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- Spatial occurrence records and distributions of tropical Asian butterflies *Eugene Yu Hin Yau¹, *Emily E. Jones¹, Toby Pak Nok Tsang^{1,2}, Shuang Xing^{1,3}, Richard T. Corlett⁴, Patrick Roehrdanz⁵, David J. Lohman⁶, Adam Kai Chi Lee¹, Catherine Wai Ching Hai¹, Shawan Chowdhury^{7,8,9,10}, Jane K. Hill¹¹, Jade A. T. Badon¹², Cheong Weei Gan¹³, Yves Basset¹⁴, I-Ching Chen¹⁵, Suzan Benedick¹⁶, Anuj Jain^{13,17}, Tiffany L.T. Ki^{11,18}, Krushnamegh Kunte¹⁹, Akihiro Nakamura²⁰, Lien Van Vu²¹, Sarah A. Scriven¹¹, Alice C. Hughes¹, Timothy C. Bonebrake^{1#} *authors contributed equally, #corresponding author ¹School of Biological Sciences, The University of Hong Kong, Pokfulam, Hong Kong SAR, CN ²The University of Toronto Scarborough, 1265 Military Trail, Scarborough, ON M1C 1A4, CA ³School of Ecology, Shenzhen Campus of Sun Yat-sen University; Shenzhen 518107, China. ⁴Center for Integrative Conservation and Yunnan Key Laboratory for the Conservation of Tropical Rainforests and Asian Elephants, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences; Honorary Research Associate, Royal Botanic Gardens, Kew. ⁵Betty and Gordon Moore Center for Science, Conservation International, Arlington, VA, USA ⁶City College of New York, City University of New York, 160 Convent Avenue New York, NY 10031, USA; PhD Program in Biology, City University of New York, 365 Fifth Avenue, New York, NY 10016; Zoology Division, National Museum of Natural History, Rizal Park, Manila 1000, Philippines ⁷Institute of Biodiversity, Friedrich Schiller University Jena, Dornburger Straße 159, 07743 Jena, Germany ⁸Department of Biodiversity and People, Helmholtz Centre for Environmental Research – UFZ, Permoserstraße 15, 04318 Leipzig, Germany ⁹German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig, Puschstraße 4, 04103 Leipzig, Germany ¹⁰Czech University of Life Sciences Prague, Faculty of Environmental Sciences, Prague, Czech Republic ¹¹Leverhulme Centre for Anthropocene Biodiversity, Department of Biology, University of York, York, YO10 5DD, UK ¹²Animal Biology Division, Institute of Biological Sciences, University of the Philippines Los Baños, Laguna 4031, Philippines ¹³Nature Society Singapore, 510 Geylang Road, 389466, Singapore ¹⁴Smithsonian Tropical Research Institute, Apartado 0843-03092, Balboa, Ancon, Panama ¹⁵Department of Life Sciences, National Cheng Kung University, Taiwan ¹⁶Faculty of Sustainable Agriculture, Universiti Malaysia Sabah, Locked Bag No. 3, 90509, Sandakan, Sabah, Malaysia ¹⁷bioSEA Pte Ltd., 68 Chestnut Avenue, 679521, Singapore ¹⁸Science Department, Natural History Museum, London SW7 5BD, United Kingdom ¹⁹National Centre for Biological Sciences (NCBS), Tata Institute of Fundamental Research (TIFR), GKVK Campus, Bellary Road, Bengaluru 560065, India ²⁰CAS Key Laboratory of Tropical Forest Ecology, Xishuangbanna Tropical Botanical
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- 55

56 Abstract

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58 Insect biogeography is poorly documented globally, particularly in the tropics. Recent intensive research in tropical Asia, combined with increasingly available records from citizen 59 science, provides an opportunity to map the distributions of tropical Asian butterflies. We 60 compiled a dataset of 724,247 occurrences of 3,591 tropical Asian butterfly species by 61 aggregating records from GBIF (651,285 records), published literature (21,271), published 62 databases (37,695), and unpublished data (13,993). Here, we present this dataset and single-63 species distribution maps of 1,520 species. Using these maps, along with records of the 2,071 64 remaining species, we identified areas of limited sampling (e.g., the Philippines, Myanmar, 65 and New Guinea) and predicted areas of high diversity (Peninsular Malaysia and Borneo). 66 67 This dataset can be leveraged for a range of studies on Asian and tropical butterflies, including 1) species biogeography, 2) sampling prioritization to fill gaps, 3) biodiversity 68 hotspot mapping, and 4) conservation evaluation and planning. We encourage the continued 69 development of this dataset and the associated code as a tool for the conservation of tropical 70 Asian insects. 71

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74 Background & Summary

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Tropical Asia, home to multiple major global biodiversity hotspots, harbors a rich assemblage
of highly range-restricted endemic species¹. Unfortunately, reliable distribution data for many
species in this region are scarce². One prominent challenge for invertebrate conservation,
known as the Wallacean shortfall, stems from our inadequate knowledge of species
distributions³. Insufficient information on species distributions impedes the identification of
vulnerable species and the efficient allocation of conservation resources across regions and
species^{3,4}.

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84 While recent global studies of butterfly biogeography have incorporated data from tropical Asia^{5,6}, they have primarily relied on coarse, country-level data to examine biogeographic 85 patterns⁵⁻⁷. The distribution information summarized based on those data is largely influenced 86 87 by political boundaries rather than relevant ecological areas and is less ideal for identifying important conservation/vulnerable areas, which requires fine-scale, biogeographic data with 88 low bias⁸. There have also been attempts to map spatial phylogenetic diversity using range 89 maps⁹, but the quality of such spatial analyses is highly dependent on the range maps used, 90 which often fail to capture distribution patterns at local scales, thereby limiting the resolution 91 92 of the spatial pattern of interest. Although fine-scale geographic distributions of several Asian butterfly groups have been mapped (e.g., *Elymnias* in Wei et al.¹⁰; *Papilio* in Condamine et 93 al.¹¹; *Polyura* in Toussaint et al.¹²; range-restricted butterflies in Scriven et al.¹³), to date, no 94 unified, fine-scale distribution dataset has been produced for the entire region – despite the 95 importance of such a tool for examining patterns of diversity within this highly biodiverse 96 region^{1,6}. Existing locality data might not be readily accessible and frequently require 97 aggregation and standardization. Fine-grained information on species distributions is an 98 99 essential first step for understanding insect biodiversity patterns and conservation needs.

- 101 The creation of regional datasets of species distributions is aided by the recent development
- 102 of large, open-source biodiversity data platforms such as the Global Biodiversity Information
- 103 Facility (hereafter, GBIF), an online database that organizes crowd-sourced data from citizen
- science platforms, scientific literature, and specimen collections¹⁴. These data, however, often
- include large spatial biases due to uneven sampling and data mobilization efforts among regions^{14,15}. Even if available, much of the fine-scale biogeographic data that could be
- regions^{14,15}. Even if available, much of the fine-scale biogeographic data that could be
 employed to reduce these biases remains buried in literature and regional databases, requiring
- 108 concerted efforts to make it analysis-ready⁷. Without unified and standardized datasets, it is
- difficult to test macroecological and macroevolutionary questions¹⁶, produce high-quality
- species distribution models¹⁷, and identify effective conservation targets^{5,6,8,18}.
- 111
- 112 The process of mapping species distributions can be accomplished either through data-driven
- 113 modeling or by relying on expert knowledge. Range maps (expert range maps) solely based 113
- on expert knowledge tend to overestimate active areas of species at the local scale^{15,17,19}. In
 addition, the quality of their source data, hence the uncertainty of the analysis, is often
- addition, the quality of their source data, hence the uncertainty of the analysis, is often
 unknown¹⁶. The dependency of range maps on expert knowledge means this method is
- available to a small subset of well-studied species⁷. In contrast, data-driven distribution maps
- 117 available to a small subset of well-studied species . In contrast, data-driven distribution map 118 offer greater transparency and reproducibility^{18,20}. Modern modeling techniques allow the
- interpolation of potential distributions into areas for which primary data collection may not
- be possible, enabling the production of more detailed and reliable distribution maps 3,21 .
- However, major data gaps exist for occurrence records of most taxa 16,22 , particularly
- invertebrates, and the non-random distribution of these gaps necessitates careful treatment within models²³.
- 123 124

Species distribution maps facilitate the identification of species ranges and hotspots of
 diversity. This provides valuable insights for local conservation planning/prioritization^{24,25}

- and policy-making, paving the way for future investigations into butterfly biogeography⁵ and
- 128 phylogeographic patterns²⁴. Specifically, species distribution maps can guide the allocation of
- 129 conservation resources, inform the strategic design of protected areas in high-
- 130 suitability/biologically diverse areas, and identify low-suitability areas in need of
- 131 management 25,26 , enabling effective conservation interventions. In conjunction with SDMs,
- 132 occurrence datasets can help inform species reintroduction programs by identifying
- potentially suitable areas^{25,27} and optimal source populations²⁸, and expedite IUCN Red List
- assessment, which has poor species coverage in Asia. Additionally, applications of SDMs 24,27,29
- include the modeling of species and community-level responses to climate change^{24,27,29} and the assessment of extinction risks³⁰.
- 137

The need for species conservation is particularly acute in tropical Asia, defined broadly here to include South Asia and Southeast Asia (see Fig. 1). The area is home to over 20,000 islands, many of which were repeatedly connected and separated from adjacent landmasses during drastic sea-level fluctuations over the past 4 Mya³¹. This dynamic past led to the

- evolution of numerous species endemic to single islands or island groups, and as such this
- region hosts some of the world's greatest biodiversity an estimated 15-25% of all well-
- studied terrestrial taxa and a large proportion of undescribed taxa^{32,33}. This highly biodiverse
- region is also one of the globe's most biologically threatened: it is estimated that 42% of Southeast Asia's biodiversity may be lost by 2100 as three superior of its arity and for the second second
- Southeast Asia's biodiversity may be lost by 2100 as three quarters of its primary forests are
 lost to agriculture, urbanization, and mineral extraction^{32,34,35}.
- 148

- We present a comprehensive dataset of tropical Asian butterflies, with more than half of the 149
- records possessing high spatial accuracy (uncertainty < 10 km). This fills a major sampling 150
- gap, given that Asia is poorly represented in global biodiversity data repositories^{15,22,36}. 151 improved datasets are urgently needed to enable effective monitoring and management of
- 152 biodiversity across the region. Leveraging the data along with tailored species distribution 153
- models (SDMs), we generate data-driven distribution maps at a resolution of 10 km x 10 km. 154
- These maps enhance a fundamental understanding of butterfly macroecological patterns in 155
- tropical Asia. Each butterfly species' distribution was individually modeled and, together 156
- with buffered occurrence points of unmodelled species, employed to assess regional patterns 157
- of species diversity. Combined with species distribution models, our aggregated data 158
- advances knowledge of butterfly macroecology and facilitates evidence-based decision-159 making for butterfly conservation in tropical Asia.
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- 161 162

Methods 163

- 164
- 165 Occurrence data
- 166 We manually extracted GBIF records for tropical Asian Papilionoidea (Lepidoptera:
- Nymphalidae, Papilionidae, Lycaenidae, Pieridae, Hesperiidae, Riodinidae; -11.426 35.64 167
- N, 67.588 174.990 E) for the years 1970-present on 15 April 2024 (Derived dataset 168 GBIF.org³⁷). The geographical extent of the study area was selected to encompass northern 169
- temperate Asia to secure sufficient data to capture the full niche breadth of all species in the 170
- subsequent SDMs. We included presence records derived from human observation, preserved 171
- specimens, material samples, or literature, provided they had associated coordinates. We 172 omitted all records with >100,000 m coordinate uncertainty, so-called "fuzzy" taxon matches, 173
- and records for which the scientific name was missing or incomplete unless nomenclature 174
- could be extracted using a BOLD identifier (boldsystems.org/). This resulted in a final 175
- 176 number of GBIF records equalling 651,285.
- 177
- Roughly 73% (472,714) of these records are 'Research-grade' observations from iNaturalist. 178 Information on how this designation is made is available at GBIF.org. The accuracy of 179 opportunistically collected data from crowd-sourced platforms like GBIF is often diminished 180 due to misidentifications, taxonomic, spatial, and temporal biases, as well as uneven 181 taxonomic validation due to lack of standardized reference data^{14,38-41}. Given these potential 182 issues, and to fill geographic gaps, we supplemented these GBIF data with expert data 183 (coauthor datasets, published literature) and harmonized binomials to a single expert dataset 184 (Lamas, 2015. Catalogue of the butterflies (Papilionoidea), available from the author.; see 185 below). 186
- 187
- We extracted data from the B2D2 Database of Butterflies for Borneo provided by JKH/the 188 Darwin Initiative (n = 19,417), a dataset for Bangladesh provided by SC (Chowdhury et al.⁴²; 189 n = 18,278), and unpublished datasets from coauthors AN, DJL, LVV, TK, and YB (n = 190 13,993). For geographic regions with relatively few records (e.g., China, Myanmar, Thailand) 191 and for species with < 10 records, we conducted targeted searches of post-1970 published 192 literature on Google Scholar in English and Chinese (simplified and traditional) (genus OR 193 genus + species + country name), producing an additional 21,271 records. Although some 194 195 publications lacked collection dates for records (e.g., checklists), we assume that the inclusion of species in recent publications is indicative of species' current localities. 196 197

- For all records in published sources, we extracted coordinates, locality name, locality type 198 (e.g., exact coordinates, city, national park, island, or province), country, and year of record 199 (where available). If exact coordinates were not provided by the source, we used Google 200 Earth Pro (v7.3.6.9345) to estimate the locality centroid for any record provided at the 201 province level or below (e.g., national park or city). For records from islands ≤ 100 km at the 202 widest dimension (e.g., localities within the Philippines and Indonesia), we estimated the 203 island or archipelago centroid. If a range of coordinates was provided (e.g., records from The 204 Butterflies of Vietnam), we selected a point within the range. Data sources for all records are 205 provided in the reference column in Occurrence Records of Tropical Asian Butterflies: 1970-206 2024 (https://doi.org/10.6084/m9.figshare.25037645). 207
- 208

Final binomial harmonization, validation, and authority assignment were conducted by DJL
using a taxonomic reference prepared by Gerardo Lamas (Lamas, 2015). Family names were
aligned by hand to GBIF.

212

The resulting database consists of 724,247 occurrence records for 3,591 species from 546

- 214 genera. These records represent approximately 20% of all described butterfly species 215 $abch = 10^{-43} \frac{44}{17} (17.280, 17.500)$ and $back = 10^{-20} \frac{20225}{100}$. Becaude of Neural all described butterfly species
- globally^{43,44} (17,280-17,500 spp.; but see Pinkert et al., 2022^5). Records of Nymphalidae
- (313,899; 1,324 spp.) comprise 43% of the dataset, followed by Lycaenidae (968 spp;
 147,277 records), Papilionidae (264 spp.; 101,500 records), Pieridae (405 spp.; 97,120
- records), Hesperiidae (611 spp.; 60,460 records), and Riodinidae (22 spp.; 3,991 records).
- 219

219 220 Of the 3,591 species in the database, 1,580 (< 31%) are represented by ≥ 10 records within

- the extent of 36° N to 10° S and 69° E to 161.6° E that are >10 km apart (see details on distribution modeling below). Most occurrence records are concentrated in a limited number
- of regions, for example, India (28.34% of all data), Taiwan (13.75% of all data), Singapore
- (8.92% of all data), Hong Kong (8.30% of all data), and Malaysia (7.16% of all data) (Fig. 1).
- Equatorial regions together with southern China are relatively underrepresented in our
- dataset. As much of the data is derived from GBIF, which contains a large proportion of
- citizen science data, we observed a clustering of our data in areas of high population and a
- 228 general lack of data in more inaccessible regions.
- 229



- **Figure 1.** Distribution of GBIF and other occurrence records in our study area. Sampling
- intensity was estimated by running kernel density on the coordinates of all available
- occurrence data of every species. Regions of Asian landmasses based on the ecoregions and
- biogeographic realms as revised by Dinerstein et al.⁴⁵, as well as Wallace's Line, Huxley's
- Line, and Weber's Line.
- 237

238 SDM methods and results

- 239 Five algorithms, Generalized Linear Model (GLM), Maximum Entropy (MaxEnt),
- 240 Multivariate Adaptive Regression Splines (MARS), Classification Tree Analysis (CTA), and
- eXtreme Gradient Boosting (XGBOOST) were selected to create an ensemble model for each
- butterfly species, using the ensemble platform "biomod2"⁴⁶ in R. We ensured that the
- 243 underlying mechanism of our selection of algorithms was diverse and relatively balanced
- between the main categories of algorithms. We used 13 predictor variables for selection by
- individual models. All modeling was conducted at 10 km x 10 km resolution.
- 246

The Generalized Linear Model (GLM) is a regression-based algorithm widely used in 247 SDMs⁴⁷. They are not as flexible when fitting complex response curve shapes, but this also 248 means that GLMs are less vulnerable to overfitting⁴⁷. Maximum Entropy (MaxEnt) in our 249 study was based on the "maxnet" R package⁴⁸, which uses penalized maximum likelihood for 250 model fitting. MaxEnt is one of the computationally less expensive algorithms that perform 251 well, making it a popular SDM algorithm⁴⁹. MaxEnt is more capable of fitting complicated, 252 non-linear response curves, enabling users to model more complex relationships by using 253 254 progressively complex statistics based on the number of samples available⁵⁰. The classification tree analysis (CTA) used by our SDM is based on the "rpart" R package⁵¹. The 255 CTA algorithm recursively splits one group of data into two subgroups using one of the 256 257 predictor variables given; therefore, the final model can be visualized as binary decision trees⁵¹. Finally, eXtreme Gradient Boosting (XGBoost) is one of the more computationally 258 efficient gradient boosting algorithms implemented in R by the "xgboost" package⁵². 259 Boosting algorithms feature an ensemble of weak models, each trained to minimize the errors 260 of the previous models^{47,53}. 261

262

For the species distribution models, we used 13 predictor variables, which comprised 8 263 Bioclim variables extracted from WorldClim⁵⁴, three soil variables extracted from SoilGrids⁵⁵ 264 through ISRIC (International Soil Reference and Information Centre)⁵⁶, and 2 vegetation 265 variables derived from satellite data. The Bioclim variables employed included annual mean 266 267 temperature (Bio 1), temperature seasonality (Bio 4), maximum temperature of warmest month (Bio 5), minimum temperature of coldest month (Bio 6), annual precipitation (Bio 12), 268 precipitation of wettest month (Bio 13), precipitation of driest month (Bio 14), precipitation 269 270 seasonality (Bio 15). The soil variables at a depth of 5-15 cm were used, including soil pH (phh2o), soil organic carbon content in the fine earth fraction (SOC), and total nitrogen 271 (nitrogen). Nitrogen is generally recognized as one of the main limiting elements for plant 272 growth⁵⁷, while soil organic carbon indicates soil quality⁵⁸. In addition, soil pH exerts 273 considerable influence on soil biogeochemical processes, ultimately impacting plant 274 growth⁵⁹. The selection of variables for our models was guided by expert knowledge to 275 276 reflect/cover the key limitations and resources relevant to both butterflies and their host plants. Knowledge of the study region and biology/ecophysiology of the species being 277 modeled allows the identification of the most ecologically relevant variables; therefore, it is 278 the preferred approach for variable selection 47,49,60,61. 279

280

The vegetation variables used were the Normalized Difference Vegetation Index (NDVI) and 281 Canopy Height. NDVI was calculated from the USGS Landsat 5 (Level 2, Collection 2, Tier 282 1, 1985 – 1999) and USGS Landsat 7 (Level 2, Collection 2, Tier 1, 2000 – 2020) datasets, 283 with a customized script to filter satellite images by cloud cover (retaining images with 15% 284 or less cloud cover over land) and to obtain the mean NDVI value. Canopy Height data was 285 retrieved from the ETH Global Sentinel-2 10 m Canopy Height dataset⁶². These vegetation 286 cover variables were directly used to model the land cover/habitat available to butterflies. 287 Mean NDVI provided information on the general greenness of an area, while Canopy Height 288 data offered structural details on vegetation to better identify different types of habitats. 289 290 Together, these variables indicate resource availability and, to some extent, habitat structure. To address potential issues associated with negative values in NDVI data, an alternative 291 variable, Corrected NDVI, which contains no negative values, was also examined. The 292 293 Corrected NDVI is derived from the equation Corrected NDVI = NDVI +1. However, the 294 SDMs using Corrected NDVI produced identical results to those using standard NDVI data, indicating that our models were unaffected by negative NDVI values. 295

296

297 The resolution of all environmental variables was set to 10 km x 10 km by averaging the values from contributing grid cells. This resolution was chosen as a result of balancing the 298 spatial accuracy of available data and computational capabilities. Our data comprises 440,731 299 300 records with coordinate uncertainty data, while an additional 283,523 records that do not have coordinate uncertainty data. Among the records with known coordinate uncertainty, 301 73,372 (18.31% of records with uncertainty data) had uncertainties ranging from 1-10 km, 302 and 39,302 (9.81% of records with uncertainty data) had uncertainties exceeding 10 km, thus 303 10 km seemed a reasonable compromise to reflect this. For the construction of SDMs, the 304 map of the study area and predicting variables were formatted to share the same extent, 305 306 resolution, and projection. We excluded entries with invalid species names (e.g., "NA" and "not present") or outside of our study area and those recorded before 1970. Data entries 307 published after 1970 but without date records were kept, assuming that their publication 308 infers their validity at the time of publication. Next, the map of tropical Asia and all 309 explanatory variable rasters were all projected to equal area projection EPSG:6933 and 310 cropped to the extent of 36° N to 10° S and 69° E to 161.6° E to fully cover the study region. 311 Our final cleaned database included 721,060 global records. 312

313

We used a function to further prepare the input files required by biomod2 and to generate SDMs individually for each species. Occurrence data of a species was first extracted from our butterfly occurrence dataset and used to produce a raster of resolution of 10 km x 10 km. A total of *n* cells in the raster were assigned a value of "1" to represent at least one occurrence record present in that cell, while cells with no record were assigned "n/a" instead of "0" since no true absence data is available.

320

Only species with n > 10 were modeled. It has been shown that SDMs based on ten 321 occurrence points can reach 90% of the maximum possible accuracy⁶³, while recent studies 322 suggest a minimum requirement of 3 to 13 occurrence points in virtual simulations and 14 to 323 25 occurrence points in real-world conditions to infer accurate SDMs⁶⁴. Therefore, n=10 was 324 chosen as the lower limit of sample size for constructing SDMs to maximize the number of 325 species modeled while maintaining a reasonably high predictive accuracy⁶³. A total of 1,580 326 species met this qualification, whereas 1951 species had fewer records. For each species, 327 328 occurrence records were split into three sets: 10% of the data was first reserved for model evaluation, and another 10% was then partitioned for model validation, leaving the remaining 329

80% of data for model calibration. The partitioning of model validation data was repeated 5
times to generate five different combinations of calibration and validation occurrence data.

332

Before SDM construction, pseudo-absence records were generated. Despite our efforts to fill 333 the spatial data gaps, the sampling effort of our dataset is still spatially biased toward highly 334 populated areas and roads due to the overwhelming number of records from GBIF and 335 iNaturalist in our dataset (more than 80%). As part of our effort to account for biases in our 336 data, we integrated the spatial bias of our dataset into the generation of pseudo-absence 337 records, assuming that all species were sampled in areas with at least one occurrence record 338 of any species. To capture such spatial bias, we created a raster layer of the spatial sampling 339 effort for all species across our study area (shown as sampling intensity in Fig. 1), which is 340 equivalent to the bias layer commonly used in the MaxEnt program. This was done by 341 342 pooling occurrence data of all species used in our models and summarising them in a raster, then performing two-dimensional kernel density estimation (kde2d) using the R package 343 "MASS"⁶⁵ with the default settings. We excluded cells with occurrence records and sampled 344 the remaining study area for pseudo-absence records based on the bias layer, giving more 345 weight to well-sampled areas, as suggested by Phillips et al.⁶⁶ and Ferrier et al.⁶⁷. Following 346 the recommendation of Barbet-Massin et al.⁶⁸, for calibration, validation, and evaluation data, 347 we produced five sets of pseudo-absence data for each species, maintaining a 1:1 ratio 348 349 between the number of pseudo-absence points and occurrence points in each set.

350

Subsequently, we constructed SDMs for each species using five different partitions of calibration and validation occurrence data, five selected algorithms, and five sets of pseudoabsence data. This resulted in a total of 125 SDM models (5 x 5 x 5). Both presence and pseudo-absence records were given equal weight during model construction to ensure a consistent prevalence of 0.5 among all species. We applied a generalized setting for all butterfly species for consistency across species, with adjustments made only to the learning

rate and the number of decision trees for the XGBoost algorithm to address overfitting. Other
 model tuning options were retained at their default.

359

We generated binary outputs by maximizing True Skill Statistics (TSS), a widely used 360 threshold-dependent index of model fit. Ensemble modeling was selected over single best 361 models for its superior performance in rare species⁶⁹, and its robustness to uncertainties in 362 individual models by capturing the central tendency among models^{47,70,71}. We constructed an 363 ensemble model using all single models with TSS values greater than 0.7, ensuring that only 364 "substantial" models were included⁷². 1,520 species out of the 1,580 modeled species 365 obtained one or more single models meeting such criteria, allowing the further construction 366 of ensemble models. The ensemble model was generated using the mean algorithm⁷¹, where 367 all candidate models' probabilistic predictions were averaged without weighting. Finally, we 368 projected the ensemble model to the current environment using the same variables when 369 constructing the SDMs. 370

371

Ensemble models were evaluated using two metrics: TSS and Boyce index. TSS and Kappa are two of the most popular SDM threshold-dependent evaluation metrics. TSS was chosen over Kappa due to the inherent dependency of Kappa on species prevalence⁷³. Since we are modeling thousands of species with differing degrees of rarity and prevalence, TSS is more appropriate for model comparison between species. TSS varies from +1 to -1, in which +1 indicates perfect agreement with evaluation data, while a TSS value close to or less than 0 indicates model performance comparable to a random model⁷³.

- Following the suggestions of Hernandez et al.⁷⁴ and Breiner et al.⁶⁹ to use multiple evaluation 380 measures when using presence-only data, we also calculated the Boyce index for all models 381 built to supplement TSS. The Boyce index is capable of providing an accurate and reliable 382 measure of model performance for models based on presence-only data⁷⁵, which is the key 383 reason for its use in our study. Another reason for the use of the Boyce index is its lower 384 sensitivity (correlation) to species prevalence relative to other metrics, including CVI. 385 MaxKappa, and adjusted $D2^{75}$, while AUC and TSS also have a negative correlation with 386 prevalence⁷³. AUC was also found to produce inflated estimates of model quality when the 387 modeled species is rare⁷⁶. Boyce index ranges from +1 to -1, in which +1 indicates the model 388 is of the highest quality and perfectly predicts evaluation data, while -1 indicates counter-389 prediction of evaluation data⁷⁵. Boyce index with a value close to 0 indicates the model 390 performs no better than a random model⁷⁵. 391
- 392

To factor biogeography into predictions and correct for biogeographic overprediction 393 generated by our SDMs (and account for differences between fundamental and realised 394 niches), we restrained the sampling of pseudo-absence records and distribution maps 395 396 produced by our models to regions that hosted more than 1% of species points (as such regions fall within species biogeographic ranges). By incorporating biogeography into model 397 predictions, we aimed to reflect the impact of oceans as dispersal barriers in the SDM outputs 398 399 to give a more realistic estimate of species' distribution and reduce false positive predictions. We first divided the landmasses of tropical Asia into 11 regions (Fig. 1) based on the 400 ecoregions and biogeographic realms as revised by Dinerstein et al.⁴⁵, as well as Wallace's 401 402 Line, Huxley's Line, and Weber's Line. For each species, we identified regions that included at least 1% of the species occurrence records, considering them to be active regions. We then 403 cropped the SDM-predicted distribution maps to include only the active regions specific to 404 405 each species. These cropped distribution maps were stacked together to generate an alpha diversity map, which illustrates the number of species present in each 10 km x 10 km cell 406 across tropical Asia. The stacked SDM predictions highlighted a number of locations with 407 relatively high diversity, exceeding 600 species in some locations (Fig. 2). 408 409





411 **Figure 2.** Projected distribution of butterfly diversity based on our species distribution

- 413
- 414 *Point buffer methods*
- For the 2,011 species (56% of all recorded species in our dataset) excluded from our species distribution modeling outputs either due to insufficient data or low quality of species
- distribution models, we plotted and buffered their occurrence records to infer alpha diversity.
- 418 We first mapped their occurrence records and created 30 km-wide polygons (buffers) around
- these points to enhance clarity. Subsequently, the buffered occurrence points were converted
- 420 into binary raster maps for each species and stacked to generate an additional alpha diversity
- 421 map, representing species with limited occurrence records.
- 422
- The diversity map derived from buffered occurrence points was then stacked with the species
 distribution model (SDM) projections to produce Fig. 3. This figure provides an overview of
 the alpha diversity of all species documented in our dataset. We identified two major
 butterfly diversity hotspots: peninsular Malaysia and the Sabah region of Borneo. We also
 found high levels of diversity predicted in Borneo, Sumatra, coastal Cambodia, southern
- 428 Thailand, the Western Ghats in peninsular India, the Assam region of India, the Cardamom
- 429 mountains in Cambodia, and Vietnam.
- 430
- 431
- 432





Figure 3. Estimated distribution of butterfly diversity based on our species distribution model
 projections and buffered occurrence points (for species not included in our SDM outputs).

- 436
- 437
- 438
- 439 *Software*
- 440 We calculated the SDMs in R^{77} , version 4.1.2. To construct and merge the SDMs into
- ensemble models, we utilized the "biomod2" package, version 4.2-4⁴⁶. The high-performance
- 442 computing cluster HPC2021 at The University of Hong Kong, operating on CentOS 8, was443 employed to run the SDMs.
- 443 ei 444
- 445

446 Data Records

- 447
- 448 All project files are publicly available in a Figshare repository
- 449 (https://doi.org/10.6084/m9.figshare.25037645). Users may access the referenced occurrence
- 450 dataset and metadata as .csv files, the SDM rasters (as file type, e.g., Fig. 4), and links to R
- 451 scripts for SDM construction and distribution map generation were published on GitHub:
- 452 https://github.com/eugeneyau/Tropical-Asian-Butterfly-Distribution/tree/main
- 453 The GBIF-derived dataset is available at GBIF³⁷ (https://doi.org/10.15468/dd.nvw5wr).
- 454 These outputs are licensed under a CC BY 4.0 license.
- 455



456

457 Figure 4. SDM-predicted distribution of *Euripus nyctelius* (Doubleday, 1845) (Nymphalidae:
458 Apaturinae) based on our occurrence dataset.

- 459
- 460

461 **Technical Validation**

463 SDM model evaluation/verification 464 The mean TSS score of all ensemble models is 0.0

The mean TSS score of all ensemble models is 0.922, with a standard deviation of 0.155,
while the Boyce index is 0.766, with a standard deviation of 0.305.

466

467 Both evaluation metrics indicate that the models constructed are of good quality. The mean 468 TSS score of our ensemble models is higher than 0.8, falling into the category of "almost

- 469 perfect" models according to the widely used division suggested by Landis & Koch⁷² (e.g.,
- 470 Capinha et al.⁷⁸; Jones et al.⁷⁹). Since we only included models with TSS values of more than
- 0.7 in our ensemble models, a high mean TSS score among the ensemble models is expected.
- 472 The mean Boyce index of our models is higher than 0.7, which has been considered an
- indicator of good models in other studies (e.g., Rupprecht et al.⁸⁰). Boyce index value of 0.5
- 474 is usually considered a cutoff for acceptable performance⁸¹.
- 475
- 476 *Collaborator Evaluation*
- 477 Our model outputs were also inspected by experts to evaluate their plausibility. Plausibility
- 478 checks form an important part of model validation by making sure the modeling results

- 479 confine to the known range and possible range of the species modeled^{49,82}, serving as a
- 480 supplement to evaluation metrics, which only measure the goodness of fit of models.
- 481

482 Experts (coauthors/collaborators) agreed that our model outputs are generally reasonable and 483 informative. However, it is important to note that some of the sampling biases persisted in the

484 final model outputs despite our efforts to address data gaps by incorporating additional

- datasets. We, therefore, encourage future data contributions to improve the coverage of our
- dataset, especially in the areas with identified data gaps.
- 487

Although the majority of data gaps can be attributed to insufficient sampling effort, the absence of data in the Philippines (and potentially other parts of tropical Asia) is primarily a result of the dominance of Facebook over other platforms like iNaturalist for citizen science data contribution. However, such data on Facebook contains limited information since EXIF data (containing GPS coordinates) of photos are removed when uploaded. Filling the Philippine data gap should be a priority, and mining Facebook data (e.g., Chowdhury et al.¹⁸)

- and other sources would be a good place to start.
- 495

Our modeling results identified the Cardamom Mountains on the Cambodian-Thai border as a butterfly diversity hotspot. During the Pleistocene when sea levels were up to 120 m lower than present, and this area was on the eastern edge of a paleoriver watershed that included the similarly diverse Malay peninsula and extended south to present-day Borneo^{83,84}. The high diversity in this area is likely relictual⁸⁵. Endemism in this area likely contributes to high butterfly diversity, which supports our models' prediction there.

502

Multiple experts pointed out the unexpected diversity differences between different parts of 503 504 Borneo. While our models identified Sabah as a hotspot for butterfly diversity, lower diversity was predicted for other parts of Borneo, such as Sarawak and Kalimantan. This 505 contradicted our expectations, as all these areas possess mountainous regions and endemic 506 species, suggesting similar levels of butterfly diversity. The heart of Borneo, characterized by 507 lower disturbance compared to other parts of the island, was also predicted to host a relatively 508 lower diversity of butterflies by our models. Such a model prediction also contradicts our 509 expectation of higher butterfly diversity in less disturbed areas. This inconsistency between 510 expected and modeled butterfly diversity in Borneo is likely attributed to sampling bias, 511 evident through the alignment of modeled butterfly diversity with political boundaries and 512 sampling intensity (Fig. 1), and the lower modeled diversity in less accessible areas such as 513 514 the heart of Borneo (Fig. 2 and 3). The lack of data in less accessible areas has been discussed by Hughes et al.¹⁵ and Boakes et al.⁸⁶, while this trend is even more obvious in citizen science 515 data¹⁵, which constitutes a considerable proportion of our dataset. 516

517

518 While some of the spatial variations in the sampling effort of our dataset are reflected in the spatial bias of our modeling results, there are several notable discrepancies between the 519 distribution of data and modeled diversity. Fig. 1 illustrates that Japan, Taiwan, and northern 520 Thailand have a relatively high intensity of sampling effort compared to their predicted 521 butterfly diversity in Fig. 2. Conversely, a reversed pattern is evident in Southern Borneo and 522 523 Southern Sumatra, where our data shows low sampling effort but our models predict high butterfly diversity. These patterns demonstrate the robustness of the models to some of the 524 spatial sampling biases present in our data. 525

526

527 To determine the variable importance in our SDMs, we calculated, for each variable, the 528 mean variable importance throughout the ensemble models of all species. Temperature

seasonality (Bio 4) emerged as the most important variable (scoring 0.280 out of 1), followed 529 by the minimum temperature of the coldest month (Bio 6, scoring 0.163 out of 1) and annual 530 mean temperature (Bio 1, scoring 0.140 out of 1). Soil pH (phh2o, scoring 0.107 out of 1), 531 precipitation of driest month (Bio 14, scoring 0.0973 out of 1), and Canopy Height (scoring 532 0.0907 out of 1) also exhibited high importance in the models. The ranking of variable 533 importance in the SDMs conforms to the hierarchical framework of Pearson & Dawson⁸⁷, in 534 which climatic variables exert greater control over species distribution at continental scales, 535 while land cover and soil variables gain influence at more localized scales. In addition, the 536 high importance of temperature variables, particularly temperature seasonality (Bio 4), is 537 consistent with the results of Carvalho et al.⁸⁸, which highlighted the strong impact of 538 temperature, especially temperature seasonality, on butterfly distribution and diversity. 539

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- 541

Usage Notes 542

543

The predictor variables considered in our SDMs, which include the 8 Bioclim variables and 544

the 3 SoilGrids variables, are products of interpolation between available point data^{54,55}. As 545

546 with most data collected without stratified sampling, these point data are likely to be spatially biased. Users should note that our SDMs inherit these biases, as well as uncertainties in the 547

interpolation result. 548

549

By generating more pseudo-absences for SDMs in well-sampled areas with the use of the bias 550 mask, we are essentially augmenting the weighting of extensively surveyed regions in our

551 models, while unsampled habitats may be presumed as suitable. Consequently, the 552

- transferability of our models to unsampled areas is limited, especially when extrapolating in 553
- novel environments not covered by training data⁶⁶ or in areas where biogeographic barriers 554

prevent dispersal. This is also one of the reasons for restraining our model predictions to the 555

- regions where a species is known to occur so that the results are not overly optimistic. Such 556
- an approach to pseudo-absence generation also assumes that the data collection method is 557 consistent throughout the entire dataset⁶⁶, while our dataset is compiled from various sources.
- 558 However, since a majority of our data is derived from a single source (GBIF), we can 559

consider the data collection method consistent in terms of the observation method. To use our 560

- data and models for the prediction of future butterfly distribution under climate change, we 561 562 suggest using the "random" method from the biomod2 package to generate pseudo-absence 563 records.
- 564

Regarding uncertainty in model results, we have limited confidence in the model predictions 565 in the Philippines and New Guinea. The scarcity of occurrence data in these two regions (Fig. 566 1) prevents strong inferences. Additionally, the presence of biogeographic barriers such as 567 Wallace's Line and Huxley's Line restricts the use of occurrence data from other regions to 568 infer butterfly distribution in these specific areas. 569

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Code Availability 572

All code used to conduct synonym harmonization, preprocess environmental variables for 574

575 SDMs, execute SDMs, process SDM outputs, and conduct point buffer analysis can be accessed in our GitHub project repository:

- 576
- https://github.com/eugeneyau/Tropical-Asian-Butterfly-Distribution 577

under the Code directory. All data, including our butterfly occurrence dataset, SDM-predicted
distribution maps, tropical Asian biogeographic regions shapefile, and buffered occurrence
points for species excluded from our species distribution modeling are available from our
FigShare repository (<u>https://doi.org/10.6084/m9.figshare.25037645</u>). Some data for Sulawesi,
provided by TK, have not been included in the data release but are available upon request to

provided by TK, have not been included in the data release but are available upon request to
TK.

584 585

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587

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599 600

601 Author Contributions

602

TCB, TPNT, SX, RTC, and PR conceptualized the study, with funding acquired by TCB, 603 TPNT, and SX. The project methodology was developed by EEJ, EYHY, TPNT, SX, AKCL, 604 RTC, PR, and ACH. TCB and ACH supervised the study, while TCB managed the project 605 administration. The occurrence dataset was compiled by EEJ with assistance from CWCH, 606 and data curation was carried out by EEJ, EYHY, and DJL. Data were contributed by DJL, 607 SC, JKH, YB, TK, LVV, and AN. EYHY conducted the species distribution modeling and 608 subsequent analyses. DJL validated scientific names in the dataset and, along with CWCH, 609 SC, JKH, ALM, JAT, ICC, GC, GCW, SB, AJ, TK, KK, and SAS, provided insights on the 610 plausibility of the species distribution model outputs. The initial draft of the manuscript was 611 612 written by EEJ, EYHY, TCB, and ACH, with EYHY responsible for data visualization. All authors contributed input and suggestions to the draft and approved the final manuscript. 613 614 615 **Additional information** 616

- 617
- 618

619 Competing interests620

- 621 The authors declare no competing interests.
- 622 623
- 624 Supplementary Information
- 625



Supplementary Figure 1. Climatic predictor variables included in our SDMs.



630 Supplementary Figure 2. Non-climatic predictor variables included in our SDMs.

References

- 634
 635 1. de Bruyn, M. et al. Borneo and Indochina are major evolutionary hotspots for Southeast
 636 Asian biodiversity. *Syst. Biol.* 63, 879-901 (2014).
- 637 2. Verde Arregoitia, L. D. Biases, gaps, and opportunities in mammalian extinction risk
 638 research. *Mammal Rev.* 46, 17-29 (2016).
- 639 3. Cardoso, P., Erwin, T. L., Borges, P. A. & New, T. R. The seven impediments in
 640 invertebrate conservation and how to overcome them. *Biol. Conserv.* 144, 2647-2655
 641 (2011).

- King, Shuang. et al. Conservation of data deficient species under multiple threats:
 Lessons from an iconic tropical butterfly (*Teinopalpus aureus*). *Biol. Conserv.* 234, 154164 (2019).
- 5. Pinkert, S., Barve, V., Guralnick, R. & Jetz, W. Global geographical and latitudinal
 variation in butterfly species richness captured through a comprehensive country-level
 occurrence database. *Glob. Ecol. Biogeogr.* 31, 830-839 (2022).
- 648
 6. Kawahara, A. Y. et al. A global phylogeny of butterflies reveals their evolutionary
 history, ancestral hosts and biogeographic origins. *Nat. Ecol. Evol.* 7, 903–913 (2023).
- Pinkert, S., Sica, Y. V., Winner, K. & Jetz, W. The potential of ecoregional range maps
 for boosting taxonomic coverage in ecology and conservation. *Ecography* 2023, (2023).
- 8. Whittaker, R. J. et al. Conservation biogeography: assessment and prospect. *Divers. Distrib.* 11, 3-23 (2005).
- Earl, C. et al. Spatial phylogenetics of butterflies in relation to environmental drivers and
 angiosperm diversity across North America. *IScience* 24, (2021).
- Wei, C.-H., Lohman, D. J., Peggie, D. & Yen, S.-H. An illustrated checklist of the genus
 Elymnias Hübner, 1818 (Nymphalidae, Satyrinae). *Zookeys* 676, 47-152. (2017).
- Condamine, F. L. et al. Fine-scale biogeographical and temporal diversification
 processes of peacock swallowtails (*Papilio* subgenus *Achillides*) in the Indo-Australian
 Archipelago. *Cladistics* 29, 88–111 (2012).
- 12. Toussaint, E. F. et al. Comparative molecular species delimitation in the charismatic
 Nawab butterflies (Nymphalidae, Charaxinae, *Polyura*). *Mol. Phylogenet. Evol.* 91, 194209 (2015).
- Scriven, S. A. et al. Assessing the effectiveness of protected areas for conserving range restricted rain forest butterflies in Sabah, Borneo. *Biotropica* 52, 380-391 (2020).
- Beck, J., Böller, M., Erhardt, A. & Schwanghart, W. Spatial bias in the GBIF database
 and its effect on modeling species' geographic distributions. *Ecol. Inform.* 19, 10-15
 (2014).
- Hughes, A. C. et al. Sampling biases shape our view of the natural world. *Ecography* 44, 1259-1269 (2021).
- 16. Meyer, C., Kreft, H., Guralnick, R. & Jetz, W. Global priorities for an effective information basis of biodiversity distributions. *Nat. Commun.* 6, 1-8 (2015).
- Peterson, A. T., Navarro-Sigüenza, A. G. & Gordillo, A. Assumption-versus data-based
 approaches to summarizing species' ranges. *Conserv. Biol.* 32, 568-575 (2016).
- 675 18. Chowdhury, S. et al. Using social media records to inform conservation planning.
 676 *Conserv. Biol.* 38, (2024).
- 19. Jetz, W., Sekercioglu, C. H. & Watson, J. E. Ecological correlates and conservation
 implications of overestimating species geographic ranges. *Conserv. Biol.* 22, 110-119
 (2008).
- Sofaer, H. R. et al. Development and delivery of species distribution models to inform decision-making. *Biosci.* 69, 544-557 (2019).
- Anderson, R. P. et al. *Ecological niches and geographic distributions* (Princeton University Press, 2011).
- Troudet, J., Grandcolas, P., Blin, A., Vignes-Lebbe, R. & Legendre, F. Taxonomic bias
 in biodiversity data and societal preferences. *Sci. Rep.* 7, (2017).
- Kramer-Schadt, S. et al. The importance of correcting for sampling bias in MaxEnt
 species distribution models. *Divers. Distrib.* 19, 1366-1379 (2013).
- 688 24. Guillera-Arroita, G. et al. Is my species distribution model fit for purpose? Matching
 689 data and models to applications. *Glob. Ecol. Biogeogr.* 24, 276-292 (2015).

- Smeraldo, S. et al. Species distribution models as a tool to predict range expansion after reintroduction: A case study on Eurasian beavers (Castor fiber). *J. Nat. Conserv.* 37, 12-20 (2017).
- 693 26. Guisan, A. et al. Predicting species distributions for conservation decisions. *Ecol. Lett.*694 16, 1424-1435 (2013).
- Araújo, M. B. et al. Standards for distribution models in biodiversity assessments. *Sci. Adv.* 5, (2019).
- Maes, D. et al. The potential of species distribution modelling for reintroduction
 projects: the case study of the Chequered Skipper in England. *J. Insect Conserv.* 23,
 419-431 (2019).
- Pacifici, M. et al. Assessing species vulnerability to climate change. *Nat. Clim. Change*5, 215-224 (2015).
- 30. Attorre, F. et al. How to include the impact of climate change in the extinction risk
 assessment of policy plant species? *J. Nat. Conserv.* 44, 43-49 (2018).
- 31. Lohman, D. J. et al. Biogeography of the Indo-Australian archipelago. *Annu. Rev. Ecol. Evol. Syst.* 42, 205-226 (2011).
- Hughes, A. C. Understanding the drivers of Southeast Asian biodiversity loss.
 Ecosphere 8, (2017).
- 33. Corlett, R. T. *The Ecology of Tropical East Asia* 3rd edn (Oxford University Press, 2019).
- 34. Sodhi, N. S., Koh, L. P., Brook, B. W. & Ng, P. K. L. Southeast Asian biodiversity: an
 impending disaster. *Trends Ecol. Evol.* 19, 654-660 (2004).
- 35. Wilcove, D. S., Giam, X., Edwards, D. P., Fisher, B., & Koh, L. P. Navjot's nightmare
 revisited: logging, agriculture, and biodiversity in Southeast Asia. *Trends Ecol. Evol.* 28,
 531-540 (2013).
- 715 36. Orr, M. C. et al. Global patterns and drivers of bee distribution. *Curr Biol.* 31, 451-458
 716 (2021).
- 717 37. *Global Biodiversity Information Facility* <u>https://doi.org/10.15468/dd.nvw5wr</u> (2024).
- 38. Ball-Damerow, J. E. et al. Research applications of primary biodiversity databases in the digital age. *PloS one* 14, (2019).
- Gaiji, S. et al. Content assessment of the primary biodiversity data published through
 GBIF network: status, challenges and potentials. *Biodiversity Informatics* 8, (2013).
- 40. Costello, M. J., Michener, W. K., Gahegan, M., Zhang, Z. Q. & Bourne, P. E.
 Biodiversity data should be published, cited, and peer reviewed. *Trends Ecol. Evol.* 28, 454-461 (2013).
- 41. Goodwin, Z. A., Harris, D. J., Filer, D., Wood, J. R. & Scotland, R. W. Widespread mistaken identity in tropical plant collections. *Curr. Biol.* 25, R1066-R1067 (2015).
- 42. Chowdhury, S. et al. Butterflies are weakly protected in a mega-populated country,
 Bangladesh. *Glob. Ecol. Conserv.* 26, (2021).
- 43. Shields, O. World numbers of butterflies. J. Lepid. Soc. 43, 178-183 (1989).
- 44. Robbins, R. K. & Opler, P. A. in *Biodiversity II: Understanding and Protecting Our Biological Resources* (eds. Reaka-Kudla, M. L., Wilson, D. E. & Wilson, E. O.) Ch. 6
 (Joseph Henry Press, 1997).
- 45. Dinerstein, E. et al. An ecoregion-based approach to protecting half the terrestrial realm. *Biosci.* 67, 534–545 (2017).
- 735 46. Thuiller, W. et al. biomod2: Ensemble Platform for Species Distribution Modeling. R
 736 package version 4.2-4. https://CRAN.R-project.org/package=biomod2 (2023).
- 47. Guisan, A., Thuiller, W. & Zimmermann, N. E. *Habitat Suitability and Distribution Models: With Applications in R* (Cambridge University Press, 2017).

49. Porfirio, L. L. et al. Improving the use of species distribution models in conservation 741 planning and management under climate change. PloS One 9, (2014). 742 50. Elith, J. & Graham, C. H. Do they? How do they? WHY do they differ? On finding 743 reasons for differing performances of species distribution models. *Ecography* 32, 66-77 744 (2009).745 51. Therneau, T., Atkinson, B. & Ripley, B. Rpart: Recursive Partitioning. R Package 746 version 4.1-3. http://CRAN.R-project.org/package=rpart (2013). 747 748 52. Chen, T. et al. xgboost: Extreme Gradient Boosting. R package version 1.7.5.1. https://CRAN.R-project.org/package=xgboost (2023). 749 53. Natekin, A. & Knoll, A. Gradient boosting machines, a tutorial. Front. Neurorobot. 7, 750 751 (2013). 54. Hijmans, R. J., Cameron, S. E., Parra, J. L., Jones, P. G. & Jarvis, A. Very high 752 resolution interpolated climate surfaces for global land areas. Int. J. Climatol. 25, 1965-753 1978 (2005). 754 755 55. Poggio, L. et al. SoilGrids 2.0: producing soil information for the globe with quantified spatial uncertainty. SOIL 7, 217-240 (2021). 756 56. International Soil Reference and Information Centre https://www.isric.org/ (2024). 757 758 57. Ågren, G. I., Wetterstedt, J. M. & Billberger, M. F. Nutrient limitation on terrestrial 759 plant growth-modelling the interaction between nitrogen and phosphorus. New Phytol. 760 194, 953-960 (2012). 761 58. Lal, R. Soil health and carbon management. Food Energy Secur. 5, 212-222 (2016). 59. Neina, D. The role of soil pH in plant nutrition and soil remediation. Appl. Environ. Soil 762 Sci. 2019, 1-9 (2019). 763 764 60. Barbet-Massin, M. & Jetz, W. A 40-year, continent-wide, multispecies assessment of relevant climate predictors for species distribution modelling. Divers. Distrib. 20, 1285-765 1295 (2014). 766 61. Zeng, Y., Low, B. W. & Yeo, D. C. Novel methods to select environmental variables in 767 MaxEnt: A case study using invasive crayfish. Ecol. Model. 341, 5-13 (2016). 768 62. Lang, N., Jetz, W., Schindler, K. & Wegner, J. D. A high-resolution canopy height 769 model of the Earth. Nat. Ecol. Evol. 7, 1778-1789 (2023). 770 63. Stockwell, D. R. & Peterson, A. T. Effects of sample size on accuracy of species 771 distribution models. Ecol. Model. 148, 1-13 (2002). 772 64. van Proosdij, A. S., Sosef, M. S., Wieringa, J. J. & Raes, N. Minimum required number 773 774 of specimen records to develop accurate species distribution models. *Ecography* 39, 775 542-552 (2016). 65. Venables, W. N. & Ripley, B. D. Modern Applied Statistics with S-PLUS 4th edn 776 777 (Springer, 2002). 66. Phillips, S. J. et al. Sample selection bias and presence-only distribution models: 778 779 implications for background and pseudo-absence data. Ecol. Appl. 19, 181-197 (2009). 780 67. Ferrier, S., Watson, G., Pearce, J. & Drielsma, M. Extended statistical approaches to modelling spatial pattern in biodiversity in northeast New South Wales. I. Species-level 781 modelling. Biodiversity Conserv. 11, 2275-2307 (2002). 782 783 68. Barbet-Massin, M., Jiguet, F., Albert, C. H. & Thuiller, W. Selecting pseudo-absences for species distribution models: How, where and how many?. Methods Ecol. Evol. 3, 784 327-338 (2012). 785 786 69. Breiner, F. T., Guisan, A., Bergamini, A. & Nobis, M. P. Overcoming limitations of modelling rare species by using ensembles of small models. *Methods Ecol. Evol.* 6, 787 1210-1218 (2015). 788

48. Phillips, S. J. maxnet: Fitting 'Maxent' Species Distribution Models with 'glmnet'. R

package version 0.1.4. https://CRAN.R-project.org/package=maxnet (2021).

739 740

- 789 70. Araújo, M. B. & New, M. Ensemble forecasting of species distributions. *Trends Ecol.* 790 *Evol.* 22, 42-47 (2007).
- 791 71. Marmion, M., Parviainen, M., Luoto, M., Heikkinen, R. K. & Thuiller, W. Evaluation of
 792 consensus methods in predictive species distribution modelling. *Divers. Distrib.* 15, 59793 69 (2009).
- 72. Landis, J. R. & Koch, G. G. The measurement of observer agreement for categorical data. *Biometrics* 33, 159-174 (1977).
- 73. Allouche, O., Tsoar, A. & Kadmon, R. Assessing the accuracy of species distribution
 models: prevalence, kappa and the true skill statistic (TSS). *J. Appl. Ecol.* 43, 1223-1232
 (2006).
- 799 74. Hernandez, P. A., Graham, C. H., Master, L. L. & Albert, D. L. The effect of sample
 800 size and species characteristics on performance of different species distribution
 801 modeling methods. *Ecography* 29, 773-785 (2006).
- Kirzel, A. H., Le Lay, G., Helfer, V., Randin, C. & Guisan, A. Evaluating the ability of habitat suitability models to predict species presences. *Ecol. Model.* 199, 142-152 (2006).
- Kobo, J. M., Jiménez-Valverde, A. & Real, R. AUC: a misleading measure of the
 performance of predictive distribution models. *Glob. Ecol. Biogeogr.* 17, 145-151
 (2008).
- R Core Team. R: A language and environment for statistical computing, version 4.1.2. R
 Foundation for Statistical Computing https://www.R-project.org/ (2021).
- 78. Capinha, C., Rocha, J. & Sousa, C. A. Macroclimate determines the global range limit of
 Aedes aegypti. *Ecohealth* 11, 420-428 (2014).
- 79. Jones, C. C., Acker, S. A. & Halpern, C. B. Combining local-and large-scale models to
 predict the distributions of invasive plant species. *Ecol. Appl.* 20, 311-326 (2010).
- 80. Rupprecht, F., Oldeland, J. & Finckh, M. Modelling potential distribution of the
 threatened tree species Juniperus oxycedrus: how to evaluate the predictions of different
 modelling approaches? J. Veg. Sci. 22, 647-659 (2011).
- 817 81. Pomoim, N., Hughes, A. C., Trisurat, Y. & Corlett, R. T. Vulnerability to climate
 818 change of species in protected areas in Thailand. *Sci. Rep.* 12, (2022).
- 819 82. Zurell, D. A standard protocol for reporting species distribution models. *Ecography* 43, 1261-1277 (2020).
- 83. Sholihah, A. et al. Impact of Pleistocene eustatic fluctuations on evolutionary dynamics
 in Southeast Asian biodiversity hotspots. *Syst. Biol.* **70**, 940-960 (2021).
- 823 84. Voris, H. K. Maps of Pleistocene sea levels in Southeast Asia: Shorelines, river systems
 824 and time durations. *J. Biogeogr.* 27, 1153-1167 (2000).
- 85. Monastyrskii, A. L. & Vane-Wright, R. I. Identity of *Euploea orontobates* Fruhstorfer,
 1910 (Lepidoptera: Nymphalidae), a milkweed butterfly from Thailand and Vietnam. *Zootaxa* 1991, 43-50 (2009).
- 86. Boakes, E. H. et al. Distorted views of biodiversity: spatial and temporal bias in species
 occurrence data. *PloS Biol.* 8, (2010).
- 87. Pearson, R. G. & Dawson, T. P. Predicting the impacts of climate change on the
 distribution of species: are bioclimate envelope models useful? *Glob. Ecol. Biogeogr.*832 12, 361-371 (2003).
- 833 88. Carvalho, A. P. S. et al. Comprehensive phylogeny of Pieridae butterflies reveals strong
 834 correlation between diversification and temperature. *iScience* 27, (2024).
- 835