

1 The genetic basis for social network traits in the wild.

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14

15 **Abstract**

16 Sociality is fundamental for many species, linked to an individual's survival and reproductive
17 success. The evolution of social behaviours has been studied, especially in complex social
18 animals such as birds. However, discerning the genetic basis of these behaviours has been
19 difficult due to the lack of ecological validity and life-history in controlled environments. In
20 wild populations, however, there are the challenges of getting accurate pedigree and
21 behavioural data. This paper used data from a long-term wild sparrow population in which
22 we tracked the social behaviour and genetic pedigree, to assemble social networks. Here,
23 we estimated the genetic architecture of three social network metrics—degree, strength, and
24 betweenness—as a method of characterising social behaviour. The results show a clear
25 genetic component, with heritability values of 8%, 6%, and 1.7% for strength, degree, and
26 betweenness respectively. This study expands our understanding of how sociality is
27 inherited from controlled settings to wild populations, showing a link between genetics and

28 social behaviour. The finding of a negative genetic correlation between strength and degree
29 indicates a more complex genetic relationship underlying these social traits, calling for more
30 research to understand the ecological and evolutionary impacts of these findings. Our results
31 clearly show a genetic basis for social interactions in sparrows, inviting more research into
32 sociality in other bird and non-bird species.

33

34

35 **Introduction**

36 The largely unexplored relationship between fitness, genetics, and social behaviour has
37 garnered significant interest within the field of evolutionary biology (Bleakley et al., 2010,
38 Lehmann & Rousset, 2014). Fitness, in evolutionary terms, refers to an individual's ability to
39 survive and reproduce within a given environment (Orr, 2009). An individual's genetic
40 makeup can influence trait variations that may be beneficial or detrimental to fitness in
41 particular environments (Dunning *et al.*, 2023). The transmission of these traits from one
42 generation to the next is central to the conception of evolution and the basis of the study of
43 genetics (Ellegren & Sheldon 2008).

44 Social behaviour, generally defined as interactions among individuals of the same species,
45 significantly affects an individual's fitness, impacting their ability to find food, mates, and
46 avoid predators (Riehl & Strong, 2018, Dunning et al., 2023). For instance, cooperative
47 behaviours can enhance survival rates by aiding in predator avoidance or resource
48 acquisition (Cornwallis et al., 2017). On the other hand, aggressive or competitive
49 behaviours might hinder an individual's fitness if they lead to injury or social isolation
50 (Arseneau-Robar et al., 2016, Szípl et al., 2019). This implies that social behaviour offer
51 evolutionary advantages and thus should have gone through selection. There is evidence of
52 intraspecific variation in social behaviour across many species, with consistent behavioural
53 differences among individuals (Bell et al., 2009, Aplin et al., 2015, Plaza et l., 2019).

54 If there is selection for these behaviours, then, if they are also heritable they can evolve.
55 Determining the genetic basis of social behaviour is pivotal, because it contributes to our
56 understanding of the evolution of sociality, and it can elucidate how social behaviours
57 contribute to an individual's fitness and, by extension, the evolutionary trajectory of species
58 (Wice & Saltz, 2021, Hardie & Cooney, 2022). Moreover, knowing the genetic architecture of
59 behaviour traits has practical applications for breeding and conservation efforts (Blumstein,
60 2010).

61 While some studies have explored the genetic basis of social behaviour in mice and
62 invertebrates, these investigations have largely been conducted in controlled laboratory
63 environments (Hammock & Young, 2005, Wice & Saltz, 2021). It is inherently difficult and
64 expensive to obtain complete and accurate pedigree data of freely roaming, and potentially
65 intermingling populations.. While laboratory experiments have their merits, they however
66 cannot replicate real-world situations and often do not represent the genetic diversity of wild
67 populations (Calisi & Bentley, 2009). Furthermore, laboratory studies cannot incorporate
68 population processes and life-history data (Pemberton, 2008). As a result, we lack data on
69 the genetic architecture of sociality from wild populations.

70 To address this knowledge gap, this paper investigates the heritability of sociality in wild
71 House sparrows (*Passer domesticus*) living on an isolated island off the coast of the
72 UK. Here, migration to and from the island is minimal, allowing for following all individuals
73 throughout their lives (Schroeder *et al.*, 2011). The project's uniqueness lies in its opportunity
74 to study the heritability of social behaviour in a wild population, which is only possible due to
75 the availability of near-complete census and long-term nature of the study (Schroeder *et al.*,
76 2011).

77 In this study, we use social network analysis (farine whitehead), to study and quantify
78 sociality . Metrics calculated using social network analysis have been evidenced as a good
79 proxy for personality traits in House sparrows, specifically in the population on Lundy Island

80 (Plaza *et al.*, 2019, Farine & Whitehead, 2015). This paper hypothesises that these traits are
81 heritable in House sparrows.

82

83 **Methods**

84 Data collection

85 The data for the study was obtained from a long-term investigation conducted on Lundy
86 Island (51°10'N, 4°40'W), located in the Bristol Channel. The study species was the House
87 sparrow, a commonly used model organism known for its extensively studied biology and life
88 history (Hansen *et al.*, 2020). House sparrows are known to form social flocks during non-
89 breeding periods and gather around food sources, making them an ideal species for
90 studying social behaviour (Tuliozi *et al.*, 2018).

91 Being located 19km off the North Devon coast limits migration, together with a near-
92 complete, annual census and breeding monitoring, enabled us to build a comprehensive
93 database of life-history traits of every individual in the population (Schroeder *et al.*, 2011,
94 Schroeder *et al.*, 2015). DNA samples were collected from the sparrows after capturing birds
95 in their natal nest box, or through mist netting. These DNA samples, along with regular nest
96 monitoring, are used to build the genetic pedigree, which was constructed using <22
97 microsatellite markers (for details see Ockendon *et al.*, 2009, Schroeder *et al.*, 2012 and
98 Dunning *et al.*, 2023). At the time of writing, the pedigree spans 20 years, from 2000-2019.

99

100 Each sparrow is equipped with a distinctive combination of coloured rings and a Passive-
101 Integrated Transponder (PIT tag, TROVANID100, for details see Schroeder *et al.*, 2011).

102 A Radio Frequency Identification (RFID) antenna measuring 19.8 x 19.8cm (DorsetID) was
103 affixed to a seed reservoir positioned that is regularly visited by the sparrows. The antenna
104 recorded instances of a tagged sparrow entering the feeder, as described in Dunning *et al.*

105 (2023). The reservoir design enables multiple birds to feed simultaneously, and PIT tags are
106 recorded every $\frac{1}{4}$ of a second. .

107

108 Social network analysis

109 Social network analysis is a process that quantifies the connections and interactions
110 between individuals within a social group, represented as nodes (individuals) and edges
111 (connections/relationships) in a network graph (Otte & Rousseau, 2002). We created social
112 networks using presence data obtained from RFID readings during the winter non-breeding
113 (NB) period of the specific year. We chose this approach to account for seasonal variations
114 in social behaviour, as sparrows exhibit territorial behaviour during breeding seasons (Tuliozi
115 *et al.*, 2018). Data during the NB period was collected for 2013, 2015-16, 2016-17, and
116 2020, resulting in a total of 37 weeks of data in the final dataset.

117 We used the arrival time method to define non-random associations between individuals,
118 where an overlap in arrival time at the bird feeder (Δt) is defined as an association (Dunning
119 *et al.*, 2023). This method is ideal for House sparrows, as other common methods tend to
120 overestimate associations for gregarious species that aggregate around resources (Dunning
121 *et al.*, 2023, Ferreira *et al.*, 2020). This method was also developed specifically on the Lundy
122 Island sparrow population and their behaviour at feeders (Dunning *et al.*, 2023). The arrival
123 time method assumes that individuals who arrive at the feeder together are socially
124 associated and were likely together before arriving (Dunning *et al.*, 2023b).

125 We conducted the computation of social networks using the arrival method in R and RStudio
126 (R Core Team, 2023, RStudio team, 2020) with a custom function, openly available in the
127 Zenodo repository (Chan and Dunning, 2023). The function incorporates two parameters: Δt ,
128 representing a predefined time interval that starts when an individual's arrival at the feeder,
129 and Δi , the time after which the individual is considered to have left the feeder. By assigning
130 Δt to each arrival, the method considers only birds arriving within one another's Δt as
131 associated, regardless of their duration at the feeder (Dunning *et al.*, 2023). Individuals could
132 only be recorded again after the Δi period. Δt and Δi were assigned fixed values of 150s and

133 300s, respectively, as these intervals are most effective at capturing non-random
134 associations of individuals who arrive in a group (Dunning *et al.*, 2023).
135 We processed the data using the arrival time functions to construct weighted, non-directional
136 social networks using the R package igraph (Csardi & Nepusz, 2006). To incorporate a
137 greater amount of data into the wild animal model, we built a social network for each week.
138 We used three node-based metrics (hereafter centrality measures), degree, strength and
139 betweenness as measurements of sociality. Degree quantifies the number of associations
140 an individual has with others and could be interpreted as how many friends an individual
141 has. Strength further considers the intensity or frequency of those relationships, or how close
142 an individual is to their friends. Betweenness is the number of shortest paths between other
143 individuals in the group that pass through the focal individual, quantifying how central an
144 individual is to the group (Sosa *et al.*, 2020).
145 We computed degree, strength, betweenness and density with the igraph package (Csardi &
146 Nepusz, 2006). Density, a global network metric referring to the ratio of associations an
147 individual has over the number of potential associations it could have, was also calculated,
148 as its use during modelling enables better comparison between different social networks
149 (Hart, 2023).

150

151 Animal model

152 We utilized an animal model to quantify quantitative genetic parameters. Specifically, a
153 generalised linear model using Markov Chain Monte Carlo (MCMC) approximation methods
154 was used, with the MCMCglmm package in R (Hadfield *et al.*, 2010).

155 Animal models are a form of mixed-effect linear regression model, leveraging genetic
156 information to estimate genetic correlations between traits as additive genetic variance
157 (Wilson *et al.*, 2010). Additive genetic variation is the measurable variance in a trait due to
158 the combined effect of shared, multiple alleles (Kruuk *et al.*, 2014). In the animal model, the
159 additive genetic effect is treated as a random effect by incorporating the inverse relatedness
160 matrix of the pedigree into the regression. This matrix mathematically portrays the extent of

161 genetic relatedness between individuals in a population, quantifying genetic relationships
162 based on the pedigree.

163 The additive genetic effect is assumed to have a variance of V_A (Kruuk *et al.*, 2014). Any
164 residual variance in phenotype is labelled as V_R , culminating in a total phenotypic variance of
165 $V_A + V_R$ (Eq. 1, Dingemanse & Dochtermann, 2014), with V_P being the total phenotypic
166 variance.

167

168 Eq 1.
$$V_P = V_A + V_R$$

169

170 The pedigree data obtained from the Lundy sparrow research team was processed to
171 include only the columns of offspring, dam, and sire. A proportion of dams and sires were
172 not present as offspring; these were added as offspring with parents labelled as NA. 10
173 individuals in the social data were not present in the pedigree, and excluded from the
174 dataset, resulting in a total of 544 informative individuals and 816 observations.

175 A tri-variate MCMCglmm was used as the animal model. Degree, strength and betweenness
176 were collectively modelled as the response variable. The inverse relatedness matrix of the
177 pedigree, denoted as “animal” in MCMCglmm, was modelled as a random effect. Bird ID
178 was also included as a random effect to account for permanent environment effects
179 (differences between individuals not driven by additive genetic effects). Additionally, the
180 network metric density was introduced as a fixed effect to improve model fit. Degree and
181 betweenness were z-transformed to standardise the model output and improve model
182 convergence. Strength was not scaled since it has a range of 0-1. All three centrality
183 measures were modelled under a gaussian distribution. The modelling approach for
184 pedigree and residuals was unrestricted to capture any potential covariance between traits
185 arising from relatedness and to explore potential interactions among these traits within
186 individuals. Priors were set as a matrix, with ones in the diagonal and zeroes elsewhere and
187 given a minimal effect on posterior probability ($\nu = 0.002$). 1,000,000 iterations, a burn-in of
188 300,000, and a thinning interval of ten were used as the parameters for the tri-variate model.

189

190 Analysis

191 For each random effect outcome within the animal model, a variance-covariance (VCV)
192 matrix was generated. The total phenotypic variance V_P was calculated as the sum of these
193 VCV matrices (Eq. 2, Wilson et al., 2010). The VCV matrix for permanent environment
194 effects (V_{PE}) was represented as variance among individuals (modelled as random effect
195 bird ID, a unique identifier of the birds).

196

197 Eq. 2
$$V_P = V_A + V_{PE} + V_R$$

198

199 - V_{PE} = Permanent environment effects

200

201 These VCV matrices were used to calculate the narrow sense heritability (h^2) and
202 repeatability of each network trait. h^2 quantifies the proportion of the variation in a trait within
203 a population that is influenced by additive genetic factors (Wilson et al., 2010). h^2 is
204 calculated as the fraction of phenotypic variance attributable to additive genetic variance,
205 which can be represented as a percentage (Eq. 3).

206

207 Eq. 3
$$h^2 = \frac{V_A}{V_P}$$

208

209 Repeatability refers to the fraction of phenotypic variance that remains consistent across
210 multiple observations in an individual and indicates the stability of a trait (Wilson et al.,
211 2010). It is often regarded as the upper limit of h^2 , as it represents the total phenotypic
212 variance observed among individuals, which includes additive genetic variance.

213 Repeatability is calculated as the sum of overall individual variance V_{ind} over total phenotypic
214 variance of a trait (Eq. 4, Dingemanse & Dochtermann, 2014).

215

216 Eq. 4 $Repeatability = \frac{V_{ind}}{V_P} = \frac{V_A + V_{PE}}{V_P}$

217

218

219 Moreover, to assess potential trait covariation, the phenotypic covariance between centrality
220 measures resulting from additive genetic effects (COV_A) was used to compute genetic
221 covariance (Wilson et al., 2010). We then computed the genetic correlation from the genetic
222 covariance, denoted as r_A (Eq. 5, Åkesson et al., 2008), where $COV_{A(x,y)}$ is the additive
223 genetic covariance between trait x and y.

224

225 Eq. 5 $r_A = \frac{COV_{A(x,y)}}{\sqrt{V_{A(x)}} \sqrt{V_{A(y)}}}$

226

227

228 **Results**

229 Significant additive genetic variance was observed for all three centrality traits (Table 1). The
230 covariance between traits due to pedigree was statistically significant for strength and
231 degree (-0.0077, 95% confidence interval = -0.0149 – -0.0003).

232 Variance due to permanent environment was also statistically significant for all three traits
233 (Table 1). Additionally, the residual variance and the covariance between traits was
234 significant for all three traits. The MCMCglmm model and its variables met all assumptions,
235 and diagnostic plots indicated convergence and independent sampling. The deviance
236 information criterion was estimated to be 3,621. MCMC model outputs are given in Appendix
237 1.

238

239 *Table 1. Variance-covariances matrices of the animal model. 95% confidence intervals are*
240 *indicated within the brackets. Genetic and phenotypic covariances are given above the*
241 *diagonal, while correlations are provided below. The diagonal displays additive genetic*
242 *variance first, followed by percentage variance.*

Additive genetic variance			
	Degree	Strength	Betweenness
Degree	0.0628 (0.0054 - 0.1226), 2.42% (0.21% - 4.72%)	-0.0077 (-0.0149 - -0.0003)	0
Strength	-0.0254 (-0.0492 - -0.0009)	0.0028 (0.0007 - 0.0051), 0.11% (0.03% - 0.20%)	0
Betweenness	0	0	0.0174 (0.0017 - 0.0399), 0.67% (0.06 - 1.53)
Permanent environment effects			
Degree	0.0194 (0.0002 - 0.0691), 0.75% (0.01% - 2.66%)	NA	NA
Strength	NA	0.0021 (0.0003 - 0.0042), 0.08% (0.01% - 0.16%)	NA
Betweenness	NA	NA	0.0067 (0.0002 - 0.2043), 0.26% (0.01% - 0.79%)
Residual variance			
Degree	0.9621 (0.8597 - 1.0663), 37.05% (33.11% - 41.06%)	0.1145 (0.0964 - 0.1326)	0.3265 (0.2543 - 0.4025)

Strength	0.0992 (0.0835 – 0.1149)	0.0301 (0.0263 – 0.0337), 1.15% (1.01% - 1.31%)	0.0507 (0.0375 – 0.0636)
Betweenness	0.1644 (0.1281 – 0.2027)	0.043 (0.0319 – 0.0540)	1.0097 (0.9120 – 1.1120), 38.88% (35.12% - 42.82%)

243

244

245 Heritability of degree, strength, and betweenness were determined to be 6.01%, 7.97% and
 246 1.68% respectively (Table 2). Genetic correlation between degree and strength was -0.8169
 247 (Table 2). As the covariance between degree and betweenness and betweenness and
 248 degree were not significant they were assumed to be 0.

249

250 *Table 2. Heritability, Repeatability and Total Phenotypic Variance (Total) Values, along with*
 251 *Genetic Correlation between Centrality Measures.*

	Heritability	Repeatability	Total
Degree	0.0601	0.0795	1.0442
Strength	0.0797	0.0818	0.0349
Betweenness	0.0168	0.0235	1.0338
Genetic correlation			
Degree – Strength	-0.8169		
Degree - Betweenness	0		
Strength - Betweenness	0		

252

253

254 **Discussion**

255 The results of this study provide compelling evidence for a genetic basis underlying social
256 centrality traits in sparrows, as all three centrality measures showed significant variance
257 attributed to pedigree, showing that more closely related individuals had more similar trait
258 measurements. This finding supports the idea that social behaviour in sparrows is heritable,
259 since social centrality measures are reliable proxies for social behaviour (Plaza *et al.*, 2019).
260 The repeatability estimates provide strong support for the existence of consistent individual
261 differences in social behaviour over time, as evidenced by significant and robust variation
262 between birds.

263 Strength appeared to be the most significantly influenced by pedigree, with a heritability
264 estimate of 8% (Table 2). On the other hand, betweenness was least affected by pedigree,
265 with a heritability estimate of 1.7%. These findings could suggest that the quality and
266 quantity of social interactions has a stronger genetic basis than the centrality of an individual
267 in its population. Furthermore, heritability made up a significant proportion of repeatability.
268 The heritability of betweenness made up 70.83% of the repeatability of betweenness, whilst
269 heritability of strength was 97.56% of the repeatability. This suggests that most of the
270 variability in social network traits could be attributed to relatedness. Further research and
271 exploration of the underlying genetic mechanisms could reveal the intriguing interplay
272 between genetics and social behaviour.

273

274 An intriguing observation was the strong negative genetic correlation (-0.8169) between
275 strength and degree (Table 2). This indicates that degree and strength co-vary in sparrow
276 populations, and that individuals with more, but weaker connections, tend to be related to
277 sparrows with fewer, but stronger associations. This might also suggest the presence of
278 pleiotropy, where a single genetic locus influences both strength and degree, or a potential
279 genetic linkage between the underlying genes for these two traits. This discovery merits
280 further investigation to explore the underlying mechanisms and potential ecological and
281 evolutionary implications.

282 While the variance attributed to pedigree was found to be statistically significant, the effect
283 sizes for degree and betweenness were relatively small, measuring only 0.0628 and 0.0174
284 standard deviations from the mean, respectively (Table 1). The genetic variance for strength
285 was also small (0.0028). The relatively wide confidence intervals, ranging from
286 approximately 0.0017 to 0.1226 for all variance estimates, further underscore the modest
287 effect sizes observed. Although the genetic influence on social centrality traits is significant,
288 caution should be exercised in interpretations, and further validation is warranted. Moreover,
289 the residuals exhibited large variances, surpassing the variation attributed to pedigree and
290 individual differences. These significant residuals, which additionally had tighter confidence
291 intervals, suggest the existence of additional unexplored factors that significantly influence
292 centrality measures beyond those considered in the current model.

293 To address this, alternative models with different variables could be used to find other
294 factors that influence social centrality measures, which could reduce residual variance and
295 improve model fit. For example, including week as a random effect. When collecting a new
296 dataset for social research, the age and sex of birds should be recorded and used since
297 individuals of different ages and gender may interact differently. Additionally, social networks
298 could be constructed on different time intervals, for example per month, or with intervals of a
299 constant sample size, to determine if these factors change the findings found in this study.

300

301 Given more time, a null model employing randomized data sets could be run to validate the
302 observed results (Farine, 2017). As the confidence intervals are relatively wide, assessing
303 whether null results yield different confidence intervals would add further robustness to the
304 significant findings.

305 Another aspect to consider is whether the variation in social traits we've observed is due to
306 genetic factors or simply a result of closely related individuals sharing similar environments.
307 This question requires more exploration, which could involve methods like cross-fostering or
308 using data that contains extra-pair offspring. Furthermore, conducting models using data

309 collected during breeding seasons could offer valuable insights into the significance of social
310 behaviour in different contexts.

311 In conclusion, despite the small impact and relatively large confidence intervals, there is
312 compelling evidence that sociality is heritable in-house sparrows, adding valuable insights
313 into the genetic basis of social behaviour. This finding warrants further investigation to
314 understand the underlying mechanisms driving social behaviour.

315

316 **Statements and Declarations**

317

318 Data Archiving Statement

319 The data used in the study was provided by the Lundy Sparrow project, a long-term research
320 initiative carried out by the Schroeder laboratory, run by Julia Schroeder, at Imperial College
321 London, in collaboration with Sheffield University and Macquarie University. The data was
322 largely cleaned before it was accessed for this study. Data sets will be available upon
323 acceptance of the manuscript. The R script can be accessed at
324 <https://github.com/JKHunter315/GeneticBasisSocialitySparrowsICL>. The arrival time method
325 of constructing social networks was developed by Jamie Dunning and Alex Chan, from the
326 Schroeder laboratory at Imperial College London, available publicly on Zenodo at
327 <https://zenodo.org/records/7527440> (Chan & Dunning, 2023).

328

329 Competing interests

330 The author declares no competing interests.

331

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487

488 **Appendix**

489

490 *Appendix 1. Summary statistics of each random effect. Covariances of interactions are*
491 *displayed as trait x – trait y. The values in the table are scaled by a factor of 100 to*
492 *accommodate the small effect sizes of the output. Significant values are denoted by *.*

Summary Statistics			
Pedigree			
	Mean variance	95% confidence interval	Effective sample size
Degree	6.28*	0.54 – 12.26	243.40
Strength	0.28*	0.07 – 0.51	338.20
Betweenness	1.74*	0.17 – 3.99	180.00
Degree - Betweenness	-2.26	-4.89 – 0.10	295.70
Degree - Strength	-0.77*	-1.49 – -0.03	433.80

Strength - Betweenness	0.46	-0.17 – 3.99	180.00
Bird ID			
	Mean variance	95% confidence interval	Effective sample size
Degree	1.94*	0.02 – 6.91	1,030.00
Strength	0.21*	0.03 – 0.42	1,112.00
Betweenness	0.67*	0.02 – 2.04	3,225.00
Residuals			
	Mean variance	95% confidence interval	Effective sample size
Degree	96.21*	85.97 – 106.63	14,649.00
Strength	3.00*	2.63 – 3.37	45,346.00
Betweenness	1.74*	0.17 – 3.99	62,995.00
Degree - Betweenness	32.65*	25.44 – 40.25	64,012.00
Degree - Strength	11.45*	9.64 – 13.26	49,850.00
Strength - Betweenness	5.07*	3.75 – 6.36	52,913.00

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