

1 **Assessing the multidimensional complexity of biodiversity using a globally** 2 **standardized approach**

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21 **Highlights**

- 22 1. The value of nature lies in its complexity.
- 23 2. We define a global framework for a holistic measure of the state of nature.
- 24 3. Inclusion of microbes and invertebrates is crucial to evaluate biodiversity.
- 25 4. Human activity consistently reduces and homogenizes biodiversity worldwide.

26

27 **Summary**

28 Quantifying biodiversity across the globe is critical for transparent reporting and assessment under
 29 the Kunming-Montreal Global Biodiversity Framework. Understanding the complexity of
 30 biodiversity requires consideration of the variation of life across genes, species, and ecosystems.
 31 Achieving this in a standardized way remains a key challenge for creating an equitable nature
 32 positive future. Here, we present the Sustainable Ecology and Economic Development (SEED)
 33 framework, which assesses the dimensions that structure biodiversity patterns worldwide (genetics,
 34 species, and ecosystems) across plants, animals, and microbial taxa, and consolidates this into a
 35 single measure of biocomplexity at every location, relative to a ‘reference’ ecosystem with minimal
 36 human disturbance. We describe the SEED methodology and highlight its features, which include
 37 seven novel measures of biodiversity intactness that are integrated into the SEED biocomplexity
 38 index. SEED will continuously integrate new datasets and maps to provide up-to-date estimates of
 39 local biocomplexity across the planet, providing a tool for decision makers to assess and improve
 40 the global state of nature.

41 **Keywords**

42 Biocomplexity; biodiversity monitoring; biodiversity intactness; genetic diversity; species
 43 diversity; ecosystem diversity

44 **Abbreviations**

SEED	Sustainable Ecology and Economic Development
GBF	Kunming-Montreal Global Biodiversity Framework
GEO BON	Group on Earth Observations Biodiversity Observation Network
EBV	Essential Biodiversity Variable
IUCN	International Union for Conservation of Nature
EII	Ecosystem Integrity Index
BII	Biodiversity Intactness Index
MSA	Mean Species Abundance
HMI	Human Modification Index
PNV	Potential Natural Vegetation

45 **1. Introduction**

46 Every species depends on other species to survive. This vast interdependence means biological
47 diversity is critical for the maintenance of life as we know it. However, humans have historically
48 valued the components of nature we can use for food, timber, and medicine over others^{1,2}. The
49 development of markets for these products has incentivized their mass propagation at the expense
50 of other components of biodiversity, driving the oversimplification of biological systems and the
51 loss of the ecosystem services on which we depend³.

52 As nature positive policy frameworks and nature markets emerge, it is critical that we learn from
53 past challenges. Mechanisms that value a single aspect of nature – such as carbon sequestration –
54 risk driving the oversimplification of the system⁴⁻⁷. This can lead to potentially counterproductive
55 outcomes, like the creation of monocultures of exotic tree species at the expense of local
56 biodiversity⁷ and human wellbeing⁸. If political and financial structures are to promote the
57 conservation of natural biodiversity, they must be underpinned by robust scientific monitoring that
58 reflects the full dimensionality of life, across genetic, species and ecosystem levels.

59 The most prominent guidelines for biodiversity monitoring are described within the Kunming-
60 Montreal Global Biodiversity Framework (GBF)⁹, which sets a series of global targets to halt and
61 reverse nature loss by 2030. Key targets in the GBF include protecting areas of high biodiversity
62 importance (Target 1), restoring 30% of degraded areas by 2030 (Target 2), and the headline
63 ‘30x30’ target that aims to conserve 30% of the Earth’s surface by 2030 (Target 3). In the financial
64 sector, the GBF requires businesses to disclose their impacts on biodiversity (Target 15), and the
65 Taskforce on Nature-related Financial Disclosures was assembled to provide guidance and outline
66 best practices for nature-related disclosures. At the heart of nature-related disclosures is a measure
67 of the “state of nature” against which impacts, dependencies, risks, and opportunities may be
68 measured¹⁰. However, it is not well defined what constitutes a suitable measure of the state of
69 nature.

70 A growing number of metrics have been developed to measure and monitor biodiversity.
71 Biodiversity metrics generally fall into two categories based on their dataset and scope: local
72 ground-sourced data (e.g. plot surveys, eDNA, bioacoustics monitors and camera traps), and global

73 remotely-sensed data (e.g. radar and multispectral imagery from satellites)¹¹. While ground-sourced
74 data are critical for providing detailed insights into the diversity of organisms in a specific study
75 area, their insights may be limited outside the local study sites^{12,13}. By contrast, remotely-sensed
76 data can provide globally-standardized assessments of ecosystem-scale characteristics like forest
77 structure¹⁴ and connectivity¹⁵, but not all aspects of nature can be measured directly from satellites.
78 Many global biodiversity metrics incorporate both ground-sourced and remote-sensed data to offer
79 a balance between local relevance and global scope. To align monitoring efforts, the Group on Earth
80 Observations Biodiversity Observation Network (GEO BON) recently proposed a worldwide
81 system of observation networks¹⁶ and established guidelines for 16 Essential Biodiversity Variables
82 (EBVs)^{17,18}. However, we lack a coherent method to consolidate these EBVs and all other
83 biodiversity metrics for a unified evaluation of nature.

84 To address the need for a unified, globally standardized measure of biodiversity across all taxa and
85 scales of variation, we first review existing global biodiversity metrics and highlight their strengths
86 and weaknesses in measuring the current state of nature (Section 2). We then present a new
87 integrative framework, the Sustainable Ecology and Economic Development (SEED), which is
88 designed to consolidate three primary hierarchical levels that underpin biodiversity – genetic,
89 species, and ecosystem – into a global measure of biocomplexity (Section 3). Nature is inherently
90 complex, and biocomplexity is defined as the emergent properties from multiple, often hierarchical
91 levels of interacting factors that “affect, sustain, or are modified by living organisms, including
92 humans”¹⁹. Recognizing this complexity, we designed SEED so that it can integrate numerous
93 global biodiversity maps and include new maps as they become available. This ensures that policy
94 and market tools will have the most up-to-date information on the state of nature.

95 **2. Current state of global biodiversity measurement**

96 Spatially-explicit biodiversity data products consist of a heterogeneous mix of calculations, model
97 predictions, metrics (standard of measure), and indices (aggregation of multiple indicators). Given
98 our exclusive focus on data products mapped over the globe, we hereafter refer to these products in
99 general as ‘maps’. We grouped existing maps into three main organizational levels of ecological
100 systems: within-species genetic diversity, among-species diversity, and ecosystem diversity. Global
101 maps at each level reflect different dimensions of diversity that are essential for a holistic

102 understanding of biocomplexity across the globe. Although some global maps correspond to a given
103 EBV²⁰, the EBVs do not consist of a set of spatially-explicit global maps²¹. Their main purpose is
104 to identify the key variables and provide standards on how to measure them¹⁷. Rather than discuss
105 this detail in depth, we identify the strengths and weaknesses of existing maps and the
106 considerations for including them in a unified measure of biodiversity. We conclude this section
107 with a brief description of a fourth group of maps that represent standardized measures of
108 biodiversity.

109 2.1 Genetic (within-species) diversity

110 Genetic diversity (heritable variation) represents variation in the genetic composition of individuals
111 within a species and among populations and is the source for adaptive responses to environmental
112 change^{22,23}. There are few global measures of genetic diversity and these are estimated from
113 mitochondrial DNA sequences stored in the National Centre for Biotechnology Information
114 GenBank and the Barcode of Life Database^{24–26}. While there are good insights about genetic
115 variation for certain plant²⁷ and animal species^{24–26}, we currently lack global predictions of genetic
116 variation for most species on the planet. Among the few taxonomic groups that have been studied,
117 loss of genetic diversity is often correlated with the loss of suitable habitat²⁸ and reduced
118 abundance²⁹. This may suggest that we could use measures of population declines to generate proxy
119 measures of decline in genetic diversity, but this population timeseries data is lacking for the vast
120 majority of organismal groups, except for the 5,200+ species in the Living Planet Database³⁰.
121 However, with modern advancements in genetic sequencing technology and widespread usage of
122 barcoding and metabarcoding³¹, new data are continually becoming available that will enable the
123 global measurement of genetic diversity across taxonomic groups²³.

124 2.2 Species, phylogenetic, and functional diversity

125 Traditionally, global biodiversity research has primarily focused on species richness (the number
126 of unique taxa). However, species diversity also includes phylogenetic (i.e., a measure of
127 evolutionary history³²) and functional diversity, and adding these dimensions adds depth to our
128 description of natural communities. Species richness maps are generally created by overlaying
129 several species range maps then summing the number of species per pixel^{33–35}, which in some

130 models is then modeled against a set of predictor variables before predicting richness^{36–38}.
131 Phylogenetic and functional diversity are calculated using a phylogeny³⁹ or a functional-trait
132 matrix⁴⁰ to quantify the unique contributions from each of the species present at each locality and
133 thus provide information about phylogenetic and functional components of diversity. Whereas a
134 species-rich community may be composed of evolutionarily and functionally redundant species,
135 other less species-rich communities may (or may not) have more evolutionarily and functionally
136 divergent species that may contribute unique functions to the community. Therefore, highlighting
137 those communities with particularly rich evolutionary roots and functional traits is key for a holistic
138 measure of biodiversity.

139 Species diversity maps inherit biases from their underlying data sources that may affect their
140 applicability. All global biodiversity databases over-represent some regions and under-represent
141 other regions, typically with a bias toward developed nations⁴¹, and there are considerable
142 disparities in taxonomic coverage, with a bias toward larger and more charismatic organisms. The
143 IUCN database – the source of many biodiversity maps^{24,25,42–45} – contains distribution data for over
144 80% of described vertebrate species and 14% of vascular plants, but only 2% of invertebrate
145 species, which represent the vast majority of animal species diversity. In addition, microbes
146 represent 88% to 99% of all species on Earth^{46,47} but are vastly underrepresented, with only 0.4%
147 of known fungi and protists included in the IUCN database. This pattern is now changing due to
148 recent advances in high-throughput sequencing technologies that enable us to observe and quantify
149 microscopic and otherwise cryptic species⁴⁸, and the availability of microbial biodiversity maps is
150 expanding exponentially^{49–52}.

151 Species diversity maps are also limited by their original scope and the underlying models. Until
152 now, most global maps of species diversity were not designed to capture fine-scale patterns in
153 species composition, or the effects of local human disturbance. With a focus on broad-scale
154 biogeographic trends, species diversity models focus climate, edaphic and topographic predictive
155 variables to predict diversity patterns across environmental gradients. The general lack of human
156 influences in these models leads to low predictive accuracy for quantifying the impacts of human
157 disturbance (Figure 1). This may be due in part to the recent emergence of global human
158 modification maps^{53–56} and also due to a paucity of biodiversity data in both heavily degraded and

159 intact landscapes. Furthermore, species diversity maps tend to be temporally static, calculated as an
160 average of observations that can span decades. Additionally, sampling methodologies are taxon
161 and/or habitat specific and may also differ by region and discipline or agency. These complexities
162 make it difficult to harmonize available data and generate well-rounded estimates that scale in space
163 and change over time in response to shifting conditions on the ground.

164 2.3 Ecosystem diversity

165 In contrast to genetic and species diversity, which rely heavily on ground-sourced data, ecosystem-
166 level maps provide a more up-to-date view of current conditions because they can be measured
167 directly from satellite imagery, or modeled based on spectral imagery (e.g., radar and multispectral
168 imaging). A wealth of global-scale remote sensing products have been designed to capture
169 information about human modifications⁵⁵, land use change⁵⁷, canopy cover⁵⁸, canopy height⁵⁹,
170 above and belowground biomass^{60,61}, soil respiration⁶², habitat heterogeneity⁶³, leaf area index⁶⁴,
171 ecosystem connectivity^{65,66}, net primary productivity⁶⁷ and ecological resilience^{68,69}. We group all
172 ecosystem-level maps that measure some aspect of biodiversity into one of three broad categories:
173 ecosystem structure, function¹⁷, or connectivity.

174 Ecosystem structure, function, and connectivity are emergent properties that arise from the
175 combination of species, landscape physiognomy, climate, and human modifications. For example,
176 the occurrence and extent of mangroves⁷⁰ and peatlands^{71,72} are due to relatively specific sets of
177 conditions, and their structural features are critical to their functional roles. The interplay between
178 living organisms and their environment are also key ecosystem properties, perhaps best exemplified
179 by plant-disperser and plant-fungal associations. Global maps of plant-disperser associations or
180 species interaction networks are not currently available, but progress in this field indicates potential
181 for such maps in the future⁷³. The field of plant-fungal associations has produced numerous global
182 maps, which predict such functional features as the relative proportions of nitrogen fixing and
183 arbuscular mycorrhizal or ectomycorrhizal associated plants⁷⁴ and the densities of their fine roots⁷⁵
184 over the landscape, which may affect the intensity of mycorrhizal colonization⁷⁵, soil moisture⁷⁶
185 and decomposition rates⁷⁷. These ecosystem properties provide a direct link to measures of
186 ecosystem services^{78,79}. Just as the three-dimensional structure of an ecosystem characterizes the
187 environmental context in which species coexist, the spatial arrangement of habitat fragments and

188 species-specific dispersal abilities and limitations determine ecosystem connectivity. Ecosystem
189 connectivity tends to decrease with habitat loss and fragmentation, and both connectivity⁸⁰ and
190 fragmentation⁸¹ indices can provide unique insights into the ecological functionality at the
191 landscape-scale⁸².

192 2.4 Standardized measures of biodiversity

193 Given the challenges of capturing dynamic changes in ecological diversity at a global scale, a few
194 analyses estimate how far the ecological community has diverged from its natural state. Using a
195 database from targeted experiments and local studies spanning disturbance gradients, it is possible
196 to identify how land use change affects ecological diversity and use these relationships to predict
197 ecological intactness across the globe. Three commonly used global indices are the Ecosystem
198 Integrity Index (EII)⁸³ by the United Nations Environment Programme World Conservation
199 Monitoring Centre, the Biodiversity Intactness Index (BII)^{84,85} by the Natural History Museum in
200 London, and the Mean Species Abundance (MSA)⁸⁶ index by the Netherlands Environmental
201 Assessment Agency.

202 The EII includes three components: structure, function, and composition. Ecosystem structure is
203 based on the human modification index (HMI)⁵⁵; ecosystem function is measured by the ratio of
204 actual to potential net primary productivity⁶⁷; and ecosystem composition is measured by the
205 BII^{84,87}. For a given location, the EII uses the lowest score of the three components to predict the
206 extent to which any ecosystem has been impacted or altered from its original state. The BII uses a
207 linear model of the impacts of land use and related pressures on two aspects of biodiversity, species
208 abundances and compositional similarity, to estimate the intactness of a community of plants,
209 vertebrates, and invertebrates. The MSA index is conceptually similar to the BII but focuses mainly
210 on species abundance, includes additional human-related pressures, and estimates an average
211 intactness value that is weighted by the land use type and its associated human pressures.

212 The MSA, BII, and EII represent the current state-of-the-art in global biodiversity modeling. Yet,
213 there are two key aspects of diversity that these indices do not capture. First, these indices are fairly
214 limited in taxonomic scope. The BII and EII (and to some degree MSA) are primarily based on the
215 PREDICTS database (Projecting Responses of Ecological Diversity In Changing Terrestrial

216 Systems)⁸⁸, which is valuable for quantifying the impact of land use on biodiversity in different
217 regions of the globe. However, at present, the data coverage of PREDICTS represents a relatively
218 limited taxonomic range, including less than 10% of the described species for most large taxonomic
219 groups like vascular plants and invertebrate animals, and a far smaller proportion for microbes^{88,89}.
220 Given that these taxa respond differently to human disturbance and show unique global distribution
221 patterns^{90,91}, the underrepresentation of important taxa may therefore bias global biodiversity
222 assessments. Second, these indices focus exclusively on species level diversity and do not include
223 genetic diversity or, except for the EII, ecosystem level properties that emerge from the web of
224 interactions among species and their environment.

225 **3. A framework for observing biological complexity on Earth**

226 With the increasing combination of ground-sourced and remotely-sensed data, we are at the
227 beginning of a data revolution in global ecology^{92,93}. The exponential growth of global ecological
228 datasets and maps across genetic, species, and ecosystem levels represents exciting new
229 opportunities for our understanding of biodiversity across the planet. Conceptual frameworks that
230 integrate and interpret this growing body of information are essential to generate a holistic
231 understanding of global biocomplexity. Our global understanding of biocomplexity will never be
232 fully complete, as emerging scientific assessments continue to capture novel information.
233 Therefore, it is important to establish flexible and dynamic frameworks that can incorporate new
234 and emerging information as it becomes available.

235 A key element in new nature-related disclosure frameworks is a vaguely defined measure of the
236 state of nature¹⁰, which would presumably represent the full multidimensional complexity of nature,
237 but for which there is currently no agreed upon metric. To address the need for a unified state of
238 nature metric, we present a holistic ecological framework (hereafter referred to as SEED) that is
239 designed to represent the multidimensionality of nature by defining nine axes of variation, nested
240 within the three hierarchical levels of diversity: genetics, species (including phylogenetic and
241 functional diversity), and ecosystems (Figure 2). Within the genetic and species levels of variation,
242 we include plants, animals, and microbes. The grouping of microbes to include archaea, bacteria,
243 protists, and fungi could be disaggregated in the future as more information becomes available.
244 Within ecosystems, we distinguish three axes: structure, function, and connectivity. The SEED

245 framework incorporates all existing biodiversity maps and will update frequently to include the best
246 available information. The SEED framework incorporates components already identified as EBVs
247 by GEO BON¹⁷ (Figure 2) and generates summary indices for the nine axes of biodiversity
248 alongside a unified biocomplexity index. We apply the term biocomplexity¹⁹ to set it apart from
249 measures of single biodiversity components, and to highlight the inclusion of multiple hierarchical
250 levels of diversity, which emerge not just from the complex interplay between biological life and
251 the environment, but also from the billions of years of physical and biological evolution on Earth.

252 To globally-standardize the biocomplexity index, we estimate the similarity (ranging from zero and
253 one) between the current state of an ecosystem and its natural state, which we measure over a suite
254 of biodiversity maps in reference areas that we identify using the Human Modification Index to
255 have only experienced minimal human disturbance (see Section 3.2). The SEED framework first
256 summarizes the intactness (i.e., similarity to natural state) of multiple underlying biodiversity
257 features in each axis, which allows each axis to be viewed separately to aid comprehension and
258 facilitate action-impact assessments. We summarized this information a radar plot (Figure 2),
259 whereby the intactness values for each axis at a site are plotted on scale from zero to one. The SEED
260 framework also calculates the mean intactness for all nine axes combined to create a unified
261 score: the SEED biocomplexity index, which also ranges between zero and one, where values near
262 zero represents the near absence of biocomplexity (e.g., an open pit mine or paved area), and one
263 represents an area that is equal to its potential natural state (i.e., a minimally-disturbed ecosystem).
264 The SEED framework therefore offers both a single standardized biocomplexity value for any area
265 of interest and nine intactness indices for each axis of biodiversity, thus allowing the user to unpack
266 this information.

267 3.1 Integrating dimensions of biodiversity

268 Consolidating the multiple hierarchical levels of biodiversity into a single value is a critical feature
269 that makes our biocomplexity index generalizable and comprehensible. All available global maps
270 are combined within each of the relevant biodiversity axes (Figure 2), using a multivariate kernel
271 estimator⁹⁴ (Eq. 1), and then the mean of these nine axes consolidates this information into a single
272 biocomplexity index.

273
$$K(\mathbf{z}, \mathbf{z}_r) = \exp[-\delta \|\mathbf{w} \circ (\mathbf{z} - \mathbf{z}_r)\|_1] \quad (\text{Eq. 1})$$

274 Here, \mathbf{z} is a n -dimensional data vector for a given location, where n represents the number of input
 275 maps involved in the calculation. The term $\|\mathbf{w}(\mathbf{z} - \mathbf{z}_r)\|_1$, represents the Manhattan distance, (or
 276 ℓ^1 distance), between the data vector \mathbf{z} and the mean values for the corresponding reference areas
 277 \mathbf{z}_r , after elementwise multiplication (\circ) by the n -dimensional vector, \mathbf{w} , which contains the
 278 normalized weight for each input map. The resulting distance value is then converted into a
 279 similarity value, which is bounded by zero and one, by applying the kernel function, K , where δ
 280 represents a scaling parameter.

281 Sensitivity of similarity is set by the scaling parameter, δ , which we set for each biodiversity axis
 282 according to two criteria. First, δ must be strictly positive to ensure that the kernel values are bound
 283 between zero and one. Second, the similarity values measured by K capture the intactness of nature
 284 and should span the full range between zero and one. We set δ to the lowest value that met these
 285 criteria, given that increasing values simply shifted the distribution further to the left toward zero
 286 intactness (see Supplement S1).

287 The weight of information is the final key consideration in our kernel estimator. We designed this
 288 framework to integrate numerous biodiversity maps, and we test it here with 75 maps (Table S2) –
 289 ecosystem structure (25), function (12), and connectivity (1); species diversity of plants (11),
 290 microbes (13), and animals (11); and genetic diversity of animals (2). We reprojected all maps to a
 291 common projection (epsg:4326) and spatial resolution (30 arc-seconds, ~926 meters at the equator)
 292 using a nearest-neighbor algorithm. The maps we reviewed vary in several aspects: coverage extent,
 293 spatial resolution, non-independence from other maps, extrapolation across regional data gaps, and
 294 in the degree to which satellite imagery or other measures of local conditions were integrated. We
 295 developed a dynamic weighting system to define the normalized weight of each map in a set,
 296 represented by \mathbf{w} . We grouped non independent layers to share equal weight, which resulted in a
 297 short hierarchical structure. We also applied a confidence score and associated decreasing
 298 confidence with decreasing weight (see Supplement Table S1 for details). Although we found no
 299 global maps of the genetic diversity of plants or microbes, we filled in two blank maps to test the
 300 full framework and set the weight of these maps to a very small nonzero number.

301 We structured the integration of the nine biodiversity axes to be an even-weighted average of the
302 intactness in each dimension. This places genetic, species, and ecosystem level diversity on equal
303 grounds in the integrated index. SEED also offers an index for each biodiversity axis for
304 independent use alongside the integrated index. Even weighting ensures equitability within genetic
305 and species diversity among plants, animals, and microbes, which deviates strongly from the more
306 common case wherein smaller, more cryptic taxa are overlooked in favor of more visible or
307 personable taxa. If, however, evidence emerges that suggests a different weighting scheme is
308 warranted, our method can be adapted in accordance with the evolving scientific landscape. In
309 practice, however, the availability and quality of available maps is not even among axes, which
310 resulted in some axes having higher relative weight and therefore more influence on the overall
311 biocomplexity index (See Supplement S1 for details on how confidence scores affect the relative
312 weights among axes). Identifying the optimal weighting for various levels of biodiversity
313 information remains a key challenge for future biodiversity research as our theoretical
314 understanding of ecological systems evolves.

315 3.2. Reference area versus a counterfactual landscape

316 To measure the intactness of biocomplexity requires establishing a baseline potential state that can
317 be used for comparison. Estimating this baseline state poses several philosophical questions
318 regarding the non-static nature of natural systems. It also poses data limitations for regions of the
319 world that have been heavily degraded and thus lack ‘pristine’ potential states, such as large regions
320 of the Northern Hemisphere⁹⁵. Thus, we opt for a more direct approach than estimating a potential
321 natural state for all biodiversity axes and underlying layers. We identify minimally-modified areas
322 as points of reference with a procedure that involves few assumptions, considers the full suite of
323 biodiversity together, and allows for a contemporary definition of natural as opposed to a historical
324 estimate⁹⁶.

325 Specifically, we developed an algorithm that uses the HMI⁵⁵ and potential natural vegetation
326 (PNV)⁵⁷ to select reference areas for each land cover class within each of the 846 ecoregions⁹⁷ on
327 Earth. This was done separately for each ecoregion to ensure ecologically relevant comparisons. In
328 each ecoregion, we identify the least impacted areas for each land cover class (from the PNV), using
329 a dynamically-defined, upper-limit HMI threshold to ensure a sufficiently large area, enabling

330 representative and robust estimates (see Figure 3a and Supplement S1). The mean value for each
331 biodiversity feature layer within this *least-impacted area* estimates \mathbf{z}_r in equation 1 and is the basis
332 for comparing all feature values in all pixels of the same vegetation class within that ecoregion.

333 Although a natural landscape with minimal anthropogenic disturbance is not a universal goal in all
334 scenarios, this minimal-disturbance baseline provides an objective, and replicable benchmark for
335 measuring the state of nature. For example, food security and financial wellbeing are the primary
336 considerations in agricultural settings, while restoration targets in conservation settings may deviate
337 from a fully natural state due to a myriad of ecological and socioeconomic factors and needs, as
338 well as previous alterations to the landscape⁹⁸. Management practices and targets will vary
339 depending on the local situation, and progress toward these targets can be evaluated against local
340 minimal-disturbance benchmarks and other landscapes in similar settings. In these contexts, the
341 SEED index provides a useful tool to benchmark local achievement against a globally-standardized
342 biodiversity metric and enables a standardized assessment of biodiversity improvement in response
343 to management practices¹⁷.

344 3.3. Illustration of the SEED biocomplexity framework

345 The biocomplexity index was developed using a comprehensive list of the most up-to-date maps of
346 genetic, species, and ecosystem diversity (see Supplement Table S2). Although, our framework is
347 designed to integrate nine biodiversity axes, the current lack of global maps to represent the genetic
348 diversity of plants and microbes limits the current index to seven functioning axes. To illustrate the
349 capabilities of the framework, we first show the viability of our dimensionality reduction
350 calculation (equation 1) and reference area (Figure 3a) approach to calculate the intactness of seven
351 biodiversity axes (Figure 3b-h). Combined, these intactness layers generate a unified measure of
352 the intactness of nature, the SEED biocomplexity index (Figure 4a). We show how SEED can be
353 aggregated across ecological boundaries and at various spatial scales to summarize biocomplexity
354 in total and along each biodiversity axis (Figure 4b-c). Finally, we conduct a cursory comparison
355 of SEED against the leading biodiversity intactness indices (Figure 5).

356 Among the seven biodiversity intactness indices that we calculated, we identified broad global
357 patterns where the indices unanimously show similar levels of intactness (Figure 3). High intactness

358 is nearly universally indicated across tundra, northern boreal forests, deserts, and the Amazon basin
359 – areas that have not experienced extensive human development. Low intactness is indicated in the
360 temperate grasslands and savannas of North America, Brazil's Atlantic Forest, narrow bands along
361 the West African coast and Sub-Saharan Africa, temperate forests across Europe and east across
362 Asia toward Siberia, Northern India, the western edge of Southeast Asia, Northeastern China, and
363 the eastern and southwestern coasts of Australia.

364 There is also notable divergence among these indices. While the intactness of plant and microbial
365 species were very similar (Figure 3f-g), and to some degree aligned with ecosystem structure
366 (Figure 3b), ecosystem function (Figure 3d) shows divergent patterns in some regions, e.g., central
367 North America, Western Ghats, and Central Deccan Plateau. Ecologically, we know very well that
368 plants and microbes share special linkages and that ecosystem structure and function result from
369 the local communities of plants, microbes, and animals. The intactness of animal species diversity
370 (Figure 3h) is also divergent and highlights that the Andes and Appalachian Mountain ranges have
371 particularly low intactness of animal species diversity relative to the other indices in these regions.
372 In the Indo-Gangetic Plains, animal species intactness remains relatively high while other indices
373 predict low intactness. Ecosystem connectivity (Figure 3c) shows a largely unique pattern.
374 Currently the SEED connectivity index is highly sensitive to fragmentation⁸¹, and therefore scores
375 land area as either highly intact or not at all. The genetic diversity of animals (Figure 3e) also shows
376 a unique pattern, but this is likely because we have only two genetic diversity maps with very coarse
377 resolution (~380-km)²⁵. This along with the absence of global models for plant and microbial
378 genetic diversity emphasizes the need for more work in global modeling of genetic diversity.

379 The value in our biocomplexity framework is twofold; it integrates information regarding multiple
380 hierarchical levels of diversity and numerous maps within each level, and it can be aggregated to
381 provide summary statistics at the desired scale. The integrated SEED biocomplexity index
382 represents a single measure of the state of nature for terrestrial land area across the globe (Figure
383 4a). This shows that, on average, the Earth is currently at 68% of its natural state (Figure 4b).
384 Aggregating by administrative boundaries⁹⁹, we can evaluate the state of nature at a level where
385 local policies may directly affect the mechanisms governing the direct human impacts on nature
386 (Figure 4c).

387 In comparison to existing global biodiversity indices, SEED (Figure 4) offers a more
388 comprehensive view of nature's state due to its inclusion of 75 global biodiversity maps, while also
389 offering disaggregated information in the form of seven intactness sub-indices (Figure 3b-h).
390 Existing global intactness indices (BII, and MSA) exclusively represent species level diversity for
391 a subset of taxa. SEED expands the taxa represented with a wealth of microbial datasets (including
392 fungi, bacteria, and archaea), while also expanding in scope to include multiple ecosystem attributes
393 that capture landscape dynamics and ecological feedback. Their inclusion in SEED may account
394 for differences between SEED and the other intactness indices (Figure S1). For example, SEED
395 may indicate low ecosystem intactness where satellite imagery detects ecosystem fragmentation,
396 while models predicting species composition may not register a change in species intactness. Given
397 the global coverage and high temporal resolution of satellite imagery, the inclusion of remote-
398 sensed ecosystem characteristics not only adds dimensionality to biodiversity estimates, but it also
399 improves the spatial and temporal resolution of biodiversity intactness predictions.

400 3.4. Spatial and temporal scalability and next steps

401 There is a growing demand for spatial and temporal scalability in global biodiversity monitoring,
402 but achieving this remains a central challenge for ecology. Determining how biodiversity scales in
403 space and time has been the focus of countless investigations, and one general result is that
404 outcomes are highly context-dependent. Meeting the demand for scalability will require
405 reimagining our approach to biodiversity modeling. Work by Map of Life¹⁰⁰ in association with
406 GEO BON exemplifies the scalable biodiversity models of the future. Map of Life provides a
407 catalog of species distributions predicted over space and time, generated using the most up-to-date
408 remote sensing technologies and species distribution models²⁰. Bringing these maps together, we
409 can reevaluate species richness, phylogenetic and functional diversity, and genetic diversity via
410 declines in species ranges; and we can generate time series for these biodiversity metrics in high
411 spatial resolution. As these new products emerge, they will be fed directly into the SEED framework
412 to update the biocomplexity index and improve the spatial and temporal resolution of the index to
413 allow for more fine-scale assessments of biodiversity and dynamic changes over time.

414 The integration of remote sensing in biodiversity modeling will be the key distinguishing factor
415 that transforms novel approaches from simply predicting general patterns to the prediction of local

416 conditions and the actual state of nature. Overall, we gain more complete spatial coverage from
417 remote sensing than would be possible from field observation, with more reliable information than
418 extrapolation of models¹⁰¹. Remote sensing can provide the means for both direct – many trees and
419 large animals are visible from space – and indirect biodiversity monitoring via the coupling of
420 remote sensing products with ecological models¹⁰². Future advances in remote sensing such as
421 hyper-spectral imaging^{103,104} are expected to provide valuable additional information, which may
422 help assess how plant species and functional biodiversity respond to different practices¹⁰⁵.

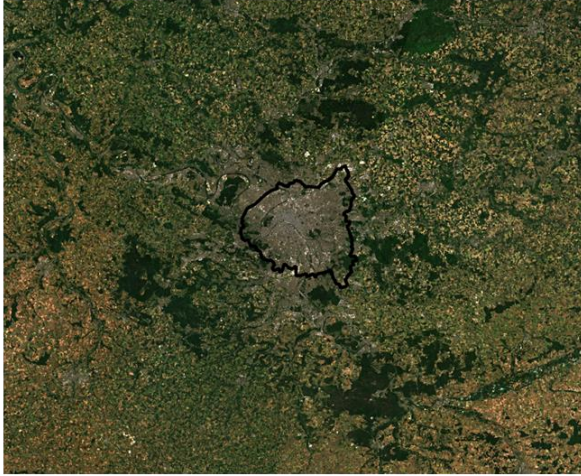
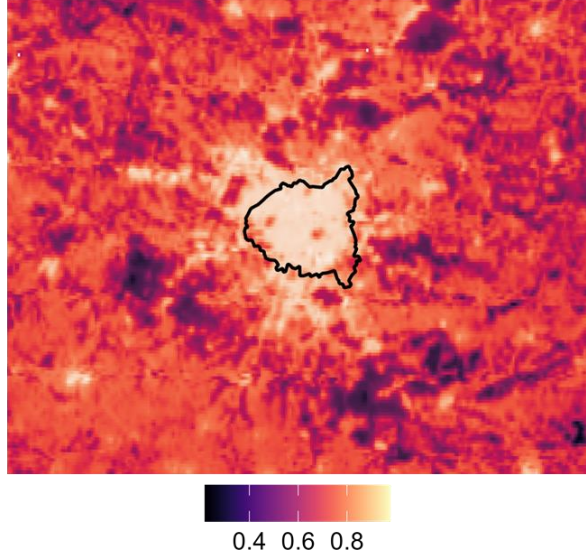
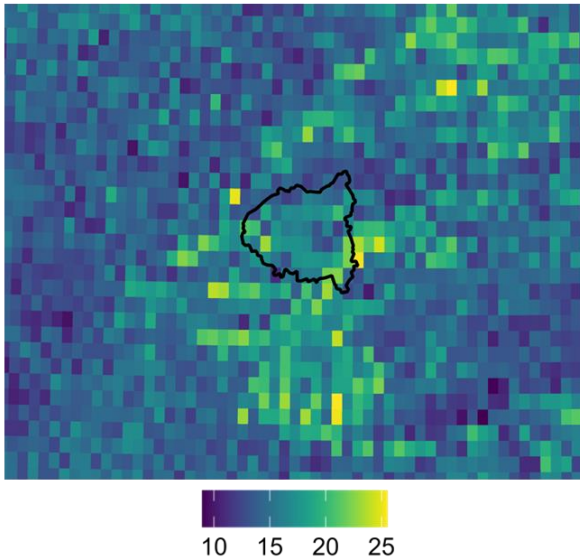
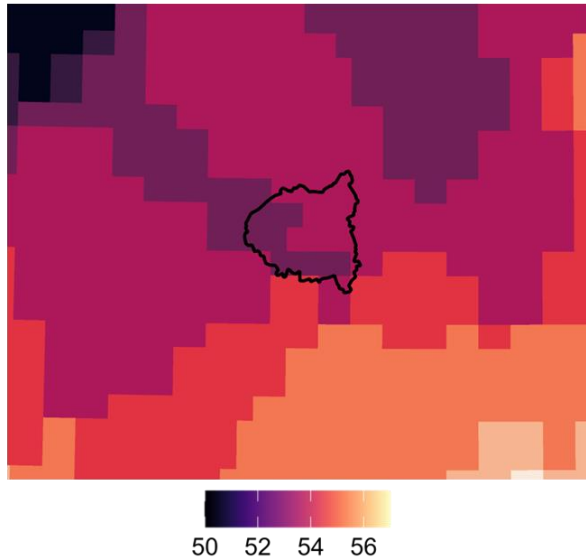
423 Global biodiversity monitoring undoubtedly requires a combination of ground-sourced and remote-
424 sensed approaches. Future on-the-ground sampling in under-sampled regions of the planet and for
425 under-sampled taxa like invertebrates is urgently needed to fill in key gaps and narrow the
426 uncertainty in global genetic and species diversity models. The emergence of next-generation
427 technologies – such as eDNA^{48,106}, bioacoustics¹⁰⁷, and camera traps – may prove to be essential
428 for scaling data collection in a cost-effective manner¹⁰⁸. Incorporation of these emerging
429 technologies is a core aim in the continual development and fine-tuning of the SEED framework to
430 ensure the best possible estimation of biodiversity.

431 **4. Conclusion and outlook**

432 There is growing recognition that we need both the political will and mechanisms to distribute
433 finance towards local efforts that promote biodiversity¹⁰⁹. To support this transition to equitable
434 biodiversity protection, we require globally standardized assessments of ecological intactness that
435 can detect change over time and are available to everyone, everywhere. Given the expansion of
436 global datasets reflecting various dimensions of biodiversity across plants, animals, and microbes,
437 frameworks for integrating this information are now within our grasp. The resulting global
438 assessments of biocomplexity are necessary to evaluate progress toward climate change and land
439 protection pledges (e.g. the GBF, Bonn Challenge¹¹⁰, and UN Sustainable Development Goals¹¹¹),
440 bring transparency to policy frameworks, and to encourage corporate accountability (under the
441 Taskforce for Nature-related Financial Disclosures (TNFD)¹⁰ and Science-Based Targets for Nature
442 (SBTN)¹¹²).

443 The SEED framework consolidates the three primary levels of biodiversity to produce a
444 standardized and comprehensible measure of biocomplexity across the globe. This framework is
445 fundamentally collaborative, incorporating biodiversity assessments and models from a diverse
446 range of biodiversity experts. In addition, it is flexible, allowing for a dynamic index that evolves
447 alongside the development of new map products and scientific advancements. We demonstrate that
448 the SEED framework captures multidimensional changes in biodiversity along seven distinct axes
449 (Figures 3) and provides a unified understanding of the state of nature at multiple scales (Figure 4).
450 Yet, there are several key challenges that need to be addressed to improve the resolution of our
451 index at finer spatial and temporal scales. Most notably, the development of models that integrate
452 remote sensing data products into maps of species richness and genetic diversity will greatly
453 improve the spatial and temporal resolution of our index.

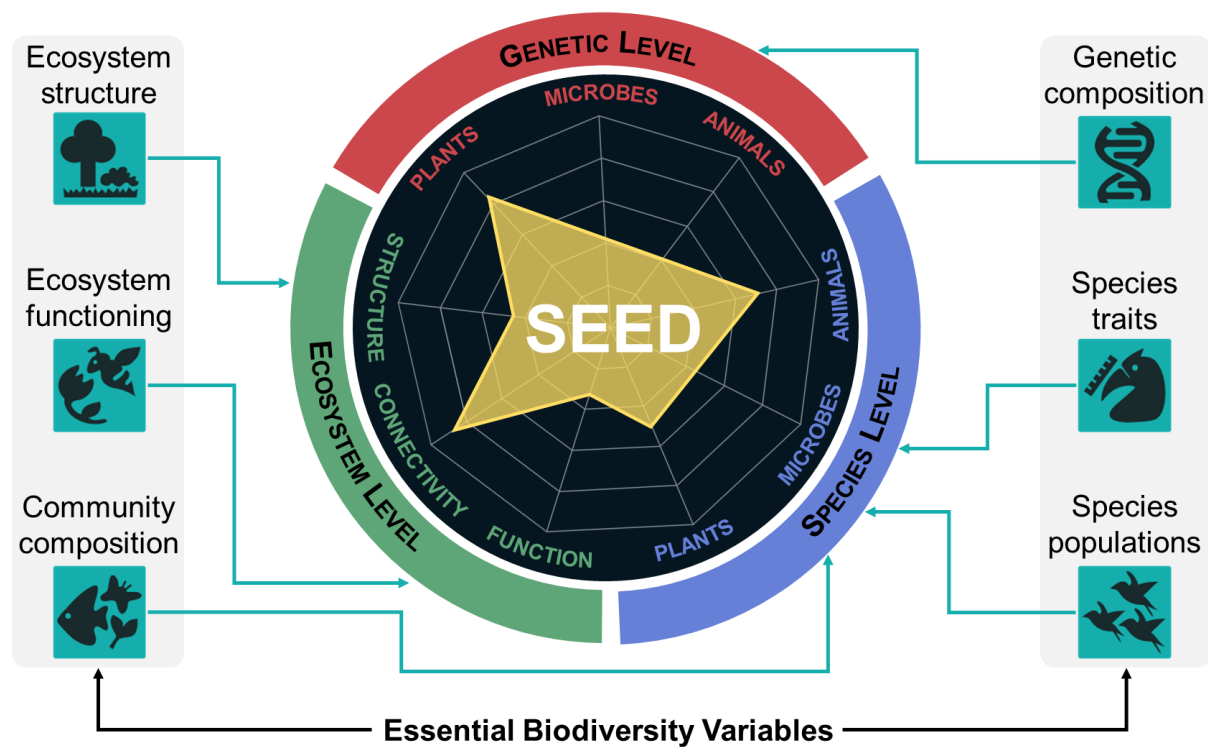
454 To ensure that the most high-integrity and up-to-date ecological information is available to policy-
455 and decision-makers, we invite the wider scientific community to collaborate on the continual
456 advancement of this biocomplexity index (www.seed-index.com). We believe that for this
457 biodiversity index to lead to positive outcomes, its satellite-based predictions must accurately
458 represent the actual biodiversity at a site. Achieving this level of accuracy presents a scientific
459 challenge that requires the collaboration of a diverse team of experts, including ecologists, remote-
460 sensing specialists, and others from around the globe. Leveraging the latest scientific and
461 technological breakthroughs, we aim to enhance and refine the biocomplexity index, thus fostering
462 better outcomes for ecosystem protection and restoration.

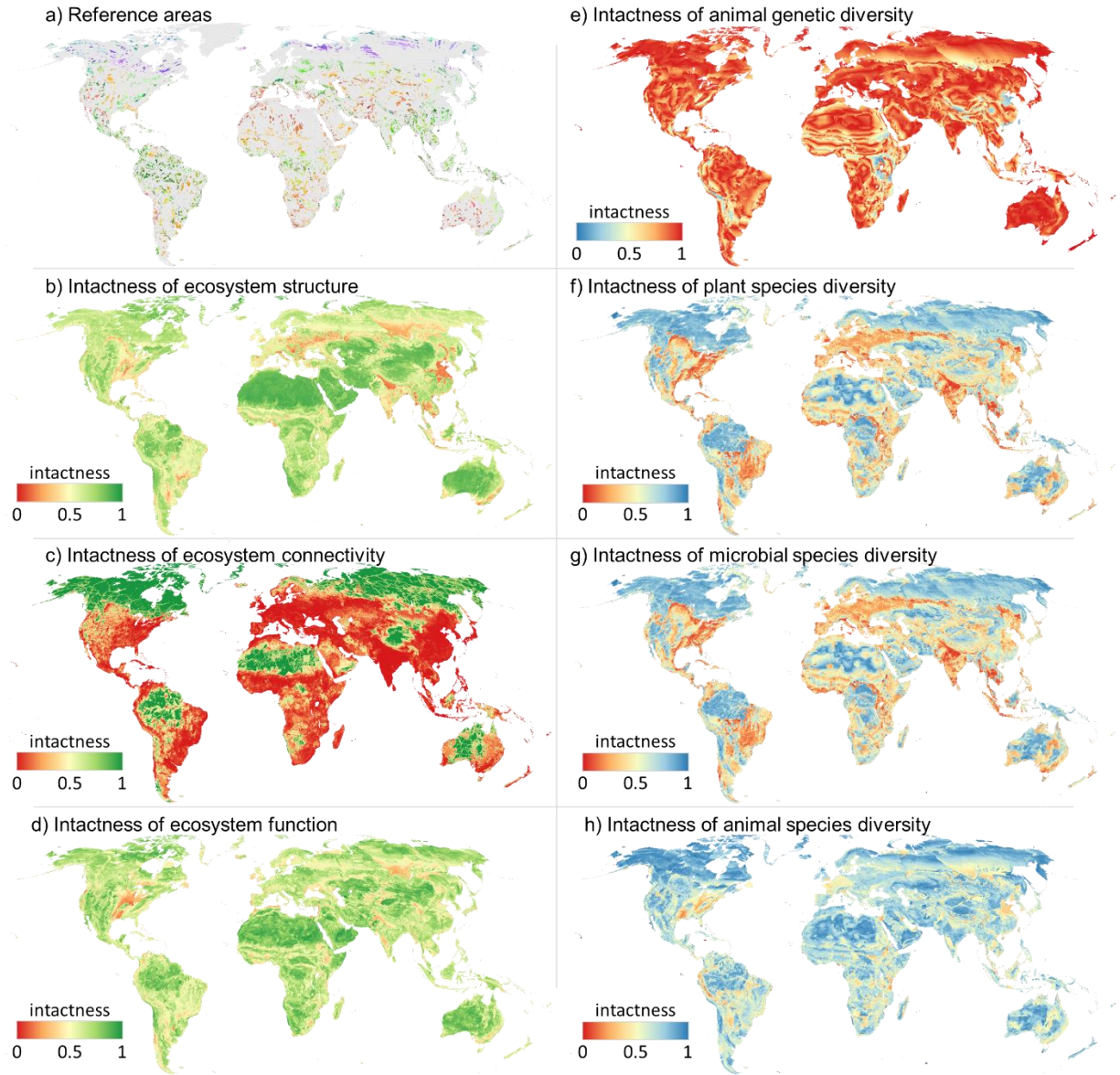
463 **Figures**a. **Aerial View**b. **Human Modification Index**c. **Plant Richness**d. **Mammal Richness**

464

465 **Figure 1. Poor performance of species richness maps in human modified landscapes.** The
 466 impacts of human activities – as is shown here for Paris (outline) and the surrounding landscape (a)
 467 in satellite imagery¹¹³ and (b) in the human modification index⁵⁵ – were not included in the
 468 development of most maps of species richness, e.g., (c) plant¹¹⁴ and (d) mammals³⁴.

469





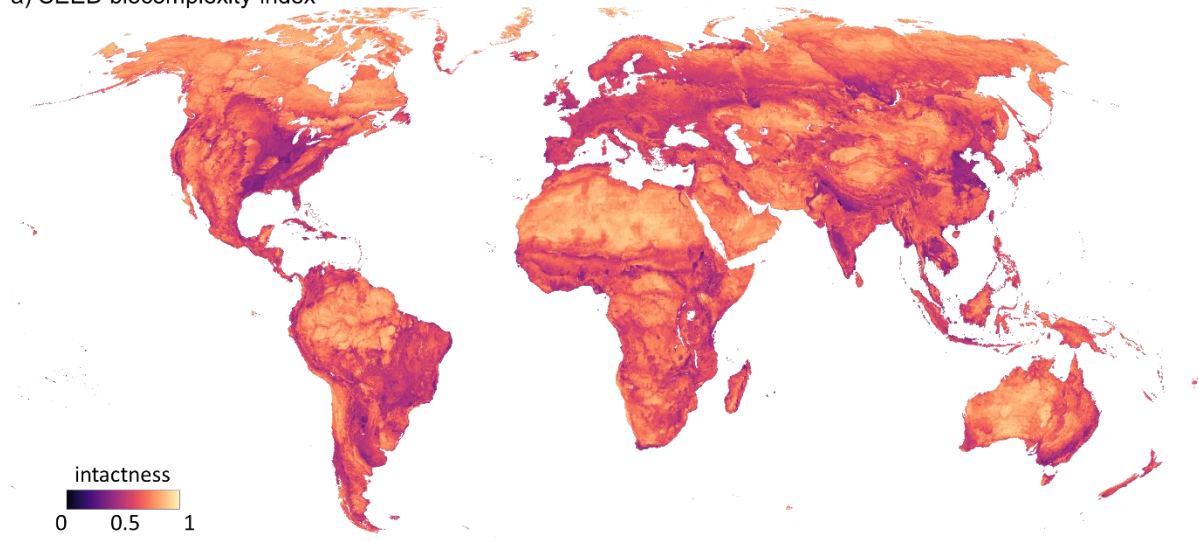
479

480 **Figure 3. A reference area approach to measuring intactness along seven axes of biodiversity.**

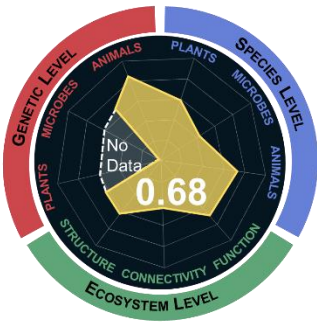
481 Series of maps showing the intactness – relative to (a) minimally impacted reference areas (colored
 482 by ecoregion⁹⁷) – of diversity along seven biodiversity axes: ecosystem (b) structure, (c)
 483 connectivity, and (d) function, genetic diversity of (e) animals, and species diversity of (f) plants,
 484 (g) microbes, and (h) animals.

485

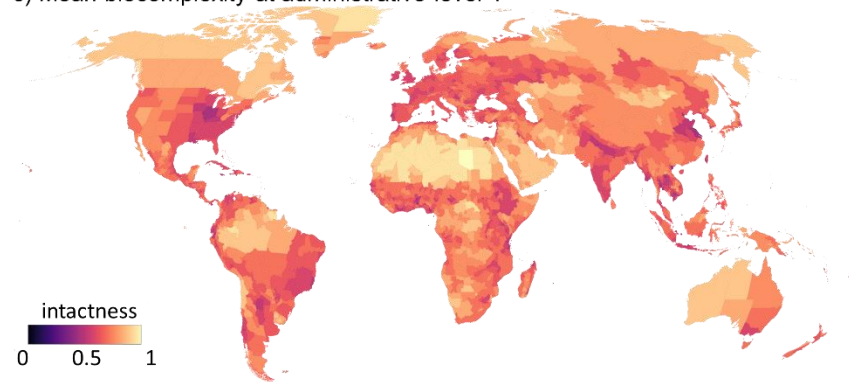
a) SEED biocomplexity index



b) Global SEED analysis



c) Mean biocomplexity at administrative level 1



486

487 **Figure 4. Global analysis of biocomplexity.** Map of the (a) SEED biocomplexity index, which
 488 measures the intactness of biodiversity for every 1-km of pixel of land. SEED can be aggregated
 489 across ecological boundaries, as is demonstrated with (b) a global analysis and (c) the summaries
 490 by administrative boundaries.

491

492 **Resource availability**

493 **Lead contact:** Requests for further information and resources should be directed to and will be
494 fulfilled by the lead contact, Robert McElderry (robert.mcelderry@usys.ethz.ch).

495 **Materials availability:** All unique biodiversity indices generated in this study may be accessed at
496 <https://doi.org/10.5281/zenodo.13799961>

497 **Data and code availability:** This paper analyzes existing, publicly available data. We compiled a
498 list of the sources where these data may be accessed [[See Excel file](#)]. All original code is currently
499 unavailable.

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505 **Author contributions:**

506 This work was conceptualized by TWC, DM, JvdH, TE, RMM, and CFL. RMM, CFL, CEL, FS,
507 PC, TWC, and JvdH developed the methodology while CEL, FS, PC, RMM, TL, CFL, and JvdH
508 composed the software. Data curation, formal analyses, and validation were performed by RMM
509 and CFL with assistance from CEL, SM, PC, JvdH and TL. Resources were curated by TL, CFL,
510 CEL, PC, and RMM. The original draft was composed by TWC, RMM, CFL, and IB, followed by
511 revisions by all authors. Visuals were envisioned by TWC, RMM, and IB, and created by CFL and
512 RMM. TWC and RMM supervised this work, and funding was acquired by TWC, TWE, RMM,
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514 **Declaration of interests:**

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518 member of Restor's Science Advisory Council.

519 **Supplemental information:**

520 **Document S1.** Additional methods, Table S1, and Figure S1

521 **Table S2.** List of layers incorporated in SEED. [See Excel file.](#)

522 Web-based map viewer. [https://robertmcelderry.users.earthengine.app/view/seed-biocomplexity-](https://robertmcelderry.users.earthengine.app/view/seed-biocomplexity-global-beta)
523 [global-beta](https://robertmcelderry.users.earthengine.app/view/seed-biocomplexity-global-beta)

524 **6. References:**

- 525 1. Henle, K., Alard, D., Clitherow, J., Cobb, P., Firbank, L., Kull, T., McCracken, D., Moritz,
526 R.F.A., Niemelä, J., Rebane, M., et al. (2008). Identifying and managing the conflicts between
527 agriculture and biodiversity conservation in Europe—A review. *Agric. Ecosyst. Environ.* *124*,
528 60–71. <https://doi.org/10.1016/j.agee.2007.09.005>.
- 529 2. Lécuyer, L., Alard, D., Calla, S., Coolsaet, B., Fickel, T., Heinsoo, K., Henle, K., Herzon, I.,
530 Hodgson, I., Quétier, F., et al. (2021). Conflicts between agriculture and biodiversity
531 conservation in Europe: Looking to the future by learning from the past. In *Advances in*
532 *Ecological Research* (Elsevier), pp. 3–56. <https://doi.org/10.1016/bs.aecr.2021.10.005>.
- 533 3. Lockwood, J.L., and McKinney, M.L. eds. (2001). *Biotic Homogenization* (Springer US)
534 <https://doi.org/10.1007/978-1-4615-1261-5>.
- 535 4. Bremer, L.L., and Farley, K.A. (2010). Does plantation forestry restore biodiversity or create
536 green deserts? A synthesis of the effects of land-use transitions on plant species richness.
537 *Biodivers. Conserv.* *19*, 3893–3915. <https://doi.org/10.1007/s10531-010-9936-4>.
- 538 5. Armenteras, D., Rodríguez, N., and Retana, J. (2015). National and regional relationships of
539 carbon storage and tropical biodiversity. *Biol. Conserv.* *192*, 378–386.
540 <https://doi.org/10.1016/j.biocon.2015.10.014>.
- 541 6. Lima, T.A., and Vieira, G. (2013). High plant species richness in monospecific tree plantations
542 in the Central Amazon. *For. Ecol. Manag.* *295*, 77–86.
543 <https://doi.org/10.1016/j.foreco.2013.01.006>.
- 544 7. Barlow, J., Gardner, T.A., Araujo, I.S., Ávila-Pires, T.C., Bonaldo, A.B., Costa, J.E., Esposito,
545 M.C., Ferreira, L.V., Hawes, J., Hernandez, M.I.M., et al. (2007). Quantifying the biodiversity
546 value of tropical primary, secondary, and plantation forests. *Proc. Natl. Acad. Sci.* *104*, 18555–
547 18560. <https://doi.org/10.1073/pnas.0703333104>.
- 548 8. Hulvey, K.B., Hobbs, R.J., Standish, R.J., Lindenmayer, D.B., Lach, L., and Perring, M.P.
549 (2013). Benefits of tree mixes in carbon plantings. *Nat. Clim. Change* *3*, 869–874.
550 <https://doi.org/10.1038/nclimate1862>.
- 551 9. Conference of the Parties to the Convention on Biological Diversity (2022). Decision 15/4.
- 552 10. TNFD – Taskforce on Nature-related Financial Disclosures TNFD. <https://tnfd.global/>.

- 553 11. Geller, G.N., Halpin, P.N., Helmuth, B., Hestir, E.L., Skidmore, A., Abrams, M.J., Aguirre,
554 N., Blair, M., Botha, E., Colloff, M., et al. (2017). Remote Sensing for Biodiversity. In *The*
555 *GEO Handbook on Biodiversity Observation Networks*, M. Walters and R. J. Scholes, eds.
556 (Springer International Publishing), pp. 187–210. [https://doi.org/10.1007/978-3-319-27288-](https://doi.org/10.1007/978-3-319-27288-7_8)
557 [7_8](https://doi.org/10.1007/978-3-319-27288-7_8).
- 558 12. Witmer, G.W. (2005). Wildlife population monitoring: some practical considerations. *Wildl.*
559 *Res.* 32, 259–263. <https://doi.org/10.1071/WR04003>.
- 560 13. Adam, E., Mutanga, O., and Rugege, D. (2010). Multispectral and hyperspectral remote
561 sensing for identification and mapping of wetland vegetation: a review. *Wetl. Ecol. Manag.*
562 18, 281–296. <https://doi.org/10.1007/s11273-009-9169-z>.
- 563 14. LaRue, E.A., Fahey, R.T., Alveshere, B.C., Atkins, J.W., Bhatt, P., Buma, B., Chen, A.,
564 Cousins, S., Elliott, J.M., Elmore, A.J., et al. (2023). A theoretical framework for the
565 ecological role of three-dimensional structural diversity. *Front. Ecol. Environ.* 21, 4–13.
566 <https://doi.org/10.1002/fee.2587>.
- 567 15. Bishop-Taylor, R., Tulbure, M.G., and Broich, M. (2018). Evaluating static and dynamic
568 landscape connectivity modelling using a 25-year remote sensing time series. *Landsc. Ecol.*
569 33, 625–640. <https://doi.org/10.1007/s10980-018-0624-1>.
- 570 16. Gonzalez, A., Vihervaara, P., Balvanera, P., Bates, A.E., Bayraktarov, E., Bellingham, P.J.,
571 Bruder, A., Campbell, J., Catchen, M.D., Cavender-Bares, J., et al. (2023). A global
572 biodiversity observing system to unite monitoring and guide action. *Nat. Ecol. Evol.*, 1–5.
573 <https://doi.org/10.1038/s41559-023-02171-0>.
- 574 17. Pereira, H.M., Ferrier, S., Walters, M., Geller, G.N., Jongman, R.H.G., Scholes, R.J., Bruford,
575 M.W., Brummitt, N., Butchart, S.H.M., Cardoso, A.C., et al. (2013). Essential Biodiversity
576 Variables. *Science* 339, 277–278. <https://doi.org/10.1126/science.1229931>.
- 577 18. Walters, M., and Scholes, R.J. eds. (2017). *The GEO Handbook on Biodiversity Observation*
578 *Networks* (Springer International Publishing) <https://doi.org/10.1007/978-3-319-27288-7>.
- 579 19. Michener, W.K., Baerwald, T.J., Firth, P., Palmer, M.A., Rosenberger, J.L., Sandlin, E.A.,
580 and Zimmerman, H. (2001). Defining and Unraveling Biocomplexity. *BioScience* 51, 1018.
581 [https://doi.org/10.1641/0006-3568\(2001\)051\[1018:DAUB\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2001)051[1018:DAUB]2.0.CO;2).
- 582 20. Jetz, W., McGeoch, M.A., Guralnick, R., Ferrier, S., Beck, J., Costello, M.J., Fernandez, M.,
583 Geller, G.N., Keil, P., Merow, C., et al. (2019). Essential biodiversity variables for mapping
584 and monitoring species populations. *Nat. Ecol. Evol.* 3, 539–551.
585 <https://doi.org/10.1038/s41559-019-0826-1>.
- 586 21. Peterson, A.T., and Soberón, J. (2018). Essential biodiversity variables are not global.
587 *Biodivers. Conserv.* 27, 1277–1288. <https://doi.org/10.1007/s10531-017-1479-5>.

- 588 22. O'Brien, D., Laikre, L., Hoban, S., Bruford, M.W., Ekblom, R., Fischer, M.C., Hall, J.,
589 Hvilsom, C., Hollingsworth, P.M., Kershaw, F., et al. (2022). Bringing together approaches
590 to reporting on within species genetic diversity. *J. Appl. Ecol.* *59*, 2227–2233.
591 <https://doi.org/10.1111/1365-2664.14225>.
- 592 23. Hoban, S., Archer, F.I., Bertola, L.D., Bragg, J.G., Breed, M.F., Bruford, M.W., Coleman,
593 M.A., Ekblom, R., Funk, W.C., Grueber, C.E., et al. (2022). Global genetic diversity status
594 and trends: towards a suite of Essential Biodiversity Variables (EBVs) for genetic
595 composition. *Biol. Rev.* *97*, 1511–1538. <https://doi.org/10.1111/brv.12852>.
- 596 24. Miraldo, A., Li, S., Borregaard, M.K., Flórez-Rodríguez, A., Gopalakrishnan, S., Rizvanovic,
597 M., Wang, Z., Rahbek, C., Marske, K.A., and Nogués-Bravo, D. (2016). An Anthropocene
598 map of genetic diversity. *Science* *353*, 1532–1535. <https://doi.org/10.1126/science.aaf4381>.
- 599 25. Theodoridis, S., Fordham, D.A., Brown, S.C., Li, S., Rahbek, C., and Nogués-Bravo, D.
600 (2020). Evolutionary history and past climate change shape the distribution of genetic
601 diversity in terrestrial mammals. *Nat. Commun.* *11*, 2557. [https://doi.org/10.1038/s41467-](https://doi.org/10.1038/s41467-020-16449-5)
602 [020-16449-5](https://doi.org/10.1038/s41467-020-16449-5).
- 603 26. French, C.M., Bertola, L.D., Carnaval, A.C., Economo, E.P., Kass, J.M., Lohman, D.J.,
604 Marske, K.A., Meier, R., Overcast, I., Rominger, A.J., et al. (2023). Global determinants of
605 insect mitochondrial genetic diversity. *Nat. Commun.* *14*, 5276.
606 <https://doi.org/10.1038/s41467-023-40936-0>.
- 607 27. Wesse, C., Welk, E., Hurka, H., and Neuffer, B. (2021). Geographical pattern of genetic
608 diversity in *Capsella bursa-pastoris* (Brassicaceae)—A global perspective. *Ecol. Evol.* *11*,
609 199–213. <https://doi.org/10.1002/ece3.7010>.
- 610 28. Exposito-Alonso, M., Booker, T.R., Czech, L., Gillespie, L., Hateley, S., Kyriazis, C.C., Lang,
611 P.L.M., Leventhal, L., Nogués-Bravo, D., Pagowski, V., et al. (2022). Genetic diversity loss
612 in the Anthropocene. *Science* *377*, 1431–1435. <https://doi.org/10.1126/science.abn5642>.
- 613 29. Canteri, E., Fordham, D.A., Li, S., Hosner, P.A., Rahbek, C., and Nogués-Bravo, D. (2021).
614 IUCN Red List protects avian genetic diversity. *Ecography* *44*, 1808–1811.
615 <https://doi.org/10.1111/ecog.05895>.
- 616 30. Ledger, S.E.H., Loh, J., Almond, R., Böhm, M., Clements, C.F., Currie, J., Deinet, S.,
617 Galewski, T., Grooten, M., Jenkins, M., et al. (2023). Past, present, and future of the Living
618 Planet Index. *Npj Biodivers.* *2*, 12. <https://doi.org/10.1038/s44185-023-00017-3>.
- 619 31. Adamowicz, S.J., and Steinke, D. (2015). Increasing global participation in genetics research
620 through DNA barcoding. *Genome* *58*, 519–526. <https://doi.org/10.1139/gen-2015-0130>.
- 621 32. Faith, D.P. (1992). Conservation evaluation and phylogenetic diversity. *Biol. Conserv.* *61*, 1–
622 10. [https://doi.org/10.1016/0006-3207\(92\)91201-3](https://doi.org/10.1016/0006-3207(92)91201-3).

- 623 33. Graham, C.H., and Hijmans, R.J. (2006). A comparison of methods for mapping species
624 ranges and species richness. *Glob. Ecol. Biogeogr.* *15*, 578–587.
625 <https://doi.org/10.1111/j.1466-8238.2006.00257.x>.
- 626 34. Jenkins, C.N., Pimm, S.L., and Joppa, L.N. (2013). Global patterns of terrestrial vertebrate
627 diversity and conservation. *Proc. Natl. Acad. Sci.* *110*, E2602–E2610.
628 <https://doi.org/10.1073/pnas.1302251110>.
- 629 35. Roll, U., Feldman, A., Novosolov, M., Allison, A., Bauer, A.M., Bernard, R., Böhm, M.,
630 Castro-Herrera, F., Chirio, L., Collen, B., et al. (2017). The global distribution of tetrapods
631 reveals a need for targeted reptile conservation. *Nat. Ecol. Evol.* *1*, 1677–1682.
632 <https://doi.org/10.1038/s41559-017-0332-2>.
- 633 36. Cai, L., Kreft, H., Taylor, A., Denelle, P., Schrader, J., Essl, F., van Kleunen, M., Pergl, J.,
634 Pyšek, P., Stein, A., et al. (2023). Global models and predictions of plant diversity based on
635 advanced machine learning techniques. *New Phytol.* *237*, 1432–1445.
636 <https://doi.org/10.1111/nph.18533>.
- 637 37. Ellis, E.C., Antill, E.C., and Kreft, H. (2012). All Is Not Loss: Plant Biodiversity in the
638 Anthropocene. *PLOS ONE* *7*, e30535. <https://doi.org/10.1371/journal.pone.0030535>.
- 639 38. Kreft, H., and Jetz, W. (2007). Global patterns and determinants of vascular plant diversity.
640 *Proc. Natl. Acad. Sci.* *104*, 5925–5930. <https://doi.org/10.1073/pnas.0608361104>.
- 641 39. Fritz, S.A., and Rahbek, C. (2012). Global patterns of amphibian phylogenetic diversity. *J.*
642 *Biogeogr.* *39*, 1373–1382. <https://doi.org/10.1111/j.1365-2699.2012.02757.x>.
- 643 40. Wolf, S., Mahecha, M.D., Sabatini, F.M., Wirth, C., Bruehlheide, H., Kattge, J., Moreno
644 Martínez, Á., Mora, K., and Kattenborn, T. (2022). Citizen science plant observations encode
645 global trait patterns. *Nat. Ecol. Evol.* *6*, 1850–1859. [https://doi.org/10.1038/s41559-022-](https://doi.org/10.1038/s41559-022-01904-x)
646 [01904-x](https://doi.org/10.1038/s41559-022-01904-x).
- 647 41. Rodrigues, A.S.L., Pilgrim, J.D., Lamoreux, J.F., Hoffmann, M., and Brooks, T.M. (2006).
648 The value of the IUCN Red List for conservation. *Trends Ecol. Evol.* *21*, 71–76.
649 <https://doi.org/10.1016/j.tree.2005.10.010>.
- 650 42. Howard, C., Flather, C.H., and Stephens, P.A. (2020). A global assessment of the drivers of
651 threatened terrestrial species richness. *Nat. Commun.* *11*, 993. [https://doi.org/10.1038/s41467-](https://doi.org/10.1038/s41467-020-14771-6)
652 [020-14771-6](https://doi.org/10.1038/s41467-020-14771-6).
- 653 43. Hughes, A.C., Orr, M.C., Yang, Q., and Qiao, H. (2021). Effectively and accurately mapping
654 global biodiversity patterns for different regions and taxa. *Glob. Ecol. Biogeogr.* *30*, 1375–
655 1388. <https://doi.org/10.1111/geb.13304>.
- 656 44. Jung, M., Arnell, A., de Lamo, X., García-Rangel, S., Lewis, M., Mark, J., Merow, C., Miles,
657 L., Ondo, I., Pironon, S., et al. (2021). Areas of global importance for conserving terrestrial

- 658 biodiversity, carbon and water. *Nat. Ecol. Evol.* 5, 1499–1509.
659 <https://doi.org/10.1038/s41559-021-01528-7>.
- 660 45. Mueller, G.M., Cunha, K.M., May, T.W., Allen, J.L., Westrip, J.R.S., Canteiro, C., Costa-
661 Rezende, D.H., Drechsler-Santos, E.R., Vasco-Palacios, A.M., Ainsworth, A.M., et al. (2022).
662 What Do the First 597 Global Fungal Red List Assessments Tell Us about the Threat Status
663 of Fungi? *Diversity* 14, 736. <https://doi.org/10.3390/d14090736>.
- 664 46. Locey, K.J., and Lennon, J.T. (2016). Scaling laws predict global microbial diversity. *Proc.*
665 *Natl. Acad. Sci.* 113, 5970–5975. <https://doi.org/10.1073/pnas.1521291113>.
- 666 47. Larsen, B.B., Miller, E.C., Rhodes, M.K., and Wiens, J.J. (2017). Inordinate Fondness
667 Multiplied and Redistributed: the Number of Species on Earth and the New Pie of Life. *Q.*
668 *Rev. Biol.* 92, 229–265. <https://doi.org/10.1086/693564>.
- 669 48. Ruppert, K.M., Kline, R.J., and Rahman, M.S. (2019). Past, present, and future perspectives
670 of environmental DNA (eDNA) metabarcoding: A systematic review in methods, monitoring,
671 and applications of global eDNA. *Glob. Ecol. Conserv.* 17, e00547.
672 <https://doi.org/10.1016/j.gecco.2019.e00547>.
- 673 49. Větrovský, T., Morais, D., Kohout, P., Lepinay, C., Algora, C., Awokunle Hollá, S.,
674 Bahnmann, B.D., Bílohnědá, K., Brabcová, V., D’Alò, F., et al. (2020). GlobalFungi, a global
675 database of fungal occurrences from high-throughput-sequencing metabarcoding studies. *Sci.*
676 *Data* 7, 228. <https://doi.org/10.1038/s41597-020-0567-7>.
- 677 50. Mikryukov, V., Dulya, O., Zizka, A., Bahram, M., Hagh-Doust, N., Anslan, S., Prylutskyi, O.,
678 Delgado-Baquerizo, M., Maestre, F.T., Nilsson, H., et al. (2023). Connecting the multiple
679 dimensions of global soil fungal diversity. *Sci. Adv.* 9, eadj8016.
680 <https://doi.org/10.1126/sciadv.adj8016>.
- 681 51. Li, P., Tedersoo, L., Crowther, T.W., Wang, B., Shi, Y., Kuang, L., Li, T., Wu, M., Liu, M.,
682 Luan, L., et al. (2023). Global diversity and biogeography of potential phytopathogenic fungi
683 in a changing world. *Nat. Commun.* 14, 6482. <https://doi.org/10.1038/s41467-023-42142-4>.
- 684 52. Delgado-Baquerizo, M., Oliverio, A.M., Brewer, T.E., Benavent-González, A., Eldridge, D.J.,
685 Bardgett, R.D., Maestre, F.T., Singh, B.K., and Fierer, N. (2018). A global atlas of the
686 dominant bacteria found in soil. *Science* 359, 320–325.
687 <https://doi.org/10.1126/science.aap9516>.
- 688 53. Sanderson, E.W., Jaiteh, M., Levy, M.A., Redford, K.H., Wannebo, A.V., and Woolmer, G.
689 (2002). The Human Footprint and the Last of the Wild. *BioScience* 52, 891.
690 [https://doi.org/10.1641/0006-3568\(2002\)052\[0891:THFATL\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2002)052[0891:THFATL]2.0.CO;2).
- 691 54. Venter, O., Sanderson, E.W., Magrath, A., Allan, J.R., Beher, J., Jones, K.R., Possingham,
692 H.P., Laurance, W.F., Wood, P., Fekete, B.M., et al. (2016). Global terrestrial Human
693 Footprint maps for 1993 and 2009. *Sci. Data* 3, 160067. <https://doi.org/10.1038/sdata.2016.67>.

- 694 55. Kennedy, C.M., Oakleaf, J.R., Theobald, D.M., Baruch-Mordo, S., and Kiesecker, J. (2019).
695 Managing the middle: A shift in conservation priorities based on the global human
696 modification gradient. *Glob. Change Biol.* 25, 811–826. <https://doi.org/10.1111/gcb.14549>.
- 697 56. Theobald, D.M., Kennedy, C., Chen, B., Oakleaf, J., Baruch-Mordo, S., and Kiesecker, J.
698 (2020). Earth transformed: detailed mapping of global human modification from 1990 to 2017.
699 *Earth Syst. Sci. Data* 12, 1953–1972. <https://doi.org/10.5194/essd-12-1953-2020>.
- 700 57. Jung, M., Dahal, P.R., Butchart, S.H.M., Donald, P.F., De Lamo, X., Lesiv, M., Kapos, V.,
701 Rondinini, C., and Visconti, P. (2020). A global map of terrestrial habitat types. *Sci. Data* 7,
702 256. <https://doi.org/10.1038/s41597-020-00599-8>.
- 703 58. Hansen, M.C., Potapov, P.V., Moore, R., Hancher, M., Turubanova, S.A., Tyukavina, A.,
704 Thau, D., Stehman, S.V., Goetz, S.J., Loveland, T.R., et al. (2013). High-Resolution Global
705 Maps of 21st-Century Forest Cover Change. *Science* 342, 850–853.
706 <https://doi.org/10.1126/science.1244693>.
- 707 59. Lang, N., Jetz, W., Schindler, K., and Wegner, J.D. (2023). A high-resolution canopy height
708 model of the Earth. *Nat. Ecol. Evol.* 7, 1778–1789. [https://doi.org/10.1038/s41559-023-](https://doi.org/10.1038/s41559-023-02206-6)
709 [02206-6](https://doi.org/10.1038/s41559-023-02206-6).
- 710 60. Spawn, S.A., Sullivan, C.C., Lark, T.J., and Gibbs, H.K. (2020). Harmonized global maps of
711 above and belowground biomass carbon density in the year 2010. *Sci. Data* 7, 112.
712 <https://doi.org/10.1038/s41597-020-0444-4>.
- 713 61. Ma, H., Mo, L., Crowther, T.W., Maynard, D.S., Van Den Hoogen, J., Stocker, B.D., Terrer,
714 C., and Zohner, C.M. (2021). The global distribution and environmental drivers of
715 aboveground versus belowground plant biomass. *Nat. Ecol. Evol.* 5, 1110–1122.
716 <https://doi.org/10.1038/s41559-021-01485-1>.
- 717 62. Warner, D.L., Bond-Lamberty, B.P., Jian, J., Stell, E., and Vargas, R. (2019). Carbon
718 Monitoring System (CMS) Global Gridded 1-km Annual Soil Respiration and Uncertainty
719 Derived from SRDB V3. Preprint at ORNL Distributed Active Archive Center,
720 <https://doi.org/10.3334/ORNLDAAC/1736> <https://doi.org/10.3334/ORNLDAAC/1736>.
- 721 63. Tuanmu, M.-N., and Jetz, W. (2015). A global, remote sensing-based characterization of
722 terrestrial habitat heterogeneity for biodiversity and ecosystem modelling. *Glob. Ecol.*
723 *Biogeogr.* 24, 1329–1339. <https://doi.org/10.1111/geb.12365>.
- 724 64. Myneni, R., Knyazikhin, Y., and Park, T. (2021). MODIS/Terra+Aqua Leaf Area Index/FPAR
725 4-Day L4 Global 500m SIN Grid V061. (NASA EOSDIS Land Processes Distributed Active
726 Archive Center). <https://doi.org/10.5067/MODIS/MCD15A3H.061>
727 <https://doi.org/10.5067/MODIS/MCD15A3H.061>.
- 728 65. Cisneros-Araujo, P., Goicolea, T., Mateo-Sánchez, M.C., García-Viñás, J.I., Marchamalo, M.,
729 Mercier, A., and Gastón, A. (2021). The Role of Remote Sensing Data in Habitat Suitability

- 730 and Connectivity Modeling: Insights from the Cantabrian Brown Bear. *Remote Sens.* *13*,
731 1138. <https://doi.org/10.3390/rs13061138>.
- 732 66. Cisneros-Araujo, P., Ramirez-Lopez, M., Juffe-Bignoli, D., Fensholt, R., Muro, J., Mateo-
733 Sánchez, M.C., and Burgess, N.D. (2021). Remote sensing of wildlife connectivity networks
734 and priority locations for conservation in the Southern Agricultural Growth Corridor
735 (SAGCOT) in Tanzania. *Remote Sens. Ecol. Conserv.* *7*, 430–444.
736 <https://doi.org/10.1002/rse2.199>.
- 737 67. Running, S.W., and Zhao, M. Daily GPP and Annual NPP (MOD17A2/A3) Products NASA
738 Earth Observing System MODIS Land Algorithm.
- 739 68. Smith, T., Traxl, D., and Boers, N. (2022). Empirical evidence for recent global shifts in
740 vegetation resilience. *Nat. Clim. Change* *12*, 477–484. [https://doi.org/10.1038/s41558-022-](https://doi.org/10.1038/s41558-022-01352-2)
741 [01352-2](https://doi.org/10.1038/s41558-022-01352-2).
- 742 69. Seddon, A.W.R., Macias-Fauria, M., Long, P.R., Benz, D., and Willis, K.J. (2016). Sensitivity
743 of global terrestrial ecosystems to climate variability. *Nature* *531*, 229–232.
744 <https://doi.org/10.1038/nature16986>.
- 745 70. Giri, C., Ochieng, E., Tieszen, L.L., Zhu, Z., Singh, A., Loveland, T., Masek, J., and Duke, N.
746 (2011). Status and distribution of mangrove forests of the world using earth observation
747 satellite data. *Glob. Ecol. Biogeogr.* *20*, 154–159. [https://doi.org/10.1111/j.1466-](https://doi.org/10.1111/j.1466-8238.2010.00584.x)
748 [8238.2010.00584.x](https://doi.org/10.1111/j.1466-8238.2010.00584.x).
- 749 71. Xu, J., Morris, P.J., Liu, J., and Holden, J. (2018). PEATMAP: Refining estimates of global
750 peatland distribution based on a meta-analysis. *CATENA* *160*, 134–140.
751 <https://doi.org/10.1016/j.catena.2017.09.010>.
- 752 72. T., G., R.M., R.-C., L.V., V., M., H., F., W., E., H., N., H., and D., M. Tropical and Subtropical
753 Wetlands Distribution version 2. <https://doi.org/10.17528/cifor/data.00058>
754 <https://doi.org/10.17528/cifor/data.00058>.
- 755 73. Mendes, S.B., Olesen, J.M., Memmott, J., Costa, J.M., Timóteo, S., Dengucho, A.L., Craveiro,
756 L., and Heleno, R. (2024). Evidence of a European seed dispersal crisis. *Science* *386*, 206–
757 211. <https://doi.org/10.1126/science.ado1464>.
- 758 74. GFBI consortium, Steidinger, B.S., Crowther, T.W., Liang, J., Van Nuland, M.E., Werner,
759 G.D.A., Reich, P.B., Nabuurs, G.J., de-Miguel, S., Zhou, M., et al. (2019). Climatic controls
760 of decomposition drive the global biogeography of forest-tree symbioses. *Nature* *569*, 404–
761 408. <https://doi.org/10.1038/s41586-019-1128-0>.
- 762 75. Barceló, M., Van Bodegom, P.M., and Soudzilovskaia, N.A. (2023). Fine-resolution global
763 maps of root biomass carbon colonized by arbuscular and ectomycorrhizal fungi. *Sci. Data*
764 *10*, 56. <https://doi.org/10.1038/s41597-022-01913-2>.

- 765 76. Rodell, M., Houser, P.R., Jambor, U., Gottschalck, J., Mitchell, K., Meng, C.-J., Arsenault,
766 K., Cosgrove, B., Radakovich, J., Bosilovich, M., et al. (2004). The Global Land Data
767 Assimilation System. *Bull. Am. Meteorol. Soc.* 85, 381–394. [https://doi.org/10.1175/BAMS-](https://doi.org/10.1175/BAMS-85-3-381)
768 85-3-381.
- 769 77. Tuomi, M., Laiho, R., Repo, A., and Liski, J. (2011). Wood decomposition model for boreal
770 forests. *Ecol. Model.* 222, 709–718. <https://doi.org/10.1016/j.ecolmodel.2010.10.025>.
- 771 78. Assis, J.C., Hohlenwerger, C., Metzger, J.P., Rhodes, J.R., Duarte, G.T., da Silva, R.A.,
772 Boesing, A.L., Prist, P.R., and Ribeiro, M.C. (2023). Linking landscape structure and
773 ecosystem service flow. *Ecosyst. Serv.* 62, 101535.
774 <https://doi.org/10.1016/j.ecoser.2023.101535>.
- 775 79. Baude, M., and Meyer, B.C. (2023). Changes in landscape structure and ecosystem services
776 since 1850 analyzed using landscape metrics in two German municipalities. *Ecol. Indic.* 152,
777 110365. <https://doi.org/10.1016/j.ecolind.2023.110365>.
- 778 80. Belote, R.T., Barnett, K., Zeller, K., Brennan, A., and Gage, J. (2022). Examining local and
779 regional ecological connectivity throughout North America. *Landsc. Ecol.* 37, 2977–2990.
780 <https://doi.org/10.1007/s10980-022-01530-9>.
- 781 81. Beyer, H.L., Venter, O., Grantham, H.S., and Watson, J.E.M. (2020). Substantial losses in
782 ecoregion intactness highlight urgency of globally coordinated action. *Conserv. Lett.* 13,
783 e12692. <https://doi.org/10.1111/conl.12692>.
- 784 82. Liu, S., Dong, Y., Deng, L., Liu, Q., Zhao, H., and Dong, S. (2014). Forest fragmentation and
785 landscape connectivity change associated with road network extension and city expansion: A
786 case study in the Lancang River Valley. *Ecol. Indic.* 36, 160–168.
787 <https://doi.org/10.1016/j.ecolind.2013.07.018>.
- 788 83. Hill, S.L.L., Fajardo, J., Maney, C., Harfoot, M., Harrison, M., Guaras, D., Jones, M., Oliva,
789 M.J., Danks, F., Hughes, J., et al. (2022). The Ecosystem Integrity Index: a novel measure of
790 terrestrial ecosystem integrity with global coverage (Ecology)
791 <https://doi.org/10.1101/2022.08.21.504707>.
- 792 84. Newbold, T., Hudson, L.N., Arnell, A.P., Contu, S., De Palma, A., Ferrier, S., Hill, S.L.L.,
793 Hoskins, A.J., Lysenko, I., Phillips, H.R.P., et al. (2016). Has land use pushed terrestrial
794 biodiversity beyond the planetary boundary? A global assessment. *Science* 353, 288–291.
795 <https://doi.org/10.1126/science.aaf2201>.
- 796 85. De Palma, A., Hoskins, A., Gonzalez, R.E., Börger, L., Newbold, T., Sanchez-Ortiz, K.,
797 Ferrier, S., and Purvis, A. (2021). Annual changes in the Biodiversity Intactness Index in
798 tropical and subtropical forest biomes, 2001–2012. *Sci. Rep.* 11, 20249.
799 <https://doi.org/10.1038/s41598-021-98811-1>.

- 800 86. Schipper, A.M., Hilbers, J.P., Meijer, J.R., Antão, L.H., Benítez-López, A., de Jonge, M.M.J.,
801 Leemans, L.H., Scheper, E., Alkemade, R., Doelman, J.C., et al. (2020). Projecting terrestrial
802 biodiversity intactness with GLOBIO 4. *Glob. Change Biol.* 26, 760–771.
803 <https://doi.org/10.1111/gcb.14848>.
- 804 87. Hill, S.L.L., Gonzalez, R., Sanchez-Ortiz, K., Caton, E., Espinoza, F., Newbold, T.,
805 Tylianakis, J., Scharlemann, J.P.W., De Palma, A., and Purvis, A. (2018). Worldwide impacts
806 of past and projected future land-use change on local species richness and the Biodiversity
807 Intactness Index (*Ecology*) <https://doi.org/10.1101/311787>.
- 808 88. Hudson, L.N., Newbold, T., Contu, S., Hill, S.L.L., Lysenko, I., De Palma, A., Phillips,
809 H.R.P., Senior, R.A., Bennett, D.J., Booth, H., et al. (2014). The PREDICTS database: a global
810 database of how local terrestrial biodiversity responds to human impacts. *Ecol. Evol.* 4, 4701–
811 4735. <https://doi.org/10.1002/ece3.1303>.
- 812 89. Hudson, L.N., Newbold, T., Contu, S., Hill, S.L.L., Lysenko, I., De Palma, A., Phillips,
813 H.R.P., Alhusseini, T.I., Bedford, F.E., Bennett, D.J., et al. (2017). The database of the
814 PREDICTS (Projecting Responses of Ecological Diversity In Changing Terrestrial Systems)
815 project. *Ecol. Evol.* 7, 145–188. <https://doi.org/10.1002/ece3.2579>.
- 816 90. Liu, L., Zhu, K., Wurzburger, N., and Zhang, J. (2020). Relationships between plant diversity
817 and soil microbial diversity vary across taxonomic groups and spatial scales. *Ecosphere* 11,
818 e02999. <https://doi.org/10.1002/ecs2.2999>.
- 819 91. Prober, S.M., Leff, J.W., Bates, S.T., Borer, E.T., Firn, J., Harpole, W.S., Lind, E.M.,
820 Seabloom, E.W., Adler, P.B., Bakker, J.D., et al. (2015). Plant diversity predicts beta but not
821 alpha diversity of soil microbes across grasslands worldwide. *Ecol. Lett.* 18, 85–95.
822 <https://doi.org/10.1111/ele.12381>.
- 823 92. Kwok, R. (2018). Ecology’s remote-sensing revolution. *Nature* 556, 137–138.
824 <https://doi.org/10.1038/d41586-018-03924-9>.
- 825 93. Farley, S.S., Dawson, A., Goring, S.J., and Williams, J.W. (2018). Situating Ecology as a Big-
826 Data Science: Current Advances, Challenges, and Solutions. *BioScience* 68, 563–576.
827 <https://doi.org/10.1093/biosci/biy068>.
- 828 94. Abe, S. (2005). Training of Support Vector Machines with Mahalanobis Kernels. In *Artificial*
829 *Neural Networks: Formal Models and Their Applications – ICANN 2005 Lecture Notes in*
830 *Computer Science.*, W. Duch, J. Kacprzyk, E. Oja, and S. Zadrozny, eds. (Springer Berlin
831 Heidelberg), pp. 571–576. https://doi.org/10.1007/11550907_90.
- 832 95. Gallagher, R., and Carpenter, B. (1997). Human-Dominated Ecosystems. *Science* 277, 485–
833 485. <https://doi.org/10.1126/science.277.5325.485>.

- 834 96. Balaguer, L., Escudero, A., Martín-Duque, J.F., Mola, I., and Aronson, J. (2014). The
835 historical reference in restoration ecology: Re-defining a cornerstone concept. *Biol. Conserv.*
836 *176*, 12–20. <https://doi.org/10.1016/j.biocon.2014.05.007>.
- 837 97. Dinerstein, E., Olson, D., Joshi, A., Vynne, C., Burgess, N.D., Wikramanayake, E., Hahn, N.,
838 Palminteri, S., Hedao, P., Noss, R., et al. (2017). An Ecoregion-Based Approach to Protecting
839 Half the Terrestrial Realm. *BioScience* *67*, 534–545. <https://doi.org/10.1093/biosci/bix014>.
- 840 98. Suding, K.N. (2011). Toward an Era of Restoration in Ecology: Successes, Failures, and
841 Opportunities Ahead. *Annu. Rev. Ecol. Evol. Syst.* *42*, 465–487.
842 <https://doi.org/10.1146/annurev-ecolsys-102710-145115>.
- 843 99. Runfola, D., Anderson, A., Baier, H., Crittenden, M., Dowker, E., Fuhrig, S., Goodman, S.,
844 Grimsley, G., Layko, R., Melville, G., et al. (2020). geoBoundaries: A global database of
845 political administrative boundaries. *PLOS ONE* *15*, e0231866.
846 <https://doi.org/10.1371/journal.pone.0231866>.
- 847 100. Jetz, W., McPherson, J.M., and Guralnick, R.P. (2012). Integrating biodiversity distribution
848 knowledge: toward a global map of life. *Trends Ecol. Evol.* *27*, 151–159.
849 <https://doi.org/10.1016/j.tree.2011.09.007>.
- 850 101. Paz, A., Silva, T.S., and Carnaval, A.C. (2022). A framework for near-real time monitoring of
851 diversity patterns based on indirect remote sensing, with an application in the Brazilian
852 Atlantic rainforest. *PeerJ* *10*, e13534. <https://doi.org/10.7717/peerj.13534>.
- 853 102. Turner, W. (2014). Sensing biodiversity. *Science* *346*, 301–302.
854 <https://doi.org/10.1126/science.1256014>.
- 855 103. Cawse-Nicholson, K., Townsend, P.A., Schimel, D., Assiri, A.M., Blake, P.L., Buongiorno,
856 M.F., Campbell, P., Carmon, N., Casey, K.A., Correa-Pabón, R.E., et al. (2021). NASA's
857 surface biology and geology designated observable: A perspective on surface imaging
858 algorithms. *Remote Sens. Environ.* *257*, 112349. <https://doi.org/10.1016/j.rse.2021.112349>.
- 859 104. European Space Agency (2020). Chime (Copernicus Hyperspectral Imaging Mission for the
860 Environment) - eoPortal. <https://www.eoportal.org/satellite-missions/chime-copernicus#eop-quick-facts-section>.
- 862 105. Cavender-Bares, J., Gamon, J.A., Hobbie, S.E., Madritch, M.D., Meireles, J.E., Schweiger,
863 A.K., and Townsend, P.A. (2017). Harnessing plant spectra to integrate the biodiversity
864 sciences across biological and spatial scales. *Am. J. Bot.* *104*, 966–969.
865 <https://doi.org/10.3732/ajb.1700061>.
- 866 106. Deiner, K., Bik, H.M., Mächler, E., Seymour, M., Lacoursière-Roussel, A., Altermatt, F.,
867 Creer, S., Bista, I., Lodge, D.M., de Vere, N., et al. (2017). Environmental DNA
868 metabarcoding: Transforming how we survey animal and plant communities. *Mol. Ecol.* *26*,
869 5872–5895. <https://doi.org/10.1111/mec.14350>.

- 870 107. Sueur, J., and Farina, A. (2015). Ecoacoustics: the Ecological Investigation and Interpretation
871 of Environmental Sound. *Biosemiotics* 8, 493–502. <https://doi.org/10.1007/s12304-015-9248->
872 x.
- 873 108. Buřivalová, Z., Yoh, N., Butler, R.A., Chandra Sagar, H.S.S., and Game, E.T. (2023).
874 Broadening the focus of forest conservation beyond carbon. *Curr. Biol.* 33, R621–R635.
875 <https://doi.org/10.1016/j.cub.2023.04.019>.
- 876 109. Dasgupta, P. (2021). *The Economics of Biodiversity: The Dasgupta Review*.
- 877 110. The Bonn Challenge | Bonchallenge <https://www.bonnchallenge.org/>.
- 878 111. THE 17 GOALS | Sustainable Development <https://sdgs.un.org/goals>.
- 879 112. Science Based Targets Network Sci. Based Targets Netw.
880 <https://sciencebasedtargetsnetwork.org/>.
- 881 113. European Space Agency (2022). Copernicus Sentinel-2 (processed by ESA), 2022, MSI
882 Level-2A BOA Reflectance Product. Collection 1.
- 883 114. Sabatini, F.M., Jiménez-Alfaro, B., Jandt, U., Chytrý, M., Field, R., Kessler, M., Lenoir, J.,
884 Schrod, F., Wiser, S.K., Arfin Khan, M.A.S., et al. (2022). Global patterns of vascular plant
885 alpha diversity. *Nat. Commun.* 13, 4683. <https://doi.org/10.1038/s41467-022-32063-z>.
- 886 115. Sanchez-Ortiz, K., Gonzalez, R.E., De Palma, A., Newbold, T., Hill, S.L.L., Tylianakis, J.M.,
887 Börger, L., Lysenko, I., and Purvis, A. (2019). Land-use and related pressures have reduced
888 biotic integrity more on islands than on mainlands. Preprint, <https://doi.org/10.1101/576546>
889 <https://doi.org/10.1101/576546>.
- 890 116. Impact Observatory and Vizzuality Biodiversity Intactness Index (BII). (Google Earth
891 Engine:projects/ebx-data/assets/earthblox/IO/BIOINTACT).
- 892

893 **Supplement S1**894 **Reference Area Methodology**

895 We developed an approach that offers a contemporary baseline which all global biodiversity maps
 896 may be evaluated. Our algorithm uses the human modification index (HMI)⁵⁵ and potential natural
 897 vegetation (PNV)⁵⁷ to create a reference area mask that delineates the relatively pristine areas on
 898 the globe. The combination of PNV classes and 846 delineated ecoregions⁹⁷ then guides the
 899 assignment of reference areas to all non-reference pixels having the same ecoregion-PNV class.
 900 Due to the large variations in the extent of human modification among different ecoregion
 901 landcover combinations, we designed a dynamic decision tree for selecting a threshold HMI to
 902 define reference pixels and then to link reference to non-reference pixels to obtain the reference
 903 mask.

904 We targeted reference pixels with minimal human modification which we defined as follows. For
 905 each PNV class, v , within each ecoregion, e , we calculated the 5th and 3rd percentile of the HMI,
 906 $P_{0.05}(HMI_{e,v})$ and $P_{0.03}(HMI_{e,v})$. The reference threshold value, $r_{e,v}$ was then set to the maximum
 907 of either $P_{0.05}(HMI_{e,v})$ or $P_{0.03}(HMI_{e,v})$ if they were less than 0.05, or 0.05 (See equation S1
 908 below).

$$909 \quad r_{e,v} = \begin{cases} P_{0.05}(HMI_{e,v}) & \text{if } P_{0.05}(HMI_{e,v}) \leq 0.05 \\ P_{0.03}(HMI_{e,v}) & \text{if } P_{0.05}(HMI_{e,v}) > 0.05 \text{ and } P_{0.03}(HMI_{e,v}) \leq 0.05 \\ 0.05 & \text{if } P_{0.05}(HMI_{e,v}) > 0.05 \text{ and } P_{0.03}(HMI_{e,v}) > 0.05 \end{cases} \quad \text{Equation S1}$$

910 We set these criteria to focus the reference areas in relative unmodified ecoregions, while allowing
 911 a more inclusion in reference where needed, but only to the point of an HMI equal to 0.05. With
 912 the threshold defined for each ecoregion-PNV class, we selected reference areas as all pixels less
 913 than $r_{e,v}$. If an ecoregion-PNV class had fewer than five pixels, we stepped down the reference
 914 threshold – from the $P_{0.05}(HMI_{e,v})$ to $P_{0.03}(HMI_{e,v})$, or from $P_{0.03}(HMI_{e,v})$ to $0.05(x_{e,v}, y_{e,v})$. If
 915 $r_{e,v}$ was 0.05, and the ecoregion-PNV class still possessed fewer than five reference pixels, then we
 916 looked beyond the ecoregion boundary and included all reference pixels of the same PNV class
 917 within the same biome.

918 **Delta calculation**

919 To calculate the delta value for each axis of the seed index, we use the following formula:

920
$$K = e^{-\delta D}$$

921 where K is the kernel, δ is the delta parameter and D is the maximum Manhattan distance of the
 922 axis for all the pixel points. This effectively maps the highest possible distance of the layers of the
 923 axis to the lowest kernel value of the axis possible. In this case we define this lowest kernel value
 924 as 0.001. By solving for δ parameter as other values in the equation are known we can get the delta
 925 value for each of the axis. We then substitute this delta value in the equation to calculate the kernel
 926 for the world of each axis. The SEED index is then calculated based on the weighted average of the
 927 individual axes.

928 **Table S1.** Table of scores, weights, and a description of the criteria guiding our judgment in rating
 929 the quality of information contained with each map that we incorporated in SEED.

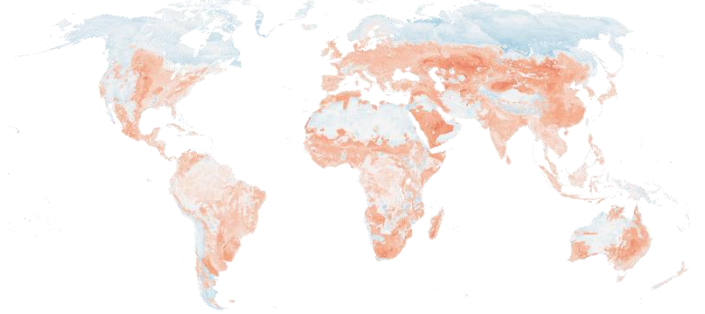
Confidence Score	Weight	Description
5	100	Fully validated model representing on the ground conditions with at least 1-km resolution
4	90	Partially validated model representing on the ground conditions with at least 1-km resolution
3	50	Model predicting on the ground conditions with at least 5-km resolution, requires validation
2	10	Model predicting general diversity patterns with some human impacts considered
1	1	Model of general diversity patterns. Does not represent human impacts on nature.

930

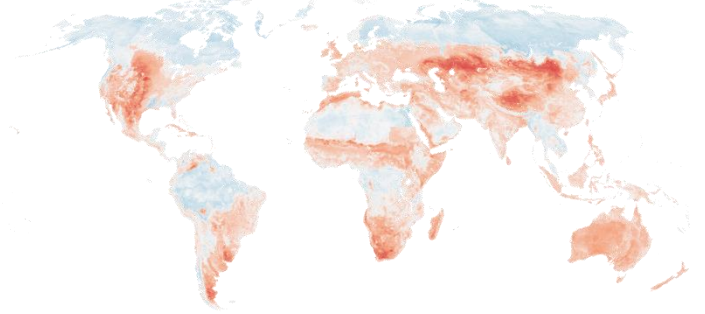
931 To compare SEED with three of the most commonly used global indices, we computed the
 932 difference between SEED and the MSA from GLOBIO⁸⁶, the BII from the Natural History
 933 Museum¹¹⁵, and a second BII (which we will call BDI) produced with a less recent version of the
 934 same core methodology^{84,116}. The broad scale differences in our index indicate that SEED may be
 935 more conservative than MSA and BII (Figure S1a-b) in boreal and desert regions, while SEED

936 estimates higher intactness across temperate regions, sub-Saharan Africa, South Africa, and parts
937 of Australia (Figure S1a-b). SEED was consistently lower than the BDI globally (Figure S1c).
938 Underlying these differences, the distribution of MSA and BII values are bimodal compared with
939 the BDI and SEED, which are unimodal. This means that MSA and BII may more commonly
940 characterize an ecosystem as either highly intact or not very intact, while SEED would more
941 commonly characterize the same ecosystem as moderately intact.

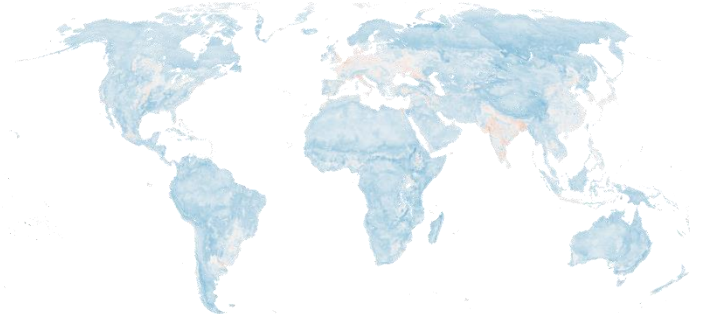
a) Mean Species Abundance (GLOBIO)



b) Biodiversity Intactness Index (Natural History Museum)



c) Biodiversity Intactness Index (Impact Observatory and Vizzuality)



SEED < BII or MSA Equal SEED > BII or MSA

943 **Figure S1. Comparison with leading global biodiversity models.** Global showing the difference
944 between SEED and (a) MSA⁸⁶, (b) BII from NHM¹¹⁵, and (c) BDI, which was created
945 independently from the BII while using the same model¹¹⁶.