

1 **Mind the lag: understanding delayed genetic erosion**

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10 ***Abstract***

11 The delay between environmental changes and the corresponding genetic responses
12 within populations is a common but surprisingly overlooked phenomenon in ecology,
13 evolutionary and conservation genetics. This time lag problem can lead to erroneous
14 conservation assessments when solely relying on genetic data. We identify population
15 size, life-history traits, reproductive strategies and the severity of population decline as
16 the main determinants of time lags, evaluate potential confounding factors affecting
17 genetic parameters during time lags, and propose methodological approaches that allow
18 controlling for them. Considering the current unprecedented rate of genetic diversity and
19 species loss, we expect our novel interpretive and methodological framework for time
20 lags to stimulate further research and discussion on the most appropriate approaches to
21 analyse genetic diversity for conservation.

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23 **Keywords**

24 conservation genomics, environmental changes, genetic diversity, genetic extinction
25 debt, life history traits

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28 **1. The time lag between environmental changes and the corresponding genetic**
29 **responses**

30 The assumption behind the use of contemporary genetic data in conservation is that
31 datasets mirror the current conservation status of a population [1]. However, genetic
32 parameters often respond to novel environmental conditions and disturbance events
33 with delay, generating time lags [2]. Failure to recognize and account for time lags in
34 genetic responses can lead to erroneous conservation assessments, misguiding the use
35 of resources for biodiversity conservation. At a time of unprecedented biodiversity loss
36 [3], understanding time lags linked to population genetic diversity is therefore not only
37 crucial in ecology and evolutionary genetics, but it is necessary to optimise conservation
38 action.

39 Environmental changes generate population genetic changes because individuals
40 respond to novel conditions with differential survival and reproduction, which can lead
41 to a decline in effective population size (N_e) and to reduced fitness over the course of a
42 single or multiple generations (Fig. 1A). If such changes are not reverted, their ultimate
43 effect will be genome-wide genetic erosion. During time lags, moderate to high levels of
44 genetic variation can persist despite deteriorated environmental conditions [2], for
45 example because of persistent individuals surviving adverse conditions. Contemporary
46 levels of population genetic diversity thus bear the legacy of past habitat and landscape
47 characteristics [4,5]. For the same reason, it is also important to account for time lags
48 when designing conservation management actions [6].

49 Time lags in the genetic response to environmental changes have also been referred to
50 as “genetic extinction debt” or “extinction debt of genetic diversity” [7–10], drawing a
51 parallel with the concept of extinction debt, which describes the delayed loss of species
52 following habitat degradation [11,12]. Extinction debts have received more attention, as
53 they affect entire communities in perturbed ecosystems and environments [6]. However,
54 an extinction debt at the community-level will depend on how environmental changes
55 have acted upon the populations of co-occurring species, and on how quickly changes in
56 the community can be detected. Put simply, extinction debts at the community-level arise
57 from genetic debts at the population level [13], and therefore, understanding genetic
58 extinction debts or time lags at the population level deserves attention as an independent
59 phenomenon.

60 Time lags may also contribute to explaining why genetic diversity is a poor predictor of
61 global IUCN threat status [14], or why threatened species do not necessarily have low
62 genetic diversity [15,16]. Such discrepancies require accounting for delayed genetic
63 responses and do not undermine the importance of exploring genetic variation for
64 conservation practice [17].

65 Conservation genetics is still lacking a framework for the interpretation of genetic
66 diversity in light of the possible occurrence of time lags. Without an organic view of the
67 time lag problem, misinterpretations of current levels of genetic variation may lead to
68 setting wrong priorities for the conservation of populations and species, with enormous
69 waste of efforts and resources.

70 In this article, we explore the biological and ecological factors determining time lags and
71 focus on situations in which time lags may be suspected or exacerbated because of
72 confounding factors. Although in some cases environmental changes improve conditions
73 for survival and have beneficial effects on genetic diversity, we only focus on cases in
74 which deteriorated conditions (e.g. due to habitat loss, fragmentation, climate change,
75 pollution, diseases; [2,11]), have detrimental effects on the survival of individuals or their
76 ability to reproduce, leading to genetic erosion. We finally propose a framework to
77 interpret genetic diversity parameters considering the possible occurrence of time lags.

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79 ***2. Determinants of time lags: factors delaying the detectability of genetic erosion***

80 Life-history traits and other biological traits (Table 1) play a crucial role in allowing
81 polymorphisms to persist even through deteriorated conditions [18–20], delaying genetic
82 erosion and supporting the build-up of a time lag. Such traits essentially both (1) extend
83 the time available for an individual to reproduce (e.g., long life span [21], vegetative
84 propagation, long generation time) and (2) increase the number of opportunities for
85 reproduction and the number of gene combinations arising from reproduction (e.g.,
86 overlapping generations, mating by outcrossing, large numbers of offspring per
87 individual (especially when they reach reproductive maturity; (Table 1)).

88 The influence of life-history traits on genetic diversity is mainly mediated by effective
89 population size (N_e), which mirrors major, long-term differences in genetic diversity
90 between species of plants and animals [20,22]. At the population-level, N_e affects the rate
91 of loss of genetic diversity by drift: large populations (large N_c , large N_e) preserve genetic
92 diversity whereas populations experiencing strong declines and those with small N_e lose
93 genetic diversity more rapidly [23]. The assumption relevant to conservation genetics is
94 that the contemporary N_e (i.e., referring to recent generations) mirrors the current
95 conservation status of the population and informs on whether this remains large enough
96 to preserve genetic diversity and adaptive potential in the long term [22,24]. The problem
97 is that some populations are slow to respond!

98 Life-history traits specifically affect how N_e changes in relationship to census size (N_c) [25],
99 with adult life span/adult mortality, age at maturity and lifetime variance in reproductive
100 success having the greatest effects [26]. In particular, life span and age at maturity
101 determine generation time, which is positively correlated with N_e , whereas lifetime

102 variance in reproductive success is inversely proportional to N_e . Predicting the ultimate
 103 influence of the whole suite of life-history traits of a species on N_e is challenging, as many
 104 life-history traits will generally affect both generation time and lifetime variance in
 105 reproductive success in the same way, generating opposite effects on N_e .

106 Species with life-history traits favouring time lags include perennial, long-lived plants and
 107 other long-lived organisms such as reptiles or sea turtles that produce large numbers of
 108 un-nurtured offspring (thereby combining the long life spans characterising *K-strategists*
 109 with high offspring numbers characterising *r-strategists*). For example, a meta-analysis in
 110 plants revealed significant negative effects of recent habitat fragmentation on genetic
 111 diversity in herbs or short-lived plants but not in trees [27], suggesting that the longer
 112 generation time of trees (Box 1) delays the negative effects of habitat fragmentation.
 113 Indeed, other authors previously found that genetic diversity was lost proportionally to
 114 the number of generations since fragmentation in a study encompassing different plant
 115 life forms [18]. Another notable example of traits favouring time lags is the survival of
 116 individuals through seed banks (e.g. in annual plants or fire-adapted species), whose
 117 genetic diversity will reflect the population dynamics of previous generations (e.g. [5]).

118 Species that lack lag-favouring traits, instead, for example short-lived species and those
 119 that frequently experience demographic changes (most *r-strategists*), might more rapidly
 120 respond to contingent threats or they might face direct extinction without any warning
 121 signals of genetic erosion [19].

122 Other biological traits such as autopolyploidy can affect the persistence of polymorphism
 123 [28] and thus the build-up of time lags. Because of their higher number of orthologous
 124 gene copies, autopolyploid species lose genetic variation by drift more slowly than diploid
 125 species [29], and this reduced loss is also mediated by a larger N_e [28].

126

127 Table 1. Traits favouring persistence of polymorphisms and delayed genomic erosion despite deteriorated
 128 environmental conditions, i.e. time lags, and their relationship with N_e . The difficulty in predicting N_e changes
 129 will generally depend on the opposite influence of generation time and lifetime variance in reproductive
 130 success, which are in turn affected by other life-history traits.

Life-history traits favouring time lags	Role in favouring time lags	Relationship between life-history traits and N_e or N_e/N_c ratio	Key references
Long generation time, as a function of age at maturity, survival rate and age-specific fecundity; inverse function of annual mortality rate.	-Persistence of individuals and increased opportunity to reproduce: genetic diversity will reflect previous generations.	- N_e increases proportionally with generation time. General principle "lengthening the pre-reproductive period increases N_e ".	[30]
	-Age at maturity (one of the main determinants of generation time) will dictate	-Increased age at maturity increases both N_e and N_e/N_c .	[26]

	how fast the progeny representative of progressively eroded genetic diversity will reproduce, all else being equal.		
Overlapping generations/iteroparity/age structure	-Increased opportunities for reproduction across age groups: genetic diversity will partially or entirely reflect previous generations.	Overlapping generations generate lifetime variance in reproductive success, thus reducing N_e .	[31]
Long life span (longevity)/High survival rate	-Persistence of individuals and increased opportunity to reproduce: genetic diversity will reflect previous generations.	The increase in survival rate is associated with a reduction in N_e/N_C (counterbalanced by an opposite effect on N_e associated with a longer generation time).	[26]
-Clonal and partially clonal reproduction (with mechanisms different from vegetative growth) -Vegetative growth	-Persistence of individuals, increased opportunity to reproduce: genetic diversity will reflect previous generations. -As above, plus increase in physical size (with associated increase in organs for sexual reproduction).	-Same as for long lifespan and long generation time, relative contribution of other life-history traits is generally difficult to disentangle (see Outstanding questions). -As above. If some (larger) individuals will consistently reproduce more (sexually), N_e and N_e/N_C will be significantly reduced because of increased lifetime variance in reproductive success.	[32,33]
Mating system and dispersal strategy	-Outcrossing and long distance dispersal will promote population connectivity, buffering or delaying genetic erosion. -Selfing might initially favour a time lag, as individuals not affected by environmental changes will continue reproducing as before: genetic diversity in the progeny of selfed individuals will reflect previous generations. -Shift from predominant outcrossing to selfing will cause a rapid drop in genetic diversity	The interactive effect of mating system and other life-history traits on N_e is generally difficult to disentangle (see Outstanding questions). -Selfing decreases N_e .	[34] [30]
Large populations / distribution ranges	Large populations in large distribution ranges have a large reservoir of genetic diversity that can compensate for local genetic diversity losses.	Large populations have large N_e .	[35]

Large number of offspring reaching reproductive maturity	-Effective reproduction will tend to buffer or delay genetic erosion, and genetic diversity will reflect previous generations, at least initially. If many offspring are generated by parents whose genetic diversity is representative of the previous generation, it will take longer for genetic parameters to reflect new environmental conditions.	Mostly dependent on variance in reproductive success. More reproducing individuals will tend to even out variance in reproductive success, increasing both N_e and N_e/N_C . Few individuals generating large numbers of offspring will increase variance in reproductive success, decreasing both N_e and N_e/N_C .	[20,22,31].
-Seed banks (e.g. in annual plants) -Diapausing eggs (e.g. in freshwater crustaceans)	-Persistence of individuals, subject to successful germination/survival: genetic diversity will reflect previous generations.	Lengthening of the juvenile life-stage increases N_e ; analogously, lengthening mean seed dormancy increases N_e .	[5,30]
Other Biological traits or selective pressure potentially favouring time lags			
Balancing selection on adaptive loci*	Polymorphism can be maintained at adaptive loci that were under past balancing selection.		[36]
Inefficient directional selection* on putatively adaptive loci under long generation times	Slow responses to selective pressures generate time lags. In addition, other life-history traits can cause a cumulative effect in the build-up of time lags.		[37]
Autopolyploidy	Loss of heterozygosity (genetic diversity) is slower in autopolyploids and heterozygosity is higher at mutation-drift equilibrium compared with diploid populations	N_e is larger in polyploid populations.	[28]

*The effect of selection only on specific loci might be considered among "confounding factors" (Section 3), as genomic erosion can be actually detected if analysing other (neutral) regions. However, balancing selection has been included among the determinants of time lags, because it can induce a long-term persistence of polymorphism at the loci it acts upon.

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135 Selective pressures generated by environmental change may also affect the build-up of
136 time lags, since they affect individuals' likelihood of survival and reproduction. For
137 example, when locally adapted populations become maladapted, the time lag in the
138 genetic response can depend on a delayed response to selection because of lag-favouring
139 life-history traits [37]. Adaptive genetic responses to selection can happen within a few
140 generations and involve, for most traits, small allele frequency shifts at many, partially
141 redundant loci [38]. If environmental changes result in strong directional selective
142 pressure on a specific trait, the frequencies of the underlying alleles will show a faster

143 loss of diversity than the genomic background and may confound the detection of a
144 possible time lag affecting genome-wide diversity [39]. Conversely, loci under balancing
145 selection will show a slower loss of genetic variation than the genomic background [36].
146 While these processes affect all populations, selection is typically most efficient in large
147 populations, whereas genetic drift hampers its efficiency in small populations [40,41].

148 Table 1 includes additional ecological and biological traits favouring time lags, but more
149 research is required to understand the entire suite of life-history traits/factors that could
150 affect time lags and to disentangle their relative contribution.

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153 **Box 1 - Trees**

154 Studies on trees have provided a great insight into the persistence of genetic diversity
155 under deteriorated habitat conditions, as these species have many of the life-history
156 traits favouring time lags. Forest tree populations are characterised by high levels of
157 genetic diversity, much higher than e.g. herbaceous species [34,42]. Tree species are
158 mostly outcrossing, have a high life-time reproductive output, are subjected to strong
159 selection pressures during early life stages and they are particularly long-lived with
160 overlapping generations [43,44]. In natural undisturbed populations, the genetic diversity
161 of dominant tree species correlates positively with the surrounding species diversity (e.g.
162 [45,46]). However, while species diversity is lower in disturbed habitats, this is not
163 necessarily the case for genetic diversity, indicating non-parallel changes after
164 disturbance events [45] possibly due to time lags. The genetic response to disturbance
165 events such as logging, fire or dieback due to invasive pathogens depend (1) on the
166 strength/rate of population size decline, and (2) specific life history traits of the tree
167 species. In the case of extensive clear-cuts, the population size of all tree species declines
168 dramatically causing increased drift which affects allele frequencies in the natural
169 regeneration. Light-demanding and fast-growing pioneer species with efficient seed
170 dispersal emerge first and gain abundance in clear-cut sites while shade-tolerant, slow-
171 growing species emerge with delay and at lower densities making them more vulnerable
172 to genetic erosion especially in tropical forest ecosystems [47,48]. Silvicultural practices,
173 such as avoiding clearcuts, raising minimum logging diameters, and rotation lengths, can
174 attenuate these effects [49,50]. However, detecting recent genetic erosion in tree
175 populations is difficult, as remnant trees will reflect the genetic diversity of the previous
176 generation, as expected in species with traits favouring time lags. Similar effects are likely
177 in some marine fish, corals, sea grasses, other partially clonal species and in general
178 species with *K-strategy* life-history traits [51,52].

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180 **3. Factors confounding the interpretation of genetic studies when time lags occur**

181 When environmental changes occur, our ability to detect their impact on genetic diversity
182 might be confounded depending on our methodological choices. As we expect that the
183 changes will not affect all individuals simultaneously, depending on reproductive
184 strategies and on the occurrence of genetic structure within a population [53,54], the
185 sampling strategy adopted becomes a major determinant of the analytical outcomes (Fig.
186 1A). Vranckx and colleagues [8] observed that long-lived species in recently fragmented
187 ecosystems displayed contrasting patterns of genetic diversity (measured as expected
188 heterozygosity and percentage of polymorphic loci) between adults and progenies, with
189 the diversity in the younger cohort being more representative of the recent
190 fragmentation. Sampling juveniles of a long-lived, age-structured orchid after a recent
191 founder event, on the contrary, produced a larger N_e estimate compared with the adult
192 cohort from the same population [55], reflecting the recent population expansion and
193 ongoing gene flow.

194 The uncertainty associated with sampling design has been extensively discussed in
195 conservation and population genomics, with the consensus being that sampling for
196 analyses based on allele frequency calculations should be representative of the genetic
197 diversity of different individuals in the population, and large sample sizes are generally
198 needed to increase statistical power, especially in large populations (e.g., [56]). Rare allele
199 frequencies, in particular, are not correctly represented in small sample sizes, and this
200 will especially bias the estimation of demographic parameters. Furthermore, sample
201 sizes need to be similar when directly comparing different populations or cohorts.

202 As changes in allele frequencies may not be simultaneously reflected in the entire
203 genome, our ability to detect the signatures of genetic erosion will also depend on the
204 choice of molecular markers or genomic regions analysed per se (Fig. 1A; [57]), on
205 whether these genomic regions are under the effect of selection or not, and on the
206 metrics used to assess genetic erosion [58]. Genomic regions with higher mutation rates
207 (e.g., microsatellites) will exhibit higher indices of genetic diversity than regions usually
208 found in two allelic states (maximum expected heterozygosity equal to 0.5), such as SNPs.
209 Because of the impact of marker choice on the magnitude of the genetic metrics
210 obtained, some authors warned against the use of thresholds for genetic diversity
211 metrics in conservation [59].

212 Genic and adaptive regions under the effect of balancing selection are expected to be
213 more conserved than neutral regions and might remain in the same state even in
214 deteriorated environmental conditions [60]. Some authors have observed that genetic
215 diversity may remain high at loci under the past effect of balancing selection, despite an
216 overall loss of genetic diversity due to genetic drift [36,41]. They referred to this

217 phenomenon as “drift debt”, following the prediction that genetic diversity will eventually
218 be eroded if balancing selection will stop acting upon these loci.

219 Demographic processes might differentially be detected depending on the genetic
220 diversity metrics considered. Since rare alleles are lost first under population decline,
221 allelic richness, number of polymorphic loci and inbreeding coefficient respond more
222 quickly than heterozygosity to changes in population size. Heterozygosity, in particular, is
223 only affected to a little extent by short bottlenecks [17,61,62].

224 Complementary information such as sample coordinates can improve the interpretation
225 of genetic data. In large populations with effective gene flow and isolation by distance
226 (e.g., in trees), where N_e is difficult to estimate [63], recent demographic changes can be
227 captured based on spatial genetic parameters such as S_p [64], which is sensitive to
228 differential management and population dynamics [65].

229 Considering the factors confounding genetic interpretations under the occurrence of
230 time lags, it becomes obvious that relatively high levels of genetic diversity may reflect
231 past conditions, and that genomic erosion may occur with a delay [16].

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234 **4. A framework for interpreting genetic parameters despite time lags**

235 Regardless of the time elapsed between the environmental changes and the onset of
236 genetic erosion, the influence of confounding factors on genetic diversity can be
237 mitigated by satisfying some methodological requirements (Fig. 1B). Researchers should
238 adopt sampling strategies that are representative of the entire target population,
239 accounting for genetic differences between life-stages in species with overlapping
240 generations, and barriers to random mating. Most importantly, consideration of
241 population ecology, life-history traits and ploidy level is essential to interpret genetic
242 diversity and the possible occurrence of time lags. When the analysis of genome-wide
243 variation is not possible [66], analyses should target as many markers as possible,
244 covering different genomic regions. Multiple genetic metrics should be used to account
245 for their differential responses to demographic processes.

246 Provided that the requirements above are satisfied, we summarise three potential
247 approaches (Fig. 1C) that might help detect genomic erosion despite the occurrence of
248 time lags.

249 (1) The joint genetic analysis of samples from contemporary populations and samples
250 collected in the past (e.g., from herbaria, museums, and archaeological sites) is
251 one of the strategies to evaluate loss of genetic variation, through the estimation
252 of delta values of genetic diversity. Historical samples may provide baseline levels

253 of variation before the onset of the environmental changes causing genetic
254 erosion [14,62,67]. The main limitation of this approach is the availability of
255 temporal samples. In addition, temporal samples may be not representative,
256 considering past population dynamics and sampling strategies (although see [67]),
257 technical pitfalls such as post-mortem damage patterns, and genotyping errors
258 associated with depth of sequencing coverage [14].

259 (2) The comparison between historical and contemporary estimates of N_e [68] and N_c
260 may reveal differences in genetic drift over time. Because of the relative simplicity
261 of estimating both historical and contemporary N_e with samples collected in a
262 single point in time, these estimates can disclose loss of genetic variation when
263 other metrics may not. The inclusion of temporal sampling of populations, may
264 provide further analytical power to detect population genetic changes, although
265 researchers should be aware of the biases associated with each estimation
266 method [68].

267 (3) Metrics that focus on processes in contemporary generations potentially
268 mirroring recent environmental change include parameters of the mating system,
269 e.g., outcrossing rates, variance in reproductive success, dispersal kernels and
270 metrics on spatial genetic structure [65,69], as well as metrics summarising rare
271 allele frequencies such as allelic richness or site frequency spectrum [70].

272 (4) Comparison of genetic or genomic parameters of a population with those of a
273 large and stable reference population may provide a surrogate for baseline levels
274 of genetic variation. Although finding a reference population may be challenging
275 because of the spatial distribution of genetic diversity (e.g., range marginality) and
276 potentially different selective pressures, the intrinsic value of having a reference
277 population may aid the conservation of the most threatened populations.

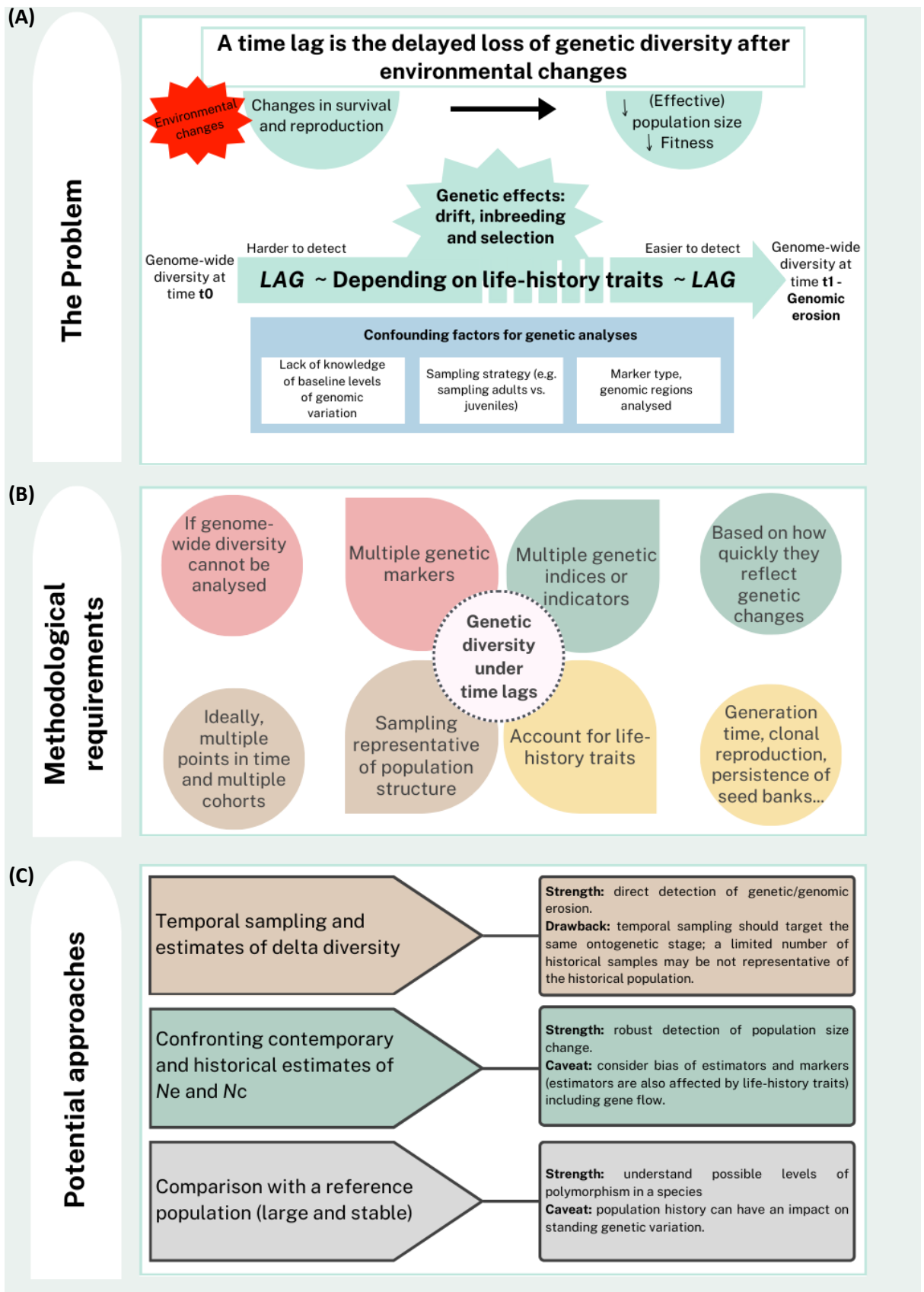
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279 ***Concluding remarks and Future Perspectives***

280 Time lags between environmental changes and the corresponding genetic changes is a
281 common but overlooked problem in ecology, evolutionary and conservation genetics.
282 With this opinion article, we offered an organic synthesis of the problem, of the potential
283 factors confounding the interpretations of genetic results, and of the possible
284 methodological approaches and solutions for a correct detection of time lags and
285 interpretations of genetic diversity levels in natural populations, especially those of
286 conservation concern. Our article also identifies **Outstanding questions** that deserve
287 exploration and open new avenues for the correct interpretations of genetic diversity
288 levels in natural populations despite the occurrence of time lags.

289 ***Declaration of interests***

290 The authors declare no competing interests.



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Figure 1. A framework to interpret and analyse delayed loss of genetic diversity after environmental changes and disturbance events. (A) The problem: genetic changes occur as a consequence of environmental changes. The main determinants of time lags include population size and life history traits, while the choice of genetic markers might mask the occurrence of a time lag or confound the

296 interpretation of genetic diversity. (B) Methodological requirements for the correct interpretation of
297 population genetic diversity under time lags. (C) Potential approaches to detect a time lag and correctly
298 interpret population genetic diversity. (B) and (C) also allow monitoring managed populations to assess
299 whether conservation interventions have been effective, when the life-history traits and reproductive
300 strategies of a species support the build-up of time lags.

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302 **References**

- 303 1. Schmidt, C. *et al.* (2023) Conservation macrogenetics: harnessing genetic data to meet
304 conservation commitments. *Trends Genet.* 39, 816–829
- 305 2. Epps, C.W. and Keyghobadi, N. (2015) Landscape genetics in a changing world: disentangling
306 historical and contemporary influences and inferring change. *Mol. Ecol.* 24, 6021–6040
- 307 3. IPBES (2019) *Summary for policymakers of the global assessment report on biodiversity and*
308 *ecosystem services*, Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem
309 Services - IPBES
- 310 4. Münzbergová, Z. *et al.* (2013) Historical habitat connectivity affects current genetic structure in a
311 grassland species. *Plant Biol.* 15, 195–202
- 312 5. Plue, J. *et al.* (2017) Does the seed bank contribute to the build-up of a genetic extinction debt in
313 the grassland perennial *Campanula rotundifolia*? *Ann. Bot.* 120, 373–385
- 314 6. Watts, K. *et al.* (2020) Ecological time lags and the journey towards conservation success. *Nat*
315 *Ecol Evol* 4, 304–311
- 316 7. Honnay, O. *et al.* (2006) Low impact of present and historical landscape configuration on the
317 genetics of fragmented *Anthyllis vulneraria* populations. *Biol. Conserv.* 127, 411–419
- 318 8. Vranckx, G. *et al.* (2012) Meta-analysis of susceptibility of woody plants to loss of genetic
319 diversity through habitat fragmentation. *Conserv. Biol.* 26, 228–237
- 320 9. Aavik, T. *et al.* (2019) Delayed and immediate effects of habitat loss on the genetic diversity of
321 the grassland plant *Trifolium montanum*. *Biodivers. Conserv.* 28, 3299–3319
- 322 10. Habel, J.C. *et al.* (2015) Fragmentation genetics of the grassland butterfly *Polyommatus coridon*:
323 Stable genetic diversity or extinction debt? *Conserv. Genet.* 16, 549–558
- 324 11. Essl, F. *et al.* (2015) Delayed biodiversity change: no time to waste. *Trends Ecol. Evol.* 30, 375–
325 378
- 326 12. Kuussaari, M. *et al.* (2009) Extinction debt: a challenge for biodiversity conservation. *Trends Ecol.*
327 *Evol.* 24, 564–571
- 328 13. Figueiredo, L. *et al.* (2019) Understanding extinction debts: spatio-temporal scales, mechanisms
329 and a roadmap for future research. *Ecography* 42, 1973–1990
- 330 14. Díez-Del-Molino, D. *et al.* (2018) Quantifying Temporal Genomic Erosion in Endangered Species.
331 *Trends Ecol. Evol.* 33, 176–185
- 332 15. Teixeira, J.C. and Huber, C.D. (2021) The inflated significance of neutral genetic diversity in
333 conservation genetics. *Proc. Natl. Acad. Sci. U.S.A.* 118
- 334 16. Kardos, M. *et al.* (2021) The crucial role of genome-wide genetic variation in conservation. *Proc.*
335 *Natl. Acad. Sci. U.S.A.* 118
- 336 17. Willi, Y. *et al.* (2022) Conservation genetics as a management tool: The five best-supported
337 paradigms to assist the management of threatened species. *Proc. Natl. Acad. Sci. U. S. A.* 119
- 338 18. Aguilar, R. *et al.* (2008) Genetic consequences of habitat fragmentation in plant populations:
339 susceptible signals in plant traits and methodological approaches. *Mol. Ecol.* 17, 5177–5188
- 340 19. Romiguier, J. *et al.* (2014) Comparative population genomics in animals uncovers the
341 determinants of genetic diversity. *Nature* 515, 261–263
- 342 20. Ellegren, H. and Galtier, N. (2016) Determinants of genetic diversity. *Nat. Rev. Genet.* 17, 422–
343 433
- 344 21. Cotto, O. *et al.* (2017) A dynamic eco-evolutionary model predicts slow response of alpine plants
345 to climate warming. *Nat. Commun.* 8, 15399
- 346 22. Waples, R.S. (2022) What is N_e , anyway? *J. Hered.* 113, 371–379
- 347 23. Frankham, R. (2019) Conservation Genetics. *Encyclopedia of Ecology* 382–390
- 348 24. Mastretta-Yanes, A. *et al.* (2023) Multinational evaluation of genetic diversity indicators for the
349 Kunming-Montreal Global Biodiversity Monitoring framework at
350 <https://ecoevorxiv.org/repository/view/6104/>
- 351 25. Charlesworth, B. (2009) Fundamental concepts in genetics: effective population size and patterns
352 of molecular evolution and variation. *Nat. Rev. Genet.* 10, 195–205
- 353 26. Waples, R.S. (2016) Life-history traits and effective population size in species with overlapping
354 generations revisited: the importance of adult mortality. *Heredity* 117, 241–250
- 355 27. González, A.V. *et al.* (2020) Meta-analysis of the differential effects of habitat fragmentation and
356 degradation on plant genetic diversity. *Conserv. Biol.* 34, 711–720
- 357 28. Moody, M.E. *et al.* (1993) Genetic variation and random drift in autotetraploid populations.
358 *Genetics* 134, 649–657
- 359 29. Monnahan, P. and Brandvain, Y. (2020) The effect of autopolyploidy on population genetic
360 signals of hard sweeps. *Biol. Lett.* 16, 20190796

- 361 30. Nunney, L. (2002) The effective size of annual plant populations: the interaction of a seed bank
362 with fluctuating population size in maintaining genetic variation. *Am. Nat.* 160, 195–204
363 31. Nunney, L. (1993) The influence of mating system and overlapping on effective population size.
364 *Evolution* 47, 1329–1341
365 32. Orive, M.E. (1993) Effective population size in organisms with complex life-histories. *Theor.*
366 *Popul. Biol.* 44, 316–340
367 33. Honnay, O. and Bossuyt, B. (2005) Prolonged clonal growth: escape route or route to extinction?
368 *Oikos* 108, 427–432
369 34. Hamrick, J.L. and Godt, M.J.W. (1997) Effects of life history traits on genetic diversity in plant
370 species. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 351, 1291–1298
371 35. Staude, I.R. *et al.* (2020) Range size predicts the risk of local extinction from habitat loss. *Glob.*
372 *Ecol. Biogeogr.* 29, 16–25
373 36. Gilroy, D.L. *et al.* (2017) Toll-like receptor variation in the bottlenecked population of the
374 Seychelles warbler: computer simulations see the “ghost of selection past” and quantify the “drift
375 debt.” *J. Evol. Biol.* 30, 1276–1287
376 37. Dauphin, B. *et al.* (2021) Genomic vulnerability to rapid climate warming in a tree species with a
377 long generation time. *Glob. Chang. Biol.* 27, 1181–1195
378 38. Barghi, N. *et al.* (2020) Polygenic adaptation: a unifying framework to understand positive
379 selection. *Nat. Rev. Genet.* 21, 769–781
380 39. Harpak, A. and Przeworski, M. (2021) The evolution of group differences in changing
381 environments. *PLoS Biol.* 19
382 40. Bijlsma, R. and Loeschcke, V. (2012) Genetic erosion impedes adaptive responses to stressful
383 environments. *Evol. Appl.* 5, 117–129
384 41. Ochoa, A. *et al.* (2020) Drift, selection and adaptive variation in small populations of a threatened
385 rattlesnake. *Mol. Ecol.* 29, 2612–2625
386 42. Chung, M.Y. *et al.* (2020) Incorporating differences between genetic diversity of trees and
387 herbaceous plants in conservation strategies. *Conserv. Biol.* 34, 1142–1151
388 43. Petit, R.J. and Hampe, A. (2006) Some evolutionary consequences of being a tree. *Annu. Rev.*
389 *Ecol. Evol. Syst.* 37, 187–214
390 44. Piovesan, G. and Biondi, F. (2021) On tree longevity. *New Phytol.* 231, 1318–1337
391 45. Wei, X. and Jiang, M. (2012) Contrasting relationships between species diversity and genetic
392 diversity in natural and disturbed forest tree communities. *New Phytol.* 193, 779–786
393 46. Raffard, A. *et al.* (2019) The community and ecosystem consequences of intraspecific diversity: a
394 meta-analysis. *Biol. Rev. Camb. Philos. Soc.* 94, 648–661
395 47. Akinagbe, A. *et al.* (2019) Towards conservation of genetic variation of tropical tree species with
396 differing successional status: the case of *Mansonia altissima* A. Chev and *Triplochiton*
397 *scleroxylon* K. Schum. *Tropical Conservation Science* 12, 1940082919864267
398 48. Kulevich, R.A. *et al.* (2020) Analysis of forests’ genetic vulnerability and arguments to reduce
399 deforestation. *Ambient. soc.* 23, e02222
400 49. Vinson, C.C. *et al.* (2015) Long-term impacts of selective logging on two Amazonian tree species
401 with contrasting ecological and reproductive characteristics: inferences from Eco-gene model
402 simulations. *Heredity* 115, 130–139
403 50. Roque, R.H. *et al.* (2023) Logging affects genetic diversity parameters in an *Araucaria*
404 *angustifolia* population: an endangered species in Southern Brazil. *For. Trees Livelihoods* 14,
405 1046
406 51. García-Castro, K.L. and Márquez, E.J. (2023) Temporal-scale assessment of population genetics
407 of the freshwater fish *Prochilodus magdalenae* in an area impacted by construction of a dam.
408 *Hydrobiologia* DOI: 10.1007/s10750-023-05396-z
409 52. Alvarado-Cerón, V. *et al.* (2023) A decade of population genetics studies of scleractinian corals:
410 A systematic review. *Mar. Environ. Res.* 183, 105781
411 53. Chikhi, L. *et al.* (2010) The confounding effects of population structure, genetic diversity and the
412 sampling scheme on the detection and quantification of population size changes. *Genetics* 186,
413 983–995
414 54. Mona, S. *et al.* (2014) Genetic consequences of habitat fragmentation during a range expansion.
415 *Heredity* 112, 291–299
416 55. Gargiulo, R. *et al.* (2023) Effective population size in a partially clonal plant is not predicted by the
417 number of genetic individuals. *Evol. Appl.* 16, 750–766
418 56. Buerkle, A.C. and Gompert, Z. (2013) Population genomics based on low coverage sequencing:
419 how low should we go? *Mol. Ecol.* 22, 3028–3035
420 57. Paz-Vinas, I. *et al.* (2021) Macrogenetic studies must not ignore limitations of genetic markers

- 421 and scale. *Ecol. Lett.* 24, 1282–1284
- 422 58. Leroy, G. *et al.* (2018) Next-generation metrics for monitoring genetic erosion within populations
423 of conservation concern. *Evol. Appl.* 11, 1066–1083
- 424 59. Zimmerman, S.J. *et al.* (2020) An empirical comparison of population genetic analyses using
425 microsatellite and SNP data for a species of conservation concern. *BMC Genomics* 21, 382
- 426 60. Koenig, D. *et al.* (2019) Long-term balancing selection drives evolution of immunity genes in. *Elife*
427 8
- 428 61. Allendorf, F.W. *et al.* (2022) *Conservation and the Genomics of Populations*, Oxford University
429 Press
- 430 62. Hoban, S. *et al.* (2014) Comparative evaluation of potential indicators and temporal sampling
431 protocols for monitoring genetic erosion. *Evol. Appl.* 7, 984–998
- 432 63. Santos-del-Blanco, L. *et al.* (2022) On the feasibility of estimating contemporary effective
433 population size (N_e) for genetic conservation and monitoring of forest trees. *Biol. Conserv.* 273,
434 109704
- 435 64. Vekemans, X. and Hardy, O.J. (2004) New insights from fine-scale spatial genetic structure
436 analyses in plant populations. *Mol. Ecol.* 13, 921–935
- 437 65. Bonnier, J. *et al.* (2023) Population genetic structure and demographic history of the timber tree
438 *Dicorynia guianensis* in French Guiana. *Tree Genet. Genomes* 20, 2
- 439 66. Hoban, S. *et al.* (2022) Global genetic diversity status and trends: towards a suite of Essential
440 Biodiversity Variables (EBVs) for genetic composition. *Biol. Rev. Camb. Philos. Soc.* 97, 1511–
441 1538
- 442 67. Wilder, A.P. *et al.* (2023) The contribution of historical processes to contemporary extinction risk
443 in placental mammals. *Science* 380, eabn5856
- 444 68. Nadachowska-Brzyska, K. *et al.* (2022) Navigating the temporal continuum of effective population
445 size. *Methods Ecol. Evol.* 13, 22–41
- 446 69. González-Martínez, S.C. *et al.* (2006) Effective gene dispersal and female reproductive success
447 in Mediterranean maritime pine (*Pinus pinaster* Aiton). *Mol. Ecol.* 15, 4577–4588
- 448 70. Excoffier, L. *et al.* (2013) Robust demographic inference from genomic and SNP data. *PLoS*
449 *Genet.* 9, e1003905