

# A Perspective on How Glyphosate and 2,4-D May Impact Climate Change

1 **Christine M Cornish<sup>1\*</sup> and Jon N Sweetman<sup>2</sup>**

2 <sup>1</sup>North Dakota State University, Biological Sciences Department, Environmental and Conservation  
3 Sciences Program, Fargo, ND, USA

4 <sup>2</sup>The Pennsylvania State University, Department of Ecosystem Science and Management, University  
5 Park, PA, USA

6 **\* Correspondence:**

7 Christine M Cornish

8 christine.cornish@ndsu.edu

9 **Keywords: glyphosate, 2,4-D, wetlands, methane, climate change**

## 10 **Abstract**

11 An increase in herbicide use is occurring due to a growing population and herbicide-resistant crops in  
12 agriculture, which has resulted in more herbicide tolerant target species. Glyphosate and 2,4-  
13 Dichlorophenoxyacetic acid (2,4-D) are two of the most commonly used herbicides worldwide and  
14 are more recently being used in combination in pre-mixed commercial formulas. Subsequently,  
15 herbicide contamination of wetlands will increase subjecting microorganisms to multiple chemical  
16 stressors. Methane is a potent greenhouse gas naturally emitted from wetlands, but herbicides may  
17 disrupt biogeochemical processes leading to an unbalanced methane cycle. This perspective  
18 examined the potential effects of herbicides on climate change using glyphosate and 2,4-D as a case  
19 study. We highlighted previous research on glyphosate-derived nutrient enrichment and 2,4-D  
20 inhibition of methane oxidation. We also explained how the concurrent effects of these herbicides  
21 could alter microbial communities leading to increased methane production in wetlands. Our  
22 perspective elucidates the potential ecosystem-level implications of herbicides in wetlands, in  
23 addition to stating the importance for research on the combined effects of herbicides.

## 24 **1 Introduction**

25 Climate change is an ongoing global concern as greenhouse gas (GHG) emissions continue to  
26 increase (IPCC, 2021). Methane (CH<sub>4</sub>) is the second most abundant GHG, after carbon dioxide  
27 (CO<sub>2</sub>), but is about 25 times more potent (Islam et al., 2018; EPA, 2023). Wetlands are a large  
28 natural source of CH<sub>4</sub> as they play a significant role in carbon (C) sequestration and cycling  
29 (Andresen et al., 2017). It has recently been suggested that GHG emissions from freshwater  
30 ecosystems may be impacted by agrochemical use (Stehle and Schulz, 2015), which has substantially  
31 increased over the past 30 years due to the introduction of herbicide-resistant crops. Herbicide-  
32 resistant crops now comprise more than 90% of crops in the United States (USDA, 2022) and are  
33 increasingly used worldwide (Peterson et al., 2018). This has unexpectedly led to an increase in  
34 herbicide use (Bai and Ogbourne, 2016; Coupe and Capel, 2016), and subsequently weeds are  
35 chronically exposed to herbicide residues resulting in the development of herbicide tolerance (Heap  
36 and Duke, 2018). For example, the introduction of Roundup Ready<sup>®</sup> (i.e., glyphosate-resistant) crops  
37 resulted in a 15-fold increase in glyphosate use (Coupe and Capel, 2016; Hébert et al., 2019), and

38 consequently, approximately 41 weed species worldwide have developed glyphosate tolerance  
39 (Jeschke and Teten, 2018). Ongoing climatic change is projected to result in intensified herbicide use  
40 (Delcour et al., 2015), creating a positive feedback loop if increasing herbicide use results in further  
41 CH<sub>4</sub> emissions. Temperature increases can also enhance toxicity and alter biodegradation processes  
42 of herbicides (Noyes et al., 2009; Koleva and Schneider, 2010; Matzrafi, 2019). Subsequently, as  
43 wetlands are subjected to a combination of more severe stressors, wetland biota, specifically  
44 microorganisms may be detrimentally affected. Pre-mixed herbicides containing multiple active  
45 ingredients (i.e., multiple modes of action), such as glyphosate and 2,4-Dichlorophenoxyacetic acid  
46 (2,4-D), are also now more commonly used to combat target species that have developed tolerance to  
47 single active ingredient herbicides (Freydier and Lundgren, 2016; Schütte et al., 2017). In this article  
48 we will present a case study on how glyphosate and 2,4-D accumulation in wetlands could impact  
49 climate change by focusing on their potential synergistic effects on microbial communities.  
50 Specifically, understanding their impacts on CH<sub>4</sub> dynamics is critical to understanding the role of  
51 wetlands in contributing to climate change.

## 52 **2 Case Study**

53 Glyphosate and 2,4-D are two of the most commonly used herbicides and are also used in pre-  
54 mixed formulas such as Enlist Duo<sup>®</sup> (1:0.95 glyphosate:2,4-D) and Landmaster<sup>™</sup> II (1:0.83  
55 glyphosate:2,4-D) (Benbrook, 2016; Zuanazzi et al., 2020; EPA, 2022). Extensive use of glyphosate  
56 and 2,4-D are cause for environmental concern within aquatic ecosystems where herbicides are  
57 substantial contributors to wetland pollution (Casado et al., 2019). As wetlands are often located in  
58 the lowest drainage points of agricultural fields, they can serve as critical sinks for herbicide residues  
59 transported through spray drift, runoff, groundwater leaching, and wind and sediment erosion (Annett  
60 et al., 2014; Bento et al., 2017). Subsequently, this can result in bioavailable residues, where  
61 microorganisms are subjected to a “pesticide cocktail” (Aparicio et al., 2013; Islam et al., 2018).  
62 Microorganisms, such as prokaryotes and algae, are important contributors to wetland ecosystem  
63 functioning, including mediating biogeochemical cycling (Sun et al., 2013; Baker et al., 2020). The C  
64 biogeochemical processes of methanogens (i.e., CH<sub>4</sub> producers) and methanotrophs (i.e., CH<sub>4</sub>  
65 consumers), in addition to plant and algal-mediated transport, play a critical role in the global CH<sub>4</sub>  
66 budget of wetlands. Agrochemical contributions to increased GHG emissions (Stehle and Schulz,  
67 2015) may be explained by an unbalanced CH<sub>4</sub> cycle via impacts to CH<sub>4</sub>-associated microorganisms.  
68 The range of effects of glyphosate and 2,4-D on wetland microorganisms can vary including direct  
69 toxicity, nutrient enrichment, and alterations to metabolic and catabolic processes.

### 70 **2.1 Glyphosate**

71 Glyphosate’s potential impacts on wetland microorganisms can be detrimental or advantageous  
72 to microbial species. While glyphosate’s mode of action was developed to target the shikimate  
73 pathway in higher plants (Hetrick and Blankinship, 2015), many archaea and bacteria also utilize this  
74 pathway resulting in non-target effects (Herrmann and Weaver, 1999). Despite the potential negative  
75 impacts on microorganisms, in many instances increased growth, respiration, and enhanced  
76 metabolism in wetland microbial communities have been observed as a result of glyphosate  
77 biodegradation (Vera et al., 2012; Lu et al., 2020) and linked with the use of glyphosate as a nutritive  
78 source (Saxton et al., 2011; Wang et al., 2016). Due to glyphosate’s chemical structure, its  
79 degradation often contributes substantial amounts of phosphorus (P), which has been found to be  
80 favored and more rapidly utilized by microorganisms compared to other sources of soil P (Hébert et  
81 al., 2019; Sun et al., 2019). Specifically, stimulated cyanobacterial growth and cyanotoxin production  
82 has been recorded from glyphosate-derived P enrichment (Vera et al., 2010; Qiu et al., 2013; Zhang

83 et al., 2016; Hernández-García and Martínez-Jerónimo, 2020; Wang et al., 2021; Lin et al., 2023).  
84 Glyphosate degradation was found to be positively correlated with total P concentrations in surface  
85 waters (Carles et al., 2019). Glyphosate additions to aquatic ecosystems can contribute to water  
86 quality issues, such as eutrophication, which has been demonstrated to be an important driver of CH<sub>4</sub>  
87 emissions (Sepulveda-Jauregui et al., 2018; Beaulieu et al., 2019; Yang et al., 2019; Bertolet et al.,  
88 2020). In addition, glyphosate-derived P loading into wetlands could stimulate algal biomass leading  
89 to eutrophic conditions, which could then increase CH<sub>4</sub> production (Carles et al., 2019). Ultimately,  
90 increased glyphosate use could shift microbial community dynamics towards copiotrophs and algae,  
91 altering important C biogeochemical processes and resulting in an indirect increase in CH<sub>4</sub>  
92 production in wetlands (Figure 1, panels A and B).

## 93 **2.2 2,4-D**

94 Despite 2,4-D being the first synthetic herbicide, compared to glyphosate, relatively little  
95 research has been conducted on its effects on aquatic microorganisms (Donald et al., 2018; Malaj et  
96 al., 2020). However, similar to glyphosate, 2,4-D can have a variety of impacts on wetland microbial  
97 communities. It targets broadleaf plants through mimicking the plant growth hormone, indol-3-yl-  
98 acetic acid (IAA or auxin), resulting in plant overgrowth (Cobb and Reade, 2010), but auxin  
99 synthesis and usage in microorganisms is also well known making them vulnerable non-target  
100 organisms (Spaepen and Vanderleyden, 2011). In addition, 2,4-D is a widespread environmental  
101 contaminant frequently detected in aquatic ecosystems (Malaj et al., 2020), and its use has increased  
102 in recent decades with the development of herbicide-resistant crops, where its use will likely continue  
103 to increase in the future (Freydier and Lundgren, 2016). Consequently, wetland microorganisms  
104 could be highly susceptible to its toxic effects with limited capacity to degrade it. Previous research  
105 has found some species use 2,4-D as a C source, whereas other species are toxicologically inhibited  
106 (Benndorf et al., 2004; Zabaloy et al., 2008; Sachu et al., 2022). Research in microcosms has also  
107 found that increased 2,4-D concentrations resulted in inhibition of CH<sub>4</sub> oxidation, decreases in CH<sub>4</sub>  
108 removal time, and increased CH<sub>4</sub> emissions (Syamsul Arif et al., 1996; Kumaraswamy et al., 1997;  
109 Top et al., 1999). Where studies from Top et al. (1999) and Seghers et al. (2003) suggested decreases  
110 in CH<sub>4</sub> removal could be due to 2,4-D inhibition of methanotroph-mediated oxidative metabolism.  
111 Research on the effects of 2,4-D on CH<sub>4</sub> oxidation is extremely limited, however these studies do  
112 indicate that 2,4-D loading into wetlands could potentially alter the CH<sub>4</sub> cycle by suppressing the  
113 removal of CH<sub>4</sub> via the food web, resulting in greater concentrations within the water column and  
114 higher emissions (Figure 1, panels C and D).

## 115 **2.3 Pesticide cocktails: Glyphosate plus 2,4-D**

116 The increased use of pre-mixed glyphosate and 2,4-D herbicides further exposes wetland  
117 microorganisms to combinations of chemical stressors, which could lead to unforeseen long-term  
118 effects. Research on the combined effects of pesticides has been conducted since the 1970's, but the  
119 majority of the focus has been on the direct toxicological impacts to aquatic flora and fauna  
120 (Lichtenstein et al., 1973; Faust et al., 1994; Gardner and Grue, 1996; Hayes et al., 2006; Relyea,  
121 2009; Moreira et al., 2020). These studies included compounds such as atrazine, chlorpyrifos,  
122 fipronil, etc., whereas research on the combined effects of glyphosate and 2,4-D is limited, especially  
123 at the aquatic microbial level. Additive and/or synergistic effects of glyphosate and 2,4-D have been  
124 found on fish and amphibian growth, fertilization, survival, and behavior (Carvalho et al., 2020;  
125 Pavan et al., 2021; Bernardi et al., 2022; Peluso et al., 2022), and zooplankton emergence (Portinho  
126 et al., 2018). Lozano et al. (Lozano et al., 2018) found additive impacts of glyphosate and 2,4-D on  
127 phytoplankton composition, abundance, and chlorophyll *a* after 7 days in microcosms, but also found

128 an antagonist effect on total and live abundance of *Staurastrum* spp. In outdoor mesocosms Lozano  
129 et al. (Lozano et al., 2018) found a decrease in phytoplankton respiration and gross primary  
130 production from a high glyphosate (applied as Roundup Max<sup>®</sup>), low 2,4-D (applied as AsiMax 50<sup>®</sup>)  
131 treatment after 4 hours. Additionally, after 7 days in mesocosms with high glyphosate, an increase in  
132 primary production, chlorophyll *a*, and micro- and nanophytoplankton was observed (Lozano et al.,  
133 2020). Sura et al. (Sura et al., 2015) researched the effects of a herbicide mixture including  
134 glyphosate, 2,4-D, and MCPA, clopyralid, dicamba, dichlorprop, mecoprop, and bromoxynil on  
135 pelagic and benthic communities in nutrient-sufficient and nutrient-deficient wetlands. They found  
136 pelagic bacterial productivity significantly increased after treatment in the nutrient-sufficient wetland,  
137 but benthic bacterial productivity did not change, which suggests the stimulatory effect of these  
138 herbicides may be related to nutrient bioavailability. These results demonstrate the complexity of the  
139 direct effects of herbicide mixtures on aquatic microorganisms, but the potential indirect effects are  
140 still poorly understood. As pre-mixed glyphosate and 2,4-D herbicides become more common it is  
141 important to consider the extent of their effects on aquatic ecosystems. Glyphosate can easily be used  
142 as a nutrient source stimulating microbial activity, specifically algal communities, whereas 2,4-D  
143 may inhibit methanotrophic communities from oxidizing CH<sub>4</sub>. As these compounds enter aquatic  
144 ecosystems their impacts on microorganisms may become synergistic and/or additive resulting in  
145 eutrophication and inhibition of methanotrophs from glyphosate and 2,4-D, respectively.  
146 Subsequently, eutrophic conditions and decreased CH<sub>4</sub> removal could cause increased CH<sub>4</sub>  
147 production via an unbalanced CH<sub>4</sub> cycle (Figure 2).

### 148 **3 Pollution-Induced Community Tolerance (PICT)**

149 Aquatic ecosystems are subjected to year-round herbicide contamination, where herbicide use  
150 differs across crop, season, habitat, and region. Microorganism structure and function can be  
151 impacted by herbicides, but toxicity is often dependent on the mode of action, concentration, and  
152 duration of exposure, as well as microbial species and environmental factors (DeLorenzo et al.,  
153 2001). For example, glyphosate stimulated *Chlorella vulgaris* growth 24-hours after exposure, but  
154 then inhibited growth after 48-hours at the same concentrations (i.e., hormesis) (Reno et al., 2014). In  
155 addition to the duration of exposure, the exposure to a different mode of action could also impact  
156 microorganisms by causing a community shift often appearing as changes in gene expression or  
157 diversity (Feld et al., 2015). Pollution-Induced Community Tolerance (PICT) refers to the response  
158 of a community to a pollutant, which results in an increased tolerance to that pollutant (Blanck,  
159 2002). The use of PICT analysis is extensive in the toxicology literature, especially on phototrophic  
160 microorganisms, which are often more susceptible to herbicidal effects due to their similarities with  
161 target species (DeLorenzo et al., 2001; Larras et al., 2016). Bérard and Benninghoff (Bérard and  
162 Benninghoff, 2001) found phytoplankton were significantly more sensitive to atrazine after one day,  
163 but then significantly less sensitive after at least 11 days. Phototrophic biofilms were found to be  
164 increasingly more sensitive to diuron as contamination levels decreased 1-3 years after its ban in the  
165 European Union (Pesce et al., 2016). It has also been shown that selection pressure from multiple  
166 stressors can lead to more opportunistic species and higher tolerances (Rotter et al., 2013).  
167 Ultimately, PICT results suggest that more sensitive species are being replaced by less sensitive  
168 species creating a more tolerant community (Blanck, 2002). This has also been seen with both  
169 glyphosate and 2,4-D. Microbial communities from sediments with high glyphosate exposure were  
170 able to degrade glyphosate faster and had higher diversity compared to sediments with low to no  
171 previous exposure (Tang et al., 2019). Zabaloy et al. (Zabaloy et al., 2008) saw an increase in a 2,4-D  
172 degrading population in soils for approximately one month after treatment and found that agricultural  
173 soils had higher 2,4-D tolerance compared to reference soils via PICT analysis. In a study by de  
174 Liphay et al. (de Liphay et al., 2002) 2,4-D treatment induced transcription of the gene responsible

175 for 2,4-D degradation (*tfdA*) which demonstrates a survival response from the microbial community.  
176 These studies demonstrate PICT can occur when communities are exposed to a herbicide, therefore  
177 contamination by an additional herbicide could further alter communities that have not been exposed  
178 before. It could be presumed that wetland microbial communities within a glyphosate-dominant  
179 region may substantially change when 2,4-D is introduced in combination with glyphosate and  
180 species are replaced. This potential shift would impact the biogeochemical functions of the  
181 community, subsequently altering herbicide degradation or metabolism.

## 182 **4 Conclusions**

183 Glyphosate and 2,4-D are frequently cited as having minimal to no environmental impacts  
184 (Peterson et al., 2016; Duke, 2020; Singh et al., 2020), however there is increasing evidence that their  
185 indirect effects may be of more substantial global concern. Wetlands naturally emit CH<sub>4</sub> via  
186 diffusion, ebullition (i.e., bubbles), and plant-mediated transport, and are the highest natural sources  
187 of CH<sub>4</sub> in the environment (Aben et al., 2017; Andresen et al., 2017), but emissions may be  
188 increasing due to agrochemical use adversely impacting CH<sub>4</sub> sink potential (Seghers et al., 2005).  
189 Glyphosate could stimulate microbial processes resulting in increased CH<sub>4</sub> production, in addition to  
190 2,4-D inhibiting CH<sub>4</sub> oxidation further resulting in increased CH<sub>4</sub> production. Ultimately, this would  
191 lead to higher CH<sub>4</sub> production versus removal from freshwater creating elevated CH<sub>4</sub> in the  
192 atmosphere. Due to the widespread and extensive use of glyphosate and 2,4-D, these contaminants  
193 are frequently found in wetlands (Islam et al., 2018; Malaj et al., 2020). To our knowledge there has  
194 been no research investigating the combined impacts of glyphosate and 2,4-D on wetland microbial  
195 communities. The potential bottom-up effects of glyphosate and 2,4-D could be detrimental to a  
196 changing climate, thus improving our understanding of how these herbicides can impact GHG  
197 emissions is crucial.

### 198 **4.1 Future Research**

199 To investigate the effects of glyphosate and 2,4-D on CH<sub>4</sub> emissions from freshwater  
200 ecosystems, micro- or mesocosm experiments could be conducted. Experiments under controlled  
201 conditions could help determine how wetland microbial communities are affected by glyphosate and  
202 2,4-D. Specifically, this research would give insight into the CH<sub>4</sub>-related mechanisms that may be  
203 enhanced or disrupted in microorganisms. In addition to in-lab research, pesticide loading data could  
204 be incorporated into GHG models. These data are currently not included in estimations of CH<sub>4</sub>  
205 emissions from wetlands, but could be an important source of variation, and could be useful for  
206 future climate modeling. These potential impacts are crucial to research as herbicide use is only  
207 expected to increase over time, where chemical selection pressure on microbial communities could  
208 contribute to climate change.

## 209 **5 References**

- 210 Aben, R. C. H., Barros, N., van Donk, E., Frenken, T., Hilt, S., Kazanjian, G., et al. (2017). Cross  
211 continental increase in methane ebullition under climate change. *Nature Communications* 8,  
212 1682. doi: 10.1038/s41467-017-01535-y.
- 213 Andresen, C. G., Lara, M. J., Tweedie, C. E., and Lougheed, V. L. (2017). Rising plant-mediated  
214 methane emissions from arctic wetlands. *Global Change Biology* 23, 1128–1139. doi:  
215 10.1111/gcb.13469.

- 216 Annett, R., Habibi, H. R., and Hontela, A. (2014). Impact of glyphosate and glyphosate-based  
217 herbicides on the freshwater environment: Impact of glyphosate-based herbicides. *J. Appl.*  
218 *Toxicol.* 34, 458–479. doi: 10.1002/jat.2997.
- 219 Aparicio, V. C., De Gerónimo, E., Marino, D., Primost, J., Carriquiriborde, P., and Costa, J. L.  
220 (2013). Environmental fate of glyphosate and aminomethylphosphonic acid in surface waters  
221 and soil of agricultural basins. *Chemosphere* 93, 1866–1873. doi:  
222 10.1016/j.chemosphere.2013.06.041.
- 223 Bai, S. H., and Ogbourne, S. M. (2016). Glyphosate: environmental contamination, toxicity and  
224 potential risks to human health via food contamination. *Environ Sci Pollut Res* 23, 18988–  
225 19001. doi: 10.1007/s11356-016-7425-3.
- 226 Baker, B. J., De Anda, V., Seitz, K. W., Dombrowski, N., Santoro, A. E., and Lloyd, K. G. (2020).  
227 Diversity, ecology and evolution of Archaea. *Nat Microbiol* 5, 887–900. doi:  
228 10.1038/s41564-020-0715-z.
- 229 Beaulieu, J. J., DelSontro, T., and Downing, J. A. (2019). Eutrophication will increase methane  
230 emissions from lakes and impoundments during the 21st century. *Nat Commun* 10, 1375. doi:  
231 10.1038/s41467-019-09100-5.
- 232 Benbrook, C. M. (2016). Trends in glyphosate herbicide use in the United States and globally.  
233 *Environ Sci Eur* 28, 3. doi: 10.1186/s12302-016-0070-0.
- 234 Benndorf, D., Davidson, I., and Babel, W. (2004). Regulation of catabolic enzymes during long-term  
235 exposure of *Delftia acidovorans* MC1 to chlorophenoxy herbicides. *Microbiology* 150, 1005–  
236 1014. doi: 10.1099/mic.0.26774-0.
- 237 Bento, C. P. M., Goossens, D., Rezaei, M., Riksen, M., Mol, H. G. J., Ritsema, C. J., et al. (2017).  
238 Glyphosate and AMPA distribution in wind-eroded sediment derived from loess soil.  
239 *Environmental Pollution* 220, 1079–1089. doi: 10.1016/j.envpol.2016.11.033.
- 240 Bérard, A., and Benninghoff, C. (2001). Pollution-induced community tolerance (PICT) and seasonal  
241 variations in the sensitivity of phytoplankton to atrazine in nanocosms. *Chemosphere* 45,  
242 427–437. doi: 10.1016/S0045-6535(01)00063-7.
- 243 Bernardi, F., Lirola, J. R., Cestari, M. M., and Bombardelli, R. A. (2022). Effects on reproductive,  
244 biochemical and genotoxic parameters of herbicides 2,4-D and glyphosate in silver catfish  
245 (*Rhamdia quelen*). *Environmental Toxicology and Pharmacology* 89, 103787. doi:  
246 10.1016/j.etap.2021.103787.
- 247 Bertolet, B. L., Olson, C. R., Szydlowski, D. K., Solomon, C. T., and Jones, S. E. (2020). Methane  
248 and Primary Productivity in Lakes: Divergence of Temporal and Spatial Relationships. *J.*  
249 *Geophys. Res. Biogeosci.* 125. doi: 10.1029/2020JG005864.
- 250 Blanck, H. (2002). A Critical Review of Procedures and Approaches Used for Assessing Pollution-  
251 Induced Community Tolerance (PICT) in Biotic Communities. *Human and Ecological Risk*  
252 *Assessment: An International Journal* 8, 1003–1034. doi: 10.1080/1080-700291905792.

- 253 Carles, L., Gardon, H., Joseph, L., Sanchís, J., Farré, M., and Artigas, J. (2019). Meta-analysis of  
254 glyphosate contamination in surface waters and dissipation by biofilms. *Environment*  
255 *International* 124, 284–293. doi: 10.1016/j.envint.2018.12.064.
- 256 Carvalho, W. F., Ruiz De Arcaute, C., Torres, L., De Melo E Silva, D., Soloneski, S., and  
257 Larramendy, M. L. (2020). Genotoxicity of mixtures of glyphosate with 2,4-  
258 dichlorophenoxyacetic acid chemical forms towards *Cnesterodon decemmaculatus* (Pisces,  
259 Poeciliidae). *Environ Sci Pollut Res* 27, 6515–6525. doi: 10.1007/s11356-019-07379-x.
- 260 Casado, J., Brigden, K., Santillo, D., and Johnston, P. (2019). Screening of pesticides and veterinary  
261 drugs in small streams in the European Union by liquid chromatography high resolution mass  
262 spectrometry. *Science of The Total Environment* 670, 1204–1225. doi:  
263 10.1016/j.scitotenv.2019.03.207.
- 264 Cobb, A. H., and Reade, J. P. H. (2010). *Herbicides and Plant Physiology: Cobb/Herbicides and*  
265 *Plant Physiology*. Oxford, UK: Wiley-Blackwell Available at:  
266 <http://doi.wiley.com/10.1002/9781444327793> [Accessed June 14, 2022].
- 267 Coupe, R. H., and Capel, P. D. (2016). Trends in pesticide use on soybean, corn and cotton since the  
268 introduction of major genetically modified crops in the United States: Pesticide use on US  
269 soybean, corn and cotton since the introduction of GM crops. *Pest. Manag. Sci.* 72, 1013–  
270 1022. doi: 10.1002/ps.4082.
- 271 de Liphthay, J. R., Aamand, J., and Barkay, T. (2002). Expression of *tfdA* genes in aquatic microbial  
272 communities during acclimation to 2,4-dichlorophenoxyacetic acid. *FEMS Microbiology*  
273 *Ecology* 40, 205–214. doi: 10.1111/j.1574-6941.2002.tb00953.x.
- 274 Delcour, I., Spanoghe, P., and Uyttendaele, M. (2015). Literature review: Impact of climate change  
275 on pesticide use. *Food Research International* 68, 7–15. doi: 10.1016/j.foodres.2014.09.030.
- 276 DeLorenzo, M. E., Scott, G. I., and Ross, P. E. (2001). Toxicity of pesticides to aquatic  
277 microorganisms: A review. *Environ Toxicol Chem* 20, 84–98. doi: 10.1002/etc.5620200108.
- 278 Donald, D. B., Cessna, A. J., and Farenhorst, A. (2018). Concentrations of Herbicides in Wetlands on  
279 Organic and Minimum-Tillage Farms. *J. Environ. Qual.* 47, 1554–1565. doi:  
280 10.2134/jeq2018.03.0100.
- 281 Duke, S. O. (2020). Glyphosate: environmental fate and impact. *Weed Sci* 68, 201–207. doi:  
282 10.1017/wsc.2019.28.
- 283 EPA (2022). Registration of Enlist One and Enlist Duo. Available at:  
284 [https://www.epa.gov/ingredients-used-pesticide-products/registration-enlist-one-and-enlist-](https://www.epa.gov/ingredients-used-pesticide-products/registration-enlist-one-and-enlist-duo)  
285 [duo](https://www.epa.gov/ingredients-used-pesticide-products/registration-enlist-one-and-enlist-duo).
- 286 EPA (2023). Overview of Greenhouse Gases. *Greenhouse Gas Emissions*. Available at:  
287 <https://www.epa.gov/ghgemissions/overview-greenhouse-gases>.
- 288 Faust, M., Altenburger, R., Boedeker, W., and Grimme, L. H. (1994). Algal toxicity of binary  
289 combinations of pesticides. *Bull. Environ. Contam. Toxicol.* 53. doi: 10.1007/BF00205150.

- 290 Feld, L., Hjelmsø, M. H., Nielsen, M. S., Jacobsen, A. D., Rønn, R., Ekelund, F., et al. (2015).  
291 Pesticide Side Effects in an Agricultural Soil Ecosystem as Measured by amoA Expression  
292 Quantification and Bacterial Diversity Changes. *PLoS ONE* 10, e0126080. doi:  
293 10.1371/journal.pone.0126080.
- 294 Freydier, L., and Lundgren, J. G. (2016). Unintended effects of the herbicides 2,4-D and dicamba on  
295 lady beetles. *Ecotoxicology* 25, 1270–1277. doi: 10.1007/s10646-016-1680-4.
- 296 Gardner, S. C., and Grue, C. E. (1996). Effects of rodeo® and garlon® 3A on nontarget wetland  
297 species in central Washington. *Environ Toxicol Chem* 15, 441–451. doi:  
298 10.1002/etc.5620150406.
- 299 Hayes, T. B., Case, P., Chui, S., Chung, D., Haeffele, C., Haston, K., et al. (2006). Pesticide  
300 Mixtures, Endocrine Disruption, and Amphibian Declines: Are We Underestimating the  
301 Impact? *Environ Health Perspect* 114, 40–50. doi: 10.1289/ehp.8051.
- 302 Heap, I., and Duke, S. O. (2018). Overview of glyphosate-resistant weeds worldwide: Overview of  
303 glyphosate-resistant weeds. *Pest. Manag. Sci* 74, 1040–1049. doi: 10.1002/ps.4760.
- 304 Hébert, M.-P., Fugère, V., and Gonzalez, A. (2019). The overlooked impact of rising glyphosate use  
305 on phosphorus loading in agricultural watersheds. *Front Ecol Environ* 17, 48–56. doi:  
306 10.1002/fee.1985.
- 307 Hernández-García, C. I., and Martínez-Jerónimo, F. (2020). Multistressor negative effects on an  
308 experimental phytoplankton community. The case of glyphosate and one toxigenic  
309 cyanobacterium on Chlorophycean microalgae. *Science of The Total Environment* 717,  
310 137186. doi: 10.1016/j.scitotenv.2020.137186.
- 311 Herrmann, K. M., and Weaver, L. M. (1999). THE SHIKIMATE PATHWAY. *Annu Rev Plant*  
312 *Physiol Plant Mol Biol* 50, 473–503. doi: 10.1146/annurev.arplant.50.1.473.
- 313 Hetrick, J., and Blankinship, A. (2015). Preliminary Ecological Risk Assessment in Support of the  
314 Registration Review of Glyphosate and Its Salts. U.S. Environmental Protection Agency:  
315 Office of Chemical Safety and Pollution Prevention.
- 316 IPCC (2021). IPCC, 2021: Climate Change 2021: The Physical Science Basis, the Working Group I  
317 contribution to the Sixth Assessment Report. Cambridge, United Kingdom and New York,  
318 NY, USA: Cambridge University Press.
- 319 Islam, F., Wang, J., Farooq, M. A., Khan, M. S. S., Xu, L., Zhu, J., et al. (2018). Potential impact of  
320 the herbicide 2,4-dichlorophenoxyacetic acid on human and ecosystems. *Environment*  
321 *International* 111, 332–351. doi: 10.1016/j.envint.2017.10.020.
- 322 Jeschke, M., and Teten, S. (2018). Glyphosate-Resistant Weeds in North America. 10.
- 323 Koleva, N. G., and Schneider, U. >A. (2010). The impact of climate change on aquatic risk from  
324 agricultural pesticides in the US. *International Journal of Environmental Studies* 67, 677–  
325 704. doi: 10.1080/00207233.2010.507477.



- 326 Kumaraswamy, S., Ramakrishnan, B., Satpathy, S. N., Rath, A. K., Misra, S., Rao, V. R., et al.  
327 (1997). Spatial distribution of methane-oxidizing activity in a flooded rice soil. *Plant and Soil*  
328 191, 241–248. doi: 10.1023/A:1004274302326.
- 329 Larras, F., Rimet, F., Gregorio, V., Bérard, A., Leboulanger, C., Montuelle, B., et al. (2016).  
330 Pollution-induced community tolerance (PICT) as a tool for monitoring Lake Geneva long-  
331 term in situ ecotoxic restoration from herbicide contamination. *Environ Sci Pollut Res* 23,  
332 4301–4311. doi: 10.1007/s11356-015-5302-0.
- 333 Lichtenstein, E. P., Liang, T. T., and Anderegg, B. N. (1973). Synergism of Insecticides by  
334 Herbicides. *Science* 181, 847–849. doi: 10.1126/science.181.4102.847.
- 335 Lin, W., Zhang, Z., Chen, Y., Zhang, Q., Ke, M., Lu, T., et al. (2023). The mechanism of different  
336 cyanobacterial responses to glyphosate. *Journal of Environmental Sciences* 125, 258–265.  
337 doi: 10.1016/j.jes.2021.11.039.
- 338 Lozano, V. L., Allen Dohle, S., Vera, M. S., Torremorell, A., and Pizarro, H. N. (2020). Primary  
339 production of freshwater microbial communities is affected by a cocktail of herbicides in an  
340 outdoor experiment. *Ecotoxicology and Environmental Safety* 201, 110821. doi:  
341 10.1016/j.ecoenv.2020.110821.
- 342 Lozano, V. L., Vinocur, A., Sabio y García, C. A., Allende, L., Cristos, D. S., Rojas, D., et al. (2018).  
343 Effects of glyphosate and 2,4-D mixture on freshwater phytoplankton and periphyton  
344 communities: a microcosms approach. *Ecotoxicology and Environmental Safety* 148, 1010–  
345 1019. doi: 10.1016/j.ecoenv.2017.12.006.
- 346 Lu, T., Xu, N., Zhang, Q., Zhang, Z., Debognies, A., Zhou, Z., et al. (2020). Understanding the  
347 influence of glyphosate on the structure and function of freshwater microbial community in a  
348 microcosm. *Environmental Pollution* 260, 114012. doi: 10.1016/j.envpol.2020.114012.
- 349 Malaj, E., Liber, K., and Morrissey, C. A. (2020). Spatial distribution of agricultural pesticide use  
350 and predicted wetland exposure in the Canadian Prairie Pothole Region. *Science of The Total*  
351 *Environment* 718, 134765. doi: 10.1016/j.scitotenv.2019.134765.
- 352 Matzrafi, M. (2019). Climate change exacerbates pest damage through reduced pesticide efficacy.  
353 *Pest Management Science* 75, 9–13. doi: 10.1002/ps.5121.
- 354 Moreira, R. A., Rocha, G. S., Da Silva, L. C. M., Goulart, B. V., Montagner, C. C., Melão, M. D. G.  
355 G., et al. (2020). Exposure to environmental concentrations of fipronil and 2,4-D mixtures  
356 causes physiological, morphological and biochemical changes in *Raphidocelis subcapitata*.  
357 *Ecotoxicology and Environmental Safety* 206, 111180. doi: 10.1016/j.ecoenv.2020.111180.
- 358 Noyes, P. D., McElwee, M. K., Miller, H. D., Clark, B. W., Van Tiem, L. A., Walcott, K. C., et al.  
359 (2009). The toxicology of climate change: Environmental contaminants in a warming world.  
360 *Environment International* 35, 971–986. doi: 10.1016/j.envint.2009.02.006.
- 361 Pavan, F. A., Samojeden, C. G., Rutkoski, C. F., Folador, A., Da Fré, S. P., Müller, C., et al. (2021).  
362 Morphological, behavioral and genotoxic effects of glyphosate and 2,4-D mixture in tadpoles  
363 of two native species of South American amphibians. *Environmental Toxicology and*  
364 *Pharmacology* 85, 103637. doi: 10.1016/j.etap.2021.103637.

- 365 Peluso, J., Furió Lanuza, A., Pérez Coll, C. S., and Aronzon, C. M. (2022). Synergistic effects of  
366 glyphosate- and 2,4-D-based pesticides mixtures on *Rhinella arenarum* larvae. *Environ Sci*  
367 *Pollut Res* 29, 14443–14452. doi: 10.1007/s11356-021-16784-0.
- 368 Pesce, S., Margoum, C., and Foulquier, A. (2016). Pollution-induced community tolerance for in situ  
369 assessment of recovery in river microbial communities following the ban of the herbicide  
370 diuron. *Agriculture, Ecosystems & Environment* 221, 79–86. doi: 10.1016/j.agee.2016.01.009.
- 371 Peterson, M. A., Collavo, A., Ovejero, R., Shivrain, V., and Walsh, M. J. (2018). The challenge of  
372 herbicide resistance around the world: a current summary. *Pest Management Science* 74,  
373 2246–2259. doi: 10.1002/ps.4821.
- 374 Peterson, M. A., McMaster, S. A., Riechers, D. E., Skelton, J., and Stahlman, P. W. (2016). 2,4-D  
375 Past, Present, and Future: A Review. *Weed technol.* 30, 303–345. doi: 10.1614/WT-D-15-  
376 00131.1.
- 377 Portinho, J. L., Nielsen, D. L., Daré, L., Henry, R., Oliveira, R. C., and Branco, C. C. Z. (2018).  
378 Mixture of commercial herbicides based on 2,4-D and glyphosate mixture can suppress the  
379 emergence of zooplankton from sediments. *Chemosphere* 203, 151–159. doi:  
380 10.1016/j.chemosphere.2018.03.156.
- 381 Qiu, H., Geng, J., Ren, H., Xia, X., Wang, X., and Yu, Y. (2013). Physiological and biochemical  
382 responses of *Microcystis aeruginosa* to glyphosate and its Roundup® formulation. *Journal of*  
383 *Hazardous Materials* 248–249, 172–176. doi: 10.1016/j.jhazmat.2012.12.033.
- 384 Relyea, R. A. (2009). A cocktail of contaminants: how mixtures of pesticides at low concentrations  
385 affect aquatic communities. *Oecologia* 159, 363–376. doi: 10.1007/s00442-008-1213-9.
- 386 Reno, U., Gutierrez, M. F., Regaldo, L., and Gagneten, A. M. (2014). The Impact of Eskoba®, a  
387 Glyphosate Formulation, on the Freshwater Plankton Community. *water environ res* 86,  
388 2294–2300. doi: 10.2175/106143014X13896437493580.
- 389 Rotter, S., Heilmeier, H., Altenburger, R., and Schmitt-Jansen, M. (2013). Multiple stressors in  
390 periphyton - comparison of observed and predicted tolerance responses to high ionic loads  
391 and herbicide exposure. *J Appl Ecol* 50, 1459–1468. doi: 10.1111/1365-2664.12146.
- 392 Sachu, M., Kynshi, B. L., and Syiem, M. B. (2022). A biochemical, physiological and molecular  
393 evaluation of how the herbicide 2, 4-dichlorophenoxyacetic acid intercedes photosynthesis  
394 and diazotrophy in the cyanobacterium *Nostoc muscorum* Meg 1. *Environ Sci Pollut Res* 29,  
395 36684–36698. doi: 10.1007/s11356-021-18000-5.
- 396 Saxton, M. A., Morrow, E. A., Bourbonniere, R. A., and Wilhelm, S. W. (2011). Glyphosate  
397 influence on phytoplankton community structure in Lake Erie. *Journal of Great Lakes*  
398 *Research* 37, 683–690. doi: 10.1016/j.jglr.2011.07.004.
- 399 Schütte, G., Eckerstorfer, M., Rastelli, V., Reichenbecher, W., Restrepo-Vassalli, S., Ruohonen-  
400 Lehto, M., et al. (2017). Herbicide resistance and biodiversity: agronomic and environmental  
401 aspects of genetically modified herbicide-resistant plants. *Environ Sci Eur* 29, 5. doi:  
402 10.1186/s12302-016-0100-y.

- 403 Seghers, D., Siciliano, S. D., Top, E. M., and Verstraete, W. (2005). Combined effect of fertilizer and  
404 herbicide applications on the abundance, community structure and performance of the soil  
405 methanotrophic community. *Soil Biology and Biochemistry* 37, 187–193. doi:  
406 10.1016/j.soilbio.2004.05.025.
- 407 Sepulveda-Jauregui, A., Hoyos-Santillan, J., Martinez-Cruz, K., Walter Anthony, K. M., Casper, P.,  
408 Belmonte-Izquierdo, Y., et al. (2018). Eutrophication exacerbates the impact of climate  
409 warming on lake methane emission. *Science of The Total Environment* 636, 411–419. doi:  
410 10.1016/j.scitotenv.2018.04.283.
- 411 Singh, S., Kumar, V., Gill, J. P. K., Datta, S., Singh, S., Dhaka, V., et al. (2020). Herbicide  
412 Glyphosate: Toxicity and Microbial Degradation. *IJERPH* 17, 7519. doi:  
413 10.3390/ijerph17207519.
- 414 Spaepen, S., and Vanderleyden, J. (2011). Auxin and Plant-Microbe Interactions. *Cold Spring  
415 Harbor Perspectives in Biology* 3, a001438–a001438. doi: 10.1101/cshperspect.a001438.
- 416 Stehle, S., and Schulz, R. (2015). Agricultural insecticides threaten surface waters at the global scale.  
417 *Proc Natl Acad Sci USA* 112, 5750–5755. doi: 10.1073/pnas.1500232112.
- 418 Sun, K., Liu, W., Liu, L., Wang, N., and Duan, S. (2013). Ecological risks assessment of  
419 organophosphorus pesticides on bloom of *Microcystis wesenbergii*. *International  
420 Biodeterioration & Biodegradation* 77, 98–105. doi: 10.1016/j.ibiod.2012.11.010.
- 421 Sun, M., Li, H., and Jaisi, D. P. (2019). Degradation of glyphosate and bioavailability of phosphorus  
422 derived from glyphosate in a soil-water system. *Water Research* 163, 114840. doi:  
423 10.1016/j.watres.2019.07.007.
- 424 Sura, S., Waiser, M. J., Tumber, V., Raina-Fulton, R., and Cessna, A. J. (2015). Effects of a herbicide  
425 mixture on primary and bacterial productivity in four prairie wetlands with varying salinities:  
426 An enclosure approach. *Science of The Total Environment* 512–513, 526–539. doi:  
427 10.1016/j.scitotenv.2015.01.064.
- 428 Syamsul Arif, M. A., Houwen, F., and Verstraete, W. (1996). Agricultural factors affecting methane  
429 oxidation in arable soil. *Biol Fertil Soils* 21, 95–102. doi: 10.1007/BF00335999.
- 430 Tang, F. H. M., Jeffries, T. C., Vervoort, R. W., Conoley, C., Coleman, N. V., and Maggi, F. (2019).  
431 Microcosm experiments and kinetic modeling of glyphosate biodegradation in soils and  
432 sediments. *Science of The Total Environment* 658, 105–115. doi:  
433 10.1016/j.scitotenv.2018.12.179.
- 434 Top, E. M., Maila, M. P., Clerinx, M., Goris, J., Vos, P., and Verstraete, W. (1999). Methane  
435 oxidation as a method to evaluate the removal of 2,4-dichlorophenoxyacetic acid (2,4-D) from  
436 soil by plasmid-mediated bioaugmentation. *FEMS Microbiology Ecology* 28, 203–213. doi:  
437 10.1111/j.1574-6941.1999.tb00576.x.
- 438 USDA (2022). Recent Trends in GE Adoption. *Adoption of Genetically Engineered crops in the U.S.*  
439 Available at: [https://www.ers.usda.gov/data-products/adoption-of-genetically-engineered-  
440 crops-in-the-u-s/recent-trends-in-ge-adoption/](https://www.ers.usda.gov/data-products/adoption-of-genetically-engineered-crops-in-the-u-s/recent-trends-in-ge-adoption/).

- 441 Vera, M. S., Di Fiori, E., Lagomarsino, L., Sinistro, R., Escaray, R., Iummato, M. M., et al. (2012).  
442 Direct and indirect effects of the glyphosate formulation Glifosato Atanor® on freshwater  
443 microbial communities. *Ecotoxicology* 21, 1805–1816. doi: 10.1007/s10646-012-0915-2.
- 444 Vera, M. S., Lagomarsino, L., Sylvester, M., Pérez, G. L., Rodríguez, P., Mugni, H., et al. (2010).  
445 New evidences of Roundup® (glyphosate formulation) impact on the periphyton community  
446 and the water quality of freshwater ecosystems. *Ecotoxicology* 19, 710–721. doi:  
447 10.1007/s10646-009-0446-7.
- 448 Wang, S., Seiwert, B., Kästner, M., Miltner, A., Schäffer, A., Reemtsma, T., et al. (2016).  
449 (Bio)degradation of glyphosate in water-sediment microcosms – A stable isotope co-labeling  
450 approach. *Water Research* 99, 91–100. doi: 10.1016/j.watres.2016.04.041.
- 451 Wang, W., Jiang, M., and Sheng, Y. (2021). Glyphosate Accelerates the Proliferation of *Microcystis*  
452 *aeruginosa*, a Dominant Species in Cyanobacterial Blooms. *Environ Toxicol Chem* 40, 342–  
453 351. doi: 10.1002/etc.4942.
- 454 Yang, Y., Chen, J., Tong, T., Li, B., He, T., Liu, Y., et al. (2019). Eutrophication influences  
455 methanotrophic activity, abundance and community structure in freshwater lakes. *Science of*  
456 *The Total Environment* 662, 863–872. doi: 10.1016/j.scitotenv.2019.01.307.
- 457 Zabaloy, M. C., Garland, J. L., and Gómez, M. A. (2008). An integrated approach to evaluate the  
458 impacts of the herbicides glyphosate, 2,4-D and metsulfuron-methyl on soil microbial  
459 communities in the Pampas region, Argentina. *Applied Soil Ecology* 40, 1–12. doi:  
460 10.1016/j.apsoil.2008.02.004.
- 461 Zhang, Q., Zhou, H., Li, Z., Zhu, J., Zhou, C., and Zhao, M. (2016). Effects of glyphosate at  
462 environmentally relevant concentrations on the growth of and microcystin production by  
463 *Microcystis aeruginosa*. *Environ Monit Assess* 188, 632. doi: 10.1007/s10661-016-5627-2.
- 464 Zuanazzi, N. R., Ghisi, N. de C., and Oliveira, E. C. (2020). Analysis of global trends and gaps for  
465 studies about 2,4-D herbicide toxicity: A scientometric review. *Chemosphere* 241, 125016.  
466 doi: 10.1016/j.chemosphere.2019.125016.

## 467 **6 Conflict of Interest**

468 This manuscript was written in the absence of commercial or financial conflicts of interest.

## 469 **7 Author Contributions**

470 Christine Cornish generated the first draft of the manuscript with revisions and comments from Jon  
471 Sweetman. Both authors have read and approved the submitted version.

## 472 **8 Funding**

473 No funding was received for this manuscript.

## 474 **9 Acknowledgments**

475 Thank you to Dr. Ted Harris for reviewing this manuscript, and for his ongoing support as a  
476 committee member to CM Cornish.

477 **10 Data Availability Statement**

478 Not applicable.

479 **Figure 1.** Conceptual diagrams of a wetland with methanogens, methanotrophs, and algae. **(A)**  
480 represents balanced CH<sub>4</sub> production; **(B)** represents glyphosate contamination stimulating  
481 methanogens and algae causing higher CH<sub>4</sub> emissions; **(C)** represents balanced CH<sub>4</sub> oxidation; **(D)**  
482 represents 2,4-D contamination inhibiting oxidation (i.e., removal) of CH<sub>4</sub> by methanotrophs causing  
483 higher CH<sub>4</sub> emissions.

484 **Figure 2.** Conceptual diagrams of a wetland with methanogens, methanotrophs, and algae. Panel **(A)**  
485 represents a balanced CH<sub>4</sub> cycle, where algae and methanogens produce CH<sub>4</sub>, methanotrophs oxidize  
486 CH<sub>4</sub>; panel **(B)** represents an unbalanced CH<sub>4</sub> cycle where glyphosate and 2,4-D contamination  
487 stimulate methanogens and algae and inhibit CH<sub>4</sub> oxidation, respectively, and thus result in higher  
488 CH<sub>4</sub> emissions.