1 Original article:

2 Forest management drives evolution of understorey herbs

3 Authors:

- 4 Charlotte Møller^{1*}, Pieter De Frenne², Martí March-Salas¹, Thomas Vanneste², Kris
- 5 Verheyen², & J. F. Scheepens¹.

6 Affiliations:

- ⁷ ¹Plant Evolutionary Ecology, Faculty of Biological Sciences, Goethe University Frankfurt,
- 8 Max-von-Laue-Str. 13, 60438 Frankfurt am Main, Germany.
- 9 ²Forest & Nature Lab, Faculty of Bioscience Engineering, Ghent University,
 10 Geraardsbergsesteenweg 267, 9090 Gontrode, Belgium.
- 11 * **Corresponding author:** Plant Evolutionary Ecology, Faculty of Biological Sciences, Goethe
- 12 University Frankfurt, Frankfurt am Main, Germany.
- 13 Email: <u>moeller@bio.uni-frankfurt.de</u>

14 **ORCID:**

- 15 CM: 0000-0002-6563-1490
- 16 PDF: 0000-0002-8613-0943
- 17 MMS: 0000-0001-5347-4056
- 18 TV: 0000-0001-5296-917X
- 19 KV: 0000-0002-2067-9108
- 20 JFS: 0000-0003-1650-2008

Abstract

Forest management has a strong impact on the forest structure and subsequently on the
 biotic and abiotic forest understorey environment. Forest understorey herbs can thus be
 expected to evolutionary respond to management-induced environmental variation
 (provided sufficient time for adaptation), but this has been little tested to date.

27 2. Here we use a common garden, to test for genetically based variation in phenotypic traits in populations of forest herbs sampled along a forest management intensity 28 gradient. Five different herbaceous species were sampled from 70-100 populations in 29 three regions in Germany and were tested for genetically based variation in flowering 30 start, proportion of flowering ramets, and plant height. Additionally, we investigated 31 the effects of management-induced environmental variation and performed structural 32 equation modelling to study how forest management drives trait differentiation via its 33 effects on the microenvironment. 34

35 3. We found that the studied forest understorey herbs varied genetically in the measured 36 functional and phenological traits among the sampled populations. Forest management 37 likely affected the traits in various directions and strengths depending on the species, 38 either directly through variation in forest structural attributes or indirectly through 39 changes in the microclimatic environment on the forest floor.

40 4. *Synthesis:* We show that forest management can have evolutionary consequences for
41 forest understorey plants. In an applied context, diverse forest management actions
42 within landscapes thus creates heterogeneity that selects for different plant traits and
43 thus helps conserving genetic diversity.

Keywords: common garden, evolutionary ecology, forest structure, genetic differentiation,
intraspecific trait variation, microclimate, structural equation modelling.

Introduction

In our rapidly changing world, land use has been recognized as having the strongest impact on 47 species and genetic diversity (IPCC, 2014; Tobias et al., 2021). The present state of temperate 48 forests in Europe is a product of decisions made by former and current management, rather 49 than natural factors and progression (Spiecker, 2003). Current silvicultural management 50 51 practices have a profound impact on forest ecosystems, as they can affect forest structure, for instance by altering crown projection area, structural complexity, and tree species composition. 52 Large, dense tree crowns cast shade, buffer against warming and reduce daytime summer 53 temperatures inside forests (De Frenne et al., 2013). Furthermore, light availability is highly 54 dependent on the composition of tree species, which in turn depends on site-specific climatic 55 factors such as water availability (Aussenac, 2000). Existing silvicultural management and 56 conservation actions supporting coppicing, selective tree extractions or other actions can lead 57 to canopy openings, potentially causing nutrient depletion of the soil, a decreasing biomass 58 59 pool, increasing nitrogen release in the soil, but also warming of the forest floor, likely causing changes in the composition of understorey herb species (De Frenne et al., 2013). 60

61 Forest understorey herbs play a key role in forest ecosystems (Augusto et al., 2003; Gilliam, 2007; Landuyt et al., 2019), harbouring the highest species diversity among all strata 62 in temperate forests (Gilliam, 2007). In temperate forests, the phenology of early-growing 63 64 understorey herbs should be attuned to abiotic cues, such as spring temperature, in order to prevent early frost or to finish their reproductive cycle before tree leaf out (Heberling et al., 65 2019). High management intensity for example can result in delayed flowering in understorey 66 67 herbs (Willems et al., 2021). More specifically, low spring temperatures were associated with intensely managed conifer plantations, with understorey herbs flowering on average two weeks 68 69 later than in unmanaged forests. However, this study is based on field observations only, and thus cannot infer whether the results are due to phenotypic plasticity or genetic differentiation. 70

71 Forest management, its influence on the forest floor environment, as well as how it impacts adaptation of understorey herbs, are still underexplored. Soil pH is highly influential 72 on plant phenology and growth, and affects traits such as germination (Gentili et al., 2018), 73 74 plant height and number of flowers (Jiang et al., 2017). Some aspects of acidic soils can be advantageous for plant growth, as more micronutrients are available to plants compared to 75 neutral-alkaline soils (Loncaric et al., 2008). Studies have found that plants germinate and 76 77 flower earlier on acidic soils, but species differences in the number of flowers (Gentili et al., 2018; Sercu et al., 2021). In general, functional trait responses to forest structural variation and 78 79 microenvironment depend on the phenology and distribution of the forest understorey herb, and can therefore be highly species-specific (De Frenne et al., 2009; Blondeel et al., 2020; 80 Willems *et al.*, 2021). 81

82 One response of forest understorey plants to silvicultural management actions and microenvironmental changes, is through the employment of genetically based phenotypic 83 84 variation to respond appropriately to stress and disturbances (Lemke et al., 2015). Mutations, genetic drift and gene flow are neutral processes affecting genetically based phenotypic 85 variation and population genetic structure, and genetic variation within populations is a 86 87 necessary prerequisite for adaption to and persistence under changing conditions (Lemke et al., 2015). As sessile organisms, plants are vulnerable to strong environmental changes, and 88 89 especially forest understorey herbs with limited dispersal capabilities may likely be adapted to 90 very specific and local environments. Because forest offer relatively temporally stable environments, we could expect understorey herbs to adapt evolutionary to spatial 91 environmental variation rather than to evolve phenotypic plasticity (Alpert and Simms, 2002; 92 Valladares et al., 2007). However, few studies have looked at genetic adaptation of forest 93 understorey herbs to their local environment (De Frenne et al., 2011; Møller et al., 2022). A 94 useful strategy to investigate evolutionary adaptation is by applying a common garden 95

96 experiment, because it can reveal genetic differentiation among sampled populations, followed
97 by correlative approaches to test for associations between trait variation and environmental
98 variation at sites of population origins (Turesson, 1922; Scheepens and Stöcklin, 2013).

Here we established a common garden using five understorey herbs sampled across a forest management intensity gradient, to (1) investigate whether populations of forest understorey herbs differ genetically in phenotypic variation; (2) If so, how does amongpopulation phenotypic trait differentiation correlate with forest structural variables and microenvironmental variables? And (3) does the forest structure have a direct effect on genetically based phenotypic variation in forest understorey herbs, or indirect effects through the microenvironment?

106

107

Material & methods

108 Study species

For this study, we chose five perennial forest understorey herbs, four forbs – Anemone *nemorosa, Galium odoratum, Oxalis acetosella* and *Viola reichenbachiana* – and one grass – *Milium effusum* (Fig. 1).

Anemone nemorosa (Ranunculaceae) and O. acetosella (Oxalidaceae) are both early-112 flowering, small forbs, reaching heights of 5-25 cm. They rely heavily on clonal spread 113 (Frederiksen and Rasmussen, 2006). By contrast, G. odoratum (Rubiaceae) and V. 114 reichenbachiana (Violaceae) flower later, grow taller, reach heights of 10-30 cm, and rely more 115 on seed dispersal (Frederiksen and Rasmussen, 2006). Lastly, M. effusum (Poaceae) is a tall-116 growing grass, reaching heights of 50-150 cm, and relies heavily on wind-dispersed seeds (De 117 Frenne et al., 2011). All species can spread vegetatively to some degree and vary in 118 colonization rates (Brunet et al., 2012). 119

120 Experimental set-up

In 2020, plants were sampled from plots set up within the framework of the Biodiversity
Exploratories (<u>www.biodiversity-exploratories.de</u>), a large-scale platform for ecological
research (Fischer *et al.*, 2010). The Biodiversity Exploratories contains three regions across
Germany: Schwäbische Alb, Hainich-Dün, and Schorfheide-Chorin (Fischer *et al.*, 2010). Each
region consists of 50 forest plots, *i.e.*, 150 plots across the three regions.

In May 2020, all forest plots were visited. Six genets were sampled for each species (hereafter 126 127 referred to as individuals), whenever they were occurring on a plot. To avoid sampling genetically identical plants, each individual was sampled with a minimum distance of 10 m to 128 the next sampled plant. Individuals were planted in multitrays (51.5 cm width, 33.5 cm length, 129 130 5.5 cm deep, 54 pots per tray) filled with potting soil ("CL T torffrei", Einheitserde, Sinntal-Altengronau, Germany) for root establishment and growth. In November 2020 all individuals 131 were transferred into 1.5 L pots with potting soil ("Typ T, Struktur 1B", Hawita, Vechta, 132 Germany). In spring 2021, all pots were relocated to a shading house (Götsch & Fälschle, 133 Alerheim, Germany), under common-garden conditions, situated at the Goethe University 134 Frankfurt (Frankfurt am Main, Germany). All individuals received water ad libitum during the 135 whole experiment. 136

137 *Plant measurements*

In total, we sampled 2474 individuals: 606 *Anemone nemorosa*, 545 *Galium odoratum*, 467 *Milium effusum*, 430 *Oxalis acetosella*, and 426 *Viola reichenbachiana*, from 70-100 populations. However, mortality caused heavy species-dependent losses after transplantation, mainly in February 2021 due to a period of deep frost following a warm spell. One year later, in the growing period of 2022, we tracked flowering phenology, counted ramets and measured vegetative traits on the remaining 1,129 individuals: 126 *A. nemorosa*, 166 *G. odoratum*, 178 *M. effusum*, 361 *O. acetosella* and 298 *V. reichenbachiana* (See Table S1 for distribution
between populations of sampled individuals).

Phenological measurements were taken three times per week (every Monday, 146 Wednesday, and Friday). Since Anemone nemorosa is a vernal geophyte, emergence date was 147 recorded only for this study species, while flowering start and flowering end were recorded for 148 149 all study species. Dates were transformed into Julian days (day of the year; DOY). Flowering peak was calculated using one-hot encoding (Lantz, 2019), transforming the data to be 150 categorical in order to count how many individuals, from each species, were flowering on a 151 given day. To calculate the proportion of flowering ramets, the number of flowering ramets 152 and total number of ramets were counted for A. nemorosa, G. odoratum and M. effusum. To 153 infer the total number of ramets for O. acetosella, we applied the formula for the frustrum of 154 cone volume measurement: 155

156
$$\frac{1}{3}\pi * h(R^2 + Rr + r^2)$$
 (1)

with h being the height of the individual, R being the radius of the lower base, and r being the 157 radius of the upper base. We consider that each leaf connects to a rhizome and is therefore 158 potentially independent; all leaves were counted for a subset of 12 pots, based on a subjective 159 stratified sampling of small, intermediate, and large plants, and a regression between the 160 frustum volume (Eq. (1)) and the number of leaves ($R^2 = 0.93$, see Fig. S1) was used to predict 161 the number of leaves in the remaining pots. The proportion of flowering ramets was then 162 calculated as the total number of flowers divided by the predicted number of leaves. For V. 163 reichenbachiana, major stems were used as a proxy for ramets, and the number of major stems 164 and number of major flowering stems were counted and used to calculate the proportion of 165 flowering stems, hereafter referred to as proportion of flowering ramets. Plant height was 166

measured for *A. nemorosa*, *G. odoratum*, *M. effusum* and *O. acetosella*, but not for *V. reichenbachiana* as height is not a meaningful variable in this species.

169 Forest structural attributes and microclimatic variables for each population of origin were extracted from the BExIS2 (www.bexis.uni-jena.de) database, maintained by the 170 Biodiversity Exploratories (Fischer et al., 2010), for the years 2018 and 2019, i.e., previous to 171 172 the plants being sampled, to catch relatively recent environmental variation leading up to the sampling in 2020. Structural complexity index (SCI), an index quantifying stand structural 173 complexity based on the fractal dimension of cross-sectional polygons (Ehbrecht et al., 2017), 174 crown projection area (m^2/ha) of mature trees, and the percentage of conifers based on crown 175 projection, was obtained for all sampled plots. These forest attributes were selected because 176 they characterize stand structure and are expected to have an influence on the biotic and abiotic 177 microclimatic conditions, as well as on light availability (Willems et al., 2021). In addition, we 178 also obtained soil pH and the annual mean spring temperature 10 cm above the forest floor 179 (calculated as the average of March, April, and May of 2018 and 2019) as microenvironmental 180 variables. 181

182 Data analyses

All statistical analyses were conducted with R version 4.1.2 (R Core Team, 2021). First, to test for population differences in genetically based phenotypic variation, we ran linear mixed-effect models (LMM) for all traits, using the function "lmer" from the package "lme4" (Bates *et al.*, 2007). The measured plant traits were used as the response variables, population, species, and their interaction as the explanatory variable, and region was included as a random factor.

188 Second, to investigate the effect of forest attributes and microclimate from the 189 population of origin on the measured traits in the common garden, we ran LMMs with 190 structural complexity, crown projection area, percentage of conifers, soil pH, spring temperature, and the two-way interaction between species and all the aforementioned as
explanatory variables. Region and population were used as random nested structure
(Region/Population). LMMs were run using the function "lmer" from the packages "lme4"
(Bates *et al.*, 2007). The model test results were obtained by applying the "Anova" function
from the package "car" (Fox *et al.*, 2012). Post hoc tests were applied to investigate significant
differences among species using the package "lsmeans" (Lenth, 2016).

Third, to investigate the direct and indirect effects of the forest attributes and 197 microclimatic variables on the measured plant traits, we performed structural equation 198 modelling (SEM) separately for each trait. Due to the significant species interactions in the 199 LMMs of the second study question, SEMs were done separately for each species. We used 200 the package "piecewiseSEM" (Lefcheck, 2016), which is particularly suitable for handling 201 LMMs with random factors. In each individual SEM, a measured plant trait (flowering start, 202 proportion of flowering ramets or plant height) was included as the response variable, with 203 204 region and population used as random nested structure. Spring temperature and pH were included as explanatory variables for the plant trait, but also as response variables for structural 205 complexity, crown projection area and percentage of conifers. Additionally, these forest 206 207 structural attributes were included as direct explanatory variables for the measured plant traits and could also correlate amongst each other. The complete data set varied between SEMs, but 208 209 each overall path model was evaluated based on a Fischer's C statistic and on its degrees of freedom and was accepted as an adequate model if the baseline model had a P-value > 0.05. 210 Marginal and conditional R²-values were calculated for each response variable, the former 211 describing the proportion of variance explained by fixed factors alone, the latter describing the 212 variance explained by both fixed and random factors. Despite starting with a model based on 213 a priori knowledge of direct and indirect interactions, pathways with a P-value > 0.1 were 214 removed from the final visualization of the SEMs. 215

217

Results

218 Population differentiation and effect of forest structure and microclimatic variables on plant
219 traits

Emergence of *A. nemorosa* showed a marginally significant difference among populations of
origin (Table 1). Flowering start and plant height differed significantly among populations of
origin across species as well as in the interaction between populations and species (Table 1).
The proportion of flowering ramets varied significantly among populations across species
(Table 1).

Flowering start was, across all species, significantly delayed in plants from sites with 225 226 higher soil pH. Additionally, the significant interaction between soil pH and species indicated that species differed in their relationship between flowering start and soil pH (Table 2). A post-227 hoc Tukey test revealed that V. reichenbachiana significantly differed from both O. acetosella 228 and *M. effusum* (Tukey P-value = < 0.001 and 0.018, respectively) and this species – in contrast 229 to the cross-species pattern – showed delayed flowering start in plants from sites with lower 230 231 soil pH (Fig. S2). Furthermore, the interaction between mean yearly spring temperature and species, showed significant variation among flowering start in species (Table 2), whereas M. 232 *effusum* was found to significantly differ from *O. acetosella* (Tukey P-value = 0.040), and stood 233 out from the other species by delaying flowering start in individuals originating from colder 234 sites (Fig. S3). 235

The proportion of flowering ramets showed significant variation among species. A post-hoc Tukey test revealed that *A. nemorosa* significantly differed from *V. reichenbachiana* and *G. odoratum* (Tukey P-values = <0.001 and 0.040, respectively). Similarly, *O. acetosella* significantly differed from *V. reichenbachiana*, *G. odoratum*, and *M. effusum* (Tukey P-values

= <0.001, 0.026, and <0.001, respectively), and together with A. nemorosa, showed the lowest 240 proportion of flowering ramets (Fig. 1, Fig. S4). At the other end of the scale, M. effusum 241 showed a higher proportion of flowering ramets, significantly differing from the rest of the 242 species (Tukey P-value: G. odoratum = <0.001), except for V. reichenbachiana. Viola 243 reichenbachiana had the highest proportion of flowering ramets of all species, and significantly 244 differed from all species except M. effusum (remaining Tukey P-value to be explained: G. 245 246 $odoratum = \langle 0.001 \rangle$. Lastly, G. odoratum differed significantly from all species and took an intermediate position (Fig. 1, Fig. S4). 247

The significant interaction between soil pH and species for plant height showed that species differed in their relationship between plant height and soil pH (Table 2). Here, *M. effusum* differed significantly from *A. nemorosa* and *O. acetosella* (Tukey P-value = 0.019 and 0.005, respectively), and increased its height with increasing soil acidity (Fig. S5).

252 Direct and indirect effects of forest management on understorey herbs

Multiple SEMs revealed that understorey herbs were affected directly by forest structural 253 attributes and indirectly through the microclimate. However, these effects showed different 254 255 patterns among species. We observed in almost all individual SEMs that our forest structural attributes had strong relationships with each other. Low structural complexity was associated 256 with small crown projection area (found in 100% of SEMs; Fig. 2-4) and with a low percentage 257 258 of conifers (100%) present at the sites of origin, and a large crown projection area was itself also associated with a high percentage of conifers (57.1%). Although significant paths varied 259 depending on the specific trait or species investigated, we saw a general pattern of soil pH 260 261 decreasing with percentage of conifers (78.6%) but increasing with structural complexity (14.3%) and crown projection area (35.7%). Furthermore, spring temperature decreased with 262

increasing percentage of conifers (78.6%) and structural complexity (21.4%) but increased with
increasing crown projection area (78.6%).

Emergence date of *A. nemorosa* was negatively associated with higher soil pH (Fig. 2A), meaning that individuals originating from populations with high soil pH emerged earlier. Individuals of *O. acetosella* flowered later if originating from populations with a high soil pH or high spring temperature (Fig. 2D). The flowering start of *G. odoratum*, *V. reichenbachiana* and *M. effusum* in the common garden was not significantly affected by any environmental variables from the sites of origin (Fig. 2BCE).

The proportion of flowering ramets decreased with increasing mean spring temperatures in *A. nemorosa* (Fig. 3A), decreased with increasing soil pH in *O. acetosella* and *G. odoratum* (Fig. 3BD), and was not affected by any variables from the sites of origin for *V. reichenbachiana* and *M. effusum* (Fig. 3CE).

The height of *O. acetosella* individuals increased with increasing mean spring temperatures (Fig. 4D). Additionally, the plant height of *G. odoratum* was directly affected by forest structure, with individuals growing taller if they originated from populations in more structurally complex forests (Fig. 4B). Neither the plant height of *A. nemorosa* nor that of *M. effusum* was significantly affected by any variables from the sites of origin (Fig. 4AC).

280

281

Discussion

Our common garden study using five forest understorey herbs revealed genetic differentiation among populations in ecologically important traits. Moreover, direct effects of forest structural attributes as well as indirect effects through forest floor microenvironment influenced variation in these traits, although most responses appeared to be species-specific. Taken together, this suggests that forest management influences the evolution of understorey herbs.

287 Genetically based phenotypic variation in five forest understorey herbs

We showed that understorey herbs harbour significant variation in functional traits depending 288 289 on their population of origin. As the measurements were obtained from individuals in a common garden rather than *in situ* in the original populations, this phenotypic variation reflects 290 genetic differentiation among populations (Turesson, 1922; Scheepens and Stöcklin, 2013), 291 292 although we should be aware that epigenetic effects may explain part of this variation (Bossdorf et al. 2008). This genetic differentiation could be the result of adaptation to local conditions, 293 so our next steps was to investigate associations between trait values and environmental values, 294 which would support adaptation as the cause for this genetic variation (Scheepens and Stöcklin, 295 2013). In particular, we investigated associations with forest structural attributes, which are 296 strongly related to forest management decisions, as well as microenvironmental variables that 297 may be influenced by forest management. However, since we did not find overall main effects 298 and since species often differ in the influence of certain environmental factors on plant traits, 299 we will instead focus on the species-specific SEMs revealing contrasting patterns. 300

301 The effects of forest structure and microenvironment

Concerning the species-specific SEMs, we will first discuss the general patterns found among the forest structural attributes, before examining the patterns found in the forest floor microenvironment. Lastly, we will dissect each species-specific SEM and discuss how the individual traits are responding to the forest structural attributes and the microenvironment at their sites of origin.

307 General patterns among forest structural attributes were found across all SEMs. 308 Structural complexity was negatively associated with both crown projection area and the 309 percentage of conifers at the sites of origin. Since coniferous forests are often associated with 310 high forest management intensity (Schall and Ammer, 2013; Willems *et al.*, 2021), we often

see cohorts of homogenous coniferous plantations leading to low structural complexity 311 (Pommerening, 2004). Similarly, a large crown projection area, caused by coniferous-312 313 dominated forests or a monospecific deciduous forest with large crowns, could be the potential cause of the negative association with structural complexity. In our dataset, conifers had larger 314 crown areas compared to deciduous trees, supporting the positive relationship between 315 percentage of conifers and crown projection area (Grote, 2003). However, it has been shown 316 317 that the crown radii of mixed stands and conifer stands are generally overestimated, while deciduous crowns are underestimated, due to overlapping of crowns in dense and structurally 318 319 complex stands (Grote, 2003).

Concurrent with previous studies (e.g., Willems et al. 2021), we found that a higher 320 percentage of conifers decreased mean spring temperatures. Most coniferous trees are 321 evergreen, and hence, the forest floor beneath them is constantly shaded, decreasing the 322 temperature experienced by the understory herb community. Large crown projection areas 323 create a bigger "forest roof", which not only buffers the temperature on the forest floor and 324 increases mean spring temperatures, but also protects against wind and other mechanistic 325 stresses (De Frenne et al., 2021). Structural complexity creates heterogenous microclimates at 326 fine spatiotemporal scales, and various canopy openings could disrupt the buffering of the 327 "forest roof", ultimately decreasing spring temperatures (De Frenne et al., 2021). 328

Conifer species are often planted on acidic soils, but they can also contribute to decrease soil pH even further by promoting soil aluminium contents (Augusto *et al.*, 2003). Stands with a high structural complexity usually harbour deciduous trees or a mix of deciduous and coniferous trees. In other words, they usually have a low percentage of conifers, which could explain the positive relationship with soil pH. Large crown projection area can be characterized by dense and overlapping deciduous tree crowns, and deciduous trees are associated with more alkaline soils compared to coniferous stands (Augusto *et al.*, 2003).

It is crucial for spring geophytes to have appropriate responses to thermal cues to avoid 336 frost damage on the one hand but also utilize the maximum amount of high light on the other 337 hand in order to successfully finish a reproductive cycle before tree leaf out (Heberling et al., 338 2019). Individuals originating from colder, often coniferous, plots might be adapted to 339 phenological cues at lower temperatures, which persists when they are transplanted to a 340 common garden in Frankfurt, thus having an advance phenological stage in the common garden 341 342 compared to individuals originating from warmer plots. In line with previous studies (Gentili et al., 2018), A. nemorosa individuals originating from more acidic soils emerged earlier in our 343 344 common garden, establishing the importance of pH as a key factor and soil property. In line with this, the phenological cue for flowering start in O. acetosella in our common garden, is 345 later in individuals originating from warmer and more alkaline sites (Sercu et al., 2021), and 346 based on our SEMs these sites mainly consist of deciduous tree species. 347

The proportion of flowering ramets can be seen as a proxy for the amount of resources 348 349 an individual chooses to allocate to sexual reproduction versus vegetative reproduction. Anemone nemorosa favours vegetative reproduction during colder spring temperatures and 350 hence conserves valuable resources for warmer periods when investment in flowers is more 351 successful owing to higher pollinator activity (Hegland et al., 2009). Additionally, we saw that 352 both O. acetosella and G. odoratum reduce their amount of flowering ramets in individuals 353 originating from acidic soils. Previous studies (e.g., Blondeel et al., 2020; Sercu et al., 2021) 354 report similar results, pointing towards the importance of pH on nutrient availability and the 355 potential importance of pH as a driver of trait variation (Gentili et al., 2018). 356

Plant height was the only measured trait that was directly affected by forest structural attributes. The structural complexity of the forest canopy heavily influences light availability, a resource causing competition within the understorey herb community, and these biotic forces could potentially cause increased height in *G. odoratum* (Pau *et al.*, 2011). Additionally, increased spring temperatures allow for greater carbon assimilation (Heberling *et al.*, 2019)
and for *O. acetosella* to invest in increased plant height.

Why do we not see any influence of forest or microenvironmental variables on the traits of *M. effusum* and *V. reichenbachiana*? Despite showing evidence for genetic differentiation, some species are not responding to the microenvironmental variables from their sites of origin. It could very well be that we picked the wrong environmental variables that could explain it, or that the variation is neutral with respect to any environmental variables.

368 Previous work by Willems et al. (2021) was carried out on the same forest plots, but as the results were based on field observations alone, no conclusion about the genetic basis could 369 be made. With our complimentary work, we now established that the effects of forest 370 371 management found under natural conditions (Willems et al., 2021) is a result of genetic 372 differentiation and thus evolution at small spatial scales. Since our plants originated from within three regions in Germany, we are not comparing forests from across a large spatial range 373 374 subject to macroclimatic variation. Instead, all observed genetic variation occurs within these three regions, despite gene flow counteracting population differentiation, especially in the 375 376 species with higher dispersal abilities (notably *M. effusum*). An important implication is that all observed genetic variation can thus also be conserved at this spatial scale. By applying 377 378 diverse forest management actions across regional scales, we create heterogeneity that likely 379 selects for different plant traits and thus helps conserving genetic diversity.

380 *Conclusion*

In conclusion, forest understorey herbs showed genetic differentiation in ecologically important traits among the sampled populations. Our results suggests that forest management, which strongly determines the forest structural variables included in this study (Ehbrecht *et al.*, 2017), drives the evolution of forest understorey herbs either directly or indirectly through its

influence on the microenvironment. Given that forest understorey herbs are important for
biodiversity and ecosystem services, it is imperative that future management considers this.
Diversifying management practices can affect intraspecific genetic diversity and thus can play
a role in the conservation of genetic diversity of understorey herbs.

389

390 Acknowledgements

We thank the current managers of the three Exploratories - Julia Bass, Max Müller, Anna K. 391 392 Franke, Miriam Teuscher, Robert Künast, Franca Marian - and all former managers for their work in maintaining the plot and project infrastructure; Christiane Fischer and Victoria 393 Grießmeier for giving support through the central office; Andreas Ostrowski for managing the 394 395 central data base, and Markus Fischer, Eduard Linsenmair, Dominik Hessenmöller, Daniel Prati, Ingo Schöning, François Buscot, Ernst-Detlef Schulze, Wolfgang W. Weisser and the 396 late Elisabeth Kalko for their role in setting up the Biodiversity Exploratories project. We thank 397 the administration of the Hainich national park, the UNESCO Biosphere Reserve Swabian Alb, 398 and the UNESCO Biosphere Reserve Schorfheide-Chorin as well as all landowners for the 399 400 excellent collaboration. We thank Oliver Bossdorf for providing facilities during the initial phase of this study in Tübingen, and Robert Anton, Susanne Pietsch, and the staff of 401 Wissenschaftsgarten at Goethe University Frankfurt for their great support. Lastly, we would 402 403 like to thank Lutz Stübing, Pascal Karitter, Marcel Glück, Silas Büse, Delia Gartner, Mariya Antsupova, and Katharina Dietrich for their help in setting up the experimental system and for 404 assisting with data collection. 405

406

407 Funding

This work has been supported by the Deutsche Forschungsgemeinschaft (DFG) Priority
Program 1374 "Biodiversity Exploratories" through grant SCHE 1899/5-1 to JFS. Field work
permits were issued by the responsible state environmental offices of Baden-Württemberg,
Thüringen, and Brandenburg.

412

413 **Data availability statement**

This work is based on data elaborated by the HerbAdapt and Forest Structure (core) projects 414 of the Biodiversity Exploratories program (DFG Priority Program 1374). The datasets are 415 publicly available Information in the Biodiversity Exploratories System 416 (http://doi.org/10.17616/R32P9Q, https://www.bexis.uni-jena.de/ddm/data/Showdata/22766, 417 418 and https://www.bexis.uni-jena.de/ddm/data/Showdata/31455).

419

420 **Conflict of interests**

421 The authors have no conflict of interest to declare.

422

423 Author contributions

CM, PDF and JFS designed the study. CM conducted the field sampling and the common
garden experiment with help from MMS. CM and PDF analysed the data with input from TV,
KV and JFS. CM wrote the first draft of the manuscript with all co-authors contributing to
revisions. Acquisition of the financial support of the project leading to this publication by PDF
and JFS.

References

430	Alpert P, Simms EL. 2002. The relative advantages of plasticity and fixity in different
431	environments: when is it good for a plant to adjust? <i>Evolutionary Ecology</i> 16 : 285–297.
432	doi:10.1023/A:1019684612767.
433	Augusto L, Dupouey J-L, Ranger J. 2003. Effects of tree species on understory vegetation
434	and environmental conditions in temperate forests. Ann. For. Sci. 60: 823-831.
435	doi:10.1051/forest:2003077.
436	Aussenac G. 2000. Interactions between forest stands and microclimate: Ecophysiological
437	aspects and consequences for silviculture. Ann. For. Sci. 57: 287-301.
438	doi:10.1051/forest:2000119.
439	Bates D, Sarkar D, Bates MD, Matrix L. 2007. The lme4 package: R package version.
440	Blondeel H, Perring MP, Depauw L, et al. 2020. Light and warming drive forest
441	understorey community development in different environments. Global Change Biology
442	26 : 1681–1696. doi:10.1111/gcb.14955.
443	Brunet J, De Frenne P, Holmström E, Mayr ML. 2012. Life-history traits explain rapid
444	colonization of young post-agricultural forests by understory herbs. Forest Ecology and
445	Management 278: 55-62. doi:10.1016/j.foreco.2012.05.002.
446	De Frenne P, Brunet J, Shevtsova A, et al. 2011. Temperature effects on forest herbs
447	assessed by warming and transplant experiments along a latitudinal gradient. Glob.
448	<i>Change Biol.</i> 17 : 3240–3253. doi:10.1111/j.1365-2486.2011.02449.x.
449	De Frenne P, Kolb A, Verheyen K, et al. 2009. Unravelling the effects of temperature,

- 450 latitude and local environment on the reproduction of forest herbs. *Global Ecology and*
- *Biogeography* **18**: 641–651. doi:10.1111/j.1466-8238.2009.00487.x.

- 452 **De Frenne P, Lenoir J, Luoto M, et al. 2021**. Forest microclimates and climate change:
- Importance, drivers and future research agenda. *Global Change Biology* 27: 2279–2297.
 doi:10.1111/gcb.15569.
- 455 De Frenne P, Rodríguez-Sánchez F, Coomes DA, et al. 2013. Microclimate moderates
- 456 plant responses to macroclimate warming. *Proceedings of the National Academy of*
- 457 *Sciences of the United States of America* **110**: 18561–18565.

458 doi:10.1073/pnas.1311190110.

- 459 Ehbrecht M, Schall P, Ammer C, Seidel D. 2017. Quantifying stand structural complexity
- 460 and its relationship with forest management, tree species diversity and microclimate.

461 *Agricultural and Forest Meteorology* **242**: 1–9. doi:10.1016/j.agrformet.2017.04.012.

- 462 Fischer M, Bossdorf O, Gockel S, et al. 2010. Implementing large-scale and long-term
- 463 functional biodiversity research: The Biodiversity Exploratories. *Basic and Applied*

464 *Ecology* **11**: 473–485. doi:10.1016/j.baae.2010.07.009.

- Fox J, Weisberg S, Adler D, *et al.* 2012. *Package 'car'*: Vienna: R Foundation for Statistical
 Computing.
- 467 Frederiksen S, Rasmussen FN. 2006. *Dansk flora*: Gyldendal A/S.
- 468 Gentili R, Ambrosini R, Montagnani C, Caronni S, Citterio S. 2018. Effect of Soil pH on
- the Growth, Reproductive Investment and Pollen Allergenicity of Ambrosia artemisiifolia
- 470 L. Front. Plant Sci. 9: 1335. doi:10.3389/fpls.2018.01335.
- 471 Gilliam FS. 2007. The Ecological Significance of the Herbaceous Layer in Temperate Forest
- 472 Ecosystems. *BioScience* **57**: 845–858. doi:10.1641/B571007.
- 473 Grote R. 2003. Estimation of crown radii and crown projection area from stem size and tree
- 474 position. Ann. For. Sci. 60: 393–402. doi:10.1051/forest:2003031.

- 475 Heberling JM, McDonough MacKenzie C, Fridley JD, Kalisz S, Primack RB. 2019.
- 476 Phenological mismatch with trees reduces wildflower carbon budgets. *Ecology letters* 22:
- 477 616–623. doi:10.1111/ele.13224.
- 478 Hegland SJ, Nielsen A, Lázaro A, Bjerknes A-L, Totland Ø. 2009. How does climate
- 479 warming affect plant-pollinator interactions? *Ecology letters* **12**: 184–195.
- 480 doi:10.1111/j.1461-0248.2008.01269.x.
- 481 IPCC. 2014. Climate change 2014: Synthesis report. Contribution of Working Groups I, II
- 482 *and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate change.*
- 483 Geneva, Switzerland.
- 484 Landuyt D, Lombaerde E de, Perring MP, et al. 2019. The functional role of temperate
- forest understorey vegetation in a changing world. *Global Change Biology* 25: 3625–
- 486 3641. doi:10.1111/gcb.14756.
- 487 **Lantz B. 2019**. *Machine Learning with R: Expert techniques for predictive modeling*.
- 488 Birmingham: Packt Publishing.
- 489 Lefcheck JS. 2016. piecewiseSEM Piecewise structural equation modelling in r for ecology,
- 490 evolution, and systematics. *Methods Ecol Evol* **7**: 573–579. doi:10.1111/2041-
- 491 210X.12512.
- 492 Lemke IH, Kolb A, Graae BJ, et al. 2015. Patterns of phenotypic trait variation in two
- temperate forest herbs along a broad climatic gradient. *Plant Ecol* **216**: 1523–1536.
- doi:10.1007/s11258-015-0534-0.
- 495 Lenth RV. 2016. Least-Squares Means: The R Package Ismeans. J. Stat. Soft. 69: 1–33.
- 496 doi:10.18637/jss.v069.i01.
- 497 Møller C, March-Salas M, Kuppler J, De Frenne P, Scheepens JF. 2022. Intra-individual
- 498 *variation in Galium odoratum is affected by experimental drought and shading*. Annals of499 Botany.

500	Pau S, Wolkovich EM, Cook BI, et al. 2011. Predicting phenology by integrating ecology,
501	evolution and climate science. Glob. Change Biol. 17: 3633-3643. doi:10.1111/j.1365-
502	2486.2011.02515.x.
503	Pommerening A. 2004. A review of the history, definitions and methods of continuous cover
504	forestry with special attention to afforestation and restocking. Forestry 77: 27-44.
505	doi:10.1093/forestry/77.1.27.
506	R Core Team. 2021. R: A language and environment for statistical computing. Vienna,
507	Austria.
508	Schall P, Ammer C. 2013. How to quantify forest management intensity in Central
509	European forests. Eur J Forest Res 132: 379–396. doi:10.1007/s10342-013-0681-6.
510	Scheepens JF, Stöcklin J. 2013. Flowering phenology and reproductive fitness along a
511	mountain slope: maladaptive responses to transplantation to a warmer climate in
512	Campanula thyrsoides. <i>Oecologia</i> 171 : 679–691. doi:10.1007/s00442-012-2582-7.
513	Sercu BK, Moeneclaey I, Goeminne B, Bonte D, Baeten L. 2021. Flowering phenology
514	and reproduction of a forest understorey plant species in response to the local
515	environment. Plant Ecol 222: 749–760. doi:10.1007/s11258-020-01104-2.
516	Spiecker H. 2003. Silvicultural management in maintaining biodiversity and resistance of
517	forests in Europe-temperate zone. Journal of environmental management 67: 55-65.
518	doi:10.1016/s0301-4797(02)00188-3.
519	Tobias JA, Durant SM, Pettorelli N. 2021. Improving Predictions of Climate Change-Land
520	Use Change Interactions. <i>Trends in Ecology & Evolution</i> 36 : 29–38.
521	doi:10.1016/j.tree.2020.08.019.
522	Turesson G. 1922 . The genotypical response of the plant species to the habitat 3 : 211–350.
523	Valladares F, Gianoli E, Gómez JM. 2007. Ecological limits to plant phenotypic plasticity
524	176(4) : 749–763.

- 525 Willems FM, Scheepens JF, Ammer C, *et al.* 2021. Spring understory herbs flower later in
- 526 intensively managed forests. *Ecological applications a publication of the Ecological*
- *Society of America* **31**: e02332. doi:10.1002/eap.2332.

Table 1: Results of mixed-effects models of emergence (for *Anemone nemorosa* only), flowering start, plant height and the proportion of flowering ramets in five different forest understorey herbs explained by population, species, and their interaction as fixed factors, and region as random variable. Chi-square (X^2) test values, degrees of freedom (df), and P-values (P) are given. Significant P-values are indicated in bold followed by * (<0.05), ** (<0.01) and *** (<0.001), and marginally significant values are indicated in bold followed by •.

532

	F	Emerger	ice	Flowering start			Proportio	on of flowe	ring ramets	Plant height			
	X^2	df	Р	X^2	df	Р	X^2	df	Р	X^2	df	Р	
Population	85.40	67	0.064 ·	160.0	116	0.004 **	162.5	127	0.018 *	258.3	123	<0.001 ***	
Species				0.946	4	0.918	3.839	4	0.428	3.135	3	0.371	
Population * Species				220.7	164	0.002 **	216.5	0.439	0.439	214.6	148	<0.001 ***	

Table 2. Results of mixed-effects models of emergence (for *Anemone nemorosa* only), flowering start, plant height and the proportion of flowering ramets in five different forest understorey herbs explained by species, environmental variables, and their interactions with species as fixed factors, and with region and populations included as nested random variables. For continuous variables, upwards arrows (\uparrow) indicate positive estimates and downwards arrows (\downarrow) indicate negative estimates. Chi-square (X^2) test values, degrees of freedom (df), and P-values (P) are given. Significant P-values are indicated in bold followed by * (<0.05), ** (<0.01) and *** (<0.001), and marginally significant values are indicated in bold followed by •.

	Emergence					Flowering start			Proportion of				Plant height			
										flowering ramets						
		X^2	df	Р		X^2	df	Р		X^2	df	Р		X^2	df	Р
Species							4	0.997		212.78	4	<0.001 ***		1.792	3	0.617
pH	\downarrow	5.663	1	0.017 *	\downarrow		1	0.021 *	\downarrow	2.032	1	0.154	\downarrow	1.633	1	0.201
Spring temperature	\downarrow	1.210	1	0.271	\downarrow		1	0.428	\downarrow	0.416	1	0.519	\downarrow	1.254	1	0.263
Conifer	\downarrow	0.113	1	0.737	\downarrow		1	0.952	\downarrow	2.237	1	0.135	\downarrow	1.373	1	0.241
Structural complexity	\downarrow	0.646	1	0.422	\downarrow		1	0.799	\downarrow	0.535	1	0.465	\downarrow	0.977	1	0.323
Crown projection area	1	0.061	1	0.805	\downarrow		1	0.331	\downarrow	0.653	1	0.419	\downarrow	0.020	1	0.888
pH * Species							4	<0.001 ***		1.243	4	0.871		13.35	3	0.004 **
Spring temperature * Species							4	0.047 *		3.440	4	0.487		3.495	3	0.321
Conifer * Species							4	0.770		8.825	4	0.066 ·		7.146	3	0.067 ·
Structural complexity * Species							4	0.954		0.781	4	0.941		5.667	3	0.129
Crown projection area * Species							4	0.958		1.448	4	0.836		1.377	3	0.711



Figure 1. Trait distribution and number of observations (N) for (A) flowering period [DOY], (B) proportion of flowering ramets, and (C) plant height [cm] (x axis logged) for *Anemone nemorosa*, *Galium odoratum*, *Milium effusum*, *Oxalis acetosella*, and *Viola reichenbachiana*. Photo credits: Franziska Willems,
 Pieter De Frenne & Charlotte Møller.



Figure 2. Effects of forest structural attributes and microenvironment of the sites of origin on phenological traits in the common garden. Shown are the results of the piecewise structural equation model (SEM) for direct and indirect relationships among forest characteristics and microclimatic variables, and the flowering start (DOY) for (A) *Anemone nemorosa*, (B) *Galium odoratum*, (C) *Milium effusum*, (D) *Oxalis acetosella*, and (E) *Viola reichenbachiana*. Arrows represent unidirectional and bidirectional relationships among variables; only marginally or more significant paths (P < 0.1) are shown. Solid lines are positive relationships, dashed lines are negative ones. The line thickness is proportional to the magnitude of the standardized estimates, plotted near each line. The R² values are given for each SEM, along with the overall model fit to the data as Fischer's C, its associated P-value and degrees of freedom (Df).







Figure 4. Effects of forest structural attributes and microclimate of the site of origin on plant height in the common garden. Shown are the results of the piecewise structural equation model (SEM) for direct and indirect relationships among forest characteristics and microclimatic variables, and plant height for (A) *Anemone nemorosa*, (B) *Galium odoratum*, (C) *Milium effusum*, and (D) *Oxalis acetosella*. Arrows represent unidirectional and bidirectional relationships among variables; only marginally or more significant paths (P < 0.1) are shown. Solid lines are positive relationships, dashed lines are negative ones. The line thickness is proportional to the magnitude of the standardized estimates, plotted near each line. The R² values are given for each SEM, along with the overall model fit to the data as Fischer's C, its associated P-value and degrees of freedom (Df).