

Should we really predict the response of wild endotherms to climate change based on thermal responses measured in captivity?

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In their recent study, Beaman et al. [1] measured the metabolic response of the most abundant Arctic seabird, the dovekie (*Alle alle*), to experimental variation in air temperature. Based on their results on energy and water expenditure in captive conditions, Beaman et al. conclude that this Arctic species should not suffer much from direct consequences of climate change. Indeed, their estimated upper critical temperature T_{UC} (*i.e.* temperature above which metabolic rate increases) was 22.4°C, which is *ca.* 15°C higher than the average air temperature, and *ca.* 2°C higher than the maximum temperature currently recorded at the breeding colony. While projecting their data under various climatic scenario, Beaman et al.[1] conclude that dovekies should even benefit of the ongoing climate warming from a thermoregulatory perspective.

While the data collection is quite standard in the field of thermal biology [2], I would like to offer my personal view on the potential perils of concluding about wild animal response to climate change based on data collected in ‘a small dark box in the lab’. Classically, the thermal response of animals is evaluated using respirometry, which enables to quantify gas exchange (*i.e.* O₂, CO₂, H₂O) between an animal and its environment in response to variation in ambient temperature within a small metabolic chamber [2]. This is usually conducted in a post-absorptive state and in the dark to limit energy expenditure linked to digestion and physical activity. While such conditions are highly relevant to evaluate the minimum energy requirements of animals (*i.e.* basal metabolic rate), I question its relevance for evaluating the thermal responses of wild animals to the heat.

First, the risk for wild animals to suffer from warm environmental conditions are very unlikely to occur in a post-absorptive state during the night. The thermal physiology and the responses to warm conditions are thus likely to differ between an animal fasting and resting within a metabolic chamber and the same animal within its natural environment. For instance, the body temperatures of doves measured by Beaman et al. [1] during their respirometry assays differ markedly from the body temperature measured on the same species while attending the breeding colony [3] as illustrated in Fig. 1A. Doves at the breeding colony at an average air temperature (T_a) of 5.4°C had a body temperature 1.5°C higher than doves measured in respirometry chambers below the thermal inflexion point (average $T_a = 14.5^\circ\text{C}$), and 1.1°C higher than those measured above the thermal inflexion point (average $T_a = 28.4^\circ\text{C}$). Of course, those animals are not fully comparable, and for instance T_b has been shown to decrease within the first hour following the return to the colony [3]. Yet, even if we select data for wild doves after more than 1 hour spent at the colony, being fully inactive (vectors of the dynamic body acceleration ≤ 0.05) and within a range of T_a comparable to captive birds (*i.e.* $T_a > 5.0^\circ\text{C}$), their body temperature remains 0.8°C higher than birds in a respirometry chamber under relative similar T_a (Fig 1B).

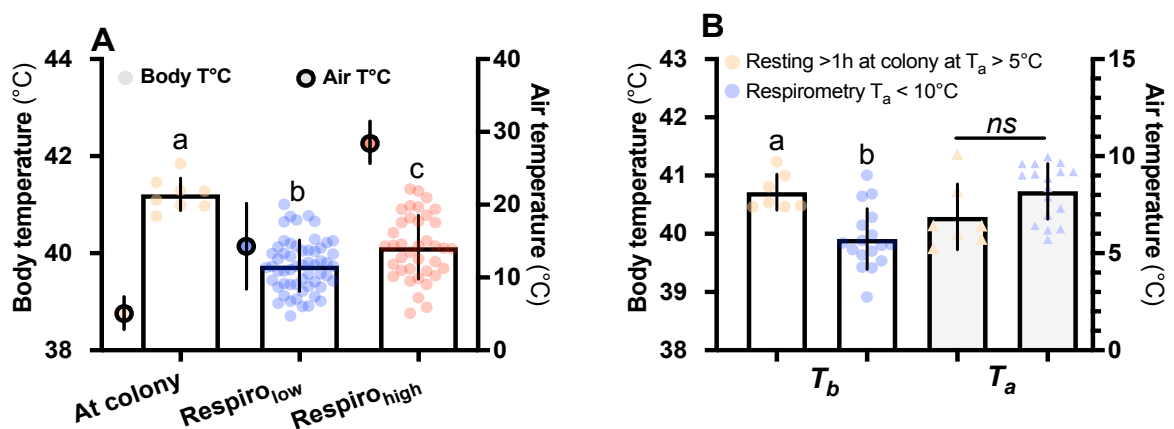


Fig. 1: Comparison of body temperature (T_b) between free-ranging doves attending the breeding colony and wild-caught individuals held in respirometry chambers. A) Doves measured at the corresponding air temperature (T_a) *in situ* while attending the breeding colony (yellow), and during respirometry assay both below (blue) and above (red) the thermal inflexion point. Data at the colony were retrieved and averaged for the $N = 8$ individuals ($n = 4352$ measures) from [3], while data for respirometry have been extracted from Fig. 2c in [1] using <https://plotdigitizer.com/> but cannot be averaged for each individual ($N = 36$, $n = 95$ measures) considering the lack of information on individual identity. Mean \pm SD are presented and have been analyzed for body temperature using non-parametric Kruskal-Wallis test ($K-W \chi^2 = 26.3$, $p < 0.001$). Different letters represent significant differences in body temperature according to Dunn's post-hoc tests (all $p < 0.020$). B) **Free-ranging and wild-caught doves measured in the most comparable status possible. Data for free-ranging individuals [3] have been restricted to datapoints gathered after more than one hour attending the colony, being in a resting state (vectors of the dynamic body acceleration ≤ 0.05) and at temperatures comparable to captive individuals (*i.e.* $T_a > 5^\circ\text{C}$; $N = 7$, $n = 707$). Data for captive individuals [1] have been restricted to measurements conducted below 10°C ($N = 17$). Mean \pm SD are presented and have been analyzed using non-parametric Mann-Whitney tests (For T_b : $p = 0.0025$; for T_a : $p = 0.11$).**

Dovekies thus seem to be in a somewhat hypometabolic state while sitting in a dark respirometry chamber in a post-absorptive state. The results of Beaman et al. [1] for body temperature are relatively similar to those obtained in dovekies measured in respirometry chambers under full-light conditions ($T_b = 40.1 \pm 0.4^\circ\text{C}$) [4], which suggests that light exposure is unlikely the main driver of the low T_b observed during respirometry. From my perspective, it seems that any conclusion on the thermal response of dovekies measured from respirometry studies should not be used, or used with extreme caution, to extrapolate the thermal response of free-living dovekies, especially to warm environmental conditions. While the precious data on body temperature collected by [3] and [1] on dovekies regarding this specific point is striking, it is likely that such assumption may apply to other wild endotherms.

Additionally, it is worth noting that the shape of the classical metabolism-ambient temperature curve measured for dovekies differ markedly between [1] and a previous study on the same species [4]. Here I put together these two studies in perspective in Figure 2 by restricting [4] to $T_a > 5^\circ\text{C}$ (*i.e.* the lower T_a used in [1]) and mean-centring all metabolic data. While the breakpoints do not differ drastically (*i.e.* 20.11, 95% C.I. [18.80-20.68] vs. 22.53 [19.63-25.00] $^\circ\text{C}$), there is notably less variation in the data of [4] (see spreading across the y axis), and the markedly steeper slope after the inflection point of this study suggests a cost of thermoregulation at high temperatures incommensurably higher than the data obtained in [1]. Such differences may potentially arise due to differences in thermal biology between populations of the same species, since birds of [1] originated from East Greenland and those of [4] from Svalbard. This seems rather important to consider when forecasting the susceptibility of a species to climate change, as done by Beaman et al. [1]. Additionally, while Beaman et al. [1] mention that “in dovekies at least, the energetic cost of heat production is greater than the energetic cost of heat dissipation”, this statement does not seem supported by their data, with the 95% confidence intervals of the slopes below [-0.153; -0.078] and above [0.040; 0.156] T_{UC} largely overlapping when converted to absolute value. Finally, the acknowledged inclusion of biologically unrealistic metabolic rate data (respiratory exchange ratio [0.45-0.65] and [1.00-1.20]) question the precision of Beaman et al. [1] respirometry data from my perspective.

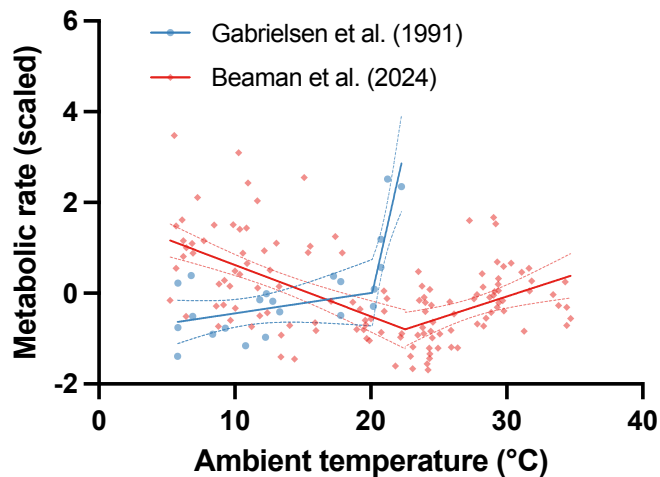


Fig. 2: Metabolic responses of captive dovekeys to experimental variation in ambient temperature measured by Beaman et al. [1] (blue) and Gabrielsen et al. [4] (red). Metabolic data on mass-corrected VO_2 (Fig. 2a in [1]) and VCO_2 (Fig. 1 in [4]) have been extracted using <https://plotdigitizer.com/> and mean-centred (*i.e.* scaled) to enable comparison of patterns between the two studies. Segmented regression analyses defining breakpoints, slopes and their confidence intervals have been performed using GraphPad Prism v10.3.0.

Secondly, Beaman et al. [1] seem to assume that wild dovekeys are only submitted to variation in air temperature. The heat load faced by wild animals depends on more than T_a alone. T_a has even been suggested to play a relatively minor role in the actual heat load faced by many wild animals [5]. While it seems obvious that our human thermal comfort depends on the combination of air temperature, solar radiations, wind speed and humidity, many studies related to the thermal biology of wild animals continue to ignore the last 3 factors in their analyses, including Beaman et al. [1]. To illustrate the importance of solar radiations for seabirds, it has for example been shown that while panting behaviour occurs at a T_a of *ca.* 24°C in the shade, it occurs at *ca.* 15°C when common guillemots (*Uria aalge*) are exposed to solar radiations [6]. It has recently been suggested that a field-friendly and informative way of measuring the heat load faced by wild animals would be to use the black globe temperature (T_g) instead of T_a [5], which integrates information related to T_a , solar radiations, wind speed and humidity. Scientists working with farmed animals have also developed and validated several indexes to quantify heat load, including the widely used Heat Load Index (*HLI*) that incorporates T_a , relative humidity, wind speed and solar radiations [8]. While measured in a respirometry chamber, animals are not exposed to wind nor to solar radiations, and are often provided with dried air that facilitates analytical calculations but also facilitates passive evaporative cooling [9]. The humidity at which Beaman et al. [1] measured dovekeys does not seem to be reported, and I thus assumed three scenarios for the following calculations, namely

dry air ($RH = 0\%$), moderate humidity ($RH = 50\%$) and saturated air ($RH = 100\%$). Based on the climatic conditions (T_a , wind speed, RH) experienced by wild doves from [3] and the solar radiations estimated at their study site during the study period (<https://open-meteo.com/en/docs/historical-weather-api>), I calculated both the T_g (based on equation from [7]) and HLI (based on equation for $T_g < 25^\circ\text{C}$ from [8]) experienced by wild doves and putted them in perspective of the T_g and HLI experienced within the respirometry chamber at the upper critical temperature determined by [1] (Fig. 3). While Beaman et al. [1] seem to conclude that wild doves should not really suffer from heat stress based on T_a alone, the data presented in Fig. 3 suggest that wild doves are most likely experiencing environmental conditions above their upper critical temperature quite frequently when solar radiations are high. It is also worth noting here that the maximum local T_a reported by [3] over their 11-days study period and used for Fig. 3 calculations is *ca.* 13.5°C , while local T_a can reach more than 20°C according to both [1] and (<https://open-meteo.com/en/docs/historical-weather-api>). This suggests that the projection of Fig. 3 is undoubtedly underestimating the actual risk of heat stress for wild doves.

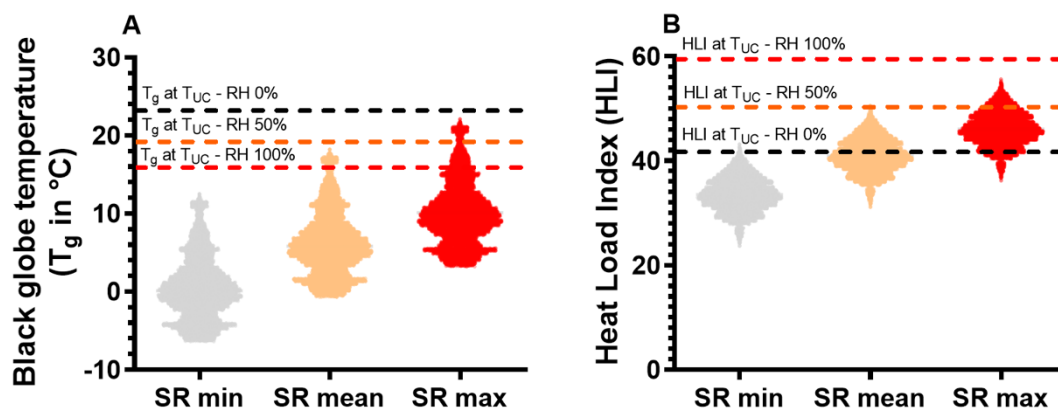


Fig. 3: Heat load indexes (A: black globe temperature T_g ; B: heat load index HLI) calculated for wild doves from [3] under three solar radiation (SR) conditions (*i.e.* minimum, mean, maximum) according to the heat load experienced by doves in a respirometry chamber at their upper critical temperature (T_{UC}) [1] under three relative humidity conditions (*i.e.* $RH = 0\%$, 50% or 100%).

Thirdly, while Beaman et al. (2024) acknowledge that their data is restricted to inactive birds, they do not seem to consider much further in their predictions the fact that wild doves are spending approximately half of their time engaged in energy expensive and heat-producing activities that are likely to increase the risks of heat stress. The *heat dissipation limit theory* posits that the capacity to dissipate metabolic heat limits the maximum rates of energy

expenditure in endotherms, and that managing hyperthermia risk constrains the biology and ecology of wild endotherms [10,11]. Therefore, I believe that caution should be taken when extrapolating the effect of warm environmental conditions on wild animals if measurements are limited to individuals at rest. If dovekeys have to rest at sea or in the colony to cool down as suggested in the discussion of Beaman et al. [1], they are likely to suffer from warm environmental conditions in terms of foraging performance, which should be costly for reproductive success and/or body maintenance, and thus impact population dynamics in response to climate change.

Fourthly, based on the relatively modest increase in water requirement projected for dovekeys under climate warming (1.1 to 2.0 the actual needs), Beaman et al. [1] suggest that water availability should not be much of an issue for wild dovekeys, especially since they have easy access to seawater and possess salt glands. This statement has to be nuanced from my perspective for three main reasons. First, the water requirements are likely under-estimated since T_a only (and not heat load) has been used in their projections. Accordingly, the dovekeys measured by Beaman et al. [1] at their T_{UC} (*i.e.* way before the inflexion point for evaporative water loss) evaporate 15.6g of H_2O /day or 16% of their total body water [4]. Those data are likely underestimating massively the water loss of wild dovekeys, since water influx in free-living individuals has been estimated to *ca.* 137 ± 31.7 g/day [4]. Second, drinking seawater is not cost-free since the metabolic cost of drinking seawater has for instance been estimated to *ca.* 15-20% of basal metabolic rate in wild shorebirds [12]. Therefore, the increase in seawater consumption will lead to a metabolic cost that has not been integrated in the projection of Beaman et al. [1]. Third, leaving the nest for reaching seawater is not cost-free if it implies letting the egg or chick unattended, and thus exposed to adverse environmental conditions and predators. It has for instance been shown that it increases the risk of reproductive failure in wild guillemots [6]. Therefore, I believe that more work is needed on the water balance of wild dovekeys (and seabirds in general) in the context of climate change, and that the easy access to seawater mentioned by Beaman et al. [1] is likely too simplistic and far from being cost-free.

While measurements under experimental conditions at various T_a are undoubtedly extremely valuable to evaluate the thermal physiology at the species or individual level, my opinion is that extreme caution should be taken when using such data to predict the response of wild animals to climate change or extreme climatic events such as heat waves. I suggest that

to really grasp the perils faced by wild animal in response to climate change and extreme climatic events, *in-situ* measurement of the biological responses to variations in heat load (encompassing not only T_a but also solar radiations, wind speed and humidity measured at the micro-climatic level [5]) should be assessed, using behavioural signs of heat stress (*e.g.* panting as in [6]), time-activity budgets (*e.g.* to gather information on reduced foraging activity or parental care under warm conditions [13]) and physiological measurements (*e.g.* body temperature, activity and energetics through heart rate (*e.g.* as in [14]), as well as dehydration status (*e.g.* as in [15])). Such approaches carry their own limitations, such as potential negative impacts of bio-logging devices on birds [16] and difficult calibration of heart rate against metabolic rate [17]. They should ideally be complemented by studying the impact of warm environmental conditions on reproductive success (see for instance the impact of warm conditions on reproductive failure in [6]), and on the potential delayed effects for survival prospects (see for instance the negative effects of warm/dry environmental conditions on telomere length [18], a biomarker of ageing and survival prospects).

Wild animals live in a complex environment and demonstrate complex responses to environmental variations. While one necessarily needs to simplify the biological complexity to model its future trends, an over-simplification of the complexity of thermal biology might lead to misleading conclusions when forecasting the effects of climate change on wild animal populations.

Data availability

The datasets used in this manuscript are available on *FigShare*:
10.6084/m9.figshare.26509378

Author's contribution

Study design: AS. Funding acquisition: AS. Data collection and analysis: AS. Writing: AS.

Ethical note

Only data previously collected by [4], [3] and [1] were used for this commentary

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References

1. Beaman JE, White CR, Clairbaux M, Perret S, Fort J, Grémillet D. 2024 Cold adaptation does not handicap warm tolerance in the most abundant Arctic seabird. *Proc. R. Soc. B* **291**, 20231887. (doi:10.1098/rspb.2023.1887)
2. Short JC, Freeman MT, McKechnie AE. 2022 Respirometry protocols for avian thermoregulation at high air temperatures: stepped and steady-state profiles yield similar results. *J. Exp. Biol.* **225**. (doi:10.1242/jeb.244166)
3. Grunst ML, Grunst AS, Grémillet D, Kato A, Gentès S, Fort J. 2023 Keystone seabird may face thermoregulatory challenges in a warming Arctic. *Sci. Rep.* **13**, 16733. (doi:10.1038/s41598-023-43650-5)
4. Gabrielsen GW, Taylor JRE, Konarzewski M, Mehlum F. 1991 Field and Laboratory Metabolism and Thermoregulation in Dovekies (*Alle alle*). *Auk* **108**, 71–78. (doi:10.1093/auk/108.1.71)
5. Mitchell D, Maloney SK, Snelling EP, Fonsêca V de FC, Fuller A. 2024 Measurement of microclimates in a warming world: problems and solutions. *J. Exp. Biol.* **227**. (doi:10.1242/jeb.246481)
6. Olin A, Dück L, Berglund P, Karlsson E, Bohm M, Olsson O, Hentati-Sundberg J. 2023 Breeding failures and reduced nest attendance in response to heat stress in a high-latitude seabird. *Mar Ecol Prog Ser* (doi:10.3354/meps14244)
7. Hajizadeh R, Dehghan SF, Golbabaie F, Jafari SM, Karajizadeh M. 2017 Offering a model for estimating black globe temperature according to meteorological measurements. *Meteorol. Appl.* **24**, 303–307. (doi:10.1002/met.1631)
8. Gaughan JB, Mader TL, Gebremedhin KG. 2011 Environmental Physiology of Livestock. , 243–265. (doi:10.1002/9781119949091.ch14)
9. Dyk M van, Noakes MJ, McKechnie AE. 2019 Interactions between humidity and evaporative heat dissipation in a passerine bird. *J. Comp. Physiol. B* **189**, 299–308. (doi:10.1007/s00360-019-01210-2)
10. Speakman JR, Król E. 2010 Maximal heat dissipation capacity and hyperthermia risk: neglected key factors in the ecology of endotherms. *Journal of Animal Ecology* **183**, no-no. (doi:10.1111/j.1365-2656.2010.01689.x)

11. Grémillet D, Meslin L, Lescroël A. 2012 Heat dissipation limit theory and the evolution of avian functional traits in a warming world. *Funct. Ecol.* **26**, 1001–1006. (doi:10.1111/j.1365-2435.2012.02048.x)
12. Gutiérrez JS, Masero JA, Abad-Gómez JM, Villegas A, Sánchez-Guzmán JM. 2011 Understanding the energetic costs of living in saline environments: effects of salinity on basal metabolic rate, body mass and daily energy consumption of a long-distance migratory shorebird. *J. Exp. Biol.* **214**, 829–835. (doi:10.1242/jeb.048223)
13. Ven TMFN van de, McKechnie AE, Cunningham SJ. 2019 The costs of keeping cool: behavioural trade-offs between foraging and thermoregulation are associated with significant mass losses in an arid-zone bird. *Oecologia* **191**, 205–215. (doi:10.1007/s00442-019-04486-x)
14. Menzies AK, Studd EK, Majchrzak YN, Peers MJL, Boutin S, Dantzer B, Lane JE, McAdam AG, Humphries MM. 2020 Body temperature, heart rate, and activity patterns of two boreal homeotherms in winter: Homeostasis, allostasis, and ecological coexistence. *Funct. Ecol.* **34**, 2292–2301. (doi:10.1111/1365-2435.13640)
15. Salaberria C, Celis P, Rull IL, Gil D. 2014 Effects of temperature and nest heat exposure on nestling growth, dehydration and survival in a Mediterranean hole-nesting passerine. *Ibis* **156**, 265–275. (doi:10.1111/ibi.12121)
16. Portugal SJ, White CR. 2022 Externally attached biologgers cause compensatory body mass loss in birds. *Methods Ecol. Evol.* **13**, 294–302. (doi:10.1111/2041-210x.13754)
17. White CR, Grémillet D, Green JA, Martin GR, Butler PJ. 2011 Metabolic rate throughout the annual cycle reveals the demands of an Arctic existence in Great Cormorants. *Ecology* **92**, 475–486. (doi:10.1890/09-1951.1)
18. Eastwood JR, Connallon T, Delhey K, Hall ML, Teunissen N, Kingma SA, Porte AML, Verhulst S, Peters A. 2022 Hot and dry conditions predict shorter nestling telomeres in an endangered songbird: Implications for population persistence. *Proc National Acad Sci* **119**, e2122944119. (doi:10.1073/pnas.2122944119)