1 Vulnerability of amphibians to global warming

- Patrice Pottier^{1,2*}, Michael R. Kearney³, Nicholas C. Wu⁴, Alex R. Gunderson⁵, Julie E. Rej⁵, A. Nayelli
 Rivera-Villanueva^{6,7}, Pietro Pollo¹, Samantha Burke¹, Szymon M. Drobniak^{1,8+}, and Shinichi Nakagawa^{1,9+}
 ¹ Evolution & Ecology Research Centre, School of Biological, Earth and Environmental Sciences,
 University of New South Wales, Sydney, New South Wales, Australia.
- ² Division of Ecology and Evolution, Research School of Biology, The Australian National University,
 Canberra, Australian Capital Territory, Australia
- 9 ³School of BioSciences, The University of Melbourne, Melbourne, Victoria, Australia
- ⁴ Hawkesbury Institute for the Environment, Western Sydney University, Richmond, New South Wales,
 Australia
- ⁵ Department of Ecology and Evolutionary Biology, Tulane University, New Orleans, Louisiana, USA
- 13 ⁶ Centro Interdisciplinario de Investigación para el Desarrollo Integral Regional Unidad Durango (CIIDIR),
- 14 Instituto Politécnico Nacional, Durango, México
- ⁷ Laboratorio de Biología de la Conservación y Desarrollo Sostenible de la Facultad de Ciencias Biológicas,
- 16 Universidad Autónoma de Nuevo León, Monterrey, México
- ⁸ Institute of Environmental Sciences, Jagiellonian University, Kraków, Poland.
- ⁹ Department of Biological Sciences, University of Alberta, Edmonton, Alberta, Canada.
- 19 *Corresponding author
- 20 ⁺These authors supervised the work equally
- 21 Corresponding author: Patrice Pottier (<u>p.pottier@unsw.edu.au</u>)

22 ORCID

- 23 Patrice Pottier https://orcid.org/0000-0003-2106-6597
- 24 Michael R. Kearney https://orcid.org/0000-0002-3349-8744
- 25 Nicholas C. Wu https://orcid.org/0000-0002-7130-1279
- 26 Alex R. Gunderson <u>https://orcid.org/0000-0002-0120-4246</u>
- 27 Julie E. Rej <u>https://orcid.org/0000-0002-3670-067X</u>
- 28 A. Nayelli Rivera-Villanueva https://orcid.org/0000-0002-9190-4317
- 29 Pietro Pollo <u>https://orcid.org/0000-0001-6555-5400</u>
- 30 Samantha Burke <u>https://orcid.org/0000-0001-6902-974X</u>
- 31 Szymon M. Drobniak <u>https://orcid.org/0000-0001-8101-6247</u>
- 32 Shinichi Nakagawa https://orcid.org/0000-0002-7765-5182

33 Main text

34 Amphibians are the most threatened vertebrates, yet their resilience to rising temperatures remains poorly understood^{1,2}. This is primarily because knowledge of thermal tolerance is taxonomically and 35 geographically biased³, compromising global climate vulnerability assessments. Here, we employed 36 37 a phylogenetically-informed data imputation approach to predict the heat tolerance of 60% of 38 amphibian species and assessed their vulnerability to daily temperature variation in thermal refugia. 39 We found that 104 out of 5203 species (2%) are currently exposed to overheating events in shaded terrestrial conditions. Despite accounting for heat tolerance plasticity, a 4°C global temperature 40 increase would create a step-change in impact severity, pushing 7.5% of species beyond their 41 physiological limits. In the Southern Hemisphere, tropical species encounter disproportionally more 42 43 overheating events, while non-tropical species are more susceptible in the Northern Hemisphere. These findings challenge evidence for a general latitudinal gradient in overheating risk⁴⁻⁶ and 44 underscore the importance of considering climatic variability in vulnerability assessments. We 45 46 provide conservative estimates assuming access to cool shaded microenvironments. Therefore, the 47 impacts of global warming will likely exceed our projections. Our microclimate-explicit analyses 48 demonstrate that vegetation and water bodies are critical in buffering amphibians during heat waves. Immediate action is needed to preserve and manage these microhabitat features. 49

50

51 Climate change has pervasive impacts on biodiversity, yet the extent and consequences of this 52 environmental crisis vary spatially and taxonomically^{7,8}. For ectothermic species, such as amphibians, the 53 link between climate warming and body temperature is clear, with immediate effects on physiological 54 processes⁹. Over 40% of amphibian species are currently listed as threatened, and additional pressures due 55 to escalating thermal extremes may further increase their extinction risk^{2,10}. Therefore, it is vital to assess 56 the resilience of amphibians to climate change to prioritise where and how conservation actions are taken.

Accurate assessments of resilience to climate change require adequate data on thermal tolerance and environmental exposure^{5,6,11}. However, the most exhaustive dataset on amphibian heat tolerance limits only covers 7.5% of known species and is geographically biased towards temperate regions³ (Fig. 1). This discrepancy is problematic, considering the high species richness in the tropics and the mounting evidence that tropical ectotherms are most susceptible to rising temperatures^{4–6,12,13}. Such sampling biases call into question the reliability of inferences in under-sampled areas and have implications for conservation strategies. Given the rapid pace of climate change and the finite resources available for research, acquiring sufficient empirical data to fill these knowledge gaps within a realistic timeframe is increasingly
untenable^{14,15}. Therefore, alternative methods to identify the populations and areas most susceptible to
thermal stress are critically needed in a rapidly warming climate.

67 Climate vulnerability assessments also require environmental data with high spatial and temporal resolution, particularly because extreme heat is more likely to trigger overheating events than increased 68 mean temperatures^{16–18}. When heat tolerance limits are known, cutting-edge approaches in biophysical 69 ecology allow fine-scale vulnerability assessments that account for morphology, behaviour, and 70 71 microhabitat setting in both historical and future climate projections^{19,20}. While broadly applicable, biophysically informed analyses are particularly relevant for amphibians, whose body temperatures depend 72 on evaporative heat loss and whose microhabitat use span terrestrial, aquatic, and arboreal environments. 73 Because microenvironmental features are essential for behavioural thermoregulation^{21,22}, modelling 74 microhabitats allow assessments of the effectiveness of different thermal refugia in buffering the impacts 75 76 of extreme heat events.

Here, we assess the global vulnerability of amphibians to extreme heat events in different climatic scenarios and thermal refugia (Extended Data Fig. 1). By integrating predicted thermal limits for 60% of amphibian species with daily operative body temperatures, our study offers the first comprehensive evaluation of the impact of heat extremes on the physiological viability of amphibians in nature.

81 Thermal limits and environmental exposure

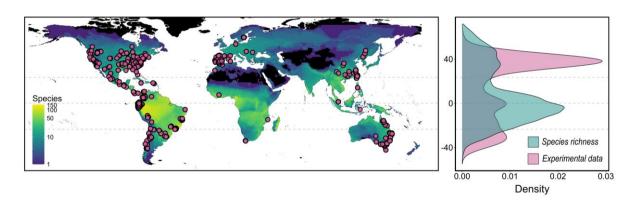
82 We first developed an approach to predict standardised thermal limits for 5,203 amphibian species using 83 data imputation based on phylogenetic niche clustering (Pagel's $\lambda = 0.95$ [0.91 - 0.98]) and known correlations between critical thermal limits (CT_{max}) and other variables (n = 2,661 estimates measured in 84 524 species; Methods). Our phylogenetic model-based imputation approach has expanded our 85 86 understanding of amphibian thermal tolerance by generating testable predictions for 4,679 unstudied 87 species, particularly in biodiversity hotspots (Fig. 1-2). We confirmed our imputation approach was likely accurate and unbiased by demonstrating a strong congruence between experimental and imputed data in 88 cross-validations (experimental mean \pm standard deviation = 36.19 \pm 2.67; imputed mean = 35.93 \pm 2.54; 89

90 n = 375; r = 0.86; Extended Data Fig. 2a,b), though, as expected, the uncertainty in imputed predictions
91 was higher in understudied clades (Extended Data Fig. 2c).

We then integrated predicted thermal limits with daily maximum operative body temperature fluctuations 92 93 estimated from biophysical models to evaluate the sensitivity of amphibians to extreme heat events in 94 terrestrial, aquatic, and arboreal microhabitats (Extended Data Fig. 1; Methods). Operative body 95 temperatures are the steady-state body temperatures that organisms would achieve in a given 96 microenvironment, which can diverge significantly from ambient air temperatures due to, for example, radiative and evaporative heat exchange processes^{19,20}. For each microhabitat, we modelled daily operative 97 98 body temperatures during the warmest quarters of 2006-2015 and across the distribution range of each species (Methods). We also used projected future climate data from TerraClimate²³ to generate projections 99 100 assuming 2°C or 4°C of global warming above pre-industrial levels. These temperatures are within the 101 range projected by the end of the century under low and intermediate/high greenhouse gas emission scenarios, respectively²⁴. Notably, recent historical CO₂ emissions most closely align with high warming 102 103 scenarios²⁵ (i.e., 4.3°C of predicted warming by 2100). All microenvironmental projections assumed access 104 to 85% of shade and sufficient humidity to maintain wet skin to simulate amphibians in thermal refugia 105 (Methods).

106 We estimated the vulnerability of amphibians by estimating daily differences between predicted thermal 107 limits and maximum hourly operative body temperatures (Extended Data Fig. 1; Methods). We also 108 adjusted daily thermal limits to assume that species were, on any given day, acclimated to local mean 109 weekly operative body temperatures, effectively accounting for plasticity throughout species' distribution 110 ranges (Methods). In total, we predicted vulnerability metrics for 203,853 local species occurrences 111 (individual species in 1° x 1° grid cells) in terrestrial conditions (5,177 species), 204,808 local species 112 occurrences in water bodies (5,203 species); and 56,210 local species occurrences (1,771 species) in above-113 ground vegetation, for each warming scenario. The number of species examined in arboreal conditions was 114 lower to reflect morphological adaptations required for climbing in above-ground vegetation. These estimates were then grouped into assemblages (all species occurring in 1° x 1° grid cells), tallying 14,090 115

- and 14,091 assemblages for terrestrial and aquatic species and 6,614 assemblages for arboreal species,
- 117 respectively.



119 Fig. 1 | Contrast between the geographical locations at which experimental data were collected and

- 120 **patterns in species richness.** Pink points denote experimental data (n = 587 species), while the colour 121 gradients refer to species richness calculated in 1 x 1 ° grid cells in the imputed data (n = 5,203 species).
- 122 Density plots on the right panel represent the distribution of experimental data ($\mu = 5,205$ species).
- species inhabiting these areas (blue) across latitudes. Dashed lines represent the equator and tropics.
- species innabiling these areas (blue) across failudes. Dashed lines represent the equator and tropies.

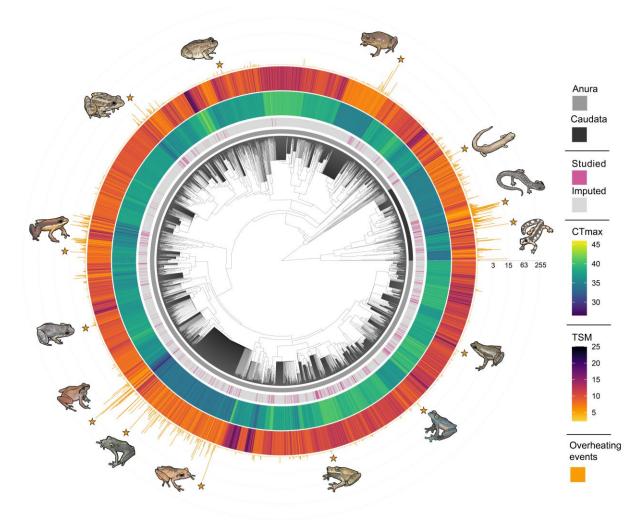
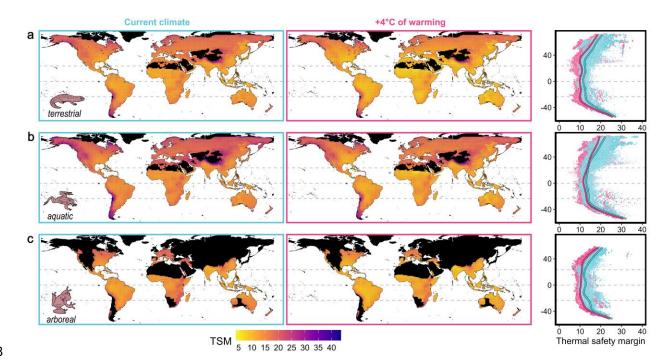


Fig. 2 | Phylogenetic coverage and taxonomic variation in climate vulnerability. Heat maps show heat 125 tolerance limits (CT_{max}) and thermal safety margins (TSM), while histograms show the number of 126 127 overheating events (days) averaged across each species' distribution range (n = 5,177 species). Pink bars 128 refer to species with prior knowledge (n = 521), while grey bars refer to entirely imputed species (n = 521) 129 4,656). This figure was constructed assuming ground-level microclimates occurring under 4°C of global warming above pre-industrial levels. Phylogeny is based on the consensus of 10,000 trees sampled from a 130 131 posterior distribution (see ²⁶ for details). Highlighted species starting from the right side, anti-clockwise: 132 Neurergus kaiseri (© Omid Mozaffari), Plethodon kiamichi (© Herps of Arkansas), Bolitoglossa 133 altamazonica (© Nick Volpe), Cophixalus aenigma (© Shane Black), Tomaptera cryptotis (© Warwick 134 Tarboton), Lithobates palustris (© Herps of Arkansas), Allobates subfolionidificans (© Albertina Pimentel 135 Lima), Phyzelaphryne miriamae (© Rafael Fraga), Barycholos ternetzi (© Werther Ramalho), Pristimantis 136 carvalhoi (© William E. Duellman), Pristimantis ockendeni (© Albertina Pimentel Lima), Boana curupi 137 (© Alfredo Sabaliauskas), Teratohyla adenocheira (©André Teles), Atelopus spumarius (© Mauricio 138 Vicariotto).

139

140 Vulnerability to historical and future heat

141 We first calculated thermal safety margins (TSM, sensu⁶) as the weighted mean difference between heat tolerance limits (CT_{max}) and the maximum daily body temperatures of the warmest quarters of 2006-2015 142 for each local species occurrence. Thermal safety margins averaged from long-term climatology are 143 routinely used in climate vulnerability analyses^{27–29}. We found evidence for a decline in TSM towards mid 144 145 to low latitudes in all microhabitats, a pattern maintained across warming scenarios (Fig. 3, Extended Data Fig. 3). However, warming substantially reduce TSM at all latitudes (Fig. 3), likely reflecting the contrast 146 between weak plastic responses in CT_{max} across latitudes^{11,15} and large variation in environmental 147 148 temperatures (Extended Data Fig. 3). Across all conditions simulated, TSM is always positive, even in the 149 highest warming scenario (Fig. 3, Extended Data Fig. 3). The mean TSM is lower for terrestrial (mean [95% confidence intervals]; current = 11.69 [8.86 - 14.43]; $+4^{\circ}C = 9.41$ [6.53 - 12.09]) and arboreal 150 conditions (current = 12.23 [9.40 - 14.96]; $+4^{\circ}C = 10.07 [7.23 - 12.80]$) than for water bodies (current = 151 $13.60 [10.71 - 16.28]; +4^{\circ}C = 11.68 [8.80 - 14.36];$ Fig. 3; Table S1). 152



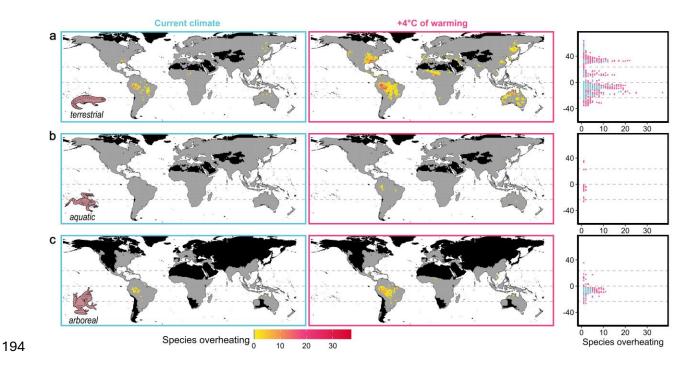
153

154 Fig. 3 | Assemblage-level patterns in thermal safety margin for amphibians in terrestrial (a), aquatic 155 (b) or arboreal (c) microhabitats. Thermal safety margins (TSM) were calculated as the weighted mean 156 difference between CT_{max} and the predicted operative body temperature in full shade during the warmest quarters of 2006-2015 in each assemblage (1-degree grid cell; n = 14,090 for terrestrial species; n = 14,091157 for aquatic species; n = 6,614 for arboreal species). Black colour depicts areas with no data. The right panel 158 depicts mean latitudinal patterns in TSM in current climates (blue) or assuming 4°C of global warming 159 above pre-industrial levels (pink), as predicted from generalised additive mixed models. Point estimates are 160 161 scaled by precision (1/s.e.), with smaller points indicating greater uncertainty. Dashed lines represent the 162 equator and tropics.

Because extreme heat events are more likely to trigger overheating events than mean temperatures 5.6.11, we 163 164 also calculated the binary probability (0/1) that operative body temperatures exceeded CT_{max} for at least 165 one day across the warmest quarters of 2006-2015 (i.e., overheating risk). Overall, overheating risk is low, 166 although numerous species are predicted to face overheating events locally (Fig. 4, Table S2). In terrestrial 167 conditions, we predict that 104 species (836 local species occurrences from 253 assemblages) are likely to 168 experience overheating events in current microclimates (Fig. 4-5). However, under 4°C of warming, 391 169 species (4,248 local species occurrences from 1,328 assemblages) are expected to overheat, which 170 represents nearly a four-fold increase relative to current conditions (Fig. 4-5; Table S2-3). The number of 171 species predicted to overheat in each grid cell also increases with warming; each assemblage comprises up 172 to 18 vulnerable species in current climates (mean [95% confidence intervals] = 3.19 [0.60 - 6.88] species) and up to 37 vulnerable species with 4°C of global warming (3.08 [0.62 - 6.56]; Fig. 4; Table S3). In 173 174 addition, the proportion of species predicted to experience overheating events in each assemblage varies

- 175 geographically and between warming scenarios (Extended Data Fig. 5; Table S4). The proportion of species
- 176 at risk is high in some areas with high species richness (e.g., Northern Australia, Southeastern United States)
- and not linearly predicted by latitude (Extended Data Fig. 5).

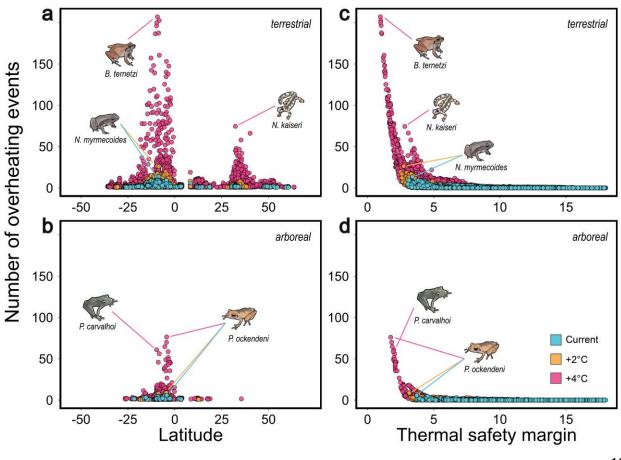
178 In current conditions for species that can shelter in trees (arboreal), 74 assemblages (comprising 1-6 species; 179 1.93 [0.05 – 5.05] species) are predicted to overheat, while 285 assemblages (comprising 1-11 species; 2.51 180 [0.31 – 5.69] species) are predicted to overheat assuming 4°C of global warming (Fig. 4; Table S3). While 181 the overheating risk is lower in arboreal conditions, considerably fewer species were examined than in 182 terrestrial conditions (1,771 vs. 5,177 species). In fact, comparing the responses of arboreal species in 183 different microhabitats revealed that occupying above-ground vegetation is only partially beneficial 184 (Extended Data Fig. 4). In current climates, up to 15 arboreal species (320 local species occurrences) are 185 predicted to experience overheating events in terrestrial conditions, whereas 13 arboreal species (152 local 186 species occurrences) are predicted to overheat in above-ground vegetation (Extended Data Fig. 4). 187 Furthermore, under 4°C of warming, 83 arboreal species (1,137 local species occurrences) are predicted to overheat in terrestrial conditions, while retreating to above-ground vegetation only reduces the number of 188 189 species exposed to overheating events by 32.5% (56 species, 748 local species occurrences) (Extended Data 190 Fig. 4). Contrary to terrestrial and arboreal conditions, no amphibian populations are predicted to overheat 191 in water bodies in current or intermediate climate warming scenarios due to the thermal buffering properties of water. However, assuming 4°C of climate warming, we predict that 11 species (56 local species 192 193 occurrences from 48 assemblages) will exceed their physiological limits in aquatic microhabitats (Fig. 4).



195 Fig. 4 | Number of species predicted to experience overheating events in terrestrial (a), aquatic (b), 196 and arboreal (c) microhabitats. The number of species overheating was assessed as the sum of species 197 overheating for at least one day in the period surveyed (warmest quarters of 2006-2015) in each assemblage (1-degree grid cell; n = 14,090 for terrestrial species; n = 14,091 for aquatic species; n = 6,614 for arboreal 198 species). Black colour depicts areas with no data, and grey colour assemblage without species at risk of 199 200 overheating. The right panel depicts latitudinal patterns in the number of species predicted to overheat in current climates (blue) or assuming 4°C of global warming above pre-industrial levels (pink). Dashed lines 201 202 represent the equator and tropics.

203 Finally, we quantified the number of days (out of 910 simulated days across the warmest quarters of 2006-204 2015) each species was predicted to locally exceed their plasticity-adjusted heat tolerance limits. This 205 metric fully integrates the frequency at which amphibians are predicted to experience temperatures beyond 206 their thermal limits. For current climates, we found that species rarely experience overheating events in 207 shaded terrestrial conditions (overall mean overheating days [95% confidence intervals] = 0.01 [0.01 - 0.01]208 (0.08); mean among overheating species = 2.15 [0.24 - 5.26] days); but these figures increase considerably 209 with global warming (Fig. 5; Table S2). Under 4°C of warming, species are predicted to overheat on as 210 many as 207.18 [182.39 – 231.97] days, representing up to 22.8% of the warmest days of the year (overall 211 mean = 0.15 [0.05 - 0.46] days; mean among overheating species = 6.75 [3.14 - 11.38] days; Fig. 5; Table 212 S2). This is noticeably more than what is predicted under 2° C of warming (overall mean = 0.02 [0.01 – 213 (0.13) days; mean among overheating species = 2.58 [0.41 - 5.86] days; Fig. 5; Table S2). In above-ground 214 vegetation, the frequency of overheating events is lower, as expected. Under current climates, arboreal

215 species are predicted to overheat on up to 5.65 [1.00 - 10.29] days in total (overall mean = 0.01 [0.01 -216 0.04] days; mean among overheating species = 1.62 [0.03 - 4.43] days; Fig. 5; Tab. Table S2). Under 4 217 degrees of warming, arboreal species are predicted to overheat on up to 76.17 [59.79 - 92.54] days (overall 218 mean = 0.08 [0.01 - 0.23] days; mean among overheating species = 5.08 [1.81 - 9.39] days; Fig. 5; Table 219 S2). Arboreal species retreating to above-ground vegetation are predicted to experience fewer overheating 220 events than those in terrestrial conditions (Extended Data Fig. 4). Interestingly, we found that species 221 predicted to overheat locally have TSMs well above zero, although some are living particularly close to 222 their heat tolerance limits during the warmest months in both terrestrial (mean [95% confidence intervals]; 223 current = 8.20 [6.91 - 9.98], range: 3.02 - 12.19; $+4^{\circ}C = 6.30 [5.02 - 8.09]$, range: 0.97 - 11.27) and above-224 ground conditions (current = 8.71 [7.20 - 10.28], range: 3.70 - 9.76; $+4^{\circ}C = 6.73$ [5.44 - 8.48], range: 1.75225 - 8.70; Fig. 5c,d). Finally, we found a strong non-linear negative association between the number of 226 overheating events and the thermal safety margin, with stark contrasts between warming scenarios (Fig. 227 5c,d; Table S5). In particular, overheating days increase rapidly as thermal safety margins fall below 5°C 228 (Figure 5c,d).



230 Fig. 5 | Latitudinal variation in the number of overheating events in terrestrial (a,c) and arboreal 231 (b,d) microhabitats as a function of latitude (a,b) and thermal safety margin (c,d). The number of 232 overheating events (days) were calculated based on the mean probability that daily maximum temperatures exceeded CT_{max} during the warmest quarters of 2006-2015 for each species in each grid cell (i.e., local 233 234 species occurrences; n = 203,853 for terrestrial species; n = 204,808 for aquatic species; n = 56,210 for 235 aquatic species). Blue points depict the number of overheating events in current microclimates, while 236 orange and pink points depict the number of overheating events assuming 2°C and 4°C of global warming 237 above pre-industrial levels, respectively. For clarity, only the species predicted to experience at least one 238 overheating event are depicted across latitudes (a,b). Highlighted species are: Neurergus kaiseri (© Omid 239 Mozaffari), Noblella myrmecoides (© Dante Fenolio), Barycholos ternetzi (© Werther Ramalho), 240 Pristimantis carvalhoi (© William E. Duellman), Pristimantis ockendeni (©Albertina Pimentel Lima).

241

242 The mounting impacts of global warming

Quantifying the resilience of biodiversity to a changing climate is one of the most pressing challenges for contemporary science^{7,8}. Here, we show that over a hundred species may already experience hourly temperatures that would likely result in death over minutes or hours of exposure in thermal refugia. This pattern is only predicted to worsen (Fig. 4-5). Assuming 4°C of global warming, the number of species and assemblages exposed to overheating events would be four to five times higher than currently, totalling 391 out of 5,203 species studied (7.5%; Fig. 4-5).

We also found striking disparities in overheating risk between the 2°C and 4°C warming projections (Fig. 249 250 5; Table S1), which are anticipated by the end of the century under low and high greenhouse gas emission scenarios, respectively²⁴. The more extreme warming scenario considerably increased the number 251 252 overheating events experienced by amphibian populations (Fig. 5), highlighting the escalating and abrupt impacts of global warming^{7,30}. Such an increase is attributable to the contrast between the rapid pace at 253 254 which temperatures are increasing and the low ability of amphibians to acclimate to new thermal environments via plasticity (Extended Data Fig. 3; species-level acclimation response ratio \pm s.d. = 0.134 255 \pm 0.008). Our study clearly demonstrates, as others have suggested^{18,28,31,32}, that physiological plasticity is 256 not a sufficient mechanism to buffer many populations from the impacts of rapidly rising temperatures. 257

258 Extreme heat events drive vulnerability

We found large spatial heterogeneity in the vulnerability of amphibians. In tropical areas, most vulnerablespecies are concentrated in South America and Australia, whereas fewer species are impacted in the African

261 and Asian tropics (Fig. 4). Tropical species also experience disproportionately more overheating events in 262 the Southern Hemisphere, while non-tropical species are more susceptible in the Northern Hemisphere (Fig. 263 5). Furthermore, the proportion of species experiencing overheating events in each assemblage was not 264 predicted by latitude (Extended Data Fig. 5). Therefore, our findings are inconsistent with the expectation of a general latitudinal gradient in overheating risk based on thermal safety margins^{4-6,13}. In fact, the 265 266 overheating risk does not increase linearly with TSM (Fig. 5c,d), and species with seemingly comparable 267 TSMs can have markedly different probabilities of overheating due to varying exposure to daily 268 temperature fluctuations (Fig. 5c,d). Therefore, TSMs alone hide critical tipping points for thermal stress 269 (Fig. 5c,d).

270 Our study questions the reliability of thermal safety margins and other climate vulnerability metrics when 271 averaged across large time scales (e.g., using the maximum temperature of the warmest quarter) for 272 detecting species most vulnerable to thermal extremes. It also challenges the general notion that lowlatitude species are uniformly most vulnerable to warming $^{4-6,13}$, revealing a far more nuanced pattern of 273 274 climate vulnerability across latitudes. While the reliability of TSM-based assessments has been questioned in previous studies¹¹, our work further emphasises the need to consider natural climatic variability and 275 276 extreme hourly temperatures^{4,16–18} when evaluating the vulnerability of ectotherms to global warming. 277 Considering alternative metrics, such as the number of predicted overheating events, may prove particularly 278 useful in identifying the most vulnerable species and populations.

279 The vital yet limited role of thermal retreats

280 Our study highlights the critical yet sometimes insufficient role that thermal retreats play in buffering the 281 impacts of warming on amphibians. Most amphibian species are predicted not to experience overheating 282 events in full shade (Fig. 4), and the availability of water bodies allows nearly all amphibians to maintain 283 their body temperatures below critical levels, apart from 11 species in the most extreme warming scenario 284 investigated. This is attributable to the higher specific heat capacity of water relative to air, delaying rapid temperature rises and affording a more stable environment during heat waves³³. Our findings add to the 285 286 growing evidence that finding access to cooler microhabitats is the main strategy amphibians and other ectotherms can use to maintain sub-lethal body temperatures^{6,21,34}. 287

However, it is crucial to emphasise that vegetated terrestrial conditions in full shade offer inadequate protection to 7.5% of species, and many arboreal species predicted to overheat at ground level face similar risks in above-ground vegetation (Fig. 4-5, Extended Data Fig. 4). In fact, although reducing the frequency of overheating events (Extended Data Fig. 4), access to shaded above-ground vegetation only reduces the number of vulnerable species by 32.5%. Moreover, although burrows offer cooler microclimates (see Extended Data Fig. 9), the ability to use underground spaces is not universal among amphibians and can greatly restrict activity, reproduction, and foraging opportunities.

295 Warming impacts may exceed projections

296 Our predictions are largely conservative, and likely overestimate the resilience of amphibians to global 297 warming in two main ways. First, we assume that microhabitats such as shaded ground-level substrates, 298 above-ground vegetation, and water bodies are available throughout a species' range, and that amphibians 299 can maintain wet skin. These assumptions will often be violated as habitats are degraded. Deforestation and 300 urbanization are diminishing vital shaded areas^{35,36}, while increased frequencies of droughts will cause water bodies to evaporate^{37,38}. These changes compromise not only habitat integrity but also local humidity 301 302 levels – key for effective thermoregulation^{39,40}. Consequently, amphibians will likely experience higher 303 body temperatures and desiccation stress events than our models predict due to inconsistent access to cooler microhabitats⁴¹, particularly in degraded systems. 304

Second, ectotherms can experience deleterious effects from heat stress before reaching their heat tolerance limits. Prolonged exposure to sub-lethal temperatures can lead to altered activity windows^{42,43}, disruptions to phenology^{44,45}, reduced reproductive fitness (fertility and fecundity)^{29,46,47}, and death^{48,49}. Although comprehensive data on thermal incapacitation times and fertility impacts are sparse in amphibians, integrating both the duration and intensity of thermal stress^{49–51} will likely point to more extreme vulnerability estimates. This represents a vital avenue for future research, albeit one requiring a large collection of empirical data.

Alternatively, species that can retreat underground during heat events are likely to experience fewer overheating events than our models predict (see Extended Data Fig. 9), and prolonged exposure to high temperatures in the permissive range (*sensu*⁴⁸) can enhance performance and fitness, thereby reducing the 315 impacts of extreme heat on natural populations. In addition, some species may adapt to changing 316 temperatures. However, evidence for slow rates of evolution and physiological constraints on thermal 317 tolerance^{52,53} challenges the likelihood of local adaptation to occur in rapidly warming climates.

318 The power of data imputation

319 Our imputation approach has generated testable predictions of the thermal limits of 5,203 species, 320 expanding the scope of previous research³ (Fig. 2). We also addressed geographical biases by generating 321 predictions in under-sampled but ecologically critical regions of Africa, Asia, and South America (Fig. 2). 322 We found that these understudied regions frequently harbor species exhibiting the highest susceptibility to 323 extreme heat events (Fig. 1,4-5), with 74% (288 out of 391) of vulnerable species remaining unstudied. 324 Targeted research efforts in these vulnerability hotspots are instrumental in validating our model predictions 325 and advancing our understanding of amphibian thermal physiology to inform their conservation. Though 326 undeniable logistical and financial challenges exist in accessing some of these remote locations, 327 collaboration with local scientists could expedite data collection and result in timely conservation measures. Exemplary initiatives to sample numerous species in South America (e.g., ^{22,54,55}) are promising steps in 328 329 this direction, and we hope our findings will catalyse research activity in these regions.

330 Amphibian biodiversity in a warming world

Our study highlights the dire consequences of global warming on amphibians. Yet it is crucial to differentiate between global extinction and local extirpations – the latter being confined extinctions within specific geographic areas. Most species will not experience overheating events throughout their entire range, and these overheating events may not occur simultaneously. Hence, most species are likely to only experience local extirpation due to overheating, according to our models. Nevertheless, local extirpations carry their own sets of ecological repercussions, such as reshuffling community compositions and eroding genetic and ecological diversity^{56,57}.

338 Some amphibian populations may also undergo range shifts, permanently or transiently relocating to 339 habitats with more hospitable weather patterns⁵⁸. However, this is only possible if suitable habitats are 340 available for establishment. Given the low dispersal rates of some amphibians and their common reliance on water bodies for reproduction and thermoregulation, opportunities for range shifts are likely to be rare for many species. Identifying which species at high risk of overheating are simultaneously predicted to have limited ability to extend their range is an interesting avenue for research. In addition, we stress that amphibians living close to their physiological limits for extended times at the warm edge of their distribution are likely to experience heat stress that could hamper activity, foraging opportunities, and reproductive success, adding layers of complexity to their survival challenges and potentially leading to population declines^{42,48}.

348 Overall, our study contributes to the evidence that climate change is a mounting threat to amphibians^{2,10} 349 and emphasises the importance of limiting global temperature rises below 2°C to minimise the risk of 350 overheating to amphibian populations. A 4°C temperature rise would not just increase these risks but create 351 a step-change in impact severity (e.g., Fig. 5c). The mechanistic basis of our species- and habitat-specific 352 predictions also leads to clear management priorities. Particularly, our analyses revealed the critical 353 importance of preserving dense vegetation cover and water bodies. These microhabitats provide conditions 354 with cooler and more stable temperatures and increase the potential for amphibians and other ectothermic species to disperse to more suitable microhabitats. Establishing protected areas and undertaking habitat 355 356 restoration initiatives may support amphibians in a changing climate and buffer additional anthropogenic 357 threats, in turn mitigating amphibian population declines^{2,10,59}. These actions are critical for the amphibians at risk and the ecosystems they support⁶⁰ in a planet undergoing perilous climatic changes. 358

360 Main references

- Carey, C. & Alexander, M. A. Climate change and amphibian declines: is there a link? *Divers*.
 Distrib. 9, 111–121 (2003).
- Luedtke, J. A. *et al.* Ongoing declines for the world's amphibians in the face of emerging threats.
 Nature 622, 308–314 (2023).
- 365 3. Pottier, P. *et al.* A comprehensive database of amphibian heat tolerance. *Sci. Data* **9**, 600 (2022).
- Pinsky, M. L., Eikeset, A. M., McCauley, D. J., Payne, J. L. & Sunday, J. M. Greater vulnerability
 to warming of marine versus terrestrial ectotherms. *Nature* 569, 108–111 (2019).
- 368 5. Deutsch, C. A. *et al.* Impacts of climate warming on terrestrial ectotherms across latitude. *Proc.*369 *Natl. Acad. Sci. U.S.A.* 105, 6668–6672 (2008).
- Sunday, J. M. *et al.* Thermal-safety margins and the necessity of thermoregulatory behavior across
 latitude and elevation. *Proc. Natl. Acad. Sci. U.S.A.* 111, 5610–5615 (2014).
- 372 7. Urban, M. C. Accelerating extinction risk from climate change. *Science* **348**, 571–573 (2015).
- 8. Walther, G.-R. *et al.* Ecological responses to recent climate change. *Nature* **416**, 389–395 (2002).
- Angilletta, M. J. Thermal adaptation: a theoretical and empirical synthesis. (Oxford University
 Press, 2009).
- 376 10. Mi, C. *et al.* Global Protected Areas as refuges for amphibians and reptiles under climate change.
 377 *Nat. Commun.* 14, 1389 (2023).
- 11. Clusella-Trullas, S., Garcia, R. A., Terblanche, J. S. & Hoffmann, A. A. How useful are thermal
 vulnerability indices? *Trends Ecol. Evol.* 36, 1000–1010 (2021).
- Huey, R. B. *et al.* Why tropical forest lizards are vulnerable to climate warming. *Proc. R. Soc. B* **276**, 1939–1948 (2009).
- Comte, L. & Olden, J. D. Climatic vulnerability of the world's freshwater and marine fishes. *Nat. Clim. Change* 7, 718–722 (2017).
- 14. Carvalho, R. L. *et al.* Pervasive gaps in Amazonian ecological research. *Curr. Biol.* 33, 34953504.e4 (2023).
- 15. Nesi, P., Luiselli, L. M. & Vignoli, L. "Heaven" of Data Deficient Species: The Conservation Status
 of the Endemic Amphibian Fauna of Vietnam. *Diversity* 15, 872 (2023).
- 388 16. Müller, J. *et al.* Weather explains the decline and rise of insect biomass over 34 years. *Nature* 628, 349–354 (2024).
- Murali, G., Iwamura, T., Meiri, S. & Roll, U. Future temperature extremes threaten land vertebrates.
 Nature 615, 461–467 (2023).
- 392 18. Gunderson, A. R., Dillon, M. E. & Stillman, J. H. Estimating the benefits of plasticity in ectotherm
 393 heat tolerance under natural thermal variability. *Funct. Ecol.* 31, 1529–1539 (2017).
- Anderson, R. O., White, C. R., Chapple, D. G. & Kearney, M. R. A hierarchical approach to
 understanding physiological associations with climate. *Glob. Ecol. Biogeogr.* 31, 332–346 (2022).

- Briscoe, N. J. *et al.* Mechanistic forecasts of species responses to climate change: The promise of
 biophysical ecology. *Glob. Change Biol.* 29, 1451–1470 (2023).
- 398 21. Kearney, M., Shine, R. & Porter, W. P. The potential for behavioral thermoregulation to buffer
 399 "cold-blooded" animals against climate warming. *Proc. Natl. Acad. Sci. U.S.A.* 106, 3835–3840
 400 (2009).
- 22. Duarte, H. *et al.* Can amphibians take the heat? Vulnerability to climate warming in subtropical and
 temperate larval amphibian communities. *Glob. Change Biol.* 18, 412–421 (2012).
- 403 23. Abatzoglou, J. T., Dobrowski, S. Z., Parks, S. A. & Hegewisch, K. C. TerraClimate, a high404 resolution global dataset of monthly climate and climatic water balance from 1958–2015. *Sci. Data*405 5, 170191 (2018).
- 406 24. Masson-Delmotte, V. *et al.* Climate change 2021: the physical science basis. *Contribution of*407 *working group I to the sixth assessment report of the intergovernmental panel on climate change* 2,
 408 (2021).
- Schwalm, C. R., Glendon, S. & Duffy, P. B. RCP8.5 tracks cumulative CO2 emissions. *Proc. Natl. Acad. Sci. U.S.A.* 117, 19656–19657 (2020).
- 411 26. Jetz, W. & Pyron, R. A. The interplay of past diversification and evolutionary isolation with present
 412 imperilment across the amphibian tree of life. *Nat Ecol Evol* 2, 850–858 (2018).
- 27. Clusella-Trullas, S., Blackburn, T. M. & Chown, S. L. Climatic Predictors of Temperature
 Performance Curve Parameters in Ectotherms Imply Complex Responses to Climate Change. *Am. Nat.* 177, 738–751 (2011).
- 416 28. Morley, S. A., Peck, L. S., Sunday, J. M., Heiser, S. & Bates, A. E. Physiological acclimation and
 417 persistence of ectothermic species under extreme heat events. *Glob. Ecol. Biogeogr.* 28, 1018–1037
 418 (2019).
- 419 29. van Heerwaarden, B. & Sgrò, C. M. Male fertility thermal limits predict vulnerability to climate
 420 warming. *Nat. Commun.* 12, 2214 (2021).
- 30. Trisos, C. H., Merow, C. & Pigot, A. L. The projected timing of abrupt ecological disruption from
 climate change. *Nature* 580, 496–501 (2020).
- 423 31. Gunderson, A. R. & Stillman, J. H. Plasticity in thermal tolerance has limited potential to buffer
 424 ectotherms from global warming. *Proc. R. Soc. B* 282, 20150401 (2015).
- 425 32. Pottier, P. *et al.* Developmental plasticity in thermal tolerance: Ontogenetic variation, persistence,
 426 and future directions. *Ecol. Lett.* 25, 2245–2268 (2022).
- 33. Denny, M. W. Thermal Properties: Body temperatures in Air and Water. in *Air and Water: The biology and physics of life's media* 145–173 (Princeton University Press, Princeton, New Jersey,
 USA, 1993).
- 430 34. Scheffers, B. R., Edwards, D. P., Diesmos, A., Williams, S. E. & Evans, T. A. Microhabitats reduce
 431 animal's exposure to climate extremes. *Glob. Change Biol.* 20, 495–503 (2014).

- 432 35. Stark, G., Ma, L., Zeng, Z.-G., Du, W.-G. & Levy, O. Cool shade and not-so-cool shade: How
 433 habitat loss may accelerate thermal stress under current and future climate. *Glob. Change Biol.* 29,
 434 6201–6216 (2023).
- 435 36. Nowakowski, A. J. *et al.* Tropical amphibians in shifting thermal landscapes under land-use and
 436 climate change. *Conserv. Biol.* **31**, 96–105 (2017).
- 437 37. Dai, A. Increasing drought under global warming in observations and models. *Nat. Clim. Change* 3,
 438 52–58 (2013).
- 439 38. McMenamin, S. K., Hadly, E. A. & Wright, C. K. Climatic change and wetland desiccation cause
 440 amphibian decline in Yellowstone National Park. *Proc. Natl. Acad. Sci. U.S.A.* 105, 16988–16993
 441 (2008).
- 39. Greenberg, D. A. & Palen, W. J. Hydrothermal physiology and climate vulnerability in amphibians. *Proc. R. Soc. B* 288, 20202273 (2021).
- 444 40. Cheng, C.-T. *et al.* Open habitats increase vulnerability of amphibian tadpoles to climate warming
 445 across latitude. *Glob. Ecol. Biogeogr.* 32, 83–94 (2023).
- 446 41. Wu, N. C. *et al.* Global exposure risk of frogs to increasing environmental dryness. Preprint at
 447 https://doi.org/10.32942/X2ZG7S (2024).
- 448 42. Kearney, M. R. Activity restriction and the mechanistic basis for extinctions under climate
 449 warming. *Ecol. Lett.* 16, 1470–1479 (2013).
- 43. Enriquez-Urzelai, U. *et al.* The roles of acclimation and behaviour in buffering climate change
 impacts along elevational gradients. *J. Anim. Ecol.* **89**, 1722–1734 (2020).
- 44. Enriquez-Urzelai, U., Nicieza, A. G., Montori, A., Llorente, G. A. & Urrutia, M. B. Physiology and
 acclimation potential are tuned with phenology in larvae of a prolonged breeder amphibian. *Oikos*2022, e08566 (2022).
- 45. Parmesan, C. Ecological and Evolutionary Responses to Recent Climate Change. *Annu. Rev. Ecol.*456 *Evol. Syst.* 37, 637–669 (2006).
- 46. Wang, W. W.-Y. & Gunderson, A. R. The Physiological and Evolutionary Ecology of Sperm
 Thermal Performance. *Front. Physiol.* 13, (2022).
- 459 47. Walsh, B. S. *et al.* The Impact of Climate Change on Fertility. *Trends Ecol. Evol.* 34, 249–259
 460 (2019).
- 461 48. Jørgensen, L. B., Ørsted, M., Malte, H., Wang, T. & Overgaard, J. Extreme escalation of heat
 462 failure rates in ectotherms with global warming. *Nature* 611, 93–98 (2022).
- 463 49. Rezende, E. L., Castañeda, L. E. & Santos, M. Tolerance landscapes in thermal ecology. *Funct.*464 *Ecol.* 28, 799–809 (2014).
- 465 50. Garcia, R. A., Allen, J. L. & Clusella-Trullas, S. Rethinking the scale and formulation of indices
 466 assessing organism vulnerability to warmer habitats. *Ecography* 42, 1024–1036 (2019).

- Jørgensen, L. B., Malte, H., Ørsted, M., Klahn, N. A. & Overgaard, J. A unifying model to estimate
 thermal tolerance limits in ectotherms across static, dynamic and fluctuating exposures to thermal
 stress. *Sci. Rep.* 11, 12840 (2021).
- 470 52. Bennett, J. M. *et al.* The evolution of critical thermal limits of life on Earth. *Nat. Commun.* 12, 1198
 471 (2021).
- 472 53. Morgan, R., Finnøen, M. H., Jensen, H., Pélabon, C. & Jutfelt, F. Low potential for evolutionary
 473 rescue from climate change in a tropical fish. *Proc. Natl. Acad. Sci. U.S.A.* 117, 33365–33372
 474 (2020).
- 475 54. May, R. von *et al.* Thermal physiological traits in tropical lowland amphibians: Vulnerability to
 476 climate warming and cooling. *PLOS One* 14, e0219759 (2019).
- 477 55. Bovo, R. P. *et al.* Beyond Janzen's Hypothesis: How Amphibians That Climb Tropical Mountains
 478 Respond to Climate Variation. *Integr. Org. Biol.* 5, obad009 (2023).
- 479 56. Arenas, M., Ray, N., Currat, M. & Excoffier, L. Consequences of Range Contractions and Range
 480 Shifts on Molecular Diversity. *Mol. Biol. Evol.* 29, 207–218 (2012).
- 481 57. Rogan, J. E. *et al.* Genetic and demographic consequences of range contraction patterns during
 482 biological annihilation. *Sci. Rep.* 13, 1691 (2023).
- 483 58. Blaustein, A. R. *et al.* Direct and Indirect Effects of Climate Change on Amphibian Populations.
 484 *Diversity* 2, 281–313 (2010).
- 485 59. Nowakowski, J. A. *et al.* Protected areas slow declines unevenly across the tetrapod tree of life.
 486 *Nature* 622, 101–106 (2023).
- 487 60. Hocking, D. & Babbitt, K. Amphibian Contributions to Ecosystem Services. *Herpetol. Conserv.*488 *Biol.* (2014).
- 489

490 Methods

491 *Reporting*

We report author contributions using the CRediT (Contributor Roles Taxonomy) statement⁶¹ and MeRIT
(Method Reporting with Initials for Transparency) guidelines⁶². We also crafted the study title, abstract and
keywords to maximise indexing in search engines and databases⁶³. All analyses were performed using R
statistical software⁶⁴ (v. 4.3.0), and most computations used the computational cluster Katana supported by
Research Technology Services at UNSW Sydney. Maps, phylogenetic trees, and data visualisations were
generated using the R packages rnaturalearthhires⁶⁵, ggtree⁶⁶, and ggplot2⁶⁷.

498 Amphibian heat tolerance limits

We leveraged the most comprehensive compilation of amphibian heat tolerance limits³ for our analyses 499 500 (Extended Data Fig. 1). Briefly, these data were collated by systematically reviewing the literature in five 501 databases and seven languages, comprising 3,095 heat tolerance limits from 616 amphibian species. To 502 facilitate the comparability and analysis of heat tolerance limits, we only included data matching four 503 specific criteria. First, we only included heat tolerance limits measured using a dynamic methodology (i.e., 504 temperature at which animals lose their motor coordination when exposed to ramping temperatures, critical thermal maximum CT_{max}⁶⁸) because it was the most used and comparable metric. Second, we only selected 505 506 data for which the laboratory acclimation temperature, or the field temperature during the month of capture, was recorded. Third, we only included data from species listed in the phylogeny from 26 . Fourth, we only 507 508 included species for which their geographical range was reported in the International Union for the Conservation of Nature red list⁶⁹ (accessed in January 2023). 509

These criteria were chosen to perform phylogenetically, climatically, and spatially informed analyses. In total, we selected 2,661 heat tolerance limits estimates with metadata for 524 amphibian species (mean = 5.08; range = 1 - 146 estimates per species; 287 species with more than one estimate). We also complemented this dataset with ecotypic data for each species. Amphibians were grouped into six major ecotypes according to ⁴¹: ground-dwelling, fossorial, aquatic, semi-aquatic, stream-dwelling and arboreal. Cave specialists were excluded because they experience unique microclimatic conditions.

516 Data-deficient species

517 Our objective was to assess the thermal tolerance of amphibians globally. However, the data compiled in ³ 518 are geographically and taxonomically biased. Therefore, we employed a data imputation procedure to infer 519 the thermal tolerance of data-deficient species, totalling 5,203 species at a broad geographical coverage (524 species + 4,679 data-deficient species; ~60% of all described amphibian species, amphibiaweb.org; 520 521 accessed in December 2023). We selected data-deficient species from a species list that matched the phylogeny from ²⁶ (7,238 species), was listed in the IUCN red list⁶⁹ along with geographic distribution data 522 523 (5,792 species), and for which ecotypes were known (6,245 species). We did not consider Caecilians (order 524 Gymnophiona) because, to our knowledge, heat tolerance limits are unknown for all Caecilian species³. Of 525 the 5,792 species for which we had distribution and phylogenetic data, 5,268 were data-deficient for CT_{max} , 526 of which 4,822 had a known ecotype. After removing Caecilians, we were left with 4,679 species to impute. 527 We also supplemented our dataset with published body mass data retrieved from literature sources or estimated based on length-mass allometries^{41,70,71}. We then estimated the geographical coordinates at which 528 all extant species occurred in their IUCN distribution range at a 1° x 1° resolution to use for biophysical 529 530 modelling (Extended Data Fig. 1).

531 Data imputation

532 We developed a phylogenetic imputation procedure, here named Bayesian Augmentation with Chained 533 Equations (BACE). The BACE procedure combines the powers of Bayesian data augmentation and multiple imputation with chain equations (MICE⁷²). Briefly, we ran multiple iterative models using 534 $MCMCglmm^{73}$ (v. 2.34) and supporting functions from the *hmi* package⁷⁴. In the first cycle, missing data 535 536 was either taken as the arithmetic mean for continuous predictors, or randomly sampled from existing 537 values for (semi)categorical predictors. Predicted (augmented) values from the models were then extracted 538 from the response variables and used as predictor variables in the next models to predict other response 539 variables. Ultimately, heat tolerance limits were predicted using augmented data from all predictors. We 540 ran 5 cycles where the data from one cycle was iteratively used in the next cycle, and estimations converged 541 after the first cycle. Although the proportion of missing data was large (89.9%), imputations based on large amounts of missing data are common^{13,75}, and although estimate uncertainty increases with the proportion 542

of missing data, as expected, simulation studies have shown estimations remain unbiased^{76,77}. Note, however, that although our approach took the uncertainty of missing data in the response or variable of interest (CT_{max}) into account, we used the most likely values for the predictors. While such an approach could underestimate the uncertainty in the response, point estimates should not be biased. In fact, our crossvalidation approach demonstrated the ability of our models to predict back known experimental estimates with reasonable error (experimental mean ± standard deviation = 36.19 ± 2.67 ; imputed mean = $35.93 \pm$ 2.54; r = 0.86; Extended Data Fig. 2).

550 Heat tolerance limits were imputed based on the species' acclimation temperatures, the duration of 551 acclimation, the ramping rate and endpoint used in assays, the medium used for measuring heat tolerance 552 limits (i.e., ambient temperatures, water/body temperatures), and the life stage of the animals (adults or 553 larvae), and their ecotype. These variables were correlated with amphibian heat tolerance limits and were 554 fitted as covariates in Bayesian linear mixed models. We also weighted heat tolerance estimates based on 555 the inverse of their sampling variance, accounted for phylogenetic non-independence using a correlation 556 matrix of phylogenetic relatedness, and fitted random intercepts for species-specific effects and 557 phylogenetic effects, as well as their correlation with acclimation temperatures (i.e., random slopes). In 558 other words, we modelled species-specific slopes (acclimation response ratio) and partitioned the variance 559 among phylogenetic and non-phylogenetic effects. We imputed data for adult amphibians assuming they were acclimated to the median, 5th, or 95th percentile operative body temperatures experienced across their 560 561 geographical range (see *Microenvironmental data and biophysical modelling*) for a duration of 10 days, 562 tested using a ramping rate of 1°C/min in a container filled with water, and for which thermal tolerance 563 endpoint was recorded as the onset of spasms. These methodological parameters were the median values in 564 the experimental dataset, or the most common values (mode). This allowed standardization of heat tolerance limits for the comparative analysis^{78–80}. In amphibians, the onset of spasms usually occurs after the loss of 565 righting response⁸¹, meaning that our estimates are conservative. While we did include data from larvae in 566 567 the training data, we only imputed data for adults to increase the comparability of our estimates.

568 For both known species and data-deficient species, we generated three ecologically relevant and 569 standardised heat tolerance estimates, and all analyses were built upon these standardised imputed estimates. In total, we generated data for 5,203 species of amphibians (Extended Data Fig. 1-2). Notably,
our imputed estimates are accompanied by standard errors, which provide estimates of uncertainty in the
imputation, and errors were propagated throughout our analyses (see *Climate vulnerability analysis*).

573 Microenvironmental data and biophysical modelling

We used the package *NicheMapR*^{82,83} (v. 3.2.1) to estimate microenvironmental temperatures and hourly operative body temperatures in current (2006-2015) and projected climatic conditions (2°C or 4°C of global warming above pre-industrial levels). Operative body temperatures are the steady-state body temperatures that organisms would achieve in a given microenvironment, which can diverge significantly from ambient air temperatures due to, for example, radiative and evaporative heat exchange processes^{19,20,84–89}.

579 For each geographic location, we generated microclimatic temperatures experienced by amphibians on i) a 580 vegetated ground-level substrate (i.e., terrestrial), ii) in above-ground vegetation (i.e., arboreal), or iii) in a 581 water body (i.e., aquatic) (Extended Data Fig, 1). For terrestrial and aquatic species, we simulated 582 microenvironmental temperatures 1 cm above the surface. For arboreal species, we simulated 583 microenvironmental temperatures 2 meters above ground, applied a reduction of 80% in windspeed to account for reduced wind due to vegetation⁹⁰, and assumed that 90% of the solar radiation was diffused due 584 to canopy cover⁸¹. All microenvironmental projections were made using 85% shade to simulate animals in 585 586 thermal refugia, i.e., the microhabitats in which animals would retreat during the hottest times of the day. 587 We did not model temperatures in the sun because ectothermic species most likely behaviourally thermoregulate by retreating to thermal refugia during extreme heat events²¹. Our calculations thus 588 589 represent conservative estimates of the vulnerability of amphibians to extreme temperature events.

For microclimatic temperature estimates, we used the *micro_ncep* function from *NicheMapR*⁸² (v. 3.2.1), which integrates 6-hourly macroclimatic data from the National Center for Environmental Predictions (NCEP). This function also inputs from the *microclima* package⁹² (v. 0.1.0) to predict microclimatic temperatures after accounting for variation in radiation, wind speed, altitude, albedo, vegetation, and topography. These data are downscaled to an hourly resolution, producing high-resolution microclimatic data. We used projected future monthly climate data from TerraClimate²³ to generate hourly projections 596 assuming 2°C or 4°C of global warming above pre-industrial levels. These temperatures are within the range projected by the end of the century under low (Shared Socioeconomic Pathway SSP 1-2.6 to SSP 2-597 4.5) and high (SSP 3-7.0 to SSP 5-8.5) greenhouse gas emission scenarios, respectively²⁴. TerraClimate 598 599 projections use monthly data on precipitation, minimum temperature, maximum temperature, wind speed, 600 vapor pressure deficit, soil moisture, and downward surface shortwave radiation. These projections impose monthly climate projections from 23 CMIP5 global circulation models, as described in ⁹³. The *micro* ncep 601 602 function then downscales monthly TerraClimate inputs to hourly by imposing a diurnal cycle to the data 603 and imposes TerraClimate offsets onto the climatic data from NCEP. Because the TerraClimate data is 604 already bias-corrected, adding future climate projections onto the NCEP data did not require further bias 605 correction. We ran all microclimatic estimations between 2005 and 2015 to match the range of pseudo-606 years available for TerraClimate future climate projections. We did not use a larger range of historical 607 records and only used climate projections available in TerraClimate (i.e., 2°C and 4°C) to reduce 608 computational demands.

609 We then used microclimate estimates to generate hourly operative body temperatures using the *ectotherm* function in NicheMapR⁸³. This modelling system has been extensively validated with field observations^{94–} 610 ⁹⁶ (see also Extended Data Fig. 10). We modelled an adult amphibian in the shape of the leopard frog 611 612 Lithobates pipiens, positioned 1 cm above ground (or 2 m for arboreal species), and assumed that 80% of 613 the skin acted as a free water surface (wet skin). Estimating body mass-specific operative body temperatures 614 for each grid cell, species, and microhabitat was too computationally extensive, given the geographic and 615 taxonomic scale of our study (464,871 local species occurrences). Therefore, we ran the ectotherm models 616 using the median body mass of the species assemblage in each geographical coordinate. When body mass 617 was unknown, we ran models assuming a body mass of 8.4 grams, the median assemblage-level body mass. Given that most amphibians in our dataset are small (median = 1.4 g, mean = 27.5 g), body temperatures 618 619 equilibrate quickly with the environment, and operative body temperatures are likely representative of core 620 body temperatures.

To model operative body temperatures in water bodies (e.g., ponds or wetlands), we used the container
 model from *NicheMapR*. Unlike previously mentioned calculations predicting steady-state temperatures,

this approach accounts for transient temperature changes, capturing lags due to thermal inertia (i.e., 623 transient heat budget model^{97,98}). For pond simulations, we modelled a container permanently filled with 624 625 water (12 m width and 1.5 m-depth) and decreased direct solar radiation to zero to simulate full shade. This 626 modelling approach serves as a proxy for estimating the body temperature of ectotherms submerged in water bodies such as ponds or wetlands, which was validated with field measurements (e.g., 40,95). Ground-627 628 level and water temperatures were modelled for all species regardless of their ecotype (apart from 629 paedomorphic salamanders that were only assessed in aquatic environments) because arboreal and 630 terrestrial species may retreat on land or in water occasionally. Temperatures in above-ground vegetation 631 were only estimated for arboreal and semi-arboreal species as reaching 2 meters height in vegetation 632 requires a morphology adapted to climbing. Our biophysical models assume that shaded microhabitats are 633 available to species throughout their range. While this may not hold true, fine-scaled distribution of these 634 microenvironments are not available at global scales. Moreover, assuming that these microenvironments 635 are available serves a functional role; it provides a best-case scenario that is useful for comparative analyses 636 and offers actionable insights for conservation. For instance, reduced exposure to overheating events in 637 aquatic relative to terrestrial environments would suggest that preserving ponds and wetlands may be 638 critical in buffering the impacts of climate change on amphibians.

639 We then estimated, for each geographical coordinate, the maximum daily body temperature and the mean 640 and maximum weekly maximum body temperature experienced in the 7 days prior to each given day to account for acclimation responses and to assess climate vulnerability metrics¹⁸ (see *Climate vulnerability* 641 analyses). We only used data for the 91 warmest days (i.e., warmest quarter) of each year, as we were 642 interested in the responses of amphibians to extreme heat events¹⁸. Note that data from the year 2005 was 643 644 excluded a posteriori as a burn-in to remove the effects of initial conditions on soil temperature, soil 645 moisture, and pond calculations. Therefore, our analyses are based on 910 days (91 days per year in the 646 range 2006-2015) for each climatic scenario (current climate, 2°C above pre-industrial levels, 4°C above 647 pre-industrial levels).

648 We also used maximum daily body temperatures on terrestrial conditions to calculate the median, 5th 649 percentile and 95th percentile maximum body temperature experienced by each species across their range of distribution. These values were used as acclimation temperatures in the training data to calibrate the data
imputation with ecologically-relevant environmental temperatures (see *Data imputation*); while
maximizing the range of temperatures used to infer the plasticity of heat tolerance limits (see *Climate vulnerability analysis*).

654 Climate vulnerability analysis

655 Using the imputed data, we fitted an individual meta-analytic model for each species to estimate the plasticity of imputed heat tolerance limits (CT_{max}) to changes in operative body temperatures using the 656 *metafor* package⁹⁹ (v. 4.2-0). CT_{max} was used as the response variable, acclimation temperature (i.e., 657 658 median, 5th percentile, or 95th percentile daily maximum body temperature experienced by a species across 659 its distribution range) was used as the predictor variable, and imputed estimates were weighted based on 660 their standard error. From these models, we used out-of-sample model predictions (using the predict function) to estimate the CT_{max} of each species in each 1° x 1° grid cell across their distribution range in 661 662 different warming scenarios, based on predicted mean weekly body temperatures. Specifically, we assumed 663 that species were, on any given day, acclimated to the mean daily body temperature experienced in the 7 days prior¹⁸. Therefore, CT_{max} was simulated as a plastic trait, which varied daily, as animals acclimate to 664 new environmental conditions (Extended Data Fig. 1). While evidence in small amphibians suggests the 665 full acclimation potential is reached within 3-4 days^{100–102}, other evidence points to some variation after 666 667 longer periods¹⁰³. Therefore, we chose 7 days to reflect that some amphibians may require longer to 668 acclimate. Because we used out-of-sample model predictions, we propagated errors from the imputation when estimating the predicted CT_{max} across geographical coordinates. Predicted CT_{max} values and their 669 670 associated standard errors thus reflect variation in both the imputation procedure and the estimation of 671 plastic responses. Our approach to accounting for plasticity assumes that plasticity is homogeneous within 672 species and ignores the possible influence of local adaptation. However, given the low variability in 673 plasticity among species (mean acclimation response ratio \pm s.d. = 0.134 \pm 0.008; range = 0.049 - 0.216; n = 5203), lack of evidence for latitudinal variation in plasticity $(^{28,31,104})$, high phylogenetic signal in thermal 674 tolerance (Pagel's $\lambda^{105} = 0.95$ [0.91 – 0.98]; see Sensitivity Analyses), and evidence for slow rates of 675

evolution and physiological constraints on $CT_{max}^{52,53}$, geographic variation in thermal tolerance and plasticity is unlikely to have a major influence on our results.

678 We then estimated the vulnerability of amphibians to global warming using three metrics (Extended Data 679 Fig. 1). First, we calculated the difference between CT_{max} and the maximum daily body temperature, i.e., 680 the thermal safety margin (i.e., TSM, sensu ⁶). We calculated weighted means and standard errors (sensu 681 106) of thermal safety margins across years to estimate the mean difference between CT_{max} and the maximum 682 temperature during the warmest quarters. Using TSM averaged from the maximum temperature of the warmest quarter is common in the literature^{27–29}. Second, we calculated the number of days the maximum 683 684 daily operative body temperature exceeded CT_{max} across the warmest quarters of 2006-2015, i.e., the 685 number of overheating events. To propagate the uncertainty, we calculated the mean probability that daily 686 operative body temperatures exceeded the predicted distribution of CT_{max} (using the *dnorm* function). Note 687 that the standard error (standard deviation of estimates) of simulated CT_{max} distributions were restricted to 688 one (i.e., simulating distributions within \sim 3°C of the mean) to avoid inflating overheating probabilities due to large imputation uncertainty (cf 75; see also Sensitivity analyses; Extended Data Fig. 8). We then 689 multiplied the mean overheating probability by the total number of simulated days (910) to estimate the 690 691 number of overheating events and their associated standard error using properties of the binomial 692 distribution. Third, we calculated the binary probability (0/1) that species overheat for at least one day 693 across the 910 days surveyed (warmest quarters of 2006-2015). The latter two metrics provide a finer resolution than TSMs, as they capture daily temperature fluctuations and potential overheating events¹⁸. 694

695 *Macroecological patterns*

The objective of this study was to characterise the vulnerability of amphibians to global warming. We investigated patterns at the level of local species occurrences (presence of a given species in a 1° x 1° grid cell based on IUCN data), allowing one to identify specific populations and species that may be more susceptible to heat stress and direct targeted research efforts. We also analysed data at the assemblage level, the species composition within a grid cell. In such case, we calculated the weighted mean and standard error of TSM (*sensu* ¹⁰⁵) across species in each grid cell. Assemblage-level analyses allow one to identify areas containing a higher number of vulnerable species, offering actionable insights for broader-scaleconservation initiatives.

704 We used the gamm4 package¹⁰⁶ to fit generalised additive mixed models (GAMM) against latitude. For 705 local species occurrences, we fitted latitude as a fixed factor, and nested genus and species identity as 706 random terms to account for phylogenetic non-independence. Note that we did not include family as a 707 random term because models failed at estimating higher taxonomic variation. While better methods exist 708 to model phylogenetic patterns, generalised additive linear models do not allow for phylogenetic correlation 709 matrices, and other functions such as brms¹⁰⁷ surpassed our computational time and memory limits. 710 Nevertheless, imputed estimates already reflect variation due to phylogeny (see *Data imputation*), and 711 phylogeny was further modelled when deriving mean estimates in each microhabitat and climatic scenario 712 (see below). We fitted models using the three metrics as response variables independently: the thermal 713 safety margin, overheating risk, and number of overheating events. The former was modelled using a 714 Gaussian distribution of residuals, overheating risk was modelled using a binomial error structure, and the 715 latter using a Poisson error structure. Note that overheating risks were rounded to integer values to fit a 716 Poisson distribution. Thermal safety margin estimates were weighted by the inverse of their sampling 717 variance to account for the uncertainty in the imputation and predictions across geographical coordinates. 718 We fitted separate models for each climatic scenario (current climate, 2°C above preindustrial levels, 4°C 719 above preindustrial levels) and microhabitat (terrestrial, aquatic, arboreal).

720 To investigate the mean TSM in each microhabitat and climatic scenario, we fitted models with the 721 interaction between microhabitat and climatic scenario as a fixed effect using MCMCglmm⁷³ (v. 2.34) and 722 flat, parameter-expanded priors. In these models, we weighted estimates based on the inverse of their 723 sampling variance, species identity was fitted as a random effect, and we accounted for phylogenetic non-724 independence using a variance-covariance matrix of phylogenetic relatedness (calculated from the consensus tree of ²⁶). To investigate the overall overheating risk and number of overheating events in each 725 726 condition, we attempted to fit models in MCMCglmm but these models failed to converge. Therefore, we fitted Poisson and binomial models using *lme4*¹⁰⁸ (v. 1.1-33) and nested genus, species, and observation as 727 728 random terms. We used similar Poisson models to investigate the relationship between the number of

overheating events and thermal safety margins. While the mean estimates from these simpler models should
be unbiased, estimate uncertainty is likely underestimated¹⁰⁹.

We also investigated patterns of climate vulnerability at the assemblage level. We calculated the weighted 731 732 average of TSM and overheating risk in each 1-degree grid cell (14.091; 14.090; or 6,614 grid cells for 733 terrestrial, aquatic, and arboreal species, respectively), and mapped patterns geographically. Averaging 734 overheating risk effectively returned the proportion of species overheating in each coordinate; and we also 735 calculated the number of species overheating in each grid cell. For assemblage-level models, we fitted 736 Gaussian, binomial or Poisson models as described above, but without taxon-level random effects because 737 these cannot be modelled at the assemblage level. All models were fitted without a contrast structure to 738 estimate mean effects in each microhabitat and climatic scenario, and with two-sided contrasts to draw 739 comparisons with current terrestrial conditions.

740 Cross-validation and sensitivity analyses

741 We assessed the accuracy of the data imputation procedure using a cross-validation approach. Specifically, 742 we removed heat tolerance estimates for 5% of the species in the experimental data and 5% of the data-743 deficient species (maintaining the same proportion of missing data) and assessed how well experimental 744 values could be predicted from the models. Of relevance, we only removed data that were comparable to 745 the data that were imputed. That is, data from adult animals tested using a ramping rate of 1°C/min, and 746 where thermal limits were recorded as the onset of spasms. While we could have trimmed any data entry 747 in the experimental data, validation of the imputation performance can only be achieved by comparing 748 comparable entries, and imputing data from species tested in unusual settings would naturally result in large 749 errors. In total, we cross-validated experimental estimates for 77 species.

We investigated alternative ways to i) calculate thermal safety margins, ii) account for acclimation responses, and iii) control for prediction uncertainty (Extended Data Fig. 6-8). In our study, we projected CT_{max} estimates assuming animals were acclimated to the mean weekly temperature experienced prior to each day. We also assessed the climate vulnerability of amphibians assuming they were acclimated to weekly maximum body temperatures (*cf.* ¹⁸), which reflects more conservative estimates (Extended Data

Fig. 7). We also calculated thermal safety margins as the difference between the maximum (or 95th 755 percentile, cf.⁴) hourly body temperature experienced by each population and their predicted CT_{max} to 756 757 investigate the consequences of averaging temperatures when calculating TSMs (Extended Data Fig. 6). To increase the comparability of our estimations with similar studies (e.g., 4), we also calculated climate 758 759 vulnerability metrics more conservatively. Specifically, we excluded temperature data falling below the 5th 760 percentile and above the 95th percentile body temperature for each population to mitigate the impact of 761 outliers (Extended Data Fig. 6). However, extreme weather events, which are typically captured by these outlier values, are the very phenomena most likely to precipitate mortality events^{16,17}. Omitting these 762 outliers could therefore obscure the ecological significance of extreme temperatures, thereby 763 764 underestimating true overheating risks. To estimate overheating probabilities, we calculated the mean daily probability that operative body temperatures exceeded the predicted distribution of CT_{max} and restricted the 765 766 standard deviation of simulated distributions to one (i.e., within ~3°C of the mean) to avoid inflating 767 overheating probability for observations with large uncertainty. We also provided alternative results (Extended Data Fig. 8) where the standard deviation of CT_{max} was restricted to the "biological range", i.e., 768 the standard deviation of the distribution of all CT_{max} estimates across species (range = 1.84 - 2.17). We 769 770 also provide a sensitivity analysis where overheating risk was positive only when the 95% confidence 771 intervals of predicted overheating days did not overlap with zero (Extended Data Fig. 8).

772 We also investigated the influence of different parameters of our biophysical models (i.e., shade and burrow 773 availability, height in above-ground vegetation, solar radiation, wind speed, pond depth) on predicted 774 vulnerability risks (Extended Data Fig. 9). Specifically, we modelled the responses of the species at highest 775 risk in terrestrial and aquatic conditions, Noblella myrmecoides, in its most vulnerable location (latitude, 776 longitude = -9.5, -69.5). For terrestrial conditions, we modelled the response of amphibians with different 777 body sizes (0.5, 4.28, or 50 grams), and with different levels of exposure to open habitat conditions. 778 Specifically, we modelled an amphibian exposed to 50% of shade to simulate an open habitat lightly 779 covered by vegetation, and inferred temperatures at different soil depths (2.5, 5, 10, 15, or 20 cm 780 underground). For aquatic conditions, we adjusted pond depths to simulate a very shallow pond (50 cm) 781 and compared it to deeper ponds (1.5- or 3-meters depth). For arboreal conditions, we modelled the

782 responses of Pristimantis ockendeni, in its most vulnerable location (-4.5, -71.5), and adjusted the height 783 in above-ground vegetation (0.5, 2, or 5 meters), the percentage of radiation diffused by vegetation (50%, 784 75%, or 90% of radiation diffused), and the percentage of wind speed reduced by vegetation (0%, 50%, or 785 80% of wind speed reduced by vegetation). We did not estimate the influence of these parameters on all 786 species and at all locations because of the scale of our study, but these results should provide insight into 787 how varying microenvironmental features and biological characteristics may impact our general 788 conclusions. Our results were generally robust to changes in model parameters, although amphibians are likely to experience more overheating events in open habitats^{6,42} and shallow ponds, and lower risks in 789 underground conditions¹¹¹ (Extended Data Fig. 9). 790

791 We also compared our predictions of operative body temperatures with field body temperature 792 measurements. We extracted night-time (18:00 - 00:30) field body temperatures measured for 11 species 793 of frogs in Mexico (21.48° N, -104.85° W; and 21.45° N, -105.03° W) between June and October of 2013 and 2015 from Table 1 of ¹¹¹. We chose this study because it provided the data and location of body 794 795 temperature measurements, covered multiple species from different sites, and matched our study timeframe 796 (2006-2015). We then compare these estimates with hourly operative body temperatures predicted in shaded 797 terrestrial conditions at the same dates and time windows (Extended Data Fig. 10). We confirmed that 798 predicted operative body temperatures were comparable to field body temperatures measured in some wild 799 frogs (Extended Data Fig. 10), and we invite additional validations with other species in different 800 geographical areas.

Finally, we confirmed the presence of a phylogenetic signal in the experimental dataset by fitting a Bayesian linear mixed model using all complete (no missing data) predictors (i.e., acclimation temperature, endpoint, acclimation status, life stage, and ecotype) in *MCMCglmm*. We accounted for phylogenetic nonindependence using a correlation matrix of phylogenetic relatedness and fitted random intercepts for nonphylogenetic species effects. The phylogenetic signal (Pagel's λ^{105} , which is equivalent to phylogenetic heritability^{112,113}) was calculated as the proportion of variance explained by phylogenetic effects relative to the total non-residual variance.

- 808 Results from all statistical models and additional data visualizations are available at https://p-
- 809 pottier.github.io/Vulnerability_amphibians_global_warming/.
- 810 Data availability
- 811 Raw and processed data are available at <u>https://github.com/p-</u>
- 812 <u>pottier/Vulnerability_amphibians_global_warming</u>, and archived permanently in Zenodo¹¹⁴. Note,
- 813 however, that some intermediate data files were too large to be shared online. These files are available
- 814 upon request. TerraClimate data is available from <u>https://www.climatologylab.org/terraclimate.html</u> and
- 815 NCEP data is available from
- 816 <u>https://psl.noaa.gov/thredds/catalog/Datasets/ncep.reanalysis2/gaussian_grid/catalog.html</u>.
- 817 *Code availability*
- 818 All code needed to reproduce the analyses is available at https://github.com/p-
- 819 <u>pottier/Vulnerability_amphibians_global_warming</u>, and archived permanently in Zenodo¹¹⁴.

821 Methods references

- McNutt, M. K. *et al.* Transparency in authors' contributions and responsibilities to promote
 integrity in scientific publication. *Proc. Natl. Acad. Sci. U.S.A.* 115, 2557–2560 (2018).
- 824 62. Nakagawa, S. *et al.* Method Reporting with Initials for Transparency (MeRIT) promotes more
 825 granularity and accountability for author contributions. *Nat. Commun.* 14, 1788 (2023).
- 826 63. Pottier, P. *et al.* Title, abstract and keywords: a practical guide to maximise the visibility and impact
 827 of academic papers. *Proc. R. Soc. B* 291, 2023.10.02.559861 (2024).
- 828 64. R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical
 829 Computing (2019).
- 830 65. South, A., Michael, S. & Massicotte, P. *rnaturalearthhires*: High resolution world vector map data
 831 from Natural Earth used in rnaturalearth. R package version 0.2.1. (2022).
- 832 66. Yu, G., Smith, D. K., Zhu, H., Guan, Y. & Lam, T. T. Y. ggtree: an R package for visualization and
 833 annotation of phylogenetic trees with their covariates and other associated data. *Methods Ecol. Evol.*834 8, 28–36 (2017).
- 835 67. Wickham, H. ggplot2: Elegant graphics for data analysis. R package version 3.5.1. (2011).
- 836 68. Lutterschmidt, W. I. & Hutchison, V. H. The critical thermal maximum: history and critique. *Can.*837 *J. Zool.* **75**, 1561–1574 (1997).
- 69. IUCN. The IUCN Red List of Threatened Species. https://www.iucnredlist.org (2021).
- 70. Johnson, J. V. *et al.* What drives the evolution of body size in ectotherms? A global analysis across
 the amphibian tree of life. *Glob. Ecol. Biogeogr.* 32, 1311–1322 (2023).
- 841 71. Santini, L., Benítez-López, A., Ficetola, G. F. & Huijbregts, M. A. J. Length-mass allometries in
 842 amphibians. *Integr. Zool.* 13, 36–45 (2018).
- 843 72. Buuren, S. van & Groothuis-Oudshoorn, K. mice: Multivariate Imputation by Chained Equations in
 844 R. *J. Stat. Softw.* 45, 1–67 (2011).
- 845 73. Hadfield, J. D. MCMC Methods for Multi-Response Generalized Linear Mixed Models: The
 846 MCMCglmm R Package. J. Stat. Softw. 33, 1–22 (2010).
- 847 74. Speidel, M., Drechsler, J. & Jolani, S. *R Package Hmi: A Convenient Tool for Hierarchical Multiple*848 *Imputation and Beyond.* https://www.econstor.eu/handle/10419/182156 (2018).
- 75. Callaghan, C. T., Nakagawa, S. & Cornwell, W. K. Global abundance estimates for 9,700 bird
 species. *Proc. Natl. Acad. Sci. U.S.A.* 118, e2023170118 (2021).
- 76. Austin, P. C. & van Buuren, S. The effect of high prevalence of missing data on estimation of the
 coefficients of a logistic regression model when using multiple imputation. *BMC Med. Res. Methodol.* 22, 196 (2022).
- Madley-Dowd, P., Hughes, R., Tilling, K. & Heron, J. The proportion of missing data should not be
 used to guide decisions on multiple imputation. *J. Clin. Epidemiol.* 110, 63–73 (2019).

- 856 78. Lutterschmidt, W. I. & Hutchison, V. H. The critical thermal maximum: data to support the onset of
 857 spasms as the definitive end point. *Can. J. Zool.* **75**, 1553–1560 (1997).
- 858 79. Camacho, A. & Rusch, T. W. Methods and pitfalls of measuring thermal preference and tolerance in
 859 lizards. *J. Therm. Biol.* 68, 63–72 (2017).
- 80. Hoffmann, A. A. & Sgrò, C. M. Comparative studies of critical physiological limits and
 vulnerability to environmental extremes in small ectotherms: How much environmental control is
 needed? *Integr. Zool.* 13, 355–371 (2018).
- 863 81. Lutterschmidt, W. I. & Hutchison, V. H. The critical thermal maximum: data to support the onset of
 864 spasms as the definitive end point. *Can. J. Zool.* **75**, 1553–1560 (1997).
- 865 82. Kearney, M. R. & Porter, W. P. NicheMapR an R package for biophysical modelling: the
 866 microclimate model. *Ecography* 40, 664–674 (2017).
- 867 83. Kearney, M. R. & Porter, W. P. NicheMapR an R package for biophysical modelling: the
 868 ectotherm and Dynamic Energy Budget models. *Ecography* 43, 85–96 (2020).
- 869 84. Pincebourde, S. & Suppo, C. The Vulnerability of Tropical Ectotherms to Warming Is Modulated
 870 by the Microclimatic Heterogeneity. *Integr. Comp. Biol.* 56, 85–97 (2016).
- 871 85. Tracy, C. R., Christian, K. A. & Tracy, C. R. Not just small, wet, and cold: effects of body size and
 872 skin resistance on thermoregulation and arboreality of frogs. *Ecology* 91, 1477–1484 (2010).
- 86. Köhler, A. *et al.* Staying warm or moist? Operative temperature and thermal preferences of
 common frogs (Rana temporaria), and effects on locomotion. *Herpetol. J.* 21, 17–26 (2011).
- 875 87. Navas, C. A., Carvajalino-Fernández, J. M., Saboyá-Acosta, L. P., Rueda-Solano, L. A. &
 876 Carvajalino-Fernández, M. A. The body temperature of active amphibians along a tropical elevation
 877 gradient: patterns of mean and variance and inference from environmental data. *Funct. Ecol.* 27,
 878 1145–1154 (2013).
- 879 88. Barton, M. G., Clusella-Trullas, S. & Terblanche, J. S. Spatial scale, topography and
 880 thermoregulatory behaviour interact when modelling species' thermal niches. *Ecography* 42, 376–
 881 389 (2019).
- 882 89. García-García, A. *et al.* Soil heat extremes can outpace air temperature extremes. *Nat. Clim. Change*883 13, 1237–1241 (2023).
- Bavies-Colley, R. J., Payne, G. W. & van Elswijk, M. Microclimate gradients across a forest edge. *N. Z. J. Ecol.* 24, 111–121 (2000).
- 886 91. Campbell, G. S. & Norman, J. M. *An Introduction to Environmental Biophysics*. (Springer Science
 887 & Business Media, 2000).
- Maclean, I. M. D., Mosedale, J. R. & Bennie, J. J. Microclima: An r package for modelling mesoand microclimate. *Methods Ecol. Evol.* 10, 280–290 (2019).
- 93. Qin, Y. et al. Agricultural risks from changing snowmelt. Nat. Clim. Change 10, 459–465 (2020).
- 891 94. Tracy, C. R. A Model of the Dynamic Exchanges of Water and Energy between a Terrestrial
 892 Amphibian and Its Environment. *Ecol. Monogr.* 46, 293–326 (1976).

- 893 95. Enriquez-Urzelai, U., Kearney, M. R., Nicieza, A. G. & Tingley, R. Integrating mechanistic and
 894 correlative niche models to unravel range-limiting processes in a temperate amphibian. *Glob.*895 *Change Biol.* 25, 2633–2647 (2019).
- 896 96. Kearney, M. R., Munns, S. L., Moore, D., Malishev, M. & Bull, C. M. Field tests of a general
 897 ectotherm niche model show how water can limit lizard activity and distribution. *Ecol. Monogr.* 88,
 898 672–693 (2018).
- 899 97. Kearney, M. R., Porter, W. P. & Huey, R. B. Modelling the joint effects of body size and
 900 microclimate on heat budgets and foraging opportunities of ectotherms. *Methods Ecol. Evol.* 12,
 901 458–467 (2021).
- 902 98. Kearney, M. *et al.* Modelling species distributions without using species distributions: the cane toad
 903 in Australia under current and future climates. *Ecography* 31, 423–434 (2008).
- 904 99. Viechtbauer, W. Conducting meta-analyses in R with the metafor package. J. Stat. Softw. 36, 1–48
 905 (2010).
- 906 100. Brattstrom, B. H. & Lawrence, P. The Rate of Thermal Acclimation in Anuran Amphibians.
 907 *Physiol. Zool.* 35, 148–156 (1962).
- 101. Layne, J. R. & Claussen, D. L. The time courses of CTMax and CTMin acclimation in the
 salamander *Desmognathus fuscus. J. Therm. Biol.* 7, 139–141 (1982).
- 910 102. Turriago, J. L., Tejedo, M., Hoyos, J. M., Camacho, A. & Bernal, M. H. The time course of
 911 acclimation of critical thermal maxima is modulated by the magnitude of temperature change and
 912 thermal daily fluctuations. *J. Therm. Biol.* 114, 103545 (2023).
- 103. Dallas, J. & Warne, R. W. Heat hardening of a larval amphibian is dependent on acclimation period
 and temperature. *J. Exp. Zool. Part A* 339, 339–345 (2023).
- 915 104. Ruthsatz, K. *et al.* Acclimation capacity to global warming of amphibians and freshwater fishes:
 916 Drivers, patterns, and data limitations. *Glob. Change Biol.* **30**, e17318 (2024).
- 917 105. Higgins, J. P. T. & Thompson, S. G. Quantifying heterogeneity in a meta-analysis. *Stat. Med.* 21, 1539–1558 (2002).
- 919 106. Wood, S. & Scheipl, F. gamm4: Generalized additive mixed models using mgcv and lme4. (2014).
- 920 107. Bürkner, P.-C. brms: An R Package for Bayesian Multilevel Models Using Stan. J. Stat. Softw. 80,
 921 1–28 (2017).
- 922 108. Bates, D., Mächler, M., Bolker, B. & Walker, S. Fitting Linear Mixed-Effects Models using lme4.
 923 Preprint at https://doi.org/10.48550/arXiv.1406.5823 (2014).
- 109. Cinar, O., Nakagawa, S. & Viechtbauer, W. Phylogenetic multilevel meta-analysis: A simulation
 study on the importance of modelling the phylogeny. *Methods Ecol. Evol.* 13, 383–395 (2022).
- 926 110. Carlo, M. A., Riddell, E. A., Levy, O. & Sears, M. W. Recurrent sublethal warming reduces
 927 embryonic survival, inhibits juvenile growth, and alters species distribution projections under
- 928 climate change. *Ecol. Lett.* 21, 104–116 (2018).

- 111. Lara-Resendiz, R. A., & Luja, V. H. Body temperatures of some amphibians from Nayarit, Mexico. *Rev. Mex. Biodivers.* 89, 577-581 (2018).
- 931 112. Hadfield, J. D. & Nakagawa, S. General quantitative genetic methods for comparative biology:
 932 phylogenies, taxonomies and multi-trait models for continuous and categorical characters. *J. Evol.*933 *Biol.* 23, 494–508 (2010).
- 113. Lynch, M. Methods for the Analysis of Comparative Data in Evolutionary Biology. *Evolution* 45, 1065–1080 (1991).
- 936 114. Pottier, P. *et al.* Data and code for: Vulnerability of amphibians to global warming. *Zenodo*. version
 937 1.0.0. https://doi.org/10.5281/zenodo.14498866 (2024).
- 115. Ivimey-Cook, E. R. *et al.* Implementing code review in the scientific workflow: Insights from
 ecology and evolutionary biology. *J. Evol. Biol.* 36, 1347–1356 (2023).
- 940 116. Agudelo-Cantero, G. A. & Navas, C. A. Interactive effects of experimental heating rates, ontogeny
 941 and body mass on the upper thermal limits of anuran larvae. *J. Therm. Biol.* 82, 43–51 (2019).
- 942 117. Alveal Riquelme, N. *Relaciones entre la fisiología térmica y las características bioclimáticas de*943 *Rhinella spinulosa (Anura: Bufonidae) en Chile a través del enlace mecanicista de nicho térmico*944 (Universidad de Concepción, 2015).
- 945 118. Alves, M. Tolerância térmica em espécies de anuros neotropicais do gênero Dendropsophus
 946 Fitzinger, 1843 e efeito da temperatura na resposta à predação (Universidade Estadual de Santa
 947 Cruz, 2016).
- 948 119. Anderson, R. C. O. & Andrade, D. V. Trading heat and hops for water: Dehydration effects on
 949 locomotor performance, thermal limits, and thermoregulatory behavior of a terrestrial toad. *Ecol.*950 *Evol.* 7, 9066–9075 (2017).
- 951 120. Aponte Gutiérrez, A. Endurecimiento térmico en Pristimantis medemi (Anura: Craugastoridae), en
 952 coberturas boscosas del Municipio de Villavicencio (Meta) (Universidad Nacional de Colombia,
 953 2020).
- 954 121. Arrigada García, K. Conductas térmica en dos poblaciones de Batrachyla taeniata provenientes de
 955 la localidad de Ucúquer en la región de O'Higgins y de la localidad de Hualpén en la región del
 956 Bío-Bío (Universidad de Concepción, 2019).
- 122. Azambuja, G., Martins, I. K., Franco, J. L. & Santos, T. G. dos. Effects of mancozeb on heat Shock
 protein 70 (HSP70) and its relationship with the thermal physiology of Physalaemus henselii
 (Peters, 1872) tadpoles (Anura: Leptodactylidae). *J. Therm. Biol.* 98, 102911 (2021).
- Bacigalupe, L. D. *et al.* Natural selection on plasticity of thermal traits in a highly seasonal
 environment. *Evol. Appl.* 11, 2004–2013 (2018).
- 962 124. Barria, A. M. & Bacigalupe, L. D. Intraspecific geographic variation in thermal limits and
 963 acclimatory capacity in a wide distributed endemic frog. *J. Therm. Biol.* 69, 254–260 (2017).

- 964 125. Beltrán, I., Ramírez-Castañeda, V., Rodríguez-López, C., Lasso, E. & Amézquita, A. Dealing with
 965 hot rocky environments: critical thermal maxima and locomotor performance in Leptodactylus
 966 lithonaetes (Anura: Leptodactylidae). *Herpetol. J.* 29, 155–161 (2019).
- 967 126. Berkhouse, C. & Fries, J. Critical thermal maxima of juvenile and adult San Marcos salamanders
 968 (Eurycea nana). *Southwest. Nat.* 40, 430–434 (1995).
- 969 127. Blem, C. R., Ragan, C. A. & Scott, L. S. The thermal physiology of two sympatric treefrogs Hyla
 970 cinerea and Hyla chrysoscelis (Anura; Hylidae). *Comp. Biochem. Physiol. A Physiol.* 85, 563–570
 971 (1986).
- 972 128. Bonino, M. F., Cruz, F. B. & Perotti, M. G. Does temperature at local scale explain thermal biology
 973 patterns of temperate tadpoles? *J. Therm. Biol.* 94, (2020).
- 974 129. Bovo, R. P. *Fisiologia térmica e balanço hídrico em anfíbios anuros* (Universidad Estadual
 975 Paulista, 2015).
- 130. Brattstrom, B. H. Thermal acclimation in Australian amphibians. *Comp. Biochem. Physiol.* 35, 69–
 103 (1970).
- 978 131. Brattstrom, B. H. & Regal, P. Rate of thermal acclimation in the Mexican salamander
 979 Chiropterotriton. *Copeia* 1965, 514–515 (1965).
- 980 132. Brattstrom, B. H. A preliminary review of the thermal requirements of amphibians. *Ecology* 44,
 981 238–255 (1963).
- 982 133. Brattstrom, B. H. Thermal acclimation in anuran amphibians as a function of latitude and altitude.
 983 *Comp. Biochem. Physiol.* 24, 93–111 (1968).
- 134. Brattstrom, B. H. & Lawrence, P. The rate of thermal acclimation in anuran amphibians. *Physiol. Zool.* 35, 148–156 (1962).
- 135. Brown, H. A. The heat resistance of some anuran tadpoles (Hylidae and Pelobatidae). *Copeia* 1969, 138 (1969).
- 988 136. Burke, E. M. & Pough, F. H. The role of fatigue in temperature resistance of salamanders. *J. Therm.*989 *Biol.* 1, 163–167 (1976).
- 137. Burrowes, P. A., Navas, C. A., Jiménez-Robles, O., Delgado, P. & De La Riva, I. Climatic
 heterogeneity in the Bolivian Andes: Are frogs trapped? *South Am. J. Herpetol.* 18, 1–12 (2020).
- 138. Bury, R. B. Low thermal tolerances of stream amphibians in the Pacific Northwest: Implications for
 riparian and forest management. *Appl. Herpetol.* 5, 63–74 (2008).
- 139. Castellanos García, L. A. Days of futures past: integrating physiology, microenvironments, and
 biogeographic history to predict response of frogs in neotropical dry-forest to global warming
 (Universidad de los Andes, 2017).
- 140. Castro, B. Influence of environment on thermal ecology of direct-developing frogs (Anura: *Craugastoridae: Pristimantis) in the eastern Andes of Colombia* (Universidad de los Andes, 2019).
- 999 141. Catenazzi, A., Lehr, E. & Vredenburg, V. T. Thermal physiology, disease, and amphibian declines
 1000 on the eastern slopes of the Andes. *Conserv. Biol.* 28, 509–517 (2014).

- 1001 142. Chang, L.-W. *Heat tolerance and its plasticity in larval Bufo bankorensis from different altitudes*1002 (National Cheng Kung University, 2002).
- 1003 143. Chavez Landi, P. A. Fisiología térmica de un depredador Dasythemis sp.(Odonata: Libellulidae) y
 1004 su presa Hypsiboas pellucens (Anura: Hylidae) y sus posibles implicaciones frente al cambio
 1005 climático (Pontificia Universidad Católica Del Ecuador, 2017).
- 1006 144. Chen, T.-C., Kam, Y.-C. & Lin, Y.-S. Thermal physiology and reproductive phenology of Buergeria
 1007 japonica (Rhacophoridae) breeding in a stream and a geothermal hotspring in Taiwan. *Zool. Sci.* 18,
 1008 591–596 (2001).
- 1009 145. Cheng, C.-B. A study of warming tolerance and thermal acclimation capacity of tadpoles in Taiwan
 1010 (Tunghai University, 2017).
- 1011 146. Cheng, Y.-J. *Effect of salinity on the critical thermal maximum of tadpoles living in brackish water*1012 (Tunghai University, 2017).
- 1013 147. Christian, K. A., Nunez, F., Clos, L. & Diaz, L. Thermal relations of some tropical frogs along an
 1014 altitudinal gradient. *Biotropica* 20, 236–239 (1988).
- 1015 148. Claussen, D. L. The thermal relations of the tailed frog, Ascaphus truei, and the Pacific treefrog,
 1016 Hyla regilla. *Comp. Biochem. Physiol. A Physiol.* 44, 137–153 (1973).
- 1017 149. Claussen, D. L. Thermal acclimation in ambystomatid salamanders. *Comp. Biochem. Physiol. A*1018 *Physiol.* 58, 333–340 (1977).
- 1019 150. Contreras Cisneros, J. *Temperatura crítica máxima, tolerancia al frío y termopreferendum del tritón* 1020 *del Montseny (Calotriton arnoldii)* (Universitat de Barcelona, 2019).
- 1021 151. Contreras Oñate, S. Posible efecto de las temperaturas de aclimatación sobre las respuestas
 1022 térmicas en temperaturas críticas máximas (TCmás) y mínimas (TCmín) de una población de
 1023 Batrachyla taeniata (Girard, 1955) (Universidad de Concepción, 2016).
- 1024 152. Cooper, R. D. & Shaffer, H. B. Allele-specific expression and gene regulation help explain
 1025 transgressive thermal tolerance in non-native hybrids of the endangered California tiger salamander
 1026 (Ambystoma californiense). *Mol. Ecol.* **30**, 987–1004 (2021).
- 1027 153. Crow, J. C., Forstner, M. R. J., Ostr, K. G. & Tomasso, J. R. The role of temperature on survival
 1028 and growth of the Barton Springs salamander (Eurycea sosorum). *Herpetol. Conserv. Biol.* 11, 328–
 1029 334 (2016).
- 1030 154. Cupp, P. V. Thermal tolerance of five salientian amphibians during development and
 1031 metamorphosis. *Herpetologica* 36, 234–244 (1980).
- 1032 155. Dabruzzi, T. F., Wygoda, M. L. & Bennett, W. A. Some like it hot: Heat tolerance of the crab-eating
 1033 frog, Fejervarya cancrivora. *Micronesica* 43, 101–106 (2012).
- 1034 156. Dainton, B. H. Heat tolerance and thyroid activity in developing tadpoles and juvenile adults of
 1035 Xenopus laevis (Daudin). *J. Therm. Biol.* 16, 273–276 (1991).
- 1036 157. Daniel, N. J. J. *Impact of climate change on Singapore amphibians* (National University of1037 Singapore, 2013).

- 1038 158. Davies, S. J., McGeoch, M. A. & Clusella-Trullas, S. Plasticity of thermal tolerance and metabolism
 1039 but not water loss in an invasive reed frog. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* 189,
 1040 11–20 (2015).
- 1041 159. de Oliviera Anderson, R. C., Bovo, R. P. & Andrade, D. V. Seasonal variation in the thermal
 1042 biology of a terrestrial toad, Rhinella icterica (Bufonidae), from the Brazilian Atlantic Forest. *J.*1043 *Therm. Biol.* 74, 77–83 (2018).
- 1044 160. de Vlaming, V. L. & Bury, R. B. Thermal selection in tadpoles of the tailed-frog, Ascaphus truei. J.
 1045 *Herpetol.* 4, 179–189 (1970).
- 1046 161. Delson, J. & Whitford, W. G. Critical thermal maxima in several life history stages in desert and
 1047 montane populations of Ambystoma tigrinum. *Herpetologica* 29, 352–355 (1973).
- 1048 162. Duarte, H. *et al.* Can amphibians take the heat? Vulnerability to climate warming in subtropical and
 1049 temperate larval amphibian communities. *Glob. Change Biol.* 18, 412–421 (2012).
- 1050 163. Duarte, H. S. *A comparative study of the thermal tolerance of tadpoles of Iberian anurans*1051 (Universidade de Lisboa, 2011).
- 1052 164. Dunlap, D. Evidence for a daily rhythm of heat resistance in cricket frogs, Acris crepitans. *Copeia*1053 1969, 852–(1969).
- 1054 165. Dunlap, D. G. Critical thermal maximum as a function of temperature of acclimation in two species
 1055 of Hylid frogs. *Physiol. Zool.* 41, 432–439 (1968).
- 1056 166. Elwood, J. R. L. Variation in hsp70 levels and thermotolerance among terrestrial salamanders of
 1057 the Plethodon glutinosus complex (Drexel University, 2003).
- 1058 167. Enriquez-Urzelai, U. *et al.* Ontogenetic reduction in thermal tolerance is not alleviated by earlier
 1059 developmental acclimation in Rana temporaria. *Oecologia* 189, 385–394 (2019).
- 1060 168. Enriquez-Urzelai, U. *et al.* The roles of acclimation and behaviour in buffering climate change
 1061 impacts along elevational gradients. *J. Anim. Ecol.* **89**, 1722–1734 (2020).
- 1062 169. Erskine, D. J. & Hutchison, V. H. Reduced thermal tolerance in an amphibian treated with
 1063 melatonin. *J. Therm. Biol.* 7, 121–123 (1982).
- 1064 170. Escobar Serrano, D. Acclimation scope of the critical thermal limits in Agalychnis spurrelli
 1065 (Hylidae) and Gastrotheca pseustes (Hemiphractidae) and their implications under climate change
- 1066 *scenarios* (Pontificia Universidad Católica Del Ecuador, 2016).
- 1067 171. Fan, X., Lei, H. & Lin, Z. Ontogenetic shifts in selected body temperature and thermal tolerance of
 1068 the tiger frog, Hoplobatrachus chinensis. *Acta Ecol. Sin.* 32, 5574–5580 (2012).
- 1069 172. Fan, X. L., Lin, Z. H. & Scheffers, B. R. Physiological, developmental, and behavioral plasticity in
 1070 response to thermal acclimation. *J. Therm. Biol.* 97, (2021).
- 1071 173. Fernández-Loras, A. *et al.* Infection with Batrachochytrium dendrobatidis lowers heat tolerance of
 1072 tadpole hosts and cannot be cleared by brief exposure to CTmax. *PLoS ONE* 14, (2019).
- 1073 174. Floyd, R. B. Ontogenetic change in the temperature tolerance of larval Bufo marinus (Anura:
- 1074 Bufonidae). Comp. Biochem. Physiol. A Physiol. 75, 267–271 (1983).

- 1075 175. Floyd, R. B. Effects of photoperiod and starvation on the temperature tolerance of larvae of the
 giant toad, Bufo marinus. *Copeia* 1985, 625–631 (1985).
- 1077 176. Fong, S.-T. *Thermal tolerance of adult Asiatic painted frog Kaloula pulchra from different*1078 *populations* (National University of Tainan, 2014).
- 1079 177. Frishkoff, L. O., Hadly, E. A. & Daily, G. C. Thermal niche predicts tolerance to habitat conversion
 1080 in tropical amphibians and reptiles. *Glob. Change Biol.* 21, 3901–3916 (2015).
- 1081 178. Frost, J. S. & Martin, E. W. A comparison of distribution and high temperature tolerance in Bufo
 americanus and Bufo woodhousii fowleri. *Copeia* 1971, 750 (1971).
- 1083 179. Gatz, A. J. Critical thermal maxima of Ambystoma maculatum (Shaw) and Ambystoma
 1084 jeffersonianum (Green) in relation to time of breeding. *Herpetologica* 27, 157–160 (1971).
- 1085 180. Gatz, A. J. Intraspecific variations in critical thermal maxima of Ambystoma maculatum.
 1086 *Herpetologica* 29, 264–268 (1973).
- 1087 181. Geise, W. & Linsenmair, K. E. Adaptations of the reed frog Hyperolius viridiflavus to its arid
 1088 environment IV. Oecological significance of water economy with comments on thermoregulation
 1089 and energy allocation. *Oecologia* 77, 327–338 (1988).
- 1090 182. González-del-Pliego, P. *et al.* Thermal tolerance and the importance of microhabitats for Andean
 1091 frogs in the context of land use and climate change. *J. Anim. Ecol.* **89**, 2451–2460 (2020).
- 1092 183. Gouveia, S. F. *et al.* Climatic niche at physiological and macroecological scales: The thermal
 1093 tolerance–geographical range interface and niche dimensionality. *Glob. Ecol. Biogeogr.* 23, 446–
 1094 456 (2014).
- 1095 184. Gray, R. Lack of physiological differentiation in three color morphs of the cricket frog (Acris
 1096 crepitans) in Illinois. *Trans. Ill. State Acad. Sci.* **70**, 73–79 (1977).
- 1097 185. Greenspan, S. E. *et al.* Infection increases vulnerability to climate change via effects on host thermal
 1098 tolerance. *Sci. Rep.* 7, (2017).
- 1099 186. Guevara-Molina, E. C., Gomes, F. R. & Camacho, A. Effects of dehydration on thermoregulatory
 1100 behavior and thermal tolerance limits of Rana catesbeiana (Shaw, 1802). *J. Therm. Biol.* 93, (2020).
- 1101187. Gutiérrez Pesquera, L. Una valoración macrofisiológica de la vulnerabilidad al calentamiento
- global. Análisis de los límites de tolerancia térmica en comunidades de anfibios en gradientes
 latitudinales y altitudinales (Pontificia Universidad Católica Del Ecuador, 2015).
- 1104 188. Gutiérrez Pesquera, M. *Thermal tolerance across latitudinal and altitudinal gradients in tadpoles*1105 (Universidad de Sevilla, 2016).
- 1106 189. Gutiérrez-Pesquera, L. M. *et al.* Testing the climate variability hypothesis in thermal tolerance
 1107 limits of tropical and temperate tadpoles. *J. Biogeogr.* 43, 1166–1178 (2016).
- 1108 190. Gvoždík, L., Puky, M. & Šugerková, M. Acclimation is beneficial at extreme test temperatures in
 1109 the Danube crested newt, Triturus dobrogicus (Caudata, Salamandridae). *Biol. J. Linn. Soc.* 90,
- 1110
 627–636 (2007).

- 1111 191. Heatwole, H., De Austin, S. B. & Herrero, R. Heat tolerances of tadpoles of two species of tropical
 anurans. *Comp. Biochem. Physiol.* 27, 807–815 (1968).
- 1113 192. Heatwole, H., Mercado, N. & Ortiz, E. Comparison of critical thermal maxima of two species of
 1114 Puerto Rican frogs of the genus Eleutherodactylus. *Physiol. Zool.* 38, 1–8 (1965).
- 1115 193. Holzman, N. & McManus, J. J. Effects of acclimation on metabolic rate and thermal tolerance in the
 1116 carpenter frog, Rana vergatipes. *Comp. Biochem. Physiol. A Physiol.* 45, 833–842 (1973).
- 1117 194. Hoppe, D. M. Thermal tolerance in tadpoles of the chorus frog Pseudacris triseriata. *Herpetologica*1118 34, 318–321 (1978).
- 1119 195. Hou, P.-C. *Thermal tolerance and preference in the adult amphibians from different altitudinal*1120 *LTER sites* (National Cheng Kung University, 2003).
- 1121 196. Howard, J. H., Wallace, R. L. & Stauffer Jr, J. R. Critical thermal maxima in populations of
 1122 Ambystoma macrodactylum from different elevations. *J. Herpetol.* 17, 400–402 (1983).
- 1123 197. Hutchison, V. H. & Ritchart, J. P. Annual cycle of thermal tolerance in the salamander, Necturus
 1124 maculosus. *J. Herpetol.* 23, 73–76 (1989).
- 1125 198. Hutchison, V. H. The distribution and ecology of the cave salamander, Eurycea lucifuga. *Ecol.*1126 *Monogr.* 28, 2–20 (1958).
- 1127 199. Hutchison, V. H. Critical thermal maxima in salamanders. *Physiol. Zool.* **34**, 92–125 (1961).
- 1128 200. Hutchison, V. H., Engbretson, G. & Turney, D. Thermal acclimation and tolerance in the
 1129 hellbender, Cryptobranchus alleganiensis. *Copeia* 1973, 805–807 (1973).
- 1130 201. Hutchison, V. H. & Rowlan, S. D. Thermal acclimation and tolerance in the mudpuppy, Necturus
 1131 maculosus. *J. Herpetol.* 9, 367–368 (1975).
- 1132 202. Jiang, S., Yu, P. & Hu, Q. A study on the critical thermal maxima of five species of salamanders of
 1133 China. *Acta Herpetol. Sin.* 6, 56–62 (1987).
- 1134 203. John-Alder, H. B., Morin, P. J. & Lawler, S. Thermal physiology, phenology, and distribution of
 1135 tree frogs. *Am. Nat.* 132, 506–520 (1988).
- 1136 204. Johnson, C. R. Daily variation in the thermal tolerance of Litoria caerulea (Anura: Hylidae). *Comp.*1137 *Biochem. Physiol. A Physiol.* 40, 1109–1111 (1971).
- 1138 205. Johnson, C. R. Thermal relations and water balance in the day frog, Taudactylus diurnus, from an
 1139 Australian rain forest. *Aust. J. Zool.* 19, 35–39 (1971).
- 1140 206. Johnson, C. R. Diel variation in the thermal tolerance of Litoria gracilenta (Anura: Hylidae). *Comp.*1141 *Biochem. Physiol. A Physiol.* 41, 727–730 (1972).
- 1142 207. Johnson, C. R. & Prine, J. E. The effects of sublethal concentrations of organophosphorus
 1143 insecticides and an insect growth regulator on temperature tolerance in hydrated and dehydrated
 1144 juvenile western toads, Bufo boreas. *Comp. Biochem. Physiol. A Physiol.* 53, 147–149 (1976).
- 1145 208. Johnson, C. R. Observations on body temperatures, critical thermal maxima and tolerance to water
- 1146 loss in the Australian hylid, Hyla caerulea (White). Proc. R. Soc. Qld. 82, 47–50 (1970).

- 1147 209. Johnson, C. R. Thermal relations and daily variation in the thermal tolerance in Bufo marinus. *J.*1148 *Herpetol.* 6, 35 (1972).
- 1149 210. Johnson, C. Thermal relations in some southern and eastern Australian anurans. *Proc. R. Soc. Qld.*1150 82, 87–94 (1971).
- 1151 211. Johnson, C. The effects of five organophosphorus insecticides on thermal stress in tadpoles of the
 1152 Pacific tree frog, Hyla regilla. *Zool. J. Linn. Soc.* 69, 143–147 (1980).
- 1153 212. Katzenberger, M., Duarte, H., Relyea, R., Beltrán, J. F. & Tejedo, M. Variation in upper thermal
 1154 tolerance among 19 species from temperate wetlands. *J. Therm. Biol.* 96, (2021).
- 1155 213. Katzenberger, M. *et al.* Swimming with predators and pesticides: How environmental stressors
 1156 affect the thermal physiology of tadpoles. *PLoS ONE* 9, (2014).
- 1157 214. Katzenberger, M., Hammond, J., Tejedo, M. & Relyea, R. Source of environmental data and
 1158 warming tolerance estimation in six species of North American larval anurans. *J. Therm. Biol.* 76,
 1159 171–178 (2018).
- 1160 215. Katzenberger, M. *Thermal tolerance and sensitivity of amphibian larvae from Palearctic and*1161 *Neotropical communities* (Universidade de Lisboa, 2013).
- 1162 216. Katzenberger, M. Impact of global warming in Holarctic and Neotropical communities of
 1163 amphibians (Universidad de Sevilla, 2014).
- 1164 217. Kern, P., Cramp, R. L. & Franklin, C. E. Temperature and UV-B-insensitive performance in
 1165 tadpoles of the ornate burrowing frog: An ephemeral pond specialist. *J. Exp. Biol.* 217, 1246–1252
 1166 (2014).
- 1167 218. Kern, P., Cramp, R. L., Seebacher, F., Ghanizadeh Kazerouni, E. & Franklin, C. E. Plasticity of
 protective mechanisms only partially explains interactive effects of temperature and UVR on upper
 thermal limits. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* 190, 75–82 (2015).
- 1170 219. Kern, P., Cramp, R. L. & Franklin, C. E. Physiological responses of ectotherms to daily temperature
 1171 variation. *J. Exp. Biol.* 218, 3068–3076 (2015).
- 1172 220. Komaki, S., Igawa, T., Lin, S.-M. & Sumida, M. Salinity and thermal tolerance of Japanese stream
 1173 tree frog (Buergeria japonica) tadpoles from island populations. *Herpetol. J.* 26, 207–211 (2016).
- 1174 221. Komaki, S., Lau, Q. & Igawa, T. Living in a Japanese onsen: Field observations and physiological
 1175 measurements of hot spring amphibian tadpoles, Buergeria japonica. *Amphibia-Reptilia* 37, 311–
 1176 314 (2016).
- 1177 222. Krakauer, T. Tolerance limits of the toad, Bufo marinus, in South Florida. *Comp. Biochem. Physiol.*1178 33, 15–26 (1970).
- 1179 223. Kurabayashi, A. *et al.* Improved transport of the model amphibian, Xenopus tropicalis, and its
 1180 viable temperature for transport. *Curr. Herpetol.* 33, 75–87 (2014).
- 1181 224. Lau, E. T. C., Leung, K. M. Y. & Karraker, N. E. Native amphibian larvae exhibit higher upper
 1182 thermal limits but lower performance than their introduced predator Gambusia affinis. *J. Therm.*
- 1183 *Biol.* **81**, 154–161 (2019).

- 1184 225. Layne, J. R. & Claussen, D. L. Seasonal variation in the thermal acclimation of critical thermal
 1185 maxima (CTMax) and minima (CTMin) in the salamander Eurycea bislineata. *J. Therm. Biol.* 7,
 1186 29–33 (1982).
- 1187 226. Layne, J. R. & Claussen, D. L. The time courses of CTMax and CTMin acclimation in the
 1188 salamander Desmognathus fuscus. *J. Therm. Biol.* 7, 139–141 (1982).
- 1189 227. Lee, P.-T. Acidic effect on tadpoles living in container habitats (Tunghai University, 2019).
- 1190 228. Longhini, L. S., De Almeida Prado, C. P., Bícego, K. C., Zena, L. A. & Gargaglioni, L. H.
- Measuring cardiorespiratory variables on small tadpoles using a non-invasive methodology. *Rev. Cubana Investig. Biomed.* 38, (2019).
- 1193 229. López Rosero, A. C. Ontogenetic variation of thermal tolerance in two anuran species of Ecuador:
 1194 Gastrotheca pseustes (Hemiphractidae) and Smilisca phaeota (Hylidae) and their relative
 1195 vulnerability to environmental temperature change (Pontificia Universidad Católica Del Ecuador,
 1196 2015).
- 1197 230. Lotshaw, D. P. Temperature adaptation and effects of thermal acclimation in Rana sylvatica and
 1198 Rana catesbeiana. *Comp. Biochem. Physiol. A Physiol.* 56, 287–294 (1977).
- 1199 231. Lu, H.-L., Wu, Q., Geng, J. & Dang, W. Swimming performance and thermal resistance of juvenile
 1200 and adult newts acclimated to different temperatures. *Acta Herpetol.* 11, 189–195 (2016).
- 1201 232. Lu, H. L., Geng, J., Xu, W., Ping, J. & Zhang, Y. P. Physiological response and changes in
 1202 swimming performance after thermal acclimation in juvenile Chinese fire-belly newts, Cynops
 1203 orientalis. *Acta Ecol. Sin.* 37, 1603–1610 (2017).
- 1204 233. Lutterschmidt, W. I. & Hutchison, V. H. The critical thermal maximum: Data to support the onset
 1205 of spasms as the definitive end point. *Can. J. Zool.* **75**, 1553–1560 (1997).
- 1206 234. Madalozzo, B. Variação latitudinal nos limites de tolerância e plasticidade térmica em anfíbios em
 1207 um cenário de mudanças climáticas: efeito dos micro-habitats, sazonalidade e filogenia
 1208 (Universidade Federal de Santa Maria, 2018).
- 1209 235. Mahoney, J. J. & Hutchison, V. H. Photoperiod acclimation and 24-hour variations in the critical
 1210 thermal maxima of a tropical and a temperate frog. *Oecologia* 2, 143–161 (1969).
- 1211 236. Maness, J. D. & Hutchison, V. H. Acute adjustment of thermal tolerance in vertebrate ectotherms
 1212 following exposure to critical thermal maxima. *J. Therm. Biol.* 5, 225–233 (1980).
- 1213 237. Manis, M. L. & Claussen, D. L. Environmental and genetic influences on the thermal physiology of
 1214 Rana sylvatica. *J. Therm. Biol.* 11, 31–36 (1986).
- 1215 238. Markle, T. M. & Kozak, K. H. Low acclimation capacity of narrow-ranging thermal specialists
 1216 exposes susceptibility to global climate change. *Ecol. Evol.* 8, 4644–4656 (2018).
- 1217 239. Marshall, E. & Grigg, G. C. Acclimation of CTM, LD50, and rapid loss of acclimation of thermal
 1218 preferendum in tadpoles of Limnodynastes peronii (Anura, Myobatrachidae). *Aust. Zool.* 20, 447–
 1219 456 (1980).

- 1220 240. Mathias, J. H. *The comparative ecologies of two species of Amphibia (B. bufo and B. calamita) on*1221 *the Ainsdale Sand Dunes National Nature Reserve* (The University of Manchester, 1971).
- 1222 241. McManus, J. J. & Nellis, D. W. The critical thermal maximum of the marine toad, Bufo marinus.
 1223 *Caribb. J. Sci.* 15, 67–70 (1975).
- 1224 242. Menke, M. E. & Claussen, D. L. Thermal acclimation and hardening in tadpoles of the bullfrog,
 1225 Rana catesbeiana. *J. Therm. Biol.* 7, 215–219 (1982).
- 1226 243. Merino-Viteri, A. R. *The vulnerability of microhylid frogs, Cophixalus spp., to climate change in*1227 *the Australian Wet Tropics* (James Cook University, 2018).
- 1228 244. Messerman, A. F. *Tales of an 'Invisible' Life Stage: Survival and Physiology Among Terrestrial*1229 *Juvenile Ambystomatid Salamanders* (University of Missouri, 2019).
- 1230 245. Meza-Parral, Y., García-Robledo, C., Pineda, E., Escobar, F. & Donnelly, M. A. Standardized
 1231 ethograms and a device for assessing amphibian thermal responses in a warming world. *J. Therm.*1232 *Biol.* 89, (2020).
- 1233 246. Miller, K. & Packard, G. C. Critical thermal maximum: Ecotypic variation between montane and
 1234 piedmont chorus frogs (Pseudacris triseriata, Hylidae). *Experientia* **30**, 355–356 (1974).
- 1235 247. Miller, K. & Packard, G. C. An altitudinal cline in critical thermal maxima of chorus frogs
 1236 (Pseudacris triseriata). *Am. Nat.* 111, 267–277 (1977).
- 1237 248. Mueller, C. A., Bucsky, J., Korito, L. & Manzanares, S. Immediate and persistent effects of
 1238 temperature on oxygen consumption and thermal tolerance in embryos and larvae of the Baja
 1239 California chorus frog, Pseudacris hypochondriaca. *Front. Physiol.* 10, (2019).
- 1240 249. Navas, C. A., Antoniazzi, M. M., Carvalho, J. E., Suzuki, H. & Jared, C. Physiological basis for
 1241 diurnal activity in dispersing juvenile Bufo granulosus in the Caatinga, a Brazilian semi-arid
 1242 environment. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* 147, 647–657 (2007).
- 1243 250. Navas, C. A., Úbeda, C. A., Logares, R. & Jara, F. G. Thermal tolerances in tadpoles of three
 1244 species of Patagonian anurans. *South Am. J. Herpetol.* 5, 89–96 (2010).
- 1245 251. Nietfeldt, J. W., Jones, S. M., Droge, D. L. & Ballinger, R. E. Rate of thermal acclimation of larval
 1246 Ambystoma tigrinum. *J. Herpetol.* 14, 209–211 (1980).
- 1247 252. Nol, Rosemarie & Ultsch, G. R. The roles of temperature and dissolved oxygen in microhabitat
 1248 selection by the tadpoles of a frog (Rana pipiens) and a toad (Bufo terrestris). *Copeia* 1981, 645–
 1249 652 (1981).
- 1250 253. Novarro, A. J. *Thermal physiology in a widespread lungless salamander* (University of Maryland,
 1251 2018).
- 1252 254. Nowakowski, A. J. *et al.* Thermal biology mediates responses of amphibians and reptiles to habitat
 1253 modification. *Ecol. Lett.* 21, 345–355 (2018).
- 1254 255. Nowakowski, A. J. *et al.* Tropical amphibians in shifting thermal landscapes under land-use and
 1255 climate change. *Conserv. Biol.* **31**, 96–105 (2017).

- 1256 256. Orille, A. C., McWhinnie, R. B., Brady, S. P. & Raffel, T. R. Positive effects of acclimation
 1257 temperature on the critical thermal maxima of Ambystoma mexicanum and Xenopus laevis. *J.*1258 *Herpetol.* 54, 289–292 (2020).
- 1259 257. Oyamaguchi, H. M. *et al.* Thermal sensitivity of a Neotropical amphibian (Engystomops
 1260 pustulosus) and its vulnerability to climate change. *Biotropica* 50, 326–337 (2018).
- 1261 258. Paez Vacas, M. I. *Mechanisms of population divergence along elevational gradients in the tropics*1262 (Colorado State University, 2016).
- 1263 259. Paulson, B. K. & Hutchison, V. H. Blood changes in Bufo cognatus following acute heat stress.
 1264 *Comp. Biochem. Physiol. A Physiol.* 87, 461–466 (1987).
- 1265 260. Paulson, B. & Hutchison, V. Origin of the stimulus for muscular spasms at the critical thermal
 1266 maximum in anurans. *Copeia* 1987, 810–813 (1987).
- 1267 261. Percino-Daniel, R. *et al.* Environmental heterogeneity shapes physiological traits in tropical direct1268 developing frogs. *Ecol. Evol.* (2021).
- 1269 262. Perotti, M. G., Bonino, M. F., Ferraro, D. & Cruz, F. B. How sensitive are temperate tadpoles to
 1270 climate change? The use of thermal physiology and niche model tools to assess vulnerability.
 1271 *Zoology* 127, 95–105 (2018).
- 1272 263. Pintanel, P., Tejedo, M., Almeida-Reinoso, F., Merino-Viteri, A. & Gutiérrez-Pesquera, L. M.
 1273 Critical thermal limits do not vary between wild-caught and captive-bred tadpoles of Agalychnis
 1274 spurrelli (Anura: Hylidae). *Diversity* 12, (2020).
- 1275 264. Pintanel, P., Tejedo, M., Ron, S. R., Llorente, G. A. & Merino-Viteri, A. Elevational and
 1276 microclimatic drivers of thermal tolerance in Andean Pristimantis frogs. *J. Biogeogr.* 46, 1664–
 1277 1675 (2019).
- 1278 265. Pintanel, P. *Thermal adaptation of amphibians in tropical mountains. Consequences of global*1279 *warming* (Universitat de Barcelona, 2018).
- 1280 266. Pintanel, P., Tejedo, M., Salinas-Ivanenko, S., Jervis, P. & Merino-Viteri, A. Predators like it hot:
 1281 Thermal mismatch in a predator-prey system across an elevational tropical gradient. *J. Anim. Ecol.*1282 (2021).
- 1283 267. Pough, F. H. Natural daily temperature acclimation of eastern red efts, Notophthalmus v.
 1284 viridescens (Rafinesque) (Amphibia: Caudata). *Comp. Biochem. Physiol. A Physiol.* 47, 71–78
 1285 (1974).
- 1286 268. Pough, F. H., Stewart, M. M. & Thomas, R. G. Physiological basis of habitat partitioning in
 1287 Jamaican Eleutherodactylus. *Oecologia* 27, 285–293 (1977).
- 269. Quiroga, L. B., Sanabria, E. A., Fornés, M. W., Bustos, D. A. & Tejedo, M. Do sublethal
 concentrations of chlorpyrifos induce changes in the thermal sensitivity and tolerance of anuran
 tadpoles in the toad Rhinella arenarum? *Chemosphere* 219, 671–677 (2019).
- 1291 270. Rausch, C. *The thermal ecology of the Red-spotted toad, Bufo punctatus, across life history*1292 (University of Nevada, 2007).

- 1293 271. Reichenbach, N. & Brophy, T. R. Natural history of the Peaks of Otter salamander (Plethodon
 1294 hubrichti) along an elevational gradient. *Herpetol. Bull.* 7–15 (2017).
- 1295 272. Reider, K. E., Larson, D. J., Barnes, B. M. & Donnelly, M. A. Thermal adaptations to extreme
 1296 freeze-thaw cycles in the high tropical Andes. *Biotropica* 53, 296–306 (2021).
- 1297 273. Richter-Boix, A. *et al.* Local divergence of thermal reaction norms among amphibian populations is
 1298 affected by pond temperature variation. *Evolution* 69, 2210–2226 (2015).
- 1299 274. Riquelme, N. A., Díaz-Páez, H. & Ortiz, J. C. Thermal tolerance in the Andean toad Rhinella
 1300 spinulosa (Anura: Bufonidae) at three sites located along a latitudinal gradient in Chile. *J. Therm.*1301 *Biol.* 60, 237–245 (2016).
- 1302 275. Ritchart, J. P. & Hutchison, V. H. The effects of ATP and cAMP on the thermal tolerance of the
 1303 mudpuppy, Necturus maculosus. *J. Therm. Biol.* 11, 47–51 (1986).
- 1304 276. Rivera-Burgos, A. C. *Habitat suitability for Eleutherodactylus frogs in Puerto Rico: Indexing*1305 *occupancy, abundance and reproduction to climatic and habitat characteristics* (North Carolina
 1306 State University, 2019).
- 1307 277. Rivera-Ordonez, J. M., Nowakowski, A. J., Manansala, A., Thompson, M. E. & Todd, B. D.
 1308 Thermal niche variation among individuals of the poison frog, Oophaga pumilio, in forest and
 1309 converted habitats. *Biotropica* 51, 747–756 (2019).
- 1310 278. Romero Barreto, P. *Requerimientos fisiológicos y microambientales de dos especies de anfibios*1311 (*Scinax ruber e Hyloxalus yasuni*) *del bosque tropical de Yasuní y sus implicaciones ante el cambio*1312 *climático* (Pontificia Universidad Católica Del Ecuador, 2013).
- 1313 279. Ruiz-Aravena, M. *et al.* Impact of global warming at the range margins: Phenotypic plasticity and
 1314 behavioral thermoregulation will buffer an endemic amphibian. *Ecol. Evol.* 4, 4467–4475 (2014).
- 1315 280. Ruthsatz, K. *et al.* Thyroid hormone levels and temperature during development alter thermal
 1316 tolerance and energetics of Xenopus laevis larvae. *Conserv. Physiol.* 6, (2018).
- 1317 281. Ruthsatz, K. *et al.* Post-metamorphic carry-over effects of altered thyroid hormone level and
 1318 developmental temperature: physiological plasticity and body condition at two life stages in Rana
 1319 temporaria. *J. Comp. Physiol. B* 190, 297–315 (2020).
- 1320 282. Rutledge, P. S., Spotila, J. R. & Easton, D. P. Heat hardening in response to two types of heat shock
 1321 in the lungless salamanders Eurycea bislineata and Desmognathus ochrophaeus. *J. Therm. Biol.* 12,
 1322 235–241 (1987).
- 1323 283. Sanabria, E. *et al.* Effect of salinity on locomotor performance and thermal extremes of
 1324 metamorphic Andean Toads (Rhinella spinulosa) from Monte Desert, Argentina. *J. Therm. Biol.* 74,
 1325 195–200 (2018).
- 1326 284. Sanabria, E. A., González, E., Quiroga, L. B. & Tejedo, M. Vulnerability to warming in a desert
 1327 amphibian tadpole community: the role of interpopulational variation. *J. Zool.* 313, 283–296 (2021).

- 1328 285. Sanabria, E. A. & Quiroga, L. B. Change in the thermal biology of tadpoles of Odontophrynus
 1329 occidentalis from the Monte desert, Argentina: Responses to photoperiod. *J. Therm. Biol.* 36, 288–
 1330 291 (2011).
- 1331 286. Sanabria, E. A., Quiroga, L. B., González, E., Moreno, D. & Cataldo, A. Thermal parameters and
 1332 locomotor performance in juvenile of Pleurodema nebulosum (Anura: Leptodactylidae) from the
 1333 Monte Desert. J. Therm. Biol. 38, 390–395 (2013).
- 1334 287. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal changes in the thermal tolerances of the
 1335 toad Rhinella arenarum (Bufonidae) in the Monte Desert of Argentina. *J. Therm. Biol.* 37, 409–412
 1336 (2012).
- 1337 288. Sanabria, E. A., Quiroga, L. B. & Martino, A. L. Seasonal changes in the thermal tolerances of
 1338 Odontophrynus occidentalis (Berg, 1896) (Anura: Cycloramphidae). *Belg. J. Zool.* 143, 23–29
 1339 (2013).
- 1340 289. Sanabria, E. A. *et al.* Thermal ecology of the post-metamorphic Andean toad (Rhinella spinulosa) at
 1341 elevation in the Monte Desert, Argentina. *J. Therm. Biol.* 52, 52–57 (2015).
- 1342 290. Sanabria, E. A., Vaira, M., Quiroga, L. B., Akmentins, M. S. & Pereyra, L. C. Variation of thermal
 1343 parameters in two different color morphs of a diurnal poison toad, Melanophryniscus rubriventris
 1344 (Anura: Bufonidae). *J. Therm. Biol.* 41, 1–5 (2014).
- 1345 291. Sanabria, E. A. & Quiroga, L. B. Thermal parameters changes in males of Rhinella arenarum
 1346 (Anura: Bufonidae) related to reproductive periods. *Rev. Biol. Trop.* 59, 347–353 (2011).
- 1347 292. Scheffers, B. R. *et al.* Thermal buffering of microhabitats is a critical factor mediating warming
 1348 vulnerability of frogs in the Philippine biodiversity hotspot. *Biotropica* 45, 628–635 (2013).
- 1349 293. Scheffers, B. R., Edwards, D. P., Diesmos, A., Williams, S. E. & Evans, T. A. Microhabitats reduce
 1350 animals' exposure to climate extremes. *Glob. Change Biol.* 20, 495–503 (2014).
- 1351 294. Schmid, W. D. High temperature tolerances of Bufo hemiophrys and Bufo cognatus. *Ecology* 46, 559–560 (1965).
- 1353 295. Sealer, J. A. & West, B. W. Critical thermal maxima of some Arkansas salamanders in relation to
 1354 thermal acclimation. *Herpetologica* 25, 122–124 (1969).
- 1355 296. Seibel, R. V. Variables affecting the critical thermal maximum of the leopard frog, Rana pipiens
 1356 Schreber. *Herpetologica* 26, 208–213 (1970).
- 1357 297. Sherman, E. Ontogenetic change in thermal tolerance of the toad Bufo woodhousii fowleri. *Comp.*1358 *Biochem. Physiol. A Physiol.* 65, 227–230 (1980).
- 1359 298. Sherman, E. Thermal biology of newts (Notophthalmus viridescens) chronically infected with a
 1360 naturally occurring pathogen. *J. Therm. Biol.* 33, 27–31 (2008).
- 1361 299. Sherman, E., Baldwin, L., Fernández, G. & Deurell, E. Fever and thermal tolerance in the toad Bufo
 1362 marinus. *J. Therm. Biol.* 16, 297–301 (1991).
- 300. Sherman, E. & Levitis, D. Heat hardening as a function of developmental stage in larval and
 juvenile Bufo americanus and Xenopus laevis. *J. Therm. Biol.* 28, 373–380 (2003).

- 301. Shi, L., Zhao, L., Ma, X. & Ma, X. Selected body temperature and thermal tolerance of tadpoles of
 two frog species (Fejervarya limnocharis and Microhyla ornata) acclimated under different thermal
 conditions. *Acta Ecol. Sin.* 32, 0465–0471 (2012).
- 302. Simon, M. N., Ribeiro, P. L. & Navas, C. A. Upper thermal tolerance plasticity in tropical
 amphibian species from contrasting habitats: Implications for warming impact prediction. *J. Therm. Biol.* 48, 36–44 (2015).
- 1371 303. Simon, M. *Plasticidade fenotípica em relação à temperatura de larvas de Rhinella (Anura:*1372 *Bufonidae) da caatinga e da floresta Atlântica* (Universidade de Sao Paulo, 2010).
- 1373 304. Skelly, D. K. & Freidenburg, L. K. Effects of beaver on the thermal biology of an amphibian. *Ecol.*1374 *Lett.* 3, 483–486 (2000).
- 1375 305. Sos, T. Thermoconformity even in hot small temporary water bodies: a case study in yellow-bellied
 1376 toad (Bombina v. variegata). *Herpetologica Romanica* 1, 1–11 (2007).
- 1377 306. Spotila, J. R. Role of temperature and water in the ecology of lungless salamanders. *Ecol. Monogr.*1378 42, 95–125 (1972).
- 1379 307. Tracy, C. R., Christian, K. A., Betts, G. & Tracy, C. R. Body temperature and resistance to
 1380 evaporative water loss in tropical Australian frogs. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.*1381 150, 102–108 (2008).
- 1382 308. Turriago, J. L., Parra, C. A. & Bernal, M. H. Upper thermal tolerance in anuran embryos and
 1383 tadpoles at constant and variable peak temperatures. *Can. J. Zool.* 93, 267–272 (2015).
- 309. Vidal, M. A., Novoa-Muñoz, F., Werner, E., Torres, C. & Nova, R. Modeling warming predicts a
 physiological threshold for the extinction of the living fossil frog Calyptocephalella gayi. *J. Therm. Biol.* 69, 110–117 (2017).
- 1387 310. von May, R. *et al.* Divergence of thermal physiological traits in terrestrial breeding frogs along a
 1388 tropical elevational gradient. *Ecol. Evol.* 7, 3257–3267 (2017).
- 1389 311. von May, R. *et al.* Thermal physiological traits in tropical lowland amphibians: Vulnerability to
 1390 climate warming and cooling. *PLoS ONE* 14, (2019).
- 1391 312. Wagener, C., Kruger, N. & Measey, J. Progeny of Xenopus laevis from altitudinal extremes display
 1392 adaptive physiological performance. *J. Exp. Biol.* 224, (2021).
- 313. Wang, H. & Wang, L. Thermal adaptation of the common giant toad (Bufo gargarizans) at different
 earlier developmental stages. J. Agric. Univ. Hebei 31, 79–83 (2008).
- 1395 314. Wang, L. The effects of constant and variable thermal acclimation on thermal tolerance of the
 1396 common giant toad tadpoles (Bufo gargarizans). *Acta Ecol. Sin.* 34, 1030–1034 (2014).
- 1397 315. Wang, L.-Z. & Li, X.-C. Effect of temperature on incubation and thermal tolerance of the Chinese
 1398 forest frog. *Chin. J. Zool.* (2007).
- 1399 316. Wang, L. & Li, X.-C. Effects of constant thermal acclimation on thermal tolerance of the Chinese
 1400 forest frog (Rana chensinensis). *Acta Hydrobiol. Sin.* 31, 748–750 (2007).

- 1401 317. Wang, L.-Z., Li, X.-C. & Sun, T. Preferred temperature, avoidance temperature and lethal
 1402 temperature of tadpoles of the common giant toad (Bufo gargarizans) and the Chinese forest frog
 1403 (Rana chensinensis). *Chin. J. Zool.* 40, 23–27 (2005).
- 1404 318. Warburg, M. R. On the water economy of Israel amphibians: The anurans. *Comp. Biochem. Physiol.*1405 *A Physiol.* 40, 911–924 (1971).
- 1406 319. Warburg, M. R. The water economy of Israel amphibians: The urodeles Triturus vittatus (Jenyns)
 1407 and Salamandra salamandra (L.). *Comp. Biochem. Physiol. A Physiol.* 40, 1055–1063 (1971).
- 320. Willhite, C. & Cupp, P. V. Daily rhythms of thermal tolerance in Rana clamitans (Anura: Ranidae)
 tadpoles. *Comp. Biochem. Physiol. A Physiol.* 72, 255–257 (1982).
- 1410 321. Wu, C.-S. & Kam, Y.-C. Thermal tolerance and thermoregulation by Taiwanese rhacophorid
 1411 tadpoles (Buergeria japonica) living in geothermal hot springs and streams. *Herpetologica* 61, 35–
 1412 46 (2005).
- 1413 322. Wu, Q.-H. & Hsieh, C.-H. Thermal tolerance and population genetics of Hynobius fuca. (No
 1414 abbreviation found; *Thermal tolerance...*) *64*, (2016).
- 1415 323. Xu, X. *The effect of temperature on body temperature and thermoregulation in different geographic*1416 *populations of Rana dybowskii* (Harbin Normal University, 2017).
- 1417 324. Yandún Vela, M. C. *Capacidad de aclimatación en renacuajos de dos especies de anuros: Rhinella*1418 *marina (Bufonidae) y Gastrotheca riobambae (Hemiphractidae) y su vulnerabilidad al cambio*1419 *climático* (Pontificia Universidad Católica Del Ecuador, 2017).
- 325. Young, V. K. H. & Gifford, M. E. Limited capacity for acclimation of thermal physiology in a
 salamander, Desmognathus brimleyorum. *J. Comp. Physiol. B* 183, 409–418 (2013).
- 1422 326. Yu, Z., Dickstein, R., Magee, W. E. & Spotila, J. R. Heat shock response in the salamanders
 1423 Plethodon jordani and Plethodon cinereus. *J. Therm. Biol.* 23, 259–265 (1998).
- 1424 327. Zheng, R.-Q. & Liu, C.-T. Giant spiny-frog (Paa spinosa) from different populations differ in
 1425 thermal preference but not in thermal tolerance. *Aquat. Ecol.* 44, 723–729 (2010).
- 1426 328. Zweifel, R. G. Studies on the critical thermal maxima of salamanders. *Ecology* **38**, 64–69 (1957).

1428 Acknowledgements

- 1429 This study was funded by UNSW Scientia Doctoral Scholarships awarded to PPottier, SB, and PPollo. SN
- 1430 was supported by the Australian Research Council (ARC) Discovery Project (DP210100812). SMD was
- supported by the ARC Discovery Early Career Award (DE180100202). MRK was supported by the ARC
- 1432 Discovery Project DP200101279. We thank the authors of the original studies who provided the
- 1433 groundwork for our analyses. We pay our respects to the Bedegal people, the traditional custodians of the
- 1434 land on which this work was primarily conducted.

1435 Authors' contributions

1436 This study was conceptualized by PPottier, MRK, SB, SMD, and SN. All data manipulation and analyses 1437 were performed by PPottier (with conceptual and technical input from SMD and SN for the imputation 1438 methods and statistical analyses, MRK, ARG, JER, and NCW for the biophysical modelling and climate vulnerability analyses). All code was reviewed by NCW, ARG, and JER following the recommendations 1439 1440 of ¹¹⁵. Ecotype information was collected by NCW, PPollo, and ANRV. PPottier, NCW, and SMD 1441 contributed to data visualization. PPottier wrote the initial draft, and all authors were involved in the review 1442 and editing. PPottier oversaw the project administration, while SMD and SN were in charge of the 1443 supervision.

1444 Inclusion & ethics statement

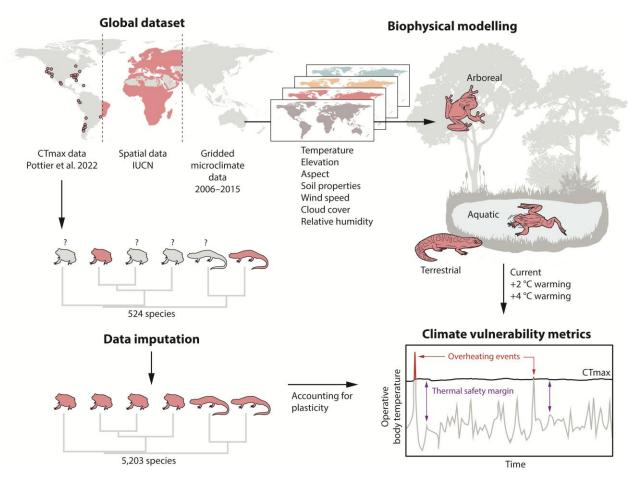
This study did not involve researchers who collected the original data. All data used for the analyses were
taken from a previous data compilation³, and original references on which all analyses were built upon are
listed in the Methods references¹¹⁶⁻³²⁸.

1448 **Competing interest declaration**

1449 The authors declare no conflict or competing interests.

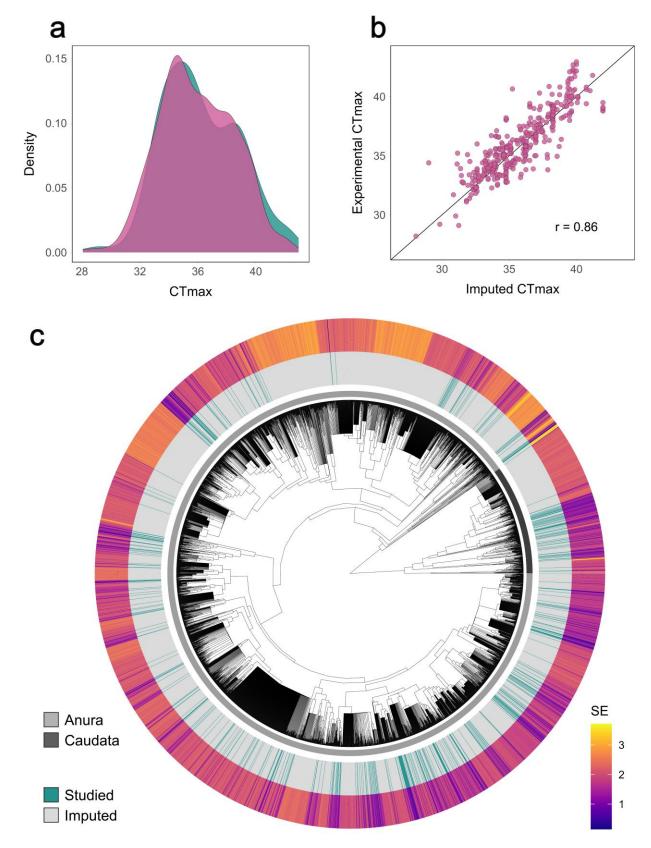
1450 Additional information

Supplementary Information is available for this paper. Correspondence and requests for materials should
be addressed to Patrice Pottier. Reprints and permissions information is available at
www.nature.com/reprints.

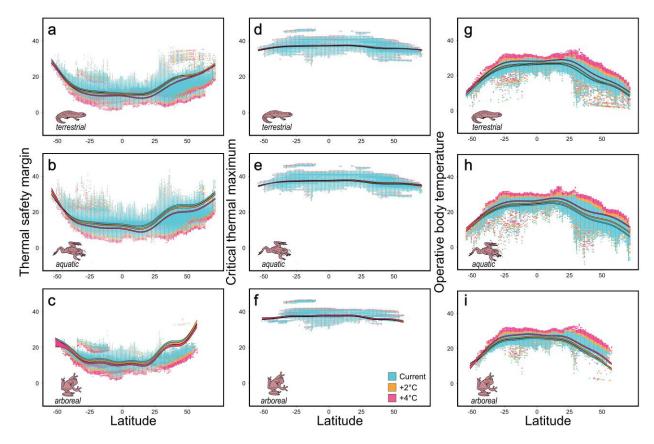


1456 Extended Data Fig. 1 | Conceptual overview of the methods employed to assess the vulnerability of

1457 amphibians to global warming.

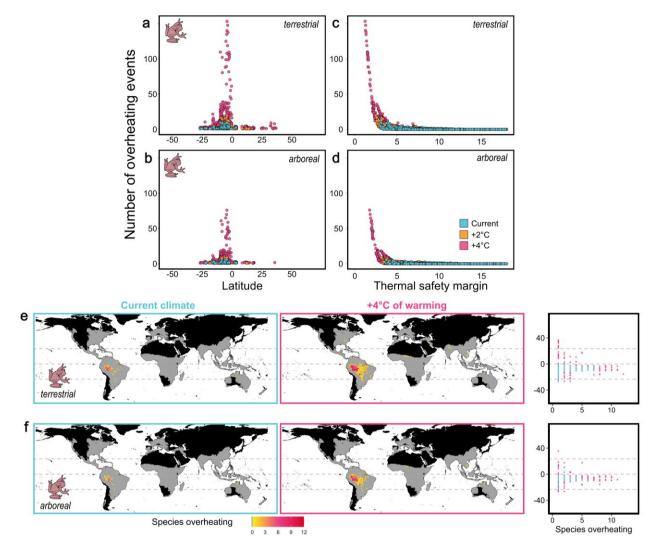


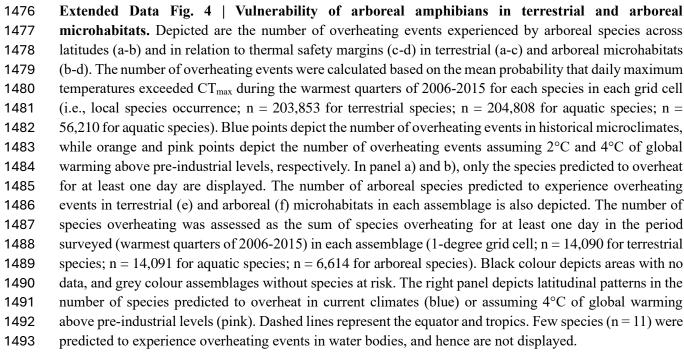
1459Extended Data Fig. 2 | Accuracy of the data imputation procedure. a) Probability density distributions1460 $(n = 375 \text{ observations}, 77 \text{ species}) of experimental CT_{max} (blue) and CT_{max} cross-validated using our data1461imputation procedure (pink). b) Correlation between experimental and imputed CT_{max} values. c) Variation1462in the uncertainty (standard error, SE) of imputed CT_{max} predictions (outer heat map) across studied (blue;1463<math>n = 524$) and imputed (grey; n = 4,679) species.

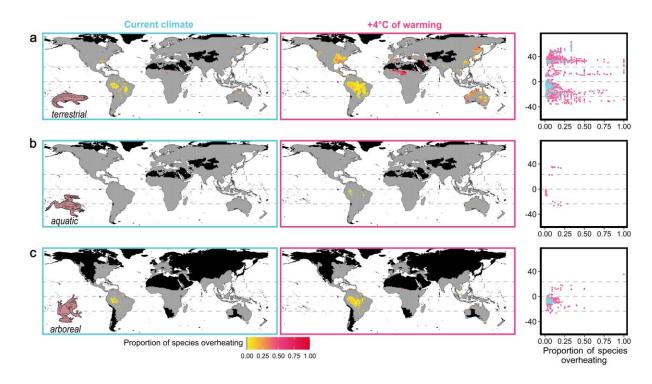




1465 Extended Data Fig. 3 | Thermal safety margin, critical thermal maximum, and operative body 1466 temperatures in different microhabitats and climatic scenarios. Weighted mean thermal safety margins (TSM; a-c), critical thermal maximum (CT_{max}; d-f) and operative body temperatures (g-i) in terrestrial 1467 1468 (a,d,g), aquatic (b,e,h) and arboreal (c,f,i) microhabitats are depicted in current microclimates (blue data 1469 points), or assuming 2°C and 4°C of global warming above pre-industrial levels (orange, and pink data 1470 points, respectively) across latitudes, for each local species occurrence (n = 203,853 for terrestrial species; 1471 n = 204,808 for aquatic species; n = 56,210 for aquatic species). Lines represent 95% confidence intervals of model predictions from generalised additive mixed models. CT_{max} and TSM estimates are scaled by 1472 1473 precision (1/s.e.), with smaller points indicating higher uncertainty. Each point represents a species in a given grid cell. 1474

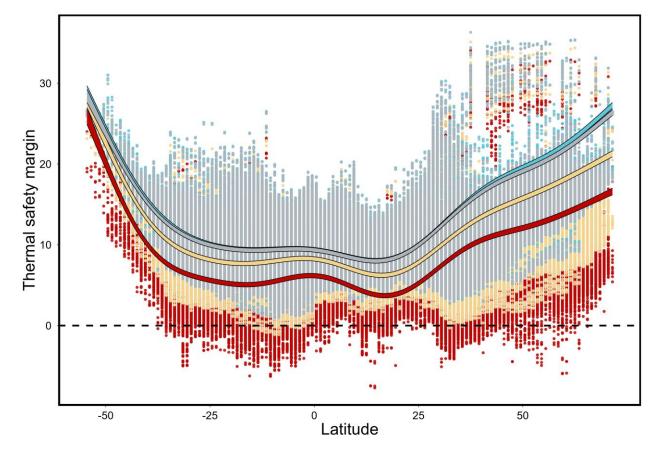






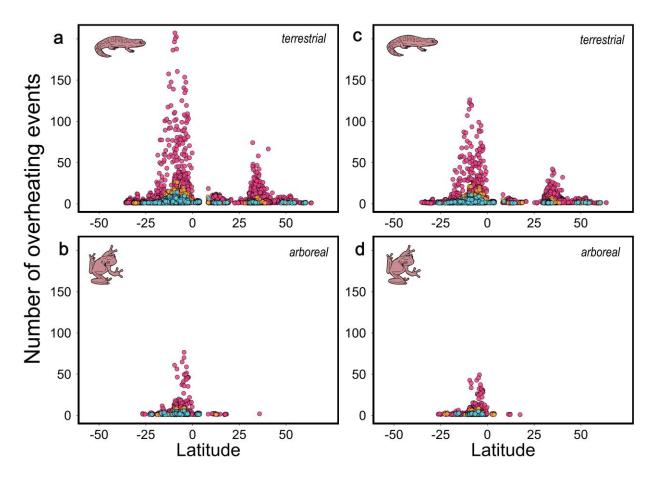
1495 Extended Data Fig. 5 | Proportion of species predicted to experience overheating events in terrestrial

1496	(a), aquatic (b), and arboreal (c) microhabitats. The proportion of species overheating was assessed as
1497	the sum of species overheating for at least one day in the period surveyed (warmest quarters of 2006-2015)
1498	divided by the number of species in each assemblage (1-degree grid cell; $n = 14,090$ for terrestrial species;
1499	n = 14,091 for aquatic species; $n = 6,614$ for arboreal species). Black colour depicts areas with no data, and
1500	grey colour assemblages without species at risk. The right panel depicts latitudinal patterns in the proportion
1501	of species predicted to overheat in current climates (blue) or assuming 4°C of global warming above pre-
1502	industrial levels (pink). Dashed lines represent the equator and tropics.



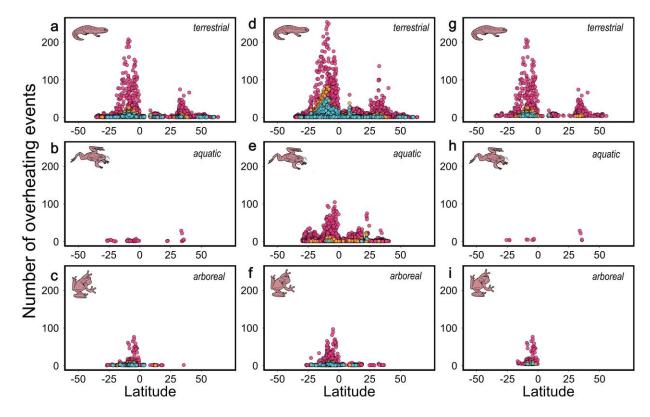


1504 Extended Data Fig. 6 | Variation in thermal safety margins calculated using different assumptions. 1505 Thermal safety margins (TSM) were calculated as the mean difference between CT_{max} and the predicted 1506 operative body temperature in full shade during the warmest quarters of 2006-2015 (grey), as the mean 1507 difference between CT_{max} and the predicted operative body temperature in full shade during the warmest 1508 quarters of 2006-2015 excluding body temperatures falling outside the 5% and 95% percentile temperatures (blue), as the difference between the 95% percentile operative body temperature and the corresponding 1509 CT_{max} (yellow), or as the difference between the maximum operative body temperature and the 1510 1511 corresponding CT_{max} (red). Lines represented 95% confidence interval ranges predicted from generalized 1512 additive mixed models. This figure was constructed assuming ground-level microclimates occurring under 1513 4°C of global warming above pre-industrial levels, for each species in each grid cell (i.e., local species 1514 occurrences; n = 203,853 for terrestrial species; n = 204,808 for aquatic species; n = 56,210 for aquatic 1515 species).



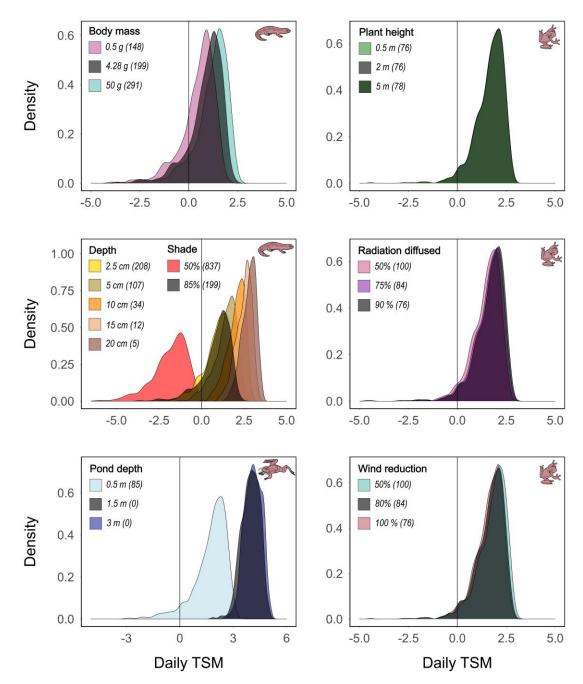


1517 Extended Data Fig. 7 | Latitudinal variation in the number of overheating events when animals are 1518 acclimated to the mean (a,b) or maximum (c,d) weekly body temperature experienced in the seven 1519 days prior in terrestrial (a,c) and arboreal (b,d) microhabitats. The number of overheating events (days) 1520 were calculated based on the mean probability that daily maximum temperatures exceeded CT_{max} during the warmest quarters of 2006-2015 for each species in each grid cell (i.e., local species occurrences; n = 1521 1522 203,853 for terrestrial species; n = 204,808 for aquatic species; n = 56,210 for aquatic species). Blue points depict the number of overheating events in historical microclimates, while orange and pink points depict 1523 1524 the number of overheating events assuming 2°C and 4°C of global warming above pre-industrial levels, 1525 respectively. For clarity, only the species predicted to experience overheating events across latitudes are 1526 depicted.



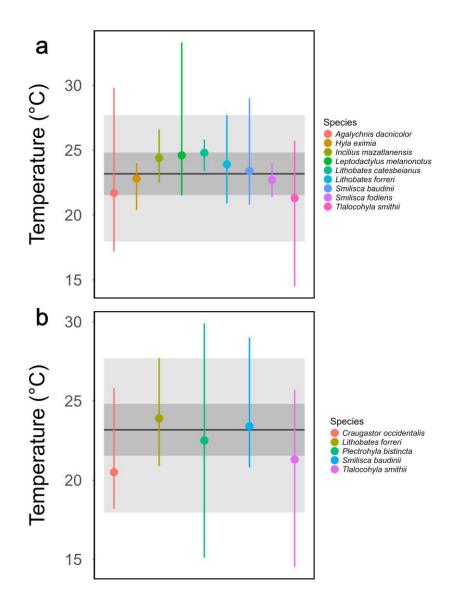


1528 Extended Data Fig. 8 | Latitudinal variation in the number of overheating events using regular (a,b,c), 1529 uncertain (d,e,f), or conservative estimates (g,h,i) in terrestrial (a,d,g), aquatic (b,e,h) and arboreal 1530 (c,f,i) microhabitats. The number of overheating events (days) were calculated based on the mean 1531 probability that daily maximum temperatures exceeded CT_{max} during the warmest quarters of 2006-2015 1532 for each species in each grid cell (i.e., local species occurrences; n = 203,853 for terrestrial species; n =1533 204,808 for aquatic species; n = 56,210 for aquatic species). Uncertain estimates are those where daily 1534 overheating probabilities were calculated based on broad predicted distributions of CT_{max} (i.e., simulated over the whole "biological range"), likely inflating overheating probabilities for observations with large 1535 1536 uncertainty. Conservative estimates are those when overheating risk was considered only when the 95% 1537 confidence intervals of the predicted number of overheating events did not overlap with zero (e,f). Blue 1538 points depict the number of overheating events in historical microclimates, while orange and pink points 1539 depict the number of overheating events assuming 2°C and 4°C of global warming above pre-industrial 1540 levels, respectively. For clarity, only the species predicted to experience overheating events across latitudes 1541 are depicted.





1543 Extended Data Fig. 9 | Influence of biophysical model parameters on the estimation of terrestrial 1544 (a,b), aquatic (c), and arboreal (d,e,f) thermal safety margins. Depicted is the variation in daily thermal 1545 safety margins (TSM) as density distributions according to body mass (a), shade availability and soil depth 1546 (b), pond depth (c), height of the animal in above-ground vegetation (d), percentage of solar radiation 1547 diffused by vegetation (e), and percentage of wind reduced by vegetation (f). All simulations were 1548 performed assuming 4°C of global warming above pre-industrial levels in a specific grid cell (latitude, 1549 longitude = -9.5, -69.5; where the highest number of overheating events was predicted), for the most 1550 vulnerable species (Noblella myrmecoides in terrestrial and aquatic microhabitats, Pristimantis ockendeni 1551 in arboreal microhabitats). Negative daily TSMs were recorded as overheating events, and conditions 1552 depicted in dark grey reflect the results presented in the manuscript. The number of predicted overheating 1553 events is indicated in brackets for each condition (n = 910 days).



Extended Data Fig. 10 | Validation of operative body temperature estimations. Terrestrial operative 1557 body temperatures estimated from biophysical models were compared to field body temperatures recorded 1558 around Tepic (21.48° N, -104.85° W; n = 11 species; panel a) and El Cuarenteño (21.45° N, -105.03° W; n 1559 = 5 species; panel b) between June and October of 2013/2015, for 11 species of frogs¹¹¹. The mean hourly 1560 operative body temperatures predicted from our models for the same date and time windows (18:00 - 01:00)1561 1562 are represented by the black horizontal line, along with their standard deviation (dark grey box), and range (light grey box). The mean (point) and range (bars) of field body temperatures recorded for each species 1563 1564 are presented in colour. Note that our analyses were based on the maximum daily temperature recorded at 1565 each site during the warmest quarters of 2006-2015, which may not match the times and dates at which 1566 field body temperatures were recorded. Nevertheless, congruence between night-time predicted and field body temperatures suggests our models are likely to capture true biological variation in operative body 1567 1568 temperatures throughout the day.

Vulnerability of amphibians to global warming

- 2 Patrice Pottier^{1,2*}, Michael R. Kearney³, Nicholas C. Wu⁴, Alex R. Gunderson⁵, Julie E. Rej⁵, A.
- 3 Nayelli Rivera-Villanueva^{6,7}, Pietro Pollo¹, Samantha Burke¹, Szymon M. Drobniak^{1,8+}, and Shinichi
- 4 Nakagawa^{1,9+}
- 5
- ¹ Evolution & Ecology Research Centre, School of Biological, Earth and Environmental Sciences,
 University of New South Wales, Sydney, New South Wales, Australia.
- ² Division of Ecology and Evolution, Research School of Biology, The Australian National
 University, Canberra, Australian Capital Territory, Australia
- 10 ³ School of BioSciences, The University of Melbourne, Melbourne, Victoria, Australia
- ⁴Hawkesbury Institute for the Environment, Western Sydney University, Richmond, New South
 Wales, Australia
- 13 ⁵ Department of Ecology and Evolutionary Biology, Tulane University, New Orleans, Louisiana, USA
- ⁶ Centro Interdisciplinario de Investigación para el Desarrollo Integral Regional Unidad Durango
- 15 (CIIDIR), Instituto Politécnico Nacional, Durango, México
- 16 ⁷ Laboratorio de Biología de la Conservación y Desarrollo Sostenible de la Facultad de Ciencias
- 17 Biológicas, Universidad Autónoma de Nuevo León, Monterrey, México
- 18 ⁸ Institute of Environmental Sciences, Jagiellonian University, Kraków, Poland.
- ⁹ Department of Biological Sciences, University of Alberta, Edmonton, Alberta, Canada.
- 20 *Corresponding author
- 21 ⁺These authors supervised the work equally
- 22
- 23 Corresponding author: Patrice Pottier (<u>p.pottier@unsw.edu.au</u>)
- 24

Table of contents26

27	Supplementary Tables	
28	Table S1	
29	Table S2	4
30	Table S3	5
31	Table S4	6
32	Table S5	7
33		
34		

35 Supplementary tables

Table S1 | Statistical model estimates for thermal safety margins calculated for local species
occurrences and assemblages Model estimates for each microhabitat (terrestrial, arboreal, aquatic)
and each climatic scenario (current, +2°C, or +4°C of global warming above pre-industrial levels) are
depicted. No contrast structure was used in the presented models. mean: mean model estimate; CI.lb:
lower bound of the 95% confidence interval; CI.ub: upper bound of the 95% confidence interval; p: pvalue; k_{sp}: number of species; k_{obs}: number of observations; Var_{sp}: variance explained by differences
between species; Var_{phy}: variance explained by shared evolutionary history; Var_{obs}: residual variance.

	Local species patterns in thermal safety margin								
	mean	CI.lb	CI.ub	р	k _{sp}	kobs	Var _{sp}	Var _{phy}	Var _{obs}
Terrestrial (current)	11.694	8.856	14.428	< 0.001	5177	203853			
Terrestrial (+2°C)	10.914	8.025	13.594	< 0.001	5177	203853			
Terrestrial (+4°C)	9.409	6.530	12.090	< 0.001	5177	203853			
Arboreal (current)	12.235	9.402	14.960	< 0.001	1771	56210			
Arboreal (+2°C)	11.517	8.660	14.236	< 0.001	1771	56210	1.295	11.960	1.828
Arboreal (+4°C)	10.073	7.229	12.797	< 0.001	1771	56210			
Aquatic (current)	13.598	10.708	16.276	< 0.001	5203	204808			
Aquatic (+2°C)	12.827	8.796	14.361	< 0.001	5203	204808			
Aquatic (+4°C)	11.682	8.796	14.361	< 0.001	5203	204808			

Assemblage-level patterns in	in thermal safety margin
------------------------------	--------------------------

			-	_		
	mean	CI.lb	CI.ub	р	k _{obs}	Var _{obs}
Terrestrial (current)	15.279	15.208	15.330	< 0.001	14090	
Terrestrial (+2°C)	14.328	14.279	14.396	< 0.001	14090	
Terrestrial (+4°C)	12.602	12.542	12.657	< 0.001	14090	
Arboreal (current)	14.279	14.191	14.381	< 0.001	6614	11.06
Arboreal (+2°C)	13.393	13.298	13.478	< 0.001	6614	
Arboreal (+4°C)	11.746	11.666	11.830	< 0.001	6614	
Aquatic (current)	17.408	17.352	17.471	< 0.001	14091	
Aquatic (+2°C)	16.528	16.468	16.581	< 0.001	14091	
Aquatic (+4°C)	15.287	15.225	15.346	< 0.001	14091	

46	Table S2 Statistical model estimates for overheating risk and the number of overheating events.
47	Model estimates for each microhabitat (terrestrial, arboreal) and each climatic scenario (current, +2°C,
48	or +4°C of global warming above pre-industrial levels) are depicted. The estimated number of
49	overheating events in species predicted to experience at least one overheating event (i.e., overheating
50	species) are also depicted. Model estimates for aquatic microhabitats are not displayed because no
51	species was predicted to experience overheating events in this microhabitat. No contrast structure was
52	used in the presented models. mean: mean model estimate; CI.lb: lower bound of the 95% confidence
53	interval; CI.ub: upper bound of the 95% confidence interval; p: p-value; k _{sp} : number of genera; k _{sp} :
54	number of species; kobs: number of observations; Vargenus: variance explained by differences between
55	genera; Var _{sp} : variance explained by differences between species; Var _{obs} : residual variance.

			01	verheati	ng risk					
	mean	CI.lb	CI.ub	р	kgenus	ksp	kobs	Vargenus	Var _{sp}	
Terrestrial (current)	9.98 x 10 ⁻⁷	5.60 x 10 ⁻⁷	1.78 x 10 ⁻⁶	< 0.001	464	5177	203853			
Terrestrial (+2°C)	1.93 x 10 ⁻⁶	1.09 x 10 ⁻⁶	3.43 x 10 ⁻⁶	< 0.001	464	5177	203853			
Terrestrial (+4°C)	9.09 x 10 ⁻⁶	5.13 x 10 ⁻⁶	1.61 x 10 ⁻⁵	< 0.001	464	5177	203853	0.306	69.653	
Arboreal (current)	4.77 x 10 ⁻⁷	2.58 x 10 ⁻⁷	8.80 x 10 ⁻⁷	< 0.001	174	1771	56210	0.300	09.055	
Arboreal (+2°C)	9.78 x 10 ⁻⁷	5.45 x 10 ⁻⁷	1.75 x 10 ⁻⁶	< 0.001	174	1771	56210			
Arboreal (+4°C)	3.72 x 10 ⁻⁶	2.08 x 10 ⁻⁶	6.67 x 10 ⁻⁶	< 0.001	174	1771	56210			
		Num	ber of over	heating	events	s (all sj	pecies)			
	mean	CI.lb	CI.ub	р	kgenus	ksp	kobs	Vargenus	Var _{sp}	
Terrestrial (current)	0.014	0.001	0.080	< 0.001	464	5177	203853			
Terrestrial (+2°C)	0.025	0.002	0.127	< 0.001	464	5177	203853) 52.500	
Terrestrial (+4°C)	0.153	0.046	0.460	< 0.001	464	5177	203853	0.110		
Arboreal (current)	0.008	0.001	0.043	< 0.001	174	1771	56210	0.110		
Arboreal (+2°C)	0.015	0.001	0.083	< 0.001	174	1771	56210			
Arboreal (+4°C)	0.076	0.012	0.230	< 0.001	174	1771	56210			
	N	umber of o	verheating	events ((among	overh	eating sp	pecies)		
	mean	CI.lb	CI.ub	р	kgenus	ksp	kobs	Vargenus	Var _{sp}	Varobs
Terrestrial (current)	2.155	0.239	5.264	< 0.001	38	104	836			
Terrestrial (+2°C)	2.576	0.410	5.857	< 0.001	61	168	1424			
Terrestrial (+4°C)	6.747	3.136	11.385	< 0.001	118	391	4248	0 252	0.187	0.210
Arboreal (current)	1.621	0.026	4.429	< 0.001	4	13	152	0.253	0.10/	0.310
Arboreal (+2°C)	1.956	0.113	4.973	< 0.001	5	16	283			
Arboreal (+4°C)	5.084	1.806	9.387	< 0.001	17	56	748			

57	Table S3 Statistical model estimates for the number of species predicted to experience
58	overheating events. Model estimates for each microhabitat (terrestrial, arboreal) and each climatic
59	scenario (current, +2°C, or +4°C of global warming above pre-industrial levels) are depicted. The
60	estimated number of species overheating in assemblages containing at least one species predicted to
61	experience at least one overheating event (i.e., overheating assemblages) are also depicted. Model
62	estimates for aquatic microhabitats are not displayed because no species was predicted to experience
63	overheating events in this microhabitat. No contrast structure was used in the presented models. mean:
64	mean model estimate; CI.lb: lower bound of the 95% confidence interval; CI.ub: upper bound of the
65	95% confidence interval; p: p-value; kobs: number of observations; Varobs: residual variance.

	Number of species overheating (all assemblages)								
-	mean	CI.lb	CI.ub	р	kobs	Varobs			
Terrestrial (current)	0.056	0.016	0.118	< 0.001	14090				
Terrestrial (+2°C)	0.096	0.029	0.199	< 0.001	14090				
Terrestrial (+4°C)	0.288	0.083	0.604	< 0.001	14090	55.47			
Arboreal (current)	0.021	0.002	0.054	< 0.001	6614	55.47			
Arboreal (+2°C)	0.040	0.006	0.094	< 0.001	6614				
Arboreal (+4°C)	0.107	0.021	0.243	< 0.001	6614				

Number of species overheating (among overheating assemblages)

-	mean	CI.lb	CI.ub	р	kobs	Var _{obs}
Terrestrial (current)	3.185	0.601	6.883	< 0.001	253	
Terrestrial (+2°C)	3.228	0.678	6.810	< 0.001	426	
Terrestrial (+4°C)	3.084	0.617	6.557	< 0.001	1328	0.601
Arboreal (current)	1.930	0.054	5.054	< 0.001	74	0.601
Arboreal (+2°C)	2.445	0.189	5.649	< 0.001	111	
Arboreal (+4°C)	2.509	0.312	5.692	< 0.001	285	

66

68	Table S4 Statistical model estimates for the proportion of species predicted to experience
69	overheating events. Model estimates for each microhabitat (terrestrial, arboreal) and each climatic
70	scenario (current, +2°C, or +4°C of global warming above pre-industrial levels) are depicted. The
71	estimated proportion of species overheating in assemblages containing at least one species predicted to
72	experience at least one overheating event (i.e., overheating assemblages) are also depicted. Model
73	estimates for aquatic microhabitats are not displayed because no species was predicted to experience
74	overheating events in this microhabitat. No contrast structure was used in the presented models. mean:
75	mean model estimate; CI.lb: lower bound of the 95% confidence interval; CI.ub: upper bound of the
76	95% confidence interval; p: p-value; kobs: number of observations; Varobs: residual variance.

	Proportion of species overheating (all assemblages)									
-	mean	CI.lb	CI.ub	р	kobs	Varobs				
Terrestrial (current)	1.22 x 10 ⁻⁵	8.96 x 10 ⁻⁶	1.66 x 10 ⁻⁵	< 0.001	14090					
Terrestrial (+2°C)	2.09 x 10 ⁻⁵	1.60 x 10 ⁻⁵	2.72 x 10 ⁻⁵	< 0.001	14090					
Terrestrial (+4°C)	8.13 x 10 ⁻⁵	6.60 x 10 ⁻⁵	1.00 x 10 ⁻⁴	< 0.001	14090	42.26				
Arboreal (current)	1.19 x 10 ⁻⁵	7.07 x 10 ⁻⁶	2.02 x 10 ⁻⁵	< 0.001	6614	42.20				
Arboreal (+2°C)	1.86 x 10 ⁻⁵	1.19 x 10 ⁻⁵	2.89 x 10 ⁻⁵	< 0.001	6614					
Arboreal (+4°C)	4.99 x 10 ⁻⁵	3.62 x 10 ⁻⁵	6.87 x 10 ⁻⁵	< 0.001	6614					

Proportion of species overheating (among overheating assemblages)

	mean	CI.lb	CI.ub	р	kobs	Varobs	
Terrestrial (current)	0.053	0.046	0.061	< 0.001	253		
Terrestrial (+2°C)	0.058	0.052	0.065	< 0.001	426	1.019	
Terrestrial (+4°C)	0.094	0.088	0.100	< 0.001	1328		
Arboreal (current)	0.038	0.029	0.050	< 0.001	74		
Arboreal (+2°C)	0.054	0.043	0.067	< 0.001	111		
Arboreal (+4°C)	0.061	0.053	0.070	< 0.001	285		

77

79 Table S5 | Statistical model estimates for the association between the number of overheating 80 events and thermal safety margins. Model estimates for each microhabitat (terrestrial, arboreal) and each climatic scenario (current, +2°C, or +4°C of global warming above pre-industrial levels) are 81 depicted. Model estimates for aquatic microhabitats are not displayed because no species was predicted 82 to experience overheating events in this microhabitat. All model estimates are on the log scale. Separate 83 84 models were fitted for each microhabitat and climatic scenario. mean: mean model estimate; se: 85 standard error; p: p-value; k_{sp}: number of genera; k_{sp}: number of species; k_{obs}: number of observations; Vargenus: variance explained by differences between genera; Varsp: variance explained by differences 86 between species; Var_{obs}: residual variance. 87

	mean	se	р	kgenus	k _{sp}	kobs	Var _{genus}	Var _{sp}	Varobs
Terrestrial (current)									
Intercept	3.723	0.390	< 0.001	464	5177	203853	5.850	3.346	0.116
Slope (TSM)	-1.201	0.031	< 0.001						
Terrestrial (+2°C)									
Intercept	6.318	0.310	< 0.001	464	5177	203853	5.272	2.380	0.078
Slope (TSM)	-1.452	0.027	< 0.001						
<i>Terrestrial (+4°C)</i>									
Intercept	7.611	0.171	< 0.001	464	5177	203853	2.954	1.025	0.248
Slope (TSM)	-1.616	0.015	< 0.001						
Arboreal (current)									
Intercept	4.929	1.091	< 0.001	174	1771	56210	0.001	15.190	0.001
Slope (TSM)	-1.511	0.094	< 0.001						
Arboreal (+2°C)									
Intercept	7.836	0.836	< 0.001	174	1771	56210	4.359	2.358	0.001
Slope (TSM)	-1.739	0.080	< 0.001						
Arboreal (+4°C)									
Intercept	10.093	0.587	< 0.001	174	1771	56210	8.789	0.917	0.001
Slope (TSM)	-2.085	0.039	< 0.001						

88