Tong Wang, Ashish B George, and Sergei Maslov. Higher-order interactions in autotroph communities enhance their resilience to resource fluctuations. *bioRxiv*, pages 2024–05, 2024.
49. Leonardo Pacciani-Mori, Andrea Giometto, Samir Suweis, and Amos Maritan. Dynamic metabolic adaptation can promote species coexistence in competitive microbial communities. *PLoS computational biology*, 16(5):e1007896, 2020.
50. Zhiyuan Li, Bo Liu, Sophia Hsin-Jung Li, Christopher G King, Zemer Gitai, and Ned S Wingreen. Modeling microbial metabolic trade-offs in a chemostat. *PLoS computational biology*, 16(8):e1008156, 2020.
51. Kapil Amarnath, Avaneesh V Narla, Sammy Pontrelli, Jiajia Dong, Jack Reddan, Brian R Taylor, Tolga Caglar, Julia Schwartzman, Uwe Sauer, Otto X Cordero, et al. Stress-induced metabolic exchanges between complementary bacterial types underly a dynamic mechanism of inter-species stress resistance. *Nature communications*, 14(1):3165, 2023.
52. Stacey Butler and James P O'Dwyer. Cooperation and stability for complex systems in resource-limited environments. *Theoretical Ecology*, 13:239–250, 2020.
53. Robert Marsland III, Wenping Cui, and Pankaj Mehta. A minimal model for microbial biodiversity can reproduce experimentally observed ecological patterns. *Scientific reports*, 10(1):3308, 2020.
54. Robert Marsland, Wenping Cui, Joshua Goldford, and Pankaj Mehta. The Community Simulator: A Python package for microbial ecology. *PLoS ONE*, 15(3):e0230430, March 2020. ISSN 1932-6203. doi: 10.1371/journal.pone.0230430.
55. Karna Gowda, Derek Ping, Madhav Mani, and Seppe Kuehn. Genomic structure predicts metabolite dynamics in microbial communities. *Cell*, 185(3):530–546, 2022.
56. Scott Devoid, Ross Overbeek, Matthew DeJongh, Veronika Vonstein, Aaron A Best, and Christopher Henry. Automated genome annotation and metabolic model reconstruction in the seed and model seed. *Systems metabolic engineering: methods and protocols*, pages 17–45, 2013.
57. Daniel Machado, Sergej Andrejev, Melanie Tramontano, and Kiran Raosaheb Patil. Fast automated reconstruction of genome-scale metabolic models for microbial species and communities. *Nucleic acids research*, 46(15):7542–7553, 2018.
58. Peter D Karp, Peter E Midford, Richard Billington, Anamika Kothari, Markus Krummenacker, Mario Latendresse, Wai Kit Ong, Pallavi Subhraveti, Ron Caspi, Carol Fulcher, et al. Pathway tools version 23.0 update: software for pathway/genome informatics and systems biology. *Briefings in bioinformatics*, 22(1):109–126, 2021.
59. Timothy J Hanly and Michael A Henson. Dynamic metabolic modeling of a microaerobic yeast co-culture: predicting and optimizing ethanol production from glucose/xylose mixtures. *Biotechnology for biofuels*, 6:1–16, 2013.
60. Beatriz García-Jiménez, José Luis García, and Juan Nogales. Flycop: metabolic modeling-based analysis and engineering microbial communities. *Bioinformatics*, 34(17):i954–i963, 2018.
61. Christian Diener, Sean M Gibbons, and Osbaldo Resendis-Antonio. Micom: metagenome-scale modeling to infer metabolic interactions in the gut microbiota. *MSystems*, 5(1):10–1128, 2020.
62. Mario E Muscarella and James P O'Dwyer. Species dynamics and interactions via metabolically informed consumer-resource models. *Theoretical Ecology*, 13(4):503–518, 2020.
63. Chen Liao, Tong Wang, Sergei Maslov, and Joao B Xavier. Modeling microbial cross-feeding at intermediate scale portrays community dynamics and species coexistence. *PLoS computational biology*, 16(8):e1008135, 2020.
64. Tim N Enke, Manoshi S Datta, Julia Schwartzman, Nathan Cermak, Désirée Schmitz, Julien Barrere, Alberto Pascual-García, and Otto X Cordero. Modular assembly of polysaccharide-degrading marine microbial communities. *Current Biology*, 29(9):1528–1535, 2019.
65. Matti Gralka, Shaul Pollak, and Otto X Cordero. Genome content predicts the carbon catabolic preferences of heterotrophic bacteria. *Nature Microbiology*, 8(10):1799–1808, 2023.
66. Jean CC Vila, Joshua Goldford, Sylvie Estrela, Djordje Bajic, Alicia Sanchez-Gorostiaga, Alejandro Damian-Serrano, Nanxi Lu, Robert Marsland III, Maria Rebolleda-Gomez, Pankaj Mehta, et al. Metabolic similarity and the predictability of microbial community assembly. *bioRxiv*, 2023.
67. James D Brunner and Nicholas Chia. Metabolic model-based ecological modeling for probiotic design. *Elife*, 13:e83690, 2024.
68. Shengbo Wu, Zheping Qu, Danlei Chen, Hao Wu, Qingge Caiyin, and Jianjun Qiao. Deciphering and designing microbial communities by genome-scale metabolic modelling. *Computational and Structural Biotechnology Journal*, 2024.
69. Germán Plata, Madan Krishnamurthy, Lukas Herron, and Purushottam Dixit. Designing host-associated microbiomes using the consumer/resource model, January 2024. Pages: 2023.04.28.538625 Section: New Results.
70. Sebastian Michel-Mata, Xu-Wen Wang, Yang-Yu Liu, and Marco Tulio Angulo. Predicting microbiome compositions from species assemblages through deep learning. *iMeta*, 1(1):e3, 2022. ISSN 2770-596X. doi: 10.1002/imt2.3. eprint: <https://onlinelibrary.wiley.com/doi/pdf/10.1002/imt2.3>.
71. Eitan E. Asher and Amir Bashan. Model-free prediction of microbiome compositions. *Microbiome*, 12(1):17, February 2024. ISSN 2049-2618. doi: 10.1186/s40168-023-01721-9.
72. Tong Wang, Xu-Wen Wang, Kathleen A. Lee-Sarwar, Augusto A. Litorjua, Scott T. Weiss, Yizhou Sun, Sergei Maslov, and Yang-Yu Liu. Predicting metabolomic profiles from microbial composition through neural ordinary differential equations. *Nat Mach Intell*, 5(3):284–293, March 2023. ISSN 2522-5839. doi: 10.1038/s42256-023-00627-3. Number: 3 Publisher: Nature Publishing Group.
73. Sebastian Michel-Mata, Xu-Wen Wang, Yang-Yu Liu, and Marco Tulio Angulo. Predicting microbiome compositions from species assemblages through deep learning. *iMeta*, 1(1):e3, 2022.
74. Kateryna Melnyk, Kuba Weimann, and Tim OF Conrad. Understanding microbiome dynamics via interpretable graph representation learning. *Scientific Reports*, 13(1):2058, 2023.
75. Jaron C. Thompson, Victor M. Zavala, and Ophelia S. Venturelli. Integrating a tailored recurrent neural network with Bayesian experimental design to optimize microbial community functions. *PLOS Computational Biology*, 19(9):e1011436, September 2023. ISSN 1553-7358. doi: 10.1371/journal.pcbi.1011436. Publisher: Public Library of Science.
76. Marco Tulio Ribeiro, Sameer Singh, and Carlos Guestrin. Why should I trust you? explaining the predictions of any classifier. In *Proceedings of the 22nd ACM SIGKDD International Conference on Knowledge Discovery and Data Mining*, pages 1135–1144. ACM, 2016. doi: 10.1145/2939672.2939778.
77. Scott M Lundberg and Su-In Lee. A unified approach to interpreting model predictions. In *Proceedings of the 31st International Conference on Neural Information Processing Systems (NIPS'17)*, pages 4768–4777. Curran Associates Inc., 2017. doi: 10.48550/arXiv.1705.07874.
78. Martina Dal Bello, Hyunseok Lee, Akshit Goyal, and Jeff Gore. Resource-diversity relationships in bacterial communities reflect the network structure of microbial metabolism. *Nature Ecology & Evolution*, 5(10):1424–1434, 2021.
79. Daniel M Weinreich, Yinghong Lan, C Scott Wylie, and Robert B Heckendorn. Should evolutionary geneticists worry about higher-order epistasis? *Current opinion in genetics & development*, 23(6):700–707, 2013.
80. Daniel M Weinreich, Yinghong Lan, Jacob Jaffe, and Robert B Heckendorn. The influence of higher-order epistasis on biological fitness landscape topography. *Journal of statistical physics*, 172:208–225, 2018.
81. Ashish B George and Kirill S Korolev. Ecological landscapes guide the assembly of optimal microbial communities. *PLoS Computational Biology*, 19(1):e1010570, 2023.
82. Alvaro Sanchez, Djordje Bajic, Juan Diaz-Colunga, Abigail Skwara, Jean C. C. Vila, and Seppe Kuehn. The community-function landscape of microbial consortia. *Cell Systems*, 14(2):122–134, February 2023. ISSN 2405-4712. doi: 10.1016/j.cels.2022.12.011.
83. Chandana Gopalakrishnappa, Karna Gowda, Kaumudi H Prabhakara, and Seppe Kuehn. An ensemble approach to the structure-function problem in microbial communities. *IScience*, 25(2), 2022.
84. Daniel S Maynard, Zachary R Miller, and Stefano Allesina. Predicting coexistence in experimental ecological communities. *Nature ecology & evolution*, 4(1):91–100, 2020.
85. Abigail Skwara, Karna Gowda, Mahmoud Yousef, Juan Diaz-Colunga, Arjun S Raman, Alvaro Sanchez, Mikhail Tikhonov, and Seppe Kuehn. Statistically learning the functional landscape of microbial communities. *Nature Ecology & Evolution*, 7(11):1823–1833, 2023.
86. Shreya Arya, Ashish B George, and James P O'Dwyer. Sparsity of higher-order landscape interactions enables learning and prediction for microbiomes. *Proceedings of the National Academy of Sciences*, 120(48):e2307313120, 2023.
87. J. Diaz-Colunga, A. Skwara, J. C. C. Vila, D. Bajic, and A. Sanchez. Global epistasis and the emergence of function in microbial consortia. *Cell*, 187(12):3108–3119, 2024.
88. David H Brookes, Amirali Aghazadeh, and Jennifer Listgarten. On the sparsity of fitness functions and implications for learning. *Proceedings of the National Academy of Sciences*, 119(1):e2109649118, 2022.
89. Ashish B George, Tong Wang, and Sergei Maslov. Functional universality in slow-growing microbial communities arises from thermodynamic constraints. *arXiv preprint arXiv:2203.06128*, 2022.
90. Benjamin H Good, Michael J McDonald, Jeffrey E Barrick, Richard E Lenski, and Michael M Desai. The dynamics of molecular evolution over 60,000 generations. *Nature*, 551(7678):45–50, 2017.
91. Zak Frenzt, Seppe Kuehn, and Stanislas Leibler. Strongly deterministic population dynamics in closed microbial communities. *Physical Review X*, 5(4):041014, 2015.
92. Jiliang Hu, Daniel R Amor, Matthieu Barbier, Guy Bunin, and Jeff Gore. Emergent phases of ecological diversity and dynamics mapped in microcosms. *Science*, 378(6615):85–89, 2022.
93. X. Shan. *Identifying functional groups in microbial communities based on ecological patterns*. PhD thesis, Massachusetts Institute of Technology, 2023.
94. Geoffrey F Chew and Steven C Frautschi. Principle of equivalence for all strongly interacting particles within the s-matrix framework. *Physical Review Letters*, 7(10):394, 1961.
95. Steven Weinberg. Conceptual foundations of the unified theory of weak and electromagnetic interactions. *Reviews of Modern Physics*, 52(3):515, 1980.

## Annotations

- 18 \* This study found that resource competition shapes ecological outcomes in a synthetic community of 15 gut strains derived from humanized mice. They demonstrate how a coarse-grained consumer-resource model can be developed from supernatant screening and untargeted metabolomics data. This model was then successfully employed to predict abundances of strains in various assemblages.
- 65 \*\* This study highlights how genome content information can be applied to predict coarse-grained descriptors of a community, in this case, the preference for glycolytic versus gluconeogenic substrates in a community of 186 bacteria on 140 carbon sources. This description of the community in terms of preference for either sugars or acid may help provide insights into interactions within this bacterial community.
- 66 \*\* This study shows that patterns in community compositions can be discerned by linking shared metabolic traits and substrate similarity. This leads to successful predictions of community compositions in novel environments.
- 81 \*\* This study develops upon, in detail, the idea of structure-function landscapes for microbiomes, inspired from fitness landscapes in evolutionary biology. It links features of a community, like niche overlap between its members, with the ruggedness of the corresponding structure-function landscape.
- 85 \*\* This study provides a middle-ground approach to study community functions and the structure-function landscape. They demonstrate for various synthetic microcosms that the structure-function landscape is smooth, which makes function prediction possible from relatively low number of observed data-points.
- 86 \*\* This study provides a middle-ground approach to study community abundances and functions and the structure-function and community- composition landscape. They show that the low ruggedness of these landscapes translates to dominance of effective, low-order interactions amongst the community members. This observation was consequently used to predict abundances and functions using a method that down-weighs higher-order interactions.