

1 **MetaR, a global database on metabolic rates of ectotherms**

3 **Authors**

4 Félix P. Leiva^{1,2} (ORCID ID: 0000-0003-0249-9274; felixpleiva@gmail.com)

5 Wilco C.E.P. Verberk² (ORCID ID: 0000-0002-0691-583X; w.verberk@science.ru.nl)

6 Piero Calosi³ (ORCID ID: 0000-0003-3378-2603; piero_calosi@uqar.ca)

7 Enrico L. Rezende⁴ (ORCID ID: 0000-0002-6245-9605; enrico.rezende@uc.cl)

8 Felix C. Mark¹ (ORCID ID: 0000-0002-5586-6704; Felix.Christopher.Mark@awi.de)

10 **Affiliations**

11 1. Department of Integrative Ecophysiology, Alfred Wegener Institute Helmholtz Centre for
12 Polar and Marine Research (AWI), 27570, Bremerhaven, Germany

13 2. Department of Animal Ecology and Physiology, Radboud Institute for Biological and
14 Environmental Sciences, Radboud University Nijmegen, Nijmegen, The Netherlands

15 3. Laboratoire de Physiologie Écologique et Évolutive Marine, Département de Biologie,
16 Chimie et Géographie, Université du Québec à Rimouski, Rimouski, Québec, Canada

17 4. Departamento de Ecología, Facultad de Ciencias Biológicas, Center of Applied Ecology
18 and Sustainability (CAPES), Pontificia Universidad Católica de Chile, Santiago, Chile

19
20 corresponding author: Félix P. Leiva (felixpleiva@gmail.com)

22 **Abstract**

23 Whole-organism metabolic rate is a key trait for understanding ectotherms' responses to
24 ongoing environmental change. It represents the interface through which organisms interact
25 with their environment and therefore allows for making predictions across various levels of
26 biological organisation. While much of the variation in metabolic rates is explained by body
27 size and temperature, a considerable part of this variation remains unexplained. Lack of
28 standard research practices, data sparsity and insufficient coverage of various taxa limit our
29 capacity to conduct a meaningful synthesis across the Tree of Life; both in the spatial and the
30 temporal dimension. To overcome these limitations, and acquire a better understanding of
31 the evolution of metabolic rates, we created MetaR: which is to date the most comprehensive
32 database on intra- and interspecific variations in ectotherms' metabolic rates. This database
33 currently comprises over 95,000 records covering more than 2,400 species across 16 phyla of
34 ectotherms, encompassing both invertebrates and small and large vertebrates from marine,
35 intertidal, freshwater and terrestrial ecosystems. MetaR also integrates methodological
36 details, which further improves our capacity, on the one hand, to detect ecological,
37 physiological and evolutionary patterns and, on the other hand, to forecast the functional
38 impacts of global environmental changes across the Tree of Life from a metabolic perspective.

40 **Background & Summary**

41 Initiatives developed for centralising and standardising the large body of literature on
42 functional traits offer great potential to achieve a broader understanding of ecological and
43 evolutionary mechanisms occurring in nature¹⁻⁴. One critical aspect is the necessity for an
44 interface facilitating the interaction between organisms and their environment, which is vital
45 for comprehending the repercussions of global environmental changes across various levels
46 of biological organisation, encompassing both aquatic and terrestrial ecosystems. In this
47 sense, whole-organism metabolic rates – usually measured through oxygen uptake, reflect the
48 energy requirements of metabolic processes associated with ATP production during aerobic
49 respiration⁵⁻⁷. The metabolic rate of an organism is a key functional trait in physiology, ecology
50 and evolution for understanding ectotherms' responses to ongoing environmental change,
51 and it has important fitness implications for organisms and demography⁸⁻¹⁰. Metabolic rates

52 govern the pace of life; i.e. the rate at which each organism processes energy and materials to
53 grow, reproduce, live and move. Using this fundamental trait therefore allows us to predict
54 most observed ecological processes at the level of organisms, populations, communities and
55 ecosystems^{9,11,12}.

56 Body size and body temperature are tightly related to metabolic rates and therefore,
57 not surprisingly, these are two key parameters of the *Metabolic Theory of Ecology* (MTE)⁹. MTE
58 proposes that metabolic rates vary with body temperature and it scales with body mass
59 according to a power relationship with an exponent of around three-quarters⁹. Although the
60 universality of the MTE predictions is debated¹³⁻¹⁸, the fact remains that most of the variation
61 in metabolic rates can be explained by differences in body mass and temperature^{9,19}. However,
62 even after accounting for these two factors, a considerable amount of intra-individual, and
63 inter- and intraspecific variation remains unexplained. For instance, there is substantial
64 variation (often ~2-fold, on a log₁₀ scale) across spatial and temporal scales²⁰, and
65 developmental stages²¹, lifestyle and activity levels²². The next natural step is to explore which
66 other traits, together with body size and body temperature, can explain this variation.
67 Reconciling the different views of this debate is, however, a serious scientific challenge, that
68 has been exacerbated by the lack of standard research practices, resulting in methodological
69 differences among studies, data sparsity, and insufficient coverage of metabolic data across
70 the Tree of Life. This in turn has limited the establishment of a meaningful synthesis to date⁵.

71 To overcome these limitations, we have created MetaR, the largest database focusing
72 on the metabolic rates of metazoans, with the scope to unveil eco-physio-evolutionary
73 patterns undescribed to date and investigate the impacts of global environmental changes on
74 this central physiological trait. We assembled and curated a large, intra- and interspecific
75 database on metabolic rates of ectotherms (invertebrates, fishes, amphibians and reptiles),
76 globally distributed across all realms (marine, freshwater, intertidal and terrestrial), climate
77 zones (polar, temperate and tropical), and from the deepest ocean to the highest
78 mountains.

79

80 **Methods**

81 In reporting and describing our methods, we adhere to the MeRIT guidelines outlined by
82 Nakagawa et al.²³ for enhanced clarity and transparency.

83

84 **Literature search.** Félix P. Leiva (FPL) searched published literature for studies quantifying
85 metabolic rates in invertebrates and vertebrates using various searching methods. We could
86 not follow a step-by-step Preferred Reporting Items for Systematic Reviews and Meta-
87 Analyses (PRISMA) approach²⁴ due to barriers to accessing data associated with the use of
88 search engines such as ISI Web of Science and Scopus at the time FPL initiated this project
89 (year 2007). Despite the mentioned limitations, the current version of MetaR far exceeds any
90 other database on this topic in terms of the number of records, metadata, number of articles,
91 and species included, as well as the spatial and temporal coverage of the data available in the
92 literature to date.

93 From 2007 to 2016, focusing specifically on invertebrates, FPL used Google Scholar
94 regularly to search for primary articles reporting metabolic rates by using the following
95 keyword combinations: (metabol* OR oxygen consumption OR metabolic rate OR respiration)
96 AND (acidification OR hypoxia OR salinity OR temperature OR scaling) AND (clade names). For
97 clade names, the following terms were employed: "invertebrates" OR "coral" OR "arthropod*"
98 OR "insect*" OR "crustacea*" OR "arach*" OR "echinoderm*" OR "molluscs" OR "gastropod".
99 Measuring oxygen uptake as a proxy for metabolic rates is considered an informative
100 physiological trait for assessing whole-organism performance in response to environmental
101 stressors^{25,26}. Therefore, the terms acidification, hypoxia, salinity, and temperature were
102 included in the search due to their potential to cause reductions in the performance or fitness
103 of organisms, especially in the context of global climate change^{27,28}. The literature search

104 resulted in >5,000 publications. After a thorough examination of their abstracts and titles,
105 1,253 full-text articles were downloaded for a more detailed screening.

106 On the 14 of March 2021, FPL used ISI Web of Science to search for literature on
107 amphibians, reptiles and fishes using a similar combination of Boolean terms as for
108 invertebrates. For amphibians, the following combination was used: (metabol* OR oxygen
109 consumption OR metabolic rate OR respiration) AND (amphibia* or salamander OR caecilian
110 OR siren OR newt* OR frog* OR toad* OR tadpoles). For reptiles the following combination:
111 (metabol* OR oxygen consumption OR metabolic rate OR respiration) AND (reptil* OR
112 crocodile* OR alligator* OR tuatara* OR lizard* OR snake* OR turtle*). For fishes the following
113 combination: (metabol* OR oxygen consumption OR metabolic rate OR respiration) AND
114 (fish*). The search was refined by focusing on specific categories such as zoology, physiology,
115 biology, ecology, biochemistry molecular biology, evolutionary biology, multidisciplinary
116 sciences, environmental sciences, marine and freshwater biology, toxicology, genetics
117 heredity, veterinary sciences, behavioural sciences, endocrinology metabolism, fisheries,
118 biodiversity conservation, respiratory system, and oceanography. The search spanned from
119 1945 to 2020 and included only primary references. Using this approach, 652 primary
120 references were obtained for amphibians, 1,265 for reptiles, and 5,510 for fishes.

121 Additionally, in an attempt to supplement this search, FPL compiled references from
122 meta-analyses²⁹⁻³³, comparative approaches^{8,34-41}, books⁴²⁻⁴⁵, and literature reviews
123 conducted on various aquatic-living taxa, including crustaceans and molluscs, among others⁴⁶⁻
124 ⁴⁸. FPL also utilized the list of references of articles that have conducted previous compilations
125 on metabolic scaling^{22,40,49,50}. In addition, on June 12, 2020, a list with a total of 348 references
126 was downloaded from FishBase⁵¹. Many of these references were not possible to access or
127 were not available digitally. Since last verified (December 2023), FishBase had not
128 incorporated additional data on metabolic rates in fishes.

129 Until January 2024, FPL contacted the corresponding authors requesting additional
130 data related to the estimation of metabolic rates for some articles published between 2001
131 and 2022, in which it was not possible to extract the data adequately from the figures. In total,
132 13 researchers responded positively (out of the 62 researchers we contacted) to this request,
133 providing their data. Their names are listed in the acknowledgements section.

134 All references were imported into Zotero for better management, and the existence
135 of duplicates was checked using the same open-source software. The complete list of
136 literature used in the current version of MetaR is publicly supporting material in the form of a
137 *.ris file (Supplementary Information).

138
139 **Structure of MetaR.** MetaR is a relational database built in Microsoft Access using nested
140 tables. Currently, the database consists of four tables that provide details on the reference,
141 taxonomy, origin, and oxygen uptake of ectotherms. The relational database, maintained and
142 curated for internal control, is shown in the supplementary information and the GitHub
143 repository (https://github.com/felixpleiva/MetaR/blob/main/DB_relationships.md). The data
144 structure undergoes a transformation into a spreadsheet file through a query, encompassing
145 an extensive table of trait records and metadata measured at the individual level, as much as
146 possible.

147
148 **Data extraction.** Combining all the searching methods, FPL screened > 12,775 references,
149 which included primary published, peer-reviewed scientific studies, as well as grey literature,
150 theses, and books. A study was included in our database if the metabolic rate was measured
151 at the individual level in any subspecies, species, or genus of metazoan. Metabolic rates data
152 from various specimens, which were aggregated, were not incorporated into the current
153 database. We attempted to consider the primary reference reporting the data as much as
154 possible. Hence, review articles were also excluded if the data were reported in a previous

155 study. Applying these criteria, we obtained a total of 2,145 papers considered for data
156 extraction.

157 We followed the classification given by the authors for life stages and type of
158 metabolic rate. Rates of oxygen uptake were categorized as routine (RMR), resting, standard
159 (SMR), maximal (MMR), or active based on the terminology used in the original publications,
160 or classified by the definitions proposed by Chabot et al.⁵² and Brett⁵³. We made several
161 assumptions in our data collection process. When a study reported only body mass or only
162 metabolic rates as an interval, we calculated a mean value based on that interval. If the papers
163 did not specify the type of body mass used (e.g., fresh mass, dry mass, ash-free dry mass) or
164 did not provide procedures for drying the animals, we assumed that the reported mass was
165 fresh mass. Whenever possible, we included this information in the comments. In studies
166 where the authors mentioned the use of freshwater, tap water, or dechlorinated water during
167 the conditions of maintenance, rearing, acclimation, and testing, we assumed a salinity of 0
168 for all these conditions. Important to mention is that the data points we used represent actual
169 observations and not estimates generated by linear models unless otherwise specified in the
170 legend of figures of the primary references. For instance, some studies included partial
171 residuals for metabolic rate data, and in such cases, those studies were not included in our
172 database. In cases where metabolic rate data were presented graphically but no information
173 was reported in the text or a table, we used the Java program Plot Digitizer, designed for
174 extracting X-Y coordinates from graphs (<http://plotdigitizer.sourceforge.net>).
175

176 **Geographical position, depth and elevation of the collection.** We harmonized the
177 geographical positions from where the species were collected using the World Geodetic
178 System (WGS84) as the reference geographical coordinate system. The geographic
179 coordinates were converted, if necessary, to decimal degrees, both for latitude and longitude.
180 In various cases, numerous researchers collected animals from different geographical
181 locations, elevations and depths. Despite these specific locations, elevations and depths being
182 explicitly mentioned in the paper, in some articles the authors chose to present the data as a
183 combined dataset, regardless of the various spatial scales at which the data was collected. In
184 these cases, we calculated the mean geographical coordinates, as well as the depth and
185 elevation concerning their respective country, region, province, county, city or locality, as
186 given in the original data sources. For those species for which we could not obtain geographical
187 coordinates from the paper, we used the description of the locality or the geographical
188 reference point mentioned by the authors in the paper. We inspected the spatial distribution
189 of the sites visually and then manually georeferenced them using Google Earth
190 (<https://www.google.com/earth/>) or GeoNames (<https://www.geonames.org/>) when the
191 reference point was clearly mentioned (e.g., Isla Chiloé). The details of which approximations
192 were used are described in the respective comments column of the database. When
193 specimens were collected from different locations and mixed during experiments, we chose
194 the central geographical reference within these locations. However, when the locality was
195 treated as a factor distinguishable in the article or its figures or tables, we included these
196 entries independently to make the distinction. As part of a forward search, additional data
197 were sought in case reference was made to another paper describing, for example, animal
198 collection sites or methodological aspects. In such cases, the additional paper was downloaded
199 and used to complement the missing information.
200

201 **Taxonomy and phylogeny.** FPL verified the scientific names of species in the database by
202 manually cross-referencing them with the Open Tree of Life^{54,55}. Subsequently, we employed
203 the same taxonomic harmonization procedure outlined by Lenoir et al.⁵⁶. This taxonomic
204 harmonization process consists of three steps: (1) searching for names in the National Center
205 for Biotechnology Information (NCBI) taxonomy database, (2) confirming any taxonomic
206 entities not found in NCBI using the Integrated Taxonomic Information System (ITIS) database,

207 and (3) checking the remaining entities not found in NCBI and ITIS against the Global
208 Biodiversity Information Facility (GBIF) database. In cases where a match was identified, the
209 corrected taxonomic entity underwent the entire procedure again, including verification in
210 NCBI and ITIS, to ensure a reliable taxonomic classification. Ultimately, only names at the
211 species and genus levels were retained for the database, with subspecies aggregated at the
212 species level. The R packages "rgbif v3.7.8"⁵⁷ and "taxize v0.9.98"⁵⁸ were utilized for this
213 harmonization procedure.

214 Before gathering information about phylogenetic relationships, the most recent
215 version of our database, as of February 2024, encompassed 2,459 entries representing taxa
216 up to the species and subspecies level. The phylogenetic tree retrieved from the OTL
217 underwent pruning to retain only those species present in our database using the package
218 "rotl"⁵⁹. Entries lacking phylogenetic information or flagged as "incertae_sedis_inherited",
219 "extinct, incertae_sedis, sibling_higher", "hidden" and "unplaced_inherited" were excluded.
220 Consequently, the final count of species in the current version (v1.0.0) of MetaR has been
221 streamlined to 2,287 distinct species, each accompanied by phylogenetic information (Fig. 3).

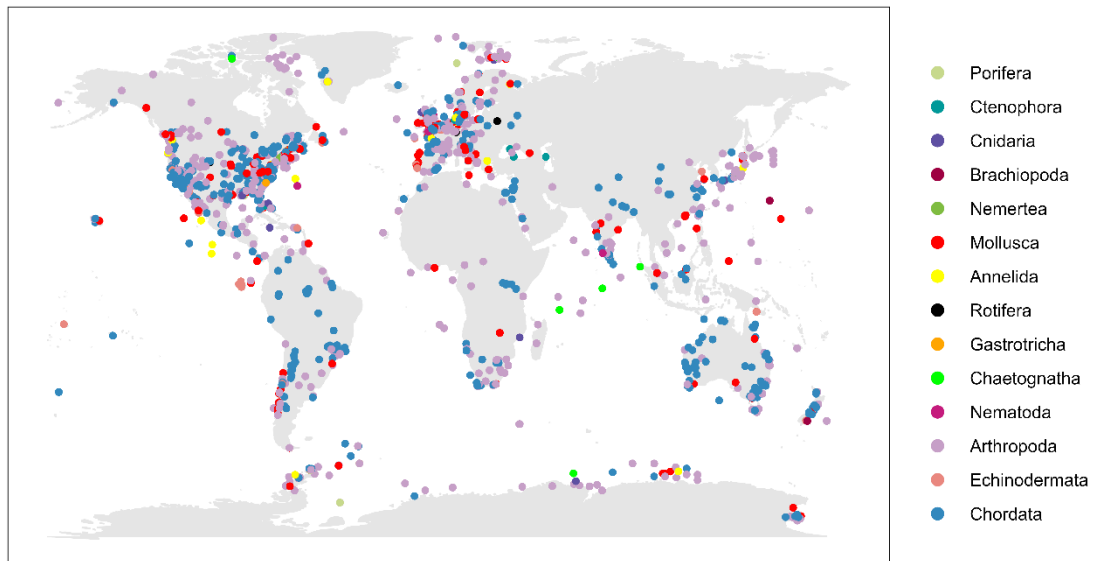
222

223 **Data Records**

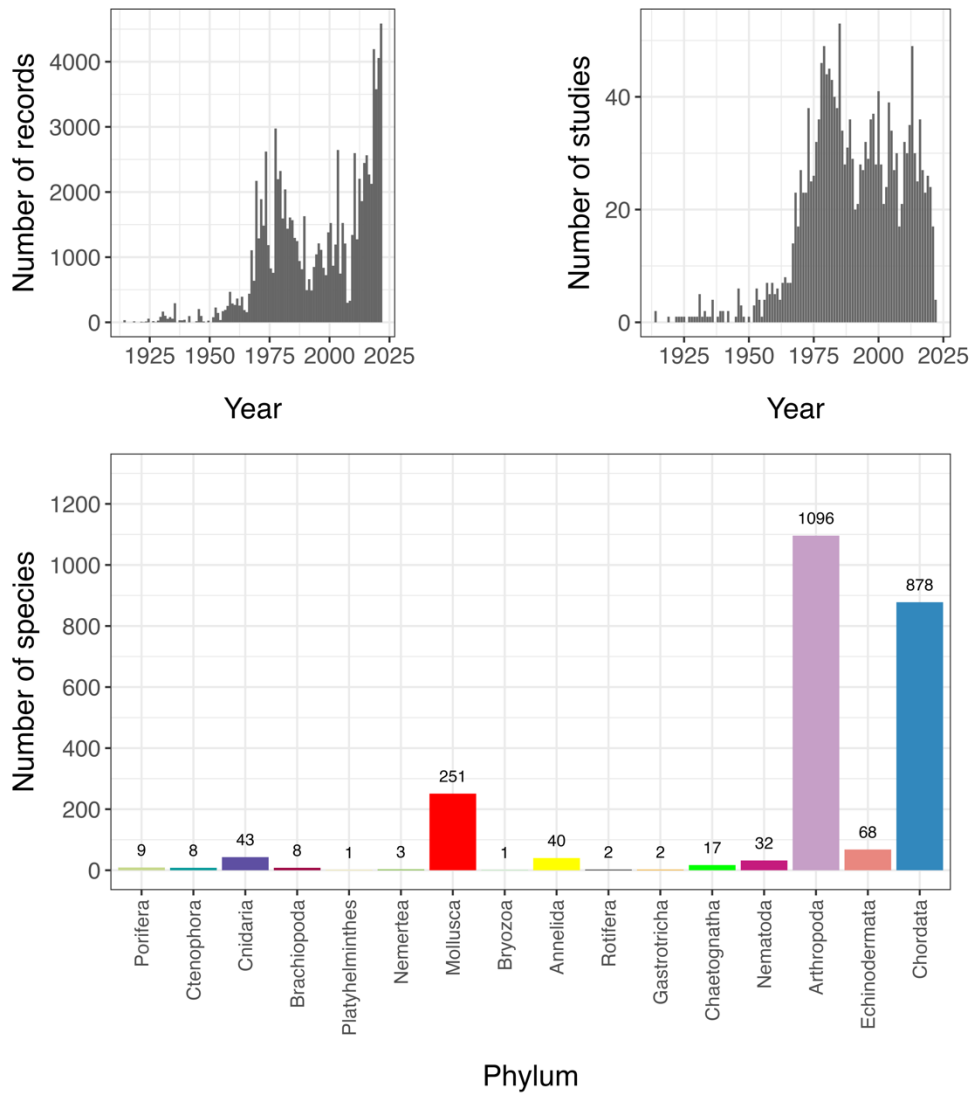
224 MetaR includes 95,373 records on metabolic rates for 2,459 species across 16 phyla, 58
225 classes, 220 orders, 780 families and 1,623 genera, ranging from invertebrates to small and
226 large vertebrates from marine, intertidal, freshwater, and terrestrial realms, extracted from
227 primary references published between 1914 to 2022 (Figs. 1-3). Based on studies that reported
228 geographical positions (N = 1,345), most of the studies (> 75 %) incorporated in our database
229 were conducted in the North Hemisphere (1,020 articles, 1,557 species, 52,130 records)
230 compared to the South Hemisphere (343 articles, 566 species, 22,081 records) (Fig. 1).

231 For each paper, we include up to 95 characteristics associated with metabolic rates,
232 when available. Taxonomically speaking, Arthropoda, Chordata, Mollusca, Echinodermata,
233 Cnidaria, Annelida, Nematoda and Chaetognatha are the main phyla present in this version of
234 MetaR. They contain, respectively, 0.10%, 1.20%, 0.29%, 0.90%, 0.32%, 0.24%, 0.17% and
235 12.88% of the species described on Earth (<https://www.catalogueoflife.org/>, accessed on the
236 22 of March 2024 and considering accepted species only). On the other hand, the phyla
237 Porifera, Ctenophora, Brachiopoda, Nemertea, Rotifera and Gastrotricha, and to a greater
238 extent, Platyhelminthes and Bryozoa, are underrepresented (Fig. 3). Records on metabolic
239 rates included in this version of MetaR show a range of variation from -7.032 to 4.621 mg O₂
240 h⁻¹ ind⁻¹, equivalent to 11.6 orders of magnitude (in log₁₀ scale). The body temperatures at
241 which these rates were measured ranged from -5°C to 47°C. On the other hand, the smallest
242 species included in this version corresponds to the nematode *Plectus palustris*, weighing 0.06
243 µg of fresh mass, while the largest species corresponds to the green sea turtle *Chelonia mydas*,
244 weighing 142 kg of fresh mass (Fig. 4). All these records were retrieved from more than 1,800
245 primary references.

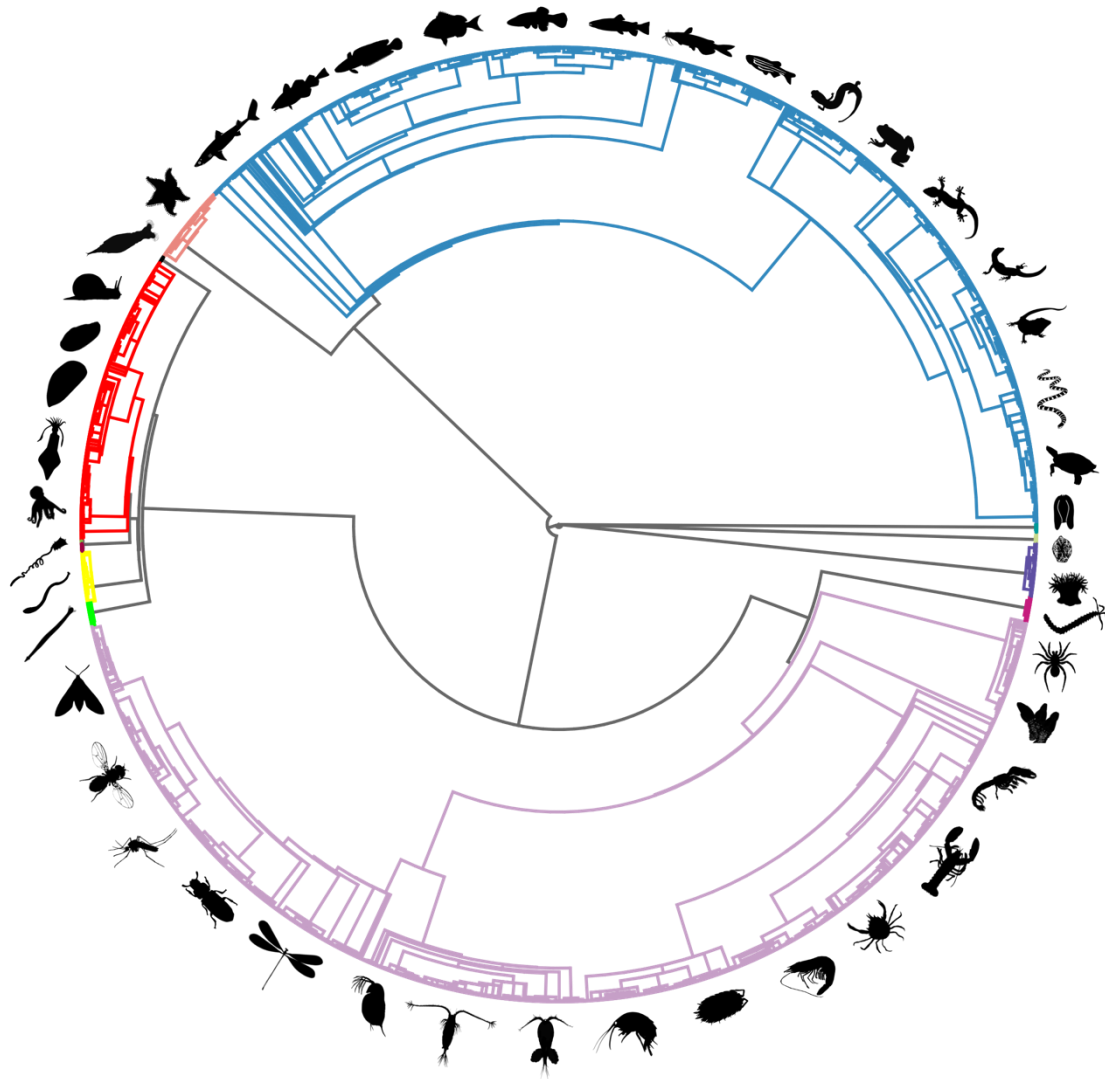
246 We foresee that MetaR will make a substantial scientific contribution by shedding light
247 on the longstanding question of what influences variation in metabolic rates. Despite the
248 widespread coverage, our database reveals evident geographic and taxonomic biases.
249 Notably, Russia, northern Canada, northern Africa, and the tropics lack studies on metabolic
250 rates. On another hand, except Chaetognatha, all other phyla included here are
251 underrepresented, and none of them exceeds 1.2% of the total number of species *per* phyla.
252 Our forthcoming step entails conducting a quantitative and qualitative analysis to map and
253 identify existing knowledge gaps. This analysis will establish a basis for addressing future
254 research questions utilizing the most comprehensive dataset on metabolic rates over the last
255 century.



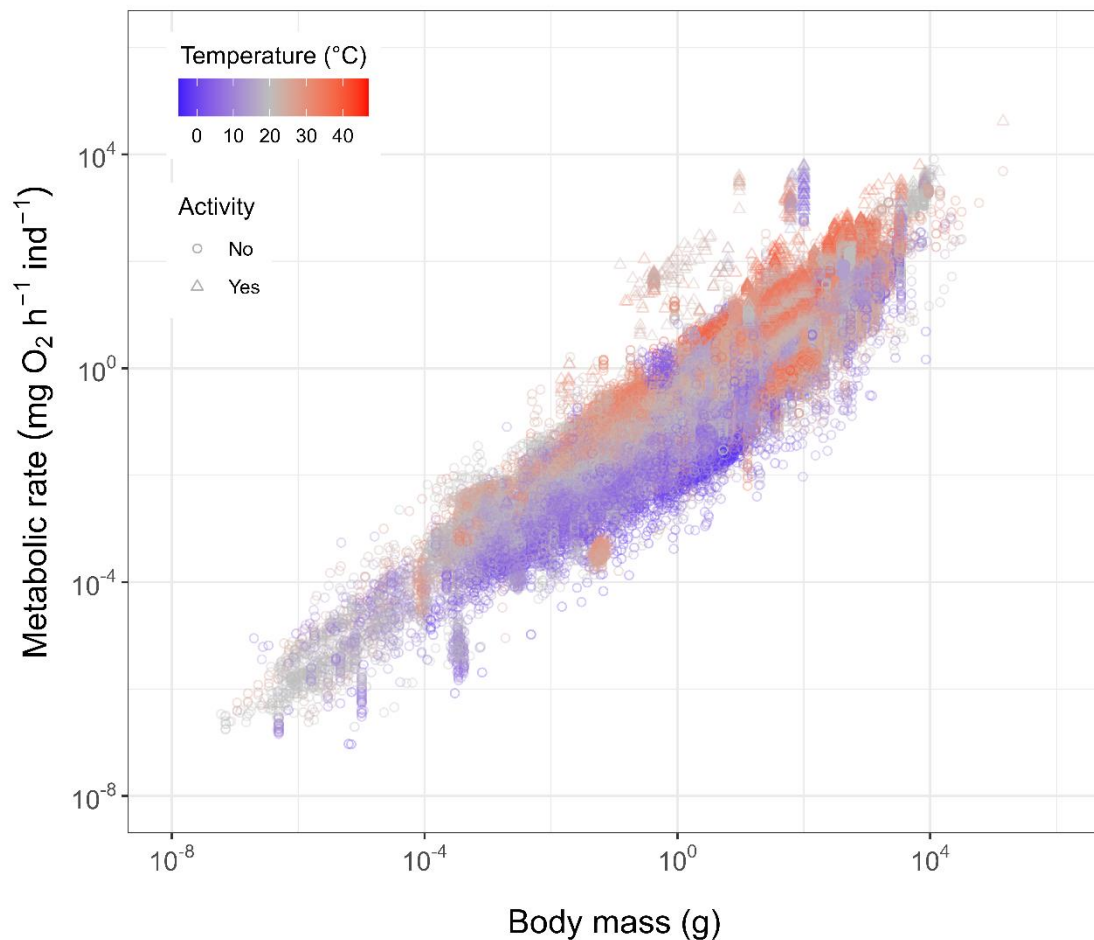
256
 257 **Figure 1.** Map illustrating the geographical locations where ectotherms species were collected.
 258 Colours define different phyla included in MetaR. The data represent 1,577 unique
 259 geographical positions reported in 1,345 studies.
 260



261
 262 **Figure 2.** (a) Total number of records, (b) studies along the years (from 1914 to 2022) and (c)
 263 number of species by Phylum included in MetaR. The colour-coded bars match the ones used
 264 in Figure 1 to represent different phyla.
 265



266
 267 **Figure 3.** Phylogenetic tree showing the relationships among the 2,287 ectotherms species
 268 included in the current version of MetaR and for which we have phylogenetic information from
 269 the Open Tree of Life. Dots are colour-coded to match the ones used in previous figures to
 270 represent different phyla. The tree is plotted using the R package "*phytools*"⁶⁰. Silhouettes are
 271 available for reuse on PhyloPic (www.phylopic.org) under the Public Domain Dedication 1.0
 272 license.
 273



274
 275 **Figure 4.** Scaling of metabolic rates with body mass using data gathered from ectotherms
 276 species across various geographical locations and realms. Measurements were conducted
 277 under diverse experimental conditions, encompassing factors such as temperature, oxygen
 278 levels, salinity, pH, CO₂, photoperiod, humidity, and food type. Additionally, observations were
 279 recorded across different sexes, life stages, and activity levels, including standard, routine,
 280 resting, minimum, and basal metabolic rates, as well as active, field, flight, maximum, and
 281 swimming activities. Coloured points represent the measurement temperature range from -
 282 5°C to 47°C, illustrating the breadth of metabolic rate variation captured in MetaR.
 283

284 **Technical Validation**

285 We have established a procedure to examine inconsistencies in our database, covering both
 286 discrete and continuous variables. We methodically detect, assess, and rectify potential
 287 inconsistencies and deviations in metabolic rate data. Specifically, these inconsistencies and
 288 deviations were evaluated in the well-known biological pattern of allometric scaling. FPL fitted
 289 a log₁₀-log₁₀ ordinary least square model between body size and metabolic rate and then
 290 calculated z-scores for the residuals of this model. These scores measure the extent of
 291 deviations from the mean, facilitating the identification of potential outliers or significant
 292 variations within the dataset based on a predefined, random threshold of 2.5. Once potential
 293 outliers were identified, the corresponding entries were examined, and any record that
 294 seemed inconsistent was rechecked against the source. Other steps included verifying the
 295 accuracy of the names of discrete variables and their units. All of these steps and potential
 296 corrections were implemented before the release of this database (v1.0.0).
 297

298 **Usage Notes**

299 The most recent versions of the MetaR database will be accessed and downloaded from the
300 GitHub repository (<https://github.com/felixpleiva/MetaR>). When citing MetaR in a
301 manuscript, users should indicate the exact version they used, together with this data paper.
302 Explicitly specifying the version is crucial when utilizing the data to guarantee the
303 reproducibility of any subsequent analyses and, if possible, citing the primary references gives
304 credit to the original publications used to build this database.

305 We provide MetaR with several metadata related to the experimental determination
306 of metabolic rates. This allows users to incorporate such information to test the robustness of
307 conclusions emerging from their analyses, but also to filter data based on their specific
308 requirements. Given the versatility of the structure of our database, as well as the variety of
309 metadata present in it, we have included many columns of information regarding
310 maintenance, rearing, acclimation, and testing conditions. Many of the studies manipulated
311 single or multiple combinations of environmental factors, such as temperature, oxygen,
312 salinity, carbon dioxide, pH, humidity, photoperiod or measuring different types of metabolic
313 rates at different life stages and sexes. MetaR contains all these data, which means users
314 should pay special attention to filtering the data of interest that allows them to address their
315 specific questions. Some studies use the term "acclimation" to refer to various aspects. For
316 example, it may denote the maintenance of animals in the laboratory, exposure to
317 environmental conditions before experiments, or during acclimation to the respirometry
318 chamber. Special attention was given to avoid misinterpretations of the experimental design
319 by incorporating such information into the corresponding column. For each paper included in
320 MetaR, there is, for instance, a column designated for maintenance temperature, rearing
321 temperature, acclimation temperature, and testing temperature. This applies to all other
322 environmental factors, such as oxygen, salinity, carbon dioxide, pH, humidity, photoperiod and
323 food type. Important to note is that many of these columns correspond to an expansion of the
324 guidelines provided by Killen et al. al.⁶¹ for aquatic respirometry, which is also applicable to
325 terrestrial respirometry. In addition, many studies simply report the means in tables or figures.
326 When this was the case, we also took the sample size and the associated error for the
327 respective measure of central tendency so that it could be used for meta-analytic approaches.
328 In an effort to help MetaR users filter and download a subset of data of interest, we have
329 developed an R script for this purpose. This R code is customizable, including options to
330 calculate mean values and to select data based on taxonomic entities and relevant covariables,
331 such as swimming speed, which could in addition serve as a metric for studies in thermal
332 performance. Additionally, the R code allows the inclusion of primary references that, as much
333 as possible, should be cited when appropriate.

334 Initially, all units of metabolic rates were converted to $\text{mg O}_2 \text{ h}^{-1} \text{ ind}^{-1}$ (milligram of
335 oxygen *per hour per individual*). To increase the reuse of MetaR and facilitate comparisons,
336 we also transformed metabolic rates from $\text{mg O}_2 \text{ h}^{-1} \text{ ind}^{-1}$ into units of concentration (mmol O_2
337 $\text{h}^{-1} \text{ ind}^{-1}$), volume ($\text{mL O}_2 \text{ h}^{-1} \text{ ind}^{-1}$) and energy (Joules). We attempted to include rates of oxygen
338 uptake (O_2) as much as possible. However, in cases where only rates of carbon dioxide
339 production (CO_2) were measured, these were converted to rates of O_2 uptake by dividing the
340 CO_2 produced by 0.85, which represents a respiratory exchange ratio based on the
341 intermediate value between the use of lipids (0.7) and carbohydrates (1) as an energy
342 substrate⁶. All body sizes were converted to fresh body mass in grams using appropriate
343 conversion factors^{8,62-66}.

344

345 **Code Availability**

346 All materials, including the database, R code, and supplementary content, are licensed under
347 the Attribution-NonCommercial-NoDerivatives 4.0 International license (CC BY-NC-ND 4.0).
348 This license requires that reusers give credit to the creator. It allows reusers to copy and
349 distribute the material in any medium or format in unadapted form and for noncommercial
350 purposes only. The MetaR project is hosted and maintained on a GitHub repository

351 (<https://github.com/felixpleiva/MetaR>) and will be assigned a DOI on Zenodo. This repository
352 comprises files containing: 1) the database, 2) metadata, 3) a list of references and 4) a
353 customizable R code. MetaR is under embargo until the 30 of June 2025 but will become open
354 to the community afterwards, and users must refer to this data paper when utilizing the
355 resource as well as the repository. We strongly suggest, if possible, citing the original articles
356 that contributed a substantial proportion to the database. We also would appreciate it if users
357 would bring to our attention errors or ambiguous values found in the database. MetaR will be
358 updated as needed with new studies, as well as any identified errors will be corrected
359 following the standard of the Semantic Versioning Specification (SemVer,
360 <https://semver.org/>).

361

362 **Acknowledgements**

363 Félix P. Leiva acknowledges financial support by the Alexander von Humboldt Foundation.
364 Piero Calosi acknowledges the support of the Natural Sciences and Engineering Research
365 Council of Canada (NSERC) Discovery programme (RGPIN-2020-05627), and his membership
366 to the FRQNT-funded research network Québec-Ocean and Ressources Aquatiques Québec.
367 Felix C. Mark acknowledges funding from the ACTNOW (HORIZON-CL6-2021-BIODIV-01
368 #101060072) and YESSS (BMBF FKZ03F0946C) projects.

369 Félix P. Leiva appreciates the work done by many researchers over the past century
370 on aspects related to the respiratory physiology of invertebrates and vertebrates. In particular,
371 the he expresses gratitude to those who provided additional information and data upon
372 request: Alexandra Steckbauer (King Abdullah University of Science and Technology, Saudi
373 Arabia), Isabel Silva Romero (University of North Carolina at Chapel Hill, USA), Fernando Rafael
374 De Grande (Universidade Federal de São Paulo, Brazil), Christophe Piscart (Université de
375 Rennes 1, France), Gisela Lannig (Alfred Wegener Institute, Germany), Daniel Lemos
376 (Universidade de São Paulo, Brazil), Katharina Ruthsatz (Technische Universitaet
377 Braunschweig, Germany), Nelly Tremblay (Université du Québec à Rimouski, Canada), Ummat
378 Somjee (Smithsonian Tropical Research Institute, Panama), Lumír Gvoždík (Czech Academy of
379 Sciences, Czech Republic), Kristjan Niitepõld (The Finnish Science Centre Heureka, Finland),
380 Margarita Brandt (Universidad San Francisco de Quito, Ecuador) and Sean Giery (Ohio
381 University, USA).

382 Finally, this paper is dedicated to the memory of Doris Abele (June 22, 1957–
383 November 21, 2021), who provided significant support to Félix P. Leiva during his funding
384 application.

385

386 **Author contributions**

387 Félix P. Leiva conceptualised the idea for this study and led all aspects related to the creation
388 and implementation of MetaR over the last 17 years. Specifically, he is responsible for creating
389 the structure of the database and ensuring its referential integrity, data entry and data
390 curation and has led the writing of the manuscript. Félix P. Leiva created all figures with inputs
391 from Wilco C.E.P. Verberk, Piero Calosi, Enrico L. Rezende and Felix C. Mark. All authors edited
392 the manuscript and gave their approval for publication.

393

394 **Competing interests**

395 The authors declare no competing financial interests.

396

397 **References**

- 398 1. Leiva, F. P., Calosi, P. & Verberk, W. C. E. P. Scaling of thermal tolerance with body mass
399 and genome size in ectotherms: A comparison between water-and air-breathers.
400 *Philosophical Transactions of the Royal Society B: Biological Sciences* **374**, 20190035
401 (2019).
- 402 2. Sunday, J. M. *et al.* Thermal tolerance patterns across latitude and elevation. *Philosophical*
403 *Transactions of the Royal Society B: Biological Sciences* **374**, (2019).
- 404 3. Verberk, W. C. E. P. *et al.* Body mass and cell size shape the tolerance of fishes to low
405 oxygen in a temperature-dependent manner. *Global Change Biology* **28**, 5695–5707
406 (2022).
- 407 4. Bennett, J. M. *et al.* GlobTherm, a global database on thermal tolerances for aquatic and
408 terrestrial organisms. *Scientific Data* **5**, 180022 (2018).
- 409 5. Metcalfe, N. B. *et al.* Solving the conundrum of intra-specific variation in metabolic rate:
410 A multidisciplinary conceptual and methodological toolkit: New technical developments
411 are opening the door to an understanding of why metabolic rate varies among individual
412 animals of a species. *BioEssays* **45**, 2300026 (2023).
- 413 6. Schmidt-Nielsen, K. *Animal Physiology: Adaptation and Environment*. (Cambridge
414 University Press, New York, USA, 1997).
- 415 7. Clarke, A. *Principles of Thermal Ecology: Temperature, Energy and Life*. (Oxford University
416 Press, Oxford and New York, 2017).
- 417 8. Brey, T. An empirical model for estimating aquatic invertebrate respiration. *Methods in*
418 *Ecology and Evolution* **1**, 92–101 (2010).
- 419 9. Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M. & West, G. B. Toward a metabolic
420 theory of ecology. *Ecology* **85**, 1771–1789 (2004).
- 421 10. White, C. R. & Kearney, M. R. Metabolic scaling in animals: methods, empirical results, and
422 theoretical explanations. *Comprehensive Physiology* (2014).
- 423 11. Enquist, B. J. *et al.* Scaling metabolism from organisms to ecosystems. *Nature* **423**, 639–
424 642 (2003).
- 425 12. West, G. B., Brown, J. H. & Enquist, B. J. A general model for the origin of allometric scaling
426 laws in biology. *Science* **276**, 122–126 (1997).
- 427 13. White, C. R. There is no single p. *Nature* **464**, 691–693 (2010).
- 428 14. Kozłowski, J. & Konarzewski, M. West, Brown and Enquist’s model of allometric scaling
429 again: the same questions remain. *Functional Ecology* **19**, 739–743 (2005).
- 430 15. White, C. R., Alton, L. A., Bywater, C. L., Lombardi, E. J. & Marshall, D. J. Metabolic scaling
431 is the product of life-history optimization. *Science* **377**, 834–839 (2022).
- 432 16. Glazier, D. S. The 3/4-power law is not universal: evolution of isometric, ontogenetic
433 metabolic scaling in pelagic animals. *BioScience* **56**, 325–332 (2006).
- 434 17. Glazier, D. S. Beyond the ‘3/4-power law’: variation in the intra- and interspecific scaling
435 of metabolic rate in animals. *Biological Reviews* **80**, 611–662 (2005).
- 436 18. Rubalcaba, J. G., Verberk, W. C. E. P., Hendriks, A. J., Saris, B. & Woods, H. A. Oxygen
437 limitation may affect the temperature and size dependence of metabolism in aquatic
438 ectotherms. *Proceedings of the National Academy of Sciences* **117**, 31963–31968 (2020).
- 439 19. Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M. & Charnov, E. L. Effects of size and
440 temperature on metabolic rate. *Science* **293**, 2248–2251 (2001).
- 441 20. Burton, T., Killen, S. S., Armstrong, J. D. & Metcalfe, N. B. What causes intraspecific
442 variation in resting metabolic rate and what are its ecological consequences? *Proc. R. Soc.*
443 *B.* **278**, 3465–3473 (2011).
- 444 21. Leiva, F. P. *et al.* Differences in the respiratory response to temperature and hypoxia
445 across four life-stages of the intertidal porcelain crab *Petrolisthes laevigatus*. *Marine*
446 *Biology* **165**, 146–156 (2018).

- 447 22. Killen, S. S., Atkinson, D. & Glazier, D. S. The intraspecific scaling of metabolic rate with
448 body mass in fishes depends on lifestyle and temperature. *Ecology letters* **13**, 184–193
449 (2010).
- 450 23. Nakagawa, S. *et al.* Method Reporting with Initials for Transparency (MeRIT) promotes
451 more granularity and accountability for author contributions. *Nature Communications* **14**,
452 1788 (2023).
- 453 24. Page, M. J. *et al.* The PRISMA 2020 statement: an updated guideline for reporting
454 systematic reviews. *International journal of surgery* **88**, 105906 (2021).
- 455 25. Schulte, P. M. What is environmental stress? Insights from fish living in a variable
456 environment. *The Journal of experimental biology* **217**, 23–34 (2014).
- 457 26. Spicer, J. I. What can an ecophysiological approach tell us about the physiological
458 responses of marine invertebrates to hypoxia? *The Journal of experimental biology* **217**,
459 46–56 (2014).
- 460 27. Pörtner, H. O., Storch, D. & Heilmayer, O. Constraints and trade-offs in climate-dependent
461 adaptation: energy budgets and growth in a latitudinal cline. *Scientia marina* **69**, 271–285
462 (2005).
- 463 28. Seibel, B. A. & Drazen, J. C. The rate of metabolism in marine animals: environmental
464 constraints, ecological demands and energetic opportunities. *Philosophical Transactions*
465 *of the Royal Society B: Biological Sciences* **362**, 2061–2078 (2007).
- 466 29. Kroeker, K. J., Kordas, R. L., Crim, R. N. & Singh, G. G. Meta-analysis reveals negative yet
467 variable effects of ocean acidification on marine organisms. *Ecology letters* **13**, 1419–1434
468 (2010).
- 469 30. Kroeker, K. J. *et al.* Impacts of ocean acidification on marine organisms: quantifying
470 sensitivities and interaction with warming. *Global change biology* **19**, 1884–1896 (2013).
- 471 31. Byrne, M. & Przeslawski, R. Multistressor impacts of warming and acidification of the
472 ocean on marine invertebrates' life histories. *Integrative and Comparative Biology* **53**,
473 582–596 (2013).
- 474 32. Harvey, B. P., Gwynn-Jones, D. & Moore, P. J. Meta-analysis reveals complex marine
475 biological responses to the interactive effects of ocean acidification and warming. *Ecology*
476 *and evolution* **3**, 1016–1030 (2013).
- 477 33. Przeslawski, R., Byrne, M. & Mellin, C. A review and meta-analysis of the effects of multiple
478 abiotic stressors on marine embryos and larvae. *Global change biology* **21**, 2122–2140
479 (2015).
- 480 34. López-Urrutia, Á., San Martín, E., Harris, R. P. & Irigoien, X. Scaling the metabolic balance
481 of the oceans. *Proceedings of the National Academy of Sciences* **103**, 8739–8744 (2006).
- 482 35. Makarieva, A. M., Gorshkov, V. G., Li, B.-L. & Chown, S. L. Size- and temperature-
483 independence of minimum life-supporting metabolic rates. *Functional Ecology* **20**, 83–96
484 (2006).
- 485 36. Makarieva, A. M. *et al.* Mean mass-specific metabolic rates are strikingly similar across
486 life's major domains: evidence for life's metabolic optimum. *Proceedings of the National*
487 *Academy of Sciences* **105**, 16994–16999 (2008).
- 488 37. Chown, S. L. *et al.* Scaling of insect metabolic rate is inconsistent with the nutrient supply
489 network model. *Functional Ecology* **21**, 282–290 (2007).
- 490 38. Hughes, S. J. M. *et al.* Deep-sea echinoderm oxygen consumption rates and an interclass
491 comparison of metabolic rates in Asterozoa, Crinozoa, Echinozoa, Holothurozoa and
492 Ophiurozoa. *Journal of Experimental Biology* **214**, 2512–2521 (2011).
- 493 39. Seebacher, F., White, C. R. & Franklin, C. E. Physiological plasticity increases resilience of
494 ectothermic animals to climate change. *Nature Climate Change* **5**, 61–66 (2015).
- 495 40. White, C. R. *et al.* The origin and maintenance of metabolic allometry in animals. *Nature*
496 *ecology & evolution* **3**, 598–603 (2019).
- 497 41. Tremblay, N. *et al.* Euphausiid respiration model revamped: Latitudinal and seasonal
498 shaping effects on krill respiration rates. *Ecological Modelling* **291**, 233–241 (2014).

- 499 42. Burggren, W. W. & McMahon, B. R. *Biology of the Land Crabs*. (Cambridge University
500 Press, 1988).
- 501 43. Gosling, E. *Bivalve Molluscs: Biology, Ecology and Culture*. (John Wiley & Sons, 2008).
- 502 44. Holdich, D. M. *Biology of Freshwater Crayfish*. vol. 702 (Blackwell Science Oxford, 2002).
- 503 45. *Advances in Marine Biology. The Biology of Northern Krill*. (Academic Press, 2010).
- 504 46. Kinne, O. Temperature: animals-invertebrates. *Marine ecology* **1**, 407–514 (1970).
- 505 47. Grieshaber, M. K., Hardewig, I., Kreutzer, U. & Pörtner, H. O. Physiological and metabolic
506 responses to hypoxia in invertebrates. *Reviews of Physiology, Biochemistry and*
507 *Pharmacology* **125**, 43–147 (1993).
- 508 48. Burnett, L. E. & Stickle, W. B. Physiological responses to hypoxia. in *Coastal hypoxia:*
509 *consequences for living resources and ecosystems* (eds. Rabalais, N. N. & Turner, R. E.) vol.
510 58 101–114 (American Geophysical Union, Washington D.C., 2001).
- 511 49. Rombough, P. J. Respiratory gas exchange, aerobic metabolism, and effects of hypoxia
512 during early life. in *Fish physiology* vol. 11 59–161 (Elsevier, 1988).
- 513 50. Zhang, L., Guo, K., Zhang, G.-Z., Lin, L.-H. & Ji, X. Evolutionary transitions in body plan and
514 reproductive mode alter maintenance metabolism in squamates. *BMC Evol Biol* **18**, 45
515 (2018).
- 516 51. Froese, R. & Pauly, D. FishBase. World Wide Web electronic publication.
517 www.fishbase.org, (ver. 10/2023). (2023).
- 518 52. Chabot, D., Steffensen, J. F. & Farrell, A. P. The determination of standard metabolic rate
519 in fishes. *Journal of Fish Biology* **88**, 81–121 (2016).
- 520 53. Brett, J. R. The metabolic demand for oxygen in fish, particularly salmonids, and a
521 comparison with other vertebrates. *Respiration physiology* **14**, 151–170 (1972).
- 522 54. Hinchliff, C. E. *et al.* Synthesis of phylogeny and taxonomy into a comprehensive tree of
523 life. *Proceedings of the National Academy of Sciences* **112**, 12764–12769 (2015).
- 524 55. Rees, J. A. & Cranston, K. Automated assembly of a reference taxonomy for phylogenetic
525 data synthesis. *Biodivers Data J* e12581 (2017) doi:10.3897/BDJ.5.e12581.
- 526 56. Lenoir, J. *et al.* Species better track climate warming in the oceans than on land. *Nature*
527 *Ecology & Evolution* **4**, 1044–1059 (2020).
- 528 57. Chamberlain, S. *et al.* Interface to the Global Biodiversity Information Facility API. (2021).
- 529 58. Chamberlain, S. A. & Szöcs, E. taxize: taxonomic search and retrieval in R. *F1000Research*
530 **2**, (2013).
- 531 59. Michonneau, F., Brown, J. W. & Winter, D. J. rotl: an R package to interact with the Open
532 Tree of Life data. *Methods in Ecology and Evolution* **7**, 1476–1481 (2016).
- 533 60. Revell, L. J. phytools: an R package for phylogenetic comparative biology (and other
534 things). *Methods in Ecology and Evolution* **3**, 217–223 (2012).
- 535 61. Killen, S. S. *et al.* Guidelines for reporting methods to estimate metabolic rates by aquatic
536 intermittent-flow respirometry. *Journal of Experimental Biology* **224**, jeb242522 (2021).
- 537 62. Wieser, W. Benthic studies in Buzzards Bay II. The meiofauna. *Limnology & Oceanography*
538 **5**, 121–137 (1960).
- 539 63. Brey, T., Müller-Wiegmann, C., Zittier, Z. M. C. & Hagen, W. Body composition in aquatic
540 organisms — A global data bank of relationships between mass, elemental composition
541 and energy content. *Journal of Sea Research* **64**, 334–340 (2010).
- 542 64. Cumminns, K. W. & Wuycheck, J. C. Caloric Equivalents for Investigations in Ecological
543 Energetics: With 2 figures and 3 tables in the text. *SIL Communications, 1953-1996* **18**, 1–
544 158 (1971).
- 545 65. Robinson, L. A. *et al.* Length–weight relationships of 216 North Sea benthic invertebrates
546 and fish. *Journal of the Marine Biological Association of the United Kingdom* **90**, 95–104
547 (2010).
- 548 66. Brey, T. Population dynamics in benthic invertebrates. A virtual handbook. Version 01.2.
549 <http://www.thomas-brey.de/science/virtualhandbook/> (2001).
- 550