1	Genetic variation of heat tolerance in a model
2	ectotherm: an approach using thermal death time curves
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25 Abstract

26 The assessment of thermal tolerance holds significant importance in predicting the 27 physiological responses of ectotherms, particularly in elucidating their capacity for evolutionary adaptation in the context of global warming. Current approaches to assessing 28 29 thermal tolerance have limitations that can lead to misleading results, especially with regard 30 to the heritability of thermal limits. In this study, we examined twenty isogenic lines of 31 Drosophila melanogaster from the DGRP panel to characterize their thermal death time (TDT) 32 curves, which account for the duration and intensity of heat stress. Furthermore, we examined the extent of genetic variation in the intercept and slope of the linear TDT curves, which are 33 34 labelled as CTmax and thermal sensitivity z. Our analysis revealed evidence of heritable 35 variation in each of the two parameters. Furthermore, simulations of the evolutionary consequences of selection on either CTmax or z indicate that selection on one parameter 36 37 induces changes in the other parameter as a correlated response. We conclude that the 38 evolution of thermosensitive or thermotolerant strategies is better achieved by directional 39 selection to decrease or increase CTmax, which may aid in mitigating the effects of global 40 warming on ectotherms.

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42 Keywords: global warming, heritability, isogenic lines, thermal death time curves

44 **1. Introduction**

45 There is substantial evidence pointing to an unprecedented rise in the temperature of our 46 planet. According to climate models, if the present warming trends persist, the surface temperature of the Earth's surface will surpass the average at the end of the 19th century by 47 48 1.5°C [1]. It is not surprising that this rise will have repercussions on the biota present on our 49 planet, particularly for animals such as ectotherms, whose physiological processes are closely linked to ambient temperature [2,3]. Climate change is already having an impact at the 50 51 demographic level. Many species are shifting their ranges, often towards cooler regions [4], 52 while others are threatened with extinction [5]. In contrast, some lineages are already able to withstand high temperatures [6], while others show phenotypic plasticity [7], and some species 53 54 may evolve in response to warming [8,9]. Assessing the adaptive capacity and heritability of 55 heat tolerance is challenging, as inaccuracies in models can lead to either under- or 56 overestimation of vulnerability to climate change [10,11].

57 The current debate about the evolutionary potential of heat tolerance in ectotherms 58 can be summarized in two main points. First, there is the notion that ectotherms exhibit limited 59 variation in heat tolerance, suggesting that heat tolerance is evolutionary constrained. Natural 60 selection appears to affect physiological responses to lower temperatures more than to higher 61 temperatures [12]. Interspecific studies have generally failed to detect genetic variability in 62 heat tolerance, variation between species and populations, and a lack of latitudinal diversity 63 [13,14]. Likewise, selection and heritability experiments on single species suggest limited 64 increases in upper thermal limits. Second, a complicating factor in understanding the genetic 65 basis of upper thermal limits is that these are to some extent affected by methodological issues 66 [for a review, see 12]. In dynamic assays, heat tolerance is typically evaluated by determining 67 the upper critical thermal maximum (CTmax). This is measured as the temperature at which 68 an ectotherm loses motor function when exposed to gradually increasing temperatures-. The CTmax represents the highest ambient temperature an ectotherm can withstand under 69 70 specific experimental conditions before succumbing to heat stress [15,16]. Studies using this 71 dynamic assays have found suggest a limited evolutionary potential for ectotherms to increase 72 their ability to tolerate high temperatures [17]. Despite the possibility of methodological issues 73 underestimating the actual evolutionary potential to withstand increasing temperatures [10], it 74 is imperative to acknowledge that a critical thermal limit is merely a component of the more 75 intricate trait "thermal tolerance" [18].

76 To circumvent this limitation, an increasing number of studies have employed thermal death time (TDT) curves to measure heat tolerance [19-24]. TDT curves [sensu 25], 77 78 acknowledges that survival probability is influenced by both temperature and exposure time 79 [26,27], and may provide a more nuanced view of the two reasons mentioned above for the 80 ongoing debate. However, the extent to which TDT curves reflect evolutionary changes within 81 a species remains unclear. In this context, we take advantage of recent research using the 82 Drosophila Genetic Reference Panel (DGRP) [18,28] to investigate the genetic variation of 83 heat tolerance in Drosophila melanogaster using TDT curves. The wild-type DGRP lines of 84 this panel are derived from a single natural population and have been inbred to homozygosity, providing extensive information on genetic variation at multiple levels [29] and offering unique 85 opportunities to quantify the genetic basis of physiological traits such as heat tolerance. 86

87 **2. Material and methods**

88 (a) Experimental flies and rearing conditions

Twenty inbred, isogenic wild-type *Drosophila melanogaster* (Meigen 1830) lines from the Drosophila Genetic Reference Panel (DGRP) were used as the study model. These lines were previously employed in a study with a different objective [21], and we assume that this subset represents an approximately random collection with respect to the focus of interest described here.

All twenty selected lines were obtained from the Bloomington Drosophila Stock Center in March 2018. They were maintained in quarantine on standard cornmeal-agar-yeast media at room temperature (approximately 22°C) for four generations until May 2018. The rearing conditions were identical to those detailed in Leiva et al. [21].

98 (b) Thermal death time (TDT) curves

99 We measured the heat tolerance of individual female and male flies using a similar 100 experimental protocol as outlined in Verspagen et al. [24]. This involved using a heating 101 circulating bath and a wireless thermometer to measure temperature consistently throughout 102 each trial. Individual virgin flies were placed in sealed 4-mL glass vials, arranged on a 103 Plexiglas[™] rack, and submerged in a 9.5-L glass aquarium filled with water set to a constant 104 temperature of 36, 37, 38, or 39°C. During each trial, a Nikon D5300 with the time-lapse 105 feature captured images at 10-second intervals. Subsequently, the compiled images were transformed into reversed videos using the open-source software Blender. Before initiating 106 107 the thermal tolerance experiments, the flies were allowed approximately 30 minutes in the 108 vials at room temperature for recovery following light CO₂ anaesthesia. This recovery period 109 proved effective, as the flies exhibited active flying or walking inside the vial.

A total of 1,686 flies underwent survival time measurements, and the parameters of the thermal death time (TDT) curves CTmax and *z* were calculated for each DGRP line and sex. The calculation utilized the equation outlined in Rezende et al. [26]:

113
$$\log_{10} t = \beta_0 + \beta_1 T = \frac{CT_{max} - T}{z}$$
(1)

where *t* represents the survival time in minutes, CTmax is the temperature (°C) where the survival time is $\log_{10} t = 0$ after 1 min of exposure to assay temperature, *T* is the assay temperature (°C), and the thermal sensitivity *z* is the temperature change (°C) required for a ten-fold difference in survival time. We controlled assay temperature *T* and measured time as the dependent variable. The estimation of CTmax and *z* for each DGRP line and sex involved regression analysis of log₁₀-transformed survival times against the four temperature treatments and a back-transformation where CTmax = β_0/β_1 and z = $1/\beta_1$.

121 (c) Estimation of variance components and broad-sense heritability of CTmax and z

For each DGRP genotype, we first assessed the effect of stressful temperatures (covariate) on survival time (in $log_{10} min$) by using for each sex the following general mixed ANCOVA model that allows separating slopes and intercepts:

Survival time =
$$\beta_0 + \beta_1 \cdot \text{stress } T^a + \mu_0 Z + \mu_1 Z \cdot \text{stress } T^a + \varepsilon$$
 (2)

126 where

127 β_0 and β_1 : intercept and slope of the fixed effect of stress temperature stress T^a ,

128 μ_0 : vector of random coefficients representing the effect of each genotype on $\beta_{0,1}$

129 μ_1 : vector of random coefficients representing the effect of each genotype on β_1 ,

130 Z: Design matrix (array of dummy variables) representing the genotypes,

131 ε: vector of random errors

We fitted linear mixed-effects models [30] and obtained various estimates of the variance components $\hat{\sigma}_{\hat{\beta}_{0i}}^2$ ($i = 1, \dots, 20$), $\hat{\sigma}_{\hat{\beta}_{1i}}^2$ and $\hat{\sigma}_{\varepsilon}^2$ (caret denotes "estimate") that refer, respectively, to the variation in the intercepts and slopes of the TDT curves for the DGRP genotypes, and the residual variation. Model coefficients and variance components were then used to estimate $CTmax_i = \frac{-\hat{\beta}_{0i}}{\hat{\beta}_{1i}}$ and $z_i = \frac{-1}{\hat{\beta}_{1i}}$, whose Taylor expanded variances [31, p. 240], became,

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$$\operatorname{Var}(\operatorname{CT}_{\max_{i}}) = \operatorname{Var}\left(\frac{-\widehat{\beta}_{0i}}{\widehat{\beta}_{1i}}\right) \approx \frac{\left(\mu_{-\widehat{\beta}_{0i}}\right)^{2}}{\left(\mu_{\widehat{\beta}_{1i}}\right)^{2}} \left[\frac{\widehat{\sigma}_{-\widehat{\beta}_{0i}}^{2}}{\left(\mu_{-\widehat{\beta}_{0i}}\right)^{2}} + \frac{\widehat{\sigma}_{\widehat{\beta}_{1i}}^{2}}{\left(\mu_{\widehat{\beta}_{1i}}\right)^{2}} - 2\frac{\operatorname{Cov}(-\widehat{\beta}_{0i},\widehat{\beta}_{1i})}{\mu_{-\widehat{\beta}_{0i}}\mu_{\widehat{\beta}_{1i}}}\right]$$

(3)

(4)

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140
$$\operatorname{Var}(z_i) \approx \frac{\hat{\sigma}_{\hat{\beta}_{1i}}^2}{\left(\mu_{\hat{z}_i}\right)^4}$$

141 We then estimated broad-sense heritability as:

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$$H_{CT_{max}}^{2} = \frac{\operatorname{Var}(\operatorname{CT}_{max_{i}})}{\operatorname{Var}(\operatorname{CT}_{max_{i}}) + \operatorname{Var}(z_{i}) + \frac{\widehat{\sigma}_{\varepsilon}^{2}}{n_{0}}},$$

142

145
$$H_z^2 = \frac{\operatorname{Var}(\hat{z}_i)}{\operatorname{Var}(\operatorname{CT}_{\max_i}) + \operatorname{Var}(z_i) + \frac{\hat{\sigma}_{\varepsilon}^2}{n_0}}$$

144

146 where

147
$$n_0 = \frac{1}{a-1} \left(\sum_{i=1}^a n_i - \frac{\sum_{i=1}^a n_i^2}{\sum_{i=1}^a n_i} \right)$$
(5)

148 is the appropriate mean value of the number of flies from each sex and DGRP line used at 149 each stressful temperature to estimate the TDT curves [32, p. 212] The reason we divided the residual variance by n_0 is because we are using line means [33]. In our case $n_0 = 10.2$ for 150 151 females and $n_0 = 10.8$ for males.

152 Delete-one-DRGP genotype at a time data resampling was also carried out to estimate the genetic components of variance and their standard errors [34]. A total of 20 pseudovalues 153 154 for each sex were obtained by dropping, in turn, each DGRP line and calculating:

 $\phi_i = N\widehat{\Theta}_N - (N-1)\widehat{\Theta}_{N-1\,i},$ 156

(6)

155

where ϕ_i is the *i*th pseudovalue, $\hat{\Theta}_N$ is the corresponding variance estimate using all N = 20157 DGRP genotypes, and $\hat{\Theta}_{N-1,i}$ is that estimate by dropping the *i*th DGRP genotype alone. The 158 159 jackknife estimate is the average of ϕ_i , and its standard error is given by

161
$$SE = \sqrt{\frac{\sum_{i=1}^{i=N} (\phi_i - \bar{\phi})^2}{N(N-1)}}.$$
160 (7)

160

Approximate 95% jackknife confidence intervals were obtained as $\bar{\phi} \pm 2$ SE. Initially, 162 the analyses for computing variance components and heritability were implemented by MS in 163 MATLAB. To enhance reproducibility, FPL and EJN replicated the analyses and implemented 164 165 them in R version 4.3.2 [35]. The data used in these analyses were based on a recently 166 reported study [21,36].

(d) Simulated selection on the TDT curves 167

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169 Appropriate estimates of the additive-genetic G and phenotypic P (co)variance matrices in an outbred Drosophila population are needed to explore the evolutionary consequences of 170 selection on CTmax and z from the multivariate breeder's equation $\Delta \mu = G\beta = GP^{-1}s$. Here, 171 172 the term $\Delta \mu$ is the vector of changes in trait means, β is the vector of selection gradients, and 173 s is the vector of selection differentials [37,38].

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175 The estimates of genetic variance-covariance components and broad-sense 176 heritability in the highly inbred DGRP lines yield only the relative contributions of CTmax and 177 z to the total genetic variance in the TDT curves [see 33]. To our knowledge, there are no 178 estimates of the narrow-sense heritability of TDT curves. Current evidence suggests that 179 upper critical limits estimated with different methodologies is moderately heritable, and it seems reasonable to assume that its narrow-sense heritability is $h^2 \approx 0.25$ [17,39,40]. Based 180 on this information and the relative contribution of CTmax and z to the total genetic variance 181 182 of TDT curves, we quantified the consequences of several simulated selective scenarios on 183 the evolution of thermal tolerance (represented by these two parameters in the TDT curves) 184 in an outbred population. Note that this will be "what-if" scenarios and more accurate estimates 185 of G and P would be needed for a more satisfactory answer.

186 **3. Results**

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(a) Determination of TDT curves and variance components

We observed substantial variation in thermal death time (TDT) curves across genetic lines for both females and males (Figure 1). At 36°C, the average survival times (\pm SD) were 150 \pm 47 minutes for females and 111 \pm 41.6 minutes for males. These durations decreased significantly at 39°C, with females surviving for 12.7 \pm 5.13 minutes and males for 14.7 \pm 5.6 minutes on average. Notably, this variation in thermal tolerance across genetic lines was consistently observed for both sexes (Figure 1).

195

196 Table 1 provides estimates of the variance-covariance components and broad-sense 197 heritability using different methods for estimating parameters in the linear mixed-effects model. 198 These estimates were highly consistent across the various methods of estimation. As 199 indicated by the jackknife 95% confidence intervals, all variance components were significantly different from zero. Furthermore, the genetic covariance between β_0 and β_1 was 200 201 negative. Broad-sense heritability was around 0.75 for CTmax and around 0.25 for z. In other 202 words, CTmax accounts for approximately 75% and z accounts for approximately 25% of the 203 total genetic variance in the TDT curves.



Figure 1. Thermal death time (*y*-axis in log₁₀-scale) curves for females (left) and males (top). Dots represent the individual survival time for females (green, left plot) and males (brown, right plot).

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Table 1. Estimation of variance-covariance components and broad-sense heritability using multiple

approaches for parameters estimation in the linear mixed-effects model described in equation (2).
 Estimates were obtained with MATLAB.

Method	Component	Females					Males			
		Estimate	Jackknife	Lower 95% CI	Upper 95% CI	Estimate	Jackknife	Lower 95% CI	Upper 95% Cl	
ANOVA	$\hat{\sigma}^2_{\hat{eta}_{0i}}$	4.176376	4.222055	0.818848	7.625262	7.894796	7.885140	1.780019	13.990261	
	$\hat{\sigma}^2_{\hat{\beta}_{1i}}$	0.003093	0.003131	0.000591	0.005671	0.005759	0.005753	0.001330	0.010175	
	$Cov(\hat{eta}_0,\hat{eta}_1)$	-0.113664	-0.114983	-0.207922	-0.022044	-0.213231	-0.212990	-0.377274	-0.048706	
	σ_c^2	0.025323	0.025350	0.017416	0.033283	0.022589	0.022626	0.016258	0.028993	
	$\sigma^2_{CT_{max}}$	0.594709	0.611843	0.095040	1.128645	2.085709	2.079167	0.604101	3.554233	
	$\sigma_{\hat{z}}$	0.163722	0.167434	0.039708	0.295159	0.689940	0.680720	0.146407	1.215034	
	$H^2_{\widehat{CT}_{max}}$	0.781584	0.790793	0.713939	0.867647	0.750866	0.750360	0.699404	0.801316	
	$H_{\hat{Z}}^2$	0.215169	0.206795	0.131965	0.281624	0.248382	0.249001	0.197763	0.300239	
	- 2									
ML	$\hat{\sigma}^2_{\hat{\beta}_{0i}}$	3.627185	3.798623	0.851691	6.745556	7.508898	7.906648	1.943793	13.869503	
	$\hat{\sigma}^2_{\hat{m{eta}}_{1i}}$	0.002667	0.002805	0.000611	0.004998	0.005478	0.005781	0.001472	0.010090	
	$Cov(\hat{eta}_0,\hat{eta}_1)$	-0.098364	-0.103366	-0.183723	-0.023008	-0.202806	-0.214053	-0.374382	-0.053725	
	σ_{ε}^2	0.025342	0.025379	0.017435	0.033323	0.022591	0.022629	0.016262	0.028995	
	$\sigma^2_{\widehat{CT}_{max}}$	0.486893	0.444964	-0.009015	0.898944	1.984456	1.900759	0.570539	3.230979	
	σ _A	0.141176	0.149578	0.038018	0.261138	0.656208	0.685757	0.170153	1.201361	
	$H^2_{\widehat{CT}_{max}}$	0.772182	0.758771	0.663605	0.853937	0.750905	0.730511	0.670472	0.790550	
	$H_{\hat{z}}^2$	0.223896	0.238107	0.145710	0.330503	0.248304	0.268794	0.208392	0.329196	
REML	$\hat{\sigma}^2_{\hat{\beta}_{0i}}$	3.855075	3.790919	0.688634	6.893203	7.934343	7.906623	1.606537	14.206708	
	$\hat{\sigma}^2_{\hat{\beta}_{1i}}$	0.002833	0.002803	0.000489	0.005117	0.005787	0.005781	0.001231	0.010332	
	$Cov(\hat{\beta}_0, \hat{\beta}_1)$	-0.104513	-0.103320	-0.188112	-0.018528	-0.214281	-0.214055	-0.383359	-0.044751	
	σ_s^2	0.025341	0.025378	0.017435	0.033321	0.022591	0.022629	0.016262	0.028995	
	$\sigma_{\widehat{CT}_{max}}^2$	0.514967	0.401291	-0.009975	0.812558	2.096738	1.893919	0.422145	3.365694	
		0.149959	0.149561	0.031902	0.267221	0.693300	0.685338	0.140733	1.229943	
	$H^2_{\widehat{CT}_{max}}$	0.771604	0.736672	0.669950	0.803394	0.750946	0.731923	0.679424	0.784421	
	$H_{\hat{z}}^2$	0.224691	0.259674	0.193723	0.325625	0.248305	0.267388	0.214568	0.320208	

216 (b) Simulated selection on the TDT curves

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We can employ the raw phenotypic (co)variance matrix of the females from the DGRP lines 218 219 as a representative of the P matrix in an outbred population, omitting any changes in 220 phenotypic variance caused by inbreeding for the sake of simplicity [41]. Let us now assume 221 the absence of epistasis and use the genetic variation in the DGRP lines for CTmax and z 222 (REML estimates for females in Table 1), together with their relative broad-sense heritability, 223 to figure out what the G matrix would look like in an outbred population. Focusing on the univariate trait CTmax, imagine we obtain a narrow-sense heritability $h_{CTmax}^2 \approx 0.25$, whereas 224 focusing on z we obtain a narrow-sense heritability $h_z^2 \approx 0.07$, i.e. about one third the 225 226 heritability for CTmax. Rescaling the genetic components from the DGRP lines to these 227 values, we could presume the subsequent (co)variance genetic, environmental, and 228 phenotypic matrices for CTmax and z in the hypothetical outbred base population: 229

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 $G = \begin{bmatrix} 0.1489 & 0.0423 \\ 0.0423 & 0.0133 \end{bmatrix}; E = \begin{bmatrix} 0.4466 & 0.2618 \\ 0.2618 & 0.1691 \end{bmatrix}; P = \begin{bmatrix} 0.5955 & 0.3041 \\ 0.3041 & 0.1823 \end{bmatrix}$

The vector of phenotypic means in this base population is $\bar{\mu} = [41.9924 \ 2.7592]^T$ (*T* stands for transpose), where the first value is for CTmax and the second for *z* (female means from the DGRP lines). We assume that these values are representative of the outbred population, which is strictly true if allelic effects are additive [42].

237 We simulated three scenarios for increasing thermoresistance (Figure 2): directional selection for CTmax ($s = \begin{bmatrix} 1 & 0 \end{bmatrix}^T$), directional selection for both CTmax and z 238 $(s = \begin{bmatrix} 1 & 0.5 \end{bmatrix}^T)$, and directional selection for $z (s = \begin{bmatrix} 0 & 0.5 \end{bmatrix}^T)$. These selection differentials 239 correspond to intensities of selection $i_{CTmax} = 1.3$ and $i_z = 1.2$. The simulated selection 240 241 regimes used to illustrate the effects of selection are much stronger than we would expect in 242 nature because directional selection tends to be weak and rarely shifts mean by more than 243 half of a phenotypic standard deviation [43,44]. However, under extreme climatic events, such 244 as heat waves, which are considered to be major triggers of evolution [45], these intensities 245 of selection may not be unrealistic [46]. 246



247

248 Figure 2. The thermal death time (TDT) curve linearly describes the relationship between test 249 temperature (T) and survival time (t, \log_{10} scale) under heat stress conditions in D. melanogaster. 250 Thermal sensitivity (z) is the reciprocal of the slope (β_1), representing the increase in temperature 251 required to reduce survival time by one order of magnitude (10-fold). CTmax is the intersection at log₁₀ 252 = 0, corresponding to the knockdown or death temperature after 1 minute of exposure. The blue line 253 represents a thermosensitive genotype, which exhibits improved tolerance to acute, intense heat stress 254 but reduced tolerance to chronic, less intense heat. Conversely, the red line depicts a thermoresistant 255 genotype, exhibiting better tolerance to chronic stress but lower tolerance to acute stress.

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257 The simulated scenarios are shown in Figure 3. It is striking that a directional selection 258 gradient always has a different sign than its selection differential, and the evolutionary 259 response can be against the selection differential (Figure 3C). The inference from these 260 hypothetical selection regimes is that directional selection to increase thermal sensitivity z261 seems to hinder evolutionary responses to increasing CTmax. In other words, directional 262 selection to increase CTmax also increases z as a correlated response (Figure 3A) and, as 263 expected, drives the population towards a more thermoresistant state (Figure 2). However, 264 directional selection to increase z results in a decrease of CTmax as a correlated response, 265 which, in turn, also decreases z, and seemingly paradoxically drives the population towards 266 increasing thermosensitivity (Figure 2).



268 Figure 3. Hypothetical strong directional selection for increased thermoresistance. In blue the 95% 269 confidence ellipses of a simulated population of N = 10,000 flies from a bivariate normal distribution 270 whose phenotypes for CTmax and z are the sum of genetic effects with mean $\bar{\mu} = [41.9924 \ 2.7592]^T$ 271 and additive-genetic (co)variance G, plus environmental effects with mean 0 and (co)variance E (see 272 text). The arrows centred on the bivariate means represent the directions in which the data vary the 273 most (i.e., the eigenvectors of the covariance matrix of the data). In black are the 95% confidence 274 ellipses after an evolutionary shift in the means (we ignore changes in variance and covariance). Panel 275 A plots directional selection for increasing CTmax; the vector of selection gradients is $\beta =$ 276 $[11.3294 - 18.8959]^T$. Panel B plots directional selection for increasing both CTmax and z, where $\beta =$ 277 -0.3958]^{*T*}. Panel C plots directional selection for increasing z, where $\beta =$ [1.8814 278 [-9.4480] $[18.5002]^T$.

279 4. Discussion

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280 Heat tolerance has traditionally been examined from a physiological perspective, with a focus 281 on the mechanisms that cause animals to succumb to heat stress, such as oxygen limitation 282 or excessive water loss [see 47 for a review]. In this study, we examine the genetic 283 components of heat tolerance through the lens of thermal death time (TDT) curves. This allows partitioning the relative contribution of CTmax and z to the more complex trait 284 "thermotolerance" (Figure 2). Our approach has relied on inbred Drosophila melanogaster 285 lines from the DGRP panel, and it would be highly desirable to extend these analyses to 286 287 outbred populations.

289 The limited number of previous studies that have assessed the thermal tolerance of 290 DGRP lines focused primarily on measuring critical thermal limits with heating assays [48,49]. 291 These, along with other studies [12,50], suggest that heat tolerance is somewhat evolutionary 292 constrained, an idea often invoked to explain the absence of strong latitudinal clines in heat 293 tolerance, at least in terrestrial ectotherms [13]. Typically, assessments of organisms' 294 vulnerability to global warming usually compare experimentally derived thermal limits using 295 dynamic trials, in which animals are exposed to increasingly higher temperatures [15,51]. 296 During these trials, the intensity and duration of stress increase concurrently. As a result, 297 animals often succumb to heat stress in rapid succession, leading to small variances and small 298 standard deviations in the measurements, making CTmax an attractive endpoint to use in 299 treatment comparisons. However, CTmax is a single point, whereas the trait of interest, 300 namely the ability of an organism to deal with heat stress, is a linear function describing how 301 stress intensity and stress duration impact survival [26]. Thus, ramping trials approach 302 overlooks the cumulative nature of heat injury and the time-dependent effects of thermal 303 tolerance [52–54], potentially underestimating organisms' vulnerability to global warming [55]. We contend that utilizing TDT curves to evaluate both CTmax and *z* parameters, as well as 304 305 their underlying genetic basis, would provide more accurate predictions. Here, we have 306 developed a methodological approach to estimate the variance components and heritability of 307 the relevant parameters in the TDT curves.

308 The hypothetical selection scenarios allow us to understand how thermosensitive and 309 thermotolerant strategies (Figure 2) could evolve. They suggest that thermosensitive or 310 thermotolerant strategies are better achieved by directional selection to decrease or increase 311 CTmax. This conclusion holds in more realistic scenarios, where the intensity of selection on 312 CTmax and/or z might be relatively weak. We acknowledge that our approach is only a rough estimate of the problem and that several caveats could be raised. For instance, although there 313 314 is a high positive correlation across species for parameters CTmax and z (r = 0.92; [26]), the 315 G matrix used to simulate the selection scenarios could overestimate the additive genetic 316 covariance in an outbred Drosophila population due to higher linkage disequilibria in the DGRP 317 lines [33]. For the time being, however, we offer the somewhat unexpected results as a 318 cautionary tale about drawing conclusions without considering the multivariate nature of the 319 trait thermal tolerance.

In summary, our findings suggest that the genetic correlation between CTmax and z impose constraints on thermal tolerance strategies. The simulations performed here highlight the importance of considering the multivariate nature of thermal tolerance traits and their genetic correlations when predicting evolutionary responses to climate change. Ultimately, this can be achieved by utilizing approaches that measure thermal tolerances considering both the duration and intensity of heat stress and its potential for evolution. Adopting such integrative approaches will enable more accurate predictions of how species might respond to increasing temperatures in a rapidly changing planet.

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330 Data accessibility

331 Data files and code supporting analyses, figures and tables of this study are publicly available on GitHub (https://github.com/felixpleiva/Genetic variation TDT). When using the code from 332 333 this manuscript, please cite it as [56]: Leiva FP, Santos M, Niklitschek EJ, Rezende EL, & Verberk WCEP. (2024). Paper data and code for: Genetic variation of heat tolerance in a 334 335 thermal death model ectotherm: an approach using time curves. Zenodo. 336 https://doi.org/10.5281/zenodo.12155988.

337 Authors' contributions

338 Félix P. Leiva: conceptualization, data curation, formal analysis, funding acquisition, 339 investigation, methodology, project administration, resources, software, validation, 340 visualization, writing – original draft preparation, writing – review and editing; Mauro Santos: 341 conceptualization, formal analysis, methodology, software, writing - original draft preparation, 342 writing – review and editing; Edwin J. Niklitschek: formal analysis, software, validation, writing 343 - review and editing; Enrico L. Rezende: conceptualization, investigation, writing - review and 344 editing; Wilco C.E.P. Verberk: conceptualization, formal analysis, funding acquisition, 345 investigation, methodology, resources, supervision, writing – review and editing. All authors 346 gave final approval for publication.

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357 References

- 2019 IPCC, 2018: Global Warming of 1.5°C. An IPCC Special Report on the impacts of global warming of 1.5°C above pre-industrial levels and related global greenhouse gas emission pathways, in the context of strengthening the global response to the threat of climate change, sustainable development, and efforts to eradicate poverty. Cambridge, UK and New York, NY, USA: Cambridge University Press.
- Huey RB, Stevenson RD. 1979 Integrating thermal physiology and ecology of ectotherms:
 a discussion of approaches. American Zoologist , 357–366.
- Rezende EL, Bozinovic F. 2019 Thermal performance across levels of biological organization. Philosophical Transactions of the Royal Society B **374**, 20180549.

- Pinsky ML, Selden RL, Kitchel ZJ. 2019 Climate-driven shifts in marine species ranges:
 scaling from organisms to communities. Annual review of marine science **12**, 153–179.
- 5. Urban MC. 2015 Accelerating extinction risk from climate change. Science **348**, 571–573.
- BioScience 50, 217–226.
 BioScience 50, 217–226.
- Gunderson AR, Dillon ME, Stillman JH. 2017 Estimating the benefits of plasticity in ectotherm heat tolerance under natural thermal variability. Functional Ecology **31**, 1529– 1539.
- Rudman Seth M., Greenblum Sharon I., Rajpurohit Subhash, Betancourt Nicolas J., Hanna Jinjoo, Tilk Susanne, Yokoyama Tuya, Petrov Dmitri A., Schmidt Paul. 2022 Direct observation of adaptive tracking on ecological time scales in Drosophila. Science **375**, eabj7484. (doi:10.1126/science.abj7484)
- Huey RB, Kingsolver JG. 1993 Evolution of resistance to high temperature in ectotherms.
 American Naturalist, S21–S46.
- Rezende EL, Tejedo M, Santos M. 2011 Estimating the adaptive potential of critical
 thermal limits: methodological problems and evolutionary implications. Functional Ecology
 25, 111–121.
- 11. Santos M, Castañeda LE, Rezende EL. 2011 Making sense of heat tolerance estimates
 in ectotherms: lessons from Drosophila. Functional Ecology 25, 1169–1180.
- Hoffmann AA, Chown SL, Clusella-Trullas S. 2013 Upper thermal limits in terrestrial
 ectotherms: how constrained are they? Functional Ecology 27, 934–949.
- Araújo MB, Ferri-Yáñez F, Bozinovic F, Marquet PA, Valladares F, Chown SL. 2013 Heat
 freezes niche evolution. Ecology Letters 16, 1206–1219.
 (doi:https://doi.org/10.1111/ele.12155)
- 391 14. Sunday JM, Bates AE, Dulvy NK. 2011 Global analysis of thermal tolerance and latitude
 392 in ectotherms. Proceedings of the Royal Society of London B: Biological Sciences 278,
 393 1823–1830. (doi:https://doi.org/10.1098/rspb.2010.1295)
- Sunday JM et al. 2019 Thermal tolerance patterns across latitude and elevation.
 Philosophical Transactions of the Royal Society B: Biological Sciences 374.
 (doi:10.1098/rstb.2019.0036)
- Hutchison VH. 1961 Critical Thermal Maxima in Salamanders. Physiological Zoology 34, 92–125. (doi:10.1086/physzool.34.2.30152688)
- 399 17. Santos M, Castañeda LE, Rezende EL. 2012 Keeping pace with climate change: what is
 400 wrong with the evolutionary potential of upper thermal limits? Ecology and Evolution 2,
 401 2866–2880. (doi:10.1002/ece3.385)
- 402 18. Leiva FP, Boerrigter JG, Verberk WCEP. 2023 The role of cell size in shaping responses
 403 to oxygen and temperature in fruit flies. Functional Ecology **37**, 1269–1279.
 404 (doi:10.1111/1365-2435.14294)

- 405 19. Burton T, Einum S. 2020 The old and the large may suffer disproportionately during
 406 episodes of high temperature: evidence from a keystone zooplankton species.
 407 Conservation Physiology 8, coaa038.
- 20. Castañeda LE, Rezende EL, Santos M. 2015 Heat tolerance in Drosophila subobscura along a latitudinal gradient: contrasting patterns between plastic and genetic responses.
 Evolution 69, 2721–2734.
- 411 21. Leiva FP, Santos M, Rezende EL, Verberk WCEP. 2024 Intraspecific variation of heat
 412 tolerance in a model ectotherm: The role of oxygen, cell size and body size. Functional
 413 Ecology 38, 439–448. (doi:10.1111/1365-2435.14485)
- 22. Truebano M, Fenner P, Tills O, Rundle SD, Rezende EL. 2018 Thermal strategies vary
 with life history stage. Journal of Experimental Biology **221**, jeb171629.
- 416 23. Verberk WCEP, Hoefnagel KN, Peralta-Maraver I, Floury M, Rezende EL. 2023 Long417 term forecast of thermal mortality with climate warming in riverine amphipods. Global
 418 Change Biology **29**, 5033–5043. (doi:10.1111/gcb.16834)
- 419 24. Verspagen N, Leiva FP, Janssen I, Verberk WCEP. 2020 Effects of developmental
 420 plasticity on heat tolerance may be mediated by changes in cell size in Drosophila
 421 melanogaster. Insect Science 27, 1244–1256. (doi:10.1111/1744-7917.12742)
- 422 25. Bigelow W. 1921 The logarithmic nature of thermal death time curves. The Journal of 423 Infectious Diseases , 528–536.
- 424 26. Rezende EL, Castañeda LE, Santos M. 2014 Tolerance landscapes in thermal ecology.
 425 Functional Ecology 28, 799–809.
- 426 27. Wang S, Tang J, Hansen JD. 2007 Experimental and simulation methods of insect thermal
 427 death kinetics. In Heat Treatments for Postharvest Pest Control: Theory and Practice (eds
 428 J Tang, EJ Mitcham, S Wang, S Lurie), pp. 105–132. Oxon, UK: CABI Publishing.
- 429 28. Leiva FP, Santos M, Rezende EL, Verberk WCEP. 2023 Intraspecific variation on heat
 430 tolerance in a model ectotherm: the role of oxygen, cell size and body size. Functional
 431 Ecology (doi:10.1111/1365-2435.14485)
- 432 29. Mackay TF et al. 2012 The Drosophila melanogaster genetic reference panel. Nature 482,
 433 173–178.
- 434 30. 2007 STATISTICA (data analysis software system).
- 435 31. Wolter KM. 2007 Introduction to Variance Estimation. New York, NY: Springer New York.
 436 (doi:10.1007/978-0-387-35099-8)
- 437 32. Sokal RR, Rohlf FJ. 1995 Biometry. New York.
- 33. Mackay TFC, Huang W. 2018 Charting the genotype–phenotype map: lessons from the
 Drosophila melanogaster Genetic Reference Panel. WIREs Developmental Biology 7,
 e289. (doi:10.1002/wdev.289)
- 441 34. Knapp SJ, Bridges-Jr WC, Yang M-H. 1989 Nonparametric confidence interval estimators
 442 for heritability and expected selection response. Genetics **121**, 891–898.

- 443 35. R Development Core Team. 2023 R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- 445 36. Leiva FP, Santos M, Rezende EL, Verberk WCEP. 2021 Paper data and code of manuscript: Intraspecific variation on heat tolerance in a model ectotherm: effects of body mass, cell size, oxygen and sex. Zenodo. (doi:https://doi.org/10.5281/zenodo.5120028)
- 448 37. Lande R. 1979 Quantitative genetic analysis of multivariate evolution, applied to brain:
 449 body size allometry. Evolution 33, 402–416.
- 38. Lande R, Arnold SJ. 1983 The measurement of selection on correlated characters.
 Evolution 37, 1210–1226.
- 452 39. Diamond SE. 2017 Evolutionary potential of upper thermal tolerance: biogeographic
 453 patterns and expectations under climate change. Annals of the New York Academy of
 454 Sciences 1389, 5–19. (doi:10.1111/nyas.13223)
- 455 40. Logan ML, Cox CL. 2020 Genetic constraints, transcriptome plasticity, and the 456 evolutionary response to climate change. Frontiers in Genetics **11**, 538226.
- 457 41. Fowler K, Whitlock MC. 1999 The distribution of phenotypic variance with inbreeding.
 458 Evolution 53, 1143–1156. (doi:10.2307/2640818)
- 42. Falconer DS, Mackay TFC. 1996 Introduction to quantitative genetics. 4th edn. Pearson
 Education India.
- 461 43. Endler JA. 1986 Natural selection in the wild. Princeton University Press.
- 462 44. Kingsolver JG, Hoekstra HE, Hoekstra JM, Berrigan D, Vignieri SN, Hill CE, Hoang A,
 463 Gibert P, Beerli P. 2001 The Strength of Phenotypic Selection in Natural Populations. The
 464 American Naturalist **157**, 245–261. (doi:10.1086/319193)
- 465 45. Grant PR, Grant BR, Huey RB, Johnson MTJ, Knoll AH, Schmitt J. 2017 Evolution caused 466 by extreme events. Phil. Trans. R. Soc. B **372**, 20160146. (doi:10.1098/rstb.2016.0146)
- 467 46. Rodríguez-Trelles F, Tarrío R, Santos M. 2013 Genome-wide evolutionary response to a 468 heat wave in Drosophila. Biol. Lett. **9**, 20130228. (doi:10.1098/rsbl.2013.0228)
- 469 47. MacMillan HA. 2019 Dissecting cause from consequence: a systematic approach to 470 thermal limits. Journal of Experimental Biology **222**, jeb191593.
- 48. Lecheta MC et al. 2020 Integrating GWAS and Transcriptomics to Identify the Molecular
 Underpinnings of Thermal Stress Responses in Drosophila melanogaster. Frontiers in
 Genetics 11, 658. (doi:10.3389/fgene.2020.00658)
- 474 49. Rolandi C, Lighton JRB, de la Vega GJ, Schilman PE, Mensch J. 2018 Genetic variation
 475 for tolerance to high temperatures in a population of Drosophila melanogaster. Ecology
 476 and Evolution 8, 10374–10383. (doi:10.1002/ece3.4409)
- 50. Kellermann V, Overgaard J, Hoffmann AA, Fløjgaard C, Svenning J-C, Loeschcke V. 2012
 Upper thermal limits of Drosophila are linked to species distributions and strongly
 constrained phylogenetically. Proceedings of the National Academy of Sciences 109,
 16228–16233.

- 481 51. Leiva FP, Calosi P, Verberk WCEP. 2019 Scaling of thermal tolerance with body mass
 482 and genome size in ectotherms: A comparison between water-and air-breathers.
 483 Philosophical Transactions of the Royal Society B: Biological Sciences **374**, 20190035.
 484 (doi:10.1098/rstb.2019.0035)
- 485 52. Rezende EL, Bozinovic F, Szilágyi A, Santos M. 2020 Predicting temperature mortality 486 and selection in natural Drosophila populations. Science **369**, 1242–1245.
- 487 53. Jørgensen LB, Malte H, Overgaard J. 2019 How to assess Drosophila heat tolerance:
 488 Unifying static and dynamic tolerance assays to predict heat distribution limits. Functional
 489 Ecology 33, 629–642. (doi:10.1111/1365-2435.132)
- 490 54. Jørgensen LB, Malte H, Ørsted M, Klahn NA, Overgaard J. 2021 A unifying model to
 491 estimate thermal tolerance limits in ectotherms across static, dynamic and fluctuating
 492 exposures to thermal stress. Scientific reports **11**, 1–14.
- 493 55. Huey RB, Kearney MR. 2020 Dynamics of death by heat. Science 369, 1163–1163.
 494 (doi:10.1126/science.abe0320)
- 495 56. Leiva FP, Santos M, Niklitschek EJ, Rezende EL, Verberk WCEP. 2024 Paper data and
 496 code for: Genetic variation of heat tolerance in a model ectotherm: an approach using
 497 thermal death time curves. Zenodo. (doi:https://doi.org/10.5281/zenodo.12155988)