1 2

3

4 5

10

12

The OptoReg system: A simple and inexpensive solution for regulating water oxygen.

Rasmus Ern^{1,*} and Fredrik Jutfelt^{1,2}

¹Department of Biology, Norwegian University of Science and Technology, Høgskoleringen 5, 7034
 Trondheim, Norway

²Department of Biology and Environmental Sciences, University of Gothenburg, Medicinaregatan 7B,
 413 90 Gothenburg, Sweden

11 *. Correspondence: rasmus@ern.dk

13 ABSTRACT

14 This paper describes an optocoupler-based regulation apparatus for saturation manipulation of oxygen 15 in water (OptoReg). This system enables control of solenoid valves for oxygen and nitrogen gases using a FireSting-O₂ meter, an optocoupler box, and an electronic switch box. The hardware components 16 17 connect to a computer through USB cables. The control software is free and has a graphical user 18 interface, making it easy to use. With the OptoReg system, any lab with a 4-channel FireSting-O₂ meter 19 can easily and cheaply set up four independently controlled systems for regulating water oxygen levels. 20 Here we describe how to assemble and run the OptoReg system and present a data set demonstrating 21 the high precision and stability of the OptoReg system during static acclimation experiments and 22 dynamic warming trials.

2324 INTRODUCTION

25 Water oxygen is, like water temperature (Brett, 1971), an "abiotic master factor" because of its 26 effects on aquatic organisms (Vaquer-Sunyer and Duarte, 2008; Chu and Tunnicliffe, 2015; Ern et al., 27 2016; Ern, 2019; Woods et al., 2022; Ern et al. 2023). Climate change and eutrophication are expanding 28 the oxygen minimum zones (OMZs) in the world's ocean and increasing the number of aquatic oxygen 29 deficient (hypoxic) "dead zones" in costal and estuarine areas (Diaz and Rosenberg, 2008; Rabalais et 30 al., 2010; Breitburg et al., 2018). Additionally, hyperoxic water can occur during aquatic warming and 31 during sunny days in shallow productive waters, but its potential effect on aquatic organisms is 32 understudied (McArley et al., 2020). Investigating the effects of deoxygenation and hyperoxygenation 33 on aquatic organisms is essential for understanding the important role of oxygen, as well as the impacts 34 of climate change, on aquatic ecosystems. Consequently, the impact of water oxygen on aquatic 35 organisms is an increasingly important research topic (IPCC, 2022).

36 Research on the effects of water oxygen levels on aquatic organisms has been partially hampered by 37 the relative unavailability of systems for controlling water oxygen in laboratory settings. Commercial 38 systems cost tens of thousands of euros, putting them out of reach for many researchers with limited 39 funding. Do It Yourself (DIY) systems based on Arduino and Raspberry Pi platforms have been 40 developed and are cheaper, but require both engineering and programming skills, which can also put 41 them out of reach for many researchers. The ability to regulate water oxygen levels in laboratory settings 42 has therefore required either large funds for available commercial systems or the technical skills to build 43 and program custom systems. To address this issue, we have designed a system for regulating water 44 oxygen levels using the common FireSting- O_2 oxygen meter in combination with commercially 45 available optocouplers, electrical switches, solenoid valves, and software with a graphical user 46 interface. Below we describe how to assemble and run this optocoupler-based regulation apparatus for 47 saturation manipulation of oxygen in water (OptoReg), and we present a data set demonstrating precise 48 oxygen regulation during a static acclimation experiment and during a dynamic warming trial.

49

50 MATERIALS AND METHODS

51 The OptoReg system uses an optocoupler to relay an electrical signal from an oxygen meter to an 52 electrical switch controlling a solenoid valve. When the water oxygen level reaches a predetermined

- 53 threshold value, the solenoid valve opens and releases a gas (nitrogen for hypoxia experiments and
- 54 oxygen for hyperoxia experiments) into the water (Figure 1).
- 55



58 59 60

56 57 FIGURE 1. A schematic drawing of the OptoReg system when regulating water oxygen levels in two tanks, one with hypoxic water and one with hyperoxic water. The FireSting-O₂ oxygen meter measures the water oxygen level using fiber-optic oxygen sensors, and the Pyro Workbench software can send signals depending on the water oxygen levels (Box 1). An optocoupler relays the signal from the oxygen meter to a computer. ClewareControl 61 software (Box 2) on the computer controls an electrical switch with a solenoid valve. The switch opens the 62 solenoid valve if the water oxygen level rises above or decreases below a set threshold value. 63

64 A solenoid valve is an electromechanical device that controls the flow of gas (or liquid). In the OptoReg 65 system, solenoid valves control the bubbling of nitrogen or oxygen gas. Nitrogen bubbling reduces the 66 water oxygen level and oxygen bubbling increases the water oxygen level. A solenoid valve has an inlet 67 port marked with a P (Pressure) and an outlet port marked with an A (Air/Atmosphere). In a normally 68 closed valve, the type used in the OptoReg system, the valve is closed when no electric current is applied 69 and opened by applying an electric current. Nitrogen is an inert gas and can, therefore, be controlled 70 using most types of Solenoid valves. By contrast, oxygen gas supports combustion and must only be 71 controlled using Solenoid valves that are suitable for use with oxygen. Controlling the flow of oxygen 72 gas using a solenoid valve that is not recommended for oxygen increases the risk of fires.

73 An optocoupler is an electronic component that uses light to transmit electrical signals between two 74 electrical circuits – an input circuit and an output circuit – while keeping them electrically isolated from 75 each other. An optocoupler consists of a light-emitting diode (LED) that is connected to the input circuit 76 and a phototransistor that is connected to the output circuit. The LED emits light when a voltage from 77 the input circuit is applied to it. The phototransistor detects the light and generates a current proportional 78 to the light intensity. Keeping the two circuits electrically isolated during signal transmission prevents 79 interference, which would otherwise cause errors in data transmission. An optocoupler is therefore used

- 80 in the OptoReg system to transmit signals between the low voltage FireSting-O2 system and the high
- 81 voltage switch.

82 The OptoReg system described below is built using oxygen meters, sensors, and control software
83 from PyroScience (pyroscience.com); optocouplers, electrical switches, and control software from
84 Cleware (cleware-shop.de); and solenoid valves from Burkert (burkert.com). All components are
85 commercially available and affordable (Table 1).

86

Table 1. Hardware specifications and purchasing details (at the time of publication) on the equipment
 used to build an OptoReg system and on FireSting-O₂ systems.

Item	Stock No	Price (€)	Quantity
Cleware-shop.de			
USB Optocoupler* (OptoIn)	54	60	1
USB Switch 4	11-3	155	1
RS-online.com			
PCB terminal block	712-4487	20	5
Wago terminal block	883-7557	10	10
Equipment wire	361-579	28	100 m
Solenoid valve (for Nitrogen)	307-0248	58	1
Power cable	490-245	9	1
Push-in fitting	212-9173	15	10
Tubing	144-7829	123	150 m
Y tube-to-tube adaptor	916-0918	22	10
Pyroscience.com			
FireSting-O ₂ (1 Channel)	FSO2-C1	2.280	1
FireSting-O ₂ (4 Channels)	FSO2-C4	4.480	1

89 * When ordering the USB Optocoupler from Cleware, add a note in the remarks field that you need

90 *2500 mV input. Then they will adapt the resistance for this setting.*

91

92 Hardware

93

94 The FireSting-O₂ Oxygen meter from PyroScience comes with up to four channels for fiber-optic 95 oxygen sensors, one temperature sensor and an USB cable for connecting it to a computer (see the 96 official manual for how to operate the FireSting oxygen meter). The back of the FireSting oxygen meter 97 has an integrated Connector X2 extension port with one ground (Pin 1) and four analog outputs (Pin 2-98 5) (Figure 2). The four analog outputs have a range of 0 mV to 2500 mV. The 8-channel USB 99 Optocoupler from Cleware has eight contacts (1-8) and a USB cable for connecting it to a computer. 100 The contacts go from right to left, with Contact 1 being furthest to the right and Contact 8 being furthest 101 to the left. Each contact has an input (positive, left clamp) and output (negative, right clamp) (Figure 102 2). A contact is open when no voltage (0 mV) is applied to the input and closed when voltage (2500 103 mV) is applied to the input. When ordering the USB Optocoupler from Cleware, add a note in the 104 remarks field that you need 2500 mV input. The 4-channel USB-Switch from Cleware has four 105 electrical switches (Socket 1-4), a USB cable for connecting it to a computer, and an electrical cord 106 with a wall plug.

- 107
- The FireSting-O₂ Oxygen meter is connected to the Cleware Optocoupler using a PCB Terminal Block,
 a Wago Terminal Block, and equipment wires (Figure 2). The signal from the Optocoupler is relayed
- 110 to the Cleware USB-Switch using the ClewareControl software as detailed below (Box 2).





112 113

FIGURE 2. How to connect the analog outputs on the FireSting-O₂ to the inputs on the Optocoupler, and outputs 114 on the Optocoupler to the ground on the FireSting-O₂. The four analog outputs (Pin 2-5) in the Connector X2 115 extension port on the FireSting are connected to the four inputs (1-4) of the first four contacts on the Optocoupler, 116 using a PCB Terminal Block (red lined wires). The four outputs (1-4) of the first four contacts on the Optocoupler 117 are all connected to the ground (Pin 1) in the Connector X2 extension port on the FireSting, using a Wago Terminal 118 Block (white lined wires). The last four contacts (5-8) on the Optocoupler are not used. Wires are attached to the 119 Optocoupler and the PCB Terminal Block using Spring Clamps, and to the Wago Terminal Block using Spring 120 Cages. The spring clamps are pushed opened using the tip of a 2 mm flathead screwdriver.

121

122 Software

123

124 Pyro Workbench software is used to control the four analog outputs on the FireSting-O₂ meter via the 125 'Alarm if out of range' mode (Box1). In this mode, the outputs can be set to change from 0 mV to 2500 126 mV if the water oxygen level falls below a Lower alarm limit or rises above an Upper alarm limit 127 (Figure 3). When the voltage of an output changes from 0 mV to 2500 mV, the connected contact on 128 the Optocoupler (c.f. Figure 2) is opened. The ClewareControl software is used to pair the first four 129 contacts on the Optocoupler with the four electrical switches on the USB-Switch, in a manner that turns 130 a switch ON when the paired contact is opened and turns in OFF when the contact is closed (Box 2, 131 Figure 4). Turning on a switch will activate the connected Solenoid valve, which will bubble the water 132 with either nitrogen or oxygen, depending on the desired water oxygen level (as detailed below).

BOX 1: Settings for the FireSting-O2 meter using Pyro Workbench software.

The 'Alarm if out of range' mode is programmed in the 'Analog output / broadcast mode' menu, which is opened via Settings \rightarrow Firesting Pro (4 Channels) (A) \rightarrow Analog output / broadcast mode. In the 'Alarm if out of range' mode, the four analog outputs in the Connector X2 extension port can output either 0 mV or 2500 mV. The 0 mV is given if the value of the oxygen level measured by the oxygen sensor is within a specific range, which can be adjusted at the Lower alarm limit and the Upper alarm limit. The analog output changes from 0 to 2500 mV if the value of the measured oxygen level falls below the Lower alarm limit or rises above the Upper alarm limit. The 'Alarm if out of range' mode is programmed in the 'Analog output / broadcast mode' menu, which is opened via Settings \rightarrow Firesting Pro (4 Channels) (A) \rightarrow Analog output / broadcast mode. In the window, the four analog outputs (Pin 2-5) in the Connector X2 extension port are designated with A, B, C, and D. Each of the four analog outputs (A-D) can be paired with one of the four optical oxygen sensor channels (Ch. 1-4). The default settings for 'Channel' and 'Value' should be left unchanged. The default settings for 'Mode' should be changed from 'Standard analog out' to 'Alarm if out of range'. The 'Lower alarm limit' is typed in the box next to the 0 mV and the 'Upper alarm limit' is typed in the box next to the 2500 mV. The default setting is 0.00 for the 'Lower alarm limit' and 5000.00 for the 'Upper alarm limit'. These settings are changed, depending on the desired water oxygen level (Figure 3).

Analog output/Broadcast mode	
Device: A, FireSting-PRO	
☑ Broadcast mode Broadcast ir	terval [ms]: 0
Analog output A-D	
А	В
Channel: Ch. 1 - Lower alarm limit y	Channel: Ch. 2
Value: Oxygen (% air sat) v 0 mV: 0.00	Value: Oxygen (% air sat) 👻 0 mV: 0.00
Mode: Standard analog out 💌 2500 mV: 5000.00	Mode: Alarm if out of range – 2500 mV: 50.00
Higher alarm limit ↑	
С	D
Channel: Ch. 3	Channel: Ch. 4
Value: Oxygen (% air sat) 🔹 0 mV: 150.00	Value: Oxygen (% air sat) 🚽 0 mV: 0.00
Mode: Alarm if out of range v 2500 mV: 5000.00	Mode: Alarm if out of range 🚽 2500 mV: 150.0

134 135

136 137

FIGURE 3. Settings for regulating water oxygen using the '*Alarm if out of range*' mode in the Pyro Workbench
control software for the FireSting-O₂ meter. The Lower alarm limit and the Higher alarm limit are indicated for
the Analog output A. Output A shown default settings; output B show settings for maintaining water oxygen level
at 50% air saturation using solenoid-controlled nitrogen bubbling; output C show settings for maintaining water
oxygen level at 150% air saturation using solenoid-controlled oxygen bubbling, and output D show settings for
maintaining water oxygen level at 150% air saturation using passive oxygen bubbling and solenoid-controlled
nitrogen bubbling. See text and Box 1 for details.

BOX 2: Connecting Optocoupler and USB-switch using ClewareControl software.

The two devices are paired in the 'Device settings' menu, which is opened via View \rightarrow Device settings. The default name for each contact is the Optocoupler serial number followed by the contact number (e.g., 1401039/1 for contact number one), and the default name for each electrical switch is the USB-Switch serial number followed by the switch number (e.g., 652418/1 for switch number one). For easy overview, it is recommended to replace the Optocoupler serial number of the contacts with 'Contact' (e.g., 1401039/1 is renamed Contact/1) and the USB-Switch serial number of the switches with 'Switch' (e.g., 652418/1 is renamed Switch/1) (c.f. Figure 4). Contact/1 is paired with Switch/1 as follows: Select **Contact/1**. Click **Add** under Switch points. Check **opened** under Contact is. Select **Switch** under Action type. Select **Switch** under Action type. Select **Switch** on under Action. Click **Ok**. This path is visualized in the Supplementary materials. With these settings, Switch/1 is turned off when Contact/1 is open (0 mV is applied) and turned on when Contact/1 is closed (2500 mV is applied) (Figure 4). The remaining Contacts (2-4) are paired with the corresponding Switches (2-4) as described for Contact/1 and Switch/1.

Device name Refresh interval	Contact/1 0.5 Seconds	Contact configuration C Input C Output
Change color disable drawing Registry base: 30-4 Switch points	b-001560cf-	1 mA
open = Switch off, S close = Switch on, S	iwitch/1 Switch/1	
Add	Delete Change	

145

146 147

- alarm level is set to the desired water oxygen level (e.g., 50% air saturation) (Figure 3, Output A). With
- this setting, the voltage from the analog output will be 0 mV when the water oxygen level is between
- 154 0% air saturation and 50% air saturation, and it will be 2500 mV when the water oxygen level is above
- 155 50% air saturation (or below 0 % air saturation). When activated, and starting with normoxic water, this
- 156 setting will bubble nitrogen gas into the water until the water oxygen level declines below 50% air

<sup>FIGURE 4 Settings for turning Switch/1 off when Contact/1 is open (0 mV is applied) and for turning Switch/1
on when Contact/1 is closed (2500 mV is applied). See test, Box 2, and Supplementary materials for details.</sup>

¹⁵⁰ Regulating water hypoxia using solenoid-controlled nitrogen bubbling

¹⁵¹ In the Pyro Workbench software, the Lower alarm level is set to 0% of air saturation and the Upper

157 saturation, at which point the solenoid valve is closed. Whenever the water oxygen level rises above158 50% air saturation, because of the passive diffusion of oxygen into the water, the solenoid valve is

opened, and the water is bubbled with nitrogen until water oxygen level again declines below 50% air saturation.

160 sa

162 *Regulating water hyperoxia* (v1) using solenoid-controlled oxygen bubbling

163 When using pure oxygen gas, the solenoid valve must be suitable for use with oxygen. In the Workbench 164 software, the Lower alarm level is set to the desired water oxygen level (e.g., 150% air sat) and the 165 Upper alarm level is set to 5000% air saturation (Figure 3, Output C). With this setting, the voltage from the analog output will be 0 mV when the water oxygen level is between 150% air saturation and 166 167 5000% air saturation, and it will be 2500 mV when the water oxygen level is below 150% air saturation. 168 When activated, and starting with normoxic water, this setting will bubble oxygen gas into the water until the water oxygen level rises above 150% air saturation, at which point the solenoid valve is closed. 169 170 Whenever the water oxygen level decreases below 50% air saturation, for example from diffusion of 171 oxygen out of the water, the solenoid valve is opened, and the water is bubbled with oxygen until water 172 oxygen level again rises above 150% air saturation.

173

174 Regulating water hyperoxia (v2) using passive oxygen bubbling and solenoid-controlled nitrogen
175 bubbling

176 Hyperoxia can alternatively be controlled without the use of oxygen solenoid valves using the following 177 setup. In the Workbench software, the Lower alarm level should be set to 0% air saturation and the 178 Upper alarm level should be set to the desired water oxygen level (e.g., 150% air sat) (Figure 3, Output 179 D). With this setting, the voltage from the analog output will be 0 mV when the water oxygen level is 180 between 0% air saturation and 150 % air saturation, and it will be 2500 mV when the water oxygen 181 level is above 150% air saturation. When activated, and starting with normoxic water, the passive 182 oxygen bubbling will raise the water's oxygen level until the oxygen level rises above 150% air 183 saturation. When this happen, the solenoid valve controlling the flow of nitrogen gas is opened, and the 184 water is bubbled with nitrogen until oxygen level decreases below 150% air saturation, at which point 185 the valve is closed and the water oxygen begins to increase because of the passive oxygen bubbling. 186 The rate of nitrogen bubbling should be large enough, that the water oxygen level decreases when the 187 solenoid valve controlling the flow of nitrogen gas is opened, despite the passive oxygen bubbling.

- 188
- **189** *Validation experiments*

190 101

The accuracy and stability of the OptoReg system were tested during a static temperature acclimation
experiment (10 days) and two dynamic warming trials (0.3°C per min). The acclimation experiment
was conducted using a 50L tank and the warming trials were performed using 12L tanks. Water mixing
during trials was achieved using Eheim universal 300 water pumps (eheim.com). Water temperature
was regulated using a SmartPID CUBE - Smart Thermostat application (smartpid.com) and Aqua Medic
500 W titanium heating elements (aqua-medic.de).

The acclimation experiment was conducted to assess the OptoReg system's ability to maintain stable oxygen levels over prolonged periods. At the start of the experiment, the system was set to maintain the water oxygen level in the acclimation tank at 50% air saturation using solenoid-controlled nitrogen bubbling (c.f., Figure 3B). A second, separate, Firesting system was used to independently record oxygen levels. The results showed that the water oxygen level (mean ± 1 SD) during the 10-day acclimation period was maintained at $50.0 \pm 0.4\%$ air saturation (Figure 5A).

The warming trials were conducted to assess the OptoReg system's ability to simultaneously maintain distinct oxygen levels in different tanks (Figure 1) and to sustain oxygen levels during fluctuations in water temperature and oxygen solubility. The ramping rate was 0.3°C per minute, which is commonly used to assess the acute upper thermal tolerance of aquatic organisms (Becker and Genoway, 1979; Morgan et al. 2018)

At the start of the trials, the OptoReg system was set to maintain one trial tank at 30% air saturation
 using solenoid-controlled nitrogen bubbling (see Figure 3B) and to maintain another trial tank at 200%
 air saturation using solenoid-controlled oxygen bubbling (see Figure 3C). The results showed that the

211 water oxygen level (mean ± 1 SD) during the warming trials was 198.9 $\pm 2.4\%$ air saturation in the 212 hyperoxia tank and 29.8 $\pm 0.6\%$ air saturation in the hypoxia tank (Figure 5B).



213

FIGURE 5 Recorded water oxygen levels during (A) a static acclimation experiment (10 days at 20°C)
 and (B) two dynamic warming trials (0.3°C per min). The embedded graph (A) shows individual O₂

recordings over a typical 20-min period during the acclimation experiment.

217218 CONCLUSION

The OptoReg system offers a simple and inexpensive solution for regulating water oxygen using the FireSting-O2 meter and sensors. Assembling and operating the system requires neither engineering nor programming skills. The system effectively maintains stable water oxygen levels over extended periods and during acute changes in water temperature. By enhancing the capabilities of laboratories to manipulate water oxygen levels, the OptoReg system is poised to facilitate an increase in studies on the responses of aquatic organisms to changes in water oxygen levels. Consequently, this will contribute valuable data to the field of conservation physiology.

227 Acknowledgement

This work was supported by a Marie Skłodowska-Curie Actions to RE (Grant number 893895) and
the European Research Council to FJ (Grant number 101003026–CLIMEVOLVE).

231 References

- Becker CD, Genoway RG (1979) Evaluation of the critical thermal maximum for determining thermal
 tolerance of freshwater fish. Environ. Biol. Fishes 4, 245-256.
 https://doi.org/doi:10.1007/BF00005481
- 236
 237 Breitburg D, Levin LA, Oschlies A, Grégoire M, Chavez FP, Conley DJ, Garçon V, Gilbert D,
 238 Gutiérrez D, Isensee K, Jacinto GS, Limburg KE, Montes I, Naqvi SWA, Pitcher GC, Rabalais
 239 NN, Roman MR, Rose KA, Seibel BA, Telszewski M, Yasuhara M, Zhang J (2018) Declining
 240 oxygen in the global ocean and coastal waters. Science 359, eaam7240.
 241 https://doi.org/10.1126/science.aam7240
- 241 https://doi.org/10.1126/science.aam7240242

246

 ²⁴³ Brett JR (1971) Energetic responses of salmon to temperatures. A study of some thermal relations in
 244 the physiology and freshwater ecology of sockeye salmon (Oncorhynchus nerka). American
 245 Zoologist 11:99-113. https://doi.org/10.1093/icb/11.1.99

- 247 Chu J, Tunnicliffe V (2015) Oxygen Limitations on Marine Animal Distributions and the Collapse of
 248 Epibenthic Community Structure during Shoaling Hypoxia. Global Change Biology 21: 2989249 3004. https://doi.org/10.1111/gcb.12898
 250
- Diaz RJ, Rosenberg R (2008) Spreading dead zones and consequences for marine ecosystems.
 Science 321, 926-929. https://doi.org/10.1126/science.1156401
- Ern R (2019) A mechanistic oxygen- and temperature-limited metabolic niche framework. Phil.
 Trans. R. Soc. B 374, 20180540. https://doi.org/10.1098/rstb.2018.0540
- 257 Ern R, Andreassen AH, Jutfelt F (2023) Physiological mechanisms of acute upper thermal tolerance
 258 in fish. Physiology 38: 141–158. https://doi.org/10.1152/physiol.00027.2022
 259
- Ern R, Norin T, Gamperl AK, Esbaugh AJ (2016) Oxygen dependence of upper thermal limits in fishes. Journal of Experimental Biology 219, 3376–3383. https://doi.org/10.1242/jeb.143495
- IPCC, 2022: Climate Change 2022: Impacts, Adaptation and Vulnerability. Contribution of Working
 Group II to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change [H.O. Pörtner, D.C. Roberts, M. Tignor, E.S. Poloczanska, K. Mintenbeck, A. Alegría, M. Craig, S.
 Langsdorf, S. Löschke, V. Möller, A. Okem, B. Rama (eds.)]. Cambridge University Press.
 Cambridge University Press, Cambridge, UK and New York, NY, USA, 3056 pp.,
 doi:10.1017/9781009325844.
- McArley, T. J., Sandblom, E., and Herbert, N. A.: Fish and hyperoxia—From cardiorespiratory and
 biochemical adjustments to aquaculture and ecophysiology implications, Fish Fish., 1–32,
 https://doi.org/10.1111/faf.12522, 2020.
- 274 Morgan R, Finnøen MH, Jutfelt F (2018) CTmax is repeatable and doesn't reduce growth in
 275 zebrafish. Scientific Reports, 8(1), 7099. https://doi.org/10.1038/s41598-018-25593-4
 276
- 277 Rabalais NN, Díaz RJ, Levin LA, Turner RE, Gilbert D, Zhang J (2010) Dynamics and distribution of natural and human-caused coastal hypoxia. Biogeosciences 7, 585-619. https://doi.org/10.5194/bg-7-585-2010
 280
- 281 Vaquer-Sunyer R, Duarte CM (2008) Thresholds of hypoxia for marine biodiversity. Proceedings of
 282 the National Academy of Sciences of the United States of America 105, 15452-15457.
 283 https://doi.org/10.1073/pnas.0803833105

284

256

269

273

285 Woods HA, Moran AL, Atkinson D, Audzijonyte A, Berenbrink M, Borges FO, Burnett KG, Burnett
286 LE, Coates CJ, Collin R, et al. (2022) Integrative approaches to understanding organismal
287 responses to aquatic deoxygenation. Biology Bulletin. 243:85–103. https://doi.org/10.1086/722899