

# How bottlenecks shape adaptive potential: from theory and microbiology to conservation biology

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## Abstract

1  
2 Wild populations frequently undergo demographic changes that can challenge their  
3 persistence and, thus, the equilibrium of ecosystems. For instance, habitat fragmentation  
4 due to human activity leads to a drastic population size reduction, a process called a  
5 bottleneck. By reducing genetic diversity, a bottleneck may prevent a population from  
6 adapting to subsequent environmental changes. In the context of climate change, it is  
7 crucial to accurately predict how a wild population evolves after a bottleneck and how it  
8 affects its persistence. Mathematical models have provided valuable insights into the im-  
9 pact of bottlenecks on the adaptive potential of populations. However, their application to  
10 wild populations requires further improvement as these theoretical predictions have been  
11 mostly experimentally tested with microbial populations. Thus, it remains unclear what  
12 the implications of the theoretical predictions are at the macroscopic scale, although these  
13 predictions are crucial for conservation biology. This review aims to determine how the  
14 knowledge acquired through evolutionary theory and experimental microbiology applies  
15 to wild populations. To achieve this aim, we address the following questions: (i) What  
16 do theory and microbiology experiments tell us about the impact of bottlenecks on the  
17 ability of populations to adapt to future environmental changes? (ii) Do these theoretical  
18 predictions apply to wild populations? and (iii) What is missing to better predict the evo-  
19 lution of wild populations after a bottleneck? We analyze how the four main evolutionary  
20 processes (i.e., mutation, genetic drift, natural selection, and gene flow) impact the fate  
21 of populations facing bottlenecks. By linking theory, microbial experiments, and empiri-  
22 cal studies on natural populations, we identify research directions that could help manage  
23 populations undergoing bottlenecks and plead for increased communication between these  
24 fields.

25 **Keywords:** Bottleneck, Adaptive potential, Demography, Evolutionary rescue, Population  
26 size, Stress, Fragmentation, Mathematical models, Microbiology, Conservation biology

## 27 **1 Introduction**

28 Most natural populations experience bottlenecks that can be caused, for example, by severe  
29 climate events, habitat loss, and overhunting (Lande, 1988; Frankham et al., 2002). The  
30 bottleneck-induced population size reduction increases the extinction risk of populations and,  
31 thus, may destabilize ecosystems (Frankham et al., 1999). As the current intensive anthro-  
32 pogenic perturbations of Earth’s systems are increasing the occurrence of bottlenecks (Barnosky  
33 et al., 2011; Ceballos et al., 2015), there is an urgent need to understand how bottlenecks impact  
34 the evolutionary fate of at-risk populations. This review examines the evolutionary fate of pop-  
35 ulations undergoing sudden random decreases in size that are selectively neutral, as opposed to  
36 those caused by directional selection. Specifically, this review focuses on bottlenecks involving  
37 random reductions in population size rather than selective bottlenecks involving non-random  
38 reductions in population size (but see section 4.2).

39 Forecasting the effects of population bottlenecks is crucial in conservation biology (Frankham  
40 et al., 2002). However, our ability to predict the long-term consequences of bottlenecks under  
41 natural conditions remains weak. First, predictions in the wild are generally made retrospec-  
42 tively, i.e., after a bottleneck has occurred and the population has gone extinct or survived  
43 (Bouzat, 2010). Second, predictions are often made case-by-case, preventing their applicability  
44 to other systems. To fill this knowledge gap and improve our understanding of bottlenecks,  
45 we need to develop a comprehensive overview combining theoretical predictions and empirical  
46 evidence. Although connections between fundamental evolutionary biology and wildlife conser-  
47 vation have slowly developed, they are increasingly strengthening, highlighting their importance  
48 (Hohenlohe et al., 2020).

49 Microbiology is a field that allows for experimentally assessing the impact of bottlenecks  
50 on the adaptive potential of microbial populations (LeClair and Wahl, 2017). A common  
51 experimental evolution technique is subjecting micro-organisms to serial passaging, which in-  
52 volves repeated bottlenecks. In such experiments, a microbial population is inoculated into a  
53 medium and grows. Then, the experimenter takes a fraction of this population, inoculates a  
54 new medium, and repeats the process several times. This common technique explains why the  
55 literature in experimental microbiology has so well documented the impact of bottlenecks on  
56 the adaptation of microbial populations (LeClair and Wahl, 2017). Micro-organisms, such as  
57 bacteria and fungi, can reproduce rapidly and reach large numbers in small spaces, allowing for  
58 highly replicated experiments and, therefore, highly accurate predictions (Elena and Lenski,  
59 2003). In summary, the simplicity of these experiments makes microbiology an excellent field  
60 for testing theoretical predictions.

61 To the best of our knowledge, no study has yet bridged the gap between what microbi-  
62 ology tells us about the impact of bottlenecks on the adaptive potential of populations and

63 conservation biology. The lack of direct links between both fields likely results from the many  
64 differences between microbial populations in controlled laboratory experiments and wild pop-  
65 ulations in natural ecosystems [see Box 4 in Kawecki et al. (2012)]:

- 66 1. Microbes used in experiments substantially differ from endangered natural species tar-  
67 geted by conservation efforts, mostly diploid and sexual.
- 68 2. The demography of microbial populations studied in laboratory conditions also differ  
69 from that of wild populations. For example, the size of microbial populations is likely  
70 larger than that of wild populations. As a result, the genetic load is likely higher in wild  
71 populations than in laboratory populations.
- 72 3. Bottlenecks in the wild likely vary in intensity and frequency, whereas they are typically  
73 periodic in microbiology experiments.
- 74 4. The number of generations before adapting to a new environment differs for microbial  
75 and wild populations.

76 Many other differences exist, such as variations in environmental constraints, and anthro-  
77 pogenic pressures, which affect wild population dynamics but are absent from laboratory con-  
78 ditions. Despite these differences, our review describes how microbiology findings apply to wild  
79 populations.

80 The evolution of wild populations involves multiple evolutionary and ecological processes  
81 that act simultaneously. Understanding the influence of each process independently is crucial  
82 for a better understanding of the overall effect of a bottleneck during demographic history on  
83 future response to selection to a new environment. Yet, empirical studies under laboratory  
84 conditions involving models other than micro-organisms have never examined the effects of  
85 bottlenecks in anything other than a holistic way [e.g., fish populations of *Heterandria formosa*  
86 in Klerks et al. (2019), insect populations of *Tribolium castaneum* in Olazcuaga et al. (2023),  
87 *Drosophila melanogaster* in Ørsted et al. (2019)]. Therefore, it is impossible to quantify the  
88 contribution of each process. Conversely, theory and microbiology have studied each ecological  
89 and evolutionary process independently [e.g., the fraction of beneficial mutations lost due to  
90 bottlenecks in Wahl et al. (2002)]. Therefore, we discuss in this review the impact on adap-  
91 tive potential of each of the following evolutionary processes occurring during a bottleneck:  
92 mutation, genetic drift, natural selection, and gene flow.

93 This review aims to enhance our comprehension of how bottlenecks impact adaptive poten-  
94 tial to a new environment of a population. This aim is achieved by synthesizing theoretical  
95 and microbiological knowledge and applying it to wild populations. For each of the evolution-  
96 ary processes of interest, we inquire: (i) What do theory and microbiology experiments tell us  
97 about the impact of bottlenecks during demographic history on the ability of populations to  
98 adapt to a future environmental change? (ii) Do these theoretical predictions apply to wild  
99 populations? and (iii) What is needed to better predict the wild population evolution after a  
100 bottleneck? Our review aims to increase the effectiveness of conservation efforts by anticipating  
101 the evolutionary consequences of demographic changes in wild ecosystems.

## 2 Disentangling the influence of each evolutionary process during a bottleneck on the future adaptive potential of populations

### 2.1 Mutation

Population bottlenecks can impact mutations' appearance, fixation, and frequency, potentially disturbing future adaptation. Indeed, mutations introduce genetic variation on which selection can act, allowing populations to adapt to their environment. A population will adapt to future environmental changes by increasing the frequency of mutations that are beneficial in the new environment. The population can adapt via (i) the appearance of new beneficial mutations, known as an adaptation from *de novo* mutations, and (ii) the pre-existence of beneficial mutations in the population, known as an adaptation from standing genetic variation. The relative importance of these two mechanisms varies depending on the population properties, such as the population size, and some timescales, such as the number of generations between the bottleneck and the environmental change. In the following section, we discuss how these two mechanisms, i.e., the appearance of *de novo* mutations and mutations from standing genetic variation (or pre-existing mutations), can be impacted during a bottleneck.

#### 2.1.1 Impact of bottleneck on *de novo* mutations

Theoretical work has shown that bottlenecks can reduce the mutation supply and the fixation probability of beneficial mutations in populations experiencing them (Wahl et al., 2002). More precisely, populations adapting mainly from *de novo* mutations have an adaptation rate limited by the mutation supply, which depends on population size and the appearance rate of adaptive beneficial mutations. These theoretical predictions were experimentally confirmed with asexual populations where adaptation depends mainly on *de novo* mutations [see, e.g., de Visser and Rozen (2005)]. Campos and Wahl (2010) even derived a more complex expression of the adaptation rate of asexuals, taking into account clonal interference. Mechanically, a bottleneck reduces the population size and, thus, limits adaptation.

Additionally, the adaptation rate of populations adapting through *de novo* mutations also depends on the distribution of fitness effects of beneficial mutations, which may also be affected by bottlenecks. Indeed, the fixation probability of all beneficial mutations is predicted to be reduced in a population undergoing bottlenecks (Wahl et al., 2002). However, mutations are affected differently depending on their rate and the fitness benefit they confer, which are usually negatively correlated. Gamblin et al. (2023) used a stochastic model to show that severe bottlenecks following a long growth phase favor rare beneficial mutations [as shown by Wahl et al. (2002)]. In contrast, relaxed bottlenecks following short growth phases favor frequent weakly beneficial ones. A similar effect has been observed in microbial experiments studying antimicrobial resistance. Garoff et al. (2020) and Schenk et al. (2022) found that antimicrobial resistance evolved through weakly beneficial mutations with large mutational targets when

139 the population size prior to the bottleneck was small. When the population size prior to  
140 the bottleneck was larger, antimicrobial resistance evolved through rarer and more beneficial  
141 mutations.

142 The theoretical and experimental studies mentioned above mostly deal with asexual popu-  
143 lations, which rely more on *de novo* mutations to adapt to environmental changes than sexual  
144 populations, the latter being more frequently genetically diverse. Even if most endangered  
145 species, which are the focus of conservation efforts, are sexual, these predictions could apply to  
146 them. Indeed, recent evidence suggests that some animal species are also limited by mutation  
147 supply on recent evolutionary scales (Rousselle et al., 2020).

### 148 **2.1.2 Impact of bottleneck on pre-existing genetic variation**

149 Populations experiencing a bottleneck are theoretically expected to have reduced genetic diver-  
150 sity (Chakraborty and Nei, 1982; Lynch and Hill, 1986; Nei et al., 1975; Tajima, 1989, 1996).  
151 This reduction can limit their ability to adapt to future changing environments (Frankham  
152 et al., 2002; Willi et al., 2006). This effect is especially important for sexual populations, as  
153 their adaptation in a short timescale is mostly driven by standing genetic variation. Indeed,  
154 sexual populations have constrained access to beneficial mutations due to Haldane’s sieve, which  
155 results from selection mainly acting on heterozygotes, thus decreasing the fixation probability  
156 of beneficial recessive mutations compared to asexual populations [see Marad et al. (2018) for  
157 a comparison in yeast]. Also, sexual reproduction allows selection to act on individual loci  
158 rather than haplotypes, thus making it possible to exploit the standing diversity [see Burke  
159 et al. (2014) for this observation in yeast]. Finally, higher organisms typically have smaller  
160 population sizes and lower mutation rates than micro-organisms, resulting in a limited supply  
161 of new mutations to rely on for adaptation.

162 Studies on asexual yeasts have shown that standing genetic variation drives adaptation along  
163 with *de novo* mutations (Vázquez-García et al., 2017; Ament-Velásquez et al., 2022). However,  
164 this aspect of microbial adaptation has not been extensively studied because experiments in-  
165 volving asexual individuals often start with a clonal population. Thus, there are no results  
166 yet from microbial experiments based on standing genetic variation that could apply to wild  
167 endangered populations.

168 Nonetheless, some observations from *Drosophila* experiments show how bottlenecks impact  
169 adaptation from standing genetic variation by disrupting allele frequencies. Rare alleles are  
170 likely to be lost during the bottleneck, resulting in a reduced allelic diversity (Allendorf, 1986;  
171 Fuerst and Maruyama, 1986). Swindell and Bouzat (2005) performed an empirical test of the  
172 drift-mutation model using *Drosophila*. This drift-mutation model aimed to predict the adap-  
173 tive potential of a population through genetic variation, which is modeled as an equilibrium  
174 between mutations and fixation due to inbreeding (Lynch and Hill, 1986; Clayton and Robert-  
175 son, 1955). In particular, this model assumes that the adaptive potential only depends on  
176 heterozygosity and not on allelic diversity (Falconer, 1960), which are two different aspects of  
177 the genetic diversity of a population. During their experiment, Swindell and Bouzat (2005)

178 observed a good agreement between model predictions and empirical observations, except after  
179 a severe bottleneck. The authors hypothesized that not considering the loss of allelic diversity  
180 during the bottleneck leads to overestimating the adaptive capacity following this event. This  
181 result suggests that the loss of heterozygosity and rare alleles are to be accounted for when  
182 predicting the effect of a bottleneck on a wild endangered population.

### 183 **2.1.3 Additive genetic variance from a quantitative genetics perspective**

184 For quantitative traits, the additive component of the genetic variance, which is denoted  $V_A$ ,  
185 is often taken as a proxy for the adaptive potential. The effect of a bottleneck on quantitative  
186 genetic variation is more complex to predict (Hoffmann et al., 2017). In theory, the genetic  
187 variance should decrease after a bottleneck as it is proportional to the effective population size  
188 (Lynch and Hill, 1986). This effect is usually observed in morphological traits (Willi et al.,  
189 2007), whereas fitness-associated life-history traits often show an increased genetic variance  
190 following a bottleneck (van Heerwaarden et al., 2008). A possible explanation is that, after a  
191 bottleneck, the disruption of allele frequencies could result in a transfer of epistatic and domi-  
192 nance variance to additive variance, especially for life-history traits, which are more influenced  
193 by these non-additive effects (Crnokrak and Roff, 1995; Roff and Emerson, 2006). However,  
194 these life-history traits also typically experience high inbreeding depression (DeRose and Roff,  
195 1999). As a result, an increase in additive variance for the genes associated with these traits  
196 may mitigate the fitness decrease. Still, it will not allow an increase in fitness compared to the  
197 pre-bottleneck level.

198 Overall, Willi et al. (2006) and Lopez-Fanjul and Villaverde (1989) concluded that genetic  
199 variance, and thus the future adaptive response, may increase in a population facing a bottleneck  
200 ("bottlenecked population" hereafter). However, this phenomenon is unlikely to result in a full  
201 fitness recovery, let alone a fitness increase.

202 The implication for endangered species management is that computing the additive genetic  
203 variance just after a bottleneck event may not reflect long-term adaptive potential but merely  
204 short-term adaptation in reaction to this event.

### 205 **2.1.4 Prospects for filling knowledge gaps**

206 Understanding the effect of bottlenecks simultaneously on pre-existing mutations and *de novo*  
207 mutations appearing during a bottleneck is important to predict the impact of bottlenecks on  
208 adaptability better. Performing microbial experiments that include initial genetic variation, al-  
209 lowing for better differentiating the effect of *de novo* mutations from standing genetic variation  
210 after a bottleneck, could improve this understanding. A recent review (Burke, 2023) suggested  
211 using yeast evolution experiments with standing genetic variation to study eukaryote adapta-  
212 tion. Indeed, yeast can combine short generation time and easy handling in the lab with sexual  
213 reproduction. Performing such experiments, particularly with small populations undergoing  
214 different types of bottlenecks, would help quantify the importance of standing genetic variation  
215 versus *de novo* mutations for eukaryote adaptation.

216 To improve our knowledge of which quantitative traits have their variance decreased after a  
217 bottleneck, we would need experiments to estimate the genetic variance in a wide range of traits  
218 (Willi et al., 2006). Estimating the genetic variance seems essential to predict the overall effect  
219 of a bottleneck on the population’s adaptive potential. Moreover, in natural populations, the  
220 link between genetic diversity and response to selection is not always clear (Pujol et al., 2018).  
221 For example, Albatross persists despite losing genetic diversity (Milot et al., 2007). Clarifying  
222 the link between genetic variation and response to selection would help understand if genetic  
223 diversity can be used to accurately predict the natural populations’ potential to adapt after  
224 one or more bottlenecks.

## 225 **2.2 Genetic drift**

226 Populations suffering from bottlenecks are particularly affected by genetic drift, which is the  
227 change in allele frequencies caused by population size fluctuations rather than by selection,  
228 mutation, or migration. Indeed, the strength of this process is inversely proportional to the  
229 effective size of the population (Kimura, 1955). These population size fluctuations caused by  
230 chance likely lead to negative impacts on a population, such as (i) the fixation of deleterious  
231 mutations, which decreases the population’s fitness; (ii) the reduction in the fixation probability  
232 of beneficial mutations, which limits adaptation, and (iii) the increase of alleles at extreme  
233 frequencies (i.e., 0 or 1), which reduces genetic variation (Falconer, 1960). The latter point has  
234 already been covered in section 2.1 about mutation. Taken together, these effects may limit  
235 the adaptation of bottlenecked populations to future environmental changes.

### 236 **2.2.1 Fixation of deleterious mutations**

237 The accumulation of deleterious mutations caused by genetic drift in a population undergoing  
238 repeated bottlenecks predicted by theoretical work was highlighted numerous times in microbial  
239 experiments (Muller, 1964). In particular, many experiments used clone-to-clone transfers to  
240 maximize the rate and the speed of accumulation (Clarke et al., 1993) [see the review Elena  
241 and Lenski (2003) for references on viruses, bacteria, and yeast]. For instance, a linear decay  
242 of the average fitness of a hypermutator *Escherichia coli* strain subject to repeated single-cell  
243 bottlenecks was observed in (Heilbron et al., 2014).

244 Sexual populations are also theoretically expected to suffer from deleterious mutation ac-  
245 cumulation (Lynch et al., 1995). This accumulation was observed in domesticated species due  
246 to bottlenecks and selective sweeps (Marsden et al., 2016; Xie et al., 2018), but also in some  
247 wild bottlenecked populations such as the Florida panther (Roelke et al., 1993). However, the  
248 deleterious mutation is potentially less common than in asexual microorganisms, where, due to  
249 the absence of recombination, the offsprings are expected to bear at least as much mutational  
250 load as their ancestors, a process called Muller’s ratchet (Muller, 1964, 1932). In wild popula-  
251 tions that reproduce sexually, recombination can break this process (McDonald et al., 2016).  
252 Therefore, the populations being the focus of conservation efforts are probably less affected by  
253 this particular bottleneck effect.

## 254 **2.2.2 Reduced fixation probability of beneficial mutations**

255 Some theoretical studies reproducing microbiology experiments showed that bottlenecks can  
256 reduce the fixation probability of a beneficial mutation. For example, Wahl et al. (2002) found  
257 that the fixation probability of a beneficial mutation in a periodically bottlenecked popula-  
258 tion was reduced by a factor accounting for the probability of losing the mutation during the  
259 dilution. As a reminder, the fixation probability of a beneficial mutation in a fixed-size pop-  
260 ulation is approximately twice the selective advantage (Haldane, 1927). Heffernan and Wahl  
261 (2002) also considered that genetic drift is increased in bottlenecked populations due to a lower  
262 size resulting from the bottleneck. This effect reduces the fixation probability by about 25%  
263 compared to previous estimates. These theoretical predictions were confirmed by experiments  
264 using microorganisms where bottlenecks and genetic drift hindered adaptation. In the case  
265 of experimental evolution of antibiotic resistance, Huseby et al. (2017) found a positive cor-  
266 relation between bottleneck size and ciprofloxacin tolerance. In addition, Garoff et al. (2020)  
267 highlighted that a low-intensity bottleneck (i.e., a small reduction in population size) leads to  
268 higher fluoroquinolone tolerance than a high-intensity bottleneck. This effect is also likely to  
269 impact wild populations of endangered species, though the extent of this impact is not clear as  
270 their adaptation relies mostly on standing genetic variation.

## 271 **2.2.3 Prospects for filling knowledge gaps**

272 As bottleneck-induced drift affects both new and existing mutations, estimating its impact on  
273 the adaptive potential of bottlenecked populations would require quantifying the respective  
274 importance of *de novo* mutations versus standing genetic variation for the adaptation of a  
275 given population on a given timescale. This problem was previously mentioned in section 2.1  
276 about mutation. The review (Barrett and Schluter, 2008) presents the relative contribution of  
277 these two sources of genetic variation in wild populations. This review suggested ways to detect  
278 molecular signatures of adaptation from standing genetic variation (Barrett and Schluter, 2008;  
279 Przeworski et al., 2005).

280 Another open question deals with the potential beneficial effect of genetic drift on the  
281 adaptive potential of populations. In a modeling study, Handel and Rozen (2009) found that  
282 small asexual populations could reach higher fitness peaks than large ones on rugged landscapes  
283 because drift prevents them from being stuck on a local maximum. The authors concluded that  
284 there is an optimal population size to maximize adaptation, depending on the fitness landscape's  
285 characteristics and the relative importance of adapting rapidly versus reaching high fitness peaks  
286 (Handel and Rozen, 2009). Assessing whether these effects are also observed in wild populations  
287 would be interesting.

## 288 **2.3 Natural selection**

289 Natural selection will act more or less effectively on the bottlenecked population depending  
290 on several factors, such as the severity of the bottleneck, its duration, and the population's



291 genetic diversity before and after the bottleneck. Some general predictions can be drawn from  
292 microbiology about the impact of natural selection after a bottleneck, regardless of the charac-  
293 teristics of the bottleneck: (i) Several mechanisms tend to reduce the population fitness, such  
294 as genetic load and inbreeding depression, but (ii) natural selection can also purge deleterious  
295 alleles in sexual populations. These mechanisms will modify the population's fitness, impacting  
296 its ability to adapt to future environmental changes.

### 297 **2.3.1 Fitness decrease due to bottlenecks**

298 Several factors may explain why a bottlenecked population experiences a fitness decline, even  
299 without any environmental change.

300 As detailed in section 2.2 about genetic drift, bottlenecks are expected to cause an accumu-  
301 lation of deleterious mutations in the population due to genetic drift, leading to a decrease in  
302 fitness if the bottlenecks are severe.

303 It is important to note a major difference between microbial populations and the endangered  
304 wild populations of diploid eukaryotes that we are considering in this review: the latter can  
305 suffer from inbreeding depression because bottlenecks reduce population sizes (Charlesworth  
306 and Charlesworth, 1987; Keller, 2002), leading to more reproductive events between related  
307 individuals. This inbreeding depression results in a loss of heterozygosity that can unmask re-  
308 cessive deleterious alleles, ultimately decreasing this population's fitness and adaptive potential  
309 (Barrett and Kohn, 1991; Ellstrand and Elam, 2003). More generally, genetic load (i.e., the  
310 actual or potential reduction in population mean fitness due to drift load, inbreeding load, and  
311 mutation load) is responsible for a direct decline in population fitness following a bottleneck  
312 (Hedrick and Garcia-Dorado, 2016; Kirkpatrick and Jarne, 2000).

313 In less well-understood ways, bottlenecks can affect other characteristics of the populations  
314 than genetics but still influence their fitness and future ability to adapt. Specifically, a bot-  
315 tleneck can impact the balanced relationship between host and microbiome in eukaryotes. For  
316 instance, Ørsted et al. (2022) showed that *Drosophila* populations that had undergone bottle-  
317 neck treatment also lost the diversity and richness of their microbiome. The direct consequence  
318 of this loss is a reduction in the fitness of individuals belonging to bottlenecked populations  
319 (Ørsted et al., 2022).

### 320 **2.3.2 Purge of deleterious alleles by natural selection increases population fitness**

321 Whereas a bottleneck can increase the frequency of deleterious alleles (see section 2.2 about  
322 genetic drift), natural selection can purge these deleterious alleles (Kirkpatrick and Jarne, 2000;  
323 Hedrick and Garcia-Dorado, 2016). If a purging process is more efficient during a bottleneck,  
324 then going through a bottleneck could be beneficial for the adaptive potential of the population  
325 (Bouzat, 2010; Bertorelle et al., 2022; Dussex et al., 2023).

326 Purifying selection can play out in microorganism experiments and yet population evolution,  
327 but inbreeding facilitates the purge in diploid eukaryotes (Hedrick and Garcia-Dorado, 2016).  
328 As mentioned above, inbreeding increases homozygosity and, thus, the unmasking of recessive

329 deleterious alleles. Whereas inbreeding depression decreases population fitness, it is also an  
330 opportunity for selection to act on these deleterious alleles and purge them.

331 The empirical evidence for a purge of deleterious alleles following a bottleneck appears to  
332 be mixed (Bouzat, 2010). There is some evidence that purge can strongly affect experimental  
333 populations (Crnokrak and Barrett, 2002) and captive populations (López-Cortegano et al.,  
334 2021; Boakes et al., 2006). In experimental yeast populations, Agrawal and Whitlock (2011)  
335 used data from the *Saccharomyces* Genome Deletion Project to estimate fitness and dominance  
336 coefficient at about 1000 loci. From this, the authors estimated that the effect of one gener-  
337 ation of purging (i.e., deliberate inbreeding) in an already partially inbred population would  
338 substantially decrease inbreeding depression.

339 Evidence of a purge process in wild endangered populations is mostly indirect (Bouzat,  
340 2010). However, some direct evidence exists. For example, the deleterious load was significantly  
341 lower for the endangered species Iberian lynx (*Lynx pardinus*) than for the widespread Eurasian  
342 lynx (*Lynx lynx*) due to a genetic purging process (Kleinman-Ruiz et al., 2022). Other examples  
343 of purging in natural environments after a bottleneck exist [e.g., the Alpine ibex Grossen et al.  
344 (2020)]. As discussed in (Bouzat, 2010), the role of purging during a bottleneck and the factors  
345 influencing its role in natural populations still need to be discovered.

### 346 **2.3.3 Prospects for filling knowledge gaps**

347 While allele purging appears to be a key process for understanding the adaptive potential of  
348 bottlenecked populations, empirical evidence is still mixed, proving that we do not yet fully  
349 understand this mechanism. Therefore, it would be useful to use diploid eukaryotic microor-  
350 ganisms to test the factors and conditions under which allele purging occurs.

351 In addition, a study suggested carefully handling the results of selection detection methods  
352 when working with bottlenecked populations (Leigh et al., 2021). Indeed, Leigh et al. (2021)  
353 found that, due to the high level of genetic drift, methods commonly used to detect selec-  
354 tion (e.g.,  $F_{ST}$  outlier scans and Genome-Environment Association analyses) presented high  
355 false positive rates when applied to bottlenecked Alpine Ibex populations. Detecting adapta-  
356 tion is essential for managing endangered populations, so testing other methods' false positive  
357 and negative rates and developing new methods to distinguish between drift and selection is  
358 necessary.

## 359 **2.4 Gene flow**

360 Human activity causes fragmentation of populations in the wild, leading to spatially structured  
361 populations divided into sub-populations (also called demes or islands) of reduced sizes between  
362 which individuals may migrate. Thus, population fragmentation results in a bottleneck that  
363 risks reducing genetic diversity within sub-populations (i.e., genetic depression), losing adap-  
364 tive potential, and accumulating deleterious mutations (Keyghobadi, 2007; Frankham et al.,  
365 2017). However, the migration of individuals between sub-populations can induce gene flow,  
366 which represents an opportunity to diversify the gene pool of the sub-populations despite the

367 fragmentation-induced bottleneck. Quantifying the genetic diversity of sub-populations is cru-  
368 cial to assessing the adaptive potential of fragmented populations, particularly in the case of  
369 changing environments threatening their persistence. This section reviews how and in what  
370 conditions gene flow can restore the adaptive potential of a bottlenecked population. Gene  
371 flow can (i) restore lost genetic variation, (ii) mitigate inbreeding depression, (iii) resulting in  
372 a decreased probability of extinction and restoration of adaptive potential, and (iv) amplify  
373 natural selection depending on the meta-population structure.

#### 374 **2.4.1 Restoration of lost genetic variation**

375 One of the main effects of gene flow in a bottleneck population is the restoration of lost genetic  
376 variation (Soulé, 1987; Franklin and Frankham, 1998). This theoretical expectation is observed  
377 experimentally (Swindell and Bouzat, 2006) and in natural populations (Jangjoo et al., 2016;  
378 Goodman et al., 2001; Chiuichi and Gibbs, 2010).

379 Habitat fragmentation can cause the extinction of bottlenecked populations. Gene flow be-  
380 tween sub-populations can mitigate the negative effects of bottlenecks by restoring lost genetic  
381 diversity (Ingvarsson, 2001). For example, Jangjoo et al. (2016) discovered that connectivity  
382 in a meta-population of the alpine butterfly *Parnassius smintheus* preserves genetic diversity  
383 before, during, and after a two-generation bottleneck. The Roseate Tern (*Sterna dougallii*  
384 *dougallii*) is an endangered Atlantic seabird population that provides another example of how  
385 connectivity and gene flow across populations can help retain genetic diversity despite a severe  
386 bottleneck (Dayton and Szczys, 2021). Seed dispersal with water facilitates gene flow between  
387 bottlenecked populations, mitigating the decrease in allelic diversity caused by a bottleneck  
388 (Yu et al., 2020). These examples are not isolated cases and are found in many endangered  
389 species. To further elaborate, gene flow between sub-populations undergoing a bottleneck can  
390 even erase the genetic variation effects of a bottleneck to the point where no negative genetic  
391 effects can be detected [e.g., *Actinidia chinensis* populations (Yu et al., 2020)].

#### 392 **2.4.2 Change in genetic load composition**

393 One less studied effect of gene flow on bottleneck populations, which could be predominant in  
394 the populations' fate, is its impact on genetic load. Gene flow is theoretically expected to reduce  
395 the deleterious effects of inbreeding in bottleneck populations. Gene flow in metapopulations  
396 can mitigate inbreeding load by preventing the fixation of deleterious alleles in bottleneck  
397 populations (Whitlock, 2003). However, gene flow can also reduce the effectiveness of the purge  
398 of these deleterious alleles by increasing heterozygosity. With individual-based simulations,  
399 a study found that an intermediate rate of gene flow can minimize the mutation load and  
400 prevent the extinction of local populations while still allowing some purging of deleterious  
401 alleles (Sachdeva et al., 2022).

### 402 2.4.3 Gene flow mitigate extinction risk

403 Bottlenecked populations are highly vulnerable to extinction via (i) demographic stochasticity  
404 (e.g., random births and deaths), demographic heterogeneity and sampling variation in sex  
405 ratios (Melbourne and Hastings, 2008), and (ii) environmental stochasticity [e.g., catastrophic  
406 events Lande (1988)]. Specifically, bottlenecked populations can fall into an "extinction vortex",  
407 often characterized by a complex interplay between genetic drift, demographic stochasticity,  
408 and environmental fluctuations (Soulé, 1986). A population bottleneck reduces fitness directly  
409 through increased genetic load and indirectly through erosion of genetic variation, leading to  
410 population decline, exacerbating the effects of genetic drift, demographic stochasticity, and  
411 environmental fluctuations until extinction (Nordstrom et al., 2023). Theoretical models have  
412 highlighted a critical level of gene flow that allows a metapopulation to survive over long  
413 timescales, even if it is often ultimately driven to extinction (Hanski and Ovaskainen, 2003;  
414 Gyllenberg and Hanski, 1992; Lande et al., 1998).

415 The process of restoring gene flow in these bottlenecked populations to alleviate genetic load,  
416 increase genetic variation, and increase persistence probability is termed genetic rescue (Bell  
417 et al., 2019; Tallmon et al., 2004; Whiteley et al., 2015). Much empirical evidence suggests that  
418 recently fragmented populations will likely receive a demographic benefit from gene flow, beyond  
419 the addition of immigrant individuals, through genetic rescue (Whiteley et al., 2015; Frankham,  
420 2015; Hufbauer et al., 2015; Fitzpatrick and Reid, 2019). A recent meta-analysis revealed the  
421 significant and consistent benefits of gene flow for the adaptive potential of endangered species  
422 experiencing a fragmentation-induced population bottleneck (Frankham, 2015).

423 When a population faces deteriorating environmental conditions and is doomed to extinc-  
424 tion, gene flow may allow its evolutionary rescue. For example, Bell and Gonzalez (2011) set up  
425 an experiment in which a yeast metapopulation was subjected to salt-induced environmental  
426 stress. This experiment showed that local yeast dispersal and gradual deterioration favored  
427 the evolutionary rescue of the metapopulation, which would otherwise die out due to envi-  
428 ronmental stress. This experimental result later led to the development of theoretical models  
429 investigating the probability of evolutionary rescue by including a hitherto overlooked ecolog-  
430 ical factor: population structure, which may result from fragmentation. Interestingly, these  
431 models showed that the probability of evolutionary rescue in an island model, in which demes  
432 deteriorate one by one, does not vary monotonically with gene flow rate (Uecker et al., 2014).  
433 In other words, there is a gene flow rate that optimizes the evolutionary rescue of a popula-  
434 tion. Gene flow allows genetic variation and the introduction of beneficial mutants necessary  
435 for adaptation. However, a too-strong gene flow risks preventing local beneficial mutants from  
436 becoming permanently established, hence the need for intermediate gene flow to optimize adap-  
437 tation (Tomasini and Peischl, 2022). Further studies showed that directed gene flow based on  
438 habitat choice could favor evolutionary rescue (Czuppon et al., 2021). Habitat choice occurs,  
439 for example, when individuals, whether mutant or wild-type, preferentially immigrate to demes  
440 whose environment has already changed. Other details of population fragmentation, such as  
441 between which sub-populations gene flow is allowed (e.g., island model, stepping-stone model)

442 or its asymmetry, impact the probability of evolutionary rescue (Tomasini and Peischl, 2020,  
443 2022). To our knowledge, the above-mentioned theoretical predictions have not been tested  
444 experimentally.

#### 445 **2.4.4 Meta-population structure can amplify or suppress natural selection**

446 The fragmentation of a population induces a bottleneck that divides the population into smaller  
447 sub-populations. This bottleneck accentuates genetic drift within sub-populations, but its effect  
448 on natural selection is more subtle. Its effect may depend on the meta-population structure  
449 resulting from fragmentation and the gene flow pattern.

450 Many scientific publications address whether population fragmentation and gene flow change  
451 the fixation probability of a mutation compared to a non-fragmented population of the same  
452 size. In (Pollak, 1966), the author focused on a population fragmented into a finite number of  
453 demes between which individuals can migrate and showed that symmetric migrations lead to  
454 the same fixation probability as in a non-subdivided population.

455 Whitlock (2003) and Whigham et al. (2008) challenged this independence of the fixation  
456 probability from the meta-population structure resulting from fragmentation. Further works  
457 showed that the meta-population structure resulting from fragmentation could either amplify  
458 or suppress natural selection (Lieberman et al., 2005; Houchmandzadeh and Vallade, 2011)  
459 (i.e., increase or decrease the efficacy of natural selection, respectively). Amplifying natural  
460 selection means reducing the fixation probability of deleterious mutations and increasing that  
461 of beneficial ones, whereas suppressing natural selection does the opposite. Importantly, the  
462 meta-population structure alone is insufficient to assess the impact of a fragmentation-induced  
463 bottleneck on natural selection (i.e., amplifier, suppressor, or without effect) as the gene flow  
464 pattern needs to be taken into account (Marrec et al., 2021). Many theoretical studies assessing  
465 the impact of fragmentation on evolutionary dynamics focused on the fixation probability, but  
466 other important quantities are impacted, such as the adaptation rates (Hindersin and Traulsen,  
467 2014). An experiment in which ciprofloxacin-resistant mutants were tracked in a *Pseudomonas*  
468 *aeruginosa* meta-population showed that for low migration rates, natural selection is ampli-  
469 fied in a star topology compared to a well-mixed population (Chakraborty et al., 2023), thus  
470 confirming a theoretical prediction made by (Marrec et al., 2021).

#### 471 **2.4.5 Prospects for filling knowledge gaps**

472 Human activity fragments populations into several sub-populations (also called demes or is-  
473 lands), which can become isolated if gene flow between them is limited. As biodiversity declines,  
474 it is crucial to understand the impact of fragmentation and gene flow on the evolutionary dy-  
475 namics of bottlenecked populations and, in particular, their adaptive potential. In this review,  
476 we have shown that there are many theoretical studies investigating this impact. However, the-  
477 oretical predictions are rarely directly tested or mostly with microbiology experiments whose  
478 design does not allow comparison with mathematical models. A stronger collaboration be-  
479 tween theory and experiment [e.g., Marrec et al. (2021) combined with Chakraborty et al.

480 (2023)] would lead to a better understanding of fragmentation and gene flow on the evolu-  
 481 tionary dynamics of meta-populations. Also, more experiments with diploid organisms would  
 482 enable better comparison with wild populations [e.g., Bakker et al. (2010)].

### 483 3 Relative importance of these processes

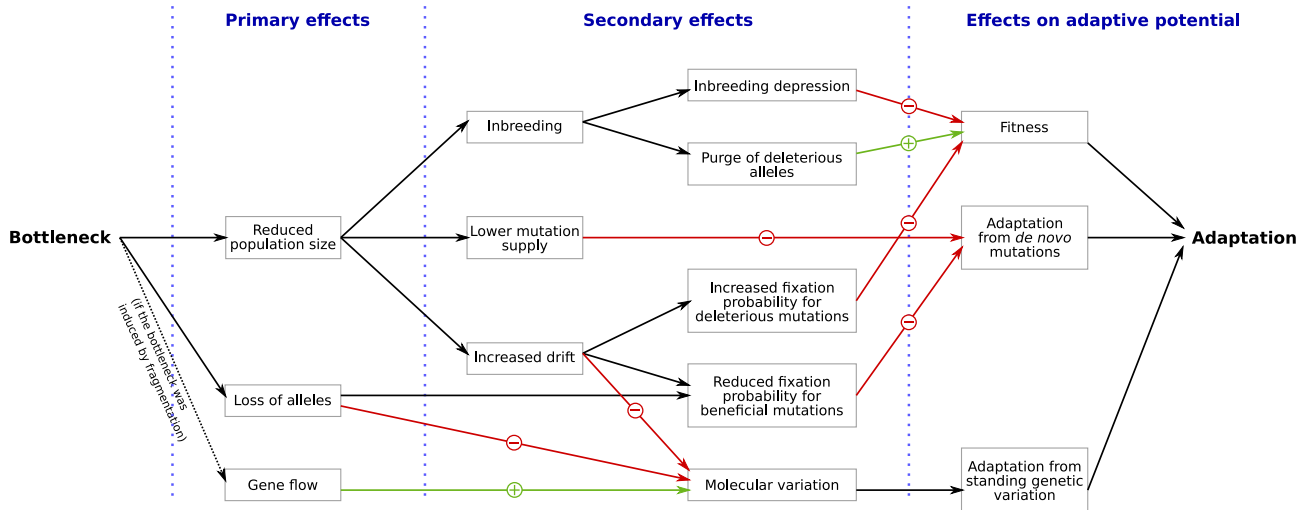


Figure 1: **Potential effects of a bottleneck on adaptive potential.** Summary of the main effects of bottlenecks on the adaptive potential of a population, as described in this review. The existence and relative importance of these different effects vary across populations, bottleneck characteristics, and environments. Black arrows represent a causal effect, green arrows represent a positive effect, and red arrows represent a negative effect.

#### 484 3.1 Summary of the previous parts

485 We have reviewed in the previous parts how the different evolutionary processes can be disrupted  
 486 during a bottleneck and how these processes shape the adaptive potential of bottlenecked  
 487 populations (see Figure 1).

488 As expected, most of these mechanisms are predicted to decrease the adaptive potential  
 489 following a bottleneck. Bottlenecks limit adaptation from *de novo* mutations by reducing  
 490 the mutation supply and the fixation probability of beneficial mutations. More importantly  
 491 for sexual populations, they also limit adaptation from standing genetic variation. Indeed,  
 492 molecular variation is decreased due to smaller population sizes, increased drift, and the loss  
 493 of rare alleles during bottlenecks. In addition, the fitness of a bottlenecked population may  
 494 decrease due to the accumulation of deleterious mutations and genetic load in general, which  
 495 for sexual diploids includes inbreeding depression. A population with lower fitness will struggle  
 496 to survive future environmental changes that may increase its probability of extinction.

497 Conversely, only two mechanisms can mitigate the negative impacts of bottlenecks. When  
 498 the population is part of a meta-population, gene flow can restore some of the lost genetic  
 499 variation by introducing new variation. In addition, in the case of sexual diploid populations,

500 inbreeding could, under some conditions, facilitate the purge of deleterious alleles and, thus,  
501 reduce the masked genetic load.

502 These findings show that knowledge transfer from microbial to endangered wild populations  
503 is possible. A collaboration between microbiology and conservation biology would be fruitful  
504 if microbial experiments were adapted to include more characteristics of these endangered  
505 populations. For example, one could use (facultative) sexual micro-organisms, such as yeast,  
506 or include standing genetic variation in evolution experiments.

## 507 **3.2 Relative importance of each of these processes**

508 Several evolutionary processes come into play when trying to predict the impact of a bottleneck.  
509 As some of these processes have opposite effects, a major concern is to estimate the relative  
510 importance of these processes to predict the fate of a population. Even when two processes  
511 negatively affect the adaptive potential of populations, it may be useful to know which one  
512 is predominant to determine the conditions threatening the persistence of wild populations  
513 precisely. Determining these conditions would help identify the key priorities in population  
514 management. In the following of this review, we discuss the relative contribution of the evolu-  
515 tionary processes seen above.

### 516 **3.2.1 Selection vs. genetic drift**

517 One of the major concerns during a bottleneck is the increase of genetic drift. The positive or  
518 negative aspect of genetic drift depends on whether the alleles are deleterious or beneficial.

519 In section 2.2 about genetic drift, we have seen that a bottleneck reduces the fixation  
520 probability of beneficial alleles and increases the chance that they are lost compared to a fixed-  
521 size population.

522 The impact of a bottleneck on deleterious alleles is more complex, as inbreeding can facilitate  
523 their purge by natural selection. As previously said, little is known about the conditions required  
524 for selection to overcome drift. Even if these conditions were known, we would still have to  
525 choose if the management priority is to purge deleterious alleles, which requires inbreeding, or  
526 to restore genetic diversity, which requires outcrossing. Conservation biology often deals with  
527 small endangered wild populations that have already experienced severe bottlenecks. For such  
528 populations, the loss of genetic diversity may be a major concern, and the impact of the purge is  
529 minor, which has been confirmed in wild populations (Bouzat, 2010) [but see van Heerwaarden  
530 et al. (2008)]. For example, wild populations of elephants in South Africa underwent a severe  
531 bottleneck due to widespread hunting. Microsatellite comparisons of current wild populations  
532 with museum specimens of this elephant population before the bottleneck confirmed the loss  
533 of genetic diversity (Whitehouse and Harley, 2001). In such wild populations, the response to  
534 selection can be expected to be less effective, as the probability of having beneficial mutations  
535 is low (Frankham, 2009). A reduced response to selection after a bottleneck has already been  
536 highlighted in experimental fish populations. For example, populations of *Heterandria formosa*  
537 having undergone a bottleneck during their demographic history showed a 50% slower response

538 to selection for heat tolerance than populations having not undergone a bottleneck (Klerks  
539 et al., 2019). An experiment showed that housefly populations that faced a short bottleneck  
540 followed by a period of expansion had better fitness and lower genetic load than populations  
541 kept at a constant size with a similar expected inbreeding score (Reed and Bryant, 2001). Reed  
542 and Bryant (2001) concluded that, when managing endangered wild populations, the priority  
543 is to act on the cause of decline to promote rapid expansion and avoid inbreeding.

544 For adaptive polymorphisms such as the Major Histocompatibility Complex (MHC), the  
545 predominance of selection over drift probably depends on the duration of the bottleneck. The  
546 MHC is a set of polymorphic genes essential to the adaptive immune system of vertebrates  
547 and can be particularly affected by bottlenecks. The potential loss of diversity at this locus  
548 is of great concern as it is associated with increased disease susceptibility (Sommer, 2005). In  
549 the meta-analysis (Sutton et al., 2011), the adaptive polymorphism of the MHC was shown  
550 to be significantly reduced after a bottleneck, and even more so than neutral polymorphisms  
551 (by 15%). One possible explanation found by Sutton et al. (2011) is that negative frequency-  
552 dependent selection is an important force shaping pre-bottleneck Major Histocompatibility  
553 Complex diversity, resulting in a high frequency of very rare alleles. As these rare alleles are  
554 more at risk of being lost during a bottleneck, this would explain the greater reduction in Major  
555 Histocompatibility Complex diversity. The authors concluded that diversifying selection cannot  
556 counter genetic drift in recently bottlenecked populations. However, this conclusion can be  
557 mitigated as the authors did not find a significant effect on Major Histocompatibility Complex  
558 diversity for short-scale bottlenecks. For instance, in a water vole population undergoing a 4-  
559 month bottleneck, the Major Histocompatibility Complex diversity was greatly reduced during  
560 this period but recovered in a few generations to reach the pre-bottleneck level (Oliver and  
561 Piertney, 2012).

### 562 **3.2.2 Loss of heterozygosity vs. loss of genetic variation**

563 Another open question is to identify the mechanism causing the more significant reduction in  
564 the adaptive potential of bottlenecked populations between the loss of heterozygosity and the  
565 loss of genetic variation. In population genetics theory, heterozygosity determines the evolution-  
566 ary potential, particularly the short-term response to selection (Falconer, 1960). Accordingly,  
567 *Drosophila* populations that faced intense or diffuse bottlenecks leading to the same level of het-  
568 erozygosity showed no difference in their response to selection (England et al., 2003). Whereas  
569 both bottleneck regimes were expected to yield different allelic diversities, the measured allelic  
570 diversities were not significantly different (England et al., 2003).

571 On the other hand, Ørsted et al. (2019), focusing on *Drosophila* populations having ex-  
572 perienceed different levels of inbreeding, showed that molecular diversity was more strongly  
573 correlated to adaptation than was the expected inbreeding coefficient. This result highlights  
574 the importance of molecular diversity for adaptation. It provides a way to summarize the  
575 history of a population, which seems more relevant than keeping track of population sizes.

576 However, to our knowledge, the methods used to restore heterozygosity are the same as



577 those used to restore diversity and consist of outcrossing (i.e., crossing the population with  
578 individuals from other populations and/or expanding the population size).

## 579 **4 Thoughts for future research directions**

580 In the future, conservation biology could benefit even more from microbiology by maintaining  
581 a close link between the two fields. With a reverse approach, evolution experiments using  
582 microorganisms could directly address the conservation biology needs. We make the following  
583 suggestions:

- 584 1. Include ecological factors within experimental evolution studies.
- 585 2. Include selective history when considering the demographic history of populations.
- 586 3. Include bottleneck characteristics and demographic history.

### 587 **4.1 Testing the influence of ecological factors on population re-** 588 **sponse**

589 As discussed in this review, most microbiology studies investigating the adaptive potential of  
590 bottlenecked populations have taken an evolutionary perspective without considering ecological  
591 factors. However, natural populations evolve in interaction with their biotic and abiotic envi-  
592 ronment. Theoretical predictions could be biased without considering these ecological factors.  
593 For instance, Nordstrom et al. (2023) showed through stochastic individual-based simulations  
594 that considering population growth with negative density dependence (i.e., intraspecific com-  
595 petition) or density independence leads to different outcomes of evolutionary rescue. This  
596 prediction regarding the impact of density dependence vs. independence was empirically tested  
597 and confirmed with flour beetles (Olazcuaga et al. in prep.). More precisely, Olazcuaga et al.  
598 showed that the effect of negative density dependence varies depending on whether the popula-  
599 tions have experienced a bottleneck in their demographic history. Osmond and de Mazancourt  
600 (2013) proved with an adaptive dynamic model that interspecific competition can favor evolu-  
601 tionary rescue by increasing the strength of selection and speeding up adaptation. Following  
602 Olazcuaga et al.'s example, examining how interspecific competition affects the probability of  
603 rescue in bottlenecked populations would be valuable.

### 604 **4.2 Testing the influence of selective and demographic history on** 605 **population response**

606 In this review, we focused on the effects of bottlenecks, which entail random reductions in  
607 population size ("random bottleneck" hereafter), rather than selective bottlenecks, which in-  
608 volve non-random decreases in population size. Wild populations can experience both random

609 and selective bottlenecks. Random bottlenecks can occur due to fragmentation, whereas se-  
610 lective bottlenecks are more likely to occur when adapting to a drastic environmental change,  
611 such as pollutants or antibiotic resistance. These selective bottlenecks can result in U-shaped  
612 population size curves during evolutionary rescue processes (Gomulkiewicz and Holt, 1995).  
613 Selective bottlenecks, as random bottlenecks, can impact how populations respond to future  
614 stress. A few studies tested how adaptation to a first environmental change, which was associ-  
615 ated with a decrease in population size, impacted the response to future adaptation to a new  
616 environment (Lachapelle et al., 2017; O'Connor et al., 2020; Samani and Bell, 2016; Gonzalez  
617 and Bell, 2013) using microorganisms: *Chlamydomonas reinhardtii*, *Pseudomonas fluorescens*,  
618 *Saccharomyces paradoxus*, and *Saccharomyces spp*, respectively). Adaptation to a new envi-  
619 ronmental change would be favored for populations that have already undergone similar stress  
620 in their demographic history (Lachapelle et al., 2017) [see O'Connor et al. (2020) for a change  
621 in the speed of future adaptation]. Conversely, if the stress is different, adaptation would be  
622 less likely (Lachapelle et al., 2017). This impact of the first dissimilar stress makes sense since  
623 the response to the selection of the first stress would reduce genetic variability (Carlson et al.,  
624 2014). Additionally, populations that have evolved under first stress during their demographic  
625 history seem to have a higher probability of extinction when they experience new and different  
626 stress (Lachapelle et al., 2017; Samani and Bell, 2016; Gonzalez and Bell, 2013). An increase in  
627 genetic load is expected during selective bottleneck (Stewart et al., 2017), which could explain  
628 this result. However, whether these deleterious mutations can be purged as efficiently as in  
629 a random bottleneck is unclear. Furthermore, the mean frequency of mutations and the ge-  
630 netic load can change in a complex way during a selective bottleneck, in contrast to a random  
631 bottleneck (Dussex et al., 2023). Overall, the evidence that adaptive bottlenecks increase the  
632 probability of extinction and impact the probability of adaptation suggests that the processes  
633 involved differ from those occurring in a random bottleneck or are not as straightforward as  
634 expected. The impacts of random bottlenecks *versus* selective bottlenecks have been studied  
635 theoretically in infection models and host-pathogen infection processes [as reviewed in Abel  
636 et al. (2015), e.g., Moxon and Kussell (2017) and De Ste Croix et al. (2020)]. However, how a  
637 selective bottleneck, compared to a random bottleneck, impacts the response to future stress  
638 has never been empirically studied. Integrating demographic and selective history can improve  
639 predictions of population response to different stresses.

640 Finally, it is essential to note that bottlenecks exist on a gradient and cannot be categorized  
641 into two binary categories. Selective and random bottlenecks represent the extreme points of  
642 this gradient, where the relative contribution of drift and selection varies. Moreover, we have  
643 discussed that natural selection can play a role in a random bottleneck process, challenging the  
644 assumption that a random bottleneck is entirely random. As a first step, it would be useful  
645 to compare the effects of selective and random bottlenecks on the probability of adaptation to  
646 future environmental conditions. Then, it would be important to study how the contribution of  
647 genetic drift and selection during demographic history influences the response of bottlenecked  
648 populations.

### 4.3 Testing the influence of bottleneck characteristics on future population response

In this review, we have focused on the impact of a single bottleneck on population response, except when considering microbial experiments that usually involve multiple bottlenecks. However, the demographic history of natural populations is never restricted to a single bottleneck (Hohenlohe et al., 2020; Gladstone et al., 2022). Therefore, it is essential to consider the entire demographic history of natural populations, including the intensity and frequency of these bottlenecks.

Microbiology informs us about the impact of the intensity and frequency of bottlenecks, which can be useful to conservation biology. Microbiologists can control the characteristics of the bottleneck, such as its frequency, intensity, and duration (LeClair and Wahl, 2017). The impact of bottleneck intensity has been tested in microbial experimental evolution. The adaptive pathways differ depending on whether the bottleneck is weak or intense (Garoff et al., 2020; Vogwill et al., 2016; Mahrt et al., 2021). Overall, empirical studies on the evolution of antibiotic resistance have observed a negative correlation between bottleneck severity and adaptive response (Garoff et al., 2020; Huseby et al., 2017; Mahrt et al., 2021). In addition, Mahrt et al. (2021) showed a decrease in parallel evolution with increasing bottleneck intensity. This result suggests that after experiencing strong bottlenecks, resistance evolves through a wider range of genetic mechanisms, likely due to increased genetic drift. Theoretical studies of bottleneck characteristics suggest that smaller population sizes before or after the bottleneck constrain evolutionary paths, thus limiting the supply of beneficial mutations and adaptation (Gamblin et al., 2023). In addition, Wein and Dagan (2019) pointed out that while bottleneck intensity is a factor in determining population evolvability, selective conditions during evolution can play a more significant role. Mahrt et al. (2021) notably examined the interaction between bottleneck intensity and intensity of selective pressure. The application of the effect of bottleneck intensity to natural populations remains unclear. Olazcuaga et al. (2023) conducted an experiment demonstrating that *Tribolium castaneum* populations responds similarly to environmental change, regardless of the intensity of the bottleneck they experienced in their demographic history. England et al. (2003) also found no difference in adaptive potential between *Drosophila melanogaster* populations that underwent an intense or diffuse bottleneck designed to produce similar inbreeding levels.

The duration for which populations can recover, which is the time between two bottlenecks, also affects the adaptive potential of populations. For instance, Moxon and Kussell (2017) showed that increasing the severity of bottlenecks or reducing the growth period leads to faster adaptation during pathogen microbial infection. Natural microbial populations that experience frequent bottlenecks, such as pathogens, can adapt to changing environmental conditions. A theoretical study predicted a high probability that some mutations acquired during growth in a given host will be passed to the next one in viruses (Sigal et al., 2018). These results could apply to conservation biology since frequent bottlenecks are commonly observed in wild populations (Hohenlohe et al., 2020). Recent genomic approaches have been used to determine the timing

689 and nature of past population bottlenecks by detecting changes in the shape of the deleterious  
690 variation landscape [see Bortoluzzi et al. (2019) for an application with chicken populations as  
691 well as Cornuet and Luikart (1996) and Peery et al. (2012) for classical approaches].

692 An important area for future research is to investigate whether the cumulative effects of  
693 multiple bottlenecks are additive or synergistic. This effect could be studied experimentally in  
694 microbiology and then applied to natural populations. In theory, multiple bottlenecks will not  
695 have the same impact on the population's ability to adapt from *de novo* mutations and from  
696 standing genetic variation. What matters for adaptation from *de novo* mutation is the current  
697 population census size, which is the result of the last bottleneck only. What matters for adap-  
698 tation from standing genetic variation is the genetic diversity of the population, which results  
699 from past variations in population size (i.e., from the last common ancestor of the population to  
700 the present). Genetic diversity is proportional to the effective population size, which is usually  
701 computed as the harmonic mean of past population sizes for populations of varying sizes (Crow  
702 and Kimura, 2009; Otto and Whitlock, 1997; Charlesworth, 2009). However, other parameters,  
703 such as population structure and selection, can also impact the effective population size. As  
704 these parameters can vary between different environments, comparisons between experimental  
705 and wild populations must be made with caution. Advances in population genomics applied to  
706 conservation biology are very useful in this case and are a fruitful avenue of research (Hohenlohe  
707 et al., 2020). Moreover, even if genetic diversity should theoretically correlate with the response  
708 to selection, this effect is rarely observed in the wild due to interference from other biological  
709 mechanisms [e.g., plasticity or coevolution, see Pujol et al. (2018)]. We would need experiments  
710 with populations undergoing bottlenecks of different severity and duration to assess the differ-  
711 ential impacts of such bottlenecks on genetic variation. Indeed, some studies have observed  
712 that genetic variation quickly recovers after a short bottleneck, with examples of both short-  
713 generation (water vole) and long-generation (white-tailed eagle) species (Oliver and Piertney,  
714 2012; Keller et al., 2001; Hailer et al., 2006). Moreover, low initial genetic variation does not  
715 seem to be limiting for the adaptation of invaders [see the review Bock et al. (2015)]. Thus,  
716 there is likely to be a threshold of severity and duration above which a population struggles to  
717 recover.

718 To conclude, in this section and throughout the review, we have proposed several research  
719 directions and suggested new experiments that could help to understand the adaptation of  
720 bottlenecked populations. The new knowledge gained from these experiments could ultimately  
721 be integrated into existing methods for detecting species most at risk of extinction due to  
722 climate change [reviewed in Hoffmann and Sgrò (2011)].

## 723 5 Perspectives

724 Our review has shown that the impact of bottlenecks on the evolutionary dynamics of popu-  
725 lations is a topic that spans several fields, such as microbiology and conservation biology, and  
726 has inspired theoretical, and empirical works. Our review emphasizes that these fields share

727 a common goal and are not as distinct as previously thought. We believe that an improved  
728 collaboration between these fields will lead to a better understanding of how bottlenecks affect  
729 the evolutionary dynamics of populations.

730 Similar to our review, Alexander et al. (2014) showed how seemingly unrelated fields address  
731 the evolution of declining populations. Specifically, Alexander et al. (2014) emphasized that  
732 evolutionary rescue is a research topic in medicine (e.g., drug resistance evolution in patients  
733 undergoing chemotherapy) and conservation biology (e.g., survival of species undergoing habitat  
734 deterioration). Similar to our review, Alexander et al. (2014) pointed out that integrating  
735 different fields could accelerate our scientific knowledge.

736 We hope that these synthesis reviews will pave the way for empirical studies that combine  
737 different fields. Given the challenges of the 21st century, such as the loss of biodiversity, it would  
738 be highly valuable to employ approaches that enhance our comprehension of biological processes  
739 and our ability to forecast the reaction of natural populations to environmental change.

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747 Loïc Marrec: Conceptualization (Equal), Writing - original draft (20% - Supporting), Writing  
748 - review & editing (Equal).

749 Laure Olazcuaga: Project administration (Lead), Supervision (Lead), Conceptualization (Lead),  
750 Writing – original draft (45% - Lead), Writing - review & editing (Equal).

## 751 **Conflict of interest declaration**

752 The authors declare they have no competing interests.

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## References

S. Abel, P. Abel zur Wiesch, B. M. Davis, and M. K. Waldor. Analysis of bottlenecks in experimental models of infection. *PLoS Pathogens*, 11(6):e1004823, June 2015. ISSN 1553-7374. doi: 10.1371/journal.ppat.1004823. URL <http://dx.doi.org/10.1371/journal.ppat.1004823>.

A. F. Agrawal and M. C. Whitlock. Inferences About the Distribution of Dominance Drawn From Yeast Gene Knockout Data. *Genetics*, 187(2):553–566, Feb. 2011. ISSN 1943-2631. doi: 10.1534/genetics.110.124560. URL <http://dx.doi.org/10.1534/genetics.110.124560>.

H. K. Alexander, G. Martin, O. Y. Martin, and S. Bonhoeffer. Evolutionary rescue: linking theory for conservation and medicine. *Evolutionary Applications*, 7(10):1161–1179, Oct. 2014. ISSN 1752-4571. doi: 10.1111/eva.12221. URL <http://dx.doi.org/10.1111/eva.12221>.

F. W. Allendorf. Genetic drift and the loss of alleles versus heterozygosity. *Zoo Biology*, 5(2):181–190, 1986. ISSN 1098-2361. doi: 10.1002/zoo.1430050212. URL <http://dx.doi.org/10.1002/zoo.1430050212>.

S. L. Ament-Velásquez, C. Gilchrist, A. Rêgo, D. P. Bendixsen, C. Brice, J. M. Grosse-Sommer, N. Rafati, and R. Stelkens. The dynamics of adaptation to stress from standing genetic variation and de novo mutations. *Molecular Biology and Evolution*, 39(11), Nov. 2022. ISSN 1537-1719. doi: 10.1093/molbev/msac242. URL <http://dx.doi.org/10.1093/molbev/msac242>.

J. Bakker, M. van Rijswijk, F. J. Weissing, and R. Bijlsma. Consequences of fragmentation for the ability to adapt to novel environments in experimental *Drosophila* metapopulations: ESF-ConGen Meeting on Integrating Population Genetics and Conservation Biology. *Conservation Genetics*, 11(2):435–448, Apr. 2010. ISSN 1566-0621. doi: 10.1007/s10592-010-0052-5. URL <http://dx.doi.org/10.1007/s10592-010-0052-5>.

A. D. Barnosky, N. Matzke, S. Tomiya, G. O. U. Wogan, B. Swartz, T. B. Quental, C. Marshall, J. L. McGuire, E. L. Lindsey, K. C. Maguire, B. Mersey, and E. A. Ferrer. Has the Earth’s sixth mass extinction already arrived? *Nature*, 471(7336):51–57, Mar. 2011. ISSN 1476-4687. doi: 10.1038/nature09678. URL <http://dx.doi.org/10.1038/nature09678>.

R. D. H. Barrett and D. Schluter. Adaptation from standing genetic variation. *Trends in Ecology & Evolution*, 23(1):38–44, Jan. 2008. ISSN 0169-5347. doi: 10.1016/j.tree.2007.09.008. URL <http://dx.doi.org/10.1016/j.tree.2007.09.008>.

- 789 S. C. H. Barrett and J. R. Kohn. Genetic and evolutionary consequences of small population  
790 size in plants: Implications for conservation. *Genetics and conservation of rare plants...*, pages  
791 3–30, 1991.
- 792 G. Bell and A. Gonzalez. Adaptation and evolutionary rescue in metapopulations experiencing  
793 environmental deterioration. *Science*, 332(6035):1327–1330, June 2011. ISSN 1095-9203. doi:  
794 10.1126/science.1203105. URL <http://dx.doi.org/10.1126/science.1203105>.
- 795 G. Bell, V. Fugère, R. Barrett, B. Beisner, M. Cristescu, G. Fussmann, J. Shapiro, and A. Gon-  
796 zalez. Trophic structure modulates community rescue following acidification. *Proceedings of*  
797 *the Royal Society B: Biological Sciences*, 286(1904):20190856, June 2019. ISSN 1471-2954.  
798 doi: 10.1098/rspb.2019.0856. URL <http://dx.doi.org/10.1098/rspb.2019.0856>.
- 799 G. Bertorelle, F. Raffini, M. Bosse, C. Bortoluzzi, A. Iannucci, E. Trucchi, H. E. Morales,  
800 and C. van Oosterhout. Genetic load: Genomic estimates and applications in non-model  
801 animals. *Nature Reviews Genetics*, 23(8):492–503, Aug. 2022. ISSN 1471-0064. doi: 10.1038/  
802 s41576-022-00448-x. URL <http://dx.doi.org/10.1038/s41576-022-00448-x>.
- 803 E. H. Boakes, J. Wang, and W. Amos. An investigation of inbreeding depression and purging  
804 in captive pedigreed populations. *Heredity*, 98(3):172–182, Dec. 2006. ISSN 1365-2540. doi:  
805 10.1038/sj.hdy.6800923. URL <http://dx.doi.org/10.1038/sj.hdy.6800923>.
- 806 D. G. Bock, C. Caseys, R. D. Cousens, M. A. Hahn, S. M. Heredia, S. Hübner, K. G. Turner,  
807 K. D. Whitney, and L. H. Rieseberg. What we still don’t know about invasion genetics.  
808 *Molecular Ecology*, 24(9):2277–2297, May 2015. doi: 10.1111/mec.13032. URL <http://dx.doi.org/10.1111/mec.13032>.
- 809
- 810 C. Bortoluzzi, M. Bosse, M. F. L. Derks, R. P. M. A. Crooijmans, M. A. M. Groenen, and  
811 H. Megens. The type of bottleneck matters: Insights into the deleterious variation landscape  
812 of small managed populations. *Evolutionary Applications*, 13(2):330–341, Sept. 2019. ISSN  
813 1752-4571. doi: 10.1111/eva.12872. URL <http://dx.doi.org/10.1111/eva.12872>.
- 814 J. L. Bouzat. Conservation genetics of population bottlenecks: The role of chance, selection,  
815 and history. *Conservation Genetics*, 11(2):463–478, Apr. 2010. ISSN 1572-9737. doi: 10.  
816 1007/s10592-010-0049-0. URL <http://dx.doi.org/10.1007/s10592-010-0049-0>.
- 817 M. K. Burke. Embracing Complexity: Yeast Evolution Experiments Featuring Standing Genetic  
818 Variation. *Journal of Molecular Evolution*, Feb. 2023. ISSN 0022-2844, 1432-1432. doi:  
819 10.1007/s00239-023-10094-4. URL <http://dx.doi.org/10.1007/s00239-023-10094-4>.
- 820 M. K. Burke, G. Liti, and A. D. Long. Standing Genetic Variation Drives Repeatable Experi-  
821 mental Evolution in Outcrossing Populations of *Saccharomyces cerevisiae*. *Molecular Biology*  
822 *and Evolution*, 31(12):3228–3239, Dec. 2014. ISSN 0737-4038. doi: 10.1093/molbev/msu256.  
823 URL <http://dx.doi.org/10.1093/molbev/msu256>.

- 824 P. R. A. Campos and L. M. Wahl. The adaptation rate of asexuals: Deleterious mutations,  
825 clonal interference and population bottlenecks. *Evolution*, 64(7):1973–1983, July 2010. doi:  
826 10.1111/j.1558-5646.2010.00981.x. URL <http://dx.doi.org/10.1111/j.1558-5646.2010.00981.x>.
- 828 S. M. Carlson, C. J. Cunningham, and P. A. Westley. Evolutionary rescue in a changing  
829 world. *Trends in Ecology & Evolution*, 29(9):521–530, Sept. 2014. ISSN 0169-5347. doi:  
830 10.1016/j.tree.2014.06.005. URL <http://dx.doi.org/10.1016/j.tree.2014.06.005>.
- 831 G. Ceballos, P. R. Ehrlich, A. D. Barnosky, A. García, R. M. Pringle, and T. M. Palmer.  
832 Accelerated modern human-induced species losses: Entering the sixth mass extinction.  
833 *Science Advances*, 1(5):e1400253, June 2015. doi: 10.1126/sciadv.1400253. URL <http://dx.doi.org/10.1126/sciadv.1400253>.
- 835 P. P. Chakraborty, L. R. Nemzer, and R. Kassen. Experimental evidence that network topology  
836 can accelerate the spread of beneficial mutations. *Evolution Letters*, 7(6):447–456, Dec. 2023.  
837 ISSN 2056-3744. doi: 10.1093/evlett/qrad047. URL <http://dx.doi.org/10.1093/evlett/qrad047>.
- 839 R. Chakraborty and M. Nei. Genetic differentiation of quantitative characters between pop-  
840 ulations or species: I. mutation and random genetic drift. *Genetical Research*, 39(3):  
841 303–314, June 1982. ISSN 1469-5073. doi: 10.1017/s0016672300020978. URL <http://dx.doi.org/10.1017/s0016672300020978>.
- 843 B. Charlesworth. Effective population size and patterns of molecular evolution and variation.  
844 *Nature Reviews Genetics*, 10(3):195–205, Mar. 2009. ISSN 1471-0064. doi: 10.1038/nrg2526.  
845 URL <http://dx.doi.org/10.1038/nrg2526>.
- 846 D. Charlesworth and B. Charlesworth. Inbreeding depression and its evolutionary consequences.  
847 *Annual Review of Ecology and Systematics*, 18(1):237–268, Nov. 1987. ISSN 0066-4162. doi:  
848 10.1146/annurev.es.18.110187.001321. URL <http://dx.doi.org/10.1146/annurev.es.18.110187.001321>.
- 850 J. E. Chiucchi and H. L. Gibbs. Similarity of contemporary and historical gene flow among  
851 highly fragmented populations of an endangered rattlesnake: Gene flow in endangered rat-  
852 tlesnakes. *Molecular Ecology*, 19(24):5345–5358, Oct. 2010. ISSN 0962-1083. doi: 10.1111/j.  
853 1365-294x.2010.04860.x. URL <http://dx.doi.org/10.1111/j.1365-294x.2010.04860.x>.
- 854 D. K. Clarke, E. A. Duarte, A. Moya, S. F. Elena, E. Domingo, E Domingo, J J Holland, J. J.  
855 Holland, and J. H. Holland. Genetic bottlenecks and population passages cause profound  
856 fitness differences in RNA viruses. *Journal of Virology*, 67(1):222–228, Jan. 1993. doi: 10.  
857 1128/jvi.67.1.222-228.1993. URL <http://dx.doi.org/10.1128/jvi.67.1.222-228.1993>.



- 858 G. Clayton and A. Robertson. Mutation and Quantitative Variation. *The American Naturalist*,  
859 May 1955. ISSN 0003-0147. doi: 10.1086/281874. URL [http://dx.doi.org/10.1086/  
860 281874](http://dx.doi.org/10.1086/281874).
- 861 J. M. Cornuet and G. Luikart. Description and power analysis of two tests for detecting recent  
862 population bottlenecks from allele frequency data. *Genetics*, 144(4):2001–2014, Dec. 1996.  
863 ISSN 1943-2631. doi: 10.1093/genetics/144.4.2001. URL [http://dx.doi.org/10.1093/  
864 genetics/144.4.2001](http://dx.doi.org/10.1093/genetics/144.4.2001).
- 865 P. Crnokrak and S. C. H. Barrett. Perspective: Purging the Genetic Load: A Review of  
866 the Experimental Evidence. *Evolution*, 56(12):2347–2358, 2002. ISSN 1558-5646. doi: 10.  
867 1111/j.0014-3820.2002.tb00160.x. URL [http://dx.doi.org/10.1111/j.  
868 tb00160.x](http://dx.doi.org/10.1111/j.0014-3820.2002.tb00160.x).
- 869 P. Crnokrak and D. A. Roff. Dominance variance: Associations with selection and fitness.  
870 *Heredity*, 75(5):530–540, Nov. 1995. ISSN 1365-2540. doi: 10.1038/hdy.1995.169. URL  
871 <http://dx.doi.org/10.1038/hdy.1995.169>.
- 872 J. Crow and M. Kimura. *An Introduction to Population Genetics Theory*. Blackburn Press,  
873 2009. ISBN 9781932846126. URL <https://books.google.ch/books?id=VWqKPwAACAAJ>.
- 874 P. Czappon, F. Blanquart, H. Uecker, and F. Débarre. The Effect of Habitat Choice on  
875 Evolutionary Rescue in Subdivided Populations. *The American Naturalist*, 197(6):625–643,  
876 June 2021. ISSN 1537-5323. doi: 10.1086/714034. URL [http://dx.doi.org/10.1086/  
877 714034](http://dx.doi.org/10.1086/714034).
- 878 J. Dayton and P. Szczys. Metapopulation connectivity retains genetic diversity following a  
879 historical bottleneck in a federally endangered seabird. *Ornithological Applications*, 123(4),  
880 Sept. 2021. ISSN 2732-4621. doi: 10.1093/ornithapp/duab037. URL [http://dx.doi.org/  
881 10.1093/ornithapp/duab037](http://dx.doi.org/10.1093/ornithapp/duab037).
- 882 M. De Ste Croix, J. Holmes, J. J. Wanford, E. R. Moxon, M. R. Oggioni, and C. D. Bayliss.  
883 Selective and non-selective bottlenecks as drivers of the evolution of hypermutable bacterial  
884 loci. *Molecular Microbiology*, 113(3):672–681, Mar. 2020. ISSN 1365-2958. doi: 10.1111/  
885 mmi.14453. URL <http://dx.doi.org/10.1111/mmi.14453>.
- 886 J. A. G. M. de Visser and D. E. Rozen. Limits to adaptation in asexual populations. *Journal*  
887 *of Evolutionary Biology*, 18(4):779–788, July 2005. doi: 10.1111/j.1420-9101.2005.00879.x.  
888 URL <http://dx.doi.org/10.1111/j.1420-9101.2005.00879.x>.
- 889 M. A. DeRose and D. A. Roff. A Comparison of Inbreeding Depression in Life-History and  
890 Morphological Traits in Animals. *Evolution*, 53(4):1288–1292, Aug. 1999. ISSN 0014-  
891 3820. doi: 10.1111/j.1558-5646.1999.tb04541.x. URL [http://dx.doi.org/10.1111/j.  
892 1558-5646.1999.tb04541.x](http://dx.doi.org/10.1111/j.1558-5646.1999.tb04541.x).

893 N. Dussex, H. E. Morales, C. Grossen, L. Dalén, and C. van Oosterhout. Purging and accu-  
894 mulation of genetic load in conservation. *Trends in Ecology & Evolution*, 0(0), June 2023.  
895 ISSN 0169-5347. doi: 10.1016/j.tree.2023.05.008. URL [http://dx.doi.org/10.1016/j.](http://dx.doi.org/10.1016/j.tree.2023.05.008)  
896 [tree.2023.05.008](http://dx.doi.org/10.1016/j.tree.2023.05.008).

897 S. F. Elena and R. E. Lenski. Evolution experiments with microorganisms: The dynamics  
898 and genetic bases of adaptation. *Nature Reviews Genetics*, 4(6):457–469, June 2003. doi:  
899 10.1038/nrg1088. URL <http://dx.doi.org/10.1038/nrg1088>.

900 N. Ellstrand and D. Elam. Population Genetic Consequences of Small Population Size: Im-  
901 plications for Plant Conservation. *Annual Review of Ecology and Systematics*, 24:217–242,  
902 Nov. 2003. doi: 10.1146/annurev.es.24.110193.001245. URL [http://dx.doi.org/10.1146/](http://dx.doi.org/10.1146/annurev.es.24.110193.001245)  
903 [annurev.es.24.110193.001245](http://dx.doi.org/10.1146/annurev.es.24.110193.001245).

904 P. R. England, G. H. R. Osler, L. M. Woodworth, M. E. Montgomery, D. A. Briscoe, and  
905 R. Frankham. Effects of intense versus diffuse population bottlenecks on microsatellite genetic  
906 diversity and evolutionary potential. *Conservation Genetics*, 4(5):595–604, Sept. 2003. doi:  
907 10.1023/a:1025639811865. URL <http://dx.doi.org/10.1023/a:1025639811865>.

908 D. S. Falconer. Introduction to quantitative genetics. *Introduction to quantitative genetics.*,  
909 1960.

910 S. W. Fitzpatrick and B. N. Reid. Does gene flow aggravate or alleviate maladaptation to  
911 environmental stress in small populations? *Evolutionary Applications*, 12(7):1402–1416, Feb.  
912 2019. ISSN 1752-4571. doi: 10.1111/eva.12768. URL [http://dx.doi.org/10.1111/eva.](http://dx.doi.org/10.1111/eva.12768)  
913 [12768](http://dx.doi.org/10.1111/eva.12768).

914 R. Frankham. Inbreeding in the wild really does matter. *Heredity*, 104(2):124–124, Nov. 2009.  
915 ISSN 1365-2540. doi: 10.1038/hdy.2009.155. URL [http://dx.doi.org/10.1038/hdy.2009.](http://dx.doi.org/10.1038/hdy.2009.155)  
916 [155](http://dx.doi.org/10.1038/hdy.2009.155).

917 R. Frankham. Genetic rescue of small inbred populations: meta-analysis reveals large and  
918 consistent benefits of gene flow. *Molecular Ecology*, 24(11):2610–2618, Mar. 2015. ISSN  
919 1365-294X. doi: 10.1111/mec.13139. URL <http://dx.doi.org/10.1111/mec.13139>.

920 R. Frankham, K. Lees, M. E. Montgomery, P. R. England, E. H. Lowe, and D. A. Briscoe.  
921 Do population size bottlenecks reduce evolutionary potential? *Animal Conservation*, 2  
922 (4):255–260, Nov. 1999. ISSN 1469-1795. doi: 10.1111/j.1469-1795.1999.tb00071.x. URL  
923 <http://dx.doi.org/10.1111/j.1469-1795.1999.tb00071.x>.

924 R. Frankham, D. Briscoe, and J. Ballou. *Introduction to Conservation Genetics*. Cambridge  
925 University Press, 2002. ISBN 9780521639859. URL [https://books.google.ch/books?id=](https://books.google.ch/books?id=F-XB8hqZ4s8C)  
926 [F-XB8hqZ4s8C](https://books.google.ch/books?id=F-XB8hqZ4s8C).

- 927 R. Frankham, J. Ballou, K. Ralls, M. Eldridge, M. Dudash, C. Fenster, R. Lacy, and P. Sun-  
928 nucks. *Genetic Management of Fragmented Animal and Plant Populations*. Oxford Uni-  
929 versity Press, 2017. ISBN 9780198783398. URL [https://books.google.ch/books?id=](https://books.google.ch/books?id=oksrDwAAQBAJ)  
930 [oksrDwAAQBAJ](https://books.google.ch/books?id=oksrDwAAQBAJ).
- 931 I. R. Franklin and R. Frankham. How large must populations be to retain evolutionary poten-  
932 tial? *Animal Conservation*, 1(1):69–70, Feb. 1998. ISSN 1469-1795. doi: 10.1111/j.1469-1795.  
933 1998.tb00228.x. URL <http://dx.doi.org/10.1111/j.1469-1795.1998.tb00228.x>.
- 934 P. A. Fuerst and T. Maruyama. Considerations on the conservation of alleles and of genic  
935 heterozygosity in small managed populations. *Zoo Biology*, 5(2):171–179, Jan. 1986. ISSN  
936 0733-3188, 1098-2361. doi: 10.1002/zoo.1430050211. URL [http://dx.doi.org/10.1002/](http://dx.doi.org/10.1002/zoo.1430050211)  
937 [zoo.1430050211](http://dx.doi.org/10.1002/zoo.1430050211).
- 938 J. Gamblin, S. Gandon, F. Blanquart, and A. Lambert. Bottlenecks can constrain and channel  
939 evolutionary paths. *GENETICS*, 224(2):iyad001, May 2023. ISSN 1943-2631. doi: 10.1093/  
940 genetics/iyad001. URL <http://dx.doi.org/10.1093/genetics/iyad001>.
- 941 L. Garoff, F. Pietsch, D. L. Huseby, T. Lilja, G. Brandis, and D. Hughes. Population Bottlenecks  
942 Strongly Influence the Evolutionary Trajectory to Fluoroquinolone Resistance in *Escherichia*  
943 *coli*. *Molecular Biology and Evolution*, 37(6):1637–1646, June 2020. doi: 10.1093/molbev/  
944 msaa032. URL <http://dx.doi.org/10.1093/molbev/msaa032>.
- 945 N. S. Gladstone, N. L. Garrison, T. Lane, P. D. Johnson, J. Garner, and N. V. Whelan. Pop-  
946 ulation genomics reveal low differentiation and complex demographic histories in a highly  
947 fragmented and endangered freshwater mussel. *Aquatic Conservation: Marine and Freshwa-*  
948 *ter Ecosystems*, 32(8):1235–1248, June 2022. ISSN 1099-0755. doi: 10.1002/aqc.3849. URL  
949 <http://dx.doi.org/10.1002/aqc.3849>.
- 950 R. Gomulkiewicz and R. D. Holt. When does evolution by natural selection prevent extinction?  
951 *Evolution*, 49(1):201, Feb. 1995. ISSN 0014-3820. doi: 10.2307/2410305. URL [http://dx.](http://dx.doi.org/10.2307/2410305)  
952 [doi.org/10.2307/2410305](http://dx.doi.org/10.2307/2410305).
- 953 A. Gonzalez and G. Bell. Evolutionary rescue and adaptation to abrupt environmental change  
954 depends upon the history of stress. *Philosophical Transactions of the Royal Society B: Biolog-*  
955 *ical Sciences*, 368(1610):20120079, Jan. 2013. ISSN 1471-2970. doi: 10.1098/rstb.2012.0079.  
956 URL <http://dx.doi.org/10.1098/rstb.2012.0079>.
- 957 S. J. Goodman, H. B. Tamate, R. Wilson, J. Nagata, S. Tatsuzawa, G. M. Swanson, J. M.  
958 Pemberton, and D. R. McCullough. Bottlenecks, drift and differentiation: the population  
959 structure and demographic history of sika deer (*cervus nippon*) in the japanese archipelago.  
960 *Molecular Ecology*, 10(6):1357–1370, June 2001. ISSN 1365-294X. doi: 10.1046/j.1365-294x.  
961 2001.01277.x. URL <http://dx.doi.org/10.1046/j.1365-294x.2001.01277.x>.

- 962 C. Grossen, F. Guillaume, L. F. Keller, and D. Croll. Purging of highly deleterious muta-  
963 tions through severe bottlenecks in alpine ibex. *Nature Communications*, 11(1), Feb. 2020.  
964 ISSN 2041-1723. doi: 10.1038/s41467-020-14803-1. URL [http://dx.doi.org/10.1038/  
965 s41467-020-14803-1](http://dx.doi.org/10.1038/s41467-020-14803-1).
- 966 M. Gyllenberg and I. Hanski. Single-species metapopulation dynamics: A structured model.  
967 *Theoretical Population Biology*, 42(1):35–61, Aug. 1992. ISSN 0040-5809. doi: 10.1016/  
968 0040-5809(92)90004-d. URL [http://dx.doi.org/10.1016/0040-5809\(92\)90004-D](http://dx.doi.org/10.1016/0040-5809(92)90004-D).
- 969 F. Hailer, B. Helander, A. O. Folkestad, S. A. Ganusevich, S. Garstad, P. Hauff, C. Koren,  
970 T. Nygård, V. Volke, C. Vilà, and H. Ellegren. Bottlenecked but long-lived: High genetic  
971 diversity retained in white-tailed eagles upon recovery from population decline. *Biology  
972 Letters*, 2(2):316–319, Mar. 2006. doi: 10.1098/rsbl.2006.0453. URL [http://dx.doi.org/  
973 10.1098/rsbl.2006.0453](http://dx.doi.org/10.1098/rsbl.2006.0453).
- 974 J. B. S. Haldane. A Mathematical Theory of Natural and Artificial Selection, Part V: Selection  
975 and Mutation. *Mathematical Proceedings of the Cambridge Philosophical Society*, 23(7):  
976 838–844, July 1927. ISSN 1469-8064, 0305-0041. doi: 10.1017/S0305004100015644. URL  
977 <http://dx.doi.org/10.1017/S0305004100015644>.
- 978 A. Handel and D. E. Rozen. The impact of population size on the evolution of asexual microbes  
979 on smooth versus rugged fitness landscapes. *BMC Evolutionary Biology*, 9(1):236, Sept.  
980 2009. ISSN 1471-2148. doi: 10.1186/1471-2148-9-236. URL [http://dx.doi.org/10.1186/  
981 1471-2148-9-236](http://dx.doi.org/10.1186/1471-2148-9-236).
- 982 I. Hanski and O. Ovaskainen. Metapopulation theory for fragmented landscapes. *Theoretical  
983 Population Biology*, 64(1):119–127, Aug. 2003. ISSN 0040-5809. doi: 10.1016/s0040-5809(03)  
984 00022-4. URL [http://dx.doi.org/10.1016/S0040-5809\(03\)00022-4](http://dx.doi.org/10.1016/S0040-5809(03)00022-4).
- 985 P. W. Hedrick and A. Garcia-Dorado. Understanding Inbreeding Depression, Purging, and  
986 Genetic Rescue. *Trends in Ecology & Evolution*, 31(12):940–952, Dec. 2016. ISSN 0169-5347.  
987 doi: 10.1016/j.tree.2016.09.005. URL <http://dx.doi.org/10.1016/j.tree.2016.09.005>.
- 988 J. M. Heffernan and L. M. Wahl. The effects of genetic drift in experimental evolution.  
989 *Theoretical Population Biology*, 62(4):349–356, Dec. 2002. ISSN 0040-5809. doi: 10.1016/  
990 S0040-5809(02)00002-3. URL [http://dx.doi.org/10.1016/S0040-5809\(02\)00002-3](http://dx.doi.org/10.1016/S0040-5809(02)00002-3).
- 991 K. Heilbron, M. Toll-Riera, M. Kojadinovic, and R. C. MacLean. Fitness Is Strongly Influenced  
992 by Rare Mutations of Large Effect in a Microbial Mutation Accumulation Experiment. *Ge-  
993 netics*, 197(3):981–990, July 2014. ISSN 1943-2631. doi: 10.1534/genetics.114.163147. URL  
994 <http://dx.doi.org/10.1534/genetics.114.163147>.
- 995 L. Hindersin and A. Traulsen. Counterintuitive properties of the fixation time in network-  
996 structured populations. *Journal of The Royal Society Interface*, 11(99):20140606, Oct. 2014.

997 ISSN 1742-5662. doi: 10.1098/rsif.2014.0606. URL <http://dx.doi.org/10.1098/rsif.>  
998 2014.0606.

999 A. A. Hoffmann and C. M. Sgrò. Climate change and evolutionary adaptation. *Nature*, 470  
1000 (7335):479–485, Feb. 2011. ISSN 1476-4687. doi: 10.1038/nature09670. URL <http://dx.>  
1001 [doi.org/10.1038/nature09670](http://dx.doi.org/10.1038/nature09670).

1002 A. A. Hoffmann, C. M. Sgrò, and T. N. Kristensen. Revisiting Adaptive Potential, Population  
1003 Size, and Conservation. *Trends in Ecology and Evolution*, 32(7):506–517, July 2017. doi:  
1004 10.1016/j.tree.2017.03.012. URL <http://dx.doi.org/10.1016/j.tree.2017.03.012>.

1005 P. A. Hohenlohe, W. C. Funk, and O. P. Rajora. Population genomics for wildlife conservation  
1006 and management. *Molecular Ecology*, 30(1):62–82, Nov. 2020. ISSN 1365-294X. doi: 10.  
1007 1111/mec.15720. URL <http://dx.doi.org/10.1111/mec.15720>.

1008 B. Houchmandzadeh and M. Vallade. The fixation probability of a beneficial mutation in  
1009 a geographically structured population. *New Journal of Physics*, 13(7):073020, July 2011.  
1010 ISSN 1367-2630. doi: 10.1088/1367-2630/13/7/073020. URL <http://dx.doi.org/10.1088/>  
1011 [1367-2630/13/7/073020](http://dx.doi.org/10.1088/1367-2630/13/7/073020).

1012 R. A. Hufbauer, M. Szűcs, E. Kasyon, C. Youngberg, M. J. Koontz, C. Richards, T. Tuff,  
1013 and B. A. Melbourne. Three types of rescue can avert extinction in a changing environ-  
1014 ment. *Proceedings of the National Academy of Sciences*, 112(33):10557–10562, Aug. 2015.  
1015 ISSN 1091-6490. doi: 10.1073/pnas.1504732112. URL <http://dx.doi.org/10.1073/pnas.>  
1016 [1504732112](http://dx.doi.org/10.1073/pnas.1504732112).

1017 D. L. Huseby, F. Pietsch, G. Brandis, L. Garoff, A. Tegehall, A. Tegehall, and D. Hughes.  
1018 Mutation Supply and Relative Fitness Shape the Genotypes of Ciprofloxacin-Resistant  
1019 *Escherichia coli*. *Molecular Biology and Evolution*, 34(5):1029–1039, May 2017. doi:  
1020 10.1093/molbev/msx052. URL <http://dx.doi.org/10.1093/molbev/msx052>.

1021 P. K. Ingvarsson. Restoration of genetic variation lost – the genetic rescue hypothesis. *Trends in*  
1022 *Ecology & Evolution*, 16(2):62–63, Feb. 2001. ISSN 0169-5347. doi: 10.1016/s0169-5347(00)  
1023 02065-6. URL [http://dx.doi.org/10.1016/s0169-5347\(00\)02065-6](http://dx.doi.org/10.1016/s0169-5347(00)02065-6).

1024 M. Jangjoo, S. F. Matter, J. Roland, and N. Keyghobadi. Connectivity rescues genetic diversity  
1025 after a demographic bottleneck in a butterfly population network. *Proceedings of the National*  
1026 *Academy of Sciences*, 113(39):10914–10919, Sept. 2016. ISSN 1091-6490. doi: 10.1073/pnas.  
1027 1600865113. URL <http://dx.doi.org/10.1073/pnas.1600865113>.

1028 T. J. Kawecki, R. E. Lenski, D. Ebert, B. Hollis, I. Olivieri, and M. C. Whitlock. Experimental  
1029 evolution. *Trends in Ecology & Evolution*, 27(10):547–560, Oct. 2012. ISSN 0169-5347. doi:  
1030 10.1016/j.tree.2012.06.001. URL <http://dx.doi.org/10.1016/j.tree.2012.06.001>.

- 1031 L. Keller. Inbreeding effects in wild populations. *Trends in Ecology & Evolution*, 17(5):230–241,  
1032 May 2002. ISSN 0169-5347. doi: 10.1016/s0169-5347(02)02489-8. URL [http://dx.doi.org/  
1033 10.1016/s0169-5347\(02\)02489-8](http://dx.doi.org/10.1016/s0169-5347(02)02489-8).
- 1034 L. F. Keller, K. J. Jeffery, P. Arcese, M. A. Beaumont, W. M. Hochachka, J. N. M. Smith,  
1035 and M. W. Bruford. Immigration and the ephemerality of a natural population bottleneck:  
1036 Evidence from molecular markers. *Proceedings of the Royal Society of London. Series B:  
1037 Biological Sciences*, 268(1474):1387–1394, July 2001. doi: 10.1098/rspb.2001.1607. URL  
1038 <http://dx.doi.org/10.1098/rspb.2001.1607>.
- 1039 N. Keyghobadi. The genetic implications of habitat fragmentation for animals. *Canadian  
1040 Journal of Zoology*, 85(10):1049–1064, Oct. 2007. ISSN 1480-3283. doi: 10.1139/z07-095.  
1041 URL <http://dx.doi.org/10.1139/Z07-095>.
- 1042 M. Kimura. Random Genetic Drift in Multi-Allelic Locus. *Evolution*, 9(4):419, Dec. 1955. ISSN  
1043 00143820. doi: 10.2307/2405476. URL <http://dx.doi.org/10.2307/2405476>.
- 1044 M. Kirkpatrick and P. Jarne. The Effects of a Bottleneck on Inbreeding Depression and the  
1045 Genetic Load. *The American Naturalist*, 155(2):154–167, Feb. 2000. ISSN 1537-5323. doi:  
1046 10.1086/303312. URL <http://dx.doi.org/10.1086/303312>.
- 1047 D. Kleinman-Ruiz, M. Lucena-Perez, B. Villanueva, J. Fernández, A. P. Saveljev,  
1048 M. Ratkiewicz, K. Schmidt, N. Galtier, A. García-Dorado, and J. A. Godoy. Purging of  
1049 deleterious burden in the endangered iberian lynx. *Proceedings of the National Academy  
1050 of Sciences*, 119(11), Mar. 2022. ISSN 1091-6490. doi: 10.1073/pnas.2110614119. URL  
1051 <http://dx.doi.org/10.1073/pnas.2110614119>.
- 1052 P. L. Klerks, G. N. Athrey, and P. L. Leberg. Response to selection for increased heat tolerance  
1053 in a small fish species, with the response decreased by a population bottleneck. *Frontiers in  
1054 Ecology and Evolution*, 7, July 2019. ISSN 2296-701X. doi: 10.3389/fevo.2019.00270. URL  
1055 <http://dx.doi.org/10.3389/fevo.2019.00270>.
- 1056 J. Lachapelle, N. Colegrave, and G. Bell. The effect of selection history on extinction risk during  
1057 severe environmental change. *Journal of Evolutionary Biology*, 30(10):1872–1883, Aug. 2017.  
1058 ISSN 1420-9101. doi: 10.1111/jeb.13147. URL <http://dx.doi.org/10.1111/jeb.13147>.
- 1059 R. Lande. Genetics and demography in biological conservation. *Science*, 241(4872):1455–1460,  
1060 Sept. 1988. ISSN 1095-9203. doi: 10.1126/science.3420403. URL [http://dx.doi.org/10.  
1061 1126/science.3420403](http://dx.doi.org/10.1126/science.3420403).
- 1062 R. Lande, S. Engen, B.-E. Sæther, and B.-E. Saether. Extinction times in finite metapopulation  
1063 models with stochastic local dynamics. *Oikos*, 83(2):383, Nov. 1998. ISSN 0030-1299. doi:  
1064 10.2307/3546853. URL <http://dx.doi.org/10.2307/3546853>.

- 1065 J. S. LeClair and L. M. Wahl. The impact of population bottlenecks on microbial adaptation.  
1066 *Journal of Statistical Physics*, 172(1):114–125, Nov. 2017. ISSN 1572-9613. doi: 10.1007/  
1067 s10955-017-1924-6. URL <http://dx.doi.org/10.1007/s10955-017-1924-6>.
- 1068 D. M. Leigh, H. E. L. Lischer, F. Guillaume, C. Grossen, and T. Günther. Disentangling  
1069 adaptation from drift in bottlenecked and reintroduced populations of Alpine ibex. *Molecular*  
1070 *Ecology Resources*, 21(7):2350–2363, 2021. ISSN 1755-0998. doi: 10.1111/1755-0998.13442.  
1071 URL <http://dx.doi.org/10.1111/1755-0998.13442>.
- 1072 E. Lieberman, C. Hauert, and M. A. Nowak. Evolutionary dynamics on graphs. *Nature*,  
1073 433(7023):312–316, Jan. 2005. ISSN 1476-4687. doi: 10.1038/nature03204. URL <http://dx.doi.org/10.1038/nature03204>.
- 1074
- 1075 C. Lopez-Fanjul and A. Villaverde. Inbreeding Increases Genetic Variance for Viability in  
1076 *Drosophila melanogaster*. *Evolution*, 43(8):1800–1804, 1989. ISSN 0014-3820. doi: 10.2307/  
1077 2409394. URL <http://dx.doi.org/10.2307/2409394>.
- 1078 M. Lynch and W. G. Hill. Phenotypic Evolution by Neutral Mutation. *Evolution*, 40(5):915–  
1079 935, Sept. 1986. ISSN 0014-3820. doi: 10.1111/j.1558-5646.1986.tb00561.x. URL <http://dx.doi.org/10.1111/j.1558-5646.1986.tb00561.x>.
- 1080
- 1081 M. Lynch, J. Conery, and R. Burger. Mutation Accumulation and the Extinction of Small  
1082 Populations. *The American Naturalist*, 146(4):489–518, Oct. 1995. ISSN 0003-0147, 1537-  
1083 5323. doi: 10.1086/285812. URL <http://dx.doi.org/10.1086/285812>.
- 1084 E. López-Cortegano, E. Moreno, and A. García-Dorado. Genetic purging in captive endangered  
1085 ungulates with extremely low effective population sizes. *Heredity*, 127(5):433–442, Sept. 2021.  
1086 ISSN 1365-2540. doi: 10.1038/s41437-021-00473-2. URL <http://dx.doi.org/10.1038/s41437-021-00473-2>.
- 1087
- 1088 N. Mahrt, A. Tietze, S. Künzel, S. Franzenburg, C. Barbosa, G. Jansen, and H. Schulenburg.  
1089 Bottleneck size and selection level reproducibly impact evolution of antibiotic resistance.  
1090 *Nature Ecology and Evolution*, 5(9):1233–1242, July 2021. doi: 10.1038/s41559-021-01511-2.  
1091 URL <http://dx.doi.org/10.1038/s41559-021-01511-2>.
- 1092 D. A. Marad, S. W. Buskirk, and G. I. Lang. Altered access to beneficial mutations slows  
1093 adaptation and biases fixed mutations in diploids. *Nature Ecology and Evolution*, 2(5):  
1094 882–889, Mar. 2018. doi: 10.1038/s41559-018-0503-9. URL <http://dx.doi.org/10.1038/s41559-018-0503-9>.
- 1095
- 1096 L. Marrec, I. Lamberti, and A.-F. Bitbol. Toward a Universal Model for Spatially Structured  
1097 Populations. *Physical Review Letters*, 127(21):218102, Nov. 2021. doi: 10.1103/PhysRevLett.  
1098 127.218102. URL <http://dx.doi.org/10.1103/PhysRevLett.127.218102>.

- 1099 C. D. Marsden, D. Ortega-Del Vecchyo, D. P. O'Brien, J. F. Taylor, O. Ramirez, C. Vilà,  
1100 T. Marques-Bonet, R. D. Schnabel, R. K. Wayne, and K. E. Lohmueller. Bottlenecks and  
1101 selective sweeps during domestication have increased deleterious genetic variation in dogs.  
1102 *Proceedings of the National Academy of Sciences*, 113(1):152–157, Jan. 2016. doi: 10.1073/  
1103 pnas.1512501113. URL <http://dx.doi.org/10.1073/pnas.1512501113>.
- 1104 M. J. McDonald, D. P. Rice, and M. M. Desai. Sex speeds adaptation by altering the dynamics  
1105 of molecular evolution. *Nature*, 531(7593):233–236, Mar. 2016. ISSN 1476-4687. doi: 10.  
1106 1038/nature17143. URL <http://dx.doi.org/10.1038/nature17143>.
- 1107 B. A. Melbourne and A. Hastings. Extinction risk depends strongly on factors contributing  
1108 to stochasticity. *Nature*, 454(7200):100–103, July 2008. ISSN 1476-4687. doi: 10.1038/  
1109 nature06922. URL <http://dx.doi.org/10.1038/nature06922>.
- 1110 E. Milot, H. Weimerskirch, P. Duchesne, and L. Bernatchez. Surviving with low genetic  
1111 diversity: the case of albatrosses. *Proceedings of the Royal Society B: Biological Sci-*  
1112 *ences*, 274(1611):779–787, Jan. 2007. ISSN 1471-2954. doi: 10.1098/rspb.2006.0221. URL  
1113 <http://dx.doi.org/10.1098/rspb.2006.0221>.
- 1114 R. Moxon and E. Kussell. The impact of bottlenecks on microbial survival, adaptation, and  
1115 phenotypic switching in host-pathogen interactions: Bottlenecks in host-pathogen interac-  
1116 tions. *Evolution*, 71(12):2803–2816, Oct. 2017. ISSN 0014-3820. doi: 10.1111/evo.13370.  
1117 URL <http://dx.doi.org/10.1111/evo.13370>.
- 1118 H. Muller. The relation of recombination to mutational advance. *Mutation Re-*  
1119 *search/Fundamental and Molecular Mechanisms of Mutagenesis*, 1(1):2–9, May 1964. ISSN  
1120 0027-5107. doi: 10.1016/0027-5107(64)90047-8. URL [http://dx.doi.org/10.1016/](http://dx.doi.org/10.1016/0027-5107(64)90047-8)  
1121 [0027-5107\(64\)90047-8](http://dx.doi.org/10.1016/0027-5107(64)90047-8).
- 1122 H. J. Muller. Some genetic aspects of sex. *The American Naturalist*, 66(703):118–138, 1932.  
1123 doi: 10.1086/280418. URL <https://doi.org/10.1086/280418>.
- 1124 M. Nei, T. Maruyama, and R. Chakraborty. The bottleneck effect and genetic variability in  
1125 populations. *Evolution*, 29(1):1, Mar. 1975. ISSN 0014-3820. doi: 10.2307/2407137. URL  
1126 <http://dx.doi.org/10.2307/2407137>.
- 1127 S. W. Nordstrom, R. A. Hufbauer, L. Olazcuaga, L. F. Durkee, and B. A. Melbourne. How  
1128 density dependence, genetic erosion and the extinction vortex impact evolutionary rescue.  
1129 *Proceedings of the Royal Society B: Biological Sciences*, 290(2011), Nov. 2023. ISSN 1471-  
1130 2954. doi: 10.1098/rspb.2023.1228. URL <http://dx.doi.org/10.1098/rspb.2023.1228>.
- 1131 L. Olazcuaga, B. Lincke, S. DeLacey, L. F. Durkee, B. A. Melbourne, and R. A. Hufbauer.  
1132 Population demographic history and evolutionary rescue: Influence of a bottleneck event.  
1133 *Evolutionary Applications*, 16(8):1483–1495, Aug. 2023. ISSN 1752-4571, 1752-4571. doi:  
1134 10.1111/eva.13581. URL <http://dx.doi.org/10.1111/eva.13581>.



- 1135 M. Oliver and S. B. Piertney. Selection maintains MHC diversity through a natural population  
1136 bottleneck. *Molecular Biology and Evolution*, 29(7):1713–1720, July 2012. doi: 10.1093/  
1137 molbev/mss063. URL <http://dx.doi.org/10.1093/molbev/mss063>.
- 1138 M. Ørsted, A. A. Hoffmann, E. Sverrisdóttir, K. L. Nielsen, and T. N. Kristensen. Genomic  
1139 variation predicts adaptive evolutionary responses better than population bottleneck history.  
1140 *PLoS Genetics*, 15(6):e1008205, June 2019. ISSN 1553-7390. doi: 10.1371/journal.pgen.  
1141 1008205. URL <http://dx.doi.org/10.1371/journal.pgen.1008205>.
- 1142 M. Ørsted, E. Yashiro, A. A. Hoffmann, and T. N. Kristensen. Population bottlenecks constrain  
1143 host microbiome diversity and genetic variation impeding fitness. *PLOS Genetics*, 18(5):  
1144 e1010206, May 2022. ISSN 1553-7404. doi: 10.1371/journal.pgen.1010206. URL <http://dx.doi.org/10.1371/journal.pgen.1010206>.
- 1146 M. M. Osmond and C. de Mazancourt. How competition affects evolutionary rescue. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 368  
1147 (1610):20120085, Jan. 2013. ISSN 1471-2970. doi: 10.1098/rstb.2012.0085. URL <http://dx.doi.org/10.1098/rstb.2012.0085>.
- 1150 S. P. Otto and M. C. Whitlock. The Probability of Fixation in Populations of Changing Size.  
1151 *Genetics*, 146(2):723–733, June 1997. ISSN 1943-2631. doi: 10.1093/genetics/146.2.723. URL  
1152 <http://dx.doi.org/10.1093/genetics/146.2.723>.
- 1153 L. M. J. O’Connor, V. Fugère, and A. Gonzalez. Evolutionary rescue is mediated by the  
1154 history of selection and dispersal in diversifying metacommunities. *Frontiers in Ecology and*  
1155 *Evolution*, 8, Dec. 2020. ISSN 2296-701X. doi: 10.3389/fevo.2020.517434. URL <http://dx.doi.org/10.3389/fevo.2020.517434>.
- 1157 M. Z. Peery, R. Kirby, B. N. Reid, R. Stoelting, E. Doucet-Béer, S. Robinson, C. Vasquez-  
1158 Carrillo, J. N. Pauli, and P. J. Palsboll. Reliability of genetic bottleneck tests for detecting  
1159 recent population declines. *Molecular Ecology*, 21(14):3403–3418, May 2012. ISSN 1365-294X.  
1160 doi: 10.1111/j.1365-294x.2012.05635.x. URL <http://dx.doi.org/10.1111/j.1365-294x.2012.05635.x>.
- 1162 E. Pollak. On the survival of a gene in a subdivided population. *Journal of Applied Probability*,  
1163 3(1):142–155, June 1966. ISSN 1475-6072. doi: 10.2307/3212043. URL <http://dx.doi.org/10.2307/3212043>.
- 1165 M. Przeworski, G. Coop, and J. D. Wall. The Signature of Positive Selection on Stand-  
1166 ing Genetic Variation. *Evolution*, 59(11):2312–2323, 2005. ISSN 1558-5646. doi: 10.  
1167 1111/j.0014-3820.2005.tb00941.x. URL <http://dx.doi.org/10.1111/j.0014-3820.2005.tb00941.x>.
- 1169 B. Pujol, S. Blanchet, A. Charmantier, E. Danchin, B. Facon, P. Marrot, F. Roux, I. Scotti,  
1170 C. Teplitsky, C. E. Thomson, and I. Winney. The Missing Response to Selection in the

- 1171 Wild. *Trends in Ecology & Evolution*, 33(5):337–346, May 2018. ISSN 0169-5347. doi:  
1172 10.1016/j.tree.2018.02.007. URL <http://dx.doi.org/10.1016/j.tree.2018.02.007>.
- 1173 D. H. Reed and E. H. Bryant. Fitness, genetic load and purging in experimental populations of  
1174 the housefly. *Conservation Genetics*, 2(1):57–61, Mar. 2001. ISSN 1572-9737. doi: 10.1023/A:  
1175 1011567018530. URL <http://dx.doi.org/10.1023/A:1011567018530>.
- 1176 M. E. Roelke, J. S. Martenson, and S. J. O’Brien. The consequences of demographic reduction  
1177 and genetic depletion in the endangered Florida panther. *Current Biology*, 3(6):340–350,  
1178 June 1993. ISSN 0960-9822. doi: 10.1016/0960-9822(93)90197-V. URL [http://dx.doi.org/10.1016/0960-9822\(93\)90197-V](http://dx.doi.org/10.1016/0960-9822(93)90197-V).
- 1180 D. A. Roff and K. Emerson. Epistasis and Dominance: Evidence for Differential Effects in  
1181 Life-History Versus Morphological Traits. *Evolution*, 60(10):1981–1990, 2006. ISSN 1558-  
1182 5646. doi: 10.1111/j.0014-3820.2006.tb01836.x. URL [http://dx.doi.org/10.1111/j.](http://dx.doi.org/10.1111/j.0014-3820.2006.tb01836.x)  
1183 [0014-3820.2006.tb01836.x](http://dx.doi.org/10.1111/j.0014-3820.2006.tb01836.x).
- 1184 M. Rousselle, P. Simion, M.-K. Tilak, E. Figuet, B. Nabholz, and N. Galtier. Is adaptation  
1185 limited by mutation? A timescale-dependent effect of genetic diversity on the adaptive substi-  
1186 tution rate in animals. *PLOS Genetics*, 16(4):e1008668, Apr. 2020. ISSN 1553-7404. doi: 10.  
1187 1371/journal.pgen.1008668. URL <http://dx.doi.org/10.1371/journal.pgen.1008668>.
- 1188 H. Sachdeva, O. Olusanya, and N. Barton. Genetic load and extinction in peripheral popu-  
1189 lations: The roles of migration, drift and demographic stochasticity. *Philosophical Trans-*  
1190 *actions of the Royal Society B: Biological Sciences*, 377(1846):20210010, Jan. 2022. doi:  
1191 10.1098/rstb.2021.0010. URL <http://dx.doi.org/10.1098/rstb.2021.0010>.
- 1192 P. Samani and G. Bell. The ghosts of selection past reduces the probability of plastic rescue but  
1193 increases the likelihood of evolutionary rescue to novel stressors in experimental populations  
1194 of wild yeast. *Ecology Letters*, 19(3):289–298, Jan. 2016. ISSN 1461-0248. doi: 10.1111/ele.  
1195 12566. URL <http://dx.doi.org/10.1111/ele.12566>.
- 1196 M. F. Schenk, M. P. Zwart, S. Hwang, P. Ruelens, E. Severing, J. Krug, and J. A.  
1197 G. M. de Visser. Population size mediates the contribution of high-rate and large-benefit  
1198 mutations to parallel evolution. *Nature Ecology & Evolution*, 6(4):439–447, Mar. 2022.  
1199 ISSN 2397-334X. doi: 10.1038/s41559-022-01669-3. URL [http://dx.doi.org/10.1038/](http://dx.doi.org/10.1038/s41559-022-01669-3)  
1200 [s41559-022-01669-3](http://dx.doi.org/10.1038/s41559-022-01669-3).
- 1201 D. Sigal, Jennifer N.S. Reid, J. N. Reid, and L. M. Wahl. Effects of Transmission Bottlenecks  
1202 on the Diversity of Influenza A Virus. *Genetics*, 210(3):1075–1088, Sept. 2018. doi: 10.1534/  
1203 genetics.118.301510. URL <http://dx.doi.org/10.1534/genetics.118.301510>.
- 1204 S. Sommer. The importance of immune gene variability (MHC) in evolutionary ecology and  
1205 conservation. *Frontiers in Zoology*, 2(1):16, Oct. 2005. ISSN 1742-9994. doi: 10.1186/  
1206 1742-9994-2-16. URL <http://dx.doi.org/10.1186/1742-9994-2-16>.

- 1207 M. Soulé. *Conservation Biology: The Science of Scarcity and Diversity*. Oxford University  
1208 Press, Incorporated, 1986. ISBN 9780878937950. URL [https://books.google.ch/books?](https://books.google.ch/books?id=Qi1jUut7JL8C)  
1209 [id=Qi1jUut7JL8C](https://books.google.ch/books?id=Qi1jUut7JL8C).
- 1210 M. Soulé. *Viable Populations for Conservation*. Cambridge University Press, 1987. ISBN  
1211 9780521333900. URL <https://books.google.ch/books?id=B9tzQgAACAAJ>.
- 1212 G. S. Stewart, M. R. Morris, A. B. Genis, M. Szűcs, B. A. Melbourne, S. J. Tavener, and R. A.  
1213 Hufbauer. The power of evolutionary rescue is constrained by genetic load. *Evolutionary*  
1214 *Applications*, 10(7):731–741, May 2017. ISSN 1752-4571. doi: 10.1111/eva.12489. URL  
1215 <http://dx.doi.org/10.1111/eva.12489>.
- 1216 J. T. Sutton, S. Nakagawa, B. C. Robertson, and I. G. Jamieson. Disentangling the roles of  
1217 natural selection and genetic drift in shaping variation at MHC immunity genes. *Molecular*  
1218 *Ecology*, 20(21):4408–4420, Nov. 2011. doi: 10.1111/j.1365-294x.2011.05292.x. URL [http:](http://dx.doi.org/10.1111/j.1365-294x.2011.05292.x)  
1219 [//dx.doi.org/10.1111/j.1365-294x.2011.05292.x](http://dx.doi.org/10.1111/j.1365-294x.2011.05292.x).
- 1220 W. R. Swindell and J. L. Bouzat. Modeling the Adaptive Potential of Isolated Populations:  
1221 Experimental Simulations Using *Drosophila*. *Evolution*, 59(10):2159–2169, Oct. 2005. doi: 10.  
1222 1111/j.0014-3820.2005.tb00925.x. URL [http://dx.doi.org/10.1111/j.0014-3820.2005.](http://dx.doi.org/10.1111/j.0014-3820.2005.tb00925.x)  
1223 [tb00925.x](http://dx.doi.org/10.1111/j.0014-3820.2005.tb00925.x).
- 1224 W. R. Swindell and J. L. Bouzat. Inbreeding depression and male survivorship in *Drosophila*:  
1225 Implications for senescence theory. *Genetics*, 172(1):317–327, Jan. 2006. ISSN 0016-6731. doi:  
1226 10.1534/genetics.105.045740. URL <http://dx.doi.org/10.1534/genetics.105.045740>.
- 1227 F. Tajima. Statistical method for testing the neutral mutation hypothesis by dna polymorphism.  
1228 *Genetics*, 123(3):585–595, Nov. 1989. ISSN 1943-2631. doi: 10.1093/genetics/123.3.585. URL  
1229 <http://dx.doi.org/10.1093/genetics/123.3.585>.
- 1230 F. Tajima. The amount of dna polymorphism maintained in a finite population when the  
1231 neutral mutation rate varies among sites. *Genetics*, 143(3):1457–1465, July 1996. ISSN 1943-  
1232 2631. doi: 10.1093/genetics/143.3.1457. URL [http://dx.doi.org/10.1093/genetics/](http://dx.doi.org/10.1093/genetics/143.3.1457)  
1233 [143.3.1457](http://dx.doi.org/10.1093/genetics/143.3.1457).
- 1234 D. Tallmon, G. Luikart, and R. Waples. The alluring simplicity and complex reality of genetic  
1235 rescue. *Trends in Ecology & Evolution*, 19(9):489–496, Sept. 2004. ISSN 0169-5347. doi:  
1236 10.1016/j.tree.2004.07.003. URL <http://dx.doi.org/10.1016/j.tree.2004.07.003>.
- 1237 M. Tomasini and S. Peischl. When does gene flow facilitate evolutionary rescue? *Evolution*,  
1238 74(8):1640–1653, Aug. 2020. ISSN 0014-3820, 1558-5646. doi: 10.1111/evo.14038. URL  
1239 <http://dx.doi.org/10.1111/evo.14038>.
- 1240 M. Tomasini and S. Peischl. The role of spatial structure in multi-deme models of evolutionary  
1241 rescue. *Journal of Evolutionary Biology*, 35(7):986–1001, July 2022. ISSN 1010-061X, 1420-  
1242 9101. doi: 10.1111/jeb.14018. URL <http://dx.doi.org/10.1111/jeb.14018>.

- 1243 H. Uecker, S. P. Otto, and J. Hermisson. Evolutionary rescue in structured populations. *The*  
1244 *American Naturalist*, 183(1):E17–35, Jan. 2014. ISSN 1537-5323. doi: 10.1086/673914. URL  
1245 <http://dx.doi.org/10.1086/673914>.
- 1246 B. van Heerwaarden, Y. Willi, T. N. Kristensen, and A. A. Hoffmann. Population Bottlenecks  
1247 Increase Additive Genetic Variance But Do Not Break a Selection Limit in Rain Forest  
1248 *Drosophila*. *Genetics*, 179(4):2135–2146, Aug. 2008. ISSN 1943-2631. doi: 10.1534/genetics.  
1249 107.082768. URL <http://dx.doi.org/10.1534/genetics.107.082768>.
- 1250 T. Vogwill, R. L. Phillips, D. R. Gifford, and R. C. MacLean. Divergent evolution peaks under  
1251 intermediate population bottlenecks during bacterial experimental evolution. *Proceedings of*  
1252 *the Royal Society B: Biological Sciences*, 283(1835):20160749, July 2016. ISSN 1471-2954.  
1253 doi: 10.1098/rspb.2016.0749. URL <http://dx.doi.org/10.1098/rspb.2016.0749>.
- 1254 I. Vázquez-García, F. Salinas, J. Li, A. Fischer, B. Barré, J. Hallin, A. Bergström, E. Alonso-  
1255 Perez, J. Warringer, V. Mustonen, and G. Liti. Clonal heterogeneity influences the fate  
1256 of new adaptive mutations. *Cell Reports*, 21(3):732–744, Oct. 2017. ISSN 2211-1247. doi:  
1257 10.1016/j.celrep.2017.09.046. URL <http://dx.doi.org/10.1016/j.celrep.2017.09.046>.
- 1258 L. M. Wahl, P. J. Gerrish, and I. Saika-Voivod. Evaluating the impact of population bottlenecks  
1259 in experimental evolution. *Genetics*, 162(2):961–971, Oct. 2002. doi: 10.1093/genetics/162.  
1260 2.961. URL <http://dx.doi.org/10.1093/genetics/162.2.961>.
- 1261 T. Wein and T. Dagan. The effect of population bottleneck size and selective regime on genetic  
1262 diversity and evolvability in bacteria. *Genome Biology and Evolution*, page evz243, Nov. 2019.  
1263 ISSN 1759-6653. doi: 10.1093/gbe/evz243. URL <http://dx.doi.org/10.1093/gbe/evz243>.
- 1264 P. A. Whigham, G. C. Dick, and H. G. Spencer. Genetic drift on networks: Ploidy and the  
1265 time to fixation. *Theoretical Population Biology*, 74(4):283–290, Dec. 2008. ISSN 0040-5809.  
1266 doi: 10.1016/j.tpb.2008.08.004. URL <http://dx.doi.org/10.1016/j.tpb.2008.08.004>.
- 1267 A. M. Whitehouse and E. H. Harley. Post-bottleneck genetic diversity of elephant populations  
1268 in south africa, revealed using microsatellite analysis. *Molecular Ecology*, 10(9):2139–2149,  
1269 Sept. 2001. ISSN 1365-294X. doi: 10.1046/j.0962-1083.2001.01356.x. URL <http://dx.doi.org/10.1046/j.0962-1083.2001.01356.x>.
- 1271 A. R. Whiteley, S. W. Fitzpatrick, W. C. Funk, and D. A. Tallmon. Genetic rescue to the  
1272 rescue. *Trends in Ecology & Evolution*, 30(1):42–49, Jan. 2015. ISSN 0169-5347. doi: 10.  
1273 1016/j.tree.2014.10.009. URL <http://dx.doi.org/10.1016/j.tree.2014.10.009>.
- 1274 M. C. Whitlock. Fixation probability and time in subdivided populations. *Genetics*, 164  
1275 (2):767–779, June 2003. ISSN 1943-2631. doi: 10.1093/genetics/164.2.767. URL <http://dx.doi.org/10.1093/genetics/164.2.767>.

- 1277 Y. Willi, J. Van Buskirk, and A. A. Hoffmann. Limits to the adaptive potential of small  
1278 populations. *Annual Review of Ecology, Evolution, and Systematics*, 37(1):433–458, Dec.  
1279 2006. ISSN 1545-2069. doi: 10.1146/annurev.ecolsys.37.091305.110145. URL [http://dx.  
doi.org/10.1146/annurev.ecolsys.37.091305.110145](http://dx.<br/>1280 doi.org/10.1146/annurev.ecolsys.37.091305.110145).
- 1281 Y. Willi, J. Van Buskirk, B. Schmid, and M. Fischer. Genetic isolation of fragmented pop-  
1282 ulations is exacerbated by drift and selection. *Journal of Evolutionary Biology*, 20(2):  
1283 534–542, 2007. ISSN 1420-9101. doi: 10.1111/j.1420-9101.2006.01263.x. URL [http:  
//dx.doi.org/10.1111/j.1420-9101.2006.01263.x](http:<br/>1284 //dx.doi.org/10.1111/j.1420-9101.2006.01263.x).
- 1285 X. Xie, Y. Yang, Q. Ren, X. Ding, P. Bao, B. Yan, X. Yan, J. Han, P. Yan, and Q. Qiu.  
1286 Accumulation of deleterious mutations in the domestic yak genome. *Animal Genetics*, 49  
1287 (5):384–392, 2018. ISSN 1365-2052. doi: 10.1111/age.12703. URL [http://dx.doi.org/10.  
1111/age.12703](http://dx.doi.org/10.<br/>1288 1111/age.12703).
- 1289 W. Yu, B. Wu, X. Wang, Z. Yao, Y. Li, and Y. Liu. Scale-dependent effects of habitat  
1290 fragmentation on the genetic diversity of *actinidia chinensis* populations in china. *Horticulture  
1291 Research*, 7(1), Oct. 2020. ISSN 2052-7276. doi: 10.1038/s41438-020-00401-1. URL [http:  
//dx.doi.org/10.1038/s41438-020-00401-1](http:<br/>1292 //dx.doi.org/10.1038/s41438-020-00401-1).