Maternal investment and early thermal conditions affect 1 performance and antipredator responses 2 3 Maider Iglesias Carrasco^{1,2*}, Jiayu Zhang², Daniel W.A. Noble² 4 5 6 ¹ Evolution and Ecology of Sexual Interactions group. Doñana Biological Station-CSIC 7 ² Division of Ecology and Evolution, Research School of Biology, The Australian National University, Canberra, Australia 8 9 10 *Corresponding author: miglesias15@gmail.com 11 12 Running title: Thermal and maternal effects in lizards 13 14 Lay summary: Early thermal conditions can interact with maternal resource investment to 15 shape offspring performance and behaviour. While anthropogenic climate change is expected 16 to alter the thermal conditions during development, other anthropogenic factors are likely to 17 impact the resource availability and stress levels of mothers. In this study we found that these 18 two factors can interact together to shape offspring responses, which could have important 19 consequences for wildlife conservation in the face of rapid environmental change. 20 21 Abstract: Exposure to increased temperatures during early development can lead to 22 phenotypic plasticity in morphology, physiology, and behaviour across a range of 23 ectothermic animals. In addition, maternal effects are known to be important contributors to 24 phenotypic variation in offspring. Whether the two factors interact to shape offspring 25 morphology and behaviour has been barely explored. This is critical since both incubation temperature and maternal resource investment are likely to change as consequence of altered 26 27 environmental conditions. Using a fully-factorial design we explored how the manipulation 28 of early thermal environment and yolk-quantity in eggs affected the morphology, 29 performance and antipredator behaviour of two sympatric Australian species (Lampropholis 30 delicata and L. guichenoti) that differ in a range of life-history traits. We found that juveniles 31 from the hot treatment were larger than those on the cold treatment in L. guichenoti but not L. 32 delicata. We also found that incubation temperature and maternal investment interacted to 33 shape performance, measured as running speed. Finally, we found that maternal investment 34 impacted antipredator behaviour, with animals from the yolk-reduced treatment incubated 35 under cold conditions resuming activity faster after a simulated predatory attack in L. 36 *delicata*, but not *L. guichenoti*. Our results highlight the importance of exploring the 37 multifaceted role that environments play across generations to understand how different 38 anthropogenic factors will impact wildlife in the future. In addition, our study shows that the 39 responses to different anthropogenic factors can be species-specific. 40 41 Keywords: Bayesian multivariate mixed effects models, incubation temperature,

- 42 *Lampropholis*, morphology, yolk-reduction.
- 43

44

45 Introduction

46

47 Gradual and sudden changes in temperature due to anthropogenic activities pose a challenge 48 to organisms, having important consequences on phenotype and fitness. Ectotherms are 49 especially vulnerable to altered thermal conditions, and exposure to increased temperatures 50 during early development have been shown to lead to phenotypic plasticity in morphology, 51 physiology and behaviour, that can impact fitness in a range of taxonomic groups (Dang et 52 al., 2015; Dayananda & Webb, 2017; Noble et al., 2018; Raynal et al., 2022; Ślipiński et al., 53 2021; Valenzuela & Lance, 2004). While research has focused mainly on the effects of early 54 life temperature on individual development, little work has addressed the multifaceted role 55 that environments play across generations. Environmental stress (e.g., changes in thermal or 56 resource conditions) experienced by mothers is expected to cascade to affect offspring 57 through maternal effects, and this may interact in complex ways with environments 58 experienced by offspring.

59

60 Maternal effects are important contributors to offspring phenotypic variation (Bernardo, 61 1996; Noble et al., 2014; Wolf & Wade, 2009), and may moderate the effects of the 62 environment experienced during development. Mothers can adjust their reproductive 63 behaviour or differentially invest in energy and resources that are deposited in eggs in 64 response to changes in the environmental conditions they experience (Carter et al., 2018; 65 Huang et al., 2013; Rutstein et al., 2005). For example, nutrient deprived mothers alter their 66 reproductive allocation compared to mothers fed with normal quality food, leading to changes in offspring phenotype and sex (Warner et al., 2007). Similarly, maternal diet can 67 affect hormone deposition in eggs, which is known to influence hatching success, and 68 69 offspring phenotype and fitness (e.g., Rosenfeld and Roberts 2004; Warner and Lovern 2014; 70 Rutstein et al. 2005; Huang et al. 2013; Carter et al. 2018). Therefore, maternal effects can 71 exacerbate, dampen or counteract negative phenotypic or fitness effects of early life 72 environments experienced by offspring, and may play an important role in explaining the 73 diversity of phenotypic responses observed within and across species (e.g., Noble et al. 74 2018). Nonetheless, the interaction between maternal and offspring environments on key 75 fitness traits is seldom explored in detail to better understand the multifaceted role 76 environments will have on phenotypic development and fitness. Quantifying such effects is 77 important in an era of climate change because maternal environments will be impacted -78 among others, their ability to acquire and invest resources in reproduction (McCarty, 2001; 79 McRae et al., 2008).

80

81 Maternal and offspring environments that impact upon behaviour are likely to have important 82 consequences in shaping how organisms interact with their environment. Such behavioural 83 changes can have a direct link to fitness (reviewed in Saaristo et al., 2018 in association with 84 chemical contamination). Antipredator responses, including predator escape and refuge 85 seeking, are a case in point since the inability to escape from predatory attacks are inevitably 86 associated with mortality. Thermal developmental conditions and maternal effects are known 87 to affect antipredator strategies. For instance, exposure to high temperatures during early 88 development (e.g., eggs or juveniles) has been shown to affect sprint speed and a range of 89 antipredator behaviours such as predator avoidance and hiding time (Brodie & Russell, 1999; 90 Dalesman & Rundle, 2010; McDonald & Schwanz, 2018; Webb et al., 2001). Similarly, 91 maternal effects can pre-adapt offspring responses to the prevalent predatory conditions by 92 changing egg composition (Sharda et al., 2021). Such effects are likely mediated by changes 93 in morphology (e.g., size, body condition etc), which in turn can influence the antipredator

- 94 strategies of individuals (Lancaster et al., 2010; Mcghee et al., 2012; Räsänen et al., 2005).
- 95 Clearly, thermal conditions and maternal effects independently have the potential to influence
- 96 a range of fitness-related behaviours, however, whether the two interact to shape offspring
- 97 responses remains unknown.
- 98

99 To test how species with different life-history traits respond to altered development

- 100 temperatures and maternal resource investment, we measured an integrated set of
- 101 morphological and behavioural traits related to antipredator responses in two closely related
- 102 lizard species, Lampropholis delicata and L. guichenoti. We apply 'phenotypic engineering'
- 103 methods (Sinervo & Basolo, 1996) to manipulate both maternal investment in eggs and
- 104 offspring temperature in a fully factorial design. We predicted that: 1) embryos experiencing high temperatures early in development would be smaller in size with longer tails and have 105
- 106 reduced performance (e.g., Sanger et al., 2018; Tiatragul et al., 2017). As a result, we expect
- 107 them to be risk adverse relative to embryos experiencing cold temperatures; 2) lower
- 108 maternal investment in eggs should exacerbate the effects of temperature; 3) Lampropholis
- 109 delicata will be more strongly impacted by temperature and reduced maternal investment
- 110 because of the smaller egg size and larger clutches compared to L. guichenoti.
- 111

112 **Methods**

- 113
- 114 Study species and housing
- 115

116 We used two sympatric skink species. The delicate skink (Lampropholis delicata) is a small 117 lizard (max. SVL 51 mm) native to south-eastern Australia (Wilson & Swan, 2010). Females

- 118 lay a single clutch of 3-6 small eggs each year. It has been used extensively in experiments to
- 119 explore how different environments affect the morphology and performance of individuals
- 120 (Bilcke et al., 2006; De Jong et al., 2023; Downes & Hoefer, 2007). In addition, it is an
- 121 invasive species on some islands (Chapple, 2016; Chapple et al., 2014), which suggests that
- 122 the species has the potential to respond and become used to novel environmental conditions.
- 123 The garden skink (Lampropholis guichenoti) is a small lizard (max. SVL 48 mm) and 124 widespread across south-eastern Australia (Wilson & Swan, 2010). Females lay one or two
- 125 clutches of 2-4 large eggs a year. The garden skink has been used in previous studies
- 126
- exploring the effect of rearing temperatures and humidity conditions on offspring phenotypic 127 plasticity (Booth et al., 2000; Qualls & Shine, 1998).
- 128
- 129 We captured gravid female L. delicata and L. guichenoti in semiurban parks in Sydney
- 130 (Australia). Animals were brought to the laboratory at XXX (not reported for blind review)
- 131 where they were housed in single-species groups of five in indoor terraria (3-4 females -
- 132 width x length: 40x55 cm) to allow them to lay eggs. Terraria were filled with approximately
- 133 8 cm deep of soil, refuge, a water container, and a container full of vermiculite for egg laying.
- 134 Terraria were heated by a lamp and had a UV lamp for UVA/UVB exposure. The heat lamp
- 135 was situated at one end to ensure a temperature gradient. Lights were set to a photoperiod of
- 136 12:12 h (light/dark). Animals were provided with water every day (both spraying the soil and
- 137 filling the water container) and with crickets dusted in calcium and multivitamin every
- second day. Females were kept in the laboratory for around two weeks for egg-laying and 138 139 were then released at their capture locations.
- 140
- 141 Experimental design
- 142

- 143 To explore how incubation temperature and maternal investment interact to affect
- 144 performance and antipredator behaviour of juveniles, we designed a fully-factorial
- 145 experiment where eggs of the two skink species were exposed to two levels of temperature
- 146 [cold $(23 \pm 3^{\circ} \text{ C})$ or hot $(28 \pm 3^{\circ} \text{ C})$] and yolk removal (yolk content reduced or a sham-
- 147 control) to simulate changes in maternal investment in eggs. Incubation temperatures were
 148 selected to mimic extreme temperatures measured in natural nests of *L. delicata* (Cheetham et
- al., 2011).
- 150
- 151 Enclosures were checked daily for eggs. We randomly allocated one egg from a clutch to
- each of our four treatments: 23°C sham-control eggs; 23°C yolk reduced eggs; 28°C sham-
- 153 control eggs; and 28°C yolk reduced eggs. Our design was a partial split-clutch design as it
- 154 was not possible to allocate eggs from a given clutch to each of the four treatments (i.e., a 155 split-clutch design) given the small clutch sizes of some individuals. Eggs allocated to the
- 156 yolk reduced treatment were weighed (to the nearest mg), and then pierced with a sterilised
- 157 insulin syringe to extract part of the yolk. Eggs were weighted again, and the difference in
- 158 weight pre- and post-extraction was used as an approximation of the percentage of yolk
- 159 extracted. Following methods in Sinervo, 1990 (Sinervo, 1990) we aimed for around 15%
- 160 yolk removal (mean =- $SD = 12.49\% \pm 2.64$). Control eggs were weighted and pierced with a
- 161 needle, but we did not remove any yolk. Eggs were then placed in a container filled with four
- 162 grams of vermiculite dampened with 12 g of water and covered with cling-wrap (Glad Wrap)
- 163 to avoid dehydration. Each egg was then placed in an incubator at the corresponding
- 164 temperature.
- 165
- 166 Measures of morphological traits, performance, and antipredator behaviour
- 167

168 We checked the eggs every day for hatchlings and each hatched lizard was individually

- housed in terraria (20 x 35 cm) heated by a heat cord. All animals also had UV lighting.
- 170 Enclosures contained paper as substrate, a water container, and a refuge. All the animals were
- housed in the same laboratory conditions. Juveniles were feed every second day withcrickets.
- 172 0 173
- 174 Juveniles were measured and their behaviour tested when they were 3-5 weeks old. We 175 ensured that there was equal representation from each of the four treatment combinations and species (e.g., eggs in cold treatment had longer incubation times) during each measurement 176 177 session (groups of 48 per day, see below). Body size and mass can influence performance and 178 behaviour (Baxter-Gilbert et al., 2018; Huey & Hertz, 1984). As such, we measured weight, 179 snout-vent length, tail length and total length at each measurement to control for these during 180 the analysis. Given the small size of hatchlings, we weighed lizards (to the nearest mg) using an Ohaus scale and took a ventral photo of each lizard. From this photo we later measured the 181
- 182 snout-vent length (SVL) as the distance (in mm) from the snout to the cloaca, and the tail
- length as the distance from the cloaca to the tip of the tail using imageJ (Abràmoff et al.,2004).
- 184 185
- 186 We set up 12 CCTV cameras with each recording four individual terraria simultaneously.
- 187 Each week we selected 48 juveniles from the correct age window a mix of the two species
- and four treatment combinations. We weighted and photographed the animals, and then
- 189 placed them randomly across the shelves to avoid any biases associated with the location in
- the laboratory. The terraria for the assays were opaque to avoid lizards viewing each other
- which could influence their behaviour. In addition, the terraria were separated from the
- 192 walking corridors by thick-opaque curtains to avoid the presence of the researchers to

193 influence lizard behaviour. All enclosures had a refuge and a water container. The heat lamp 194 was placed on one side of the enclosure to ensure a thermal gradient of at least 6 degrees

- 195 between the lamp and the refuge. The same group of animals were housed in these terraria during the 6 days of the trials.
- 196 197

198 Over the six days animals were under cameras we took three measurements of performance 199 and antipredator behaviour for each animal with one day between subsequent measurements. 200 More specifically, we collected the following behavioural variables:

- 201 (1) Running speed (in seconds): We measured running performance as the total time 202 needed to run the one-metre-long straight racetrack. We also recorded burst speed as 203 the fastest 25 cm section (the racetrack had a detector and time tracker every 25cm). 204 For analysis, we used the total time used to cover the full one meter and the fastest 205 time taken to run a 25 cm interval. Fifteen minutes before each performance measure, 206 lizards were placed in an incubator at 28°C to ensure constant body temperature 207 across lizards. After the trial, each animal was placed back in their corresponding terraria. We repeated the running trial 3 times for each individual, on alternate days. 208
- 209 (2) Activity: We measured the distance travelled (in cm) as a proxy for the activity level of individuals. To record activity, we removed the refuge and the water container 210 from the terraria to avoid animals hiding. We then switched on the camera and left the 211 animals to behave and move freely in their terrariums for 20 minutes. The distance 212 covered was later calculated using the software EthoVision XT (vers. 12.0). 213
- (3) Antipredator behaviour. Immediately after the activity trial, we replaced the refuge. 214 After 30 minutes, we simulated a predatory attack. To do this, we approached the 215 216 terraria and tapped the animals with a painting brush near the tail until they took 217 refuge. The same person (JZ) performed all the predatory attacks. The only identification in the terraria was the randomly allocated ID number provided to the 218 219 eggs at the beginning of the experiment such that JZ was blind to the treatment. After 220 the simulated attack, we recorded each lizard's response for 90 minutes. From the videos we calculated (1) the time (in seconds) each lizard took to seek refuge since 221 222 the first tape in the tail (hereafter "time to hide") (2) the time between the moment the animal took refuge (time to hide) until the animal's head appeared at the entrance and 223 was clearly visible in the videos (hereafter "hiding time", in seconds) and (3) the time 224 225 elapsed since the animal took refuge (time to hide) to the moment the animal left the refuge to start their normal activity after the predatory attack (hereafter "time to 226 227 activity"). We consider this to be the moment when the back forelimbs left the refuge. 228 We repeated the activity and antipredator assays three times in alternate days from 229 performance trails.
- 230 231 All trials took place between 9 and 12 am, during the period when activity was the highest. 232 We recorded 22 individuals per species and treatment. We discarded from the analysis any 233 individual that lost their tail during the experiment (n = 5 out of 176) to avoid any bias 234 associated with impaired running ability. The final sample sizes were 22 for hot-control L. 235 delicata, and 23:control and 28:yolk-reduced L. guichenoti, and 21 for the rest of the 236 combinations.
- 237
- 238 Statistical analysis
- 239

240 We used Bayesian Multivariate Mixed Effects Models using rstan (Team, 2020) in the

- 241 package brms (Bürkner, 2018) to explore whether incubation temperature and maternal
- 242 investment impacted morphology (tail length, SVL and weight), performance (running speed

- and activity level) and antipredator behaviour. We also estimated the correlations between the
- variables measured at the between- and within-individual level. We first checked for
- normality of the data by visualizing the residuals of intercept only random effects model. To
- 246 meet the assumptions of normality running speed (both 25 cm burst and 1 m long) was log-
- transformed for the two species. For all models, we ran four MCMC chains, with each chain
- being run for 4000 iterations with a warmup of 1000 and used default priors. We retained
- each sample (thinning of 1) from each chain. We checked that MCMC chains were mixing well by visualising trace plots, checked that all chains had converged ($R_{hat} < 1.01$), and that
- 250 wen by visualising trace plots, checked that an chains had converged ($K_{hat} > 1.0$ 251 the effective sample size for each parameter was greater than 1000.
- 252
- 253 Both species were analysed separately. We ran two separate multivariate mixed models for 254 each species, one with morphological traits, and the other with performance and antipredator 255 behaviour as response variables. We separated morphology from performance and behaviour 256 because the latter variables were measured 3 times allowing us to decompose between and 257 within-individual variation (O'Dea et al., 2022). Missing data resulted from video failures for 258 some assays. Instead of a complete case analysis, we retained missing data and using data 259 augmentation methods during model fitting which can be more powerful than complete case analyses (Noble & Nakagawa, 2021). Models contained fixed effects (explanatory variables) 260 of incubation temperature and maternal investment treatment along with their interaction. We 261 262 also included individual and clutch identity as random effects (intercepts). In the morphology 263 model, only clutch was added as random effect given that we only had a single measurement 264 for each individual. In the behaviour model, SVL was included as covariate to control for any 265 potential effect of body size on the traits measured. We repeated the behaviour/performance 266 model without SVL as covariate to explore for any indirect effect of temperature and maternal treatments on behaviour that might have been influenced by body size. Most of the 267 268 performance and antipredator variables showed moderate to high repeatability between the three different measures taken (Table S1). Using the posterior distributions from these 269 270 models, we derived the key interaction comparison of interest – whether the difference 271 between control and yolk removal treatments was amplified or subdued in response to temperature. In addition, we used the posterior distribution to calculate the overall 272 273 temperature and maternal investment effect by pooling the posteriors across the second 274 factor. We present the posterior mean and 95% credible intervals (CI) for these parameters of 275 interest. Credible intervals not overlapping zero were considered significant and we calculate 276 and present the probability (pMCMC) of obtaining this effect under a null hypothesis of no 277 effect. 278
- 279 **Results**
- 280
- Maternal investment and early thermal environment affected morphology in L guichenoti but
 not L. delicata
- We did not find any effect of temperature, maternal investment, or their interaction on morphology in *Lampropholis delicata* [tail length: (Fig1a), snout-vent-length (SVL, Fig 1b) or weight (Fig 1c) (See Table 1)].
- In contrast, maternal investment and temperature treatment interacted to affect weight
 in *L guichenti* (Table 1). The weight difference between control and yolk removal treatment
 was larger in the cold incubation temperature compared to the difference under warm
 incubation temperatures for *L guichenoti* (Table 1). We also found a statistically significant
 effect of incubation temperature in juvenile size of *L. guichenoti* (Table 1). Generally,
- 292 individuals coming from eggs incubated at hotter temperatures had longer tails (estimate =

- 293 6.347, 95% CI = 9.619-3.079, Table 1, Fig 1d), larger SVL (estimate = 2.973, 95% CI = 294 4.715 - 1.241, Table 1, Fig 1e) and were heavier (estimate = 0.112, 95% CI = 0.207 - 0.019, 295 Table1, Fig1f) than those hatched from eggs at colder temperatures. 296 In both species, there was a positive correlation between SVL, tail length and body 297 mass, with individuals with larger bodies also being heavier and having longer tails (Supp 298 Info Table S2). 299 300 Impacts of maternal investment on running performance are mediated by early thermal 301 environment in both species 302 303 We found a statistically significant interaction between maternal yolk investment and 304 incubation temperature on 25cm burst speed of juvenile L. guichenoti (Fig 2d, Table 1). This 305 effect persisted even when controlling for SVL (Table S3). There was a significantly bigger difference in 25 cm burst speed between control and yolk removal eggs in lizards incubated 306 307 under hot conditions compared to cold conditions (Table 2 – estimate = -0.470, 95% CI: -308 0.854 - -0.081, pMCMC = 0.018). 309 Although not significant, we also found a similar interaction between maternal yolk 310 investment and incubation temperature on 1m sprint speed in L. delicata (Table 2 & Table 311 S3). However, in contrast to L. guichenoti, there was a significantly smaller difference in 1 312 cm burst speed between control and yolk removal eggs in lizards incubated under hot 313 conditions compared to cold conditions (Table 2 – estimate = 0.335, 95% CI: -0.023 - 0.689, 314 pMCMC = 0.068). 315 In both species, trials where individuals had a faster burst speed also took shorter to 316 travel the full 1m track (L. delicata: within-individual correlation \pm SE = 0.42, 95% CI = 0.30-0.54; L. guichenoti: within-individual correlation + SE = 0.51, 95% CI = 0.4-0.61). In 317 318 addition, individuals that has faster burst speed also tended to run the full 1m faster (L. delicata: between-individual correlation = 0.82, 95% CI = 0.63-0.95; L. guichenoti: within-319 320 individual correlation = 0.95, 95% CI = 0.89-0.99). 321 322 Weak evidence that antipredator behaviour is affected by early thermal environment and 323 *maternal investment* 324 325 Antipredator behaviours were weakly integrated with performance measures at the 326 between and within-individual levels for most traits (Table S4). At the between-individual 327 level, there was a strong correlation (r = 0.90, 95% CI: 0.78–0.97) between the hiding time 328 and the time to activity as well as between the time to activity and burst speed (r = 0.32, 95%329 CI: 0.04 - 0.57) in L. guichenoti. At the within-individual level trials lizards with shorter 330 hiding times also resumed their activity faster overall for both species (L. guichenoti: r = 331 0.69, 95% CI: 0.58 – 0.77; *L. delicata*: r = 0.82, 95% CI: 0.73 – 0.88). 332 Changes in the time to activity after a simulated predatory attack between control and 333 yolk removal eggs depended on temperature in L. delicata when controlling for body size 334 (interaction estimate = -1,003.752, 95% CI = -1,988.452--33.590, pMCMC = 0.044, Table 335 S3). Similar effects were observed when not controlling for body size, but it was not 336 significant (Table 2). Yolk reduced lizards appeared to resume activity faster compared to lizards hatching from control eggs when incubated at cold temperatures whereas there was no 337 338 difference between control and yolk removed eggs under hot temperatures (Fig. 3b). We did 339 not find strong evidence that other behavioural traits involved in antipredator responses were 340 impacted by temperature, maternal investment, or their interaction in L. delicata or L. 341 guichenoti (Table 2 & Table S3; Figure 3).
- 342

343 **Discussion**

344

345 Environmental conditions experienced by offspring are expected to interact in complex ways with the environments experienced by their parents. Anthropogenic climate change and other 346 347 human-associated stressors will simultaneously alter temperatures while impacting other factors such as maternal food availability and stress levels that could affect the maternal 348 349 investment in eggs. In ectotherms, temperature conditions during early life are known to have 350 important effects on individuals (Noble et al., 2018; Singh et al., 2020; While et al., 2018). but little work has explored the interaction between maternal investment and temperatures; 351 352 even though there have been suggestions of these factors explaining variation in thermal 353 effects (Noble et al., 2018). Understanding how maternal effects interact with offspring 354 environments to affect development and fitness is important for ascertaining whether climate 355 induced changes are likely to be exacerbated or dampened by impacts in parental generations 356 and help to explain why responses vary so dramatically across populations and species with 357 respect to early thermal conditions. Here we explored whether cold and hot incubation 358 temperatures lead to different responses in morphology, performance and antipredator 359 behaviour in two sympatric skink species (L. delicata and L. guichenoti), and whether a 360 controlled manipulation in the amount of yolk in eggs moderated such responses. We found that hot incubation temperatures affect morphology, and that temperature interacts with 361 362 maternal yolk treatment to moderate performance and the associated antipredator responses. 363 The effects observed appear to be species-specific, and possibly the result of different life-

364 history strategies.

365 *Effect of early life thermal conditions and maternal investment on morphology*

366 Contrary to our expectations we found that juveniles of *L. guichenoti* emerging from eggs incubated at hot temperatures were larger and heavier than those reared at cold temperatures, 367 368 however incubation temperature did not significantly affect morphology in L. delicata (see 369 also de Jong et al., 2022). Since we did not find any effect of incubation temperature on body 370 size on the day of hatch in either of the two species (data collected for another study), our 371 results show that the difference in body size between individuals coming from cold and hot 372 incubation regimes in L. guichenoti appeared during early juvenile development. Our 373 findings suggest that, rather than a change in metabolic and developmental rates during 374 embryo growth of this species, incubation temperature programmed post-hatching 375 metabolism, potentially affecting feeding and growth rates, as observed in previous studies in lizards exposed to high temperatures (Singh et al., 2020). 376

377 We did not find any effect of the maternal investment treatment on body length for 378 either L. guichenoti or L. delicata, but body mass was affected in L. guichenoti. Although we 379 found that individuals of both species hatched smaller from the yolk reduced treatment than 380 from the control (again, data collected for another study, but see also e.g., Warner and Lovern 381 2014), maternal investment effects on morphology seem to have disappeared by the age of 3-382 5 weeks. This finding contrasts with previous studies where juveniles from yolk-reduced 383 treatments hatched smaller but also showed slower growth rates than those from control 384 treatments (Warner & Lovern, 2014). Our results are not completely surprising, however, 385 since many of the impacts of incubation conditions on morphological and behavioural traits 386 observed in recently hatched individuals often disappear as the individuals age (McDonald & 387 Schwanz, 2018; Pearson & Warner, 2016). This could also be the case for many maternal 388 effects. In our study, the fact that juveniles of both yolk-reduced and control eggs attain a

- 389 similar body size a few weeks after hatching suggests that individuals can, to some extent,
- compensate for the poor start in life by accelerating their growth during the first few weeks,
- probably by increasing their feeding rates. This might be key, since in a range of species,
- larger juvenile body size has been associated with better survival (e.g., Einum & Fleming,
- 2000; Webb et al., 2006), although this relationship is sometimes complex and dependant of
 other ecological and biological factors (Langkilde & Shine, 2005; Sinervo et al., 1992;
- 395 Warner & Andrews, 2002).
- 396

397 *Effect of early life thermal conditions and maternal investment on performance and*398 *behaviour*

399 The growth compensation observed in the analyses of morphology might be associated with

- 400 changes in the allocation of resources and trade-offs with other fitness-associated traits
- 401 during development, such as the observed slower running speed of juveniles of *L. delicata*
- 402 hatched from yolk-reduced eggs and of *L. guichenoti* hatched from yolk-reduced eggs in hot
- 403 incubation conditions. This result suggests that, even though animals from poor
- 404 developmental conditions are able to morphologically catch up with individuals from more
- 405 beneficial early-life conditions, this might come at a cost in performance. This is in
- 406 accordance with previous studies on *Sceloporus undulatus* showing that clutches with
- individuals with fast grow rates, but slow runners, had lower survival than slow growing butfast runners (Warner & Andrews, 2002).
- 409

410 Impaired performance was expected to lead to more risk adverse individuals to compensate 411 for a potential increased vulnerability to predators. In contrast to our prediction, antipredator 412 behaviour of L. guichenoti was not affected by our treatments. In addition, we found that 413 juvenile L. delicata from the yolk-reduced treatments, and especially when reared at cold 414 temperatures, took shorter to resume activity after a simulated predatory attack than those 415 from the control treatment – although this was not a strong effect. Our results show that 416 despite their impaired performance, juveniles from the yolk-reduced treatment leave the refuge quicker than those from the control treatment. This contrasts with previous studies that 417 have found that individuals with perceived higher vulnerability alter their antipredator 418 419 behaviour to hide for longer to successfully avoid a predatory attack (e.g., Cooper, 2007; 420 Iglesias-Carrasco et al., 2016; Martin & López, 1999). Instead, our results suggest that the 421 benefits of resuming activities, such as basking and feeding, might potentially outweigh the

- 422 potential survival costs in *L. delicata*, at least in a laboratory setting where real predators are
- 423 absent. However, from our experiment we cannot know whether the change in antipredator
- response observed confers a fitness advantage regarding, for example, quicker growth, or
- 425 instead would lead to costs in terms of increased predation risk in the wild. Further
- 426 experiments would benefit from studying how incubation temperature and maternal427 investment interact to affect behaviour, and the consequent fitness payoffs, in a more natural
- 427 investm428 setting.
- 429
- 430 Species-specific responses

431 The two skink species studied differed in their morphological, performance and antipredator

- 432 responses to the incubation temperature and maternal investment. These differences could be
- in part associated with some life-history traits, such as the size of the egg. Eggs of *L. delicata*
- 434 are smaller than those of *L. guichenoti*, which might make these eggs more sensitive to small
- alterations in the incubation environment, strongly impacting the phenotype and behaviour of

- 436 juveniles, as observed in our study (Thompson et al., 2001). While this result suggests that
- 437 environmental changes in early thermal environment coupled with reduced maternal
- 438 investment will impact *L* delicata more negatively compared to *L* guichenoti, we caution over
- 439 interpretation because phenotypic and behavioural plasticity could provide juveniles with
- increased environmental tolerance that may confer a fitness benefit in face of climate change
 (DeWitt et al., 1998; Fox et al., 2019; Yeh & Price, 2004, but see Oostra et al., 2018). The
- 442 ability to plastically respond to different environmental conditions could also explain L
- 443 *delicata's* success as an invasive species (see e.g., (Davidson et al., 2011). In contrast, the
- 444 lack of behavioural responses in *L. guichenoti* could be a sign of the inability of the species to
- adaptively react to environmental challenges, or rather suggest that in this species the
- explored behavioural responses might not impose a fitness cost. Since the incubation
- temperatures used in our experiment overlap with the those occurring in the wild (Cheetham
- et al., 2011), it will be interesting to explore how more extreme thermal incubationconditions, expected as consequence of anthropogenic climate change, will interact with
- 450 maternal condition to shape hatchling performance and survival in the future.
- 451 Conclusions
- 452

453 We have shown that exploring the complex interaction between offspring and maternal

454 environments is critical to predict how anthropogenic activities will affect individual

455 performance and ultimately fitness. Although maternal yolk investment did not buffer the

456 effects of higher incubation temperature in all the morphological and behavioural traits

457 measured, the general pattern suggests that a reduction in the resources allocated by mothers

- to eggs exacerbates the response triggered by warmer temperatures. Future research willbenefit from studying whether such plastic responses are adaptive in novel environment
- 459 benefit from studying whether such plastic responses are adaptive in novel environment 460 conditions, whether more extreme temperatures predicted by climate change will exert
- 461 stronger reactions, and whether species sensitivity depends on species-specific life-history
 462 traits.
- 463

464 Acknowledgements: We thank Kai Dewar, Eoin Noble and Rose Zhang for help in the
465 laboratory. Lizards were captured under the NSW SL102296 and ACT LT201917 permits,
466 and the handling and procedures used with animals complied with the ethics approval
467 ARA2019/17 from the Australian National University. This work was supported by the
468 Australian Research Council, though an ARC Discovery Project to DWAN (grant number
469 DP210101152).

470

471 Author contribution: MI-C and DN conceived and designed the experiment; MI-C and JZ
472 collected the data; MI-C and DN analysed the data. MI-C drafted the first version of the
473 manuscript; all authors contributed to the draft and gave final approval for publication.

- 474
- 475 **Data availability**: Data used in this article can be accessed in
- 476 https://github.com/daniel1noble/lampro_yolk_behav
- 477

478

479 **References**

- Abràmoff, M. D., Magalhães, P. J., & Ram, S. J. (2004). Image processing with imageJ. *Biophotonics International*, 11(7), 36–41.
- Baxter-Gilbert, J., Riley, J. L., & Whiting, M. J. (2018). Runners and fighters: clutch effects
 and body size drive innate antipredator behaviour in hatchling lizards. *Behavioral*
- 484 *Ecology and Sociobiology*, 72, 97. https://doi.org/10.1007/s00265-018-2505-7

- 485 Bernardo, J. (1996). Maternal effects in animal ecology. *American Zoologist*, 36(2), 83–105.
 486 https://doi.org/10.1093/icb/36.2.83
- Bilcke, J., Downes, S., & Büscher, I. (2006). Combined effect of incubation and ambient
 temperature on the feeding performance of a small ectotherm. *Austral Ecology*, *31*(8),
 937–947. https://doi.org/10.1111/j.1442-9993.2006.01663.x
- Booth, D. T., Thompson, M. B., & Herring, S. (2000). How incubation temperature
 influences the physiology and growth of embryonic lizards. *Journal of Comparative Physiology B Biochemical, Systemic, and Environmental Physiology*, 170(4), 269–276.
- 493 https://doi.org/10.1007/s003600000097
- Brodie, E. D., & Russell, N. H. (1999). The consistency of individual differences in
 behaviour: Temperature effects on antipredator behaviour in garter snakes. *Animal Behaviour*, 57(2), 445–451. https://doi.org/10.1006/anbe.1998.0990
- Bürkner, P. C. (2018). Advanced Bayesian multilevel modeling with the R package brms. *R Journal*, 10(1), 395–411. https://doi.org/10.32614/rj-2018-017
- 499 Carter, A. W., Bowden, R. M., & Paitz, R. T. (2018). Evidence of embryonic regulation of
 500 maternally derived yolk corticosterone. *Journal of Experimental Biology*, 221(22).
 501 https://doi.org/10.1242/jeb.182600
- 502 Chapple, D. G. (2016). Origin, Spread and Biology of the Invasive Plague Skink
 503 (*Lampropholis delicata*) in New Zealand. In New Zealand Lizards (pp. 341–360).
 504 https://doi.org/10.1007/978-3-319-41674-8
- 505 Chapple, D. G., Miller, K. A., Chaplin, K., Barnett, L., Thompson, M. B., & Bray, R. D.
 506 (2014). Biology of the invasive delicate skink (*Lampropholis delicata*) on Lord Howe
 507 Island. *Australian Journal of Zoology*, 62(6), 498–506. https://doi.org/10.1071/ZO14098
- 508 Cheetham, E., Doody, J. S., Stewart, B., & Harlow, P. (2011). Embryonic mortality as a cost
 509 of communal nesting in the delicate skink. *Journal of Zoology*, 283(4), 234–242.
 510 https://doi.org/10.1111/j.1469-7998.2010.00764.x
- 511 Cooper, W. E. (2007). Compensatory changes in escape and refuge use following autotomy
- in the lizard Sceloporus virgatus. Canadian Journal of Zoology, 85(1), 99–107.
 https://doi.org/10.1139/Z06-200
- 514 Dalesman, S., & Rundle, S. D. (2010). Influence of rearing and experimental temperatures on
 515 predator avoidance behaviour in a freshwater pulmonate snail. *Freshwater Biology*,
 516 55(10), 2107–2113. https://doi.org/10.1111/j.1365-2427.2010.02470.x
- 517 Dang, W., Zhang, W., & Du, W. G. (2015). Incubation temperature affects the immune
 518 function of hatchling soft-shelled turtles, *Pelodiscus sinensis*. *Scientific Reports*, 5, 1–9.
 519 https://doi.org/10.1038/srep10594
- Davidson, A., Jennions, M., & Nicotra, A. (2011). Do invasive species show higher
 phenotypic plasticity than native species and, if so, is it adaptive? A meta-analysis.
 Ecology Letters, 14, 419–431. https://doi.org/10.1111/j.1461-0248.2011.01596.x
- 523 Dayananda, B., & Webb, J. K. (2017). Incubation under climate warming affects learning
 524 ability and survival in hatchling lizards. *Biology Letters*, 13(3), 20170002.
 525 https://doi.org/10.1098/rsbl.2017.0002
- De Jong, M. J., Alton, L. A., White, C. R., O'Bryan, M. K., Chapple, D. G., & Wong, B. B.
 M. (2023). Long-term effects of incubation temperature on growth and thermal
 physiology in a small ectotherm. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 378(1884), 20220137.
 https://doi.org/10.1098/rstb.2022.0137
- de Jong, M. J., White, C. R., Wong, B. B. M., & Chapple, D. G. (2022). Univariate and
 multivariate plasticity in response to incubation temperature in an Australian lizard. *Journal of Experimental Biology*, 225(22), jeb244352.
- 534 https://doi.org/10.1242/jeb.244352

- 535 DeWitt, T. J., Sih, A., & Wilson, D. S. (1998). Cost and limits of phenotypic plasticity.
 536 *Trends in Ecology & Evolution*, *13*(97), 77–81. https://doi.org/10.1111/j.1558 537 5646.2009.00647.x
- Downes, S., & Hoefer, A. M. (2007). An experimental study of the effects of weed invasion
 on lizard phenotypes. *Oecologia*, 153(3), 775–785. https://doi.org/10.1007/s00442-0070775-2
- 541 Einum, S., & Fleming, I. A. (2000). Selection against late emergence and small offspring in
 542 Atlantic salmon (*Salmo salar*). *Evolution*, 54(2), 628–639.
 543 https://doi.org/10.1111/j.0014-3820.2000.tb00064.x
- Fox, R. J., Donelson, J. M., Schunter, C., Ravasi, T., & Gaitán-Espitia, J. D. (2019). Beyond
 buying time: The role of plasticity in phenotypic adaptation to rapid environmental
 change. *Philosophical Transactions of the Royal Society B: Biological Sciences*,
 374(1768). https://doi.org/10.1098/RSTB.2018.0174
- Huang, V., Bowden, R. M., & Crews, D. (2013). Yolk-albumen testosterone in a lizard with
 temperature-dependent sex determination: Relation with development. *General and Comparative Endocrinology*, 186, 67–71. https://doi.org/10.1016/j.ygcen.2013.02.019
- Huey, R. B., & Hertz, P. E. (1984). Effects of body size and slope on acceleration of a lizard
 (*Stellio stellio*). Journal of Experimental Biology, 110(1), 113–123.
- 553 https://doi.org/10.1242/jeb.110.1.113
- Iglesias-Carrasco, M., Head, M. L., & Cabido, C. (2016). Habitat dependent effects of
 experimental immune challenge on lizard anti-predator responses. *Behavioral Ecology and Sociobiology*, 70(11), 1931–1939. https://doi.org/10.1007/s00265-016-2199-7
- Lancaster, L. T., McAdam, A. G., & Sinervo, B. (2010). Maternal adjustment of egg size
 organizes alternative escape behaviors, promoting adaptive phenotypic integration.
 Evolution, 64(6), 1607–1621. https://doi.org/10.1111/j.1558-5646.2009.00941.x
- Langkilde, T., & Shine, R. (2005). Different optimal offspring sizes for sons versus daughters
 may favor the evolution of temperature-dependent sex determination in viviparous
 lizards. *Evolution*, 59(10), 2275. https://doi.org/10.1554/05-239.1
- Martin, J., & López, P. (1999). When to come out from a refuge: Risk sensitive and statedependent decisions in an alpine lizard. *Behavioral Ecology*, 10(5), 487–492.
 https://doi.org/10.1093/beheco/10.5.487
- McCarty, J. P. (2001). Ecological consequences of recent climate change. *Conservation Biology*, 15(2), 320–331. https://doi.org/10.1046/j.1523-1739.2001.015002320.x
- McDonald, S., & Schwanz, L. E. (2018). Thermal parental effects on offspring behaviour and
 their fitness consequences. *Animal Behaviour*, 135, 45–55.
 https://doi.org/10.1016/j.anbehav.2017.11.007
- Mcghee, K. E., Pintor, L. M., Suhr, E. L., & Bell, A. M. (2012). Maternal exposure to
 predation risk decreases offspring antipredator behaviour and survival in threespined
 stickleback. *Functional Ecology*, 26(4), 932–940. https://doi.org/10.1111/j.13652435.2012.02008.x
- McRae, B. H., Schumaker, N. H., McKane, R. B., Busing, R. T., Solomon, A. M., & Burdick,
 C. A. (2008). A multi-model framework for simulating wildlife population response to
 land-use and climate change. *Ecological Modelling*, *219*(1–2), 77–91.
 https://doi.org/10.1016/j.ecolmodel.2008.08.001
- Noble, D. W. A., Mcfarlane, S. E., Keogh, J. S., & Whiting, M. J. (2014). Maternal and
 additive genetic effects contribute to variation in offspring traits in a lizard. *Behavioral Ecology*, 25(3), 633–640. https://doi.org/10.1093/beheco/aru032
- 582 Noble, D. W. A., & Nakagawa, S. (2021). Planned missing data designs and methods:
- 583 Options for strengthening inference, increasing research efficiency and improving

- animal welfare in ecological and evolutionary research. *Evolutionary Applications*,
 14(8), 1958–1968. https://doi.org/10.1111/eva.13273
- Noble, D. W. A., Stenhouse, V., & Schwanz, L. E. (2018). Developmental temperatures and
 phenotypic plasticity in reptiles: a systematic review and meta-analysis. *Biological Reviews*, 93(1), 72–97. https://doi.org/10.1111/brv.12333
- 589 O'Dea, R. E., Noble, D. W. A., & Nakagawa, S. (2022). Unifying individual differences in
 590 personality, predictability and plasticity: A practical guide. *Methods in Ecology and*591 *Evolution*, 13(2), 278–293. https://doi.org/10.1111/2041-210X.13755
- Oostra, V., Saastamoinen, M., Zwaan, B. J., & Wheat, C. W. (2018). Strong phenotypic
 plasticity limits potential for evolutionary responses to climate change. *Nature Communications 2018 9:1, 9*(1), 1–11. https://doi.org/10.1038/s41467-018-03384-9
- Pearson, P. R., & Warner, D. A. (2016). Habitat- and season-specific temperatures affect
 phenotypic development of hatchling lizards. *Biology Letters*, *12*(10), 20160646.
 https://doi.org/10.1098/rsb1.2016.0646
- Qualls, F. J., & Shine, R. (1998). Geographic variation in lizard phenotypes: Importance of
 the incubation environment. *Biological Journal of the Linnean Society*, 64(4), 477–491.
 https://doi.org/10.1006/bijl.1998.0236
- Räsänen, K., Laurila, A., & Merilä, J. (2005). Maternal investment in egg size: Environmentand population-specific effects on offspring performance. *Oecologia*, 142(4), 546–553.
 https://doi.org/10.1007/s00442-004-1762-5
- Raynal, R. S., Noble, D. W. A., Riley, J. L., Senior, A. M., Warner, D. A., While, G. M., &
 Schwanz, L. E. (2022). Impact of fluctuating developmental temperatures on phenotypic
 traits in reptiles: a meta-analysis. *Journal of Experimental Biology*, 225, jeb243369.
 https://doi.org/10.1242/jeb.243369
- Rosenfeld, C. S., & Roberts, R. M. (2004). Maternal diet and other factors affecting offspring
 sex ratio: A review. *Biology of Reproduction*, 71(4), 1063–1070.
 https://doi.org/10.1095/biolreprod.104.030890
- Rutstein, A. N., Gilbert, L., Slater, P. J. B., & Graves, J. A. (2005). Sex-specific patterns of
 yolk androgen allocation depend on maternal diet in the zebra finch. *Behavioral Ecology*, 16(1), 62–69. https://doi.org/10.1093/beheco/arh123
- Saaristo, M., Brodin, T., Balshine, S., Bertram, M. G., Brooks, B. W., Ehlman, S. M.,
 McCallum, E. S., Sih, A., Sundin, J., Wong, B. B. M., & Arnold, K. E. (2018). Direct
 and indirect effects of chemical contaminants on the behaviour, ecology and evolution
 of wildlife. *Proceedings of the Royal Society B: Biological Sciences*, 285(1885),
 20181297. https://doi.org/10.1098/rspb.2018.1297
- Sanger, T. J., Kyrkos, J., Lachance, D. J., Czesny, B., & Stroud, J. T. (2018). The effects of
 thermal stress on the early development of the lizard *Anolis sagrei*. *Journal of Experimental Zoology Part A: Ecological and Integrative Physiology*, 329(4–5), 244–

622 251. https://doi.org/10.1002/jez.2185

- Sharda, S., Zuest, T., Erb, M., & Taborsky, B. (2021). Predator-induced maternal effects
 determine adaptive antipredator behaviors via egg composition. *Proceedings of the National Academy of Sciences of the United States of America*, 118(37), e2017063118.
 https://doi.org/10.1073/pnas.2017063118
- Sinervo, B. (1990). The evolution of maternal investment in lizards: an experimental and
 comparative analysis of egg size and its effects on offspring performance. *Evolution*,
 44(2), 279–294.
- 630 Sinervo, B., & Basolo, A. (1996). Testing adaptation using phenotypic manipulations. In M.
 631 Rose & G. Lauder (Eds.), *Adaptation* (pp. 149–185). CA: Academic.
- 632 Sinervo, B., Doughty, P., Huey, R. B., & Zamudio, K. (1992). Allometric engineering: a
 633 causal analysis of natural selection on offspring size. *Science*, 258, 1927–1930.

- 634 Singh, S. K., Das, D., & Rhen, T. (2020). Embryonic temperature programs phenotype in
 635 reptiles. *Frontiers in Physiology*, 11, 35. https://doi.org/10.3389/fphys.2020.00035
- Slipiński, P., Trigos-Peral, G., Maák, I., Wojciechowska, I., & Witek, M. (2021). The
 influence of age and development temperature on the temperature-related foraging risk
 of Formica cinerea ants. *Behavioral Ecology and Sociobiology*, 75(7), 107.
- 639 https://doi.org/10.1007/s00265-021-03029-w
- 640 Team, S. D. (2020). *RStan: The R interface to Stan.*
- Thompson, M. B., Speake, B. K., Russell, K. J., & McCartney, R. J. (2001). Utilisation of
 lipids, protein, ions and energy during embryonic development of Australian oviparous
 skinks in the genus *Lampropholis*. *Comparative Biochemistry and Physiology A*, *129*,
 313–326. https://doi.org/10.1016/S1095-6433(00)00349-4
- Tiatragul, S., Kurniawan, A., Kolbe, J. J., & Warner, D. A. (2017). Embryos of non-native
 anoles are robust to urban thermal environments. *Journal of Thermal Biology*, 65, 119–
 124. https://doi.org/10.1016/j.jtherbio.2017.02.021
- Valenzuela, N., & Lance, V. A. (2004). *Temperature-Dependent Sex Determination in Vertebrates.* Smithsonian Books.
- Warner, D. A., & Andrews, R. M. (2002). Laboratory and field experiments identify sources
 of variation in phenotypes and survival of hatchling lizards. *Biological Journal of the Linnean Society*, 76(1), 105–124. https://doi.org/10.1046/j.1095-8312.2002.00054.x
- Warner, D. A., & Lovern, M. B. (2014). The maternal environment affects offspring viability
 via an indirect effect of yolk investment on offspring size. *Physiological and Biochemical Zoology*, 87(2), 276–287. https://doi.org/10.1086/674454
- Warner, D. A., Lovern, M. B., & Shine, R. (2007). Maternal nutrition affects reproductive
 output and sex allocation in a lizard with environmental sex determination. *Proceedings of the Royal Society B: Biological Sciences*, 274(1611), 883–890.
 https://doi.org/10.1098/rspb.2006.0105
- Webb, J. K., Brown, G. P., & Shine, R. (2001). Body size, locomotor speed and antipredator
 behaviour in a tropical snake (*Tropidonophis mairii*, colubridae): The influence of
 incubation environments and genetic factors. *Functional Ecology*, 15(5), 561–568.
 https://doi.org/10.1046/j.0269-8463.2001.00570.x
- Webb, J. K., Shine, R., & Christian, K. A. (2006). The adaptive significance of reptilian
 viviparity in the tropics: testing the maternal manipulation hypothesis. *Evolution*, 60,
 115. https://doi.org/10.1554/05-460.1
- While, G. M., Noble, D. W. A., Uller, T., Warner, D. A., Riley, J. L., Du, W. G., & Schwanz,
 L. E. (2018). Patterns of developmental plasticity in response to incubation temperature
 in reptiles. *Journal of Experimental Zoology Part A: Ecological and Integrative Physiology*, 329(4–5), 162–176. https://doi.org/10.1002/jez.2181
- Wilson, S., & Swan, G. (2010). A complete guide to reptiles of Australia (3rd ed.). Reed New
 Holland, Sydney.
- Wolf, J. B., & Wade, M. J. (2009). What are maternal effects (and what are they not)? *Philosophical Transactions of the Royal Society B: Biological Sciences*, 364(1520),
 1107–1115. https://doi.org/10.1098/rstb.2008.0238
- Yeh, P. J., & Price, T. D. (2004). Adaptive phenotypic plasticity and the successful colonization of a novel environment. *American Naturalist*, *164*(4), 531–542.
 https://doi.org/10.1086/423825
- 679
- 680
- 681





685 Figure 1. Effect of temperature (cold, 23 °C vs hot, 28 °C) and maternal resource investment (yolk reduced vs control) in morphological traits of *L. delicata* (a, b and c) and *L. guichenoti*

- (d, e and f).





Figure 2. Effect of temperature (cold, 23 °C vs hot, 28 °C) and maternal resource investment (yolk reduced vs control) in lizard performance: distance moved and running speed (25 cm burnst and 1 m). L. delicata panels a, b and c, and L. guichenoti panels d, e and f. Note: 3 datapoints (raw data > 10s and < 0.01s) in the 25 burst of *L*. *delicata* were removed for visualization reasons.



698 699

700 Figure 3. Effect of temperature (cold, 23 °C vs hot, 28 °C) and maternal resource investment

701 (yolk reduced, control) in lizard antipredator behaviour: time hiding (time, in seconds, since
702 the lizard hided in the refuge until the head was visible) and time to active (time before

resuming activity). *L. delicata* panels a and b, and *L. guichenoti* panels c and d.

Tables

Table 1– Posterior means and 95% credible intervals for the interaction between temperature (Temp) and maternal investment (Invest) along with the main effects of temperature and maternal investment on morphological traits for *Lampropholis delicata* and *Lampropholis guichenoti*. Main effects are pooled posterior means over each level of the second predictor variable (either temperature or maternal investment treatments depending on the focal variable). Posterior distributions are estimated from a multi-response model that accounts for the correlation between morphological traits. Bold indicates significant effects.

Species	Trait	Term	Estimate	2.5%	97.5%	рМСМС
L. delicata	SVL	Interaction [(C23 - A23) - (C28 - A28)]	-0.198	-1.651	1.288	0.790
		Temp (23-28)	-0.192	-1.327	0.913	0.738
		Invest (C-A)	0.404	-0.663	1.442	0.438
	Weight	Interaction [(C23 - A23) - (C28 - A28)]	0.010	-0.035	0.057	0.665
		Temp (23-28)	-0.006	-0.042	0.031	0.728
		Invest (C-A)	0.002	-0.031	0.038	0.918
	Tail	Interaction [(C23 - A23) - (C28 - A28)]	-2.898	-6.238	0.516	0.089
		Temp (23-28)	-1.299	-4.979	2.128	0.565
		Invest (C-A)	-0.777	-4.348	2.475	0.768
L. guichenoti	SVL	Interaction [(C23 - A23) - (C28 - A28)]	1.324	-0.501	3.124	0.150
		Temp (23-28)	-2.973	-4.715	-1.241	0.000
		Invest (C-A)	0.348	-1.394	2.072	0.739
	Weight	Interaction [(C23 - A23) - (C28 - A28)]	0.089	0.006	0.172	0.035
		Temp (23-28)	-0.112	-0.207	-0.019	0.012
		Invest (C-A)	0.014	-0.080	0.108	0.862
	Tail	Interaction [(C23 - A23) - (C28 - A28)]	2.174	-1.420	5.776	0.241
		Temp (23-28)	-6.347	-9.619	-3.079	0.000
		Invest (C-A)	0.902	-2.361	4.147	0.627

Table 22 – Posterior means and 95% credible intervals for the interaction between temperature (Temp) and maternal investment (Invest) along with the main effects of temperature and maternal investment on behavioural and performance traits for *Lampropholis delicata* and *Lamprpholis guichenoti*. Main effects are pooled posterior means over each level of second predictor variable. Estimates are from a Bayesian multivariate (multi-response) model not controlling for SVL. See Supplement for model with SVL controlled. Bold estimates are significant and italics indicated effects with less than a 10% chance of being observed.

Species	Trait	Term	Estimate	Q2.5	Q97.5	рМСМС
L. delicata	Time to Activity (s)	Interaction [(C23 - A23) - (C28 - A28)]	-864.379	-1,847.746	103.120	0.080
		Temp (23-28)	-676.650	-1,696.061	325.761	0.239
		Invest (C-A)	233.230	-826.597	1,216.182	0.731
	Hiding Time (s)	Interaction [(C23 - A23) - (C28 - A28)]	-718.025	-1,631.020	192.257	0.117
		Temp (23-28)	-502.792	-1,406.149	388.546	0.328
		Invest (C-A)	115.504	-810.196	985.614	0.830
	Distance Moved (cm)	Interaction [(C23 - A23) - (C28 - A28)]	35.435	-174.231	252.904	0.748
		Temp (23-28)	33.968	-121.384	187.916	0.660
		Invest (C-A)	-50.048	-204.844	109.218	0.528
	log 1m Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	0.335	-0.023	0.689	0.068
		Temp (23-28)	-0.028	-0.405	0.349	0.928
		Invest (C-A)	-0.262	-0.644	0.116	0.233
	log Burst Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	0.214	-0.178	0.591	0.279
		Temp (23-28)	-0.062	-0.398	0.270	0.737
		Invest (C-A)	-0.095	-0.435	0.238	0.610
L. guichenoti	Time to Activity (s)	Interaction [(C23 - A23) - (C28 - A28)]	-93.237	-1,263.710	1,052.332	0.876
		Temp (23-28)	-42.896	-870.269	778.343	0.929
		Invest (C-A)	133.420	-681.068	951.159	0.750
	Hiding Time (s)	Interaction [(C23 - A23) - (C28 - A28)]	166.056	-694.735	989.883	0.683
		Temp (23-28)	-2.416	-631.727	619.633	0.992
		Invest (C-A)	90.790	-519.904	697.379	0.767
	Distance Moved (cm)	Interaction [(C23 - A23) - (C28 - A28)]	127.921	-185.868	444.379	0.423
		Temp (23-28)	4.077	-249.201	263.186	0.976
		Invest (C-A)	-25.340	-282.031	232.134	0.843
	log 1m Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	-0.179	-0.549	0.206	0.348

Species	Trait	Term	Estimate	Q2.5	Q97.5	рМСМС
		Temp (23-28)	0.116	-0.198	0.432	0.489
		Invest (C-A)	-0.027	-0.347	0.281	0.878
	log Burst Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	-0.470	-0.854	-0.081	0.018
		Temp (23-28)	0.134	-0.327	0.603	0.774
		Invest (C-A)	-0.034	-0.502	0.424	0.953

Supplementary material for

Maternal resources and early thermal conditions affect performance and antipredator responses Supplementary results

Table S1. Re	epeatability	of the thre	e performanc	e and antip	redatory	behaviour measures.
--------------	--------------	-------------	--------------	-------------	----------	---------------------

		L. delicata		l	. guichenoi	ti
	R	1	U	R	1	u
Hiding time	0.2807	0.0222	0.5025	0.2391	0.0612	0.4072
Time to activity	0.0724	0.0001	0.2431	0.2315	0.0307	0.4093
Distance moved	0.2644	0.0406	0.4859	0.4824	0.3425	0.6092
Running velocity 1m	0.4580	0.3128	0.5893	0.5579	0.4339	0.6698
Running velocity 25cm	0.3440	0.2055	0.4812	0.6310	0.5223	0.7294

Table S2. Within-individual correlations between morphological traits estimated from Bayesian multivariate models for <u>morphology</u> in *Lampropholis delicata and L. guichenoti*.

L. delicata

Within-individual correlations in morphological traits

Estimate SE 1-95% CI U-95% CI Rhat Bulk_ESS Tail_ESS

rescor(SVL, Weight)	0.8	0.04	0.7	0.87	1	12001	10099
rescor(SVL, Tail length)	0.53	0.09	0.33	0.7	1	11828	10260
rescor(Weight, Tail length)	0.56	0.09	0.36	0.72	1	12109	10709

L. guichenoti

Within-individual correlations in morphological traits

	Estimate	Est.Error	l-95% CI	U-95% CI	Rhat	Bulk_ESS	Tail_ESS
rescor(SVL, Weight)	0.69	0.06	0.55	0.8	1	6372	7408
rescor(SVL, Tail length)	0.63	0.08	0.45	0.77	1	5176	6727
rescor(Weight, Tail length)	0.88	0.04	0.79	0.93	1	3383	6556

Table 3S3. Posterior means and 95% credible intervals for the interaction between temperature (Temp) and maternal investment (Invest) along with the main effects of temperature and maternal investment on behavioural and performance traits for *Lampropholis delicata* and *Lampropholis guichenoti*. Main effects are pooled posterior means over each level of second predictor variable. Estimates are from a Bayesian multivariate (multi-response) model controlling for SVL (Z-transformed).

Species	Trait	Term	Estimate	Q2.5	Q97.5	рМСМС
L. delicata	Time to activity (s)	Interaction [(C23 - A23) - (C28 - A28)]	-1,003.752	-1,988.452	-33.590	0.044
		Temp (23-28)	-468.643	-1,589.133	597.340	0.543
		Invest (C-A)	122.220	-1,014.488	1,158.751	0.873
	Hiding time (s)	Interaction [(C23 - A23) - (C28 - A28)]	-896.713	-1,803.445	24.917	0.057
		Temp (23-28)	-423.236	-1,444.311	558.994	0.544
		Invest (C-A)	62.101	-961.269	1,022.564	0.922
	Distance Moved (cm)	Interaction [(C23 - A23) - (C28 - A28)]	54.939	-174.283	295.328	0.642
		Temp (23-28)	56.787	-120.822	231.572	0.522
		Invest (C-A)	-35.953	-206.360	140.925	0.668
	log 1m Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	0.312	-0.071	0.692	0.109
		Temp (23-28)	-0.054	-0.439	0.334	0.826
		Invest (C-A)	-0.267	-0.646	0.109	0.198
	log Burst Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	0.184	-0.206	0.577	0.352
		Temp (23-28)	-0.129	-0.454	0.210	0.457
		Invest (C-A)	-0.098	-0.423	0.223	0.566
L. guichenoti	Time to activity (s)	Interaction [(C23 - A23) - (C28 - A28)]	-105.815	-1,301.414	1,101.039	0.863
		Temp (23-28)	43.518	-893.814	998.022	0.935
		Invest (C-A)	108.002	-750.916	976.971	0.809
	Hiding time (s)	Interaction [(C23 - A23) - (C28 - A28)]	298.455	-574.277	1,159.178	0.499
		Temp (23-28)	-65.151	-808.196	660.997	0.867
		Invest (C-A)	49.173	-628.295	718.879	0.884
	Distance Moved (cm)	Interaction [(C23 - A23) - (C28 - A28)]	130.568	-198.951	467.296	0.440
		Temp (23-28)	8.658	-287.891	299.148	0.943
		Invest (C-A)	-53.818	-323.685	212.532	0.697
	log 1m Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	-0.135	-0.528	0.265	0.493
		Temp (23-28)	0.024	-0.307	0.358	0.887
		Invest (C-A)	-0.008	-0.318	0.301	0.962

Species	Trait	Term	Estimate	Q2.5	Q97.5	рМСМС
	log Burst Speed (cm/s)	Interaction [(C23 - A23) - (C28 - A28)]	-0.413	-0.810	-0.006	0.046
		Temp (23-28)	-0.015	-0.483	0.452	0.956
		Invest (C-A)	0.008	-0.432	0.455	0.991

Table 4S4. Posterior means and 95% credible intervals for the between and within-individual correlations among behavioural traits in *Lampropholis delicata* and *Lampropholis guichenoti*. Estimates are from a Bayesian multivariate (multi-response) model. Bold estimates indicate ones where the 95% credible interval does not overlap zero.

Species - level	Correlation	Estimate	2.5 % CI	97.5 % CI
L. delicata - Between Individual	cor(Time to Activity (s), Hiding Time(s))	0.29159827	-0.60960494	0.868097048
	cor(Time to Activity (s), Distance Moved (cm))	-0.13655106	-0.72829302	0.620152880
	cor(Hiding Time(s), Distance Moved (cm))	-0.13688198	-0.70275908	0.579493448
	cor(Time to Activity (s), 1m Speed (cm/s))	0.27195884	-0.38825792	0.776064363
	cor(Hiding Time(s), 1m Speed (cm/s))	0.15041240	-0.46974706	0.688090745
	cor(Distance Moved (cm), 1m Speed (cm/s))	0.24238409	-0.13979502	0.628382729
	cor(Time to Activity (s), Burst Speed (cm/s))	0.22520672	-0.46104064	0.762376365
	cor(Hiding Time(s), Burst Speed (cm/s))	0.19915158	-0.42866231	0.739702151
	cor(Distance Moved (cm), Burst Speed (cm/s))	0.26857069	-0.12762202	0.664960881
	cor(1m Speed (cm/s), Burst Speed (cm/s))	0.82102166	0.62546459	0.952074963
L. delicata - Within Individual	cor(Time to Activity (s), Hiding Time(s))	0.81659503	0.73344677	0.880468668
	cor(Time to Activity (s), Distance Moved (cm))	-0.15311562	-0.33138147	0.034234536
	cor(Hiding Time(s), Distance Moved (cm))	-0.07303817	-0.24215358	0.098905974
	cor(Time to Activity (s), 1m Speed (cm/s))	0.07619062	-0.11077008	0.260270547
	cor(Hiding Time(s), 1m Speed (cm/s))	0.01950419	-0.15776967	0.196633984
	cor(Distance Moved (cm), 1m Speed (cm/s))	0.01958401	-0.13491722	0.169922742
	cor(Time to Activity (s), Burst Speed (cm/s))	0.02807736	-0.18700479	0.239544933
	cor(Hiding Time(s), Burst Speed (cm/s))	-0.04548645	-0.24981026	0.159714536
	cor(Distance Moved (cm), Burst Speed (cm/s))	0.01922480	-0.12981225	0.169160469
	cor(1m Speed (cm/s), Burst Speed (cm/s))	0.42413076	0.29703469	0.542793049
L. guichenoti - Between Individual	cor(Time to Activity (s), Hiding Time(s))	0.90369565	0.77529782	0.978227159
_	cor(Time to Activity (s), Distance Moved (cm))	-0.45171409	-0.69358613	-0.177708817
	cor(Hiding Time(s), Distance Moved (cm))	-0.29881986	-0.57921282	-0.002829739
	cor(Time to Activity (s), 1m Speed (cm/s))	0.24722197	-0.03334559	0.511867690
	cor(Hiding Time(s), 1m Speed (cm/s))	0.19628177	-0.08423812	0.469426798
	cor(Distance Moved (cm), 1m Speed (cm/s))	-0.03441905	-0.33088469	0.271057143
	cor(Time to Activity (s), Burst Speed (cm/s))	0.31610099	0.03933083	0.571799305
	cor(Hiding Time(s), Burst Speed (cm/s))	0.25109151	-0.02586077	0.510175923
	cor(Distance Moved (cm), Burst Speed (cm/s))	-0.15763758	-0.43004251	0.126905402
	cor(1m Speed (cm/s), Burst Speed (cm/s))	0.95286559	0.89131640	0.989537957
L. guichenoti - Within Individual	cor(Time to Activity (s), Hiding Time(s))	0.68721300	0.58149348	0.771981041
	cor(Time to Activity (s), Distance Moved (cm))	-0.03812672	-0.20063612	0.121080431
	cor(Hiding Time(s), Distance Moved (cm))	0.06659344	-0.08967582	0.221268294

Species - level	Correlation	Estimate	2.5 % CI	97.5 % CI
	cor(Time to Activity (s), 1m Speed (cm/s))	-0.01538997	-0.16973859	0.140063113
	cor(Hiding Time(s), 1m Speed (cm/s))	-0.04596171	-0.19493574	0.108994499
	cor(Distance Moved (cm), 1m Speed (cm/s))	-0.03168564	-0.17659253	0.116113479
	cor(Time to Activity (s), Burst Speed (cm/s))	-0.04886429	-0.21126303	0.117981964
	cor(Hiding Time(s), Burst Speed (cm/s))	-0.02058361	-0.17741319	0.138176209
	cor(Distance Moved (cm), Burst Speed (cm/s))	0.04028649	-0.10947845	0.189531771
	cor(1m Speed (cm/s), Burst Speed (cm/s))	0.50825642	0.39813324	0.609206642