

Temporal changes in streamflow regime and host-related factors affect parasitism of freshwater prawn in Southeast Brazil

Alison Carlos Wunderlich¹; Esli Emanoel Domingues Mosna¹;

Marcelo Antonio Amaro Pinheiro^{1,2}

¹ Universidade Estadual Paulista (UNESP), Instituto de Biociências – Campus do Litoral Paulista (IB/CLP) – Departamento de Ciências Biológicas e Ambientais, Laboratório de Biologia da Conservação de Crustáceos e Ambientes Costeiros (LBS) – Grupo de Pesquisa em Biologia de Crustáceos (CRUSTA), São Vicente (SP), Brazil – Email: ACW

(awunderlich@gmail.com); EEDM (e.mosna@unesp.br); MAAP

(marcelo.pinheiro@unesp.br).

² Programa de Pós-Graduação em Biodiversidade de Ambientes Costeiros (PPG-BAC), UNESP IB/CLP, São Vicente (SP) – Programa de Pós-Graduação em Ecologia, Evolução e Biodiversidade (PPG-EcoEvoBio), UNESP Campus de Rio Claro (IB/RC), Rio Claro (SP).

Correspondence

Alison C. Wunderlich, Universidade Estadual Paulista (UNESP), Instituto de Biociências – Campus do Litoral Paulista (IB/CLP) - Grupo de Pesquisa em Biologia de Crustáceos (CRUSTA), São Vicente (SP), Brazil

Email: awunderlich@gmail.com

Keywords: Cymothoid; *Macrobrachium*; stream; Rainfall; *Telotha henselii*.

Abstract

1. Understanding how changes in hydrological regime and intrinsic host factors drive parasite loads and dynamics remains a challenging issue in aquatic parasitology. Many host-parasite systems have shown that host-related factors can influence parasitism in freshwater systems. Temporal changes in the hydrological regime are also one key factor that could alter the streamflow and habitat area and indirectly affect the interactions between ectoparasite and host. However, there are still few studies on the combined effects of both streamflow regime and intrinsic host factors to investigate variations in parasite loads on stream ecosystems.

2. Here, we used a cymothoid-palaemonid prawn model to test whether host-related traits and the temporal changes in streamflow and area caused by the rainfall regime could influence ectoparasite loads in stream habitats. Prawns were collected seasonally over 4 years in a tropical stream in southeast Brazil. We also tested the hypothesis of whether the prawn body size covaries with cymothoid body size, and also whether this change between sex and seasons.

3. We found a strong relationship between the ectoparasite and host body sizes, which confirms Harrison's rule of this host-ectoparasite system. Our best model showed an effect of host body size, host sex, and host molt stage, but not host age, on ectoparasite loads. Prawn females have higher ectoparasite loads than males, and there was a positive association with the ecdysis stage. Our model also predicts that decreased rainfall could increase parasitism in dry seasons by reducing the natural flow regime and habitat area in these tropical stream systems.

4. Our findings show that temporal variability in streamflow is a key environmental factor combined with host-related characteristics to drive the parasite dynamics of a cymothoid-prawn system in stream habitats.

5. Identifying how the interactions between aquatic invertebrates and parasites respond to the variability in hydrological conditions can help to understand how infections and parasitism may affect the benthic invertebrates play in stream systems.

1. INTRODUCTION

Parasites represent a significant component of biodiversity, with key roles in most ecosystems of the world (Poulin, 2007; Hechinger *et al.* 2011; Timi and Poulin, 2020). Although the relevance of parasites in ecosystem functions and conservation has risen in recent years (Poulin, 2021; Gagne *et al.* 2022), this subject has been neglected in studies on crustacean biology and ecology, particularly in the neotropical freshwater environment (Wunderlich *et al.* 2011; Smit *et al.* 2014; Smit *et al.* 2019). For example, most studies on parasitic isopod crustaceans are limited to localities in the Caribbean, Indo-Pacific and Mediterranean (e.g., Smit *et al.* 2019; Welicky *et al.*, 2019), with few in the Neotropical freshwater region (Wunderlich *et al.* 2011; Andrade *et al.* 2020; Ribeiro and Horch, 2023). Furthermore, freshwater ecosystems are among the most diverse parasite species (Poulin, 2016), and face increasing high anthropogenic pressure and hydrological disturbance (Boos *et al.* 2019; Zanghi *et al.* 2023; Rumschlag *et al.* 2023), which calls for more studies to understand how these environmental changes interact with the properties of hosts to affect parasite infestations.

The history of host-parasite interaction involving epicaridean isopods and prawns has been observed in fossils from the early Cretaceous and the late Jurassic (163–110 Mya), indicating a long evolutionary association (Klompaker *et al.* 2021; Lima *et al.* 2023). Since then, the dynamic of host-parasite systems is driven by host-related factors that can influence the likelihood of infection on several scales of observation (Poulin, 2007; Castrillo, 2018; Schmid-Hempel, 2021). For example, host body size often correlates positively with parasite abundance or parasite species richness (Poulin, 2007; Kamiya *et al.* 2014a). Also, the parasite body size can increase with the host body size; a pattern termed “Harrison’s rule” (Harrison, 1915; Maestri *et al.* 2020). Although this rule has been reported in several taxa (Poulin, 2007), we still lack knowledge on whether Harrison's rule holds among seasons and between host sex,

and also for cymothoid isopods (Welicky *et al.* 2019). Other host characteristics such as host age, sex, and molt are also correlated with parasite variations between individuals within populations (Christe *et al.* 2007; Ben-Ami, 2019, Izhar *et al.* 2020), which can be age-dependent (Wunderlich *et al.* 2022) or sex-biased (Poulin and Forbes, 2012). In crustaceans, the molting stage is a key growth factor, while it can expose hosts to parasite infections (Duneau and Ebert, 2012) or reduce the likelihood of parasite penetration (Izhar *et al.* 2020). Furthermore, these host factors can also correlate with changes in the hydrological regime, which might increase susceptibility to parasitic infections and alter the dynamics of the disease (Groner *et al.* 2018; Xu *et al.* 2020).

Temporal changes in the hydrological regime act as a strong drive to sustain and alter the structure of benthic invertebrate communities (Poff and Zimmerman, 2010; Northington and Webster, 2017; Poff *et al.* 2018; Herbst *et al.* 2019; Palmer and Ruhi, 2019), as well as the infectious disease and parasite dynamics in freshwater ecosystems (Altizer *et al.* 2006; Groner *et al.* 2018; Schmid-Hempel, 2021; Tadiri and Ebert, 2023). Variability in temperature and rainfall are important aspects in understanding host-parasite interactions (Marcogliese, 2016; Shearer and Ezenwa, 2020), and can alter host susceptibility and exposure to parasites, and as a consequence, it can increase or decrease parasite loads (Groner *et al.* 2018; Vestbo *et al.* 2019; Shearer and Ezenwa, 2020). In rivers and stream habitats, the rainfall regime plays a central role because it can change the streamflow and reduce the habitat area of the stream many times (Paull *et al.* 2012; Botter *et al.* 2013). As a consequence, this reduction in streamflow might increase host density and the chances of parasite encounters and transmission (Brunner and Ostfeld, 2008; Paull *et al.* 2012; Shearer and Ezenwa, 2020). Furthermore, most studies have concentrated on the effects of increasing temperature rather than altered hydrology (Marcos-López *et al.* 2010; Löhmus and Björklund, 2015; Marcogliese, 2016; Lymbery *et al.* 2020). Therefore, it is fundamental in aquatic parasitology to disentangle the contribution of

these sources of variation, particularly in streamflow changes and habitat area, to better understand how these hydrological changes correlate with host-related factors to affect parasite loads over space and time (Poulin, 2007; Kamiya *et al.* 2014b; Cattadori *et al.* 2019; Poulin, 2021; Tadiri and Ebert, 2023).

To test whether the hydrological changes and host-related factors affect the parasite infestation, in this study we examined the effects of host size, sex, age, molt stage, rainfall, streamflow, habitat area, and their interactions on ectoparasite loads using the palaemonid prawn, *Macrobrachium brasiliense* (Heller, 1862), and a cymothoid isopod, *Telotha henselii* (Von Martens, 1869), as our model.

Cymothoid isopods are a diverse group of parasitic crustaceans with a broad distribution throughout the world (Williams and Boyko 2012). Most cymothoids are parasites of marine, freshwater, and brackish teleosts (Smit *et al.* 2014), but also may parasitise freshwater palaemonid prawns in tropical South American river systems (Wunderlich *et al.* 2011; Andrade *et al.* 2020). Adults of *Telotha henselii* are a well-known haematophagous ectoparasites cymothoid on fish, mostly found on skin and gills, and juvenile males use palaemonid prawns as optional intermediate hosts besides final hosts (Trilles and Öktener 2004; Wunderlich *et al.* 2011; Andrade *et al.* 2020; Anaguano-Yancha and Pilatasig, 2022; Fujita *et al.* 2023). Therefore, the cymothoid-palaemonid system is a tractable model that could facilitate our understanding of how changes in the hydrological regime and host-related factors can influence host-ectoparasite interactions in freshwater ecosystems.

Here, we investigate the cymothoid isopod in palaemonid prawns from freshwater streams in the Brazilian Southern. First, we analyse the effects of host-related factors on parasite infestation. Since host-related traits are crucial to driving parasite dynamics, we expect a potential effect of body size, sex, age, and molt stage on cymothoid isopod infestations in *M. brasiliense* populations inhabiting a stream habitat with hydrological variation during the

seasons and years. We predict that a reduction in streamflow and habitat area in dry seasons could indirectly facilitate the chances of parasite encounters and transmission due to the greater aggregation of hosts during this season. Thus, we also expect that prawns collected in dry seasons (i.e., low rainfall and reduced streamflow) would have higher levels of cymothoid isopod infestation than in wet seasons (i.e., high rainfall and normal streamflow). Finally, we examine the hypothesis of whether the body size of prawns correlates with the body size of cymothoids (i.e., a pattern termed “Harrison’s rule”), and whether this pattern could vary between sex and seasons (dry vs. wet). Since previous studies have shown that host body size positively covaries with parasite body size, we expected to find similar parasite-specific patterns in this study.

2. METHODS

2.1. Study area and sampling design

As part of a long-term study of ecology and parasitism, palaemonid prawns (*Macrobrachium brasiliense*) were collected at three sites in the Águas Claras stream, the Pardo River basin (21 ° 18 30 S, 47 ° 36 16 W), during 1995 to 1998 in southwest Brazil (Figure 1). The Pardo River has a drainage basin of 10.694 km² and a course of approximately 550 km, with the dry season occurring between April and September and the wet season between October and March (Alves *et al.* 2018). We collected prawns over four years from January 1995 to June 1998 monthly and bimonthly, except in 1996 and 1997, which we collected in a month only each year. The years 1996 and 1997 were excluded from the hydrological variation analysis, but we kept to examine the correlation between prawn body size and cymothoid body size. Prawn samples were obtained from three sites at the same time, for 2 hours each by two people, during the day. At each site, the prawns were collected manually by sweeping sieves (50 cm in diameter

and 4 mm mesh size) near the riverbank, in places with marginal aquatic vegetation from the stream. The prawns were kept in plastic boxes with water from the stream sites until their arrival in the laboratory, where the specimens were euthanised by thermal shock (at 3°C, for 30 min), fixed and preserved in 70% ethanol.

2.2. Measures of host-related trait and rainfall data

At the laboratory, the prawn specimens were identified according to Melo (2003) and examined to check the presence of isopod cymothoids externally. Posteriorly, prawns were submitted to biometry based on body measures (CL, carapace length – from the base of the rostral spine to the opposite region of the carapace; and TL, total length – comprising total body size, from the base of the rostral spine to the extremity of telson) with a calliper of 0.05 mm precision, and weighing (W, weight – with a digital balance of 0.01 g, in grams), and also characterized according to the age, sex, and molt stage. In this study we consider age in function of the developmental phase (juvenile and adult), using as a proxy the size at onset of morphological sexual maturity at 9 mm CL for males and 8 mm CL for females (Nogueira *et al.* 2019); sex by checking the presence of the appendix masculina in the endopod of the second pair of pleopods (Nogueira *et al.* 2020); and molt stages (intermolt, premolt, and molting or ecdysis) determined by inspection of the exoskeleton hardness, especially the carapace (Drach and Tchernigovtzeff 1967).

The monthly rainfall, streamflow, and habitat area data were obtained from an archival hydrological data set from the Department of Waters and Electric Energy of the State of São Paulo (DAEE, 2023). We used a rainfall station near the sampling sites to obtain total rainfall (mm), average monthly streamflow (m^3/s), and area (m^2). We used three rainfall variables; total rainfall (mm) for the month in which the sample was collected (R_t), total rainfall for the month before sample collection (R_{t-1}), and total rainfall two months before sample collection (R_{t-2}).

The three different measures were used to test whether current (immediate effect) or prior (delayed effect) rainfall was more strongly associated with ectoparasite loads. Together, changes in streamflow and habitat area are associated with changes in rainfall.

2.3. Parasitological screening

The palaemonid prawns were examined and the ectoparasites found were collected from each animal and measured to obtain the total length (TL) under a stereomicroscope (Zeiss® Stemi® SV-6) attached to a computer image analysis system using Zeiss® KS-100 3.0 software (0.01 mm precision). Posteriorly, the ectoparasites were fixed in 70% ethanol and subsequently identified through specific characteristics (e.g., 2nd pereopod showing five spines in the inner margin of the propodus and the 6th pereopod showing four spines in the propodus and one in merus), according to Lemos de Castro (1985) and Andrade *et al.* (2020). The abundance of ectoparasites was considered as the total number of cymothoid isopods in each host individual, which can be categorised in terms of infestation levels as follows: 0 = no infestation; 1=low infestation; 2-3=high infestation (Horton and Okamura, 2001; Kawanishi *et al.* 2016). The prevalence was calculated as the proportion of prawns infected by cymothoid isopods in relation to the total number of hosts analysed, and the mean intensity was calculated as the total number of cymothoids in a sample of infected host species divided by the number of infected individuals of the host species in the sample (Bush *et al.* 1997).

2.4. Data analysis

We used generalised linear models (GLM) to test for the relationship between prawn body size and cymothoid body size (i.e. a pattern called “Harrison's rule”), and whether this pattern can vary between sex and seasons (dry vs. wet). To estimate the effects of host traits (i.e., total length, sex, age, and molt stage) and hydrological factors (i.e., rainfall, streamflow, and habitat

area), on ectoparasite abundances (i.e., response variable), we fit generalised linear mixed models (GLMMs), using Poisson and negative binomial distributions (Zuur *et al.* 2013). Host traits, hydrological factors and their interaction were the main predictors, while month and year were included as random intercepts. Individual IDs (i.e. prawn ID) nested within each month were used as random factors in all models. We used prawn ID as an observation-level random effect to control overdispersion in our parasite count data (Zuur and Ieno 2016). Before fitting the model, we evaluated the multicollinearity of the predictors using the variance inflation factor (VIFs) with the R performance packages (Lüdecke *et al.* 2021), and excluded those with VIFs >10, which represent high multicollinearity among predictors (Borcard *et al.* 2011). We also used diagnostic plots to test for normal distribution and equal variance of residuals with the R package DHARMA (Hartig, 2022).

We used the Akaike Information Criterion (AIC) and Bayesian Information Criterion (BIC) to select the best competitive model using the R performance packages (Lüdecke *et al.* 2021). After the choice of the best model, we used the semipartial coefficients of determination (partR2) to check the variation to individual predictors of this model (Stoffel *et al.* 2021). The partR2 can decompose the variance of R^2 into components uniquely explained by individual predictors (Stoffel *et al.* 2021). GLM and GLMM analyses were performed in R v. 4.2.1 (R Development Core Team, 2022) with packages lme4 (Bates *et al.* 2015) and glmmTMB (Magnusson *et al.* 2020).

3. RESULTS

We found a general prevalence of 16.4 % of a total of 1,012 prawns collected over two years (Table 1). The body size between cymothoid and prawn was positively correlated regardless of sex and seasons (GLM: estimate \pm se: 0.054 ± 0.007 , $p < 0.0001$, $R^2_{\text{adj}} = 0.709$). However, we

did not find a significant difference of this pattern (i.e. positive correlation) in our model between sex (GLM: estimate \pm se: 0.003 ± 0.011 , Wald = 0.298 $p = 0.766$; Figure 2a) and seasons (GLM: estimate \pm se: 0.013 ± 0.016 , Wald = 0.828, $p = 0.408$; Figure 2b). We also found a significant effect of host traits on the abundances of ectoparasites (Figure 3, Table 2), in relation to the size of the host's body (one to three parasites in prawns size ≤ 35.0 mm TL – Figure 3a); host sex (parasitism in female hosts more abundant than in males – Figure 3c); and the stage of molt of the host (parasite abundance following this hierarchical sequence of stages of molt: intermolt > ecdysis > premolt – Figure 3d). However, no effect of age on parasitism was found (parasite abundance did not differ between juveniles and adults – Figure 3b, Table 2). In 1995, the prevalence of ectoparasites was twice in autumn and winter (i.e. 16.6 and 18.9%, respectively) rather than in spring and summer (i.e. 11.9 and 9.2%, respectively; Table 1). This pattern was also observed in 1998, with a higher prevalence in autumn (i.e. 34.1%) and a lower prevalence in summer (i.e. 14.2%; Table 1). The abundance of ectoparasites varied between seasons, with the highest isopod infestation in prawns (i.e. two to three parasites per infested prawn) in the autumn seasons in both years (Figure 4a, Table 1), but there are no differences between the two years in our model (Wald = -1.849; $p = 0.064$; Table 2, Figure 4b). Of the five models that we compared, the one that included three hydrological factors such as concurrent rainfall (R_t), streamflow, and habitat area received the most support (Table 2). Of all fixed variables included in the GLMM models, season showed high multicollinearity with $VIF > 10$ (Table 3). We found that the highest isopod infestations (i.e. two to three parasites per infested prawn) were observed in dry rather than wet seasons (Figure 5). The relative importance of the predictors used in the GLMM was decomposed to check the variation with the individual predictors of the best model (Figure 6). Of three hydrological factors, streamflow seems to be the best predictor of ectoparasite infestation, presenting the largest inclusive R^2

(Figure 6A and B), structure coefficient (Figure 6C) and negative beta weight (Figure 6D). Host length, sex (female) and ecdysis contributed positively to the predictors (Figure 6C-D).

4. DISCUSSION

Our results reveal a combined effect of hydrological factors and host-related characteristics as a key driver of seasonal variation in parasite loads and dynamics of a cymothoid-palaemonid prawn system. We also found strong evidence of the Harrison rule (HR) in the ectoparasite host system evaluated in this study. Our results demonstrated the importance of size in this parasite-host system, which states that both body sizes (parasite and host) tend to be positively and significantly correlated (Harrison, 1915; Clayton *et al.* 2016; Maestri *et al.* 2020). Although the pattern has often been corroborated in several parasite taxa (Nagler *et al.* 2017; Maestri *et al.* 2020), for cymothoid isopods, there is still little example to support this rule (see Welicky *et al.* 2019). Our results also corroborated the data registered by Welicky *et al.* (2019) and Kottarathil *et al.* (2019) with other cymothoid isopod species, where a positive and significant correlation was confirmed between the sizes of these parasites and the size of the fish. However, depending on the host-parasite system, the adaptive significance of this match size may vary (Poulin, 2007; Clayton *et al.* 2016; Lim *et al.* 2022; Lisnerová *et al.* 2022). Lim *et al.* (2022), also confirm Harrison's rule in Nomadinae (cleptoparasitic insects), supporting that body size dynamics influence host shifts of cleptoparasitic bees. Although this pattern between the size of the ectoparasites and their hosts has been well supported in many systems, recently an interesting work demonstrated that some environmental factors (e.g., temperature and precipitation) also affect this pattern, leading to a deceptive correlation between the sizes of the fleas and their hosts across assemblages (Maestri *et al.* 2020). Although we have found a

positive correlation between the sizes of the cymothoids and their prawns, we did not observe a significant difference of this pattern between seasons and sex in our study.

Heterogeneity in host-related factors (e.g., size, sex, age, and molt stage) can strongly influence parasite transmission, susceptibility, tolerance, and resistance to infection (Johnson and Hoverman, 2014; Izhar *et al.* 2020; Clark *et al.* 2021; Schmid-Hempel, 2021). We found that host-related traits that involved body length, sex, and molt stage (but not host age), influenced the abundance and dynamic of ectoparasites. The host body size is a critical characteristic that influences several host-parasite systems (Poulin, 2007; Schmid-Hempel, 2021). For example, parasite abundance (or loads) often correlates positively with host body size (Kamiya *et al.* 2014b; Schmid-Hempel, 2021). The theory suggests that large-bodied hosts should have more parasites (Poulin, 2007; Walker *et al.* 2017). Furthermore, body size correlates with other host traits that affect the likelihood of acquiring infections (Cooper *et al.* 2012; Blasco-Costa and Poulin 2013; Schmid-Hempel, 2021), such as longevity.

The host sex has also been an important factor in explaining sex-biased parasitism (Poulin and Forbes, 2012). In many parasite-host systems, males are more parasitised than females (Cozzarolo *et al.* 2019), an expressive contrast to our study, where female prawns have a higher abundance of ectoparasites than males. In general, the prevalence or intensity of parasites in arthropods is also higher in males than females (Klein *et al.* 2004), but in some studies it was not observed (e.g. Sheridan *et al.*, 2000). One potential explanation of a female bias might be caused by differences in immunity and exposure to parasites between the sex (Poulin, 2007, Cozzarolo *et al.* 2019; Schmid-Hempel, 2021). Males and females are likely to exhibit differences in behaviour and spatial aggregation in the host population (Nogueira *et al.* 2019; Bardera *et al.* 2020), which can influence parasite exposure and susceptibility and favours sex-biased parasitisms (Poulin, 2007; Christe *et al.* 2007). For example, ovigerous

females are more cryptic than males (Nogueira *et al.* 2019), which could favour higher exposure to parasites (i.e. to manca exposure) during the dry seasons in our study.

Although we did not find an effect of host age on ectoparasite abundance, age has been correlated with parasite variation among individuals within invertebrates (Ben-Ami, 2019, Izhar *et al.* 2020) and vertebrates host populations (Wunderlich *et al.* 2022). For example, Izhar *et al.* (2020) found that older adults have better immune responses than younger adults and juveniles of *Daphnia magna*. However, a decrease in susceptibility in older fish has also been suggested to affect host infestation outcomes (Tinsley *et al.* 2020; Bailey *et al.* 2021). For example, the severity of the disease was greater in young than in older adults due to an ontogenetic decline in susceptibility in older individuals (Bailey *et al.* 2021).

Another key factor that can interfere with infestation levels is the molting process (Groner *et al.* 2018; Izhar *et al.* 2020). Our results suggest that parasite abundance was positively associated with the ecdysis stage. In crustaceans, the molting stage is a key growth factor, while it can expose hosts to parasite infections (Duneau and Ebert, 2012) or reduce the likelihood of parasite infestations (Izhar *et al.* 2020). Furthermore, these host factors can also vary with environmental changes, which might increase susceptibility to parasitic infections and alter the disease dynamic (Groner *et al.* 2018; Xu *et al.* 2020). For example, in the American lobster (*Homarus americanus*), increasing seawater temperatures can lead to less molting during summer, when lobsters are more susceptible to peaks of epizootic shell disease (Groner *et al.* 2018).

In addition to these host-related factors, hydrological factors can also exert a strong effect on parasitism in freshwater ecosystems (Altizer *et al.* 2006; Paull *et al.* 2012). Freshwater organisms are exposed to variable rainfall regimes and streamflow, potentially altering their behaviour and subsequently parasite loads among hosts (Paull *et al.* 2012; Reynolds *et al.* 2019). Our results showed that the highest prevalences and abundances of ectoparasites were

found in dry rather than wet seasons. Rainfall has been suggested to be a key driver of seasonality in parasitism (Aneesh *et al.* 2013; Gagne and Blum, 2016; Jemi *et al.* 2020; Shearer and Ezenwa, 2020). Some authors registered higher parasite loads associated with low rainfall, suggesting a gradual increase in host susceptibility during the dry season (Gagne and Blum, 2016; Shearer and Ezenwa, 2020). Cymothoids parasitizing fish also were more abundant during the autumn season (i.e., dry season to India), suggesting that this season can facilitate the ectoparasite infestation (Aneesh *et al.* 2013; Jemi *et al.* 2020). Another interesting study showed that drought in stream fish communities can intensify parasitic copepod infestations (Medeiros and Maltchik, 1999). Furthermore, recent work suggests that the reproductive peaks of cymothoids in the free-swimming stage (called ‘mancae’) occurred in autumn (Fujita *et al.* 2023), which could support our study where a more intense infestation of the prawns by juvenile cymothoid isopods during dry seasons. Another important aspect is the effect of low rainfall that can reduce river flow and habitat area during the driest seasons and promote an increase in parasite transmission and encounter with prawns, probably the same as occurs with ectoparasite loads (Paull *et al.* 2012; Shearer and Ezenwa, 2020). This reduction in streamflow and habitat area can affect host behaviour and aggregation into stream habitats, increasing the chances of acquiring the parasite through host contact (Paull *et al.* 2012; Reynolds *et al.* 2019; Schmid-Hempel, 2021). Studies have shown that a high density of hosts can accelerate contact between those most susceptible, as well as intensify the risk of infection for host individuals (Lafferty *et al.* 2015; Buck *et al.* 2017). The same can occur with parasites because they are also more clumped together across the environment, which results in hosts experiencing a higher infection level (Brunner and Ostfeld, 2008; Paull *et al.* 2012). For example, in an interesting experiment with fish, the parasite transmission rate was higher in shoals exposed to interrupted flow (Reynolds *et al.* 2019). According to these authors, a significantly higher ectoparasite gyrodactylid transmission rate was observed in interrupted flow, resulting in a

higher parasite prevalence when compared to a system with continuous water flux. Furthermore, some authors have observed that slow water flow in downstream may be able to accumulate more free-living infective stages, which can influence the probability of host and parasite encounter, resulting in high parasitism in this part of the stream (Blasco-Costa *et al.* 2013; Resetais and Byers, 2023). However, during the wet seasons, excessive rainfall can drag away and reduce the abundance of the free-living infective stages of the parasites due to higher water volume, a fact verified in several helminths and crustacean parasitic (especially in 'mancae' of cymothoids) (Stromberg 1997; Altizer *et al.* 2006). In addition, low levels of resource availability and environmental stress caused in dry seasons can lead to poor host body conditions and immune function, with an increase in parasite prevalence and intensity (Marshall *et al.* 2008; Shearer and Ezenwa, 2020). Furthermore, a study that integrates differences between types of stream habitat and resource supplementation regimes on parasite dynamics could yield new broadly applicable strategies to help parasitism management and conservation in freshwater ecosystems (Civitello *et al.* 2018). Although our study was not designed to address host body condition differences among the seasons, we suggest the need for additional experiments to test the hypothesis that host body conditions should change among the seasons, modifying thus the ectoparasite-host dynamics.

Overall, our study supports that changes in hydrological factors combined with host-related factors can influence host-parasite dynamics in stream ecosystems. More studies are needed to understand how hydrological disturbances as a result of climate change that have occurred in the last decades can impact the natural flow regime and the benthic invertebrate communities in the stream ecosystems. Aspects such as those are very relevant for the conservation of stream biota and also promote a better understanding of their parasite biodiversity.

AUTHOR CONTRIBUTIONS

Conceptualisation, data interpretation, writing: AW, MP. Conducting the research, data analysis, preparation of figures and tables: AW, EM.

ACKNOWLEDGEMENTS

We thank members of the CRUSTA's research group (Gustavo Y. Hattori, Lucimari M. Seto, and Fabiano G. Taddei), during field expeditions to Águas Claras stream (Serra Azul municipality, SP), and help with their biometry. MAAP thanks to 'Fundação de Amparo à Pesquisa do Estado de São Paulo' – FAPESP, which indirectly funded this work during the execution of the *Arenaeus* Project (# 1995/09495-2) and to the 'Conselho Nacional de Desenvolvimento Científico e Tecnológico' – CNPq, due to the research fellowship grant (# 303286/2016-4 and # 305957/2019-8). ACW also thanks São Paulo State University (UNESP/PROPe) for a postdoctoral research fellowship (Edital 13/2022 - Programa Unesp de Pós-Doutorado- Chamada Pública para Seleção de Bolsistas - Apoio a Grupos de Pesquisa) and indirectly to FAPESP (# 2017/16650–5) due to previous financial support to statistical courses (e.g., Mixed effects modelling) that makes possible the data analyses in this study.

CONFLICT OF INTEREST STATEMENT

The authors declare that they have no conflicts of interest.

DATA AVAILABILITY STATEMENT

Data supporting the findings of this study are available upon request from the corresponding author.

ORCID

Alison Carlos Wunderlich (<https://orcid.org/0000-0001-9222-8536>)

Esli Emanuel Domingues Mosna (<https://orcid.org/0000-0002-6330-1586>)

Marcelo Antonio Amaro Pinheiro (<https://orcid.org/0000-0003-0758-5526>)

REFERENCES

- Altizer, S., Dobson, A., Hosseini, P., Hudson, P., Pascual, M., & Rohani, P. (2006). Seasonality and the dynamics of infectious diseases. *Ecol Lett*, *9*(4), 467-484. DOI:10.1111/j.1461-0248.2005.00879.x
- Alves, R. I. S., Machado, C. S., Beda, C. F., Fregonesi, B. M., Nadal, M., Sierra, J., . . . Segura-Munoz, S. I. (2018). Water Quality Assessment of the Pardo River Basin, Brazil: A Multivariate Approach Using Limnological Parameters, Metal Concentrations and Indicator Bacteria. *Arch Environ Contam Toxicol*, *75*(2), 14. DOI:10.1007/s00244-017-0493-7
- Anaguano Yancho, F. & Pilatasig Chusin, A. L. (2022). Nuevos registros y hospederos del isópodo *Telotha henselii* (Isopoda: Cymothoidae) en la Amazonia ecuatoriana. *Biota Colombiana*, *23*(1). DOI:10.21068/2539200x.920
- Andrade, K. S. F., Silva, L. V. C. & Pralon, B. G. N. (2020). First record of *Telotha henselii* (Isopoda: Cymothoidae) on *Macrobrachium jelskii* (Decapoda: Palaemonidae) from Brazil. *Pan-American Journal of Aquatic Sciences*, *2*, 6.
- Aneesh, P. T., Sudha, K., Arshad, K., Anilkumar, G., & Trilles, J. P. (2013). Seasonal fluctuation of the prevalence of cymothoids representing the genus *Nerocila* (Crustacea, Isopoda), parasitizing commercially exploited marine fishes from the Malabar Coast, India. *Acta Parasitol*, *58*(1), 80-90. DOI:10.2478/s11686-013-0112-3
- Bailey, C., Strepparava, N., Ros, A., Wahli, T., Schmidt-Posthaus, H., Segner, H., & Tafalla, C. (2021). It's a hard knock life for some: Heterogeneity in infection life history of salmonids influences parasite disease outcomes. *J Anim Ecol*, *90*(11), 2573-2593. DOI:10.1111/1365-2656.13562
- Bardera, G., Owen, M. A. G., Façanha, F. N., Sloman, K. A., & Alexander, M. E. (2020). The influence of sex on feeding behaviour in Pacific white shrimp (*Litopenaeus vannamei*). *Applied Animal Behaviour Science*, *224*. DOI:10.1016/j.applanim.2020.104946
- Bates, D., Mächler, M., Bolker, B., & Walker, S. (2015). Fitting Linear Mixed-Effects Models Using lme4. *Journal of Statistical Software*, *67*(1), 1 - 48. DOI:10.18637/jss.v067.i01
- Ben-Ami, F. (2019). Host Age Effects in Invertebrates: Epidemiological, Ecological, and Evolutionary Implications. *Trends Parasitol*, *35*(6), 466-480. DOI:10.1016/j.pt.2019.03.008
- Blasco-Costa, I., Koehler, A. V., Martin, A., & Poulin, R. (2013). Upstream-downstream gradient in infection levels by fish parasites: a common river pattern? *Parasitology*, *140*(2), 266-274. DOI:10.1017/S0031182012001527
- Bojko, J., Lipp, E. K., Ford, A. T., & Behringer, D. C. (2020). Pollution can drive marine diseases. In *Marine Disease Ecology* (pp. 0): Oxford University Press. Retrieved from <https://doi.org/10.1093/oso/9780198821632.003.0006>. doi:10.1093/oso/9780198821632.003.0006
- Boos, H., Salge, P. G., & Pinheiro, M. A. A. (2019). Conservation status and threats of Aegliidae: beyond the assessment. In S. B. Santos, S.L.S. (Ed.), *Aegliidae: life history and conservation status of unique freshwater anomuran decapods* (1 ed., pp. 233-255). New York: CRC—Taylor & Francis Group.
- Borcard, D., Gillet, F., & Legendre, P. (2011). *Numerical ecology with R*: Springer.

- Botter, G., Basso, S., Rodriguez-Iturbe, I., & Rinaldo, A. (2013). Resilience of river flow regimes. *Proc Natl Acad Sci U S A*, *110*(32), 12925-12930. DOI:10.1073/pnas.1311920110
- Brunner, J. L. & Ostfeld, R. S. (2008). Multiple causes of variable tick burdens on small-mammal hosts. *Ecology*, *89*(8), 2259-2272. DOI:<https://doi.org/10.1890/07-0665.1>
- Buck, J. C., Hechinger, R. F., Wood, A. C., Stewart, T. E., Kuris, A. M., & Lafferty, K. D. (2017). Host density increases parasite recruitment but decreases host risk in a snail–trematode system. *Ecology*, *98*(8), 2029-2038. DOI:<https://doi.org/10.1002/ecy.1905>
- Bush, A. O., Lafferty, K. D., Lotz, J. M., & Shostak, A. W. (1997). Parasitology Meets Ecology on Its Own Terms: Margolis et al. Revisited. *The Journal of Parasitology*, *83*(4), 575-583. DOI:10.2307/3284227
- Cattadori, I. M., Pathak, A. K., & Ferrari, M. J. (2019). External disturbances impact helminth–host interactions by affecting dynamics of infection, parasite traits, and host immune responses. *Ecology and Evolution*, *9*(23), 13495-13505. DOI:<https://doi.org/10.1002/ece3.5805>
- Christe, P., Glaizot, O., Evanno, G., Bruyndonckx, N., Devevey, G., Yannic, G., . . . Arlettaz, R. (2007). Host sex and ectoparasites choice: preference for, and higher survival on female hosts. *J Anim Ecol*, *76*(4), 703-710. DOI:10.1111/j.1365-2656.2007.01255.x
- Civitello, D. J., Allman, B. E., Morozumi, C., & Rohr, J. R. (2018). Assessing the direct and indirect effects of food provisioning and nutrient enrichment on wildlife infectious disease dynamics. *Philos Trans R Soc Lond B Biol Sci*, *373*(1745). DOI:10.1098/rstb.2017.0101
- Clark, J., McNally, L., & Little, T. J. (2021). Pathogen Dynamics across the Diversity of Aging. *Am Nat*, *197*(2), 203-215. DOI:10.1086/712380
- Clayton, D. H., Bush, S. E., & Johnson, K. P. (2016). *Coevolution of life on hosts: integrating ecology and history*. Chicago: The University of Chicago Press.
- Cozzarolo, C. S., Sironi, N., Glaizot, O., Pigeault, R., & Christe, P. (2019). Sex-biased parasitism in vector-borne disease: Vector preference? *PLoS One*, *14*(5), e0216360. DOI:10.1371/journal.pone.0216360
- DAEE. (2023). Hydrological Database. Retrieved 01/05/2023, from Department of Water and Electricity of the State of São Paulo, Brazil <http://www.hidrologia.dae.sp.gov.br/>
- Drach, P. & Tchernigovtzeff, C. (1967). Sur la methode de determination des stades d'intermue et son application general aux Crustaces. *Vie et Milieu*, *18*, 16.
- Duneau, D. & Ebert, D. (2012). The role of moulting in parasite defence. *Proc Biol Sci*, *279*(1740), 3049-3054. DOI:10.1098/rspb.2012.0407
- Fujita, H., Kawai, K., Deville, D., & Umino, T. (2023). Quatrefoil light traps for free-swimming stages of cymothoid parasitic isopods and seasonal variation in their species compositions in the Seto Inland Sea, Japan. *Int J Parasitol Parasites Wildl*, *20*, 12-19. DOI:10.1016/j.ijppaw.2022.12.002
- Gagne, R. B. & Blum, M. J. (2016). Parasitism of a native Hawaiian stream fish by an introduced nematode increases with declining precipitation across a natural rainfall gradient. *ECOLOGY OF FRESHWATER FISH*, *25*(3), 476-486.
- Groner, M. L., Shields, J. D., Landers, D. F., Jr., Swenarton, J., & Hoenig, J. M. (2018). Rising Temperatures, Molting Phenology, and Epizootic Shell Disease in the American Lobster. *Am Nat*, *192*(5), E163-E177. DOI:10.1086/699478
- Harrison, L. (1915). Mallophaga from Apteryx, and their Significance; with a Note on the Genus Rallicola. *Parasitology*, *8*(1), 88-100. DOI:10.1017/S0031182000010428
- Hartig, F. (2019). DHARMA: Residual Diagnostics for Hierarchical (Multi-Level / Mixed) Regression Models. *R package version 0.3.2.0*, 54.
- Hechinger, R. F., Lafferty, K. D., Dobson, A. P., Brown, J. H., & Kuris, A. M. (2011). A common scaling rule for abundance, energetics, and production of parasitic and free-living species. *Science*, *333*(6041), 445-448. DOI:10.1126/science.1204337
- Herbst, D. B., Cooper, S. D., Medhurst, R. B., Wiseman, S. W., & Hunsaker, C. T. (2019). Drought ecohydrology alters the structure and function of benthic invertebrate communities in mountain streams. *Freshwater Biology*, *64*(5), 886-902. DOI:10.1111/fwb.13270

- Horton, T. & Okamura, B. (2001). Cymothoid isopod parasites in aquaculture. *DISEASES OF AQUATIC ORGANISMS*, 46, 8.
- Izhar, R., Gilboa, C., Ben-Ami, F., & Priest, N. (2020). Disentangling the steps of the infection process responsible for juvenile disease susceptibility. *Functional Ecology*. DOI:10.1111/1365-2435.13580
- Jemi, J. N., Hatha, A. A. M., & Radhakrishnan, C. K. (2020). Seasonal variation of the prevalence of cymothoid isopod *Norileca indica* (Crustacea, Isopoda), parasitizing on the host fish *Rastrelliger kanagurta* collected from the Southwest coast of India. *Journal of Parasitic Diseases*, 44(0971-7196 (Print)), 5. DOI:10.1007/s12639-020-01208-6
- Johnson, P. T. & Hoverman, J. T. (2014). Heterogeneous hosts: how variation in host size, behaviour and immunity affects parasite aggregation. *J Anim Ecol*, 83(5), 1103-1112. DOI:10.1111/1365-2656.12215
- Kamiya, T., O'Dwyer, K., Nakagawa, S., & Poulin, R. (2014). What determines species richness of parasitic organisms? A meta-analysis across animal, plant and fungal hosts. *Biol Rev Camb Philos Soc*, 89(1), 123-134. DOI:10.1111/brv.12046
- Kamiya, T., O'Dwyer, K., Nakagawa, S., & Poulin, R. (2014). Host diversity drives parasite diversity: meta-analytical insights into patterns and causal mechanisms. *Ecography*, 37(7), 689-697. DOI:10.1111/j.1600-0587.2013.00571.x
- Kawanishi, R., Sogabe, A., Nishimoto, R., & Hata, H. (2016). Spatial variation in the parasitic isopod load of the Japanese halfbeak in western Japan. *Dis Aquat Organ*, 122(1), 13-19. DOI:10.3354/dao03064
- Klein, S. L. (2004). Hormonal and immunological mechanisms mediating sex differences in parasite infection. *Parasite Immunology*, 26, 18. DOI: <https://doi.org/10.1111/j.0141-9838.2004.00710.x>
- Klomp maker, A. A., Robins, C. M., Portell, R. W., & De Angeli, A. (2021). Crustaceans as Hosts of Parasites Throughout the Phanerozoic. In K. De Baets & J. W. Huntley (Eds.), *The Evolution and Fossil Record of Parasitism: Coevolution and Paleoparasitological Techniques* (pp. 121-172). Cham: Springer International Publishing.
- Kottarathil, H. A., Sahadevan, A. V., Kattamballi, R., & Kappalli, S. (2019). *Norileca indica* (Crustacea: Isopoda, Cymothoidae) Infects *Rastrelliger kanagurta* Along the Malabar Coast of India - Seasonal Variation in the Prevalence and Aspects of Host-parasite Interactions. *Zool Stud*, 58, e35. DOI:10.6620/ZS.2019.58-35
- Lafferty, K. D. D., G.; Briggs, C.J.; Dobson, A.P.; Gross, T.; Kuris, A.M. (2015). A general consumer-resource population model. *Science*, 349(6250), 854-857. DOI:10.1126/science.aaa6224
- Lemos de Castro, A. (1985). Ectoparasitism of *Telotha henselii* (Von Martens) (Isopoda, Cymothoidae) on *Macrobrachium brasiliense* (Heller) (Decapoda, Palaemonidae). *Crustaceana*, 49(2), 2.
- Lim, K., Lee, S., Orr, M., & Lee, S. (2022). Harrison's rule corroborated for the body size of cleptoparasitic cuckoo bees (Hymenoptera: Apidae: Nomadinae) and their hosts. *Sci Rep*, 12(1), 10984. DOI:10.1038/s41598-022-14938-9
- Lima, D., Alencar, D. R., Santana, W., Oliveira, N. C., Saraiva, A. Á. F., Oliveira, G. R., . . . Pinheiro, A. P. (2023). 110-million-years-old fossil suggests early parasitism in shrimps. *Scientific Reports*, 13(1), 14549. DOI:10.1038/s41598-023-40554-2
- Lisnerova, M., Lisner, A., Cantatore, D. M. P., Schaeffner, B. C., Peckova, H., Tymi, T., . . . Holzer, A. S. (2022). Correlated evolution of fish host length and parasite spore size: a tale from myxosporeans inhabiting elasmobranchs. *Int J Parasitol*, 52(2-3), 97-110. DOI:10.1016/j.ijpara.2021.05.008
- Löhmus, M. & Björklund, M. (2015). Climate change: what will it do to fish—parasite interactions? *Biological Journal of the Linnean Society*, 116(2), 397-411. DOI:10.1111/bij.12584
- Lüdecke, D., Ben-Shachar, M., Patil, I., Waggoner, P., & Makowski, D. (2021). performance: An R Package for Assessment, Comparison and Testing of Statistical Models. *Journal of Open Source Software*, 6(60). DOI:10.21105/joss.03139

- Lymbery, A. J., Lymbery, S. J., & Beatty, S. J. (2020). Fish out of water: Aquatic parasites in a drying world. *Int J Parasitol Parasites Wildl*, 12, 300-307. DOI:10.1016/j.ijppaw.2020.05.003
- Maestri, R., Fiedler, M. S., Shenbrot, G. I., Surkova, E. N., Medvedev, S. G., Khokhlova, I. S., & Krasnov, B. R. (2020). Harrison's rule scales up to entire parasite assemblages but is determined by environmental factors. *J Anim Ecol*, 89(12), 2888-2895. DOI:10.1111/1365-2656.13344
- Magnusson, A., Skaug, H., Nielsen, A., Berg, C., Kristensen, K., Maechler, M., van Benthem, K., Bolker, B., & Brooks, M. (2020). glmmTMB: Generalized Linear Mixed Models using Template Model Builder. *R package version 1.0.2.1*, 37.
- Marcogliese, D. J. (2016). The Distribution and Abundance of Parasites in Aquatic Ecosystems in a Changing Climate: More than Just Temperature. *INTEGRATIVE AND COMPARATIVE BIOLOGY*, 56(4), 611-619. DOI:10.1093/icb/icw036
- Marcos-Lopez, M., Gale, P., Oidtmann, B. C., & Peeler, E. J. (2010). Assessing the impact of climate change on disease emergence in freshwater fish in the United Kingdom. *Transbound Emerg Dis*, 57(5), 293-304. DOI:10.1111/j.1865-1682.2010.01150.x
- Medeiros, E. S. F. & Maltchik, L. (1999). The effects of hydrological disturbance on the intensity of infestation of *Lernaea cyprinacea* in an intermittent stream fish community. *Journal of Arid Environments*, 43, 6. DOI:<https://doi.org/10.1006/jare.1999.0545>
- Melo, G. A. S. d., Magalhães, C., Bond-Buckup, G., & Buckup, L. (2003). *Manual de identificação dos crustacea decapoda de água doce do Brasil*: Museu de Zoologia USP.
- Nogueira, C. d. S., Oliveira, M. S. d., Jacobucci, G. B., & Almeida, A. C. d. (2019). Relative growth of freshwater prawn *Macrobrachium brasiliense* (Decapoda, Palaemonidae) and its implications for reproduction. *Iheringia. Série Zoologia*, 109. DOI:10.1590/1678-4766e2019005
- Nogueira, C. S., Pantaleão, J. A. F., Almeida, A. C., & Costa, R. C. (2020). Male morphotypes of the freshwater prawn *Macrobrachium brasiliense* (Decapoda: Caridea: Palaemonidae). *Invertebrate Biology*, 139(1). DOI:10.1111/ivb.12279
- Northington, R. M. & Webster, J. R. (2017). Experimental reductions in stream flow alter litter processing and consumer subsidies in headwater streams. *Freshwater Biology*, 62(4), 737-750. DOI:10.1111/fwb.12898
- Palmer, M. & Ruhi, A. (2019). Linkages between flow regime, biota, and ecosystem processes: Implications for river restoration. *Science*, 365(6459). DOI:10.1126/science.aaw2087
- Paull, S. H., Song, S., McClure, K. M., Sackett, L. C., Kilpatrick, A. M., & Johnson, P. T. (2012). From superspreaders to disease hotspots: linking transmission across hosts and space. *Front Ecol Environ*, 10(2), 75-82. DOI:10.1890/110111
- Poff, N. L. (2018). Beyond the natural flow regime? Broadening the hydro-ecological foundation to meet environmental flows challenges in a non-stationary world. *Freshwater Biology*, 63(8), 1011-1021. DOI:10.1111/fwb.13038
- Poff, N. L. & Zimmerman, J. K. H. (2010). Ecological responses to altered flow regimes: a literature review to inform the science and management of environmental flows. *Freshwater Biology*, 55(1), 194-205. DOI:<https://doi.org/10.1111/j.1365-2427.2009.02272.x>
- Poulin, R. (2007). *Evolutionary Ecology of Parasites* (2nd ed.). Princeton: Princeton University Press.
- Poulin, R. (2016). Greater diversification of freshwater than marine parasites of fish. *International Journal for Parasitology*, 46(4), 275-279. DOI:<https://doi.org/10.1016/j.ijpara.2015.12.002>
- Poulin, R. (2021). The rise of ecological parasitology: twelve landmark advances that changed its history. *Int J Parasitol*, 51(13-14), 1073-1084. DOI:10.1016/j.ijpara.2021.07.001
- Poulin, R. & Forbes, M. R. (2012). Meta-analysis and research on host-parasite interactions: past and future. *Evolutionary Ecology*, 26(5), 1169-1185. DOI:10.1007/s10682-011-9544-0
- R Core Team. (2022). R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. Retrieved from <http://www.r-project.org/>
- Resetarits, E. J. & Byers, J. E. (2023). The role of small-scale environmental gradients on trematode infection. *Freshwater Biology*, 68(8), 1453-1461. DOI:10.1111/fwb.14140

- Reynolds, M., Hockley, F. A., Wilson, C. A. M. E., & Cable, J. (2019). Assessing the effects of water flow rate on parasite transmission amongst a social host. *Hydrobiologia*, 830(1), 201-212. DOI:10.1007/s10750-018-3863-x
- Ribeiro, F. B. & Horch, A. P. (2023). Checklist of parasitic isopods from Brazil: Bopyroidea and Cryptoniscoidea (Isopoda: Cymothoidea: Epicaridea). *Zootaxa*, 5325(2), 151-185. DOI:10.11646/zootaxa.5325.2.1
- Rumschlag, S. L. M., M.B.; Jones, D.K.; Battaglin, W.; Behrens, J.; Bernhardt, E.S.; Bradley, P.; Brown, E.; De Laender, F.; Hill, R.; Kunz, S.; Lee, S.; Rosi, E.; Schäfer, R.; Schmidt, T.S.; Simonin, M.; Smalling, K.; Voss, K.; Rohr, J.R. (2023). Density declines, richness increases, and composition shifts in stream macroinvertebrates. *Sci. Adv.*, 9, 9.
- Schmid-Hempel, P. (2021). *Evolutionary Parasitology: The Integrated Study of Infections, Immunology, Ecology, and Genetics*: Oxford University Press.
- Shearer, C. L. & Ezenwa, V. O. (2020). Rainfall as a driver of seasonality in parasitism. *Int J Parasitol Parasites Wildl*, 12, 8-12. DOI:10.1016/j.ijppaw.2020.04.004
- Smit, N. J., Bruce, N. L., & Hadfield, K. A. (2014). Global diversity of fish parasitic isopod crustaceans of the family Cymothoidae. *Int J Parasitol Parasites Wildl*, 3(2), 188-197. DOI:10.1016/j.ijppaw.2014.03.004
- Smit, N. J., Bruce, N. L., & Hadfield, K. A. (2019). Introduction to Parasitic Crustacea: State of Knowledge and Future Trends. In N. J. Smit, N. L. Bruce, & K. A. Hadfield (Eds.), *Parasitic Crustacea: State of Knowledge and Future Trends* (pp. 1-6). Cham: Springer International Publishing.
- Stoffel, M. A., Nakagawa, S., & Schielzeth, H. (2021). partR2: partitioning R(2) in generalized linear mixed models. *PeerJ*, 9, e11414. DOI:10.7717/peerj.11414
- Taberner, R., Volonterio, O., & De León, R. P. (2003). Description Of The Pulli Stages Of *Telotha Henselii* (von Martens, 1869) (isopoda, Cymothoidae), With New Hosts And Locality Records From Uruguay And Argentina. *Crustaceana*, 76(1), 27-37. DOI:<https://doi.org/10.1163/156854003321672809>
- Tadiri, C. P. & Ebert, D. (2023). The role of temperature in the start of seasonal infectious disease epidemics. *Oikos*, e10014. DOI:10.1111/oik.10014
- Timi, J. T. & Poulin, R. (2020). Why ignoring parasites in fish ecology is a mistake. *Int J Parasitol*, 50(10-11), 755-761. DOI:10.1016/j.ijpara.2020.04.007
- Tinsley, R. C., Rose Vineer, H., Grainger-Wood, R., & Morgan, E. R. (2020). Heterogeneity in helminth infections: factors influencing aggregation in a simple host-parasite system. *Parasitology*, 147(1), 65-77. DOI:10.1017/S003118201900129X
- Trilles, J.-P., Öktener, A. (2004). *Livoneca sinuata* (Crustacea; Isopoda; Cymothoidae) on *Loligo vulgaris* from Turkey, and unusual cymothoid associations. *Dis Aquat Organ*, 61, 6.
- Vestbo, S., Hindberg, C., Forbes, M. R., Mallory, M. L., Merkel, F., Steenweg, R. J., . . . Provencher, J. F. (2019). Helminths in common eiders (*Somateria mollissima*): Sex, age, and migration have differential effects on parasite loads. *Int J Parasitol Parasites Wildl*, 9, 184-194. DOI:10.1016/j.ijppaw.2019.05.004
- Walker, J. G., Hurford, A., Cable, J., Ellison, A. R., Price, S. J., & Cressler, C. E. (2017). Host allometry influences the evolution of parasite host-generalism: theory and meta-analysis. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 372(1719), 20160089. DOI:10.1098/rstb.2016.0089
- Welicky, R. L., Malherbe, W., Hadfield, K. A., & Smit, N. J. (2019). Understanding growth relationships of African cymothoid fish parasitic isopods using specimens from museum and field collections. *Int J Parasitol Parasites Wildl*, 8, 182-187. DOI:10.1016/j.ijppaw.2019.02.002
- Williams, J. D. & Boyko, C. B. (2012). The Global Diversity of Parasitic Isopods Associated with Crustacean Hosts (Isopoda: Bopyroidea and Cryptoniscoidea). *PLoS One*, 7(4), e35350. DOI:10.1371/journal.pone.0035350

- Wunderlich, A., Simioni, W., Zica, E., & Siqueira, T. (2022). Experimental evidence that host choice by parasites is age-dependent in a fish-monogenean system. *Parasitol Res*, *121*(1), 115-126. DOI:10.1007/s00436-021-07356-9
- Wunderlich, A. C., Hattori, G. Y., & Trilles, J.-P. (2011). A New Host Record, *Palaemonetes Carteri* (Gordon, 1935) (Decapoda, Palaemonidae), for *Telotha Henselii* (Von Martens, 1869) (Isopoda, Cymothoidae). *Crustaceana*, *84*(11), 1403-1409. DOI:10.1163/156854011x603794
- Xu, Z., Liu, A., Li, S., Wang, G., & Ye, H. (2020). Hepatopancreas immune response during molt cycle in the mud crab, *Scylla paramamosain*. *Sci Rep*, *10*(1), 13102. DOI:10.1038/s41598-020-70139-2

Table 1. Total length (*TL*, mm) of *Macrobrachium brasiliense* prawn, *Telotha henselii* cymothoid isopod, and their parasitological indices for each season/year. Average±standard deviation (=Mean±SD).

Season/Year	Total length		Total length		Prevalence	Mean	Intensity	Total length	
	Male		Female			Intensity	range	Parasite	
	n	Mean±SD	n	Mean±SD	%	Mean±SD	Min-Max	n	Mean±SD
1995									
Winter	81	15.3±4.8	167	15.1±3.8	18.9	1.04±0.20	1-2	47	4.4±1.1
Spring	82	17.5±4.3	120	15.7±3.6	11.9	1	1	24	5.3±1.5
Summer	59	15.3±4.5	71	15.7±3.6	9.2	1.08±0.29	1-2	12	3.2±1.1
Autumn	99	14.2±3.8	100	15.5±3.8	16.6	1.12±0.41	1-3	33	3.8±0.8
Total	321	15.5±4.5	458	15.4±3.7	14.9	1.06±0.27	1-3	116	4.3±1,3
1998									
Winter	-	-	-	-	-	-	-	-	-
Spring	-	-	-	-	-	-	-	-	-
Summer	43	28.3±17.1	105	19.7±9.5	14.2	1.23±0.44	1-2	21	2.8±0.8
Autumn	16	31.7±11.8	69	23.9±8.2	34.1	1.24±0.51	1-3	29	3.4±0.8
Total	59	29.2±15.7	174	21.3±9.4	21.5	1.24±0.48	1-3	50	3.2±0.8
TOTAL	380	17.7±8.9	632	17.1±6.5	16.4	1.11±0.35	1-3	166	3.9±1.3

Table 2. Results of generalised linear mixed models describing the total abundance of ectoparasites (i.e., response variables) as a function of host-related traits (host body size, host sex, host age, and host molt stage) and hydrological factors (rainfall, streamflow, and habitat area). Parameter estimates, standard error (SE), Wald z-values, p-value and 95% confidence interval for the parameters of explanatory variables describing variation in ectoparasite abundances are provided. The bold terms are statistically significant. The confidence intervals (CI) that exclude zero are also reported in bold.

Parameter	Parameter estimate	SE	z-value	p-value	95% CI	
					Lower	Upper
Intercept	-3.354	0.610	-5.499	<0.001	-4.550	-2.159
Host body size	0.051	0.007	6.893	<0.001	0.036	0.065
Host sex (Female)	0.546	0.186	2.925	0.003	0.180	0.912
Host age (Adult)	0.459	0.608	0.755	0.755	-0.732	1.651
Host molt stage (Premolt)	-0.333	0.424	-0.786	0.432	-1.165	0.498
Host molt stage (Ecdysis)	0.511	0.200	2.551	0.010	0.118	0.904
Rainfall (Rt)	-0.003	0.001	-2.887	0.003	-0.005	-0.001
Streamflow	-0.985	0.300	-3.275	0.001	-1.575	-3.957
Habitat Area	0.409	0.175	2.328	0.019	0.064	0.754
Year	-0.244	0.132	-1.849	0.064	-0.503	0.014

Table 3. Variable inflation scores used to select variables included in the GLMM models. Predictor terms (term), variance inflation factor (VIF) and 95% confidence interval for VIF (VIF 95% CI). We excluded those with VIF > 10 (bold).

Term	VIF	VIF 95% CI	
		Lower	Upper
Host body size	1.06	1.02	1.19
Host sex	1.04	1.01	1.20
Host age	1.07	1.03	1.19
Host molt stage	1.04	1.01	1.22
Season	98.92	87.71	111.59
Rainfall (Rt)	5.68	5.09	6.35
Year	6.48	5.80	7.26
Streamflow	9.51	8.11	11.24
Area	8.58	7.66	9.63

Figure Captions

Figure 1. Map of the study area showing the geographic location of the Águas Claras stream, the Pardo River basin (Serra Azul municipality, São Paulo State, Brazil), where the freshwater prawns *Macrobrachium brasiliense* and the cymothoid *Telotha henselli* were captured at three sites over four years (1995, 1996, 1997 and 1998).

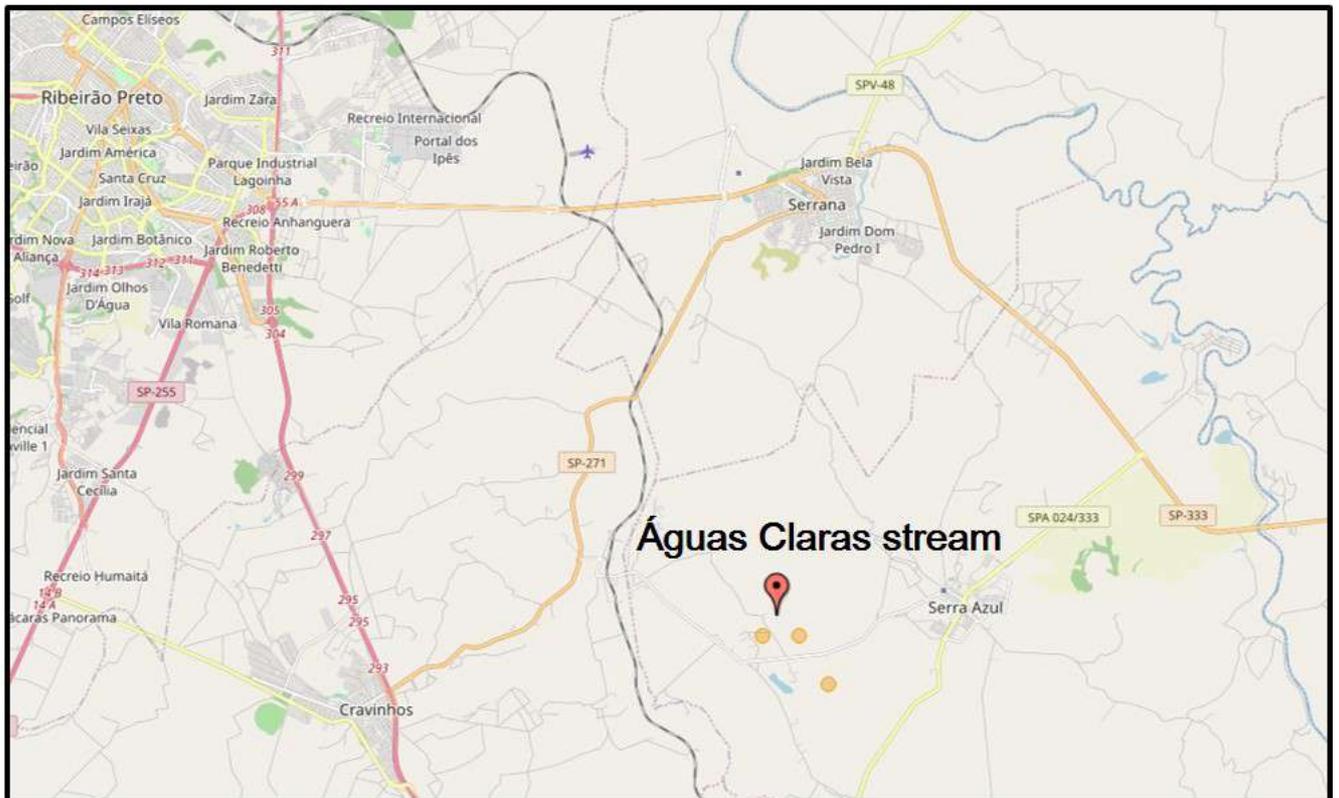
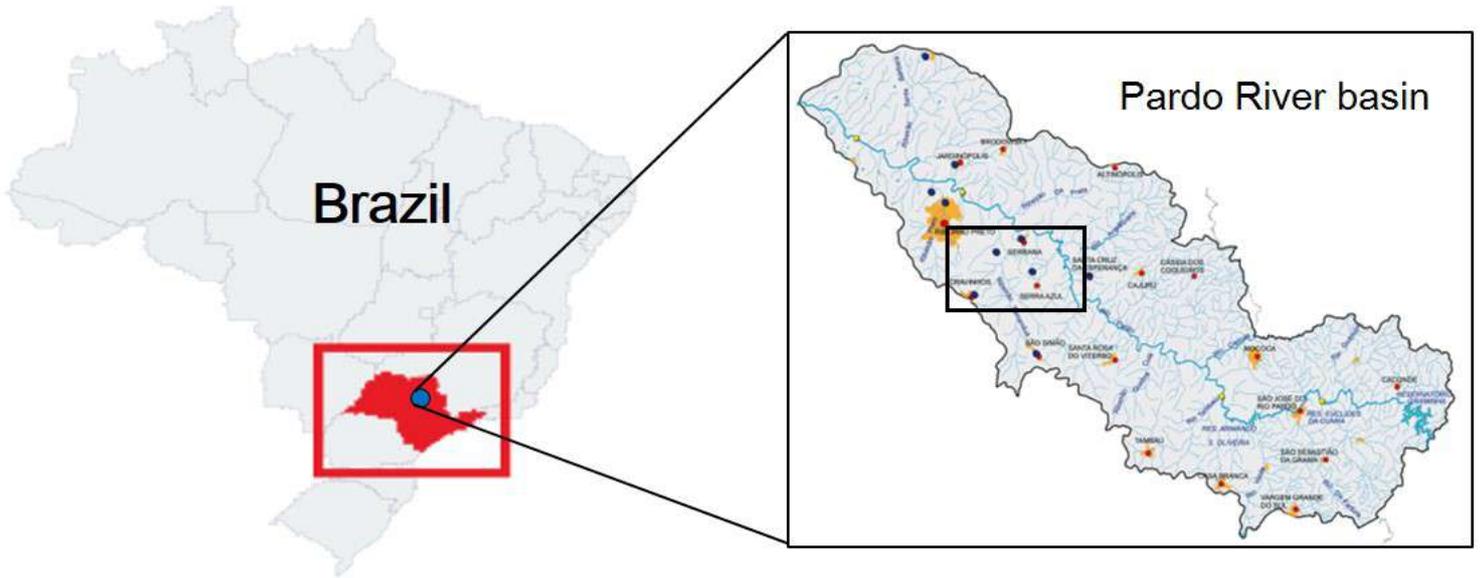
Figure 2. Relationship between host size and parasite size based on the generalised linear model (GLM) between sex (GLM: estimate \pm se: 0.003 ± 0.011 , Wald = 0.298 p = 0.766; Fig. 2a) and seasons (GLM: estimate \pm se: 0.013 ± 0.016 , Wald = 0.828, p = 0.408; Fig. 2b).

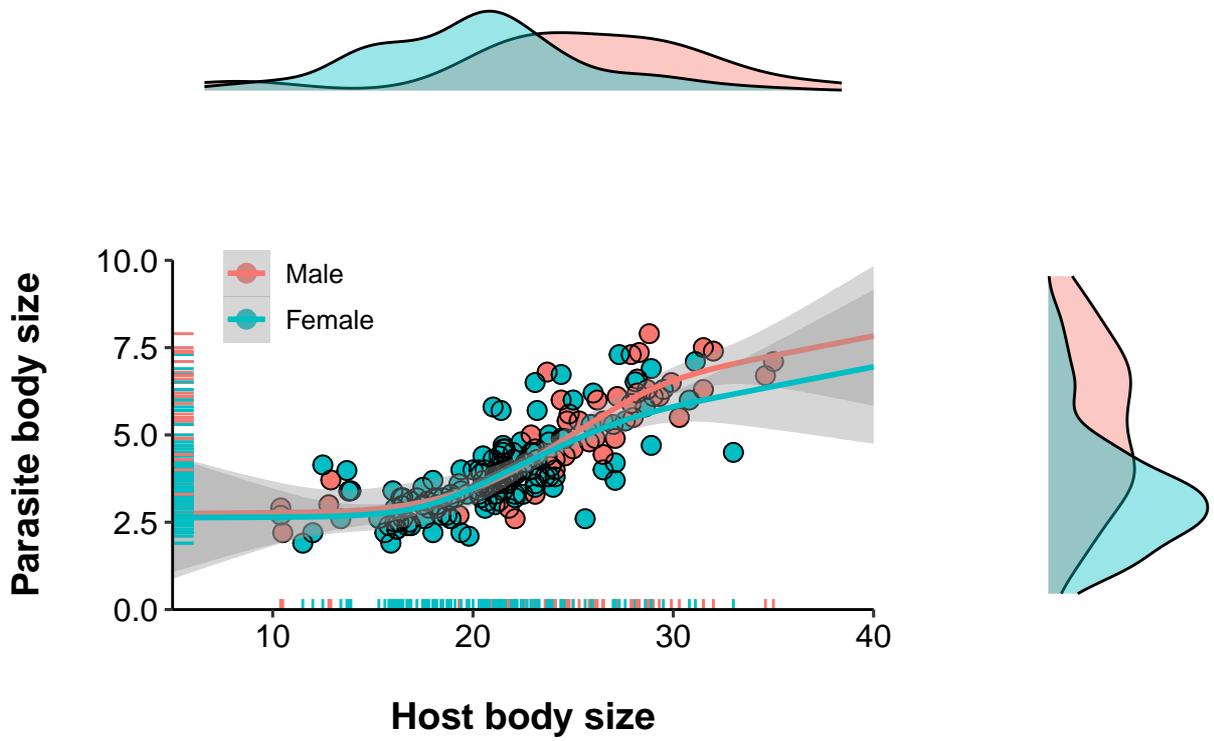
Figure 3. Variation in the total abundance of ectoparasites on *Macrobrachium brasiliense* between host size (a), host age (b), host sex (c) and host molt stage (d).

Figure 4. Variation in total ectoparasite abundance on *Macrobrachium brasiliense* between seasons (a) and years (b).

Figure 5. Variation in the total abundance of ectoparasites on *Macrobrachium brasiliense* between dry and wet seasons based on rainfall (mm).

Figure 6. Forest plots for comparison of part R^2 (coefficient of determination) for model predictors (A), inclusive R^2 (B), structure coefficients (C) and beta weights (D) including confidence intervals (CI) for the ectoparasite infestation model.



A**B**