

A Perspective on How Glyphosate and 2,4-D May Impact Climate Change

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10 **Abstract**

11 An increase in herbicide use is occurring due to a growing population and herbicide-resistant crops in
12 agriculture, which has resulted in more herbicide tolerant target species. Glyphosate and 2,4-
13 Dichlorophenoxyacetic acid (2,4-D) are two of the most commonly used herbicides worldwide and
14 are more recently being used in combination in pre-mixed commercial formulas. Subsequently,
15 herbicide contamination of wetlands will increase subjecting microorganisms to multiple chemical
16 stressors. Methane is a potent greenhouse gas naturally emitted from wetlands, but herbicides may
17 disrupt biogeochemical processes leading to an unbalanced methane cycle. This perspective
18 examined the potential effects of herbicides on climate change using glyphosate and 2,4-D as a case
19 study. We highlighted previous research on glyphosate-derived nutrient enrichment and 2,4-D
20 inhibition of methane oxidation. We also explained how the concurrent effects of these herbicides
21 could alter microbial communities leading to increased methane production in wetlands. Our
22 perspective elucidates the potential ecosystem-level implications of herbicides in wetlands, in
23 addition to stating the importance for research on the combined effects of herbicides.

24 **1 Introduction**

25 Climate change is an ongoing global concern as greenhouse gas (GHG) emissions continue to
26 increase (IPCC, 2021). Methane (CH₄) is the second most abundant GHG, after carbon dioxide
27 (CO₂), but is about 25 times more potent (Islam et al., 2018; EPA, 2023). Wetlands are a large
28 natural source of CH₄ as they play a significant role in carbon (C) sequestration and cycling
29 (Andresen et al., 2017). It has recently been suggested that GHG emissions from freshwater
30 ecosystems may be impacted by agrochemical use (Stehle and Schulz, 2015), which has substantially
31 increased over the past 30 years due to the introduction of herbicide-resistant crops. Herbicide-
32 resistant crops now comprise more than 90% of crops in the United States (USDA, 2022) and are
33 increasingly used worldwide (Peterson et al., 2018). This has unexpectedly led to an increase in

34 herbicide use (Bai and Ogbourne, 2016; Coupe and Capel, 2016), and subsequently weeds are
35 chronically exposed to herbicide residues resulting in the development of herbicide tolerance (Heap
36 and Duke, 2018). For example, the introduction of Roundup Ready[®] (i.e., glyphosate-resistant) crops
37 resulted in a 15-fold increase in glyphosate use (Coupe and Capel, 2016; Hébert et al., 2019), and
38 consequently, approximately 41 weed species worldwide have developed glyphosate tolerance
39 (Jeschke and Teten, 2018). Ongoing climatic change is projected to result in intensified herbicide use
40 (Delcour et al., 2015), creating a positive feedback loop if increasing herbicide use results in further
41 CH₄ emissions. Temperature increases can also enhance toxicity and alter biodegradation processes
42 of herbicides (Noyes et al., 2009; Koleva and Schneider, 2010; Matzrafi, 2019). Subsequently, as
43 wetlands are subjected to a combination of more severe stressors, wetland biota, specifically
44 microorganisms may be detrimentally affected. Pre-mixed herbicides containing multiple active
45 ingredients (i.e., multiple modes of action), such as glyphosate and 2,4-Dichlorophenoxyacetic acid
46 (2,4-D), are also now more commonly used to combat target species that have developed tolerance to
47 single active ingredient herbicides (Freydier and Lundgren, 2016; Schütte et al., 2017). In this article
48 we will present a case study on how glyphosate and 2,4-D accumulation in wetlands could impact
49 climate change by focusing on their potential synergistic effects on microbial communities.
50 Specifically, understanding their impacts on CH₄ dynamics is critical to understanding the role of
51 wetlands in contributing to climate change.

52 **2 Case Study**

53 Glyphosate and 2,4-D are two of the most commonly used herbicides and are also used in pre-
54 mixed formulas such as Enlist Duo[®] (1:0.95 glyphosate:2,4-D) and Landmaster[™] II (1:0.83
55 glyphosate:2,4-D) (Benbrook, 2016; Zuanazzi et al., 2020; EPA, 2022). Extensive use of glyphosate
56 and 2,4-D are cause for environmental concern within aquatic ecosystems where herbicides are
57 substantial contributors to wetland pollution (Casado et al., 2019). As wetlands are often located in
58 the lowest drainage points of agricultural fields, they can serve as critical sinks for herbicide residues
59 transported through spray drift, runoff, groundwater leaching, and wind and sediment erosion (Annett
60 et al., 2014; Bento et al., 2017). Subsequently, this can result in bioavailable residues, where
61 microorganisms are subjected to a “pesticide cocktail” (Aparicio et al., 2013; Islam et al., 2018).
62 Microorganisms, such as prokaryotes and algae, are important contributors to wetland ecosystem
63 functioning, including mediating biogeochemical cycling (Sun et al., 2013; Baker et al., 2020). The C
64 biogeochemical processes of methanogens (i.e., CH₄ producers) and methanotrophs (i.e., CH₄
65 consumers), in addition to plant and algal-mediated transport, play a critical role in the global CH₄

66 budget of wetlands. Agrochemical contributions to increased GHG emissions (Stehle and Schulz,
67 2015) may be explained by an unbalanced CH₄ cycle via impacts to CH₄-associated microorganisms.
68 The range of effects of glyphosate and 2,4-D on wetland microorganisms can vary including direct
69 toxicity, nutrient enrichment, and alterations to metabolic and catabolic processes.

70 **2.1 Glyphosate**

71 Glyphosate's potential impacts on wetland microorganisms can be detrimental or advantageous
72 to microbial species. While glyphosate's mode of action was developed to target the shikimate
73 pathway in higher plants (Hetrick and Blankinship, 2015), many archaea and bacteria also utilize this
74 pathway resulting in non-target effects (Herrmann and Weaver, 1999). Despite the potential negative
75 impacts on microorganisms, in many instances increased growth, respiration, and enhanced
76 metabolism in wetland microbial communities have been observed as a result of glyphosate
77 biodegradation (Vera et al., 2012; Lu et al., 2020) and linked with the use of glyphosate as a nutritive
78 source (Saxton et al., 2011; Wang et al., 2016). Due to glyphosate's chemical structure, its
79 degradation often contributes substantial amounts of phosphorus (P), which has been found to be
80 favored and more rapidly utilized by microorganisms compared to other sources of soil P (Hébert et
81 al., 2019; Sun et al., 2019). Specifically, stimulated cyanobacterial growth and cyanotoxin production
82 has been recorded from glyphosate-derived P enrichment (Vera et al., 2010; Qiu et al., 2013; Zhang
83 et al., 2016; Hernández-García and Martínez-Jerónimo, 2020; Wang et al., 2021; Lin et al., 2023).
84 Glyphosate degradation was found to be positively correlated with total P concentrations in surface
85 waters (Carles et al., 2019). Glyphosate additions to aquatic ecosystems can contribute to water
86 quality issues, such as eutrophication, which has been demonstrated to be an important driver of CH₄
87 emissions (Sepulveda-Jauregui et al., 2018; Beaulieu et al., 2019; Yang et al., 2019; Bertolet et al.,
88 2020). In addition, glyphosate-derived P loading into wetlands could stimulate algal biomass leading
89 to eutrophic conditions, which could then increase CH₄ production (Carles et al., 2019). Ultimately,
90 increased glyphosate use could shift microbial community dynamics towards copiotrophs and algae,
91 altering important C biogeochemical processes and resulting in an indirect increase in CH₄
92 production in wetlands (Figure 1, panels A and B).

93 **2.2 2,4-D**

94 Despite 2,4-D being the first synthetic herbicide, compared to glyphosate, relatively little
95 research has been conducted on its effects on aquatic microorganisms (Donald et al., 2018; Malaj et
96 al., 2020). However, similar to glyphosate, 2,4-D can have a variety of impacts on wetland microbial

97 communities. It targets broadleaf plants through mimicking the plant growth hormone, indol-3-yl-
98 acetic acid (IAA or auxin), resulting in plant overgrowth (Cobb and Reade, 2010), but auxin
99 synthesis and usage in microorganisms is also well known making them vulnerable non-target
100 organisms (Spaepen and Vanderleyden, 2011). In addition, 2,4-D is a widespread environmental
101 contaminant frequently detected in aquatic ecosystems (Malaj et al., 2020), and its use has increased
102 in recent decades with the development of herbicide-resistant crops, where its use will likely continue
103 to increase in the future (Freydier and Lundgren, 2016). Consequently, wetland microorganisms
104 could be highly susceptible to its toxic effects with limited capacity to degrade it. Previous research
105 has found some species use 2,4-D as a C source, whereas other species are toxicologically inhibited
106 (Benndorf et al., 2004; Zabaloy et al., 2008; Sachu et al., 2022). Research in microcosms has also
107 found that increased 2,4-D concentrations resulted in inhibition of CH₄ oxidation, decreases in CH₄
108 removal time, and increased CH₄ emissions (Syamsul Arif et al., 1996; Kumaraswamy et al., 1997;
109 Top et al., 1999). Where studies from Top et al. (1999) and Seghers et al. (2003) suggested decreases
110 in CH₄ removal could be due to 2,4-D inhibition of methanotroph-mediated oxidative metabolism.
111 Research on the effects of 2,4-D on CH₄ oxidation is extremely limited, however these studies do
112 indicate that 2,4-D loading into wetlands could potentially alter the CH₄ cycle by suppressing the
113 removal of CH₄ via the food web, resulting in greater concentrations within the water column and
114 higher emissions (Figure 1, panels C and D).

115 **2.3 Pesticide cocktails: Glyphosate plus 2,4-D**

116 The increased use of pre-mixed glyphosate and 2,4-D herbicides further exposes wetland
117 microorganisms to combinations of chemical stressors, which could lead to unforeseen long-term
118 effects. Research on the combined effects of pesticides has been conducted since the 1970's, but the
119 majority of the focus has been on the direct toxicological impacts to aquatic flora and fauna
120 (Lichtenstein et al., 1973; Faust et al., 1994; Gardner and Grue, 1996; Hayes et al., 2006; Relyea,
121 2009; Moreira et al., 2020). These studies included compounds such as atrazine, chlorpyrifos,
122 fipronil, etc., whereas research on the combined effects of glyphosate and 2,4-D is limited, especially
123 at the aquatic microbial level. Additive and/or synergistic effects of glyphosate and 2,4-D have been
124 found on fish and amphibian growth, fertilization, survival, and behavior (Carvalho et al., 2020;
125 Pavan et al., 2021; Bernardi et al., 2022; Peluso et al., 2022), and zooplankton emergence (Portinho
126 et al., 2018). Lozano et al. (Lozano et al., 2018) found additive impacts of glyphosate and 2,4-D on
127 phytoplankton composition, abundance, and chlorophyll *a* after 7 days in microcosms, but also found
128 an antagonist effect on total and live abundance of *Staurastrum* spp. In outdoor mesocosms Lozano

129 et al. (Lozano et al., 2018) found a decrease in phytoplankton respiration and gross primary
130 production from a high glyphosate (applied as Roundup Max[®]), low 2,4-D (applied as AsiMax 50[®])
131 treatment after 4 hours. Additionally, after 7 days in mesocosms with high glyphosate, an increase in
132 primary production, chlorophyll *a*, and micro- and nanophytoplankton was observed (Lozano et al.,
133 2020). Sura et al. (Sura et al., 2015) researched the effects of a herbicide mixture including
134 glyphosate, 2,4-D, and MCPA, clopyralid, dicamba, dichlorprop, mecoprop, and bromoxynil on
135 pelagic and benthic communities in nutrient-sufficient and nutrient-deficient wetlands. They found
136 pelagic bacterial productivity significantly increased after treatment in the nutrient-sufficient wetland,
137 but benthic bacterial productivity did not change, which suggests the stimulatory effect of these
138 herbicides may be related to nutrient bioavailability. These results demonstrate the complexity of the
139 direct effects of herbicide mixtures on aquatic microorganisms, but the potential indirect effects are
140 still poorly understood. As pre-mixed glyphosate and 2,4-D herbicides become more common it is
141 important to consider the extent of their effects on aquatic ecosystems. Glyphosate can easily be used
142 as a nutrient source stimulating microbial activity, specifically algal communities, whereas 2,4-D
143 may inhibit methanotrophic communities from oxidizing CH₄. As these compounds enter aquatic
144 ecosystems their impacts on microorganisms may become synergistic and/or additive resulting in
145 eutrophication and inhibition of methanotrophs from glyphosate and 2,4-D, respectively.
146 Subsequently, eutrophic conditions and decreased CH₄ removal could cause increased CH₄
147 production via an unbalanced CH₄ cycle (Figure 3).

148 **3 Pollution-Induced Community Tolerance (PICT)**

149 Aquatic ecosystems are subjected to year-round herbicide contamination, where herbicide use
150 differs across crop, season, habitat, and region. Microorganism structure and function can be
151 impacted by herbicides, but toxicity is often dependent on the mode of action, concentration, and
152 duration of exposure, as well as microbial species and environmental factors (DeLorenzo et al.,
153 2001). For example, glyphosate stimulated *Chlorella vulgaris* growth 24-hours after exposure, but
154 then inhibited growth after 48-hours at the same concentrations (i.e., hormesis) (Reno et al., 2014). In
155 addition to the duration of exposure, the exposure to a different mode of action could also impact
156 microorganisms by causing a community shift often appearing as changes in gene expression or
157 diversity (Feld et al., 2015). Pollution-Induced Community Tolerance (PICT) refers to the response
158 of a community to a pollutant, which results in an increased tolerance to that pollutant (Blanck,
159 2002). The use of PICT analysis is extensive in the toxicology literature, especially on phototrophic
160 microorganisms, which are often more susceptible to herbicidal effects due to their similarities with

161 target species (DeLorenzo et al., 2001; Larras et al., 2016). Bérard and Benninghoff (Bérard and
162 Benninghoff, 2001) found phytoplankton were significantly more sensitive to atrazine after one day,
163 but then significantly less sensitive after at least 11 days. Phototrophic biofilms were found to be
164 increasingly more sensitive to diuron as contamination levels decreased 1-3 years after its ban in the
165 European Union (Pesce et al., 2016). It has also been shown that selection pressure from multiple
166 stressors can lead to more opportunistic species and higher tolerances (Rotter et al., 2013).
167 Ultimately, PICT results suggest that more sensitive species are being replaced by less sensitive
168 species creating a more tolerant community (Blanck, 2002). This has also been seen with both
169 glyphosate and 2,4-D. Microbial communities from sediments with high glyphosate exposure were
170 able to degrade glyphosate faster and had higher diversity compared to sediments with low to no
171 previous exposure (Tang et al., 2019). Zabaloy et al. (Zabaloy et al., 2008) saw an increase in a 2,4-D
172 degrading population in soils for approximately one month after treatment and found that agricultural
173 soils had higher 2,4-D tolerance compared to reference soils via PICT analysis. In a study by de
174 Liphay et al. (de Liphay et al., 2002) 2,4-D treatment induced transcription of the gene responsible
175 for 2,4-D degradation (*tfdA*) which demonstrates a survival response from the microbial community.
176 These studies demonstrate PICT can occur when communities are exposed to a herbicide, therefore
177 contamination by an additional herbicide could further alter communities that have not been exposed
178 before. It could be presumed that wetland microbial communities within a glyphosate-dominant
179 region may substantially change when 2,4-D is introduced in combination with glyphosate and
180 species are replaced. This potential shift would impact the biogeochemical functions of the
181 community, subsequently altering herbicide degradation or metabolism.

182 **4 Conclusions**

183 Glyphosate and 2,4-D are frequently cited as having minimal to no environmental impacts
184 (Peterson et al., 2016; Duke, 2020; Singh et al., 2020), however there is increasing evidence that their
185 indirect effects may be of more substantial global concern. Wetlands naturally emit CH₄ via
186 diffusion, ebullition (i.e., bubbles), and plant-mediated transport, and are the highest natural sources
187 of CH₄ in the environment (Aben et al., 2017; Andresen et al., 2017), but emissions may be
188 increasing due to agrochemical use adversely impacting CH₄ sink potential (Seghers et al., 2005).
189 Glyphosate could stimulate microbial processes resulting in increased CH₄ production, in addition to
190 2,4-D inhibiting CH₄ oxidation further resulting in increased CH₄ production. Ultimately, this would
191 lead to higher CH₄ production versus removal from freshwater creating elevated CH₄ in the
192 atmosphere. Due to the widespread and extensive use of glyphosate and 2,4-D, these contaminants

193 are frequently found in wetlands (Islam et al., 2018; Malaj et al., 2020). To our knowledge there has
194 been no research investigating the combined impacts of glyphosate and 2,4-D on wetland microbial
195 communities. The potential bottom-up effects of glyphosate and 2,4-D could be detrimental to a
196 changing climate, thus improving our understanding of how these herbicides can impact GHG
197 emissions is crucial.

198 **4.1 Future Research**

199 To investigate the effects of glyphosate and 2,4-D on CH₄ emissions from freshwater
200 ecosystems, micro- or mesocosm experiments could be conducted. Experiments under controlled
201 conditions could help determine how wetland microbial communities are affected by glyphosate and
202 2,4-D. Specifically, this research would give insight into the CH₄-related mechanisms that may be
203 enhanced or disrupted in microorganisms. In addition to in-lab research, pesticide loading data could
204 be incorporated into GHG models. These data are currently not included in estimations of CH₄
205 emissions from wetlands, but could be an important source of variation, and could be useful for
206 future climate modeling. These potential impacts are crucial to research as herbicide use is only
207 expected to increase over time, where chemical selection pressure on microbial communities could
208 contribute to climate change.

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467 **Conflict of Interest**

468 This manuscript was written in the absence of commercial or financial conflicts of interest.

469 **6 Author Contributions**

470 Christine Cornish generated the first draft of the manuscript with revisions and comments from Jon
471 Sweetman. Both authors have read and approved the submitted version.

472 **7 Funding**

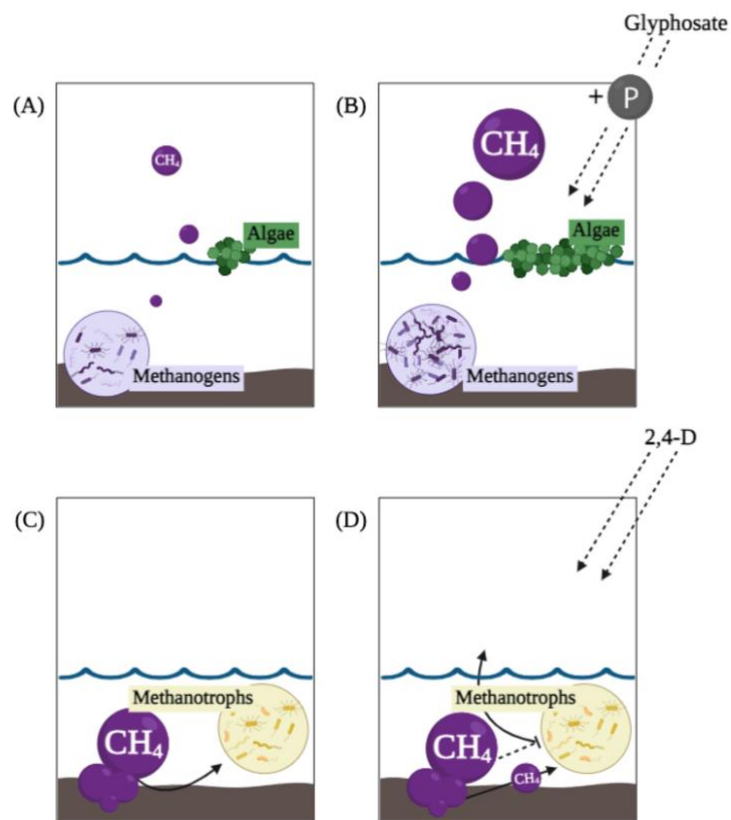
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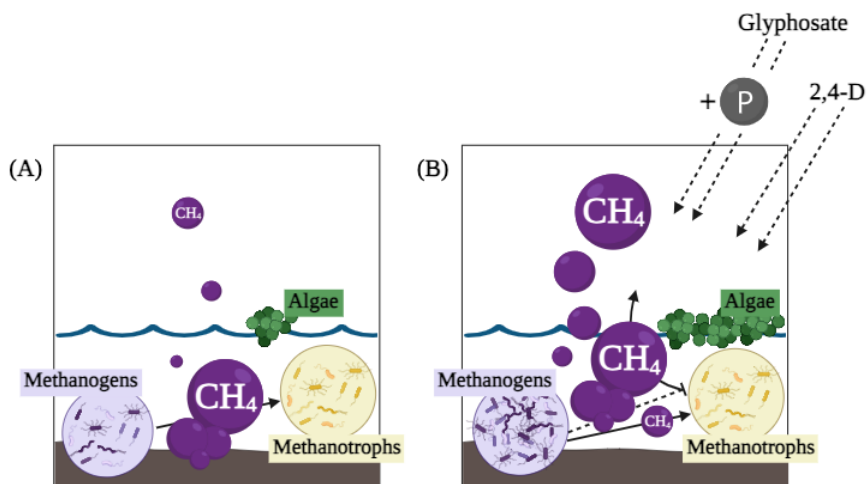
477 **9 Data Availability Statement**

478 Not applicable.



479

480 **Figure 1.** Conceptual diagrams of a wetland with methanogens, methanotrophs, and algae. (A)
 481 represents balanced CH₄ production; (B) represents glycosate contamination stimulating
 482 methanogens and algae causing higher CH₄ emissions; (C) represents balanced CH₄ oxidation; (D)
 483 represents 2,4-D contamination inhibiting oxidation (i.e., removal) of CH₄ by methanotrophs causing
 484 higher CH₄ emissions.



485

486 **Figure 2.** Conceptual diagrams of a wetland with methanogens, methanotrophs, and algae. Panel (A)
 487 represents a balanced CH₄ cycle, where algae and methanogens produce CH₄, methanotrophs oxidize
 488 CH₄; panel (B) represents an unbalanced CH₄ cycle where glycosate and 2,4-D contamination

489 stimulate methanogens and algae and inhibit CH₄ oxidation, respectively, and thus result in higher
490 CH₄ emissions.