

# Fixation probability and fixation time under strong recurrent mutation

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## Abstract

When the mutation rate is high and/or the population size is large, recurrent mutation can lead to multiple, independently generated copies of the same beneficial allele spreading through the population. However, classical analyses of fixation probability and time assume that the mutation rate is low and therefore, that fixation and extinction of a beneficial allele occur faster than the appearance of additional copies. We developed a diffusion equation approximation for the fixation probability and time that accounts for recurrent mutation, incomplete fixation, and fixation from standing genetic variation. Our results show that when the number of new beneficial alleles per generation in the population is greater than one, fixation is guaranteed, and fixation time is significantly lower than expected by the standard approximation. Moreover, we show that fixation time is significantly shorter if the initial allele frequency is greater than 0, or if fixation is defined for an allele frequency lower than 1.

**Keywords.** mutation, fixation probability, fixation time, diffusion equation approximation, wright-fisher model, evolutionary theory

**Competing Interests.** The authors declare no competing interests.

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# Introduction

30 Analyses of the fixation probability and time of beneficial mutations have mostly focused on scenarios  
31 in which the appearance of the beneficial allele is rare enough so that it becomes extinct or fixed  
32 before the next beneficial allele appears (Kimura, 1962, Kimura and Ohta, 1969). However, adaption  
33 sometimes occur in large populations and/or with high mutation rates, leading to high mutation supply  
34 (mutation rate  $\times$  population size), such that multiple new mutants are produced in every generation,  
35 rather than many generations passing between the occurrence of each mutant. Some examples are  
36 the rapid appearance of many, selectively similar, beneficial alleles (Karasov et al., 2010, Barroso-  
37 Batista et al., 2014, Levy et al., 2015, Nguyen Ba et al., 2019); the quick appearance and fixation of  
38 large-effect, rapidly forming, copy number variants (Egan et al., 2007, Yona et al., 2012, Payen et al.,  
39 2014, Avecilla et al., 2022); and evidence for mutation rate variation across the genome (Egan et al.,  
40 2007, Harpak et al., 2016). Indeed, it has been suggested that “recurrent, parallel mutation modes  
41 can profoundly shape the paths taken by evolution and undermine common models of evolutionary  
42 genetics” (Press et al., 2019).

Here, we derive an approximation for the fixation probability and the fixation time under the  
43 effect of mutation, selection, and genetic drift. Comparing our approximations to simulation results,  
44 we find they perform very well. We find that when the mutation supply is high enough (roughly  
45 one mutant individual per generation), fixation is certain and rapid. Importantly, it is significantly  
46 faster than implied by approximations that neglect mutation, as such approximations perform poorly,  
47 underestimating the fixation probability and overestimating the fixation time. We also compare our  
48 approximations to those previously derived by Hermisson and Pfaffelhuber (2008) in their analysis  
49 of genetic hitchhiking. We find that our approximation is more flexible: it allows one to assume any  
50 initial frequency for the beneficial allele—which is likely to appear in more than one individual if  
51 the mutation supply is large—and it also allows to approximate the time for the beneficial allele to  
52 reach any upper frequency, rather than 1, which is useful for applying the approximation to results of  
53 evolutionary experiments or simulations, in which researchers may not want to wait until the beneficial  
54 allele is completely fixed in the population.

55 In the following we present a simple Wright-Fisher model with a single, bi-allelic locus under the  
56 effects of selection, recurrent mutation, and drift; we derive diffusion equation approximations for the  
57 fixation probability and time in this model; compare these approximations to simulation results and  
58 previous approximations; and discuss our results.

## 60 Model and Results

### Wright-Fisher model

62 Here, we consider a Wright-Fisher model of a haploid population with non-overlapping generations  
63 and a constant population size  $N$ . We focus on a single bi-allelic locus with alleles  $A$  and  $a$ . The  
64 frequencies of alleles  $A$  and  $a$  are  $x$  and  $1 - x$ , respectively. The fitness of allele  $A$  relative to  $a$  is  $1 + s$ ,  
65 such that  $s > 0$  is the selection coefficient of  $A$ , and the effect of selection is given by

$$66 \quad x^s = \frac{x(1+s)}{x(1+s) + (1-x)}. \quad (1)$$

We assume that mutations from  $a$  to  $A$  occur with rate  $\mu$  and neglect back-mutations from  $A$  to  $a$  (the  
68 case with back mutations was analyzed by Ewens (2004)). Thus, the effect of mutation is given by

$$x^m = x^s + (1 - x^s)\mu. \quad (2)$$

70 Finally, the effect of random genetic drift is given by

$$x' = n/N, \quad n \sim \text{Bin}(N, x^m), \quad (3)$$

72 such that  $x'$  is the frequency of allele  $A$  in the next generation.

### Diffusion equation approximation

74 Let  $a(x) = \mathbf{E}[x' - x]$  and  $b(x) = \mathbf{Var}[x' - x]$  be the expectation and variance of the change in frequency  
of allele  $A$ . We define the scaled selection parameter as  $\alpha = Ns$  and the scaled mutation parameter as  
76  $\theta = 2N\mu$ , which is also called the *mutation supply*. We therefore have (see Ewens, 2004, eq. 5.6)

$$a(x) = \alpha x(1 - x) + \theta(1 - x)/2, \quad (4a)$$

$$78 \quad b(x) = x(1 - x). \quad (4b)$$

Next, we follow Ewens (2004, ch. 4 and 5) to determine the fixation probability and the fixation  
80 time of allele  $A$ . In the following results,  $\Gamma(x)$  and  $\Gamma(x, y, z)$  are the *gamma function* and the  
*generalized incomplete gamma function*, given by the integral equations  $\Gamma(x) = \int_0^\infty t^{x-1} e^{-t} dt$  and  
82  $\Gamma(x, y, z) = \int_y^z t^{x-1} e^{-t} dt$ . When  $x = 1$ , the generalized incomplete gamma function can be simplified,  
 $\Gamma(1, y, z) = e^{-y} - e^{-z}$ .

### 84 Analysis of fixation probability

The probability that a beneficial mutant allele fixates, i.e., takes over the population, is an important  
86 quantity in population genetics and evolutionary theory (reviewed in Patwa and Wahl, 2008, McCan-  
dlish and Stoltzfus, 2014). Early approximations of the fixation probability used branching processes  
88 (Haldane, 1927, Eshel, 1981). Modern and more accurate approximations usually use a diffusion  
process to calculate the probability that the allele frequency  $x$  reaches 1 before it reaches 0. However,  
90 to the best of our knowledge, the literature does not have an approximation for the fixation probability  
under directional mutation (i.e., mutation from  $a$  to  $A$ ) with a high mutation rate. The following result  
92 provides such an approximation, with proof in Appendix A.

94 **Result 1.** *The fixation probability of a beneficial allele initially at a frequency  $p > 0$ ,  $\pi(p)$ , is the  
probability that the allele reaches frequency  $x = 1$  before it reaches frequency  $x = 0$ . So,  $\pi(p)$  is  
96 approximated by one of the following:*

1. *Low mutation supply: If  $0 \leq \theta < 1$ , then*

$$98 \quad \pi(p) = \frac{\Gamma(1 - \theta, 0, 2p\alpha)}{\Gamma(1 - \theta, 0, 2\alpha)}. \quad (5)$$

2. *High mutation supply: If  $\theta \geq 1$ , then  $\pi(p) = 1$ .*

100 **Remark 1.** Note that without mutation,  $\theta = 0$ , the expression simplifies to the classical approximation  
(Kimura, 1962),

$$102 \quad \pi(p) = \frac{1 - e^{-2p\alpha}}{1 - e^{-2\alpha}}. \quad (6)$$

**Remark 2.** The case distinction in Result 1 follows from the fact that the boundary  $x = 0$  is inaccessible  
104 if  $1 \leq \theta$  (see Appendix A.3).

## Analysis of fixation time

106 Next we focus on the fixation time (conditioned on fixation), which has also been previously estimated  
 107 using a diffusion equation approximation, e.g., by Kimura and Ohta (1969) and Ewens (2004, eq. 4.21).  
 108 The following result gives an approximation for our model with unidirectional mutation, with proof  
 in Appendix A.

110

**Result 2.** *The conditional fixation time of a beneficial allele initially at a frequency  $p > 0$ ,  $\bar{t}(p)$ , is  
 112 the waiting time for the allele to reach frequency  $x = 1$  conditioned on reaching  $x = 1$  before reaching  
 $x = 0$ ,*

$$114 \quad \bar{t}(p) = \int_0^1 t(x, p) dx, \quad (7)$$

using one of the following Green's functions,  $t(x, p)$ :

116 1. *Low mutation supply: If  $0 \leq \theta < 1$ , then Green's function is*

$$t(x, p) = \frac{2^\theta e^{2\alpha x} (\alpha x)^{\theta-1}}{(1-x)\Gamma(1-\theta, 0, 2\alpha)} \times \begin{cases} t_0(x, p), & \text{if } 0 \leq x \leq p < 1 \\ t_1(x, p), & \text{if } 0 < p \leq x \leq 1 \end{cases}, \quad (8)$$

118 where

$$t_0(x, p) = \frac{\Gamma(1-\theta, 2p\alpha, 2\alpha) \Gamma(1-\theta, 0, 2x\alpha)^2}{\Gamma(1-\theta, 0, 2\alpha p)}, \quad (9a)$$

$$120 \quad t_1(x, p) = \Gamma(1-\theta, 2x\alpha, 2\alpha) \Gamma(1-\theta, 0, 2x\alpha). \quad (9b)$$

2. *High mutation supply: If  $\theta \geq 1$ , then Green's function is*

$$122 \quad t(x, p) = \frac{2^\theta e^{2\alpha x} (\alpha x)^{\theta-1}}{1-x} \times \begin{cases} \Gamma(1-\theta, 2p\alpha, 2\alpha), & \text{if } 0 \leq x \leq p < 1, \\ \Gamma(1-\theta, 2x\alpha, 2\alpha), & \text{if } 0 < p \leq x \leq 1. \end{cases} \quad (10)$$

**Remark 3.** Note that due to the change of parameters from  $s$  to  $\alpha$  and from  $\mu$  to  $\theta$ , to get the fixation  
 124 time in generations we have to multiply the result by the population size,  $N$ .

**Remark 4.** In general, it is not possible to get analytical expressions for  $\bar{t}(p)$  (eq. (7)). However,  
 126 one can numerically integrate these expressions, which serves many practical applications. However,  
 this requires the integration of gamma functions, and should be performed with care. For example,  
 128 numerical integration with *Wolfram Mathematica* was successful, but using Python with *scipy* failed  
 for some parameter values.

130 **Remark 5.** Without mutation,  $\theta = 0$ , the fixation time  $\bar{t}(p)$  has been approximated previously by  
 Kimura and Ohta (1969) (see also Ewens, 2004, Durrett, 2008). We present this result and a closed-  
 132 form solution that uses exponential integral functions in Appendix B.

## Approximation for strong selection

134 Hermisson and Pfaffelhuber (2008, abbreviated *HP* henceforth) have approximated the conditional  
 fixation time in our model with recurrent mutation, assuming strong selection (large  $\alpha$ ).

136 In the following,  $H_x = \sum_{n=1}^{\infty} \frac{x}{n(n+x)}$ , which is an analytic continuation of the harmonic numbers for  
 real-values  $x$ :

$$138 \quad H_x = \sum_{k=1}^x \frac{1}{k} = 1 + \frac{1}{2} + \dots + \frac{1}{x} + \left( \frac{1}{1+x} + \dots \right) - \left( \frac{1}{1+x} + \dots \right) = \sum_{n=1}^{\infty} \frac{1}{n} - \frac{1}{n+x} = \sum_{n=1}^{\infty} \frac{x}{n(n+x)}.$$

We now repeat their result using our notation.

140 **Result 3.** Suppose the beneficial allele is initially absent ( $p = 0$ ). Then the fixation time  $\bar{t}(0)$  can be approximated by one of the following.

142 1. Low mutation supply: If  $0 < \theta < 1$ , then

$$\bar{t}(0) = \frac{2}{\alpha} (\log(2\alpha) + \gamma_e) + \mathcal{O}\left(\frac{\log(\alpha)}{\alpha^2}\right). \quad (11)$$

144 2. High mutation supply: If  $\theta \geq 1$ , then

$$\bar{t}(0) = \frac{2}{\alpha} (\log(2\alpha) + \gamma_e) - \frac{H_{\theta-1}}{\alpha} + \mathcal{O}\left(\frac{\log(\alpha)}{\alpha^2}\right) + \frac{1}{\theta} \mathcal{O}(\alpha e^{-\alpha}). \quad (12)$$

146 Intrigued by the Figure 3b, we assumed that the reduction in fixation time is independent of  $\mu$  if we assume fixation to be at a value  $z < 1$ . By informed guessing and testing various parameter combinations against eq. (7) with the appropriate Green's function, we conclude that the difference is roughly

150 
$$t^- = \frac{\log(2(1-z)\alpha) + (2-z)\gamma_e}{\alpha}. \quad (13)$$

152 Thus, to quickly calculate the time until the population reaches a fixation cut-off  $z$ , one can use the formula

$$\bar{t}(0) - t^- = \frac{\log\left(\frac{2\alpha}{1-z}\right) + z\gamma_e + \mathbb{1}_{\theta \in (1, \infty)} H_{\theta-1}}{\alpha}, \quad (14)$$

154 where  $\mathbb{1}_{x \in A}$  denotes the indicator function that is one if  $x$  is an element of the set  $A$  and zero otherwise. The insets of Figure 3 show the very small relative error of eq. (14) compared to eq. (7).

156 Unfortunately, we did not find an adjustment for an initial number of mutants  $n_0 > 1$ .

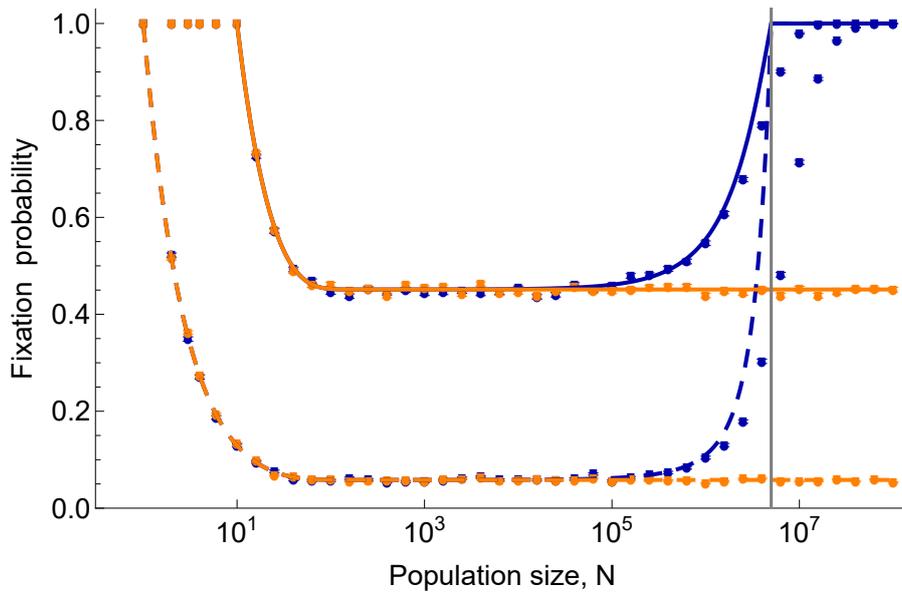
## Numerical results: Fixation probability

158 We quantitatively compare our approximation (eq. (5),  $\theta \geq 0$ ), the classical approximation (eq. (6),  $\theta = 0$ ), and simulation results (eqs. (1) to (3)), in Figure 1. For small population sizes, the fixation probability goes to 1, because the initial number of copies of the beneficial allele is not far from the population size. For intermediate population sizes, roughly,  $100 < N < 100,000$  the fixation probability is roughly  $2s \cdot pN$  (Haldane, 1927, eq. 1.0). Importantly, for larger population sizes in which  $\theta \geq 1$ , our approximation (blue lines) increases to 1 as the mutational supply ( $\theta = 2N\mu$ ) increases, whereas Kimura's approximation (orange lines) remains constant, underestimating the fixation probability (compare with markers).

## 166 Numerical results: Fixation time

Again, we provide a quantitative comparison of the above approximations and simulation results (eqs. (1) to (3)) in Figure 2. Several observations are worth mentioning. First, the classical approximation (eq. (B1)) is very good when the population size is small (i.e., mutation load is low,  $\theta \leq 1$ , left of the vertical line). Second, with a large population size (i.e., high mutation load,  $\theta \geq 1$ , right of the line), the classical approximation overestimates the fixation time. Third, both our approximation (Result 2) and the *HP* approximation (Result 3) perform well, even when the classical approximation fails. Fourth, although the *HP* approximation assumes strong selection, it performs well even for very weak selection ( $s = 3 \cdot 10^{-6}$ ), although not as well as our approximation (compare blue and orange lines in Figure 2b).

176 However, the *HP* approximation overestimates the fixation time when two implicit assumptions are not met. First, it approximates the time for the beneficial allele to reach 100% frequency. However,



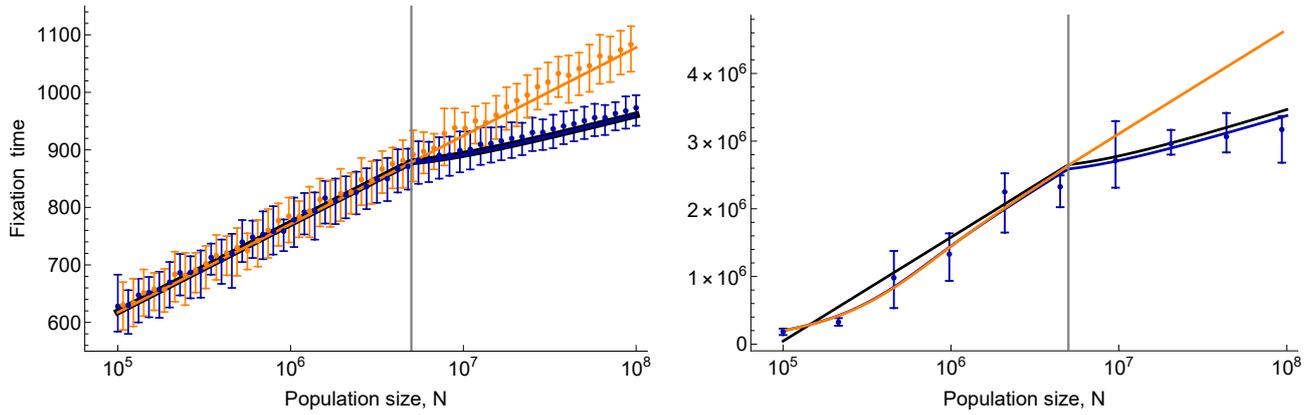
**Figure 1: Fixation probability for increasing population size.** Markers for results of simulations of the Wright-Fisher model (eqs. (1) to (3); error bars denote 50% CI.), lines for diffusion equation approximations: blue and orange for model with mutation ( $\mu = 1 \cdot 10^{-7}$ ; Result 1) and without mutation ( $\mu = 0$ ; eq. (6)), respectively. Lines and markers at the bottom are for fixation from a single copy of the beneficial allele ( $p = 1/N$ ); lines and markers at the top are for fixation from 10 copies of the beneficial allele ( $p = 10/N$ ). Gray vertical line indicates the population size  $N = \mu/2$  at which  $\theta = 1$ . Here, the selection coefficient is  $s = 0.03$ .

178 waiting for complete fixation of the beneficial allele may take too long in many applications, including  
 180 both simulations and experiments. In such cases, our approximation can still be used by simply  
 182 changing the upper bound of the integral in eq. (7) from 1 to the required frequency  $z$ , such that we  
 compute  $\bar{t}_z(p) = \int_0^z t(x, p) dx$ . Comparing this and the *HP* approximation to results of simulations  
 184 in which we wait for the beneficial allele to reach 95% frequency, rather than 100%, we find that our  
 approximation performs well whereas the *HP* approximation overestimates so much that it is actually  
 186 better to use the classical approximation (eq. (B1)), again by changing the upper bound of the second  
 integral from 1 to  $z$  (Figure 3).

186 In addition, the *HP* approximation also assumes that the beneficial allele is initially absent, that is,  
 $p = 0$ . However, this may be unrealistic with a high mutation supply, which is expected to generate  
 188 high standing variation. Consider a population evolving under stress, such as heat or drug treatment,  
 until the fixation of a beneficial allele relieve the stress. Assuming this allele was neutral or deleterious  
 190 before the stress, its frequency would have been roughly  $\mu$  or  $\mu/s$ , respectively. Under strong mutation,  
 both values could be much larger than zero, meaning that fixation proceeds from an initial frequency  
 192  $x \gg 0$  to  $x = 1$ . Our approximation (Result 2) explicitly takes the positive initial frequency of the  
 beneficial allele into consideration as a parameter  $p$ , and indeed, it performs well when the initial  
 194 frequency is greater than zero (Figure 4). However, the fixation time decreases as the initial frequency  
 increases, and thus the *HP* approximation overestimates the fixation time in these cases, whereas the  
 196 classical approximation performs well, as long as the mutation supply is low ( $\theta \leq 1$ ), as it can also  
 take the initial frequency as parameter (eq. (B1)).

## 198 Discussion

Recurrent mutation is defined here as the repeated production of a mutant genotype such that multiple  
 200 independently produced mutants are co-segregating in the population. This can occur when the

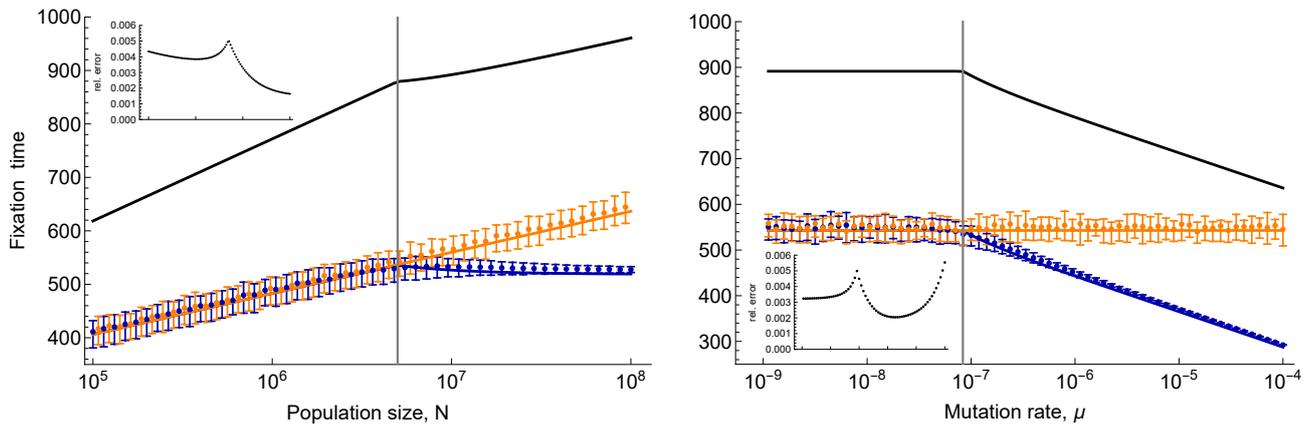


**Figure 2: Fixation time for increasing population size.** Time for the beneficial allele to go from one copy,  $x = 1/N$ , to full fixation,  $x = 1$ , conditioned on fixation (i.e.,  $x \neq 0$ ). Markers for results of simulations of the Wright-Fisher model (eqs. (1) to (3); error bars denote 50% CI), lines for diffusion equation approximations: blue and orange for model with ( $\mu = 1 \cdot 10^{-7}$ ; Result 2) and without mutation ( $\mu = 0$ ; eq. (B1)), respectively. Black line (sometimes overlapping the blue line) is the *HP* approximation (eqs. (11) and (12)). In both panels the fixation time with and without mutation diverges at  $\theta = 1$ , with the former being lower. **(a)** Strong selection,  $s = 0.03$ . **(b)** Weak selection,  $s = 0.000003$ . *HP* approximation fits well, only overestimating by a bit for majority of population sizes.

202 mutation supply is high due to large population sizes and/or high mutation rates. Note that here the  
 203 term “mutation” includes not only single-nucleotide substitutions, but any genetic modification, such  
 204 as focal or segmental duplications, aneuploidy, transposition, or short tandem repeats, all of which  
 205 may have high mutation rates compared to point mutations (Press et al., 2019). Recurrent mutation  
 206 has been somewhat neglected in the evolutionary theory literature due to the common assumption  
 207 that mutations are rare, although previous studies have investigated its effect on the site-frequency  
 208 spectrum (Harpak et al., 2016, Jenkins and Song, 2011, Wakeley et al., 2023), soft vs. hard selective  
 209 sweeps (Pennings and Hermisson, 2006), genetic hitchhiking (Hermisson and Pfaffelhuber, 2008),  
 210 and estimation of recombination rates (McVean et al., 2002).

210 Here we have developed a diffusion equation approximation for the fixation probability and time  
 211 of a beneficial mutant allele under the effects of selection, drift, and importantly, recurrent mutation  
 212 ( $\theta = 2N\mu > 1$ ). We find that if recurrent mutations occur, then our approximation (Results 1 and 2)  
 213 for the fixation probability and time is superior to the classic approximations (Kimura, 1962, Kimura  
 214 and Ohta, 1969), which can significantly overestimate both quantities. Our approximation is flexible,  
 215 as it considers both the evolutionary parameters (selection coefficient  $s$ , mutation rate  $\mu$ , population  
 216 size  $N$ ), and the boundary conditions: the initial and final mutant frequencies ( $p$  and  $z$ , respectively).  
 217 Nevertheless, if these boundary conditions are chosen such that the mutant increases in frequency from  
 218 absence ( $p = 0$ ) to full fixation ( $z = 1$ ), then the previously developed *HP* approximations (eqs. (11)  
 219 and (12)) are sufficient. Otherwise, our approximations are more accurate and flexible.

220 The diffusion equation is often used in evolutionary theory to describe evolutionary dynamics, that  
 221 is, to describe the change in allele frequency over time in a population due to the effects of natural  
 222 selection, mutation, and random genetic drift. Diffusion equation approximations have been derived  
 223 for the probability and waiting time for an allele to fix in a population since the classical studies by  
 224 Kimura (Kimura, 1962, Kimura and Ohta, 1969) under various models (Ewens, 2004, Durrett, 2008).  
 225 However, the literature is missing an explicit analysis of the diffusion equation approximation in the  
 226 case of a beneficial allele under the effect of strong mutation, in which the mutant allele continues to be  
 227 generated even after the first copy has appeared; existing analyses assume selection without mutation  
 228 (Kimura, 1962, Kimura and Ohta, 1969), uni-directional mutation without selection, and bi-directional

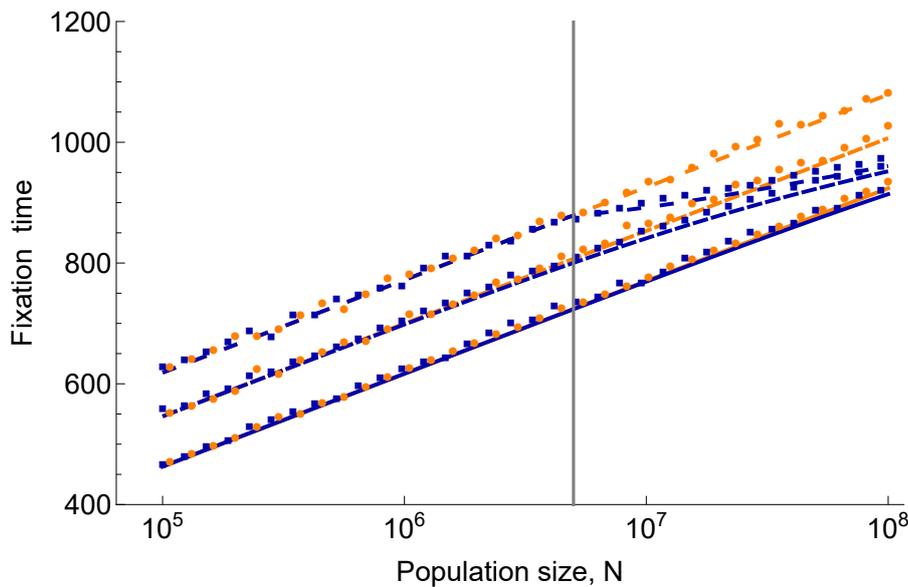


**Figure 3: Time to reach 95% for increasing population size and mutation rate.** Time for the beneficial allele to go from one copy,  $x = 1/N$ , to 95%, or  $x = 0.95$ , rather than full fixation. Markers for results of simulations of the Wright-Fisher model (eqs. (1) to (3)); error bars denote 50% CI), lines for diffusion equation approximations. Blue and orange for model with ( $\mu = 1 \cdot 10^{-7}$ ; Result 2) and without mutation ( $\mu = 0$ ; eq. (B1)), respectively, integrated numerically from 0 to 0.95 rather than to 1. Black line (sometime overlapping the blue line) is the  $HP$  approximation (eqs. (11) and (12)), which assumes full fixation (100%) rather than high frequency (95%), and therefore overestimates the fixation time. Here, the selection coefficient is  $s = 0.03$ ; (a) Mutation rate is  $\mu = 1 \cdot 10^{-7}$ ; (b) Population size is  $N = 6 \cdot 10^6$ . The insets show the relative error of the adjusted  $HP$  approximation eq. (14) compared to the blue line.

230 weak mutation and selection (Ewens, 2004). Thus, our results close a gap in the theoretical literature on the fixation of beneficial alleles.

232 When the frequency of the mutant is low, a high mutation rate may be high enough to be a major factor in its evolution (i.e. its increase in frequency). When the mutant reaches a high frequency, its further evolution will be driven by natural selection. However, in some scenarios, the high mutation rate implies that other mutants are also likely to appear, leading to rapid adaptation, clonal interference, and soft sweeps. In these cases, the focal mutant is unlikely to reach fixation before the appearance of additional mutations, either on other backgrounds, leading to clonal interference, or on the background of the focal mutant, leading to further adaptation. Thus, the assumptions underlying our approximations may not be met. Still, these approximations are superior to the classic approximations, and can be used to estimate if the focal mutant has enough time to fix or reach some target establishment frequency before other beneficial mutants appear.

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236  
238  
240



**Figure 4: Fixation time with various initial frequencies.** From top to bottom are shown fixation times for initial frequencies of the beneficial allele: top,  $p = 1/N$ ; middle,  $p = 100/N$ ; bottom,  $p = 1,000/N$ . Markers for results of simulations of the Wright-Fisher model (eqs. (1) to (3); error bars denote 50% CI), lines for diffusion equation approximations. Blue and orange for model with ( $\mu = 10^{-7}$ ; Result 2) and without mutation ( $\mu = 0$ ; eq. (B1)), respectively. Importantly, the fixation time decreases as the initial frequency increases. The approximations fit the simulations well, and the difference between models with and without mutation shrink as the initial frequency increases. Error bars are small and similar to those in Figure 2, and not shown to avoid over-plotting. Here, the selection coefficient is  $s = 0.03$ .

## Appendices

### 242 Appendix A Proofs of Result 1 and Result 2

#### A.1 Boundary types

244 There are two important properties that characterize the boundary: (i) a boundary is *accessible* if  
 245 there is a positive probability that it can be reached in finite time from a given interior point, and (ii)  
 246 is *absorbing* if the process remains forever at the boundary after reaching the boundary.

Here, we determine the boundary type of  $x = 0$  and  $x = 1$ . This is nicely demonstrated by Durrett  
 248 (2008), which not only differs in notation from Ewens (2004), but also provides helpful examples.  
 We focus on a specific example (Durrett, 2008, example 7.33, pg. 295) that studies a Wright-Fisher  
 250 diffusion with mutations in both directions, but without selection. It is explained that adding selection  
 does not change the (in)finiteness of the integrals. If  $0 \leq \theta < 1$ , then  $x = 0$  is accessible but not  
 252 absorbing, whereas it is not accessible for  $\theta \geq 1$ . In our model there is no mutation in the other  
 direction, from  $A$  to  $a$ , so the boundary  $x = 1$  is absorbing.

254 Therefore, for  $0 \leq \theta < 1$  there is a positive probability  $1 - \pi(p) > 0$  that population initially with  
 frequency  $p$  of allele  $A$  reaches  $x = 0$  before it reaches  $x = 1$ . However, mutation will eventually  
 256 introduce the extinct  $A$  allele and thus this state,  $x = 0$ , is not absorbing. Therefore, to continue with  
 the derivation of the fixation times, we have to consider the two cases:  $0 \leq \theta < 1$  and  $1 \leq \theta$ .

## 258 **A.2 Boundary $x = 0$ is accessible, $0 \leq \theta < 1$**

An important quantity that we will use in the following analyses is  $\psi(x)$ . It is given in eq. 4.16 in  
 260 (Ewens, 2004). Substituting  $a(x)$  and  $b(x)$  from eq. (4) we have

$$\psi(x) = \exp\left(-2 \int_x^1 \frac{a(y)}{b(y)} dy\right) = e^{-2\alpha x} x^{-\theta}, \quad (\text{A1})$$

262 Note that  $\psi$  is different from  $\Psi$ , which denotes the digamma function.

If  $\theta < 1$ , we use eqs. 4.22 and 4.23 of Ewens (2004) to calculate Green's function,

$$264 \quad t(x, p) = \frac{2}{b(x)\psi(x)} \times \begin{cases} (1 - \pi(p)) \int_0^x \psi(y) dy, & \text{if } 0 \leq x \leq p < 1, \\ \pi(p) \int_x^1 \psi(y) dy, & \text{if } 0 < p \leq x \leq 1, \end{cases} \quad (\text{A2})$$

where  $\pi(p)$  is the fixation probability. Substituting  $\pi(p)$  from eq. (5) into eq. (A2) gives the following  
 266 Green's function,

$$t^*(x, p) = \frac{2^\theta e^{2x\alpha} (\alpha x)^{\theta-1}}{(1-x) \Gamma(1-\theta, 0, 2\alpha)} \times \begin{cases} t_0(x, p), & \text{if } 0 \leq x \leq p < 1, \\ t_1(x, p), & \text{if } 0 < p \leq x \leq 1, \end{cases} \quad (\text{A3})$$

268 where

$$t_0(x, p) = \Gamma(1-\theta, 2p\alpha, 2\alpha) \Gamma(1-\theta, 0, 2x\alpha) \quad (\text{A4a})$$

$$270 \quad t_1(x, p) = \Gamma(1-\theta, 2x\alpha, 2\alpha) \Gamma(1-\theta, 0, 2p\alpha). \quad (\text{A4b})$$

All derivations are done with the help of the computer algebra software *Wolfram Mathematica*.

272 So far the fixation time was calculated for a population that randomly reaches one of the absorbing  
 states, either  $x = 0$  or  $x = 1$ . In this study we want to focus on the time until the population hits and is  
 274 absorbed in  $x = 1$ , that is, we are interested in the conditional fixation time, conditioned on reaching  
 $x = 1$  before  $x = 0$ .

276 In equation 4.48 in Ewens (2004, pg. 146) we have the formula to compute the sojourn times  
 conditioned that the process hits  $x = 1$  first, which gives the following Green's function,

$$278 \quad t(x, p) = t^*(x, p) \frac{\pi(x)}{\pi(p)} = t^*(x, p) \frac{\Gamma(1-\theta, 0, 2x\alpha)}{\Gamma(1-\theta, 0, 2p\alpha)}, \quad (\text{A5})$$

by substituting eq. (5) for  $\pi(p)$ . Substituting eq. (A3) into eq. (A5) gives eq. (8).

## 280 **A.3 Boundary $x = 1$ is not accessible, $1 \leq \theta$**

If  $\theta \leq 1$ , only  $x = 1$  is an accessible boundary state and we have to use eqs. 4.40 and 4.41 from Ewens  
 282 (2004),

$$t(x, p) = \frac{2}{b(x)\psi(x)} \times \begin{cases} \int_p^1 \psi(y) dy, & \text{if } 0 \leq x \leq p < 1, \\ \int_x^1 \psi(y) dy, & \text{if } 0 < p \leq x \leq 1. \end{cases} \quad (\text{A6})$$

284 Inserting eq. (4) and eq. (A1) into eq. (A6) and solving with *Wolfram Mathematica*, we get eq. (10).

## **A.4 Green's function $t(x, p)$ at $\theta = 1$**

286 We showed above that there have to be two different derivations for Green's functions, depending on  
 $\theta$ . However, at  $\theta = 1$ , the expressions coincide; the limit from below is a bit more complex, but can  
 288 be calculated with *Wolfram Mathematica*. We get

$$\lim_{\theta \uparrow 1} t(x, p) = \lim_{\theta \downarrow 1} t(x, p) = \frac{2e^{2x\alpha}}{1-x} \times \begin{cases} \Gamma(0, 2p\alpha, 2\alpha), & \text{if } 0 \leq x \leq p < 1, \\ \Gamma(0, 2x\alpha, 2\alpha), & \text{if } 0 < p \leq x \leq 1. \end{cases} \quad (\text{A7})$$

## 290 Appendix B Approximation of fixation time without mutation

We set  $\theta = 0$  in eqs. (A5), (8) and (9) and use *Wolfram Mathematica* to find a representation of Green's  
 292 function,  $t(x, p)$ , with hyperbolic functions, which is shorter than a representation with exponentials  
 (e.g. Kimura and Ohta (1969, eq. 17)). Thus, the expected conditional fixation time without mutation  
 294 is approximated by

$$\bar{t}(p) = \int_0^p I_1 dx + \int_p^1 I_2 dx, \quad (\text{B1})$$

296 where

$$I_1 = \frac{2 (\coth(p\alpha) - \coth(\alpha)) \sinh^2(x\alpha)}{(1-x)x\alpha},$$

$$I_2 = \frac{2\text{csch}(\alpha) \sinh(x\alpha) \sinh((1-x)\alpha)}{(1-x)x\alpha}.$$

*Wolfram Mathematica* further gives an explicit result for eq. (B1) in terms of hyperbolic integrals,  
 300 but we prefer to show a non-hyperbolic version,

$$\int_0^p I_1 dx = \int_0^p \frac{\exp(-2x\alpha) (\exp(2p\alpha) - \exp(2\alpha)) (1 - \exp(2x\alpha))^2}{(1 - \exp(2\alpha)) (1 - \exp(2p\alpha)) (1-x)x\alpha} dx$$

$$= \frac{\exp(2p\alpha) - \exp(2\alpha)}{(1 - \exp(2\alpha))(1 - \exp(2p\alpha))\alpha} (E_1 + L_1),$$

$$\int_p^z I_2 dx = \int_p^z \frac{\exp(-2x\alpha) (\exp(2x\alpha) - \exp(2\alpha)) (-1 + \exp(2x\alpha))}{(1 - \exp(2\alpha)) (1-x)x\alpha} dx$$

$$= \frac{E_2^z + L_2^z}{(1 - \exp(2\alpha))\alpha}$$

where

$$E_1 = \exp(-2\alpha)\Gamma(0, -2\alpha, -2(1-p)\alpha) + \exp(2\alpha)\Gamma(0, 2\alpha, 2(1-p)\alpha) - (Ei(2p\alpha) + Ei(-2p\alpha))$$

$$L_1 = 2(\log\left(\frac{2p\alpha}{1-p}\right) + \gamma_e)$$

$$E_2^z = \exp(2\alpha)\left(\Gamma(0, 2p\alpha, 2(1-p)\alpha) - \Gamma(0, 2z\alpha, 2(1-z)\alpha)\right) + \Gamma(0, -2\alpha, -2(1-p)\alpha) - \Gamma(0, -2z\alpha, -2(1-z)\alpha)$$

$$L_2^z = (1 + \exp(2\alpha)) \log\left(\frac{p(1-z)}{z(1-p)}\right)$$

$$E_2 = \exp(2\alpha)\left(\Gamma(0, 2p\alpha, 2(1-p)\alpha) + Ei(-2\alpha)\right) + \Gamma(0, -2p\alpha, -2(1-p)\alpha) + Ei(2\alpha)$$

$$L_2 = (1 + \exp(2\alpha))\left(\log\left(\frac{p}{2\alpha(1-p)}\right) - \gamma_e\right)$$

314 Here,  $E_2^z$  and  $L_2^z$  represent the expressions if the upper bound is  $z < 1$ . Since just setting  $z = 1$  does  
 not work, one needs to take the limit  $z \rightarrow 1$  in  $E_2^z + L_2^z$ . This results in  $E_2 + L_2$ .

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