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Environmental insults and compensative responses

when microbiome meets cancer

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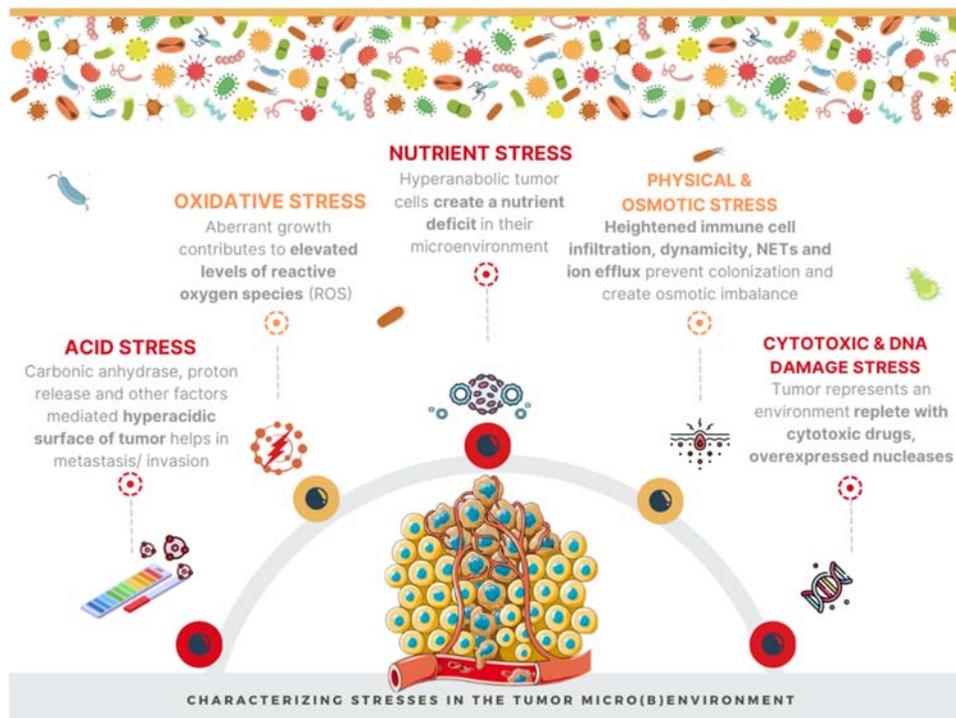
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23 **Abstract**

24 Tumor microenvironment has recently been ascribed a new hallmark – the polymorphic microbiome.
25 Accumulating evidence regarding the tissue specific territories of tumor-microbiome have opened
26 new and interesting avenues. A pertinent question is regarding the functional consequence of the
27 meeting of the host-microbiome with cancer. Given microbial communities have predominantly been
28 explored through an ecological perspective, it is important that the foundational aspects of ecological
29 stress and the fight to ‘survive and thrive’ are accounted for tumor-micro(b)environment as well.
30 Understanding the potential events leading to the synapse between the microbiome and the cancer,
31 and characterizing the subsequent environmental insults faced by the (infiltrating and intra-tumoral)
32 microbes is therefore important. Current work, building on existing evidence, aims to characterize
33 the ecological stresses and compensative responses of microbes to describe this underdiscussed
34 ecological interface between tumor and microbiota. It is hoped that a larger scientific thought on the
35 importance of microbial competition sensing vis-à-vis tumor-microenvironment would be stimulated.

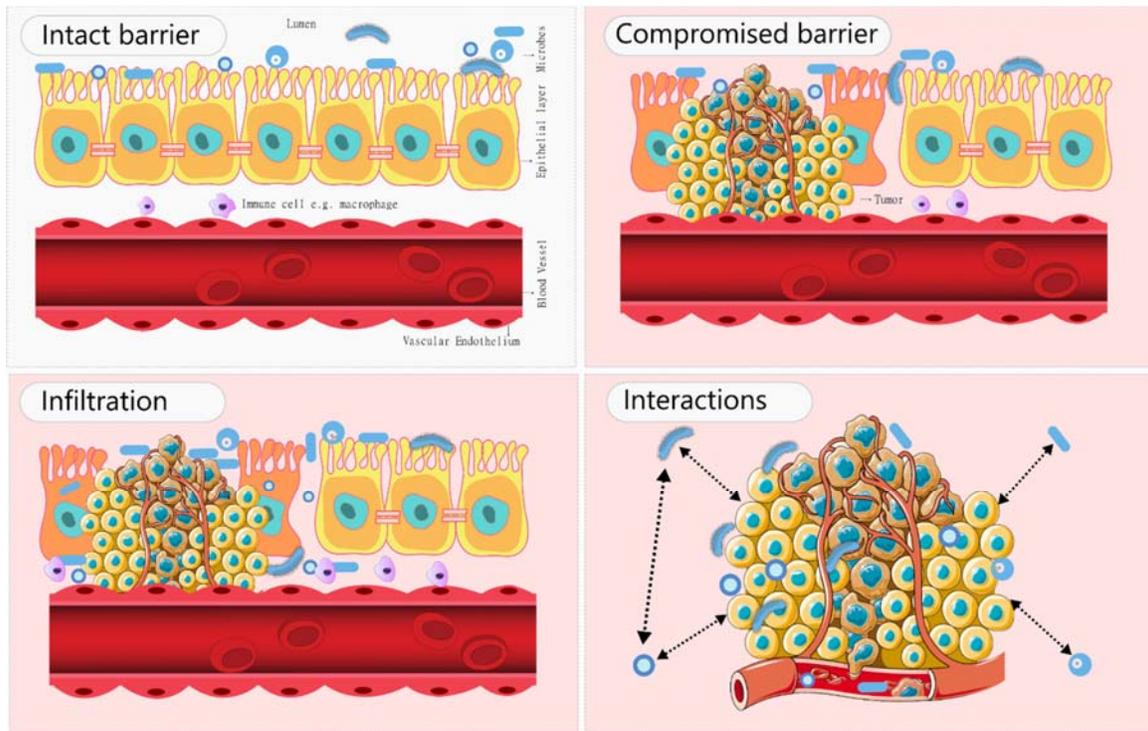


37 1. Introduction – the cancer-microbe interface

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39 Microbial association with oncopathology has been discussed for decades, with reports of anti-
40 cancerous activity of bacterial toxins dating back to a century ago ¹. Discovery of specific
41 microorganisms inside various tumors and their causal associations have consistently been reported
42 for past several decades ^{2,3}. However, it was not until recently that a successful and comprehensive
43 characterization of the microbiome associated with different human tumor types was achieved at a
44 large scale (amassing more than 1500 samples) ⁴. It laid the foundation for what may be termed as
45 the tissue specific territories of tumor microbiome. Importantly, the breakthrough quashed many
46 prevailing doubts pertaining to the contamination linked discoveries ^{4,5}. Several reports characterizing
47 the intratumoral microbiota have now emerged in the last 3 years alone, consolidating the existence
48 and importance of the tumor micro(b)environment ^{4,6-18}. Previously, reports of success in building an
49 onco-diagnostic tool using tissue and blood associated microbial-signatures in treatment-naive cancer
50 patients had also ignited interest towards looking into the sparse microbial content of the tumors ¹⁹.
51 While these pioneering studies provide guiding evidence towards differential microbial community
52 compositions in and around cancer cells ⁴ and preference of the microbes to inhabit microniches ¹⁸,
53 the functional models for tumor associated ‘communities of microbes’ warrant further research.

54 Success of colonization of tumors by microbes is expected to depend primarily on two factors
55 (i) an influx of the micro-organisms, and (ii) availability of conducive conditions for them to survive,
56 thrive and co-exist in the tumor microenvironment. While the influx can be driven by factors like
57 luminal infiltrations (Figure 1) through compromised epithelial/mucosal barrier ^{20,21}, inheritance from
58 normal adjacent tissues or NAT ⁴, zipper/trigger mechanisms of bacterial invasion ²² and circulatory
59 contributions from leaky vasculature of the tumor ^{23,24}, survival/thrival/co-existence is not only
60 dependent on the availability of favourable micro-niches in the tumor microenvironment ¹⁸ but also
61 on the activation of microbial stress responses against the perceived unfavourable ‘environmental
62 insults’ (including the inter/intraspecies competition).



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64 **Figure 1. Graphical representation of a scenario, showcasing events that can contribute to intratumoral**
65 **microbiome.** Once the host-microbiota enters the jungle of tumor micro-environment, its quest for surviving and thriving
66 begins (represented by microbe-tumor/microenvironment and microbe-microbe interactions).

67 2. The environmental insults inside the tumor microenvironment

68 Tumor microenvironment in fact offers several challenges/insults to the visiting microbes as
69 summarised in the graphical abstract and the Figure 2. These include –

70 2.1. Nutrient Stress

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Two key hallmarks of tumor are the hyperproliferation and hyperanabolism ²⁵. The unregulated proliferation leads to heightened energy and anabolic needs ^{25,26}. Consequently, the tumor-microenvironment is always nutrient deprived. While the adaptively programmed cancer cells are always hungry for glucose to utilize it ‘effectively and rapidly’ through the Warburg effect ^{26,27}, the oncogenic mutations generally lead to a heterogenous cancerous mass dependent on ‘not one but various limiting substrates’, leading to a continuous pressure on a variety of nutrients in the milieu of the tumor ²⁶. This is further aggravated in the Cancer stem

79 cells (CSCs) which represent a subpopulation in the tumor microenvironment, and are
80 undifferentiated and highly aggressive²⁸. The infiltrating and intratumor microorganisms are
81 therefore expected to encounter a perpetually hungry and aggressive competitor as soon as
82 they enter the tumor-microenvironment. How the visitors (microbes) would respond to this
83 nutrient stress, can potentially guide the development of meaningful functional models of the
84 tumor-micro(b)environment. Notably, the necrotic regions in the tumor however represent an
85 exception, offering a less competitive, nutrient rich hypoxic microniche for the growth and
86 proliferation of the microorganisms^{18,29}.

87 2.2. Oxidative stress

88 Reactive oxygen species (ROS), the free radicals, bearing unpaired reactive electron in their
89 valence shells, are normal byproducts of cellular respiration (oxidative phosphorylation).
90 Redox homeostasis is critical for maintaining a balance between the reactive oxygen species
91 (ROS) and antioxidants³⁰. This antioxidant-enzymes (e.g. superoxide dismutase or SOD)
92 mediated redox balance prevents the normal cells from cytotoxic damage and checks the
93 tumorigenic effects of ROS as well³⁰. The balance of redox homeostasis however doesn't
94 prevail in the tumor microenvironment which is replete with the ROS (the oxidative stress)
95 due to hyperproliferation, hyper-metabolism, mitochondrial dysfunction, infiltrating immune
96 cells, genetic (oncogenic) alterations, upregulated oxidases, peroxisome activity and among
97 more³¹. While primarily tumorigenic, ROS can inhibit tumors as well owing to their cytotoxic
98 nature^{30,31}. Cancer cells therefore employ adaptive metabolic modes of managing the high
99 ROS levels through NADPH accumulation, glutamine and folate metabolism etc³¹. The
100 incoming microorganisms would also need independent intrinsic mechanisms to fend this
101 insult off or perish due to the deleterious effects of free radicals on various macromolecules
102 (DNA, proteins, lipids and more), including an eventual cell death. The collateral impact of

103 said adaptive mechanisms on the tumor (microenvironment) would be interesting to probe
104 and understand.

105 2.3. Physical and Osmotic stress

106 Tumors are like wounds that never heal ³². Unlike normal tissues with a stable structure,
107 composition and biochemistry, tumor microenvironment is highly dynamic and unstable. This
108 dynamicity is attributed to the continuous angiogenesis, leaky vasculature, plasma
109 extravasation, a progression towards desmoplasia or solid tumors, among more ³².
110 Furthermore, the compressive stress faced by solid tumors while invading and navigating
111 through the normal adjacent tissue, causes increased intracellular tonicity (osmotic pressure),
112 triggering the upregulation of sodium efflux by tumors into the TME ³³. Consequently,
113 tumoral microbes are expected to face significant (i) *mechanical stress* due to the dynamic
114 spatio-temporal composition of tumor, preventing surface attachment or promoting
115 detachment, hence challenging the colonization of the TME and (ii) *osmotic stress* due to the
116 efflux of ions challenging microbial survival under the perturbed osmo-homeostasis. The
117 continuous infiltration of inflammatory and immune cells ^{32,34}, including macrophages and
118 neutrophils, in the never healing wounds of tumor, can further aggravate the physical stress
119 on the microbes seeking a firm attachment or colonization. A notable example of immune
120 surveillance mediated physical stress pertains to the expression of neutrophil extracellular
121 traps (NETs) in the tumor microenvironment ³⁵. NETs are extracellular complexes containing
122 fibres of decondensed chromatin (DNA), decorating protein granules, antimicrobial proteins
123 and histones used as a self-sacrificing defence mechanism (NETosis) by the neutrophils to
124 trap and kill invading microbial pathogens too large to engulf ³⁶. There are mixed evidence
125 towards the impact of NETs on tumors. Studies have indicated an anti-cancer role of NETs
126 through apoptosis, necrosis, ROS and H₂O₂ mediated cytotoxicity ³⁵. Evidence are also
127 accumulating that tumors are more inclined to leverage the NETs for proliferation and micro-

128 metastasis^{37,38}. It is however invariably well-founded that NETs function to inhibit or kill
129 invading microbes. The strategies adopted by microbes to adapt against or address these
130 environmental stresses interfering with colonization would therefore be additionally critical
131 in understanding the microbe-tumor interplay, especially from a spatio-temporal standpoint.

132 2.4. Acid stress

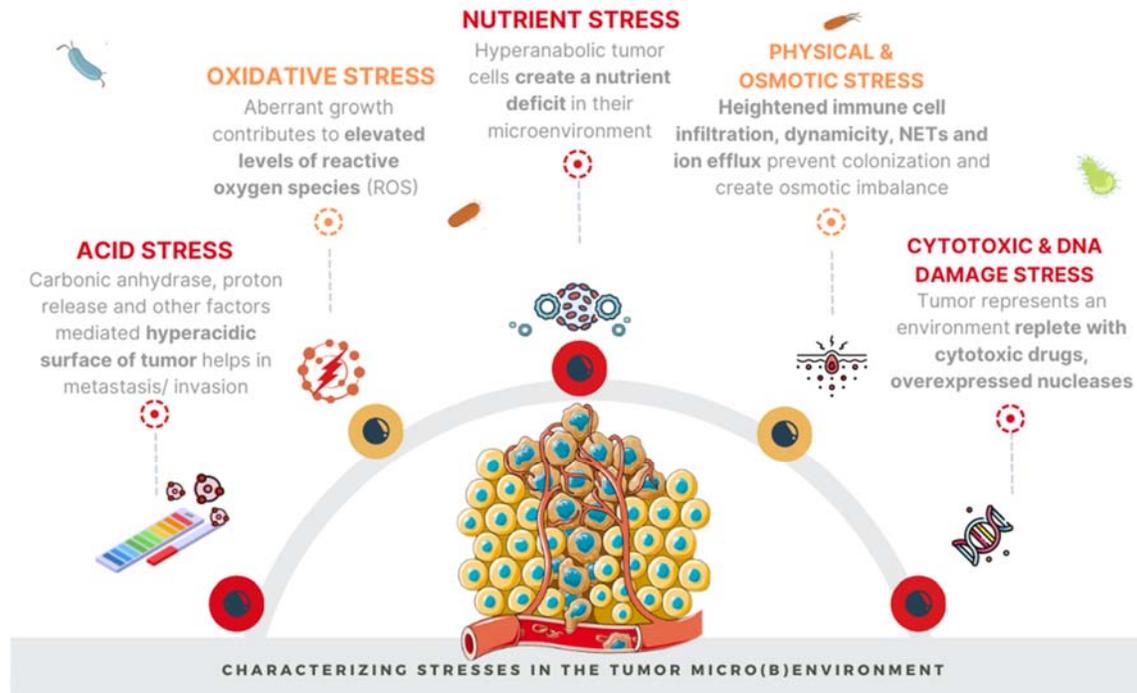
133 The Warburg-effect or the preference for glycolytic metabolism is known to lower the pH of
134 tumor-microenvironment^{27,39}. This is attributed to the rapid extrusion of accumulated lactate
135 to the extracellular environment. Additionally, the acidosis is also promoted by the
136 membrane-bound carbonic anhydrases through the release of protons while sequestering
137 carbon dioxide³⁹. Both these acidification promoting mechanisms are essentially ‘adaptive
138 responses’ of the cancer cells towards heightened energy needs (glycolytic metabolism) and
139 hypoxia (over expressed carbonic anhydrases). As a result, tumor-microenvironment exhibits
140 an inverted pH gradient ($\text{pH}_{\text{extracellular}} < \text{pH}_{\text{intracellular}}$), opposite to the normal tissues/cellular
141 environments, where extra-cellular pH is higher than the intracellular pH. An alkaline
142 intracellular pH helps tumors to continue proliferate and evade apoptosis within the
143 physiological pH range (7.2-7.4), while an acidic microenvironment (6.3-7.0) enables
144 activation of proteases and metastatic pathways, enabling cellular dispersion, immune-
145 evasion, drug-resistance, and invasion of healthy tissues³⁹. Given the heterogenous nature of
146 tumors, a stable pH gradient cannot be expected in the tumor-microenvironment. Moreover,
147 the steepness in the pH changes between the normal cellular environment and the tumor-
148 microenvironment can also be dictated by the biogeography of the host (e.g. normal
149 extracellular pH in: airway mucosa ~ 5.5-7.9, stomach ~ 1.5-3.5, colon: 6.1-7.5)^{40,41}. It would
150 be interesting to understand how the dynamic, slightly acidic pH environment of tumors can
151 affect the survival of the infiltrating microbes, which can have diverse pH sensitivities. The
152 acidosis driven dispersion/metastasis of cancer cells can additionally exert a physical stress

153 on the existing colonies or the microbes seeking a site of attachment ^{39,42}. Tumor-associated
154 pH gradients and associated heterogeneity can therefore potentially influence colonization and
155 subsequent interactions between the tumor and the microbiome, warranting further research.

156 2.5. Xenobiotic and DNA damage stress

157 In addition to the intrinsic hallmarks of cancer offering a variety of stresses to the visiting
158 microbiota, the extrinsic interventional regimens exert tremendous stress on the tumor, normal
159 tissues, and the native microbiome in and beyond tumor-microenvironment. Cytotoxic and
160 inhibitory effects of the xenobiotic chemotherapeutic agents on microbes, much of which are
161 attributed to the DNA damaging traits of these chemicals, are in fact well founded ^{43,44}. Given
162 that antibiotics have consistently been employed in many chemotherapies for their anti-cancer
163 properties, the DNA damaging/inhibitory/microbicidal action of the chemotherapeutic
164 regimens are rather expected ⁴⁵. Maier and colleagues however also demonstrated, through
165 in-vitro studies, the inhibitory effects of even the non-antibiotic chemotherapeutic agents on
166 well-known commensal microorganisms of the human gut ⁴⁶. It has also been recently proven
167 that even the conventional myelosuppressive chemotherapy disrupts intestinal microbiome ⁴⁷.
168 The heterogeneity added to the tumor-microenvironment by the (often) harsh therapeutic
169 regimens, is therefore expected to add to the insults faced by the visiting microbes.
170 Understanding the microbial response towards exposure to this stressful microenvironment
171 replete with the chemotherapeutic agents can not only (potentially) describe the ecological
172 basis of the consolidation of tumor-microbiome, but also the microbe-drug-tumor interplay.
173 Furthermore, microbial genetic material can also be stressed by the ROS (as described earlier)
174 and the pool of nucleases expressed in the tumor-microenvironment. Nucleases, the enzymes
175 that can hydrolyse nucleic acids, have consistently been perceived as promising biomarkers
176 for cancer. This is attributed to their frequently observed overexpression, with some reports
177 of interindividual variability, in the cancers of various types ⁴⁸. Nucleases however are also

178 critical towards establishing innate immunity against bacteria and viruses. This is achieved
179 through pattern recognition receptor (PRR) mediated pathways, which are aberrantly
180 expressed in tumors ⁴⁹. These nucleic acid degraders, ranging from exonucleases to
181 endonucleases, are known to be expressed intracellularly, extracellularly as well as ‘on the
182 membrane’ of cancer cells, marking their omnipresence in the tumor-microenvironment
183 (Yang 2011). While the functional significance of the largely overexpressed tumoral
184 nucleases remain to be fully understood, studies have associated the overexpression of
185 nucleases like Flap endonuclease1 (FEN1), Human apurinic/aprimidinic endonuclease1
186 (APE1), Excision repair cross-complementing group 1 xeroderma pigmentosum
187 complementation group F (ERCC1-XPF), Three prime repair exonuclease (TREX2), and
188 more with aggravated tumor growth and digressive response to chemotherapy (poor prognosis
189 and survival) ⁴⁸. Nucleases can also have bacterial origin, predominantly employed in the
190 bacterial warfare for survival in the competitive environments, targeting the non-self microbes
191 and host cells. Regardless of their origin, nucleases can target the genetic material and other
192 accessible nucleic acids of the tumoral microbiome, exposing them to heightened DNA
193 damage stress and immune surveillance. Microbial response to these multipronged stresses on
194 their genetic material is an important factor deserving attention, for an overall functional
195 understanding of tumor-microbiome’s response to its meeting with the cancer.



196

197 **Figure 2. Characterization of the key environmental insults offered by tumor-**
198 **microenvironment to the infiltrating/intratumoral microbes.** Nutrient stress, oxidative stress, acid stress,
199 physical & osmotic stress and DNA damaging/cytotoxic stress in combination are expected to offer significant and
200 persistent insults to the incoming/prevaling microbes in the tumor microenvironment.

201 3. Responding to the insults – microbial (counter) interactions

202 Microorganisms have evolved over billions of years to develop regulatory machineries for mitigating
203 the environmental stresses through well-orchestrated gene regulatory networks⁵¹. The stringent stress
204 response and the general stress response are two key well-founded hallmarks of the stress regulatory
205 responses in microbes^{51,52}. Depending upon the nature of stress ‘perceived’, as described in the
206 subsequent sections, microbes can switch to an appropriate response mechanism for survival.
207 Survival (and resilience) however is a function of ‘facilitation’ under a harsh environment and
208 ‘persistence’ through the complex intra/interspecies interactions (competition/cooperation)⁵³. This is
209 also described by Chesson in the species co-existence theory, attributing a stabilized community
210 structure to the influence of the environment on inter/intraspecies interactions including the
211 consequent tolerance of invaders/stabilized community to the mutual competition^{54,55}. The

212 competitive phenotypes of microbes broadly fall into two categories - (i) interference phenotypes and
213 (ii) exploitative phenotypes ^{56,57}. Interference competition occurs when the ability of a microbe to
214 survive or attain resources is directly thwarted by interfering phenotypes or antagonistic interactions
215 like chemical warfare and contact dependent-killing. Production of broad-spectrum antibiotics and
216 strain-specific bacteriocins to eliminate rival microorganisms is a typical example of this chemical
217 warfare mediated interference competition ^{56,58}. Exploitative competition on the other hand is an
218 indirect competition, experienced when microbes attempt to survive in a resource limited
219 environment among competitors with overlapping nutrient requirements ⁵⁶. This entails phenotypes
220 like secretion of nutrient-harvesting molecules (e.g. siderophores for iron sequestration), upregulation
221 of transport or uptake pathways, secretion of digestive proteases/nucleases and even secretion of
222 toxins like bacteriocins to specifically inhibit microorganisms with overlapping nutrient needs ^{52,56,57}.
223 An insight into the competition sensing mechanisms in the microorganisms in fact rationally indicates
224 that exploitative competition generates the interference competition between the microbes, with the
225 larger goal of ruling out any contest for the resources by adopting strategies which can inhibit,
226 displace, or kill the competitors ⁵². As Cornforth and Foster propose, an umbrella term of
227 “competition sensing” is less restrictive. It allows an emphasis on the ability of the microbes to sense
228 any harmful stimulus or stressor, perceiving its origins in potential competitors, self or non-self ⁵².
229 The suitability and strength of the response to the perceived stimuli would therefore dictate the fate
230 and function(s) of a microbial ecosystem. Given the heterogenous nature of tumor-microenvironment,
231 the dynamics governing the multi-species stress response and competition under the harsh/variable
232 environment of cancer ^{54,55} potentially hold an important key to understand tumor-microbe interplay.
233 Simply put, the balance of *‘the stress, the stress response and survival’* in the tumor
234 micro(b)environment can govern the dynamics of crosstalk between *‘the cancer and the microbes’*.
235 Notably though, despite the microbial stress response being defensive and compensative in nature, it
236 may not necessarily inhibit the cause of stress, i.e., cancer. This is unlike the response against

237 competing microorganisms, where one microbe or community tries to win against the other (the world
238 of microbe-kills-microbe). The composition of microbial community, density of the microbial
239 populations, tumor physiology, the nature and the quantum of the evoked microbial stress response
240 and the immunological response against microbial invasion is expected to decide the anti-tumor or
241 tumorigenic role of the tumor microbiome.

242 For simplicity in describing the overarching theme of this article (environmental insults and
243 compensative responses), bacterial ecology and stress response mechanisms will primarily be
244 emphasized in the subsequent sections. The terms ‘microbes and bacteria’ would therefore be used
245 interchangeably. Bacteria after all are prolifically studied microorganisms offering well founded and
246 valuable models for understanding microbial response to environmental stresses.

247 3.1. Doing collateral damage - Tumor targeting response of microbes

248 The stringent stress response (SSR) is an evolutionary conserved specific stress response
249 mechanism, mediated by the alarmone ‘guanosine tetraphosphate (ppGpp)’, that allows bacteria
250 to reprogram their transcriptional activities when faced with nutrient stress (particularly amino-
251 acid, fatty acid and iron limitations)^{59,60}. This entails a switch from translation and biosynthesis
252 to upregulated accumulation of limited resources^{52,59}. The state of nutrient stress offered by hyper
253 anabolic cancer cells, aggravated by the overlapping nutrient requirements of the tumoral
254 microbes, can evoke the SSR in the tumor-microbiota. This can reciprocate nutrient stress on
255 cancer, limiting its proliferation by competing for the nutrients critical for tumor progression,
256 particularly BCAA, acetate and iron⁶¹⁻⁶³. Ecologically, a quasi-exploitative competition between
257 the microbes sensing the competitive nutrient environment can elicit secretion of antimicrobial
258 peptides like bacteriocins and other antibiotics. These microbiome derived molecules, primarily
259 produced to fend off the perceived competition from the microbes with overlapping nutrient
260 requirements may potentially inhibit the cancer cells in collateral damage^{64,65}. A significantly
261 high production of colicins and microcins (anti-cancer bacteriocins) by mucosal microbiome in

262 CRC patients provides encouraging evidence in this regard ⁶⁶. The evidence pertaining to the
263 ability of bacteriocins to cross epithelial and vascular endothelial cells add to the plausibility of a
264 targeted response not only by the intra-tumoral microbes, but by the luminal, mucosal, NAT or
265 stromal microbiome as well ⁶⁷.

266 The presence of a global ‘General Stress Response (GSR)’ mechanism in bacteria, is however a
267 key weapon in their arsenal of defence against a broad range of environmental insults ⁶⁸. It is
268 mediated by the specialized transcriptional sigma (σ) factor(s) that compete with the house
269 keeping sigma factor to redirect transcription towards hundreds of prokaryotic stress response
270 genes, collectively called the general stress regulon. ^{68,69} Physio-biochemical stresses triggering
271 the expression of this regulon are rather well founded. These include bacterial exposure to nutrient
272 starvation, free radicals, heat, osmotic imbalance, acids, alcohols, membrane & DNA damaging
273 environmental stimuli and more that (threaten to) compromise the integrity/survival of a microbial
274 cell ⁶⁹. Given the association of GSR with a regulon consisting of hundreds of compensative
275 genes, the phenotypic output of this defence mechanism is multi-pronged and confers a broad
276 cross-resistance against a variety of rather unrelated stresses ⁶⁸. Accumulation of nutrients (e.g.
277 glycogen, amino acids, acetate, iron etc), shift to fermentation and biofilm formation, expression
278 of enzymes like catalases and oxidoreductases, accumulation or synthesis of osmoprotectants (e.g.
279 trehalose, amino acids, K^+), heightened expression of ‘amino acid decarboxylases, deaminases,
280 proton pumping, biofilm formation’ for acid tolerance are few classical examples of GSR
281 phenotypes ⁶⁸⁻⁷³. It is also pertinent to note the association of GSR with transition to the stationary
282 growth phase which is marked by a metabolic switch to the accumulation of inhibitory by-
283 products/secondary metabolites like antibiotics, toxins and even complex behaviours like biofilm
284 formation ⁶⁸.

285 The diverse environmental insults offered by tumor microenvironment to the visiting/thriving
286 microbes are expected to trigger the expression of aforementioned general stress regulon. This is

287 particularly true for nutrient and oxidative stress (abundantly prevailing in the TME) which are
 288 known to confer a broad cross-protectivity through the activation of general stress response ⁶⁹.
 289 Table 1, backed by literature evidence, is compiled to describe the the key GSR linked phenotypic
 290 outcomes that can (potentially) inflict a collateral reciprocation of insults on the cancer cells. The
 291 relevant tumorigenic/tumor-promoting outcomes of the said GSR expression are summarised in
 292 the Figure 3 and in the subsequent sections of this article.

GSR target	Phenotype	Mechanism of collateral damage for cancer	Reference
<i>Nutrient stress</i>	<ul style="list-style-type: none"> Accumulation of nutrients (e.g. glycogen, amino acids, acetate, iron etc) shift to fermentation biofilm formation 	<ul style="list-style-type: none"> Resource limitation for hyperanabolic cancer cells (cancers need glycogen, acetate, iron, BCAA etc) anti-mitotic role of SCFAs metastasis distraction by biofilms through secretion of exopolysaccharides, preventing cancer cell binding to the endothelial cells 	70,74–76
<i>Oxidative stress</i>	<ul style="list-style-type: none"> Expression of free radical scavenging enzymes like catalases, oxidoreductases, Superoxide dismutase Damage repairing proteins like thioredoxins, glutaredoxins, and methionine sulfoxide reductases 	<ul style="list-style-type: none"> Free radical clearance and release of damage repairing proteins limits DNA damage, inflammatory cytokines, metastasis, and oncogenic mutagenesis 	31,77
<i>Acid Stress</i>	<ul style="list-style-type: none"> expression of amino acid (Arginine and Glutamate) decarboxylases activation of Arginine deaminase system proton pumping increased glycolytic activity biofilm formation 	<ul style="list-style-type: none"> Cancer cells are arginine addict (deprivation leads to cancer cell death) Glutamate is a key substrate for cancer cells Proton release by intra-tumor microbes can disrupt pH of cancer cells Heightened microbial glycolytic activity and biofilm formation can compete for energy metabolism and prevent metastasis 	26,39,78

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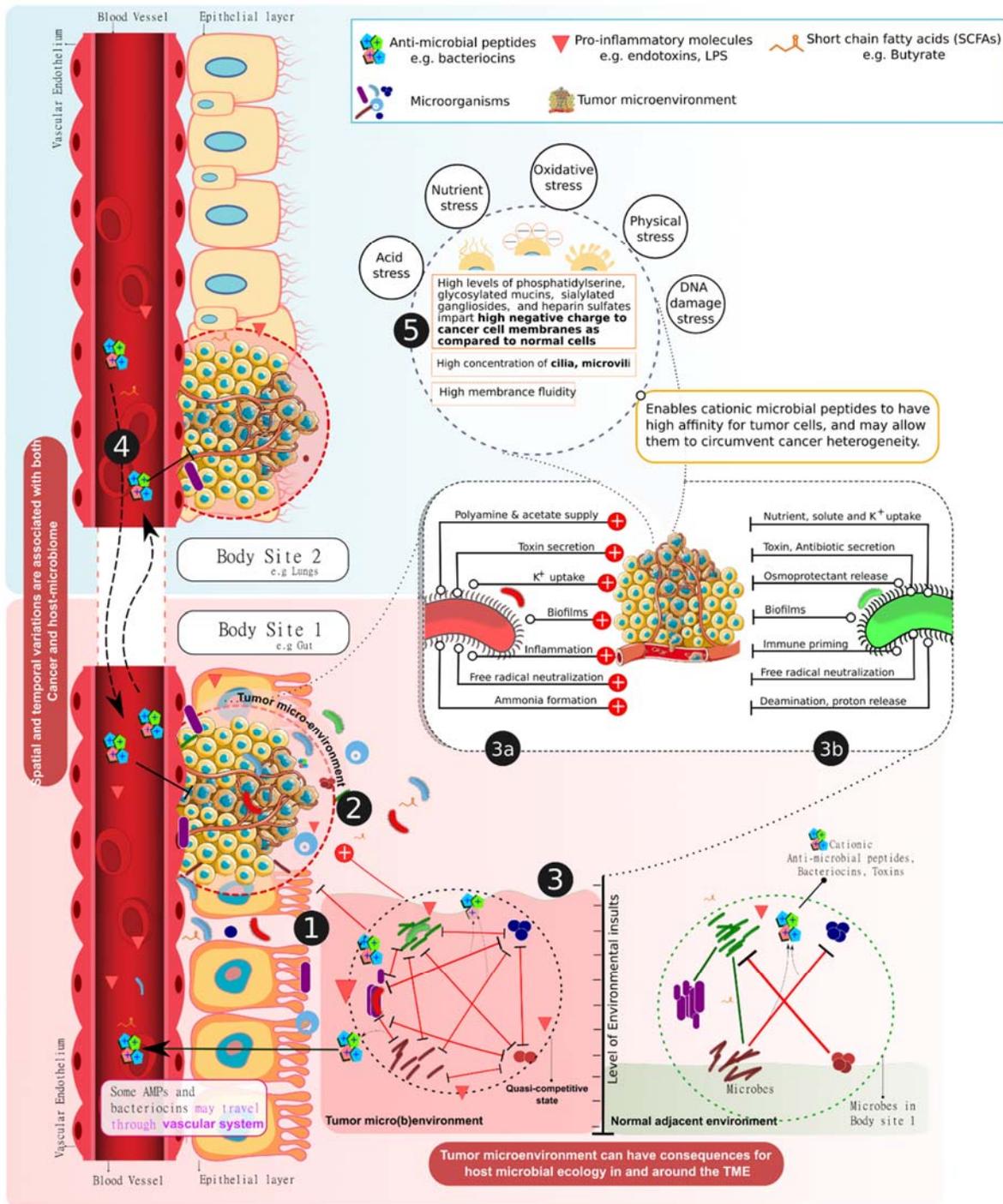
<p>Physical Stress</p>	<ul style="list-style-type: none"> • upregulated MSCRAMMs* and biofilm formation • expression of autolysin like enzymes and release of eDNA*, teichoic acid and other cytoplasmic contents • upregulation of virulence factors like surface endonucleases 	<ul style="list-style-type: none"> • MSCRAMMs mediate covalent binding leading to persistent biofilms that can compete for nutrition and arrest metastasis • eDNA and teichoic acids can mediate non-covalent binding of microbes to cancer, and can also trigger immune surveillance for collateral recognition of cancer cells • Degradation of NETs* and other entrapments by surface endonucleases can prevent metastasis 	<p>36,37,42,79,80</p>
<p>Osmotic stress</p>	<ul style="list-style-type: none"> • Solute uptake including amino acids, potassium ions (K⁺) • Synthesis and accumulation of Trehalose 	<ul style="list-style-type: none"> • Uptake of amino acids as solutes can limit cancer energy metabolism • Trehalose released through mechanosensitive channels and upon bacterial lysis can reduce inflammation, limit free radicals, enhance apoptosis • Uptake of the storm of K⁺ ions released by dying cancer cells can limit suppression of cancer killing T-cell effector function 	<p>61,81,82</p>
<p>DNA damage</p>	<p>SOS response upregulates –</p> <ul style="list-style-type: none"> • biofilms with (drug resistant) persister population • Intraspecies competition and consequent toxin secretion • Toxin-anti toxin (TA) system activation • Horizontal gene transfer 	<ul style="list-style-type: none"> • Persistent biofilms can compete for energy metabolism and prevent metastasis • Toxins against intraspecies competition (e.g. colicins) can inhibit cancers • TA systems can specifically cause cancer cell death (e.g. MazF-MazE toxin–antitoxin of E.coli against pancreatic and colorectal cancers) • Anti-cancer toxins/antibiotics encoded by plasmids can promote population level phenotype through HGT 	<p>52,64,83–86</p>
<p>* <i>MSCRAMM: Microbial Surface Components Recognizing Adhesive Matrix Molecules</i> are microbial surface proteins that adhere specifically to host extra-cellular matrix (ECM); eDNA: extracellular DNA; NETs: Neutrophil Extracellular Traps</p>			

293 **Table 1.** Potential responses of tumor invading/inhabiting microbiota mediated by the expression of GSR regulon under diverse
 294 environmental insults of the TME. Notably, the GSR targets can be ameliorated by any stressor that can activate entire general stress
 295 regulon (conferring cross protectivity).

296 Under the right tumor microbial composition (native or interventional), this GSR and stationary
 297 phase linked in-vivo production of compensative products may even support cancer-therapy by

298 priming the onco-immune system towards anti-tumor effects. The reported role of intra-tumoral
299 probiotic gut-microbes in facilitating immunotherapy through the secondary metabolite mediated
300 triggering of the STING signalling (stimulator of interferon genes), highlights this significance of
301 tumoral colonization by commensal bacteria like *Bifidobacterium sp.* ⁸⁷.

302 From an ecological point of view, insults like oxidative stress, DNA damage stress, physical
303 stress and acid stress are perceived as instances of direct challenges interfering with the ability of
304 the microbes to survive and thrive. This calls for an activation of interference competitive
305 phenotype and hence release of antibiotics and strain-specific bacteriocins towards the microbe-
306 kill-microbe response ⁵⁶. The collateral damage inflicted on the cancer cells by this chemical
307 warfare started by microbes under the perceived interference competition is plausible and
308 therefore deserves exploration. The molecular mechanistic details underpinning this warfare may
309 be described by the evolutionary matured stress response mechanisms as described earlier. In
310 addition to the development of functional models, this would be important for the design of live
311 biotherapeutics or dietary interventions aiming to favourably customize the microbial and
312 metabolite composition of tumor invading/prevaling microbiota. **Figure 3** provides a graphical
313 overview of the (aforementioned) events that may ensue in tumor micro(b)environment.



314

315 **Figure 3. Microbial quest for survival in the tumor affected ecosystem.**

316 Microbes and microbial products may infiltrate to tumors through dysfunctional epithelial barrier, adjacent tissues or

317 circulatory system. (2) Stressful tumor environment can trigger Stringent and general stress response in microbes. (3)

318 Environmental insults can lead to quasi-exploitative and quasi-interference competition between tumor microbes. (3a)

319 Competitive environment and resultant stress response manifests in the form of upregulation of nutrient and ion uptake,

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320 synthesis of anti-microbial peptides/toxins, shift to fermentation, biofilm formation, redox balance and more causing
321 collateral damage to cancer. (3b) Microbial responses can be oncogenic/promoting too (e.g. toxin secretion, polyamine
322 metabolism, ammonia formation, inflammatory LPS, lowered oxidative stress on tumor, potassium influx). (4) Microbial
323 metabolites including toxins and AMPs can access circulatory system for potential systemic effects. (5) Properties of
324 cancer cell membranes can enable targeted attack by cationic anti-microbial peptides

325 3.2. Into the wounds that never heal - Tumor promoting response of microbes

326

327 While Rudolph Virchow first linked chronic inflammation with tumor development ⁸⁸, Harold
328 Dvorak's comparison of tumors with the 'Wounds that never heal' notified similarities between
329 tumor stroma generation (essential for tumor growth) and wound healing ³². Microbial invasion
330 of these wounds can spur the inflammation process ⁸⁹, supporting the tumor elicited inflammation
331 characterised by an accelerated recruitment of immune cells and up-regulation of pro-
332 inflammatory cytokines and growth factors ⁸⁹⁻⁹². This can not only promote tumor progression
333 but also aggravate the associated adverse symptoms. Notably, in-addition to the immune-
334 regulating components of microbial anatomy like flagellin and lipopolysaccharide (LPS), the
335 secondary metabolic products of microbial stress response like toxins (e.g. colibactin, endotoxin)
336 can be pro-inflammatory and oncogenic ^{3,89,90,93}. These, as interjected earlier, are expected to be
337 elicited in response to the diverse environmental insults faced by the invading microorganisms
338 (Table 1).

339 The responses controlled by the general stress regulon may additionally support tumor
340 progression (Figure 3). This includes - (i) the neutralization of oxidative stress by microbes in
341 the tumor microenvironment, thereby lowering the compensative load on tumor cells which are
342 also sensitive to redox imbalance ^{31,77} (ii) acid stress management by microbial urease system
343 leading to the formation of normally cytotoxic, proinflammatory but a potent nitrogen-reservoir
344 for cancer cells - ammonia ^{73,94} (iii) influx of potassium ions upon activation of osmotic stress
345 response in the microbes, lowering intracellular tonicity of tumors and limiting T-cell stemness
346 that enables cancer clearance ^{33,95,96} and (iv) the reported role of stress resilient bacterial biofilms,

347 a phenotypic response expected against nutrient, physical, DNA damage and acid stress, in
348 initiation and progression of cancer through polyamine metabolism, toxin secretion and other well
349 founded pro-oncogenic responses is worth consideration as well ⁹⁷. Furthermore, the fermentative
350 state of microbial growth under anoxic and nutrient depleted environment of tumors and normal
351 adjacent tissue (e.g. gut epithelium and lumen) may contribute acetate (the most abundant SCFA),
352 which, even though is reported for its anticancer potential, is also a key energy molecule for
353 proliferating cancer cells ⁶².

354 Unsurprisingly, the molecular basis of ecological interactions of tumor prevailing/invading
355 microbes with the potentially insulting environmental conditions dictate that the meeting of
356 microbes with cancer can have both deleterious and advantageous consequences for the tumor.
357 Where the balance would weigh more, can only be determined by the stabilized (or tweaked)
358 microbial population and its functional potential. It is therefore important, as we next discuss, to
359 ponder over the directions that can branch out of this school of thought and potential limitations
360 in assuming the native microbial populations of tumors, including any microbe-tumor cross-talk.

361 4. Future directions and limitations

362
363 Human body essentially serves as an ecosystem to the colonizing microbes. The organ and tissue
364 specific (spatio-temporal) territories of host microbiome are governed by the myriad of physiological,
365 physical, metabolic and nutritional conditions specific to the sites of microbial colonization. Tumor
366 development needs to be viewed as an ecological disturbance and its micro-environment as a
367 perturbed niche capable of reshaping the structure of individual microbial populations through
368 systemic and localized environmental pressures. How prevailing microbiota responds and survives
369 against the ecological stresses offered by tumor development/progression is expected to drive the
370 compositional and metabolic variations observed in different individuals, across different types of
371 tumors. Such an understanding is critical to drive the development of in-silico models of tumor
372 micro(b)environment through due attention to the dynamics of underlying metabolic fluxes and multi-

373 species interactions ('host-microbe, tumor-microbe, microbe-microbe and even tumor-tumor'). A
374 functional gradation and classification of key microbial players (e.g. drivers, passengers) identified
375 inside the tumor micro-environment may enable validation of the well founded driver-passenger
376 models of various types of cancer ^{98,99}. Importantly, the functional understanding of microbial
377 response to tumor micro-environment can aid development of therapeutic regimens aimed at
378 modulating microbial populations and function thereof inside and around the cancer. This includes,
379 but not limited to the probiotic and prebiotic formulations that can assist an accelerated reshaping of
380 host and tumor microbiome towards an 'anti-cancer' community ^{66,100,101}.

381 Cancer however is a complex disease characterised not only by abnormally dividing hyper-
382 anabolic cells, unique micro-environment and location or site-specific manifestations but
383 multifactorial confounders like specialized care and aggressive therapeutic regimens (e.g.
384 chemotherapy, radiotherapy) etc ¹⁰²⁻¹⁰⁴. This milieu of confounding factors can significantly impact
385 the systemic as well as the localized host microbial ecology which may not overlap with the expected
386 or characteristic response of microbes inside and in vicinity of a treatment-naive tumor environment.
387 Additionally, the personalized nature of host microbiome, governed by spatio-temporal dynamics adds
388 to the complexity of factors that need to be accommodated for arriving at in-silico models or
389 translatable interventions. The systemic implications of surgical (like Ostomy) and case-dependent
390 dosages and durations of invasive therapeutic regimens like radiation or chemotherapy only add to
391 the associated complications of the disease and its ecosystem ^{102,105,106}. Worth consideration are the
392 challenges associated with reproducing the results of microbiome studies (the reproducibility crisis),
393 especially considering the compositionally sparse microbiota of tumor ⁴. Given the extremely low
394 microbial load of tumor associated samples, contaminants become an additional and key bottleneck
395 to address against innumerable sources of contamination throughout the lengthy workflow of
396 microbiome study. Nevertheless, under all these variables, microbial response to stimuli is a

397 significant constant and deserves attention for any research that intends to decipher the functional
398 models of cancer micro(b)environment.

399 5. Conclusion

400
401 Surviving and thriving are key to organismal existence in the living world, microbes are no exception.
402 Appreciating the challenges associated with colonization of an environment as complex and
403 heterogeneous as tumor and linking them with what is well founded in microbial ecology can drive
404 foundational understanding of microbial role in modulating the tumor microenvironment. Here an
405 effort was made to characterize the relevant stresses in the tumor microenvironment that may serve
406 as insults compromising the colonization and survival of microbes in the harsh environment of the
407 tumors. Upon revisiting the classical evidence of microbial ecology/competition and stress response,
408 it becomes encouragingly clear that collateral impact of microbial compensative responses to the
409 consistent insults of the TME could hold an important key for developing functional models of tumor-
410 microbe interaction. The success of various dietary regimens and microbial interventions (e.g.
411 pre/probiotics), that attempt to channelize the host-microbial arsenal for cancer prevention or
412 treatment may after all have roots in the basic concept of microbial competition sensing and their
413 response to the environmental stimuli^{52,107,108}. Understanding such stimuli in tumor
414 micro(b)environment and microbial responses to the same, is critical to throw light on what happens
415 (and can happen), when microbiota meets cancer.

416

417

418

419

420

421 **Abbreviations**

422 **NAT:** Normal Adjacent Tissue; **GSR:** General Stress Response/Regulon; **SSR:** Stringent Stress
423 Response; **BCAA:** Branched Chain Amino Acids; **SCFA:** Short Chain Fatty Acids; **STING:**
424 Stimulator of Interferon Genes; **LPS:** Lipopolysaccharide

425

426 **Declaration**

427 No part of this article was written or designed with the aid of any automated or generative tool like
428 ChatGPT or DALL.E. All ideas and content of this work are authors' own work, including a sincere
429 effort to manually go through various research articles that served as reference or evidence towards
430 building the presented perspective(s).

431

432 **Author contribution**

433 Conceived the idea: SN; Manuscript draft and Figures' design: SN; Supervision: SSM; Proofreading:
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435

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441

442 **Competing interests**

443 Authors are salaried research Scientists at TCS Research. TCS holds a portfolio of patents in
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447

448 **References**

449

- 450 1. Coley, W. B. The Treatment of Inoperable Sarcoma by Bacterial Toxins (the Mixed Toxins
451 of the Streptococcus erysipelas and the Bacillus prodigiosus). *Proc R Soc Med* **3**, (1910).
- 452 2. Parida, S. & Sharma, D. The microbiome and cancer: Creating friendly neighborhoods and
453 removing the foes with in A C. *Cancer Research* vol. 81 Preprint at
454 <https://doi.org/10.1158/0008-5472.CAN-20-2629> (2021).
- 455 3. Wong-Rolle, A., Wei, H. K., Zhao, C. & Jin, C. Unexpected guests in the tumor
456 microenvironment: microbiome in cancer. *Protein and Cell* vol. 12 Preprint at
457 <https://doi.org/10.1007/s13238-020-00813-8> (2021).
- 458 4. Nejman, D. *et al.* The human tumor microbiome is composed of tumor type-specific
459 intracellular bacteria. *Science (1979)* **368**, (2020).
- 460 5. Robinson, K. M., Crabtree, J., Mattick, J. S. A., Anderson, K. E. & Hotopp, J. C. D.
461 Distinguishing potential bacteria-tumor associations from contamination in a secondary data
462 analysis of public cancer genome sequence data. *Microbiome* **5**, (2017).
- 463 6. Atreya, C. E. & Turnbaugh, P. J. Probing the tumor micro(b)environment. *Science (1979)*
464 **368**, (2020).
- 465 7. Choi, J. K. *et al.* Cross-talk between cancer and *Pseudomonas aeruginosa* mediates tumor
466 suppression. *Commun Biol* **6**, 16 (2023).
- 467 8. Yuan, L. *et al.* Tumor microbiome diversity influences papillary thyroid cancer invasion.
468 *Commun Biol* **5**, 864 (2022).
- 469 9. Hermida, L. C., Gertz, E. M. & Ruppin, E. Analyzing the tumor microbiome to predict
470 cancer patient survival and drug response. *Cancer Res* **81**, (2021).
- 471 10. Feng, Z. *et al.* In situ imaging for tumor microbiome interactions via imaging mass
472 cytometry on single-cell level. *Cytometry Part A* **101**, (2022).
- 473 11. Thyagarajan, S. *et al.* Comparative analysis of racial differences in breast tumor microbiome.
474 *Sci Rep* **10**, (2020).
- 475 12. Murphy, C. L. *et al.* Mapping the colorectal tumor microbiota. *Gut Microbes* **13**, (2021).
- 476 13. Okuda, S. *et al.* Profiling of host genetic alterations and intra-tumor microbiomes in
477 colorectal cancer. *Comput Struct Biotechnol J* **19**, (2021).
- 478 14. Bahig, H. *et al.* Longitudinal characterization of the tumoral microbiome during radiotherapy
479 in HPV-associated oropharynx cancer. *Clin Transl Radiat Oncol* **26**, (2021).
- 480 15. Livyatan, I., Nejman, D., Shental, N. & Straussman, R. Characterization of the human tumor
481 microbiome reveals tumor-type specific intra-cellular bacteria. *OncImmunity* vol. 9
482 Preprint at <https://doi.org/10.1080/2162402X.2020.1800957> (2020).
- 483 16. Guo, W. *et al.* Tumor microbiome contributes to an aggressive phenotype in the basal-like
484 subtype of pancreatic cancer. *Commun Biol* **4**, (2021).
- 485 17. Zwinsová, B. *et al.* Colorectal tumour mucosa microbiome is enriched in oral pathogens and
486 defines three subtypes that correlate with markers of tumour progression. *Cancers (Basel)* **13**,
487 (2021).
- 488 18. Niño, J. L. G. *et al.* Effect of the intratumoral microbiota on spatial and cellular heterogeneity
489 in cancer. *Nature* (2022) doi:10.1038/s41586-022-05435-0.

- 490 19. Poore, G. D. *et al.* Microbiome analyses of blood and tissues suggest cancer diagnostic
491 approach. *Nature* **579**, (2020).
- 492 20. Mullin, J. M. Epithelial barriers, compartmentation, and cancer. *Science's STKE: signal*
493 *transduction knowledge environment* vol. 2004 Preprint at
494 <https://doi.org/10.1126/stke.2162004pe2> (2004).
- 495 21. Soler, A. P. *et al.* Increased tight junctional permeability is associated with the development
496 of colon cancer. *Carcinogenesis* **20**, (1999).
- 497 22. O Cróinín, T. & Backert, S. Host epithelial cell invasion by *Campylobacter jejuni*: trigger or
498 zipper mechanism? *Frontiers in cellular and infection microbiology* vol. 2 Preprint at
499 <https://doi.org/10.3389/fcimb.2012.00025> (2012).
- 500 23. Cummins, J. & Tangney, M. Bacteria and tumours: Causative agents or opportunistic
501 inhabitants? *Infectious Agents and Cancer* vol. 8 Preprint at [https://doi.org/10.1186/1750-](https://doi.org/10.1186/1750-9378-8-11)
502 [9378-8-11](https://doi.org/10.1186/1750-9378-8-11) (2013).
- 503 24. Hashizume, H. *et al.* Openings between defective endothelial cells explain tumor vessel
504 leakiness. *American Journal of Pathology* **156**, (2000).
- 505 25. Hanahan, D. Hallmarks of Cancer: New Dimensions. *Cancer Discovery* vol. 12 Preprint at
506 <https://doi.org/10.1158/2159-8290.CD-21-1059> (2022).
- 507 26. Sullivan, M. R. & vander Heiden, M. G. Determinants of nutrient limitation in cancer.
508 *Critical Reviews in Biochemistry and Molecular Biology* vol. 54 Preprint at
509 <https://doi.org/10.1080/10409238.2019.1611733> (2019).
- 510 27. Liberti, M. v. & Locasale, J. W. The Warburg Effect: How Does it Benefit Cancer Cells?
511 *Trends in Biochemical Sciences* vol. 41 Preprint at <https://doi.org/10.1016/j.tibs.2015.12.001>
512 (2016).
- 513 28. Yadav, U. P. *et al.* Metabolic Adaptations in Cancer Stem Cells. *Frontiers in Oncology* vol.
514 10 Preprint at <https://doi.org/10.3389/fonc.2020.01010> (2020).
- 515 29. Zhou, S., Gravekamp, C., Bermudes, D. & Liu, K. Tumour-targeting bacteria engineered to
516 fight cancer. *Nature Reviews Cancer* vol. 18 Preprint at [https://doi.org/10.1038/s41568-018-](https://doi.org/10.1038/s41568-018-0070-z)
517 [0070-z](https://doi.org/10.1038/s41568-018-0070-z) (2018).
- 518 30. Valko, M. *et al.* Free radicals and antioxidants in normal physiological functions and human
519 disease. *International Journal of Biochemistry and Cell Biology* vol. 39 Preprint at
520 <https://doi.org/10.1016/j.biocel.2006.07.001> (2007).
- 521 31. Hayes, J. D., Dinkova-Kostova, A. T. & Tew, K. D. Oxidative Stress in Cancer. *Cancer Cell*
522 vol. 38 Preprint at <https://doi.org/10.1016/j.ccell.2020.06.001> (2020).
- 523 32. Dvorak, H. F. Tumors: Wounds that do not heal-redux. *Cancer Immunol Res* **3**, (2015).
- 524 33. McGrail, D. J. *et al.* Osmotic Regulation Is Required for Cancer Cell Survival under Solid
525 Stress. *Biophys J* **109**, (2015).
- 526 34. Flier, J. S., Underhill, L. H. & Dvorak, H. F. Tumors: Wounds That Do Not Heal. *New*
527 *England Journal of Medicine* **315**, (1986).
- 528 35. de Meo, M. L. & Spicer, J. D. The role of neutrophil extracellular traps in cancer progression
529 and metastasis. *Seminars in Immunology* vol. 57 Preprint at
530 <https://doi.org/10.1016/j.smim.2022.101595> (2021).
- 531 36. Brinkmann, V. *et al.* Neutrophil Extracellular Traps Kill Bacteria. *Science (1979)* **303**,
532 (2004).
- 533 37. Yang, L. *et al.* DNA of neutrophil extracellular traps promotes cancer metastasis via
534 CCDC25. *Nature* **583**, (2020).
- 535 38. Masucci, M. T., Minopoli, M., del Vecchio, S. & Carriero, M. V. The Emerging Role of
536 Neutrophil Extracellular Traps (NETs) in Tumor Progression and Metastasis. *Frontiers in*
537 *Immunology* vol. 11 Preprint at <https://doi.org/10.3389/fimmu.2020.01749> (2020).

- 538 39. Lee, S. H. & Griffiths, J. R. How and why are cancers acidic? Carbonic anhydrase ix and the
539 homeostatic control of tumour extracellular ph. *Cancers* vol. 12 Preprint at
540 <https://doi.org/10.3390/cancers12061616> (2020).
- 541 40. Evans, D. F. *et al.* Measurement of gastrointestinal pH profiles in normal ambulant human
542 subjects. *Gut* **29**, (1988).
- 543 41. Fischer, H. & Widdicombe, J. H. Mechanisms of acid and base secretion by the airway
544 epithelium. *Journal of Membrane Biology* vol. 211 Preprint at
545 <https://doi.org/10.1007/s00232-006-0861-0> (2006).
- 546 42. Otto, M. Physical stress and bacterial colonization. *FEMS Microbiology Reviews* vol. 38
547 Preprint at <https://doi.org/10.1111/1574-6976.12088> (2014).
- 548 43. Maurice, C. F., Haiser, H. J. & Turnbaugh, P. J. Xenobiotics shape the physiology and gene
549 expression of the active human gut microbiome. *Cell* **152**, (2013).
- 550 44. Johnson, N. P., Razaka, H., Wimmer, F., Defais, M. & Villani, G. Toxicity, mutagenicity and
551 drug resistance in *Escherichia coli* treated with platinum antitumor compounds. *Inorganica*
552 *Chim Acta* **137**, (1987).
- 553 45. Shapiro, R. S. Antimicrobial-Induced DNA Damage and Genomic Instability in Microbial
554 Pathogens. *PLoS Pathogens* vol. 11 Preprint at <https://doi.org/10.1371/journal.ppat.1004678>
555 (2015).
- 556 46. Maier, L. *et al.* Extensive impact of non-antibiotic drugs on human gut bacteria. *Nature* **555**,
557 (2018).
- 558 47. Papanicolas, L. E. *et al.* Conventional myelosuppressive chemotherapy for non-
559 haematological malignancy disrupts the intestinal microbiome. *BMC Cancer* **21**, (2021).
- 560 48. Balian, A. & Hernandez, F. J. Nucleases as molecular targets for cancer diagnosis.
561 *Biomarker Research* vol. 9 Preprint at <https://doi.org/10.1186/s40364-021-00342-4> (2021).
- 562 49. Nagi, R. S., Bhat, A. S. & Kumar, H. Cancer: A tale of aberrant PRR response. *Front*
563 *Immunol* **5**, (2014).
- 564 50. Yang, W. Nucleases: Diversity of structure, function and mechanism. *Q Rev Biophys* **44**,
565 (2011).
- 566 51. Foster, P. L. Stress responses and genetic variation in bacteria. *Mutation Research -*
567 *Fundamental and Molecular Mechanisms of Mutagenesis* vol. 569 Preprint at
568 <https://doi.org/10.1016/j.mrfmmm.2004.07.017> (2005).
- 569 52. Cornforth, D. M. & Foster, K. R. Competition sensing: The social side of bacterial stress
570 responses. *Nature Reviews Microbiology* vol. 11 Preprint at
571 <https://doi.org/10.1038/nrmicro2977> (2013).
- 572 53. Hart, S. P. & Marshall, D. J. Environmental stress, facilitation, competition, and coexistence.
573 *Ecology* **94**, (2013).
- 574 54. Chesson, P. Multispecies Competition in Variable Environments. *Theor Popul Biol* **45**,
575 (1994).
- 576 55. Chesson, P. Updates on mechanisms of maintenance of species diversity. *Journal of Ecology*
577 **106**, (2018).
- 578 56. Hibbing, M. E., Fuqua, C., Parsek, M. R. & Peterson, S. B. Bacterial competition: Surviving
579 and thriving in the microbial jungle. *Nature Reviews Microbiology* vol. 8 Preprint at
580 <https://doi.org/10.1038/nrmicro2259> (2010).
- 581 57. Ghoul, M. & Mitri, S. The Ecology and Evolution of Microbial Competition. *Trends in*
582 *Microbiology* vol. 24 Preprint at <https://doi.org/10.1016/j.tim.2016.06.011> (2016).
- 583 58. Bauer, M. A., Kainz, K., Carmona-Gutierrez, D. & Madeo, F. Microbial wars: Competition
584 in ecological niches and within the microbiome. *Microbial Cell* vol. 5 Preprint at
585 <https://doi.org/10.15698/mic2018.05.628> (2018).

- 586 59. Irving, S. E., Choudhury, N. R. & Corrigan, R. M. The stringent response and physiological
587 roles of (pp)pGpp in bacteria. *Nature Reviews Microbiology* vol. 19 Preprint at
588 <https://doi.org/10.1038/s41579-020-00470-y> (2021).
- 589 60. Boutte, C. C. & Crosson, S. Bacterial lifestyle shapes stringent response activation. *Trends in*
590 *Microbiology* vol. 21 Preprint at <https://doi.org/10.1016/j.tim.2013.01.002> (2013).
- 591 61. Sivanand, S. & vander Heiden, M. G. Emerging Roles for Branched-Chain Amino Acid
592 Metabolism in Cancer. *Cancer Cell* vol. 37 Preprint at
593 <https://doi.org/10.1016/j.ccell.2019.12.011> (2020).
- 594 62. Schug, Z. T., vande Voorde, J. & Gottlieb, E. The metabolic fate of acetate in cancer. *Nature*
595 *Reviews Cancer* vol. 16 Preprint at <https://doi.org/10.1038/nrc.2016.87> (2016).
- 596 63. Manz, D. H., Blanchette, N. L., Paul, B. T., Torti, F. M. & Torti, S. v. Iron and cancer:
597 Recent insights. *Ann N Y Acad Sci* **1368**, (2016).
- 598 64. Karpiński, T. M. & Adameczak, A. Anticancer activity of bacterial proteins and peptides.
599 *Pharmaceutics* vol. 10 Preprint at <https://doi.org/10.3390/pharmaceutics10020054> (2018).
- 600 65. Dobson, A., Cotter, P. D., Paul Ross, R. & Hill, C. Bacteriocin production: A probiotic trait?
601 *Applied and Environmental Microbiology* vol. 78 Preprint at
602 <https://doi.org/10.1128/AEM.05576-11> (2012).
- 603 66. Kohoutova, D. *et al.* Bacteriocin production by mucosal bacteria in current and previous
604 colorectal neoplasia. *BMC Cancer* **20**, (2020).
- 605 67. Dreyer, L., Smith, C., Deane, S. M., Dicks, L. M. T. & van Staden, A. D. Migration of
606 Bacteriocins Across Gastrointestinal Epithelial and Vascular Endothelial Cells, as
607 Determined Using In Vitro Simulations. *Sci Rep* **9**, (2019).
- 608 68. Gottesman, S. Trouble is coming: Signaling pathways that regulate general stress responses
609 in bacteria. *Journal of Biological Chemistry* vol. 294 Preprint at
610 <https://doi.org/10.1074/jbc.REV119.005593> (2019).
- 611 69. Boor, K. J. Bacterial stress responses: What doesn't kill them can make them stronger. *PLoS*
612 *Biology* vol. 4 Preprint at <https://doi.org/10.1371/journal.pbio.0040023> (2006).
- 613 70. Dauer, P. & Lengyel, E. New Roles for Glycogen in Tumor Progression. *Trends in Cancer*
614 vol. 5 Preprint at <https://doi.org/10.1016/j.trecan.2019.05.003> (2019).
- 615 71. Gottschlich, L., Geiser, P., Bortfeld-Miller, M., Field, C. M. & Vorholt, J. A. Complex
616 general stress response regulation in *Sphingomonas melonis* Fr1 revealed by transcriptional
617 analyses. *Sci Rep* **9**, (2019).
- 618 72. Bearson, S., Bearson, B. & Foster, J. W. Acid stress responses in enterobacteria. *FEMS*
619 *Microbiology Letters* vol. 147 Preprint at [https://doi.org/10.1016/S0378-1097\(96\)00503-4](https://doi.org/10.1016/S0378-1097(96)00503-4)
620 (1997).
- 621 73. Guan, N. & Liu, L. Microbial response to acid stress: mechanisms and applications. *Applied*
622 *Microbiology and Biotechnology* vol. 104 Preprint at [https://doi.org/10.1007/s00253-019-](https://doi.org/10.1007/s00253-019-10226-1)
623 10226-1 (2020).
- 624 74. Deberardinis, R. J. & Chandel, N. S. Fundamentals of cancer metabolism INTRODUCTION
625 AND OVERARCHING PRINCIPLES. *Adv Sci* (2016).
- 626 75. Ohara, T. & Mori, T. Antiproliferative Effects of Short-chain Fatty Acids on Human
627 Colorectal Cancer Cells via Gene Expression Inhibition. *Anticancer Res* **39**, (2019).
- 628 76. Sieow, B. F. L., Wun, K. S., Yong, W. P., Hwang, I. Y. & Chang, M. W. Tweak to Treat:
629 Reprogramming Bacteria for Cancer Treatment. *Trends in Cancer* vol. 7 Preprint at
630 <https://doi.org/10.1016/j.trecan.2020.11.004> (2021).
- 631 77. Ezraty, B., Gennaris, A., Barras, F. & Collet, J. F. Oxidative stress, protein damage and
632 repair in bacteria. *Nature Reviews Microbiology* *2017 15:7* **15**, 385–396 (2017).
- 633 78. Al-Koussa, H., el Mais, N., Maalouf, H., Abi-Habib, R. & El-Sibai, M. Arginine deprivation:
634 A potential therapeutic for cancer cell metastasis? A review. *Cancer Cell International* vol.
635 20 Preprint at <https://doi.org/10.1186/s12935-020-01232-9> (2020).

- 636 79. Beiter, K. *et al.* An endonuclease allows *Streptococcus pneumoniae* to escape from
637 neutrophil extracellular traps. *Curr Biol* **16**, 401–407 (2006).
- 638 80. Ho, S. S., Michalek, S. M. & Nahm, M. H. Lipoteichoic acid is important in innate immune
639 responses to gram-positive bacteria. *Infect Immun* **76**, (2008).
- 640 81. S N Chaitanya, N., Devi, A., Sahu, S. & Alугоju, P. Molecular mechanisms of action of
641 Trehalose in cancer: A comprehensive review. *Life Sciences* vol. 269 Preprint at
642 <https://doi.org/10.1016/j.lfs.2020.118968> (2021).
- 643 82. Eil, R. *et al.* Ionic immune suppression within the tumour microenvironment limits T cell
644 effector function. *Nature* **537**, (2016).
- 645 83. Žgur-Bertok, D. DNA Damage Repair and Bacterial Pathogens. *PLoS Pathog* **9**, (2013).
- 646 84. Podlesek, Z. & Žgur Bertok, D. The DNA Damage Inducible SOS Response Is a Key Player
647 in the Generation of Bacterial Persister Cells and Population Wide Tolerance. *Front*
648 *Microbiol* **11**, 1785 (2020).
- 649 85. Inglis, R. F., Bayramoglu, B., Gillor, O. & Ackermann, M. The role of bacteriocins as selfish
650 genetic elements. *Biol Lett* **9**, (2013).
- 651 86. Shapira, S. *et al.* Innovative dual system approach for selective eradication of cancer cells
652 using viral-based delivery of natural bacterial toxin–antitoxin system. *Oncogene* **40**, (2021).
- 653 87. Shi, Y. *et al.* Intratumoral accumulation of gut microbiota facilitates CD47-based
654 immunotherapy via STING signaling. *Journal of Experimental Medicine* **217**, (2020).
- 655 88. David, H. Rudolf Virchow and Modern Aspects of Tumor Pathology. *Pathol Res Pract* **183**,
656 (1988).
- 657 89. Grivennikov, S. I. *et al.* Adenoma-linked barrier defects and microbial products drive IL-
658 23/IL-17-mediated tumour growth. *Nature* **491**, (2012).
- 659 90. Yang, W. & Cong, Y. Gut microbiota-derived metabolites in the regulation of host immune
660 responses and immune-related inflammatory diseases. *Cellular and Molecular Immunology*
661 vol. 18 Preprint at <https://doi.org/10.1038/s41423-021-00661-4> (2021).
- 662 91. Wang, K. & Karin, M. Tumor-Elicited Inflammation and Colorectal Cancer. in *Advances in*
663 *Cancer Research* vol. 128 (2015).
- 664 92. Hou, J., Karin, M. & Sun, B. Targeting cancer-promoting inflammation — have anti-
665 inflammatory therapies come of age? *Nature Reviews Clinical Oncology* vol. 18 Preprint at
666 <https://doi.org/10.1038/s41571-020-00459-9> (2021).
- 667 93. Shalapour, S. & Karin, M. Cruel to Be Kind: Epithelial, Microbial, and Immune Cell
668 Interactions in Gastrointestinal Cancers. *Annual Review of Immunology* vol. 38 Preprint at
669 <https://doi.org/10.1146/annurev-immunol-082019-081656> (2020).
- 670 94. Li, X. *et al.* Role of glutamine and its metabolite ammonia in crosstalk of cancer-associated
671 fibroblasts and cancer cells. *Cancer Cell International* vol. 21 Preprint at
672 <https://doi.org/10.1186/s12935-021-02121-5> (2021).
- 673 95. Vodnala, S. K. *et al.* T cell stemness and dysfunction in tumors are triggered by a common
674 mechanism. *Science (1979)* **363**, (2019).
- 675 96. Csonka, L. N. Physiological and genetic responses of bacteria to osmotic stress. *Microbiol*
676 *Rev* **53**, (1989).
- 677 97. Li, S., Konstantinov, S. R., Smits, R. & Peppelenbosch, M. P. Bacterial Biofilms in
678 Colorectal Cancer Initiation and Progression. *Trends in Molecular Medicine* vol. 23 Preprint
679 at <https://doi.org/10.1016/j.molmed.2016.11.004> (2017).
- 680 98. Garza, D. R. *et al.* Metabolic models predict bacterial passengers in colorectal cancer.
681 *Cancer Metab* **8**, (2020).
- 682 99. Geng, J. *et al.* Co-occurrence of driver and passenger bacteria in human colorectal cancer.
683 *Gut Pathog* **6**, (2014).

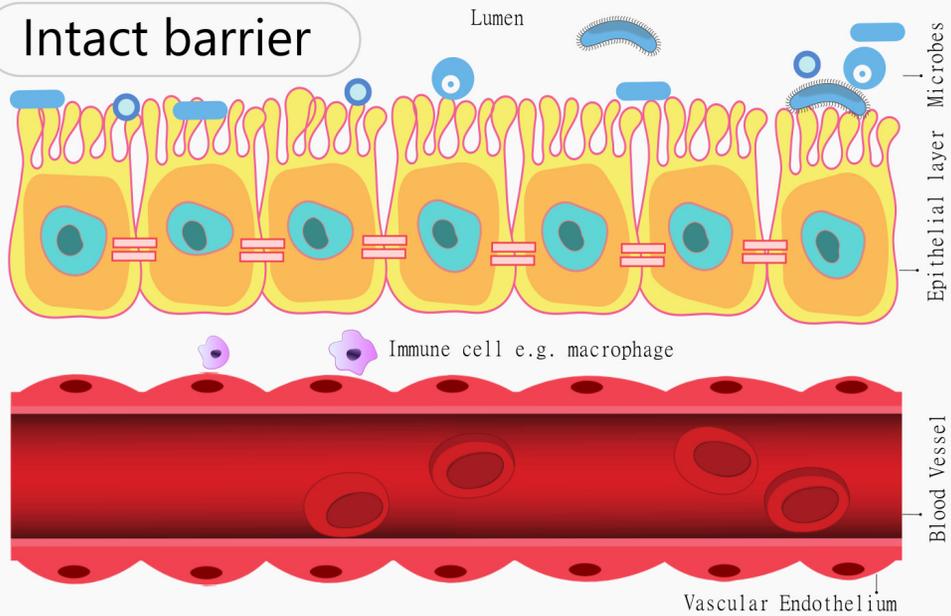
- 684 100. Hols, P., Ledesma-García, L., Gabant, P. & Mignolet, J. Mobilization of Microbiota
685 Commensals and Their Bacteriocins for Therapeutics. *Trends in Microbiology* vol. 27
686 Preprint at <https://doi.org/10.1016/j.tim.2019.03.007> (2019).
- 687 101. Chakrabarty, A. M. Microorganisms and cancer: Quest for a therapy. *Journal of Bacteriology*
688 vol. 185 Preprint at <https://doi.org/10.1128/JB.185.9.2683-2686.2003> (2003).
- 689 102. Park, D. S. *et al.* The goldilocks window of personalized chemotherapy: Getting the immune
690 response just right. *Cancer Res* **79**, (2019).
- 691 103. Boedtkjer, E. & Pedersen, S. F. The Acidic Tumor Microenvironment as a Driver of Cancer.
692 *Annual Review of Physiology* vol. 82 Preprint at [https://doi.org/10.1146/annurev-physiol-](https://doi.org/10.1146/annurev-physiol-021119-034627)
693 [021119-034627](https://doi.org/10.1146/annurev-physiol-021119-034627) (2020).
- 694 104. de Berardinis, R. J. & Chandel, N. S. Fundamentals of cancer metabolism. *Science Advances*
695 vol. 2 Preprint at <https://doi.org/10.1126/sciadv.1600200> (2016).
- 696 105. Beamish, E. L. *et al.* Loop ileostomy-mediated fecal stream diversion is associated with
697 microbial dysbiosis. *Gut Microbes* **8**, (2017).
- 698 106. Helmink, B. A., Khan, M. A. W., Hermann, A., Gopalakrishnan, V. & Wargo, J. A. The
699 microbiome, cancer, and cancer therapy. *Nature Medicine* vol. 25 Preprint at
700 <https://doi.org/10.1038/s41591-019-0377-7> (2019).
- 701 107. Kolodziejczyk, A. A., Zheng, D. & Elinav, E. Diet–microbiota interactions and personalized
702 nutrition. *Nature Reviews Microbiology* vol. 17 Preprint at [https://doi.org/10.1038/s41579-](https://doi.org/10.1038/s41579-019-0256-8)
703 [019-0256-8](https://doi.org/10.1038/s41579-019-0256-8) (2019).
- 704 108. Lee, C. & Longo, V. D. Fasting vs dietary restriction in cellular protection and cancer
705 treatment: From model organisms to patients. *Oncogene* vol. 30 Preprint at
706 <https://doi.org/10.1038/onc.2011.91> (2011).

707

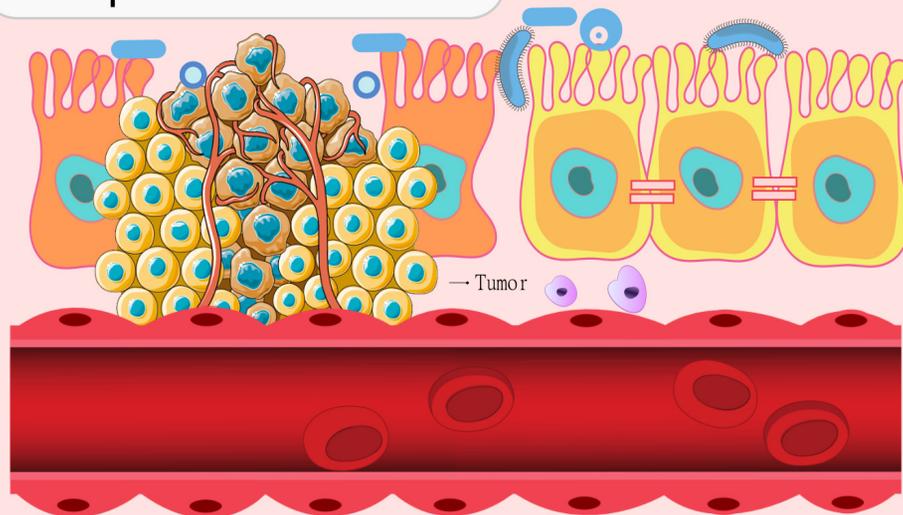
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Intact barrier



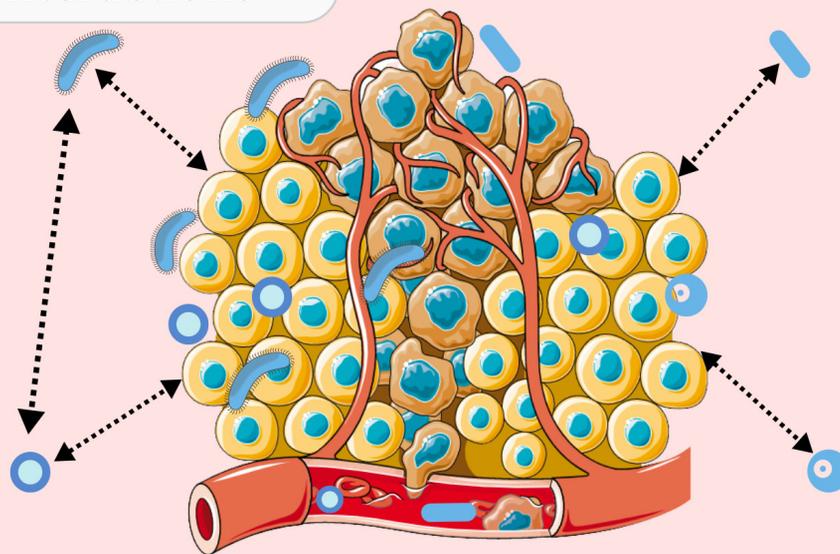
Compromised barrier



Infiltration



Interactions



NUTRIENT STRESS

Hyperanabolic tumor cells **create a nutrient deficit** in their microenvironment

PHYSICAL & OSMOTIC STRESS

Heightened immune cell **infiltration, dynamicity, NETs and ion efflux** prevent colonization and create osmotic imbalance

OXIDATIVE STRESS

Aberrant growth contributes to **elevated levels of reactive oxygen species (ROS)**

ACID STRESS

Carbonic anhydrase, proton release and other factors mediated **hyperacidic surface of tumor** helps in metastasis/ invasion

CYTOTOXIC & DNA DAMAGE STRESS

Tumor represents an environment **replete with cytotoxic drugs, overexpressed nucleases**

CHARACTERIZING STRESSES IN THE TUMOR MICRO(B)ENVIRONMENT

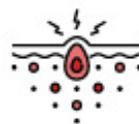
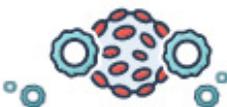
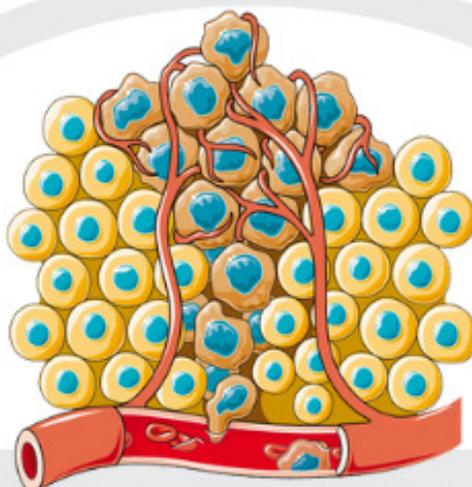


Table 1. Potential responses of tumor invading/inhabiting microbiota mediated by the expression of GSR regulon under diverse environmental insults of the TME. Notably, the GSR targets can be ameliorated by any stressor that can activate entire general stress regulon (conferring cross protectivity).

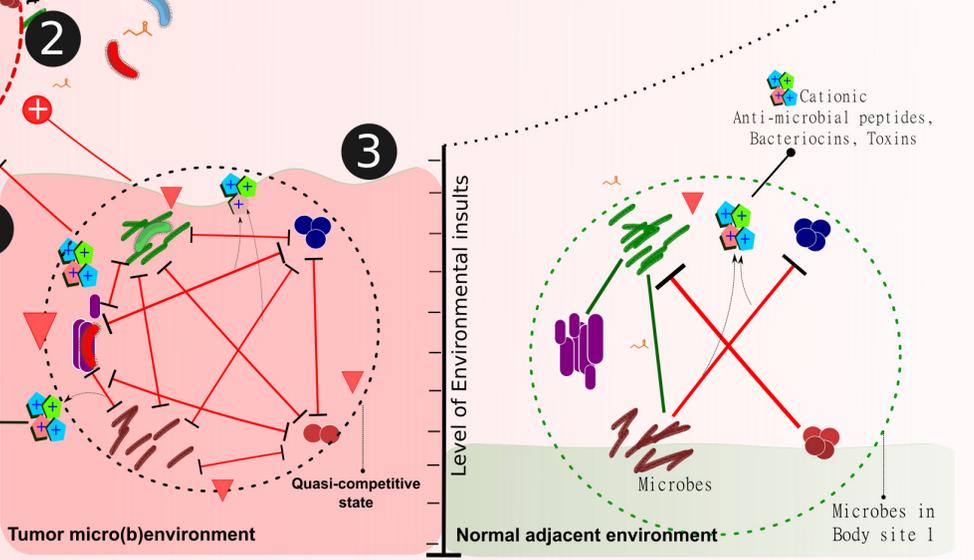
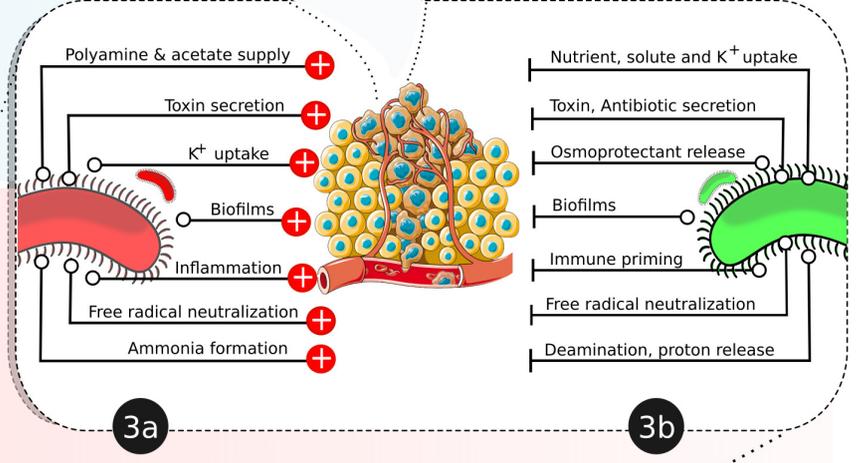
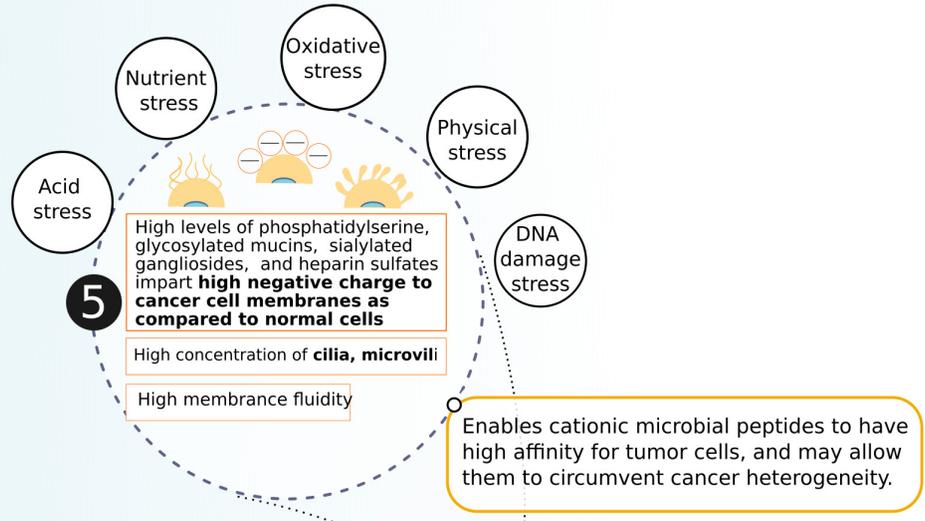
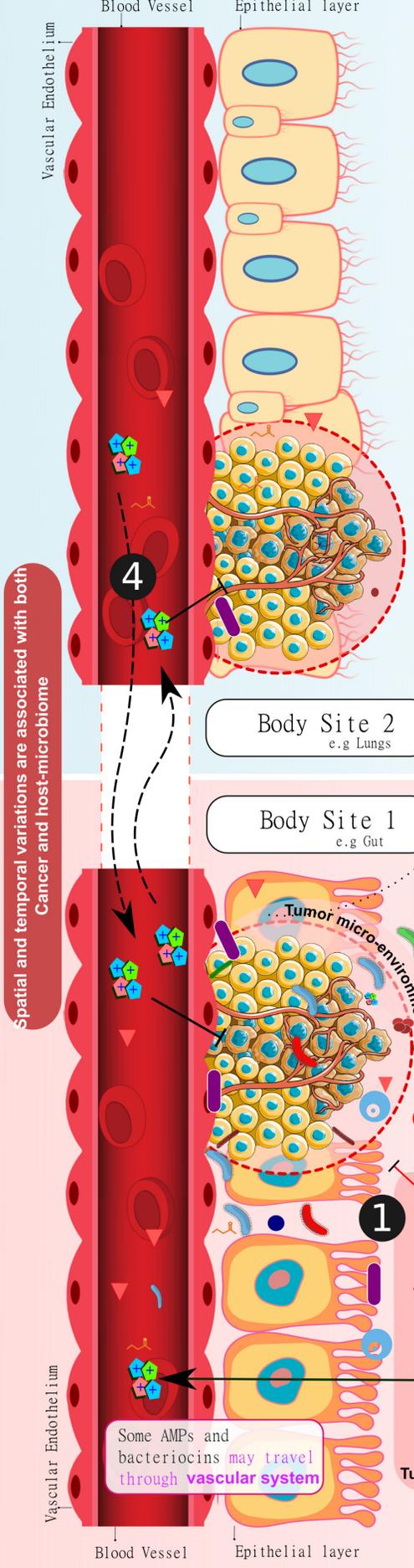
GSR target	Phenotype	Mechanism of collateral damage for cancer	Reference
<i>Nutrient stress</i>	<ul style="list-style-type: none"> Accumulation of nutrients (e.g. glycogen, amino acids, acetate, iron etc) shift to fermentation biofilm formation 	<ul style="list-style-type: none"> Resource limitation for hyperanabolic cancer cells (cancers need glycogen, acetate, iron, BCAA etc) anti-mitotic role of SCFAs metastasis distraction by biofilms through secretion of exopolysaccharides, preventing cancer cell binding to the endothelial cells 	59,63–65
<i>Oxidative stress</i>	<ul style="list-style-type: none"> Expression of free radical scavenging enzymes like catalases, oxidoreductases, Superoxide dismutase Damage repairing proteins like thioredoxins, glutaredoxins, and methionine sulfoxide reductases 	<ul style="list-style-type: none"> Free radical clearance and release of damage repairing proteins limits DNA damage, inflammatory cytokines, metastasis, and oncogenic mutagenesis 	20,66
<i>Acid Stress</i>	<ul style="list-style-type: none"> expression of amino acid (Arginine and Glutamate) decarboxylases activation of Arginine deaminase system proton pumping increased glycolytic activity biofilm formation 	<ul style="list-style-type: none"> Cancer cells are arginine addict (deprivation leads to cancer cell death) Glutamate is a key substrate for cancer cells Proton release by intra-tumor microbes can disrupt pH of cancer cells Heightened microbial glycolytic activity and biofilm formation can compete for energy metabolism and prevent metastasis 	15,28,67

<p>Physical Stress</p>	<ul style="list-style-type: none"> • upregulated MSCRAMMs* and biofilm formation • expression of autolysin like enzymes and release of eDNA*, teichoic acid and other cytoplasmic contents • upregulation of virulence factors like surface endonucleases 	<ul style="list-style-type: none"> • MSCRAMMs mediate covalent binding leading to persistent biofilms that can compete for nutrition and arrest metastasis • eDNA and teichoic acids can mediate non-covalent binding of microbes to cancer, and can also trigger immune surveillance for collateral recognition of cancer cells • Degradation of NETs* and other entrapments by surface endonucleases can prevent metastasis 	<p>25,26,31,68,69</p>
<p>Osmotic stress</p>	<ul style="list-style-type: none"> • Solute uptake including amino acids, potassium ions (K⁺) • Synthesis and accumulation of Trehalose 	<ul style="list-style-type: none"> • Uptake of amino acids as solutes can limit cancer energy metabolism • Trehalose released through mechanosensitive channels and upon bacterial lysis can reduce inflammation, limit free radicals, enhance apoptosis • Uptake of the storm of K⁺ ions released by dying cancer cells can limit suppression of cancer killing T-cell effector function 	<p>50,70,71</p>
<p>DNA damage</p>	<p>SOS response upregulates –</p> <ul style="list-style-type: none"> • biofilms with (drug resistant) persister population • Intraspecies competition and consequent toxin secretion • Toxin-anti toxin (TA) system activation • Horizontal gene transfer 	<ul style="list-style-type: none"> • Persistent biofilms can compete for energy metabolism and prevent metastasis • Toxins against intraspecies competition (e.g. colicins) can inhibit cancers • TA systems can specifically cause cancer cell death (e.g. MazF-MazE toxin–antitoxin of E.coli against pancreatic and colorectal cancers) • Anti-cancer toxins/antibiotics encoded by plasmids can promote population level phenotype through HGT 	<p>41,53,72–75</p>

* *MSCRAMM: Microbial Surface Components Recognizing Adhesive Matrix Molecules are microbial surface proteins that adhere specifically to host extra-cellular matrix (ECM); eDNA: extracellular DNA; NETs: Neutrophil Extracellular Traps*

 Anti-microbial peptides
e.g. bacteriocins
  Pro-inflammatory molecules
e.g. endotoxins, LPS
  Short chain fatty acids (SCFAs)
e.g. Butyrate

 Microorganisms
  Tumor microenvironment



Tumor microenvironment can have consequences for host microbial ecology in and around the TME

Spatial and temporal variations are associated with both Cancer and host-microbiome