

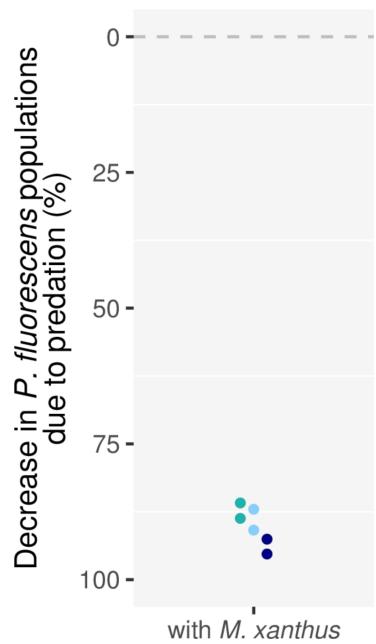
Supporting information for Killer prey: Pre-interaction ecology reverses bacterial predation

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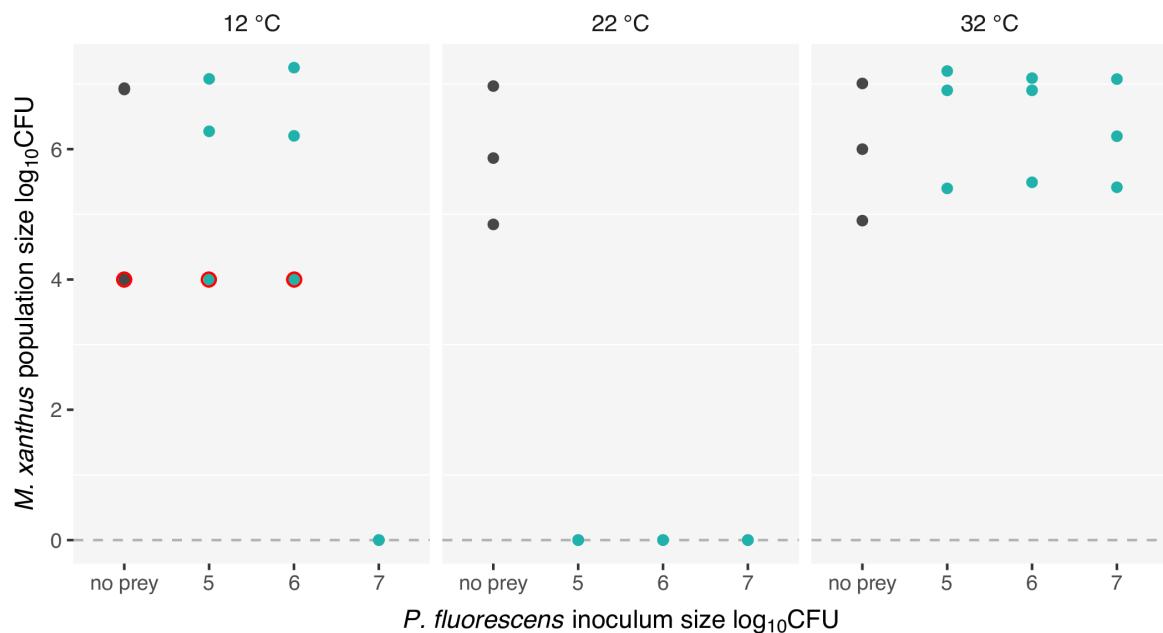
This document contains the Supplementary figures S1-6 and Supplementary tables S1 and S2.

Figure S1.



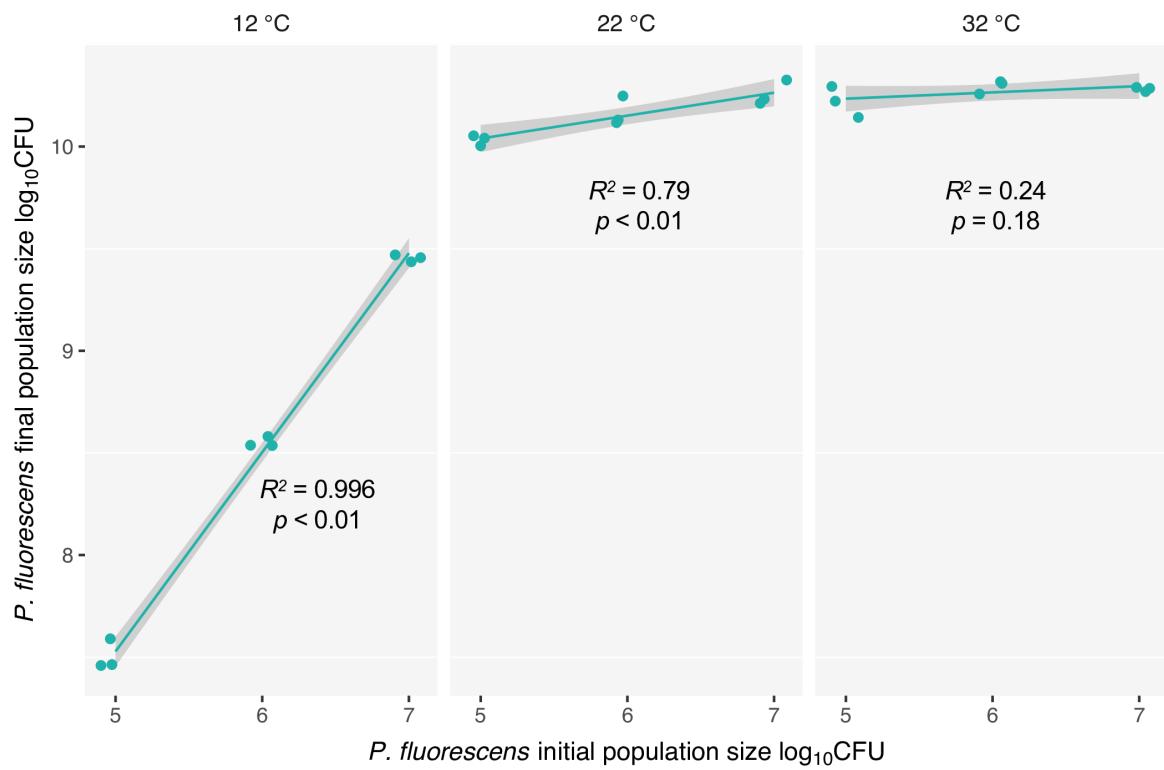
***M. xanthus* kills *P. fluorescens* grown on M9cas agar at 32 °C.** Percentage reduction of *P. fluorescens* population size after four days in the presence of *M. xanthus* relative to in the absence of *M. xanthus*. Colors correspond to three independent replicates each run with two technical replicates.

Figure S2.



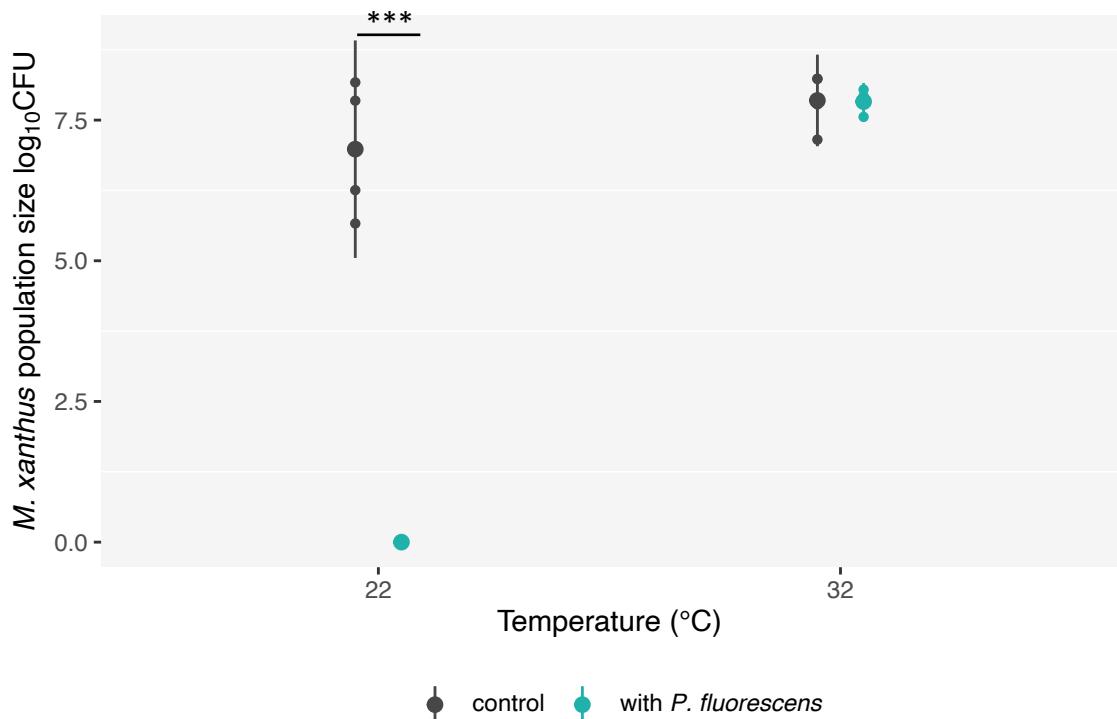
***P. fluorescens* grown at 22 °C kills *M. xanthus* after 30 minutes of interaction.** *M. xanthus* population sizes 30 minutes after inoculation onto *P. fluorescens* lawns grown overnight from one of three inoculum sizes and at one of three temperatures (green dots) or onto bacteria-free control plates that had been incubated overnight at one of the same three temperatures prey (black dots). Means of log₁₀-transformed CFU + 1 values and 95% confidence intervals are shown. Note that, due to a technical issue with dilution-plating for one replicate of the 12 °C treatment, the highest available plated dilution was too low to accurately count colonies; the corresponding plates had more colonies than could feasibly be counted. We therefore attributed counts of 1000 for these plates, which was clearly a substantial underestimate in each case. These underestimated values are identified in the graph with red circles around corresponding data points. Lighter dots are biological replicates ($n = 3$).

Figure S3.



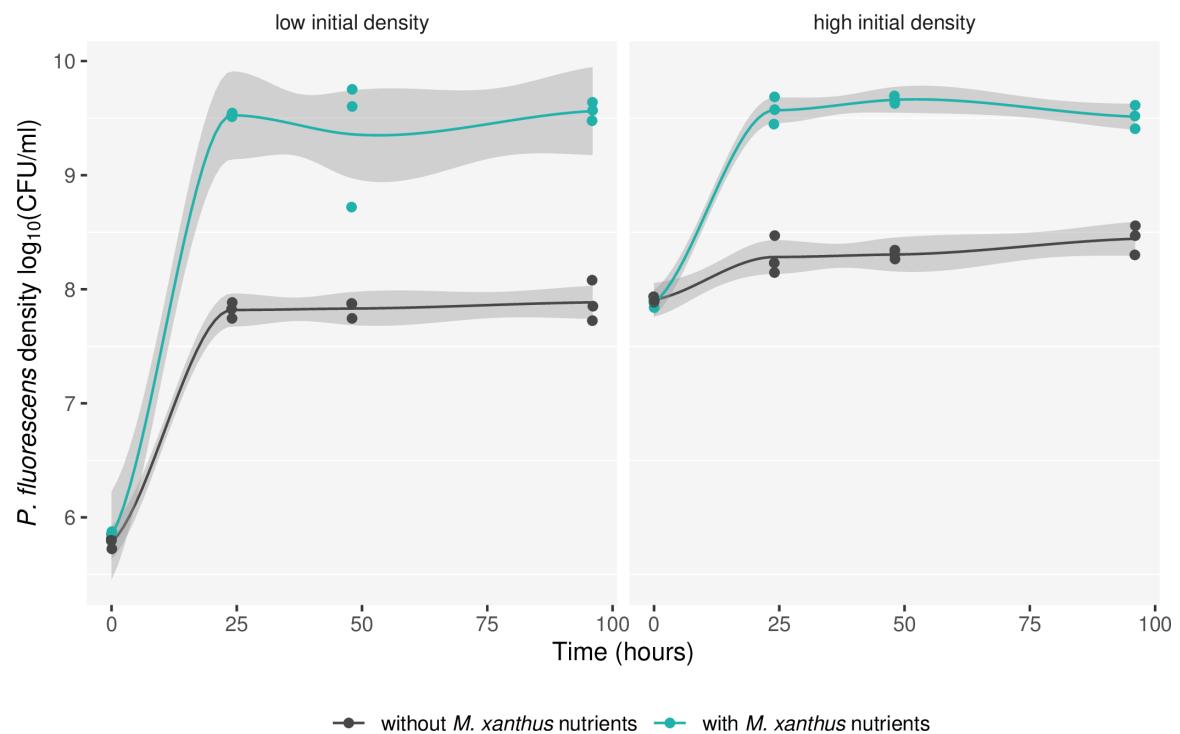
Final *P. fluorescens* population size correlates positively with initial population size after growth at 12 and 22 °C but not at 32 °C. Relationship between *P. fluorescens* initial and final population sizes after 22 hours of growth at different temperatures 12, 22 or 32 °C. Log₁₀-transformed CFU values ($n = 3$), linear fits and 95% confidence intervals about the linear fits are shown.

Figure S4.



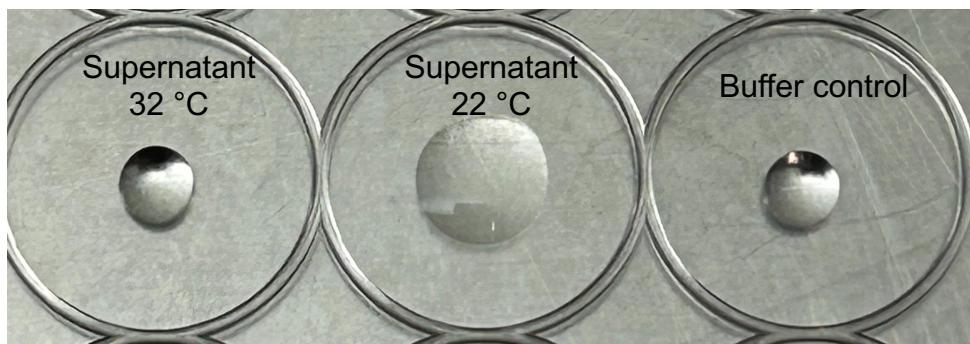
***P. fluorescens* kills *M. xanthus* when both species are pre-grown and interact at 22 °C but not when both are pre-grown and interact at 32 °C.** DK3470 population sizes are shown 24 hours after interaction with *P. fluorescens* when the two species interact at the same temperature at which both had been reared prior to interaction. Means of \log_{10} -transformed CFU + 1 values and 95% confidence intervals are shown. Lighter dots are biological replicates ($n = 3$). *** $p < 0.001$ (Tukey-adjusted contrasts) for the difference between *M. xanthus* population size after interaction with *P. fluorescens* (green dots) vs in the control treatment (black dots) when the two species were reared and interacted at 22 °C.

Figure S5.



Reversed predation decomposes *M. xanthus* cells into diffusible nutrients. Remains of *M. xanthus* killed by 22 °C-reared *P. fluorescens* sufficient to fuel large *P. fluorescens* population growth pass through 0.2-µm filters within six hours of inter-species interaction. Estimated densities (log-transformed CFU/ml, $n = 3$) of *P. fluorescens* populations over time inoculated at two initial densities (~ 10^6 and ~ 10^8 CFU/ml) into supernatant from 22 °C-reared *P. fluorescens* lawns to which *M. xanthus* cells were either added (and which killed those *M. xanthus* cells, green dots) or not (black dots). Trend lines show local polynomial regression fitting and dark-gray bands represent 95% confidence regions.

Figure S6.



Drop-collapse assay with *P. fluorescens* supernatants. Picture of 20- μ l aliquots of supernatants from liquid suspensions of *P. fluorescens* grown on M9cas agar at 32 °C (left) and 22 °C (center), and of M9 buffer control (right). The three replicates of this assay yielded visually indistinguishable results; one replicate is shown here.

Table S1. Statistical analysis of *M. xanthus* swarming data on prey lawns. Linear model and Type III ANOVA for swarming data using prey identity, predator identity, temperature treatment and their interactions as explanatory variables. Posthoc contrasts between temperature treatments are computed for each predator-prey combination.

A. Linear model

```
model <- lm(formula = swarm_diameter ~ prey * predator * temperature)
```

Multiple R-squared: 0.9325, Adjusted R-squared: 0.908

F-statistic: 38.08 on 62 and 171 DF, p-value: < 2.2e-16

Anova Table (Type III tests)

Response: swarm_diameter

	Sum Sq	Degree of freedom	F value	Pr(>F)
(Intercept)	52.926	1	442.719	< 2.2e-16 ***
prey	22.681	6	31.621	< 2.2e-16 ***
predator	8.065	2	33.733	4.478e-13 ***
temperature	0.035	2	0.146	0.864
prey:predator	10.384	12	7.238	1.234e-10 ***
prey:temperature	7.294	12	5.084	3.151e-07 ***
predator:temperature	0.022	4	0.047	0.996
prey:predator:temperature	1.439	24	0.501	0.975
Residuals	20.442	171		

Signif. codes: 0 = ***, 0.001 = **, 0.01 = *, 0.05 = .

B. Tukey-adjusted contrasts on temperature treatment for each predator-prey combination

```
pairs(emmeans::emmeans(model, "temperature", by = c("prey", "predator")))
```

prey = NO, predator = GJV1:

```
contrast estimate SE df. t.ratio p.value
12 - 22 0.1250 0.244 171 0.511 0.8660
12 - 32 0.1000 0.244 171 0.409 0.9120
22 - 32 -0.0250 0.244 171 -0.102 0.9943
```

prey = AG, predator = GJV1:

```
contrast estimate SE df. t.ratio p.value
12 - 22 0.1625 0.244 171 0.665 0.7843
12 - 32 0.2875 0.244 171 1.176 0.4691
22 - 32 0.1250 0.244 171 0.511 0.8660
```

prey = BB, predator = GJV1:

```
contrast estimate SE df. t.ratio p.value
12 - 22 -0.1250 0.244 171 -0.511 0.8660
12 - 32 -0.0125 0.244 171 -0.051 0.9986
22 - 32 0.1125 0.244 171 0.460 0.8899
```

prey = EC, predator = GJV1:

```
contrast estimate SE df. t.ratio p.value
12 - 22 -0.2250 0.244 171 -0.920 0.6282
12 - 32 -0.3750 0.244 171 -1.534 0.2777
22 - 32 -0.1500 0.244 171 -0.614 0.8129
```

prey = ML, predator = GJV1:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	-0.2875	0.244	171	-1.176	0.4691	
12 - 32	-0.0750	0.244	171	-0.307	0.9495	
22 - 32	0.2125	0.244	171	0.869	0.6604	

prey = PF, predator = GJV1:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.7250	0.244	171	2.965	0.0096	
12 - 32	-1.1875	0.244	171	-4.857	<.0001	
22 - 32	-1.9125	0.244	171	-7.823	<.0001	

prey = RV, predator = GJV1:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.0250	0.346	171	0.072	0.9971	
12 - 32	0.0750	0.346	171	0.217	0.9744	
22 - 32	0.0500	0.346	171	0.145	0.9885	

prey = NO, predator = A75:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.0000	0.244	171	0.000	1.0000	
12 - 32	0.0875	0.244	171	0.358	0.9319	
22 - 32	0.0875	0.244	171	0.358	0.9319	

prey = AG, predator = A75:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.2250	0.244	171	0.920	0.6282	
12 - 32	0.4125	0.244	171	1.687	0.2130	
22 - 32	0.1875	0.244	171	0.767	0.7238	

prey = BB, predator = A75:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.0875	0.244	171	0.358	0.9319	
12 - 32	0.0625	0.244	171	0.256	0.9646	
22 - 32	-0.0250	0.244	171	-0.102	0.9943	

prey = EC, predator = A75:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	-0.1375	0.244	171	-0.562	0.8402	
12 - 32	-0.1375	0.244	171	-0.562	0.8402	
22 - 32	0.0000	0.244	171	0.000	1.0000	

prey = ML, predator = A75:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.2000	0.244	171	0.818	0.6924	
12 - 32	0.0625	0.244	171	0.256	0.9646	
22 - 32	-0.1375	0.244	171	-0.562	0.8402	

prey = PF, predator = A75:

	contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.5125	0.244	171	2.096	0.0936	
12 - 32	-1.8625	0.244	171	-7.618	<.0001	
22 - 32	-2.3750	0.244	171	-9.714	<.0001	

prey = RV, predator = A75:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.0250	0.346	171	0.072	0.9971
12 - 32	-0.0250	0.346	171	-0.072	0.9971
22 - 32	-0.0500	0.346	171	-0.145	0.9885

prey = NO, predator = SO1:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.1125	0.244	171	0.460	0.8899
12 - 32	0.1125	0.244	171	0.460	0.8899
22 - 32	0.0000	0.244	171	0.000	1.0000

prey = AG, predator = SO1:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.1375	0.244	171	0.562	0.8402
12 - 32	0.2750	0.244	171	1.125	0.5001
22 - 32	0.1375	0.244	171	0.562	0.8402

prey = BB, predator = SO1:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.1500	0.244	171	0.614	0.8129
12 - 32	0.1625	0.244	171	0.665	0.7843
22 - 32	0.0125	0.244	171	0.051	0.9986

prey = EC, predator = SO1:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.0000	0.244	171	0.000	1.0000
12 - 32	-0.1000	0.244	171	-0.409	0.9120
22 - 32	-0.1000	0.244	171	-0.409	0.9120

prey = ML, predator = SO1:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.0125	0.244	171	0.051	0.9986
12 - 32	0.0625	0.244	171	0.256	0.9646
22 - 32	0.0500	0.244	171	0.205	0.9772

prey = PF, predator = SO1:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.5625	0.244	171	2.301	0.0584
12 - 32	-0.9375	0.244	171	-3.835	0.0005
22 - 32	-1.5000	0.244	171	-6.135	<.0001

prey = RV, predator = SO1:

contrast	estimate	SE	df.	t.ratio	p.value
12 - 22	0.2250	0.346	171	0.651	0.7922
12 - 32	0.1000	0.346	171	0.289	0.9549
22 - 32	-0.1250	0.346	171	-0.362	0.9305

P value adjustment: tukey method for comparing a family of 3 estimates

Table S2. Statistical analysis of *M. xanthus* growth data. Linear model and Type III ANOVA for *M. xanthus* growth data using *P. fluorescens* inoculum size, temperature treatment, their interaction and time as explanatory variables. Posthoc contrasts between inoculum sizes are computed for each temperature-time combination.

A. Linear model

```
model <- lm(log10(predator_number + 1) ~ prey_inoculum_size * rearing_temperature + time)
```

Multiple R-squared: 0.9407, Adjusted R-squared: 0.9286

F-statistic: 77.93 on 12 and 59 DF, p-value: < 2.2e-16

Anova Table (Type III tests)

Response: log10(predator_number + 1)

	Sum Sq	Degree of freedom	F value	Pr(>F)
(Intercept)	244.066	1	315.709	< 2.2e-16 ***
prey_inoculum_size	192.800	3	83.131	< 2.2e-16 ***
rearing_temperature	0.049	2	0.032	0.969
time	10.421	1	13.480	0.001
prey_inoculum_size:rearing_temperature	195.261	6	42.096	< 2.2e-16 ***
Residuals	45.611	59		

Signif. codes: 0 = ***, 0.001 = **, 0.01 = *, 0.05 = .

B. Tukey-adjusted contrasts on prey inoculum size for each temperature-time combination

```
pairs(emmeans::emmeans(model, "prey_inoculum_size", by = c("rearing_temperature", "time")))
```

temperature = 12, time = 30 min:

contrast	estimate	SE	df.	t.ratio	p.value
no prey - 5	0.8001	0.508	59	1.576	0.3999
no prey - 6	0.7414	0.508	59	1.461	0.4675
no prey - 7	7.0188	0.508	59	13.826	<.0001
5 - 6	-0.0587	0.508	59	-0.116	0.9994
5 - 7	6.2187	0.508	59	12.250	<.0001
6 - 7	6.2773	0.508	59	12.366	<.0001

temperature = 22, time = 30 min:

contrast	estimate	SE	df.	t.ratio	p.value
no prey - 5	6.9464	0.508	59	13.684	<.0001
no prey - 6	6.9464	0.508	59	13.684	<.0001
no prey - 7	6.9464	0.508	59	13.684	<.0001
5 - 6	0.0000	0.508	59	0.000	1.0000
5 - 7	0.0000	0.508	59	0.000	1.0000
6 - 7	0.0000	0.508	59	0.000	1.0000

temperature = 32, time = 30 min:

contrast	estimate	SE	df.	t.ratio	p.value
no prey - 5	0.4443	0.508	59	0.875	0.8176
no prey - 6	0.4176	0.508	59	0.823	0.8435
no prey - 7	0.6910	0.508	59	1.361	0.5284
5 - 6	-0.0267	0.508	59	-0.053	0.9999
5 - 7	0.2467	0.508	59	0.486	0.9619
6 - 7	0.2734	0.508	59	0.539	0.9492

temperature = 12, time = 7 days:

contrast	estimate	SE	df.	t.ratio	p.value
no prey - 5	0.8001	0.508	59	1.576	0.3999
no prey - 6	0.7414	0.508	59	1.461	0.4675
no prey - 7	7.0188	0.508	59	13.826	<.0001
5 - 6	-0.0587	0.508	59	-0.116	0.9994
5 - 7	6.2187	0.508	59	12.250	<.0001
6 - 7	6.2773	0.508	59	12.366	<.0001

temperature = 22, time = 7 days:

contrast	estimate	SE	df.	t.ratio	p.value
no prey - 5	6.9464	0.508	59	13.684	<.0001
no prey - 6	6.9464	0.508	59	13.684	<.0001
no prey - 7	6.9464	0.508	59	13.684	<.0001
5 - 6	0.0000	0.508	59	0.000	1.0000
5 - 7	0.0000	0.508	59	0.000	1.0000
6 - 7	0.0000	0.508	59	0.000	1.0000

temperature = 32, time = 7 days:

contrast	estimate	SE	df.	t.ratio	p.value
no prey - 5	0.4443	0.508	59	0.875	0.8176
no prey - 6	0.4176	0.508	59	0.823	0.8435
no prey - 7	0.6910	0.508	59	1.361	0.5284
5 - 6	-0.0267	0.508	59	-0.053	0.9999
5 - 7	0.2467	0.508	59	0.486	0.9619
6 - 7	0.2734	0.508	59	0.539	0.9492

P value adjustment: tukey method for comparing a family of 4 estimates