

# 1 **Faster than expected: Release of nitrogen and phosphorus from decomposing wood**

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## 13 14 **Summary**

- 15 ● Deadwood represents globally important carbon, nitrogen, and phosphorus pools.  
16 Current wood nutrient dynamics models are extensions of those developed for leaf  
17 litter decomposition. However, tissue structure and dominant decomposers differ  
18 between deadwood and litter, and recent evidence suggests that decomposer  
19 stoichiometry in combination with litter quality may affect nutrient release.  
20  
21 ● We quantified decomposition and release of carbon and nutrients from wood for two  
22 stem sizes of 22 tree species in a phosphorus-limited temperate forest near Sydney,  
23 Australia, and compared these to estimates from leaf litter literature.  
24  
25 ● Following theory, nitrogen and phosphorus accumulated during early decomposition,  
26 but began to decline earlier than expected from work on leaves. Deadwood  
27 converged on higher carbon:nitrogen (50) and nitrogen:phosphorus (80) ratios than  
28 in leaf litter studies. Carbon:nitrogen at which nitrogen was released was higher in  
29 large stems (~135) than small stems (~95); both being higher than leaf litter.  
30  
31 ● Drawing from the literature, these differences in nitrogen and phosphorus dynamics  
32 may be due to the identity of wood decomposers. Carbon:nitrogen of wood  
33 decomposers is higher than mean carbon:nitrogen of leaf litter decomposers, and  
34 this difference in stoichiometry may have important flow-on effects for nutrient cycles  
35 in forests.  
36

## 37 **Keywords**

38

39 Decomposition, Deadwood, Nutrient cycling, Microbes, Stoichiometry, Temperate forest

40

## 41 **Introduction**

42

43 Deadwood represents up to 8% of forest carbon (C) stocks and is a crucial component of  
44 nutrient stores (Harmon *et al.*, 1986; Pan *et al.*, 2011). Deadwood has low concentrations of  
45 nitrogen (N) and phosphorus (P) compared with other dead vegetation components, such as  
46 leaf litter, making wood a low quality resource for decomposers (Laiho & Prescott, 2004).  
47 However, some decomposing microbes, specifically wood saprotrophic fungi, have adapted  
48 to exploit deadwood despite its low quality, promoting nutrient cycling within forests (Lindahl  
49 *et al.*, 2002). Compared with leaf litter, nutrients stored within deadwood exhibit a slower  
50 turnover due to wood's slower decomposition rates providing a more long-lasting source of  
51 nutrients to the ecosystem. Currently, most knowledge of nutrient cycling within dead  
52 vegetation pools comes from leaf litter (Parton *et al.*, 2007; Manzoni *et al.*, 2010). However,  
53 because of the large biomass and unique structural properties and contributions to the  
54 nutrient and C cycles as compared to leaf litter, it is important to understand the factors that  
55 determine the rate at which C and nutrients are released from deadwood.

56

57 Previous studies showed that deadwood retains, and even increases, nutrient  
58 concentrations such as N and P during the decomposition process (Harmon *et al.*, 1986;  
59 Herrmann & Prescott, 2008; Palviainen *et al.*, 2008; Meriem *et al.*, 2016; Smyth *et al.*, 2016;  
60 Castillo *et al.*, 2023). Such nutrient accumulation is known as immobilisation and is primarily  
61 driven by the foraging behaviour of microbes, which have adapted different strategies to  
62 efficiently process C while retaining nutrients (Manzoni *et al.*, 2021). These strategies  
63 include the alteration of C use efficiency (Manzoni *et al.*, 2008), import of nutrients from the  
64 surrounding environment (Wells & Boddy, 1995) and increased retention time of elements  
65 within microbes after senescence (Spohn & Widdig, 2017). The nutrients accumulate and  
66 are immobilised within the deadwood as C concentrations decrease until a certain threshold  
67 of nutrient:C ratio is reached. Once this threshold is met, microbes and their substrates are  
68 at a stoichiometric balance meaning that nutrients are no longer limiting the growth of  
69 microbes and can be exported to other mycelium or released to the soil (Zechmeister-  
70 Boltenstern *et al.*, 2015). In addition, other mechanisms, although less influential, such as N  
71 fixation or physico-chemical processes including wicking can contribute to the accumulation  
72 of N (Smyth *et al.*, 2016). As N concentrations increase while C concentrations decrease,  
73 the C:N ratio will also decrease during the decomposition process until it stabilises ~20  
74 (Manzoni *et al.*, 2010).

75

76 Deadwood resources vary significantly in nutrient concentrations depending on the age and  
77 species of a fallen tree (Harmon *et al.*, 1986), which may lead to differences in the amount of  
78 time nutrients are immobilised within the resource leading to different lag times of when  
79 there are net releases of nutrients. Theory developed for leaf litter decomposition suggests  
80 that the initial nutrient concentration of litter is the main determinant for an observed lag in  
81 the release of nutrients from a substrate (Parton *et al.*, 2007; Manzoni *et al.*, 2010, 2021).  
82 Nutrients within leaf litter with low nutrient availability tend to be released later in the  
83 decomposition process than from more nutrient rich litter. In concurrence, microbes have  
84 developed adaptations such as a reduction in C use efficiency (Manzoni *et al.*, 2008) and  
85 extraction of additional nutrients from the surrounding soil to cope with low nutrient  
86 availability (Frey *et al.*, 2000; Parton *et al.*, 2007; Manzoni *et al.*, 2021). Studies found that  
87 these adaptations lead to critical changes in the stoichiometric balance of litter at which N is  
88 released ( $Crit_{CN}$ ), which tends to increase as initial N increases (Manzoni *et al.*, 2008; Ågren  
89 *et al.*, 2013). For litter,  $Crit_{CN}$  is typically between 30 and 50 (Parton *et al.*, 2007) but this  
90 value is likely higher in deadwood due to its higher initial C:N ratios (Manzoni *et al.*, 2010;  
91 Smyth *et al.*, 2016). While these processes are fairly well understood within leaf litter, the  
92 factors controlling the accumulation and release patterns in N and P for deadwood are not.

93

94 In translating nutrient cycling theory for leaf litter to deadwood, besides differences in their  
95 stoichiometry, the other main differences between leaf litter and deadwood are size and  
96 geometric structure. Contrary to litter which tends to have a two-dimensional structure and  
97 low variation in surface area:volume, deadwood is three-dimensional and variable in volume  
98 depending on the diameter and length of the branches and stems falling to the ground.  
99 Different sized wood may influence decomposition rates and C loss; for instance, smaller  
100 diameter stems decompose faster (Mackensen *et al.*, 2003; Weedon *et al.*, 2009; Hu *et al.*,  
101 2018; Lee *et al.*, 2022). Such differences are thought to be because of the higher surface  
102 area-to-volume ratios and higher accessible nutrient concentrations in small as compared to  
103 large stems, which allows for a faster and more efficient colonisation of the substrate by  
104 microbes (Cornwell *et al.*, 2009). In light of these expectations, we predicted a faster  
105 colonisation process and a higher proportional nutrient concentration in small stems, which  
106 could lead these stems to immobilise nutrients for shorter periods than larger stems of the  
107 same species.

108

109 The assimilation and release patterns of different nutrients such as N and P may also  
110 depend on how saprotrophs exploit the resources and availability of these resources in the  
111 surrounding environment. For instance, N is expelled from organisms and reabsorbed as it is

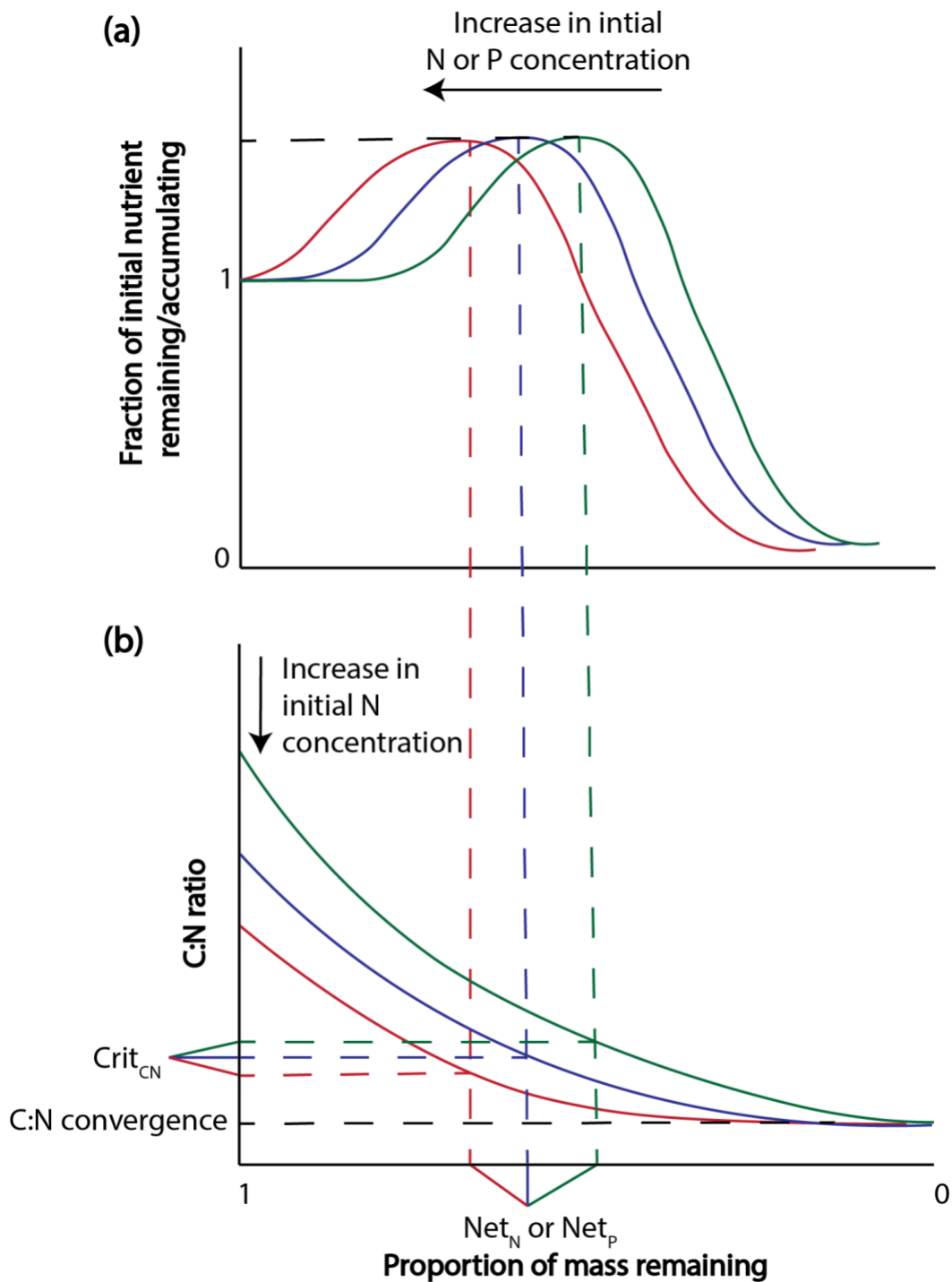
112 crucial for the functioning of extracellular enzymes (Walker & White, 2017). In contrast, P is  
113 mostly retained inside saprotrophic organisms (Beever & Burns, 1981). We predicted P to be  
114 released earlier in the decomposition process than N as it is more efficiently retained by the  
115 saprotrophs and not as dependent on exterior stoichiometry of the resource to be  
116 reabsorbed. N-limited or P-limited ecosystems may also lead to different strategies in how  
117 organisms process these resources. As the concentration of P in the surrounding  
118 environment is lower, we also predicted that it would be exported earlier from deadwood as  
119 there is higher competition for P and other organisms may be actively syphoning the  
120 nutrients (Zechmeister-Boltenstern *et al.*, 2015).

121

122 To better understand the dynamics of nutrients within deadwood through time, we used a  
123 decomposition experiment including 22 woody species in a sclerophyll woodland in Australia  
124 where P limits productivity (Ellsworth *et al.*, 2017). The nutrient concentrations of deadwood  
125 for each species were measured at five time points over a five-year window to accurately  
126 track their changes through time. Following theory based on litter decomposition (Parton *et*  
127 *al.*, 2007; Manzoni *et al.*, 2010) and previous studies looking at changes in nutrients through  
128 time for deadwood (Fig. 1), we formulated four hypotheses:

- 129 - H<sub>1</sub>: We expected nutrient (N and P) concentrations in deadwood to initially increase  
130 until a certain mass remaining threshold is reached at which point a net nutrient  
131 release ( $Net_N$  for N and  $Net_P$  for P) will occur. We expected  $Net_N$  and  $Net_P$  of wood to  
132 be similar to that of low-quality leaf litter, i.e. ~40% of mass remaining. We also  
133 expected N to be released at a C:N ratio above 50 and P to be released before N.
- 134 - H<sub>2</sub>: We expected deadwood with higher initial N and P to release nutrients earlier in  
135 the decomposition process (i.e., at higher mass remaining) and at a lower  $Crit_{CN}$ .
- 136 - H<sub>3</sub>: We expected smaller size wood to have a higher N and P concentration and  
137 release nutrients earlier in the decomposition process while N would be released at a  
138 lower  $Crit_{CN}$ .
- 139 - H<sub>4</sub>: Given the P limitation of our site, we expected N:P ratios of stems to be >30  
140 (Zechmeister-Boltenstern *et al.*, 2015) and decrease during the decomposition  
141 process up to that of saprotrophic fungi at ~13 (Zhang & Elser, 2017). We expected  
142 C:N ratios to decrease until a threshold of ~20 (Manzoni *et al.*, 2010) as fungi create  
143 a favourable environment for C assimilation throughout the decomposition process.

144



145

146 Figure 1 - (a) Nutrient release curves and (b) C:N ratio of wood against proportion of mass  
 147 remaining. Each coloured line represents changes in (a) fraction of initial nutrient  
 148 remaining/accumulating and (b) C:N ratios of a species against proportion of mass  
 149 remaining with species ranging in initial N or P concentration (green < blue < red). The  
 150 intersection of the peak of the nutrient release curve with the proportion mass remaining  
 151 represents the mass remaining threshold when there is a net release of the nutrient ( $Net_N$  for  
 152 N and  $Net_P$  for P). The intersection of  $Net_N$  with the corresponding C:N ratio corresponds to  
 153  $Crit_{CN}$ .

154

## 155 **Materials and Methods**

156

### 157 *Study design*

158

159 We measured the decomposition of wood from 22 tree species in Cumberland Plain  
160 Woodland on the Western Sydney University Hawkesbury campus (33°37'13"S  
161 150°44'16"E). The tree species were selected from three sites including Agnes Banks  
162 Nature Reserve, Castlereagh Nature Reserve and the Cumberland Plain SuperSite of the  
163 Terrestrial Ecosystem Research Network. More information regarding these sites and the  
164 selected species can be found in Lee *et al.* (2019) and Lee *et al.* (2022). All species are  
165 found within sclerophyll woodlands with mean annual rainfall of 728.1 mm and mean annual  
166 temperatures of 24.3 (max) to 11.1 (min) °C. For each species, we collected a live stem from  
167 at least 3 individuals belonging to a smaller (1–2 cm diameter) size class. For 12 of these  
168 species, we were also able to collect a live stem from at least three individuals from a larger  
169 (5-9 cm diameter) size class. The species for which we did not have a larger size class do  
170 not naturally produce stems of these sizes. For both size classes, each stem was cut into  
171 individual pieces of wood, each 10 cm long.

172

173 At the start of the decomposition study, 30 randomly selected stems for each combination of  
174 species and size class were set out in the field on 4-5 July 2013. The stems were set out  
175 across three 7 m x 1 m plots, each separated by 1 m, in a completely randomised design.  
176 We raked the plots before the deployment of the wood stems to remove litter and coarse  
177 woody debris. We checked that there were no trees or woody shrubs within 1 m of any plot.  
178 Within each plot, pieces were set out at a density of 50 per m<sup>2</sup> with five woody stems placed  
179 lengthwise in rows parallel to the short edge of the plot. We left roughly 10 cm of space  
180 between the ends of each stem. The stem placement was completely randomised. Finally,  
181 we used aluminium tags fastened with plastic cable ties to each stem, and these were  
182 fastened to the ground with a steel pin looping around the stem (Lee *et al.*, 2022). Up to six  
183 randomly selected pieces of wood from each species were collected at five sampling points  
184 (February 2014, August 2014, August 2015, August 2016 and June 2018) for the large size  
185 class and four sampling occasions (all dates except June 2018) for the small size class.  
186 Particularly late in the experiment, we were unable to collect six pieces due to the wood  
187 being too decomposed to recover. At each time point, the stems were collected from the field  
188 and placed in an oven at 105°C for five days at which point dry mass was obtained. We then  
189 calculated the proportion of mass remaining for each piece of wood (equation 1).

190

191 
$$x(t) = \frac{m(t)}{m(t_1)} \tag{1}$$

192

193 Where  $m$  represents the dry mass at time  $t$  and initial mass  $t_1$ .

194

195 *Wood chemistry*

196

197 At the start of the experiment, elemental concentration measurements (for C, N and P) were  
198 made on 1-3 undeployed pieces of wood coming from the same branches and individuals as  
199 used in the experiment. These were collected for each combination of species and size class  
200 where possible (Lee *et al.*, 2022). The elemental concentrations were then measured on  
201 three individual pieces of wood per harvest, except for the final harvest after five years  
202 where measurements were taken on up to six individual pieces of wood, with the number  
203 depending on how many could be recovered.

204

205 For each piece of wood, 150 mg of dried and milled wood was analysed for elemental  
206 concentrations. C and N were determined using a TruMac CN Macro analyzer (Leco, St.  
207 Joseph, Michigan, USA) and P was measured using an Epsilon 3X X-ray fluorescence  
208 spectrometer from PANalytical (Sydney, Australia). In total, we obtained 543 samples with N  
209 and C measurements and 481 samples with P measurements.

210

211 *Analysis*

212

213 To look at the differences in initial N and P concentration of small and large stems, we used  
214 a linear mixed effects model. We averaged N and P concentrations for each combination of  
215 species and size class from the undeployed pieces of wood and used these as a response  
216 variable. Size class was used as a fixed explanatory variable and species as a random  
217 variable to account for variability in responses across species.

218

219 The speed of decomposition for each species and size class combination has been  
220 examined in Lee *et al.* (2022). Decomposition rates ranged between 1.7 to 4.8 years at 50%  
221 mass remaining in small stems, while ranging from 2.2 to 6 years at 50% mass remaining in  
222 large stems. Here we focused on changes in chemistry through the decomposition process.

223 To quantify such changes, we calculated the fraction of each element remaining in relation to  
224 the mass remaining of the wood stem (equation 2).

225

226 
$$f(t) = \frac{(n(t) * x(t))}{(n(t_1) * x(t_1))} \tag{2}$$

227

228 Where  $n$  is the concentration of the element at time  $t$  and  $x$  is the proportion of initial mass  
229 remaining of the wood stem at time  $t$  or  $t_1$ . To quantify the lag in release of each element  
230 through the decomposition process of wood, we used the equation in Parton *et al.* (2007),  
231 (equation 3).

232

$$233 \quad f(x(t), a, b) = \frac{(x(t)*100)/b}{\sqrt{(2*a*(x(t)*100)/b)^2 + (1-((x(t)*100)/b)^2)^2}}$$

234 (3)

235

236 where  $a$  is the control factor for the peak value of the function and  $b$  is the location on  $x$  for  
237 the peak value of the function. Using the package *litterfitter* (Cornwell & Weedon, 2014) in R  
238 (v 4.2.1), we computed the optimal values of  $a$  and  $b$  which generates the function with the  
239 best fit to the fraction of the element remaining against mass remaining.  $Net_N$  and  $Net_P$  were  
240 calculated as  $b$  divided by 100 to standardise these to proportions (equation 4).

241

$$242 \quad Net_N / Net_P = \frac{b}{100} \quad (4)$$

243

244 We extracted  $Net_N$  or  $Net_P$  for each species and size class combination. Using the *glmmPQL*  
245 package, we ran generalised linear mixed effects models with  $Net_N$  or  $Net_P$  as response  
246 variables against size and initial nutrient concentration as fixed factors. In the case of the  
247 model with  $Net_N$  as a response, we multiplied initial N concentration by 10 to model changes  
248 in probability with every 0.1% change in initial N. This was done given the initial N content of  
249 deadwood ranged between 0.08 and 0.8. Additionally, for models using  $Net_N$  or  $Net_P$ , we  
250 scaled the response around the mean to obtain a mean-centred intercept. Such intercepts  
251 correspond to the modelled probability for the mean values of initial N and P concentration,  
252 respectively. Species was used as a random effect to account for variability in responses  
253 across species. We applied a quasibinomial distribution to the model as  $Net_N$  or  $Net_P$  are  
254 proportions which are found within a value of 0 and 1.

255

256 To compute the critical C:N ratio at which N was released, we extracted the mass remaining  
257 at which N was released ( $Net_N$ ) for each combination of species and size class. We then  
258 quantified the change in C:N ratio throughout the decomposition process assuming an  
259 exponential increase in the ratio against proportion mass remaining (i.e. an exponential  
260 decrease throughout the decomposition process), (equation 5).

261

$$262 \quad f(x) = r^x + i \quad (5)$$



263

264 Where  $r$  controls the rate of increase in C:N and  $i$  determines the point of convergence of  
265 C:N at 0% mass remaining (i.e. the output of the equation at  $x = 1$ ). We generated the line of  
266 best fit using the *litterfitter* package which extracted the optimal  $r$  and  $i$  values for each  
267 species and size class combination from which we computed the C:N ratio corresponding to  
268  $Net_N$  ( $Crit_{CN}$ ) as in equation 6.

269

$$270 \quad Crit_{CN} = c^{NetN} + i \quad (6)$$

271

272 For each piece of wood, we also calculated the C:N and N:P ratios. We then ran linear  
273 mixed effects models to look at trends in C:N and N:P ratios through time against proportion  
274 mass remaining and size of stems as explanatory variables. Similar to the other models,  
275 species were used as a random variable. The ratios were log-transformed to account for  
276 their right skew towards lower C:N and N:P values.

277

## 278 **Results**

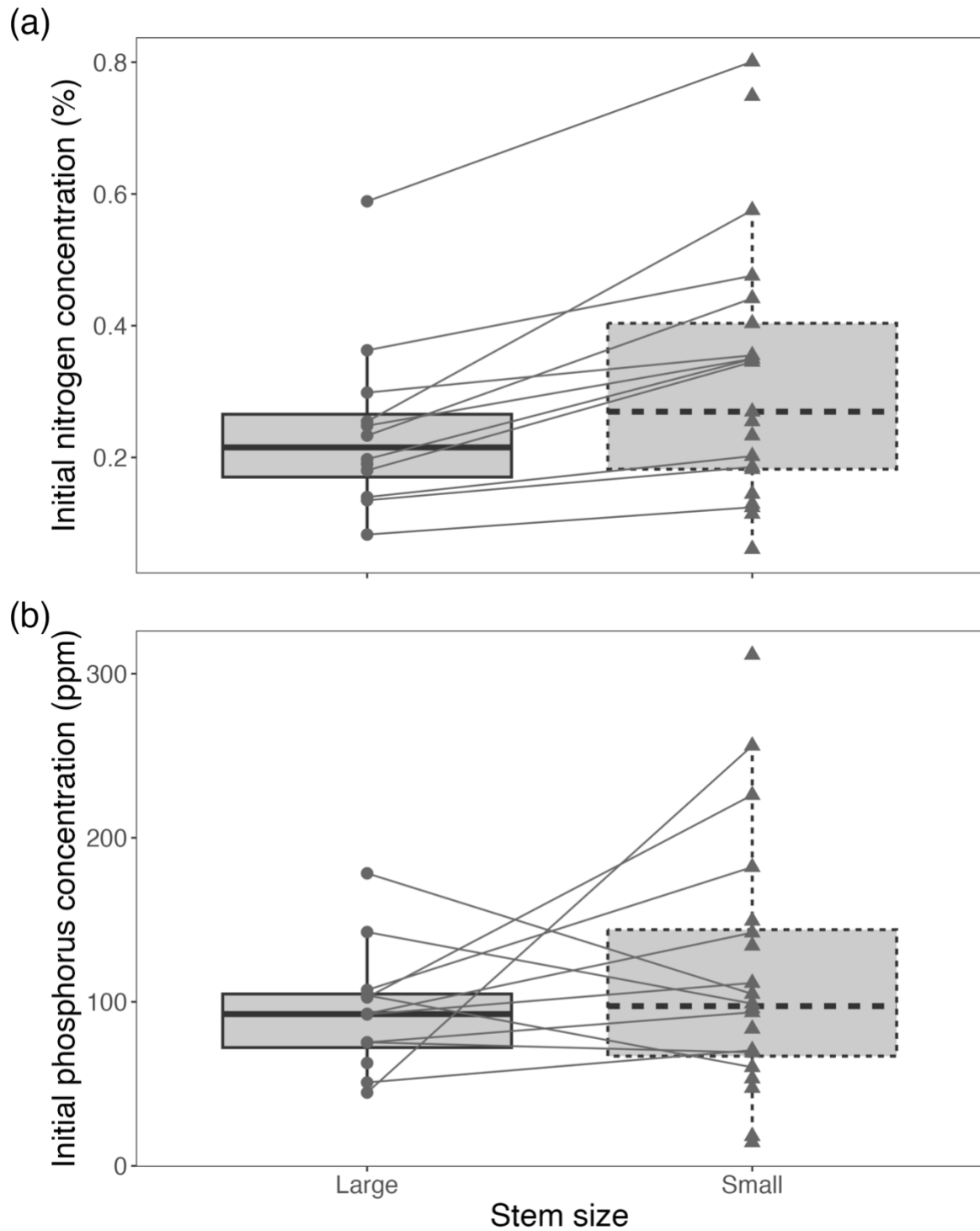
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### 280 *Characteristics of wood*

281

282 Smaller stems had a higher initial N concentration than larger stems (Estimate (Small) =  
283 0.13, 95% CI [0.08-0.18],  $p < 0.001$ ) while there was no difference in initial P concentration  
284 between stem sizes (Estimate (Small) = 22.09, 95% CI [-27.2-71.43],  $p = 0.37$ ), (Fig. 2).

285



286

287 Figure 2 – (a) Initial N concentration and (b) initial P concentration of large (solid contour  
 288 line) and small (dashed contour line) wood stems. Each point represents a species average  
 289 and lines link averages for the same species of different sizes.

290

291

292 *Nutrient release patterns and stoichiometry*

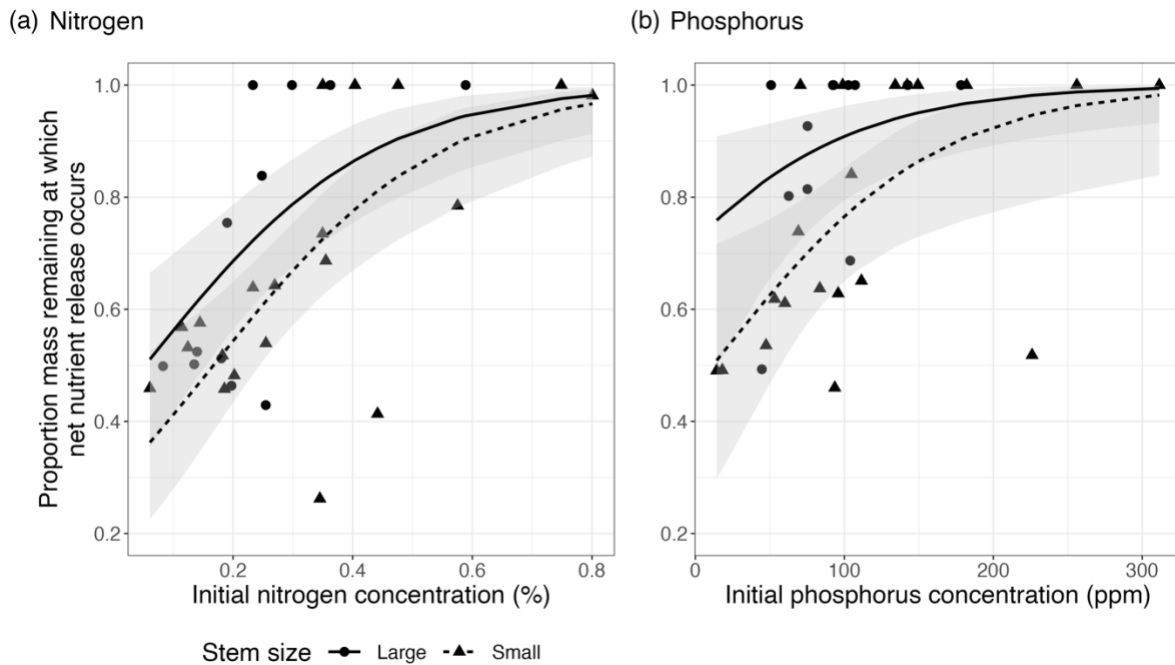
293

294 The fraction of N and P increased in the first stages of the decomposition for most but not all  
295 wood species (Fig. S1, Fig. S2, Fig. S3, Fig. S4). N and P started to be released at a mean  
296 ( $\pm$  SD) of 0.68 ( $\pm$  0.23) and 0.81 ( $\pm$  0.2) of proportion mass remaining, respectively.

297

298 Wood species with a lower initial N concentration released N at a lower mass remaining  
299 (Table 1, Fig. 3a). Similarly, wood species with a lower P concentration released P at a lower  
300 mass remaining (Table 1, Fig. 3b). However, stem size was not observed to influence the  
301 proportion of mass remaining at which either N or P were released (Table 1).

302



303

304 Figure 3 - Proportion mass remaining at which (a) N ( $Net_N$ ) and (b) P ( $Net_P$ ) were released  
305 from wood against initial N or P concentration respectively in large (solid line) and small  
306 (dashed line) stems.

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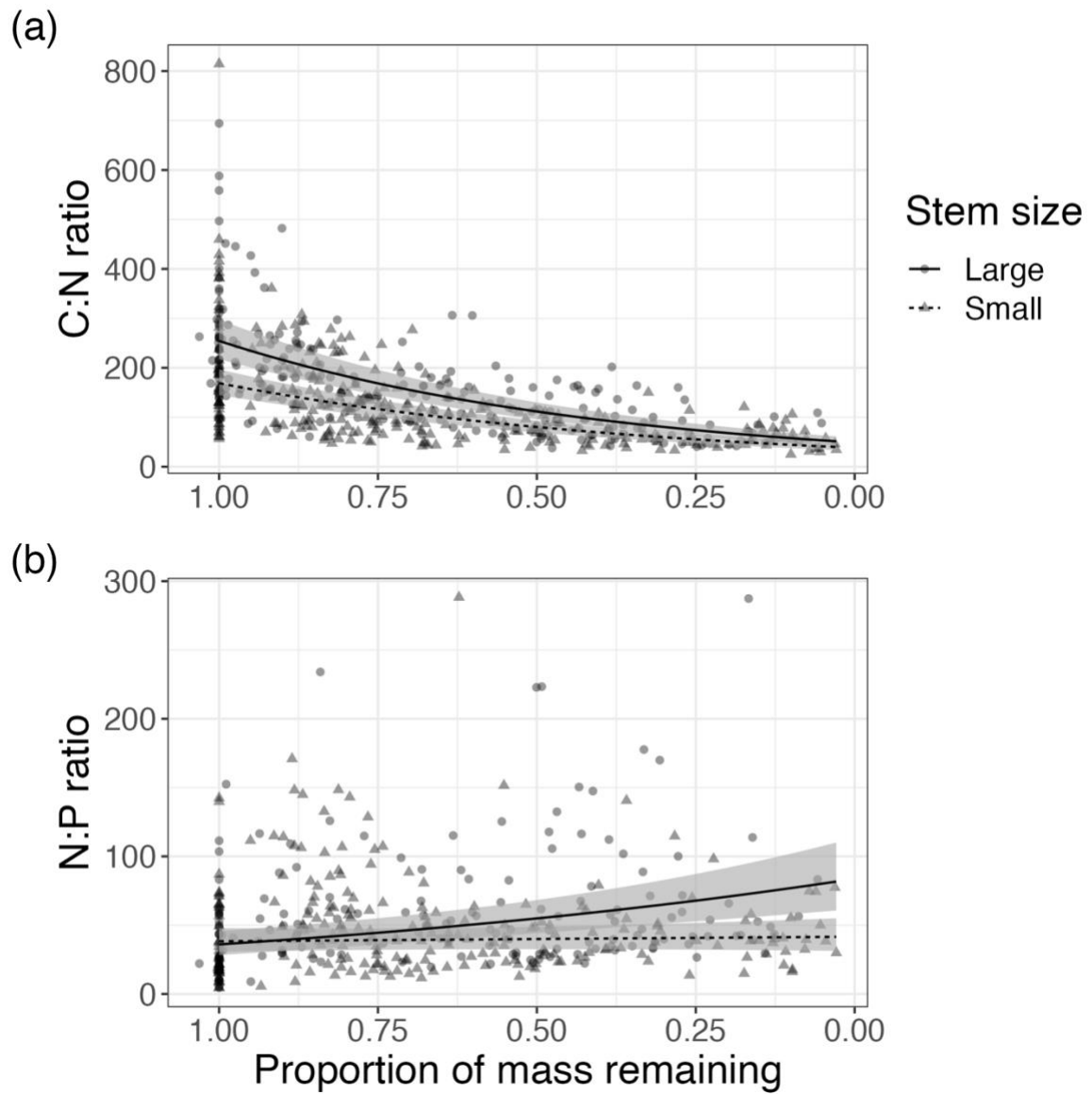
316 Table 1 - Linear mixed effect model output of proportion mass remaining at which N (Net<sub>N</sub>) or  
 317 P (Net<sub>P</sub>) started being released against initial N or P concentration, respectively, and size of  
 318 stem. Species of wood were specified as a random factor. P-values in bold were significant  
 319 at >0.05. To account for the low variation in nutrient concentration among wood species  
 320 (ranging from 0.08 to 0.8) and calculate the probability for every increase in 0.1% initial N  
 321 concentration, initial N concentration was multiplied by 10 and scaled around the mean to  
 322 obtain a mean-centred intercept. Initial P concentration was also scaled around the mean.  
 323 The mean-centred intercepts correspond to the modelled probability for the mean values of  
 324 initial N and P concentration, respectively.

<i>Response</i>	<i>Predictors</i>	<i>Probability</i>	<i>CI</i>	<i>p</i>
<b>Net<sub>N</sub></b>	<i>(Mean-centred intercept)</i>	0.79	0.67 – 0.87	<b>&lt;0.001</b>
	<i>Initial N concentration*10 (%)</i>	0.72	0.60 – 0.82	<b>0.004</b>
	<i>Size [Small]</i>	0.35	0.2 – 0.54	0.125
<b>Net<sub>P</sub></b>	<i>(Mean-centred intercept)</i>	0.92	0.80 – 0.97	<b>&lt;0.001</b>
	<i>Initial P concentration (ppm)</i>	0.71	0.53 – 0.84	<b>0.036</b>
	<i>Size [Small]</i>	0.25	0.09 – 0.53	0.086

325 Notes: Random effect outputs for Net<sub>N</sub>:  $\sigma^2 = 3.29$ ,  $T_{00 \text{ species}} = 0.03$ , ICC = 0.01,  $N_{\text{species}} = 21$ . Observations = 32,  
 326  $R^2 = 0.21$ . Random effect outputs for Net<sub>P</sub>:  $\sigma^2 = 3.29$ ,  $T_{00 \text{ species}} = 0$ , ICC = 0,  $N_{\text{species}} = 21$ . Observations = 32,  $R^2$   
 327 = 0.22.

328  
 329 C:N ratios slowly decreased throughout the decomposition process from a high of ~250 for  
 330 large stems and ~180 for small stems at the start of the decomposition process to a value of  
 331 ~50 at the end regardless of size (Fig. 4a,  $p < 0.001$ ). On the contrary, N:P ratios were ~40  
 332 and similar for both sized stems at the start of the decomposition process. However, N:P  
 333 ratios increased with decomposition to ~80 for large stems while staying similar throughout  
 334 the decomposition process for small stems (Fig. 4b,  $p < 0,001$ ). N was released at an  
 335 average Crit<sub>CN</sub> of 110. Crit<sub>CN</sub> was higher in large stems (~135) than small stems (~95) but  
 336 was not influenced by the initial concentration of N (Fig. 5, Table 2).

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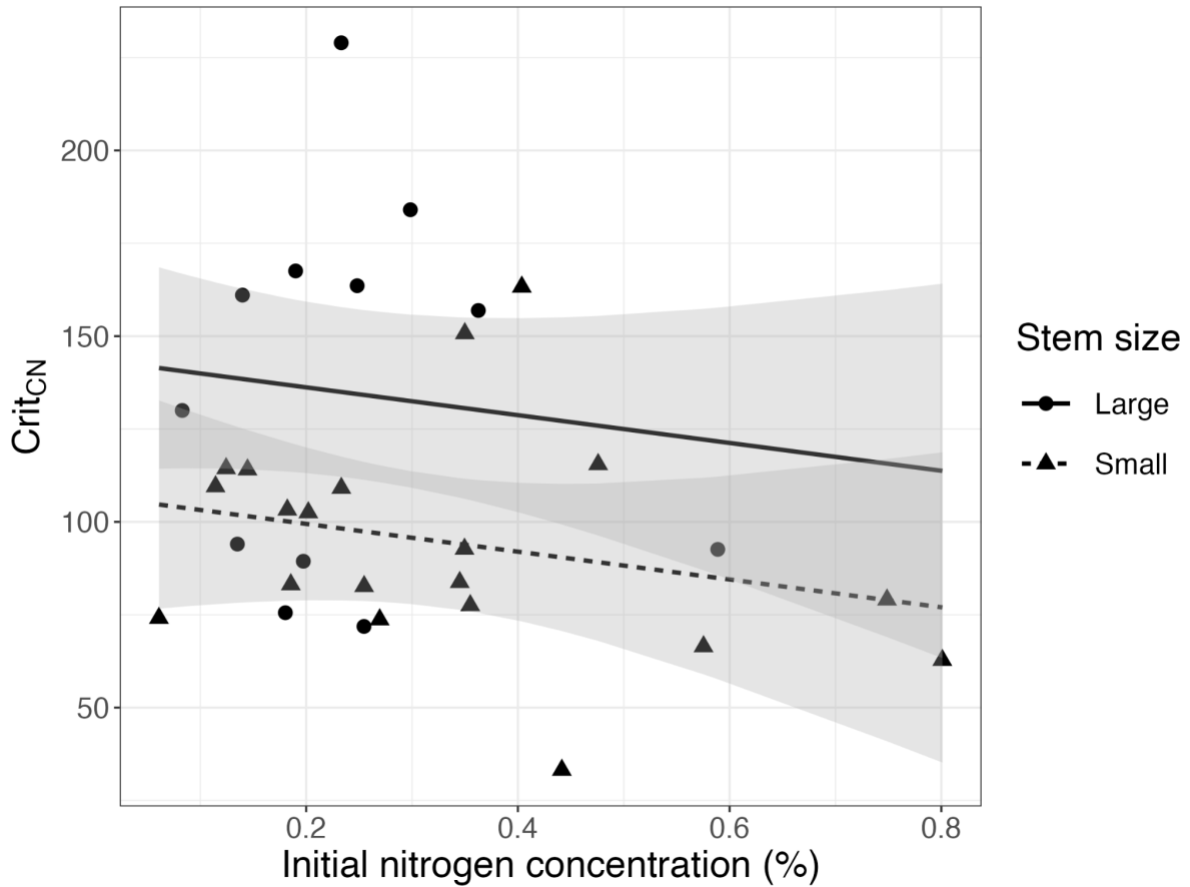


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339 Figure 4 – (a) C:N ratio and (b) N:P of wood stems against proportion mass remaining in

340 large (solid) and small (dashed) stems.

341



342

343 Figure 5 - C:N ratio at which N started being released ( $Crit_{CN}$ ) against initial N concentration  
 344 in large (solid line) and small (dashed line) stems.

345

346 Table 2 - Linear mixed effect model output of C:N ratio at which N is released against size of  
 347 stem and initial N concentration. Species of wood were specified as a random factor. P-  
 348 values in bold are considered significant.

<i>Predictors</i>	<i>Estimates</i>	<b><math>Crit_{CN}</math></b>	
		<i>CI</i>	<i>p</i>
(Intercept)	143.70	113.68 – 173.73	<b>&lt;0.001</b>
Size [Small]	-36.76	-66.48 – -7.04	<b>0.017</b>
Initial N concentration (%)	-37.42	-117.86 – 43.02	0.348

349

Notes: Random effect outputs:  $\sigma^2 = 1483.83$ ,  $T_{00 \text{ species}} = 0$ ,  $N_{\text{species}} = 21$ . Observations = 32,  $R^2 = 0.23$ .

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351

## 352 Discussion

353

354 We investigated the factors influencing the accumulation and release of nutrients from  
355 decomposing wood during the decomposition process. Given the dearth of similar studies in  
356 wood, our predictions were based on studies on leaf litter and we found that these were only  
357 partially supported (Table 3). In line with hypothesis 1, nutrients accumulated until a certain  
358 mass remaining threshold was reached; however, nutrients were released earlier in the  
359 decomposition process for wood compared to even low quality litter (comparison with Moore  
360 *et al.*, (2006) and Parton *et al.*, (2007)), with deadwood from several species showing no lag  
361 in release. In accordance with hypothesis 2, the release of N ( $Net_N$ ) and P ( $Net_P$ ) occurred at  
362 a lower mass remaining for stems with lower initial N and initial P concentrations. However,  
363 contrary to litter studies (Manzoni *et al.*, 2008; Ågren *et al.*, 2013),  $Crit_{CN}$  did not decrease  
364 with initial N. Contrary to our expectations from hypothesis 3, stem size did not influence  
365 mass remaining at which N and P were released from the wood, although stems with a  
366 larger size had a higher  $Crit_{CN}$ . Finally, while general trends in C:N and N:P ratios agreed  
367 with our expectations under hypothesis 4, C:N and N:P converged to a higher (~50 or 80,  
368 respectively) value than expected based on average fungal stoichiometry and previous  
369 studies on leaf litter (Moore *et al.*, 2006; Manzoni *et al.*, 2010). The unique properties of  
370 wood, which attract specially adapted organisms (Wijas *et al.*, 2024), likely lead to singular  
371 and surprisingly dynamic properties in the release of nutrients within forest ecosystems.  
372 Throughout these next sections, we discuss our results in the context of the state of the field,  
373 largely shaped by studies carried out on leaf litter.

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390 Table 3 - Different characteristics in the release of N and P from leaf litter and deadwood.  
 391 Findings for leaf litter from the literature as noted and findings for deadwood from this study.

Hypothesis	Characteristic	Leaf litter	Deadwood
H <sub>1</sub>	Net <sub>N</sub>	40-50% mass remaining (forest data from Parton <i>et al.</i> , (2007))	68% mass remaining
H <sub>1</sub>	Net <sub>P</sub>	57% mass remaining (Tamarack needles in Moore <i>et al.</i> , (2006))	81% mass remaining
H <sub>1</sub>	Crit <sub>CN</sub>	~40 (Parton <i>et al.</i> , 2007)	110
H <sub>2</sub> , H <sub>3</sub>	Decrease in Net <sub>N</sub> with initial N concentration	Yes (Parton <i>et al.</i> , 2007; Manzoni <i>et al.</i> , 2010)	Yes (72% probability of increase in Net <sub>N</sub> for every 0.1% increase in initial N)
H <sub>2</sub> , H <sub>3</sub>	Decrease in Net <sub>P</sub> with initial P concentration	Yes (Parton <i>et al.</i> , 2007; Manzoni <i>et al.</i> , 2010)	Yes (71% probability of increase in Net <sub>P</sub> for every increase in 1 ppm of initial P)
H <sub>2</sub> , H <sub>3</sub>	Decrease in Crit <sub>CN</sub> with increased initial N content	Yes (Ågren <i>et al.</i> , 2013)	No, but smaller stems with a higher N concentration had a lower Crit <sub>CN</sub> compared with larger stems
H <sub>4</sub>	C:N convergence	20 (Manzoni <i>et al.</i> , 2010) 30 (Moore <i>et al.</i> , 2006)	50 (regardless of stem size)
H <sub>4</sub>	N:P convergence	20 (Manzoni <i>et al.</i> , 2010) 16 (Moore <i>et al.</i> , 2006)	40 in small stems, 80 in large stems

392

393

394 *Variation in immobilisation and release: What may be driving it?*

395



396 We found that N and P increased in concentration until an earlier threshold of mass  
397 remaining, 68% and 81% mass remaining, respectively, compared to previous work on leaf  
398 litter (Moore *et al.*, 2006; Parton *et al.*, 2007). These results suggest that export of N and P  
399 balances and then exceeds import at an earlier stage in the decomposition trajectory. These  
400 results appear surprising: why does wood with a lower initial N than leaf litter release N  
401 sooner in the decomposition trajectory compared to leaf litter? One possible explanation for  
402 this incongruity is the recognition that different decomposers break down leaf litter and  
403 deadwood and the stoichiometric needs of them may be different, i.e., the mean C:N of  
404 wood decomposers is much higher than the mean C:N of leaf litter decomposers. For  
405 instance, the C:N ratios at which white rot fungi, which are adapted to degrade lignin in  
406 deadwood, function optimally are estimated to be between 160 and 400 (Eriksson *et al.*,  
407 1980). Under this explanation convergence on decomposer C:N occurs at a relatively higher  
408 mass remaining of deadwood, leading to the surprising release of N (and P) at a high mass  
409 remaining. It is worth noting two things in this context. First, the decomposition process of  
410 wood is significantly slower than that of leaf litter (Pietsch *et al.*, 2014), therefore the release  
411 dynamics are occurring at different time scales (across months for leaf litter compared to  
412 years to decades for deadwood). Second, N and P may not be externally available during  
413 the “release” phase; a portion of these could be exported to belowground fungal mycelium in  
414 a similar way that import occurs early in the decomposition process.

415

416 Although our study was conducted in a P-limited ecosystem, we found that P was released  
417 at a higher mass remaining of deadwood than N. This finding may be indicative of the  
418 dominance of fungi within the decomposition of our substrates given their lower reliance on  
419 P and their higher tolerance to P-limitations than other saprotrophs such as bacteria  
420 (Güsewell & Gessner, 2009). Unlike N, P is not required for the functioning of extracellular  
421 enzymes meaning these can be retained within the cells of fungi (Walker & White, 2017).  
422 Through the transport of P within their cells, fungi can target specific areas to improve  
423 stoichiometric balance and favour the intake of further nutrients (Beever & Burns, 1981).  
424 Additionally, the low availability of P within the soil may increase competition for P and lead  
425 to an early export through the mycelial networks of fungi rather than imports from the soil  
426 (Boddy, 1999). In contrast, the higher availability of N allows the element to accumulate for  
427 longer periods of time. Increases in N:P ratios during the decomposition process, especially  
428 for larger stems, further suggests that higher N is required for the functioning of microbes  
429 compared with P, whether P is being transported out or N is being imported during  
430 decomposition. An increase in N:P ratios in large stems during the decomposition process  
431 also goes against what is generally assumed for leaf litter (Manzoni *et al.*, 2010) in which it  
432 stays unchanged, as we found within small stems.

433

434 Tree species with higher initial wood N and P concentrations released their respective  
435 nutrients at a higher proportion mass remaining. These results were similar to those for leaf  
436 litter from Parton *et al.* (2007) and Manzoni *et al.* (2010) and are consistent with models  
437 showing that stoichiometric balance is obtained at a higher mass remaining in tissue with  
438 higher initial nutrients. Contrary to leaf litter results, however, Crit<sub>CN</sub> did not decrease as  
439 initial N increased (Manzoni *et al.*, 2008, 2010). Microbial communities within deadwood  
440 stems with low nutrient concentrations attained a similar C:N ratio before the net release of  
441 N to those of microbial communities with high nutrient concentrations. Our finding suggests  
442 that stems with low N concentrations either accumulated larger amounts of N and/or  
443 released more C (e.g., via respiration) than stems with high N concentration before they  
444 began releasing N. Stems with lower N concentration could therefore have outsized  
445 contributions to N or C cycling in forest ecosystems.

446

447 *Deadwood size: When does it matter?*

448

449 Small stems did not show a release of N or P at higher mass remaining than large stems  
450 although they contained higher concentrations of N (although P did not differ between stem  
451 sizes; Fig. 2 and Fig. 3). It may seem surprising that higher initial N concentrations did not  
452 lead to earlier release of N in the decomposition process; however, Crit<sub>CN</sub> was also lower in  
453 these stems. Integrating these two results suggests that the higher Crit<sub>CN</sub> in larger stems  
454 may lead to a similar Net<sub>N</sub> between small and large stems compensating for the initial  
455 difference in N concentrations. The size of the substrate therefore may have modulated how  
456 microbes exploited their resources as well as which microbe species could exploit the  
457 resource. Indeed, it is well documented that deadwood with different diameters support  
458 different microbial communities (Juutilainen *et al.*, 2011; Krah *et al.*, 2018). Our results  
459 suggest decomposition was more nuanced than initially expected, depending on the relative  
460 initial and changing concentrations of multiple elements and may mean we need to consider  
461 multiple metrics, e.g., both Crit<sub>CN</sub> and initial N concentrations, when trying to predict the  
462 cycling of nutrients from deadwood within forest ecosystems.

463

464 Overall, as decomposition progressed, the stoichiometry of wood stems changed, although  
465 these effects varied depending on the size of the stems. Similar to studies on litter  
466 decomposition, we showed that C:N decreased as decomposition progressed (Moore *et al.*,  
467 2006; Manzoni *et al.*, 2010), which is most probably due to a combination of N  
468 immobilisation and C respiration. However, while previous estimates based mainly on leaf  
469 litter decomposition suggest that C:N ratios converged ~15-20, we found that C:N ratios

470 converged ~50 in our study. Interestingly, C:N ratios decreased at a higher rate within large  
471 stems compared with small stems. Given N release occurred at similar stages of  
472 decomposition between small and large stems, this suggests C was respired at higher rates  
473 out of larger than smaller stems which may be due to the lower C use efficiency of microbes  
474 (Manzoni *et al.*, 2008). Additionally, we found that N:P increased unequally across large and  
475 small stems as the decomposition process progressed. N:P ratios increased and stabilised  
476 at a value of 80 within large stems while staying unchanged and stabilising ~40 within small  
477 stems. The unequal dynamics in C:N and N:P ratios throughout the decomposition process  
478 between small and large stems suggest that microbial communities are exploiting their  
479 resources differently depending on the size of their resource. Microbial communities within  
480 wood include many specialists (McGuire & Treseder, 2010) and there are different species  
481 of fungi that target larger and smaller stems (Juutilainen *et al.*, 2011; Krah *et al.*, 2018). Such  
482 specialisation may have led to different stoichiometric needs and preferences in different  
483 sizes of wood.

484

#### 485 *Conclusions*

486

487 We show that element accumulation and release patterns within deadwood have important  
488 similarities and differences with leaf litter, which could have consequences for ecosystem  
489 function including for N and P cycles in forests worldwide. Our novel results provide new  
490 insight into how microbes may be responding to the deadwood substrate by showcasing the  
491 stoichiometric balance and resource structural requirements for their optimal function. As C  
492 and N ecosystem models such as the Microbial-MIneral Carbon Stabilization model (Wieder  
493 *et al.*, 2014) or the Community Land Model (Lawrence *et al.*, 2019) increasingly incorporate  
494 details about N and C cycling within different substrates, moving beyond just leaf litter, our  
495 findings on the uniqueness of deadwood may be an early important step. If the results found  
496 here for 22 wood species hold across other taxa and systems, then the similarities and  
497 differences in wood C and nutrient release should be included. Future models could  
498 incorporate nutrient dynamics in relation to deadwood by assuming, for instance, that critical  
499 C:N at which N is released is ~110 and that decomposition stabilises at a C:N ratio of ~50.  
500 Additionally, nutrients are released at a mass remaining of ~40-100% depending on the  
501 nutrient considered and the initial concentrations of that nutrient within the wood. Finally, N:P  
502 ratios can increase during the decomposition process. These characteristics may emerge in  
503 conjunction with modern modelling frameworks if decomposers are also represented for their  
504 substrate preference e.g., leaf litter versus small stems versus large stems of deadwood.  
505 Such approaches would parallel the distinction in models among different plant growth forms

506 and reflect the growing body of knowledge on microbial functional ecology (Zanne *et al.*,  
507 2020).

508

### 509 **Acknowledgments**

510 Funding was received in the form of an Australian Research Council Discovery Grant  
511 (DP160103765). We thank Marissa Lee who provided help with code and data processing.  
512 We thank Brendan Choat and Peter Reich for contributing to the initial project design. We  
513 also thank Kylie Brice, Bethanie Coleman, Coline Deveautour, Gillian Powell, Marc  
514 Rosenfield and Laura Super for help with field and laboratory work.

515

### 516 **Competing interests**

517 We declare no competing interests.

518

### 519 **Author contributions**

520 AEZ, WKC, BO and JRP designed the experiment. WKC, JRP and BO performed the field  
521 data collection. BW performed the data analysis and interpreted the results with guidance  
522 from WKC and AEZ. BW wrote the manuscript with input from all the authors who have  
523 edited the draft.

524

### 525 **Data availability**

526 The data and code will be made available on a public repository.

527

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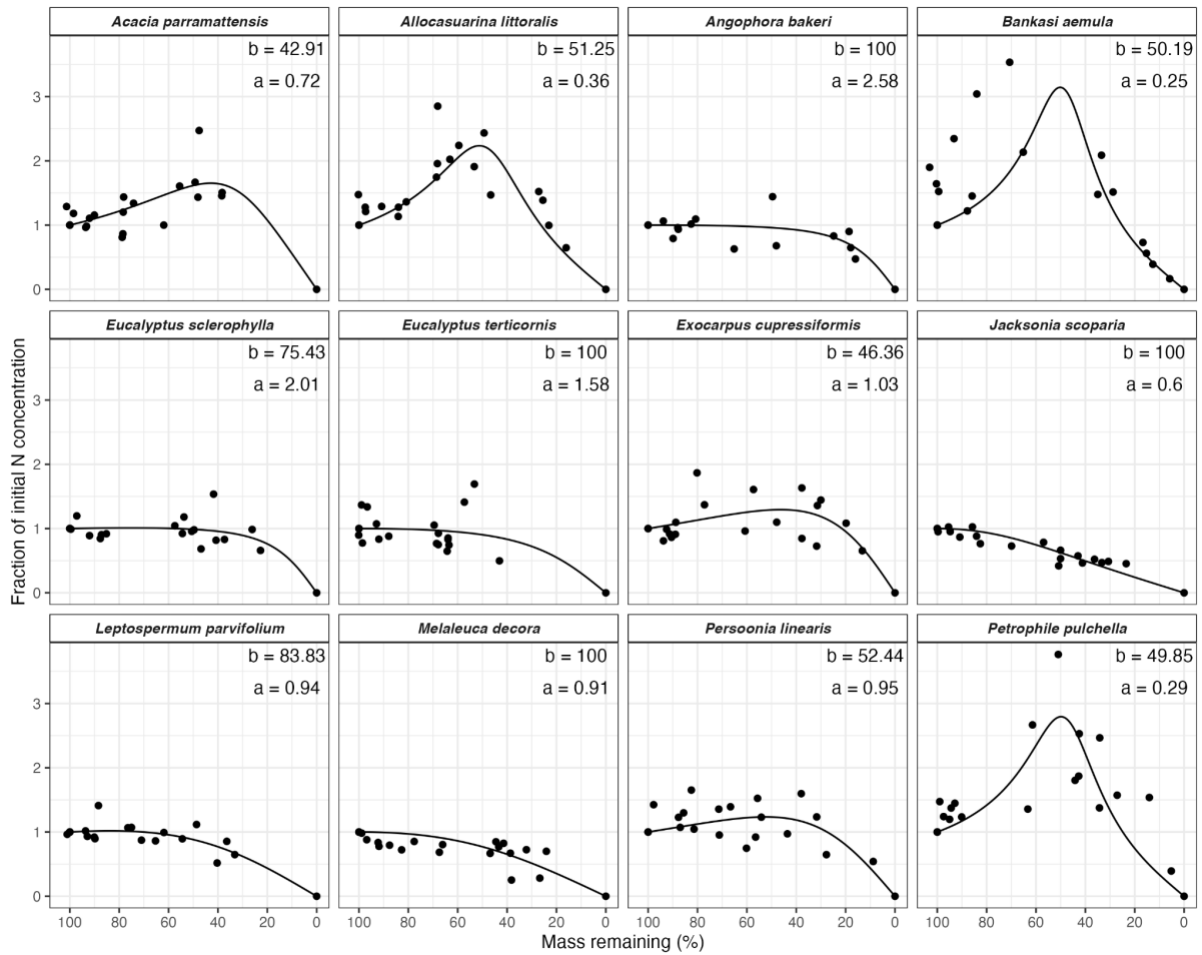
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655 Figure S1 - Fraction of initial nitrogen concentration against mass remaining for each  
 656 species within the larger stem category. The a and b values represent the outputs from the  
 657 line of best fit for each species using equation 3.

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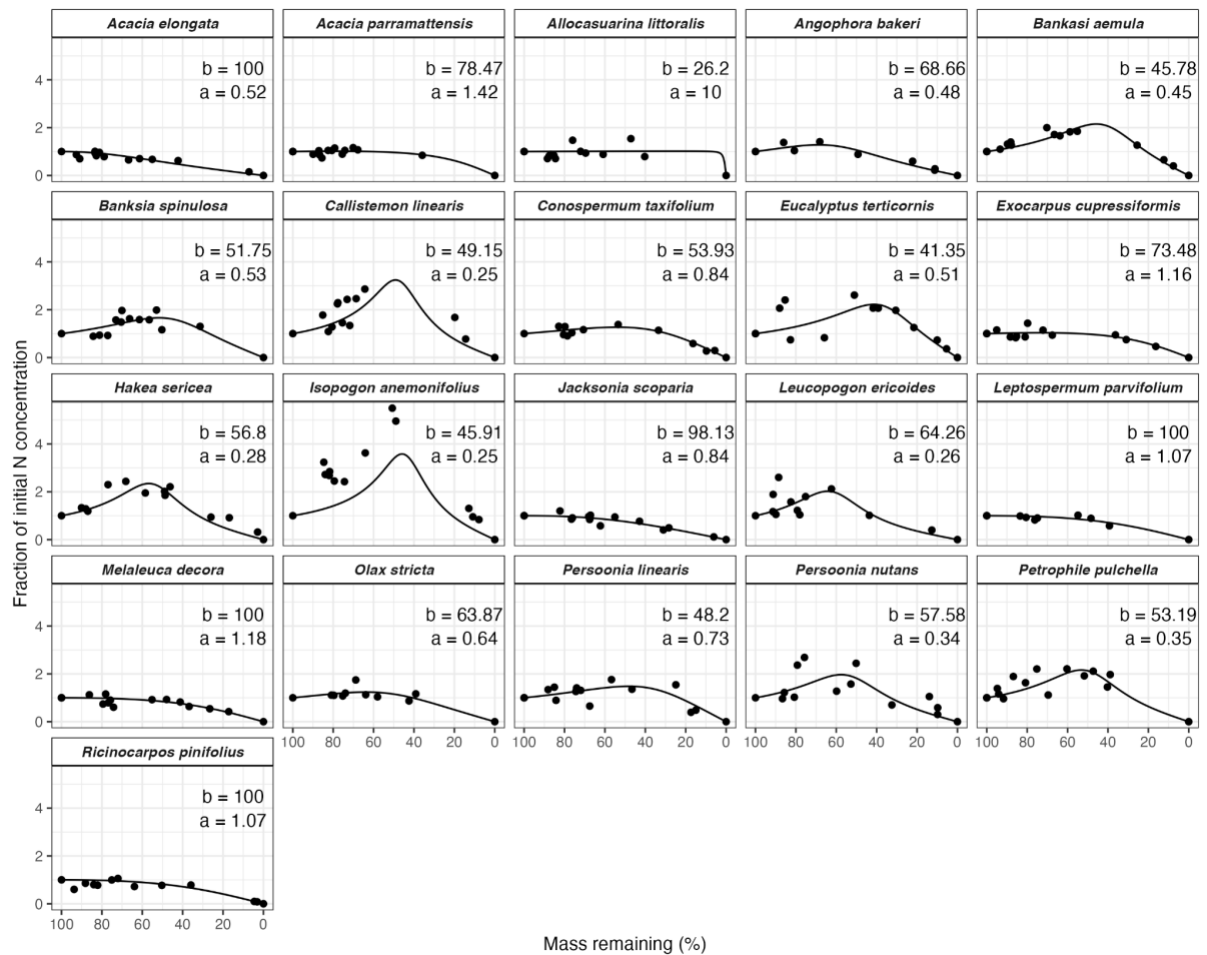
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671 Figure S2 - Fraction of initial nitrogen concentrations for each species within the smaller  
 672 stem category. The a and b values represent the outputs from the line of best fit for each  
 673 species using equation 3.

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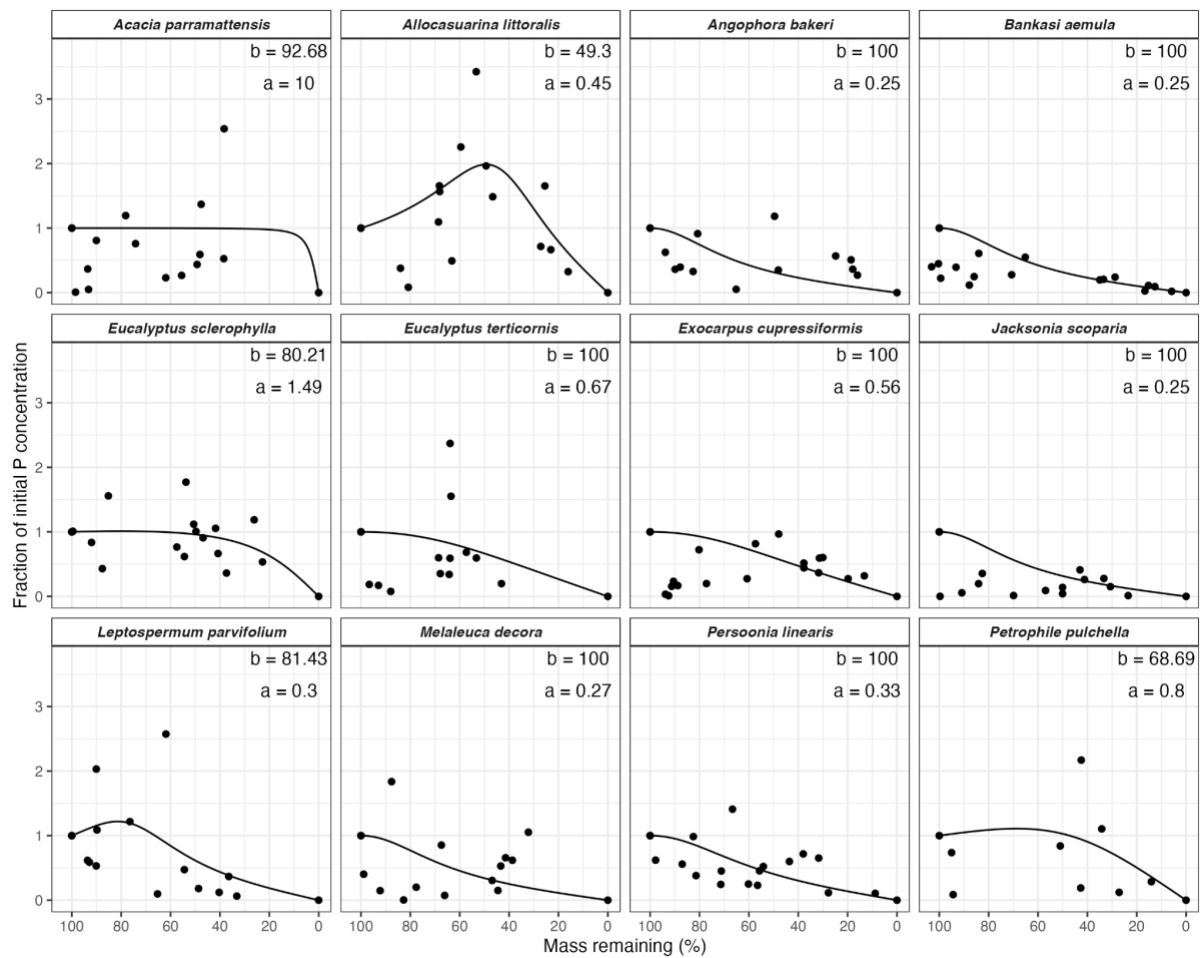
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690 Figure S3 - Fraction of initial phosphorus concentration against mass remaining for each  
 691 species within the larger stem category. The  $a$  and  $b$  values represent the outputs from the  
 692 line of best fit for each species using equation 3.

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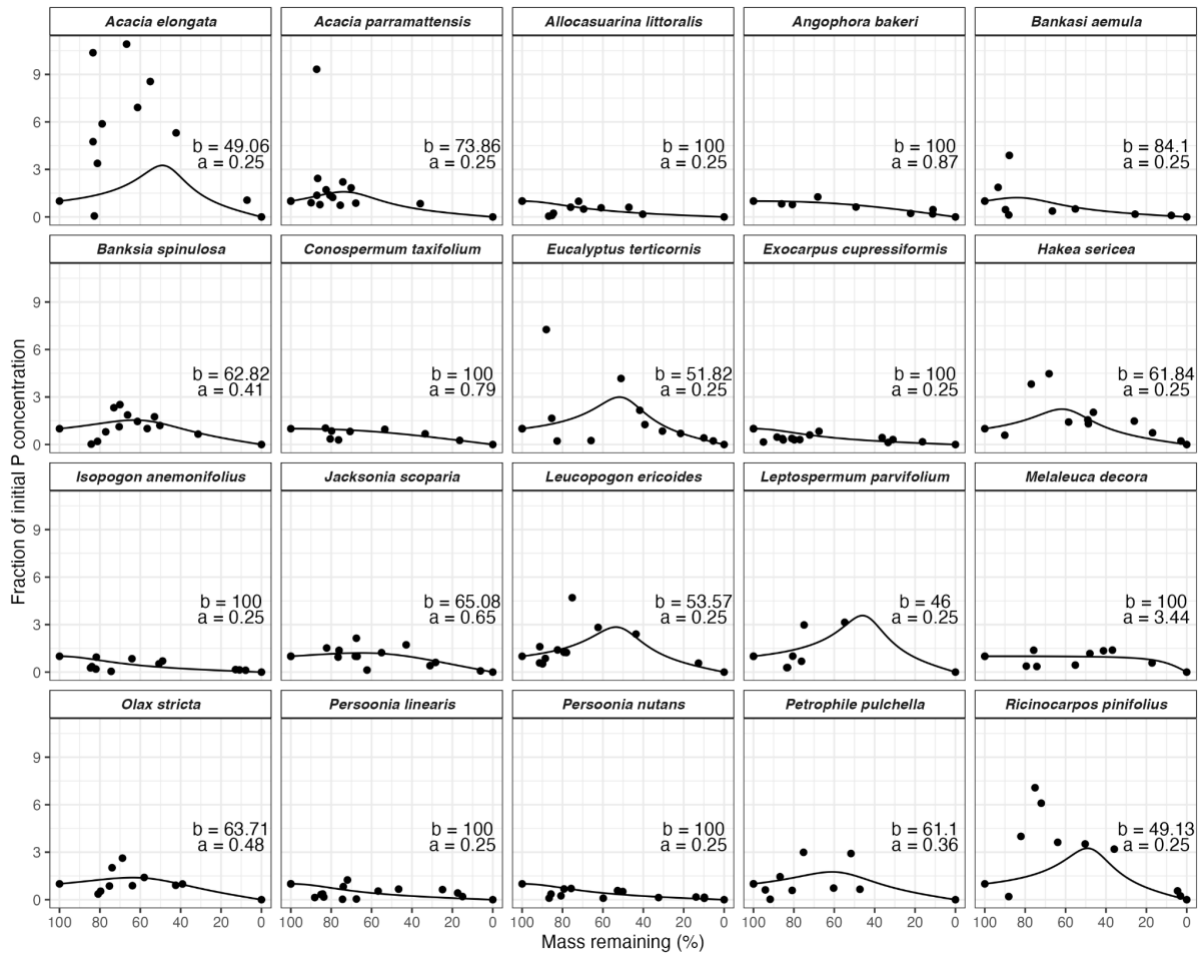
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707 Figure S4 - Fraction of initial phosphorus concentrations for each species within the smaller  
 708 stem category. The a and b values represent the outputs from the line of best fit for each  
 709 species using equation 3.

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