

1 **Opinions/Position Paper:**

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4 **Selection versus Transmission: Quantitative and Organismic**  
5 **Biology in Antibiotic Resistance**

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24 **ABSTRACT**

25 We aimed to determine the importance of selection (mostly dependent on the anthropogenic use  
26 of antimicrobials) and transmission (mostly dependent on hygiene and sanitation) as drivers of the  
27 spread of antibiotic-resistant bacterial populations. The first obstacle to estimating the relative  
28 weight of both independent variables is the lack of detailed quantitative data concerning the  
29 number of bacterial cells, potentially either pathogenic or harmless, and bacterial species exposed  
30 to antimicrobial action in the microbiotas of specific environments. The second obstacle is the  
31 difficulty of considering the importance of the various parameters influencing antibiotic resistance  
32 across eco-biological levels that constitute an “organism-like” entity. As a consequence, advances  
33 are urgently required in quantitative biology and organismic biology of antimicrobial resistance.  
34 The absolute number of humans exposed to antibiotics and the absolute number of potentially  
35 pathogenic and commensal bacteria in their microbiomes should influence both the selection and  
36 transmission of resistant bacterial populations. The “whole Earth” microbiome, with astonishingly  
37 high numbers of bacterial cells and species, which are also exposed to anthropogenic  
38 antimicrobials in various biogeographical spaces, shape the antibiotic resistance landscape. These  
39 biogeographical spaces influence various intensities of selection and transmission of potentially  
40 pathogenic bacteria. While waiting for more precise data, biostatistics analysis and mathematical  
41 or computational modeling can provide proxies to compare the influence of selection and  
42 transmission in resistant bacteria. In European countries with lower sanitation levels, antibiotic  
43 consumption plays a major role in increasing antibiotic resistance; however, this is not the case in  
44 countries with high sanitation levels. Although both independent variables are linked, their relative  
45 influence on the level of antibiotic resistance varies according to the particular location. Therefore,

46 although interventions directed to decrease antibiotic resistance should be designed “à la carte,”  
47 the global effects of antibiotic exposure and resistance on local sites cannot be underestimated.

## 48 **Introduction**

49 The classical approach to answering the question of why antibiotic-resistant microorganisms  
50 increase in frequency has been hospital-based, with the initial response based on selection. The  
51 more antibiotics that are used in an environment (independent variable), the more resistant  
52 organisms (dependent variable) that survive in comparison with the susceptible ones. However,  
53 we can easily conceive (and this has been observed in the field) that even in a location with low  
54 antibiotic use, antibiotic-resistant organisms might invade and propagate, particularly if conditions  
55 favor their transmission. Similarly, locations with high rates of antibiotic consumption might not  
56 result in high rates of antibiotic resistance if the conditions for transmission from individual to  
57 individual are limited; for instance, by extensive sanitation or by individual isolation. Any  
58 intervention attempting to correct the trend toward increased resistance should consider the local  
59 conditions; however, generalizations are frequently useless. Quantitation of the variable elements  
60 involved in selection and transmission of antibiotic-resistant microorganisms is in its infancy, and  
61 more studies should be performed (González-Candelas et al., 2011). In this regard, further  
62 quantitative biology research is needed to construct “precision” models adapted to the local  
63 conditions and to therefore foster useful interventions (Howard, 2014). We remain, however, “not  
64 so much concerned with the number of organisms, as such, as we are with another quantity”  
65 (Elsasser, 1964). Unfortunately, many of the parameters required to determine the precise “space  
66 and size of antibiotic resistance” remain poorly established (Martinez and Baquero, 2014). In this  
67 review, we suggest that progress in the quantitative biology of antibiotic resistance should be  
68 integrated into an organismic biology perspective. The concept of “organism” as an individual

69 entity composed of integrated functional parts (organ-like), was conceived by the anti-mechanistic  
70 philosopher John Scott Haldane in 1917 (Herring and Radick, 2019; Sturdy, 1988) and later applied  
71 by Ridley to general biology (see below). From this perspective, selection and transmission are  
72 probably the more general causal mechanisms explaining the effect we are targeting: the spread of  
73 antibiotic resistance in an “organism-like” entity constituted by the ensemble of bacterial and  
74 anthropogenic functions.

## 75 **Selection**

### 76 **1. Antibiotic consumption is a major driver for selection**

#### 77 **1.1. The absolute number of individual hosts exposed to antibiotics and the** 78 **absolute number of antibiotic-exposed potential pathogens**

79 Comparing rates of antibiotic consumption among countries is frequently based on the daily-  
80 defined-dose/1000 habitants per day, roughly reflecting the number of individuals exposed to  
81 antimicrobial treatment each day. However, the selective consequences of such exposure in a  
82 particular region depends on the absolute number of individuals exposed to antibiotics, or in other  
83 terms, the absolute number of bacteria of the various species located in these individuals’  
84 microbiota. Each 70 kg “reference” human carries approximately  $3.8 \times 10^{13}$  bacterial cells (Sender  
85 et al., 2016), an estimated ratio of 1.3 bacterial cells for every human cell in the body (Gilbert et  
86 al., 2018). In the same study, only in the gut, a rough estimate of 1000 bacterial species was  
87 proposed, with an average of 2000 genes per species, yielding approximately 2,000,000 genes, 100  
88 times the number of approximately 20,000 human genes. This distinction has been poorly  
89 explored, because it depends on the knowledge of the relative density of the various bacterial

90 populations. Even if we reduce this search to the more frequent antibiotic-resistant and potentially  
91 pathogenic (“antibiotic threat”) organisms in the intestine (such as *Escherichia coli* or  
92 *Enterococcus faecium*), the absolute density in the human or animal intestine is high and extremely  
93 variable, typically below 1% of the total human microbiota. This figure is consistent with  
94 quantitative polymerase chain reaction studies of bacteria adhered to gastrointestinal mucosa  
95 (Huijsdens, 2002). This proportion can increase in pathological conditions (including  
96 undernutrition), in which *E. coli* can increase by over  $10^5$  cells/ml in the upper gut, a volume  
97 roughly equivalent to the human large intestine. Despite its potential interest, there is scarce data  
98 on the total number of cells in the *E. coli* population (as an example) in human and animal  
99 microbiomes. Similarly, although the estimated number of cells in local environments (including  
100 sewage) remains poorly studied, it can be suggested that the number of cells/ml of many potentially  
101 pathogenic bacterial species in sewage does not differ significantly from the number in the large  
102 intestine in humans and animals (Silkie and Nelson, 2009). However, given that the absolute  
103 volume of contaminated water is extremely large, the number of cells provided by sewage is  
104 important for any estimation of the total number of potentially pathogenic individual bacterial cells  
105 in the species. The total number of bacterial cells on Earth (most with an “intrinsic resistome” is  
106 approximately  $5.10^{30}$  (<https://www.worldatlas.com/how-much-bacteria-is-on-earth.html>),  
107 constituting (after plants) the “heaviest” biomass on the planet, including approximately 70  
108 gigatons of carbon (Bar-On et al., 2018). In terms of “species,” or operational taxonomic units  
109 (OTUs) assembling closely related prokaryotic genomes, it has been estimated that there are more  
110 than 1 million (perhaps reaching 1.6 million) prokaryotic OTUs on the Earth (Louca et al., 2019).  
111 The distribution of these OTUs is environment dependent. However, a classic fundamental topic  
112 in ecology is the relationship between frequency (the proportion of samples where a particular

113 species appears) and population density (the number of individuals belonging to this species in a  
114 particular space). In principle, in a given environment, the denser the population the greater the  
115 frequency with which the species will appear, so that both parameters appear to be correlated (Dice,  
116 1948; Fisher, 1928). However, there are sampling biases due to the random distribution of some  
117 species, probably more in the minority ones. For instance, the absolute density (number) of  
118 potentially pathogenic intestinal bacteria in a given environment is dependent on 1) the density of  
119 these pathogens in the gut of species present in the environment, considering the volume of the  
120 colonized intestine in each species; 2) the local density of individuals of the various species; 3) the  
121 volume and periodicity of dropping fecal material; 4) the conditions for survival and growth of the  
122 pathogenic bacteria in the soil or water contaminated with fecal material and other fluids of human  
123 origin; and 5) the extension (volume) and physical structure of the contaminated environment. This  
124 last point will be developed in the next section. What we want to highlight here is the unimaginable  
125 number of global bacterial cells or species potentially exposed to anthropogenic antimicrobial  
126 agents, and therefore the vast opportunities to evolve to antibiotic resistance. Such an evolutionary  
127 trend could have consequences for human treatment of infections, but also for the equilibrium not  
128 only of the microbiosphere, but of the entire biosphere (Baquero et al., 2021).

## 129 **1.2. The space of antibiotic selection**

130 The space for antibiotic selection is the ensemble of bacterially colonized areas inside or outside  
131 the host where antibiotics can exert a selective effect, resulting in a local increase in antibiotic-  
132 resistant bacterial populations. This space is physically, chemically and biologically  
133 heterogeneous, thus the selective events in which these heterogeneities associate necessarily differ  
134 into subspaces.

### 135 **1.2.1. Gradients and granularity in selective spaces**

136 Physical differences that influence antibiotic activity include space compartmentalization,  
137 granularity (proportion of particles in the space), fluidity and viscosity, temperature differences  
138 and other physicochemical parameters. An important point is that these conditions determine the  
139 formation of more or less steep gradients of antibiotic concentrations, with various selective  
140 consequences. For bacteria, there are numerous possibilities for becoming antibiotic resistant.  
141 Many mutations allow the cells to resist small concentrations of antibiotics and to be selected by  
142 these concentrations. This selection increases the possibility of climbing up in the gradient and  
143 tolerating increasing antibiotic concentrations (Baquero and Negri, 1997). Very small antibiotic  
144 concentrations, far below the classic “minimal inhibitory concentration” used in susceptibility  
145 testing, are indeed very selective (Hughes and Andersson, 2012). Each subspace along the gradient  
146 where a particular concentration occurs might select for a particular resistant variant (Baquero et  
147 al., 1998; Negri et al., 2000). The space where low antibiotic concentrations occur is much larger  
148 (exponentially) than the space where the bacteria are exposed to high antibiotic concentrations;  
149 therefore, the absolute number of bacteria confronting a selective effect is considerably more  
150 abundant (Baquero and Coque, 2014).

151 Selection also benefits from the extension of environmental surfaces in the physical space where  
152 bacteria and antibiotics meet. Bacterial cells, as well as nutrient molecules, tend to accumulate on  
153 surfaces, particularly in particulate granular material in water and soil. This accumulation produces  
154 microbiotic particles where bacteria colonize the surface of physical granules (organic or  
155 inorganic), such as interbacterial aggregates, microfungus particles, protozoa, phytoplankton,  
156 zooplankton, biodetritus (such as plant remains), humus, mineral particles (clay, carbonates,  
157 silicates) and particles of anthropogenic origin, such as wastewater particles and microplastics

158 (Baquero et al., 2022). The density of bacterial populations in microbiotic particles depends on the  
159 size (i.e., diameter) of the basic particle and the environmental conditions allowing multi-layered  
160 bacterial multiplication. Antimicrobial agents also accumulate at the surface of these microbiotic  
161 particles, which can exert selection for antibiotic resistance.

### 162 **1.2.2. Antibiotic eco-pharmacokinetics and eco-pharmacodynamics**

163 The environmental variation in drug concentrations over time, and its biological effect on the  
164 components of ecosystems, are the essential parts of environmental pharma-ecology (Jjemba,  
165 2019). There is a potentially fertile microecological field of research on the kinetics of the  
166 antibiotic-bacteria interaction and antibiotic resistance selection in coalescent microbiotic  
167 particles. On one hand, bacteria frequently form more or less stable micro-biofilms after surface  
168 attachment (Puri et al., 2022), which is eventually followed by the shedding of planktonic bacteria.  
169 On the other hand, potentially selective antibiotic molecules might adsorb or de-adsorb from  
170 microbiotic particles and could also be detoxified when absorbed by clay minerals (degradative  
171 reactions influenced by light, metals or pH), charcoal, cellulose and chitin. Also can be sequestered  
172 by living and dead cells or cell components or biodegraded by microbial populations. Antibiotics  
173 have heterogeneous detoxification kinetics, and therefore their selective power for antibiotic-  
174 resistant populations might differ (Baquero et al., 2022). Once again, there are a lack of  
175 quantitative biological data (number and type of microbiotic particles, antibiotic kinetics and  
176 dynamics influencing the selective power for particular resistant bacteria) to help us understand  
177 the antibiotic selective processes.

### 178 **1.3. Does the increase in resistant populations replace the susceptible ones?**



179 Even after a very long period of exposure—more than a century—to antimicrobial agents around  
180 the world, and considering the high density and high replication rate of bacterial populations,  
181 resistant populations of pathogenic bacteria have not, with a few exceptions, replaced the  
182 susceptible ones. Once more, the lack of reliable data on the frequency of resistance (at least for  
183 the pathogenic bacteria) in non-exposed humans makes it difficult to estimate the size of the  
184 “susceptible reservoir,” a critical parameter to predict the possibility of interventions directed to  
185 the biorestitution of naturally susceptible organisms, at least in the Anthropocene Operating Space  
186 (Jorgensen et al., 2018). Fortunately, the environmental limits within which humanity can safely  
187 operate are limited, meaning that most antibiotic resistance genes in the microbiosphere are placed  
188 in non-pathogenic bacteria. Therefore, if antibiotics enter a particular environment (such as the  
189 intestine, or in water and soil environments), selection acts on the dominant non-pathogenic  
190 organisms, which can also protect the susceptible pathogens (for instance, the release of beta-  
191 lactamases by anaerobic bacteria). Most “antibiotic resistance genes” have not originated or  
192 evolved as genes to confer antibiotic resistance, but to serve other functions (the “intrinsic  
193 resistome”), antibiotic resistance being a “secondary phenotype.” Some of these genes can indeed  
194 occasionally be transmitted to kin pathogens. Given that resistance genes are listed without criteria  
195 in most databases, rules need to be established for estimating the risks associated with genes that  
196 are present in metagenomic resistomes by evaluating the likelihood of their introduction into  
197 human pathogens, as well as the consequences of such events for the treatment of infections  
198 (Martínez et al., 2015). On the other hand, every microorganism on Earth is subjected to  
199 environmental changes resulting in stress. The expression of antibiotic resistance, and the carriage  
200 of mobile genetic elements carrying antibiotic resistance genes, supposes an extra cost for  
201 individual resistant bacteria so that susceptible bacteria might have a competitive advantage over

202 the resistant ones. Long-term (decades) evolutionary experiments with a single *E. coli* in the  
203 absence of antibiotics show the progressive dominance of variant strains that are more sensitive to  
204 various antibiotics than their common ancestor(s) (Card et al., 2019). A possible way to accelerate  
205 this process and help susceptible bacteria prevail is to increase the fitness cost of resistant  
206 populations in the environment, including the gut (Baquero et al., 2024). Unfortunately, although  
207 the resistance fitness cost can be compensated by genetic variation, such variation might alter long-  
208 term bacterial physiology. In fact, the historical contingency of compensatory variation is a critical  
209 point in predicting the dominance of resistant pathogenic organisms (Card, 2019).

210

## 211 **2. Transmission in the spatial spread of antibiotic resistance**

212 Antibiotic resistance is a major health problem because bacteria that have been selected by  
213 antibiotic exposure are transmitted across various environments, allowing host-to-host direct or  
214 indirect transmission (Baquero, 2018). A host population with low rates of antibiotic consumption,  
215 and therefore low selective forces for antibiotic resistance, can be invaded by resistant bacterial  
216 clones for reasons unrelated to antibiotic use. Typically, this is the case (linked to extensive  
217 globalization) in countries with low antibiotic consumption, either because of stringent antibiotic-  
218 use policies (frequent in high-income countries) or due to drug unavailability (low-income  
219 countries).

### 220 **2.1. The causes of transmission**

221 Transmission is frequently a consequence of selection, ultimately on the local selective  
222 reproduction of resistant cells. Only if a critical number of antibiotic-resistant cells accumulate in  
223 a given spatial site can transmission occur, propagating these cells to other spatial locations.

224 However, successful transmission from a source can only be achieved if the antibiotic-resistant  
225 cells can multiply in the receiving patch. This dynamic has been likened to information theory as  
226 requiring “emitters” and “receivers” (Baquero, 2018). Transmission, or emission, requires leaving  
227 the niche of the original population and seeking other places for similar niches to be colonized. In  
228 such a process, resistant bacteria most frequently are obliged to cross “death valleys” in the  
229 adaptive landscape, where replication is difficult or impossible; therefore, the population density  
230 decreases and consequently the transmission might be aborted. Therefore, a significant initial  
231 density (as in the case of plant seeds, or fish eggs) is required. If such density is not achieved, or  
232 if we increase the difficulties for survival in “death valley,” the spread of resistance will be reduced.  
233 An important point is that natural “empty niches” to be colonized are rare or nonexistent. This lack  
234 of empty niches implies that antibiotic-resistant populations should “invade” areas (patches, in  
235 terms of metapopulation biology) containing high-carrying capacity niches that are already  
236 colonized, as occurs in the human or animal intestine. How resistant bacteria at very low  
237 population sizes can invade heavily colonized areas (established microbiotas) in a new host in the  
238 absence of selection is a fundamental area of research that has, so far, scarcely been investigated.  
239 There are some possible answers.

240 **First**, there could be a pre-colonization area allowing multiplication of the resistant population,  
241 either in the environment (e.g., contaminated food or water), in the host itself (e.g., mouth, upper  
242 intestine), or in medical devices (e.g., ventilators in the hospital’s intensive care units). The hospital  
243 setting can be invaded by clones that have previously successfully spread in the community,  
244 associated or not with disease (“hidden epidemics”), reaching high numbers in hosts outside  
245 hospitals (Rodríguez et al., 2021). Certainly, the “absolute population size” of resistant organisms  
246 exponentially increases during epidemics, so that epidemic events feed its propagation.

247 **Second**, the antibiotic-resistant bacteria might be endowed with “attack mechanisms” to replace  
248 resident bacteria (frequently phylogenetically close), as in the case of the very spreadable *E. coli*  
249 phylogroup B2, in which several high-risk clones (such as 025b-ST131), carry antibiotic peptides  
250 (microcins) (Micenková et al., 2017). In their turn, resistant clones that have succeeded in the  
251 colonization process use their former attack mechanisms as defense mechanisms to protect their  
252 conquered niche from re-invasion in formerly established non-resistant clones. Another possibility  
253 derives from the changes in the microecology of the gut, resulting from pathogenic effects on the  
254 host. A resistant pathogenic organism in the intestine, even at low densities, might produce reactive  
255 changes in the host, altering intestinal signals as well as secretion of antibacterial substances of  
256 innate immunity and changing the populational composition of the healthy microbiota so that the  
257 new colonizer gains in relative fitness (Chowdhury et al., 2023). Also, if the resident community  
258 is already disturbed by principally antibiotic medical interventions, the colonization resistance  
259 capacity (due to the unavailability of nutrients for the incoming organisms) is diminished, favoring  
260 colonization by this antibiotic-resistant colonizer (Pérez-Cobas et al., 2023; Spragge et al., 2023).

261 **Third**, small quantities of antibiotic-resistant bacteria might invade areas previously colonized by  
262 susceptible populations following cumulative events of transmission. Continuous contact of hosts  
263 colonized with susceptible bacteria with other hosts carrying antibiotic-resistant populations leads  
264 to a final cumulative-competitive number of resistant organisms. That is the case of transmission  
265 within families in which one of the members is carrying a resistant clone (Valverde et al., 2008),  
266 or of the farmers and veterinarians colonized by antibiotic-resistant animal strains (Gao et al.,  
267 2024). A similar case is human individuals chronically exposed to water or food contaminated by  
268 antibiotic-resistant organisms in low-income countries. Interestingly, this exposure might also  
269 occur in high-income countries; intensive care units in hospitals might contain “built

270 environments” where antibiotic resistant clones might persist for long periods of time (Aracil-  
271 Gisbert et al., 2024).

272 **Fourth**, the success of the spread of antibiotic resistance is also due to a process of “conversion”,  
273 in which a particular resistant clone is transmitted to a particular patch containing antibiotic  
274 susceptible strains, “converting” these strains into resistant ones by horizontal gene transmission,  
275 most frequently mediated by conjugative plasmids carrying antibiotic resistance genes (Castañeda-  
276 Barba et al., 2024). In the absence of antibiotic selection this process of emergence of new resistant  
277 organisms, most frequently among commensals (Dionisio et al., 2023), entirely depends on the  
278 spontaneous plasmid conjugation rate. Computational simulations indicate that only high rates of  
279 transmission (such as  $10^{-3}$ ) might have a small transient effect on the acquisition of resistance  
280 (Campos et al., 2020). However, this process of genetic transmission might be facilitated by  
281 conditions in which susceptible and resistant bacteria enter into contact, such as in microbiotic  
282 particles, those present in the gut, or in free environments (see 1.2.1.).

## 283 **2.2. Transmission barriers**

284 There are several barriers to reducing the spread of antibiotic resistance in human and animal  
285 environments.

286 **First**, appropriate general sanitation. If resistant organisms colonizing a host are prevented from  
287 having access to another host, transmission does not take place. In the case of the main pathogenic  
288 intestinal microorganism, inadequate access to water, sanitation and hygiene is a key component  
289 of the spread of resistance in lower- and middle-income countries (Walsh et al., 2023; Nadimpalli  
290 et al., 2018). Water and food contamination with resistant organisms facilitates human and animal  
291 colonization, and following vicious circle dynamics, such colonization is followed by

292 environmental contamination. In the hospital setting, barriers impeding cross-colonization  
293 between patients, involving health care workers (including hand-washing) and medical device  
294 decontamination or preventing access to colonized built environments (see 2.1), reduces the spread  
295 of antibiotic resistance.

296 **Second**, vaccination against antibiotic-resistant clones. By far, the most effective documented  
297 intervention in the fight against antibiotic resistance has been vaccination against antibiotic-  
298 resistant clones of *Streptococcus pneumoniae* or *Haemophilus influenzae*. Developments in this  
299 field, encompassing antibiotic resistant Gamma-Proteobacteria (mostly Enterobacteriaceae), or  
300 Bacillota (mostly *Staphylococcus*, *Enterococcus*) high-risk clones are in the pipeline (Frost et al.,  
301 2023). The efficacy of vaccines might differ when targeting other pathogens; however, even  
302 imperfect vaccines might be useful in decreasing antibiotic resistance (Joice and Lipsitch, 2013).  
303 In any case, a limitation of this type of intervention is derived from the fact that many antibiotic-  
304 resistant high-risk clones are also commensal antibiotic-susceptible clones, coexisting and  
305 coevolving with humans, and thus part of the “normal microbiota.” If vaccination contributes to  
306 their clearance (Kubinak and Round, 2016), the consequences are difficult to predict. Note that the  
307 spread of antibiotic-susceptible clones is competing with the spread of resistant ones.

308 **Third**, boosting the selection of antibiotic-susceptible clones. The ideal “magic intervention” to  
309 reduce or eliminate antibiotic-resistant clones is to facilitate the selection of antibiotic-susceptible  
310 kin organisms. This biorestorative task can be approached in various ways. One is based on  
311 microbiota transplantation with feces of antibiotic resistance-free individuals (Woodworth et al.,  
312 2023). In general, ensuring a high biodiversity in the microbiota, resulting in a wealth of  
313 interbacterial interactions, should reduce selection for antibiotic resistance (Nair and Andersson,  
314 2023). Another possibility (proposed, not explored) is taking advantage of the fitness costs

315 associated with the expression of antibiotic resistance, which might be additive (synergistic in  
316 some cases?) with the “natural stresses” of the bacterial cell during the passage or stay in the  
317 intestine. Boosting such stresses might favor antibiotic-susceptible clones (Baquero et al., 2024)

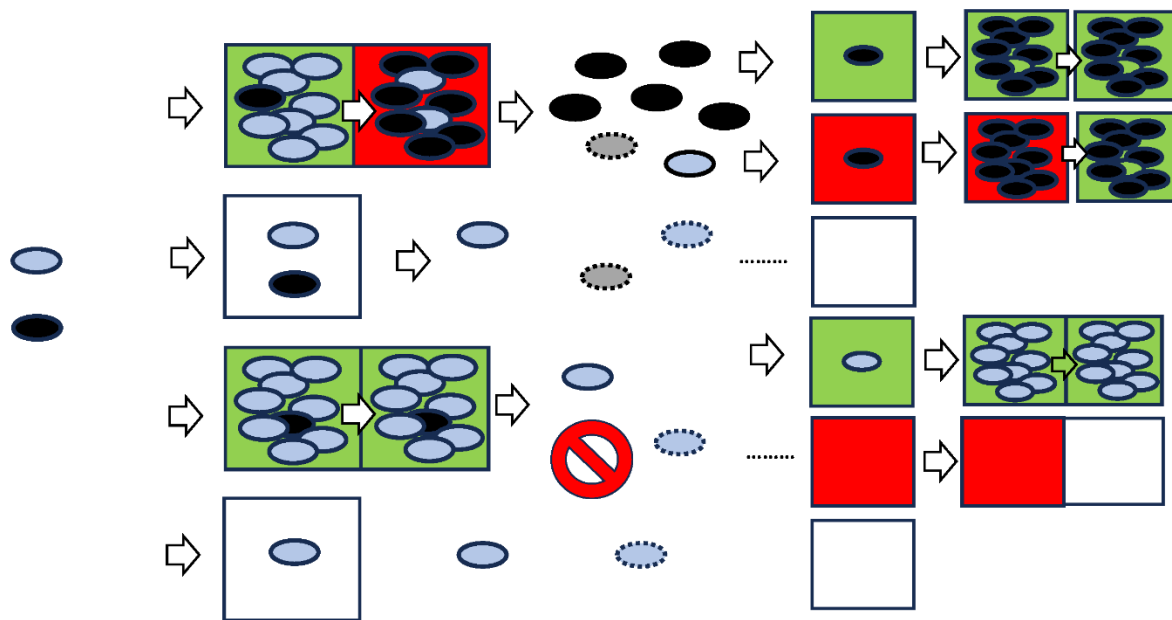
318

## 319 **Organismic biology and antibiotic resistance**

320 As stated earlier, the notion of organismic biology is probably on the grounds of the contemporary  
321 One Health approach. However, this concept was coined more than a century ago, in 1919, by the  
322 zoologist W.E. Ritter, following the track of John Scott Haldane; he proposed that “the organism  
323 in its totality is as essential to an explanation of its elements as its elements are to an explanation  
324 of the organism” (Ritter, 1919; Herring and Radick, 2019). Around the middle of the last century,  
325 the term “organismic biology” was contrasted with pure “mechanistic biology” (Nagel, 1961;  
326 Elsasser, 1964; Milam, 2010). An organism is an “individual” that can be composed of other  
327 “individuals” of lower hierarchical ranges. In evolutionary theory, every individual is a “unit of  
328 selection” (Baquero, 2011). For the case we are considering, we can conceive a bacterial clone of  
329 a given antibiotic-resistant pathogen as a biological individual, composed of many bacterial cells,  
330 some of them genetic variants of their ancestors, so that the clone is a “clonal complex” acting as  
331 a single individual.

332 The biological functions *constitute* the organism as a biological individual, and therefore functions  
333 are the effectors determining paths and trajectories of evolution (what reproduces, what evolves)  
334 (Baquero, 2021) and transmission (what is transmitted) (Baquero, 2017). These are the basic  
335 functions shaping a biological individual: reproduction and transmission, respectively representing  
336 the biological conquest of the time and space dimensions. Necessarily, these basic functions are

337 associated: only if an organism reproduces does transmission take place; however, transmission  
 338 assures the exploration of new spaces where new reproductive bursts can occur, overcoming local  
 339 environmental instabilities. In a sense, this appears to be a case of mutual causation, as in Lois  
 340 Frankel’s metaphor of the “King and Queen of Hearts,” playing cards that lean against one another,  
 341 forming a simple card house. If either moves, the other will fall (Frankel, 1986). To ensure  
 342 continuous adaptation to environmental variation, the organism either *changes its structure* to be  
 343 selected (as a genetic or a phenotypic variant) or *changes its location* in the space. In such a way,  
 344 we can compare transmission with genetic variation. Both are basic organismal functions that  
 345 justify their functional unity under the organismic biology perspective (Pepper and Herron, 2008).  
 346 A schematic illustration of this is presented in Figure 1. As in the “King and Queen” cards  
 347 metaphor, efficient interventions against either selection and transmission of antibiotic resistant  
 348 organisms (or both) should collapse the house of antibiotic resistance.



349



350 **Figure 1. Selection and transmission.** Square boxes represent ecological patches with different  
351 carrying capacities (conditions for bacterial exploitation): low (white), high (green, such as  
352 mammalian hosts, or sewage in warm-humid countries), or high but non-permissive (red, such as  
353 hosts or sewage with antimicrobial agents). White arrows are symbols for transmission, and the  
354 no-entry traffic sign represents high sanitation reducing transmission between patches. Light blue  
355 and black ovals, respectively, represent antibiotic susceptible (S) and resistant (R) bacteria. The  
356 **first row** depicts a patch highly colonized with S organisms with a minority of Rs. If this patch is  
357 in close contact with another one, this time under high antibiotic exposure, bacterial populations  
358 are transmitted and invade, but selection for Rs takes place. The high density of R populations  
359 assures a more efficient transmission in the inter-patch environment so that these Rs reach and  
360 invade other remote patches, either under the absence (green) or presence (red) of antibiotics. In  
361 the **second row**, S or R organisms enter into a low carrying capacity patch (white), so that  
362 reproduction is limited, and therefore organisms transmitted to remote patches, such as bacteria,  
363 die (discontinuous oval) during the transmission process or are too diluted to invade. **In the third**  
364 **row**, S-bacteria invade a patch with high carrying capacity, which transmits the same population  
365 to another neighbor patch. High densities of Ss reached in the patches might eventually overcome  
366 sanitation barriers, reaching new colonizable patches (green), thus expanding the S bacteria.  
367 Although they might not reach patches with antibiotics, if that were the case, they would become  
368 extinct. The possibility of reaching such spaces for R organisms is negligible with good sanitation.  
369 In the **fourth row**, high sanitation makes the transmission of organisms with low population  
370 densities almost impossible.

371

372

373 Let's now move upward. The concept of organismic biology expands the notion of "organism" to  
374 ensembles of individual entities. If bacterial cells compose the clonal complex "organism" or  
375 species, then ensembles of interacting organisms constitute the microbiota "organism." The notion  
376 of "intestinal microbiota as a human organ" had long ago been proposed (Baquero and Nombela,  
377 2012). Can we conceive an ensemble of interacting microbiotas of human individuals (and health

378 workers?) and the microbiotas of the “built environment” from a hospital intensive care unit (ICU)  
379 as an organism, or just a particular ecosystem? It is certainly an ecosystem, with some organismic  
380 features, such as the permanence in time of many bacterial organisms for years despite the flux of  
381 patients, and its reproduction in analogous environments (such as a new ICU), which implies the  
382 effect of similar selective and transmission dynamics (Aracil-Gisbert et al., 2024). From an  
383 operational perspective, aiming to detect global changes in antibiotic resistance genes present in  
384 the ICU over time, we can consider an “ICU global intestinal microbiota” composed of all  
385 intestinal microbiota from patients attended during a given time. This consideration explores the  
386 evolution in time of antibiotic resistance by “pooling” procedures (mixing samples from the  
387 patients), to be used in metagenomic studies (Aracil-Gisbert et al., 2022).

388

## 389 **Selection versus transmission: quantitative biology and modeling**

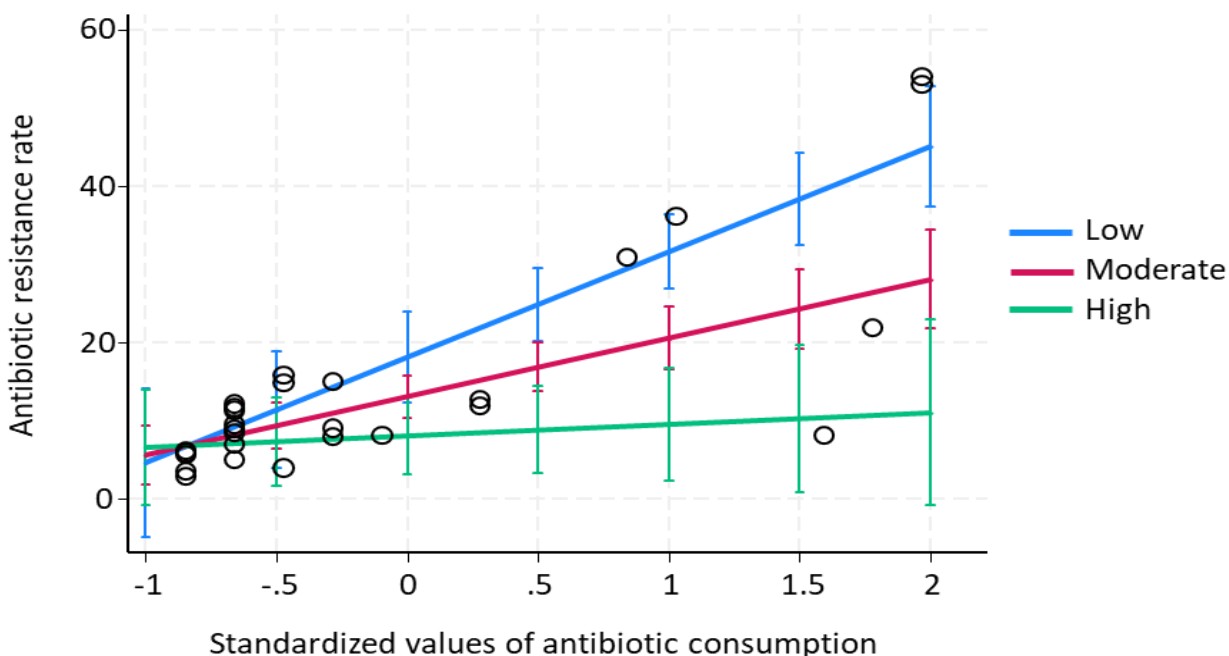
### 390 **approaches**

391 In the previous sections, we noted the need for more precise quantitative experimental data to  
392 understand the dynamics of antibiotic resistance. However, we also mentioned the overwhelming  
393 complexity of the task of defining the parameters shaping the various selective and transmission  
394 spaces (Martínez and Baquero, 2014). Most importantly, we are very short of knowing the critical  
395 issues to determine; for instance, the total number of bacterial cells of a given species or particular  
396 clones in a given population, the changes in the diversity of bacterial subpopulations or the size of  
397 the selective and transmission exposome (Wei et al, 2022). We need to know (or to realistically  
398 estimate), the absolute number of clinically relevant resistant bacterial populations, versus the  
399 absolute number of resistant populations rarely associated with infections or the absolute number

400 of individuals hosting resistant bacteria in a particular environment. Such an ambitious task should  
401 be studied in well-described ecological systems and followed over time. We need to expand the  
402 quantitative biology of antibiotic resistance based on a deeper analysis of the microbial  
403 composition of our colonized habitat. Similarly, the quantitative effects of interventions, such as  
404 specific restrictions on the use of particular antibiotics in a hospital to decrease antibiotic  
405 resistance, are based on low-quality evidence (Chatzopolou and Reynolds, 2020). Another case is  
406 the quantitative insufficiency of data on the transmission of antibiotic resistance from the  
407 environment to humans, given that the abundance of resistant bacteria and antibiotic resistance  
408 genes does not necessarily provide accurate risk assessment (Manaia, 2017).

409 We can argue that, as in the case of quantitative cell biology, quantitative environmental biology  
410 should not only be oriented to obtain precise observational and experimental results but to test the  
411 plausibility of testing theories using statistical, mathematical and computational models (Howard,  
412 2014). An example of this approach is the application of membrane computing procedures to  
413 predict the effect of changes in selection (antibiotic consumption) and transmission (host-to-host  
414 cross-contamination with resistant organisms) (Campos et al., 2019). This computing methodology  
415 (derived from natural computing) considers the elements of the system (such as susceptible and  
416 resistant bacteria, antimicrobial agents, mobile genetic elements carrying antibiotic resistance and  
417 infected or colonized hosts) as “objects” defined by virtual membranes. Each object can be  
418 endowed with deterministic or stochastic rules by which they can replicate, propagate, dissolve  
419 and be transmitted and selected when interacting with another “object”. The application of this  
420 model to the selection-transmission dynamics of antibiotic resistance indicates that in hospitalized  
421 patients, reductions in antibiotic use are much more effective for preventing the rise of antibiotic  
422 resistance in the intestinal microbiota than reductions in host-to-host transmission. A possible

423 interpretation is that high transmission rates *also* facilitate the spread of antibiotic-susceptible  
 424 bacteria. If antibiotic consumption (selection) is high, resulting in high rates of antibiotic  
 425 resistance, a decrease in transmission will be highly effective. For example, using publicly  
 426 available datasets (Wolf et al., 2022; Versporten et al., 2021), Figure 2 illustrates the interaction  
 427 between selection (antibiotic consumption) and transmission (sanitation, hygiene) in 5 European  
 428 Regions (Mediterranean, North, West, East and Central Europe) countries, shaping the local  
 429 frequency of third-generation cephalosporin resistance in *E. coli*. This figure reveals a dynamic  
 430 and complex relationship regarding the drivers of antibiotic resistance. The extent of the  
 431 association between increased antibiotic consumption and higher antibiotic resistance is influenced  
 432 by the hygiene index of the countries composing a region (circles in the figure). Both variables  
 433 were standardized to be compared. Regions with suboptimal hygiene indices experience a more  
 434 pronounced rise in antibiotic resistance with increased consumption. Conversely, in regions with  
 435 better hygiene indices, the impact of antibiotic consumption on resistance is mitigated.



436

437 **Figure 2. Antibiotic consumption versus sanitation.** Influence of low, moderate and high levels  
438 of sanitation in various European countries on the effect of antibiotic consumption (third-  
439 generation cephalosporins) in *Escherichia coli* resistance to cefotaxime-ceftriaxone. In countries  
440 with low sanitation levels, antibiotic consumption plays a major role in increasing antibiotic  
441 resistance.

442

## 443 **The biogeographical patterns of selection and transmission: the** 444 **anthropogenic causation of resistance exposome**

445 As discussed previously, both selection and transmission of antibiotic resistance are mutually  
446 synergistic in the emergence and spread of antibiotic resistance. The weight of such synergy  
447 depends on the intensity of each one of these variables in particular environments, and the time  
448 during which selection and transmission are exerting their effects in the microbiosphere, the  
449 “exposome” temporal dimension (Wei et al., 2022). The “cumulative history of antimicrobial use”  
450 over the last century (or more if heavy metal-based anti-infectives were considered) in a particular  
451 location could be critical to understanding the present scenarios (Coque et al., 2023). However, the  
452 current rates of antibiotic consumption and the rates of sanitation widely vary in different parts of  
453 the world, and therefore we could predict to a certain extent the main biogeographical areas where  
454 antibiotic resistance will emerge and spread. The main reason for the various combinations of  
455 selection and transmission is anthropogenic, encompassing economy and culture, which in turn  
456 are tightly associated. In other words, inequality is the main difficulty in designing interventions  
457 directed at limiting antibiotic resistance (Nadimpalli et al., 2021). As stated in a previous  
458 paragraph, areas with low antibiotic consumption (low selection) and high sanitation (low

459 transmission) have lower antibiotic resistance rates, followed by those with low consumption  
460 because of poor accessibility. However, the resistance burden in areas with medium-high  
461 consumption depends on the rate of sanitation (particularly water and soil fecal pollution); high  
462 antibiotic consumption and low sanitation provide the “perfect storm” for antibiotic resistance. As  
463 stated previously, these conditions depend on the economic background of the various areas, which  
464 influences and is influenced by the sociocultural landscape. For instance, the application of the  
465 Inglehart-Welzel World Cultural Map to the consumption of antibiotics reveals that countries with  
466 more “traditional” versus “secular” values and “survival” versus “self-expression” values tend to  
467 have higher rates of antibiotic consumption, except when accessibility to drugs is very limited  
468 (Dionisio et al., 2023). With this exception, low sanitation probably also correlates with  
469 “traditional” and “survival” cultural dimensions. As is clear, climatology influences the possibility  
470 of economic development of particular countries; in warmer-humid countries, the persistence of  
471 antibiotic resistance in the environment is probably longer (MacFadden et al., 2018; McGough et  
472 al., 2020). Concerns about the effect of global warming, increasing transmission of antibiotic  
473 resistant pathogens and the number of antibiotic-treatable infections (increasing selection) are well  
474 justified. These changes also influence the extensive international exchange of human and animal  
475 hosts, food, and also geological phenomena, such as oceanic currents (Zhang et al., 2012),  
476 contributing to the Earth's dispersion of antibiotic resistant populations. We can conclude that  
477 interventions directed to decrease antibiotic resistance should be designed “à la carte” (Rahbe et  
478 al., 2023), but the global effects of antibiotic exposure and resistance on local sites cannot be  
479 underestimated.

480

481

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488

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