

Assessing species interactions using integrated predator-prey models

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Abstract

1
2 Inferring the strength of species interactions from demographic data is a challenging task. The
3 Integrated Population Modelling (IPM) approach, bringing together population counts, capture-
4 recapture, and individual-level fecundity data into a unified model framework, has been extended
5 from single species to the community level. This allows to specify IPMs for multiple species with
6 interactions specified as links between vital rates and stage-specific densities. However, there is no
7 evaluation of such models when interactions are actually absent—while any interaction inference
8 method runs the risk of producing false positives. We investigate here whether multispecies
9 IPMs could output interactions where there are in fact none, building on an existing predator-
10 prey IPM. We show that interspecific density-dependence estimates are centered on zero when
11 simulated to be zero, and therefore their estimation is unbiased. Their coverage probability,
12 quantifying how many times credible intervals include zero, is also satisfactory. We further
13 confirm that adding random temporal variation to multispecies density-dependent link functions
14 does not alter these results. This study therefore reaffirms the potential of multispecies IPMs
15 to infer correctly how biotic interactions influence demography, although future studies should
16 investigate model misspecifications.

17 **Keywords:** Integrated Population Model; data assimilation; species interactions; predation;
18 density-dependence.

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21 1 Introduction

22 Estimating ecological interactions between and within species through models of their joint popula-
23 tion dynamics is a task which requires large amounts of data. Indeed, with potentially as many as
24 q^2 interaction parameters for q model compartments (combination of species and age classes), the
25 number of parameters to estimate can climb very rapidly. Therefore, ecological statistics searches
26 for improved ways to infer such population-level interaction strengths. A recently developed tech-
27 nique consists in combining data sources in multispecies Integrated Populations Models (IPMs)
28 including interspecific interactions (Péron & Koons, 2012; Barraquand & Gimenez, 2019; Quéroué
29 *et al.*, 2021). Because Integrated Population Models (IPMs, Besbeas *et al.*, 2002) combine data on
30 demographic rates (e.g., capture recapture, breeding data) with data on population size (typically
31 from counts), they allow: (a) estimating both demographic rates and population size (and hence
32 their inter-dependencies) in a joint analysis, (b) an improved precision in parameter estimates,
33 compared to separate analyses of component datasets, since the information contained in several
34 datasets combine into estimated parameters (e.g., count data and capture recapture data both con-
35 tain information on survival rates), and in some cases (c) to estimate parameters for which there is
36 no dedicated data stream, that can only be estimated through inverse estimation of a demographic
37 model (Kéry & Schaub, 2011; Abadi *et al.*, 2010). This last property is particularly useful to esti-
38 mate population-level species interactions strengths, since population-level interactions are always
39 indirectly inferred. Although inverse estimation can in theory be performed using a single data
40 source such as population counts, such inverse estimation is a difficult task fraught with identifi-
41 ability issues. Asking whether multispecies IPMs performed better than classical inverse estimation
42 from count data alone, Barraquand & Gimenez (2019) have shown that better estimates of interac-
43 tion parameters could be obtained by combining data sources. Additionally, an empirical study in
44 a bird predator-prey system Quéroué *et al.* (2021) was able to detect the expected bottom-up de-
45 mographic linkages from prey to predator but not the expected top-down relationships, suggesting
46 that those may be too weak to be detected.

47 In these multispecies IPM studies estimating interspecific interactions, between-species linkages
48 have always been considered to be present in the simulations or in the underlying reality (based on
49 background knowledge). Other choices are possible: some multispecies IPMs do not assume inter-
50 specific interactions to be present a priori (Lahoz-Monfort *et al.*, 2017), but they do not estimate
51 them either and focus instead on environmental effects. However, multispecies IPMs with inter-

52 specific interactions could also be used in situations where it is not clear whether population-level
53 interactions between species are possible. This is all the more true that interactions are specified as
54 links between vital rates and stage-specific densities, and while some of these relationships may be
55 known a priori, others may not. The issue was raised but not tackled by [Barraquand & Gimenez](#)
56 (2019): a natural follow-up is therefore to ask what happens whenever we try to estimate interac-
57 tions that are actually absent, to make sure that multispecies IPMs do not yield false positives.

58 Let us note that when estimating or predicting interspecific interactions in general—not just
59 with multispecies IPMs—whether methods could output false positives is a key concern (e.g., with
60 multivariate autoregressive models, [Mutshinda et al. 2009](#); [Barraquand et al. 2021](#); dynamic bayesian
61 networks, [Sander et al. 2017](#); or other machine learning tools, [Strydom et al. 2021](#)). The fact that
62 all interaction inference methods run the risk of creating false positives of interspecific interactions
63 at exaggerated rates only reinforces the need to evaluate it in multispecies IPMs.

64 An additional concern is temporal stochasticity in the functions linking vital rates of a given
65 stage of species i to the densities of a given stage of species j . In the simulation-based study of
66 [Barraquand & Gimenez \(2019\)](#), it was assumed that such stochasticity was absent, while empirical
67 studies ([Péron & Koons, 2012](#); [Quéroué et al., 2021](#)) assumed its presence in order to partition
68 variation in vital rates due to species densities vs other factors changing over time. We therefore
69 still need to understand whether theoretical performances hold in this more empirically realistic
70 context, where environmental factors can perturb demographic rates, and those are not solely
71 deterministic functions of species densities.

72 To sum up, we follow-up here on the multispecies IPM study of [Barraquand & Gimenez \(2019\)](#)
73 by asking whether (1) inter-species interactions are truly estimated to be zero when species have
74 in fact independent dynamics and (2) how species interaction strengths estimates can be affected
75 by the absence and presence of environmental stochasticity (random year effects on demographic
76 rates).

77 2 Methods

78 2.1 General description of the multispecies IPM

79 The deterministic skeleton can be described as a density-dependent matrix population model

$$\mathbf{n}_{t+1} = \mathbf{A}(\mathbf{n}_t)\mathbf{n}_t. \tag{1}$$

Eq. 1 describes in discrete-time the dynamics of abundances of two species and two stages per species, with projection matrix

$$\mathbf{A}(\mathbf{n}_t) = \begin{pmatrix} 0 & \frac{1}{2}f_{V,t} \left(n_{V,t}^A \right) \phi_{V,t}^J \left(n_{P,t}^A \right) & 0 & 0 \\ \phi_{V,t}^A & \phi_{V,t}^A & 0 & 0 \\ 0 & 0 & 0 & \frac{1}{2}f_{P,t} \left(n_{V,t}^J \right) \phi_{P,t}^J \left(n_{P,t}^A \right) \\ 0 & 0 & \phi_{P,t}^A & \phi_{P,t}^A \end{pmatrix}$$

and abundance vector

$$\mathbf{n}_t = \begin{pmatrix} n_{V,t}^J \\ n_{V,t}^A \\ n_{P,t}^J \\ n_{P,t}^A \end{pmatrix}$$

80 where $n_{V,t}^J, n_{V,t}^A, n_{P,t}^J$ and $n_{P,t}^A$ are respectively the abundances of juvenile prey (denoted V as 'vic-
81 tim'), adult prey, juvenile predators and adult predators, at time t . The fecundities $f_{V,t}, f_{P,t}$ are the
82 expected number of juvenile prey and predator produced by an adult prey and predator, respec-
83 tively. Survival probabilities between t and $t + 1$ are denoted with ϕ , so that $\phi_{V,t}^J, \phi_{V,t}^A, \phi_{P,t}^J$ and $\phi_{P,t}^A$
84 are the survival probabilities of the juvenile prey, adult prey, juvenile predator and adult predator.

85 2.1.1 Count data

86 To simulate and account for demographic stochasticity, we modelled yearly (st)age specific abun-
87 dances \mathbf{n}_t using Binomial and Poisson distributions as in Barraquand & Gimenez (2019) eqs. (2)–(5).

88 Regarding the observation process for count data, the 2019 model assumed a negligible obser-
89 vation error ($\sigma^2 = 10^{-5}$). The reason was that in absence of replicated counts at each time unit,
90 observation error variance is notoriously difficult to disentangle from process error variance (Knape,
91 2008; Auger-Méthé *et al.*, 2016). While in some cases it could be possible to remove observation error
92 altogether, because total population sizes of each species (summed numbers of juveniles and adults)
93 are the observed count variables (as in most IPMs), they need to appear in the model as drawn from
94 some probability distribution—they need to be a stochastic node in the MCMC representation. It
95 was therefore decided to keep the formulation of the model in its state-space version, but forcing it
96 to observe true population size almost with certainty (negligible process error variance). However,
97 we uncovered in the present work that stage-specific abundances could not be estimated properly.

98 Because correctly reproducing stage-specific abundances when fitting a stage-structured model is
 99 desirable, and that there is in most wildlife surveys some measure of observation error on counts,
 100 we assumed in the present article a non-negligible, positive observation error variance. As we do
 101 not have replicated counts at any given time, we do not attempt to estimate observation error vari-
 102 ance, and assume that it is known and classically set on the logarithmic scale (i.e., the coefficient
 103 of variation of observed population size is constant). For predator counts (denoted P) we have

$$y_{P,t}|\mathbf{n}_t \sim \mathcal{LN}(\log(n_{P,t}^J + n_{P,t}^A), \sigma_{obs}^2) \quad (2)$$

104 and similarly for prey counts

$$y_{V,t}|\mathbf{n}_t \sim \mathcal{LN}(\log(n_{V,t}^J + n_{V,t}^A), \sigma_{obs}^2), \quad (3)$$

105 with \mathcal{LN} the log-Normal distribution and its associated variance on the log-scale $\sigma_{obs}^2 = 0.1$. Other
 106 choices of observation model are possible but this one is standard for abundance values that are not
 107 too small (Besbeas *et al.*, 2002; Dennis *et al.*, 2006).

108

109 2.1.2 Survival data

110 To increase computational efficiency (particularly true for the scenarios with more individuals cap-
 111 tured and a shorter time series), we simulated and fitted the capture-mark-recapture data in the
 112 m-array format, using a multinomial likelihood (Burnham, 1987). The data is in the form of two
 113 $(T-1) \times T$ matrices \mathbf{M}^J and \mathbf{M}^A , one for each age class, with $\mathbf{M}^{(a)} = (m_{t,j}^{(a)})$, with $m_{t,j}^{(a)} = 0$, $\forall j < t$,
 114 where T is the total number of years of capture recapture history. $m_{t,t}^{(a)}$ is the number of individuals
 115 of released at age class (a) at time t that were re-sighted the following year, and the last column
 116 $m_{t,T}^{(a)}$ is the number of individuals released at age class (a) at time t that were never re-sighted. We
 117 then have:

$$\mathbf{m}_{t,\bullet}^{(a)} = (m_{t,t}^{(a)}, m_{t,t+1}^{(a)}, \dots, m_{t,T}^{(a)}) \sim \text{Multinomial} \left(R_t^{(a)}, (\theta_{t,t}^{(a)}, \dots, \theta_{t,T}^{(a)}) \right) \quad (4)$$

118 with $R_t^{(a)} = \sum_{k=t}^T m_{t,k}^{(a)}$ the number of individuals of age class (a) released at time t .

119 It is important to note that for the matrix of released juveniles \mathbf{M}^J , R_t^J corresponds to the
 120 number of juveniles newly marked at time t . However, R_t^A corresponds to the number of newly

121 marked adults (lets denote it $R_{m,t}^A$), but *also* of all previously marked juveniles and adults that were
 122 released at time t . That is,

$$R_1^A = R_{m,1}^A, \quad (5)$$

123 but

$$R_{t+1}^A = R_{m,t}^A + \sum_{k=1}^t m_{k,t}^J + m_{k,t}^A. \quad (6)$$

124 Therefore, unless individuals are not released when marked (e.g., killed, or taken to be released
 125 outside of the study population), one needs to provide data and model the number of released
 126 adults re-sighted, even if no individuals are first marked as adults. As no individuals are marked as
 127 adults here, $R_{m,1}^A = 0$, and so that Equations 5 and 6 can be simplified accordingly.

128 Note also that in such case were no adults are newly marked, no data on R_t^A is needed to
 129 simulate and fit \mathbf{M}^A . Since $R_1^A = 0$, we have:

$$\mathbf{m}_{1,\bullet}^A = 0, \quad (7)$$

130

$$\mathbf{m}_{2,\bullet}^A \sim \text{Multinomial} (m_{1,1}^J + m_{1,1}^A, (\theta_{1,t}^A, \dots, \theta_{1,T}^A)), \quad (8)$$

131

$$\mathbf{m}_{3,\bullet}^A \sim \text{Multinomial} \left(\sum_{k=1}^2 m_{k,2}^J + m_{k,2}^A, (\theta_{2,t}^A, \dots, \theta_{2,T}^A) \right), \quad (9)$$

132 and so on.

133 For juveniles, diagonal elements of the $\boldsymbol{\theta}^J$ matrix write:

$$\theta_{t,t}^J = \phi_t^J p,$$

134 with ϕ_t^J the first year (i.e. juvenile) survival probability from year t to year $t + 1$ (for the species
 135 considered), and p the recapture (or re-sighting) probability set as constant among years and age
 136 classes, and for $t < j < T$

$$\theta_{t,j}^J = \phi_t^J \left(\prod_{k=t+1}^j \phi_k^A \right) (1 - p)^{j-t} p,$$

137 with ϕ_t^A the adult survival probability from year t to year $t + 1$ (for the species considered). The

138 last element pertains to individuals never recaptured

$$\theta_{t,T}^J = 1 - \sum_{k=t}^{T-1} \theta_{t,k}^J.$$

139 Similarly for θ^A , the above mentioned equations are identical to the exception that ϕ^J is replaced
140 by ϕ^A , which leads to:

$$\theta_{t,t}^A = \phi_t^A p$$

141 for the diagonal elements of the θ^A matrix, and for $t < j < T$:

$$\theta_{t,j}^A = \left(\prod_{k=t}^j \phi_k^A \right) (1-p)^{j-t} p.$$

142 The last element again pertains to individuals never recaptured

$$\theta_{t,T}^A = 1 - \sum_{k=t}^{T-1} \theta_{t,k}^A.$$

143 **2.1.3 Fecundity data**

144 Fecundity was modelled using a Poisson regression:

$$F_t \sim \text{Poisson}(f_t R_t) \tag{10}$$

145 with F_t the total number of offspring counted, R_t the number of surveyed broods/litters, and f_t the
146 expected number of offspring (male + female) per adult female each year t .

147 **2.2 Alternative scenarios and parameter values**

148 **2.2.1 Density dependence and random temporal variation on demographic rates**

149 Intra- and inter-species density dependence of survival rates $\phi_{i,t}^j$ (with $i \in \{V, P\}$ and $j \in \{J, A\}$)
150 and fecundities $f_{i,t}$ were modelled on the logit and log scale, respectively. We initially used the
151 same equations as the 2019 model, which are:

$$\text{logit}(\phi_{P,t}^J) = \alpha_1 + \alpha_2 n_{P,t}^A \tag{11}$$

152 where the number of adult predators negatively affects juvenile predator survival (negative intraspe-
 153 cific density dependence),

$$\text{logit}(\phi_{V,t}^J) = \alpha_3 + \alpha_4 n_{P,t}^A \quad (12)$$

where the number of adult predators negatively affects juvenile prey survival (predation),

$$\text{logit}(\phi_{P,t}^A) = \alpha_{\phi_P^A} \quad (13)$$

$$\text{logit}(\phi_{V,t}^A) = \alpha_{\phi_V^A} \quad (14)$$

154 (no density dependence on adult survival)

$$\log(f_{P,t}) = \alpha_5 + \alpha_6 n_{V,t}^J \quad (15)$$

155 where the number of juvenile prey positively affects predator fecundity, and

$$\log(f_{V,t}) = \alpha_7 + \alpha_8 n_{V,t}^A \quad (16)$$

156 where the number of adult prey individuals negatively affect prey fecundity (negative intraspecific
 157 density dependence). Associated results can be found in Supplementary Information Table [S1](#) and
 158 Figures [S5](#) to [S8](#).

159 However, to limit posterior correlation between intercept and slope parameters and improve
 160 their estimation, we centered the abundances in the density dependent functions. While centering
 161 is typically done and most efficient on mean values, mean abundances varied here from a simulation
 162 to the next due to stochasticity. Therefore, intercept parameter values would have to be redefined for
 163 each simulation to maintain equivalent mean demographic rate values and asymptotic stage specific
 164 abundance equilibria for all simulation. To avoid these complications, we centered by subtracting
 165 the corresponding fixed point equilibria estimated in [Barraquand & Gimenez \(2019\)](#) as $\bar{n}_P^A = 21$,
 166 $\bar{n}_V^J = 101$ and $\bar{n}_V^A = 152$. The new α intercept parameters obey the following centered formulas:

$$\text{logit}(\phi_{P,t}^J) = \alpha_1 + \alpha_2(n_{P,t}^A - \bar{n}_P^A) \quad (17)$$

$$\text{logit}(\phi_{V,t}^J) = \alpha_3 + \alpha_4(n_{P,t}^A - \bar{n}_P^A) \quad (18)$$

$$\text{logit}(\phi_{P,t}^A) = \alpha_{\phi_P^A} \quad (19)$$

$$\text{logit}(\phi_{V,t}^A) = \alpha_{\phi_V^A} \quad (20)$$

$$\log(f_{P,t}) = \alpha_5 + \alpha_6(n_{V,t}^J - \bar{n}_V^J) \quad (21)$$

$$\log(f_{V,t}) = \alpha_7 + \alpha_8(n_{V,t}^A - \bar{n}_V^A). \quad (22)$$

167 To maintain equivalent dynamics to parameter set 1 of the 2019 model, we calculated the
 168 intercepts α_1 , α_3 , α_5 and α_7 as their original values plus the original slope multiplied by the
 169 estimated fixed point equilibrium of the n responsible for density dependence. For example, we now
 170 use whenever simulating $\alpha_3 = 0.5 - 0.025 \times 21 = -0.025$ and $\alpha_5 = 0 + 0.004 \times 101 = 0.404$ (Table 1).

171 In addition, we introduced scenarios with inter-annual random variation in the intercepts of
 172 density-dependent links, such that

$$\text{logit}(\phi_{P,t}^J) = \alpha_1 + \alpha_2(n_{P,t}^A - \bar{n}_P^A) + \sigma_{\phi_P^J} \epsilon_{\phi_P^J} \quad (23)$$

$$\text{logit}(\phi_{V,t}^J) = \alpha_3 + \alpha_4(n_{P,t}^A - \bar{n}_P^A) + \sigma_{\phi_V^J} \epsilon_{\phi_V^J} \quad (24)$$

$$\text{logit}(\phi_{P,t}^A) = \alpha_{\phi_P^A} + \sigma_{\phi_P^A} \epsilon_{\phi_P^A} \quad (25)$$

$$\text{logit}(\phi_{V,t}^A) = \alpha_{\phi_V^A} + \sigma_{\phi_V^A} \epsilon_{\phi_V^A} \quad (26)$$

with $\epsilon \sim \mathcal{N}(0, 1)$ i.i.d. and

$$\log(f_{P,t}) \sim \mathcal{N}(\alpha_5 + \alpha_6(n_{V,t}^J - \bar{n}_V^J), \sigma_{f_P}^2) \quad (27)$$

$$\log(f_{V,t}) \sim \mathcal{N}(\alpha_7 + \alpha_8(n_{V,t}^A - \bar{n}_V^A), \sigma_{f_V}^2). \quad (28)$$

173 Although mathematically identical, we used a parameterisation of the form $\mu + \epsilon\sigma$, $\epsilon \sim \mathcal{N}(0, \sigma^2)$
 174 (sometimes called non-centered) for survival estimates and a centered parameterisation ($\mathcal{N}(\mu, \sigma^2)$)
 175 for fecundity estimates as it was found to be optimal for the mixing of the MCMC chains. As we
 176 were primarily interested in the ability of multispecies IPMs to estimate species interactions when
 177 these were in fact absent, inter species density dependence parameter values for α_2 and α_4 were

Table 1: Model parameters with their values. Values of α_4 and α_6 in the scenarios with true presence of species interactions are presented in parentheses. Temporal standard deviations (SD) are only present in the scenarios with random temporal variation. For interpretation, note that α_i and temporal SD parameters are within exponential functions. For instance, $\alpha_5 = 0.404$ corresponds to a mean fecundity of $e^{0.404} \approx 1.5$.

Parameter	Value	Interpretation
α_1	0.29	juvenile predator survival – intercept
α_2	-0.01	juvenile predator survival – slope
α_3	-0.025	juvenile prey survival – intercept
α_4	0 (-0.025)	juvenile prey survival – slope – inter species density dependence
α_5	0.404	predator fecundity – intercept
α_6	0 (0.004)	predator fecundity – slope – inter species density dependence
α_7	1.24	prey fecundity – intercept
α_8	-0.005	prey fecundity – slope
p	0.7	recapture probability
$\alpha_{\phi_P^A}$	logit(0.7)	adult predator survival – intercept
$\alpha_{\phi_V^A}$	logit(0.6)	adult prey survival – intercept
σ_{obs}^2	0.1	observation error
σ_{f_P}	0.1	temporal SD of predator fecundity
σ_{f_V}	0.1	temporal SD of prey fecundity
$\sigma_{\phi_P^J}$	0.1	temporal SD of juvenile predator survival
$\sigma_{\phi_P^A}$	0.1	temporal SD deviation of adult predator survival
$\sigma_{\phi_V^J}$	0.1	temporal SD deviation of juvenile prey survival
$\sigma_{\phi_V^A}$	0.1	temporal SD deviation of adult prey survival

178 either set to zero for the simulations, or at the same value as the 2019 model. Parameter values
 179 used to simulate data and their interpretation can be found in Table 1.

180 2.2.2 Initial values and monitoring setup

For all simulation scenarios in the main text, we used the initial population size vector

$$\begin{pmatrix} n_{V,1}^J \\ n_{V,1}^A \\ n_{P,1}^J \\ n_{P,1}^A \end{pmatrix} = \begin{pmatrix} 100 \\ 100 \\ 20 \\ 20 \end{pmatrix}$$

181 , a study period of $T = 30$ years, the yearly number of monitored prey and predator broods/litters
 182 respectively $R_t^V = 50$ and $R_t^P = 20$, and the yearly number of marked juveniles was 100 for
 183 both species. Results using the monitoring setups of Barraquand & Gimenez (2019) with either
 184 100 marked juveniles per species per year for $T = 10$ years, or 20 marked juveniles per species
 185 per year for $T = 30$ years (and the non centered density-dependencies) are also presented in the

186 Supplementary Information B.

187 We consider two alternative situations without interspecific interactions: with or without ran-
188 dom temporal noise. To compare model performances in the no-interactions setting to cases with
189 interspecific interactions, we also simulated and fitted data in presence of species interactions using
190 the same α_i values as Barraquand & Gimenez (2019) under the four above-mentioned scenarios (i.e.,
191 with/without interactions \times with/without stochasticity on interactions; see Supplementary Table 2
192 and Figures S1 and S3 in addition to main text results). For each of these four combinations of
193 parameter sets, we simulated 100 datasets using the Nimble package (de Valpine *et al.*, 2017, 2022,
194 version 0.12.2) in R (R Core Team, 2022, version 4.2.1).

195 2.3 Priors specification and model fitting

196 Multispecies IPMs were implemented in a Bayesian framework, hence the need to specify priors.
197 When fitting the models to simulated data, we used $\mathcal{N}(100, 10)$ and $\mathcal{N}(20, 10)$ priors for the initial
198 stage-specific prey and predator population sizes (truncated to be positive). These priors also
199 differed from the 2019 model where they were all set to $\mathcal{N}(25, 10^{-5})$.

200 Priors for standard deviations were chosen as $\sigma \sim \text{Exp}(1)$, which corresponds to priors with max-
201 imum entropy on the log and logit scales (e.g., McElreath, 2020). Prior probabilities of recapture
202 were drawn as $p \sim \text{Unif}(0, 1)$ and vital rate/interaction parameters were given weakly informative
203 priors $\alpha_k \sim \mathcal{N}(0, 1)$ ($k \in \{1, \dots, 8\}$).

204 Data were both simulated and fitted using the Nimble R package (R Core Team, 2022; de
205 Valpine *et al.*, 2017, 2022, version 0.12.2). To improve their mixing and minimize their posterior
206 correlations, intercepts, slopes and temporal SD were block sampled using automated factor slice
207 samplers (Tibbits *et al.*, 2014; Ponisio *et al.*, 2020). For each simulated dataset, we fitted the same
208 multispecies IPM that was used to generate the data (e.g., no random temporal noise estimated on
209 data without temporal noise), except in that species interactions were estimated even in absence of
210 such interactions. Two MCMC chains were run for 60200 iterations and we sampled the last 60000
211 iterations every 60th iteration leading to 2000 posterior samples saved per dataset. Real parameter
212 values were used as initial values to minimise time to convergence (see Appendix Section C for
213 an evaluation of the influence of initial values on parameter estimation). We assess convergence
214 and mixing of the chains for all α_i by calculating the potential scale reduction factor (\hat{R} , Brooks &
215 Gelman 1998; Gelman & Rubin 1992) and effective sample size ($n_{eff.}$) using the "gelman.diag()" and
216 the "effectiveSize()" functions of the *coda* package (Plummer *et al.*, 2006, version 0.19-4). We only

217 used outputs from models for which all α_i had $\hat{R} < 1.1$ and $n_{eff.} > 50$, that is, 100/100 models for
218 the scenario without random temporal variation and 94/100 models for the scenario with random
219 temporal variation. The computer code is provided at [https://github.com/MatthieuPaquet/
220 multi_species](https://github.com/MatthieuPaquet/multi_species).

221 **3 Results**

222 Overall, estimates of density dependence curves were unbiased, regarding interspecific density de-
223 pendence (either absent, Figures 1 and 3, or present, Figures S1 and S3) as well as intraspecific
224 density dependence. This was true without and with temporal stochasticity (Figures 1 to 4).

225 This absence of bias extends to the alternative data designs with smaller sample sizes considered
226 in Barraquand & Gimenez (2019) (shown in Supplementary Information in Figures S5 to S8).
227 Estimated α_i parameters also did not show sign of bias in any scenario (Table 2 and Table S1).

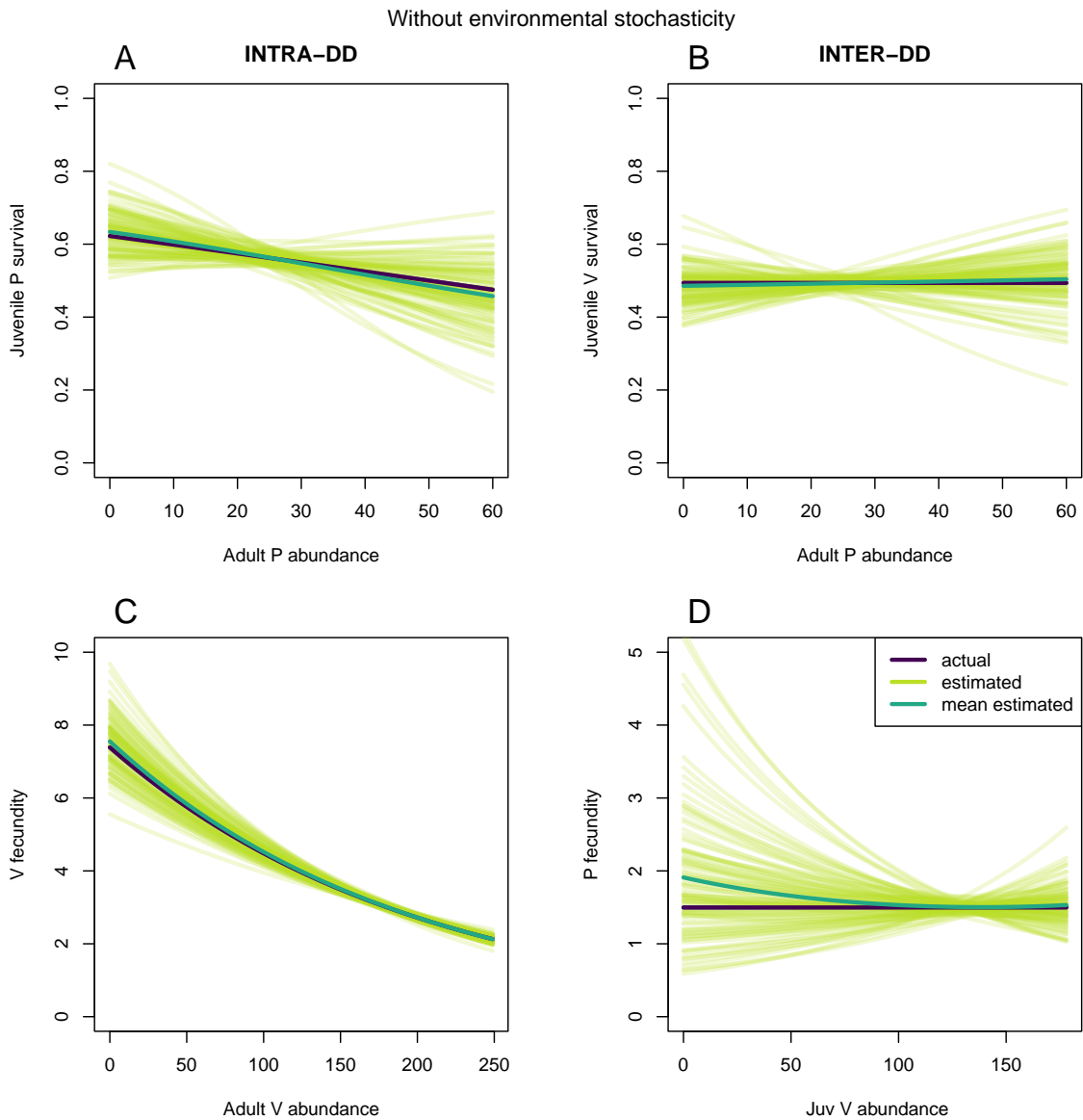


Figure 1: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario without random time variation. Purple: simulated relationships, light green: posterior mean relationships for all 100 fitted models, dark green: average of the posterior mean relationships. True inter species density-dependencies (right panels) were set to be absent.

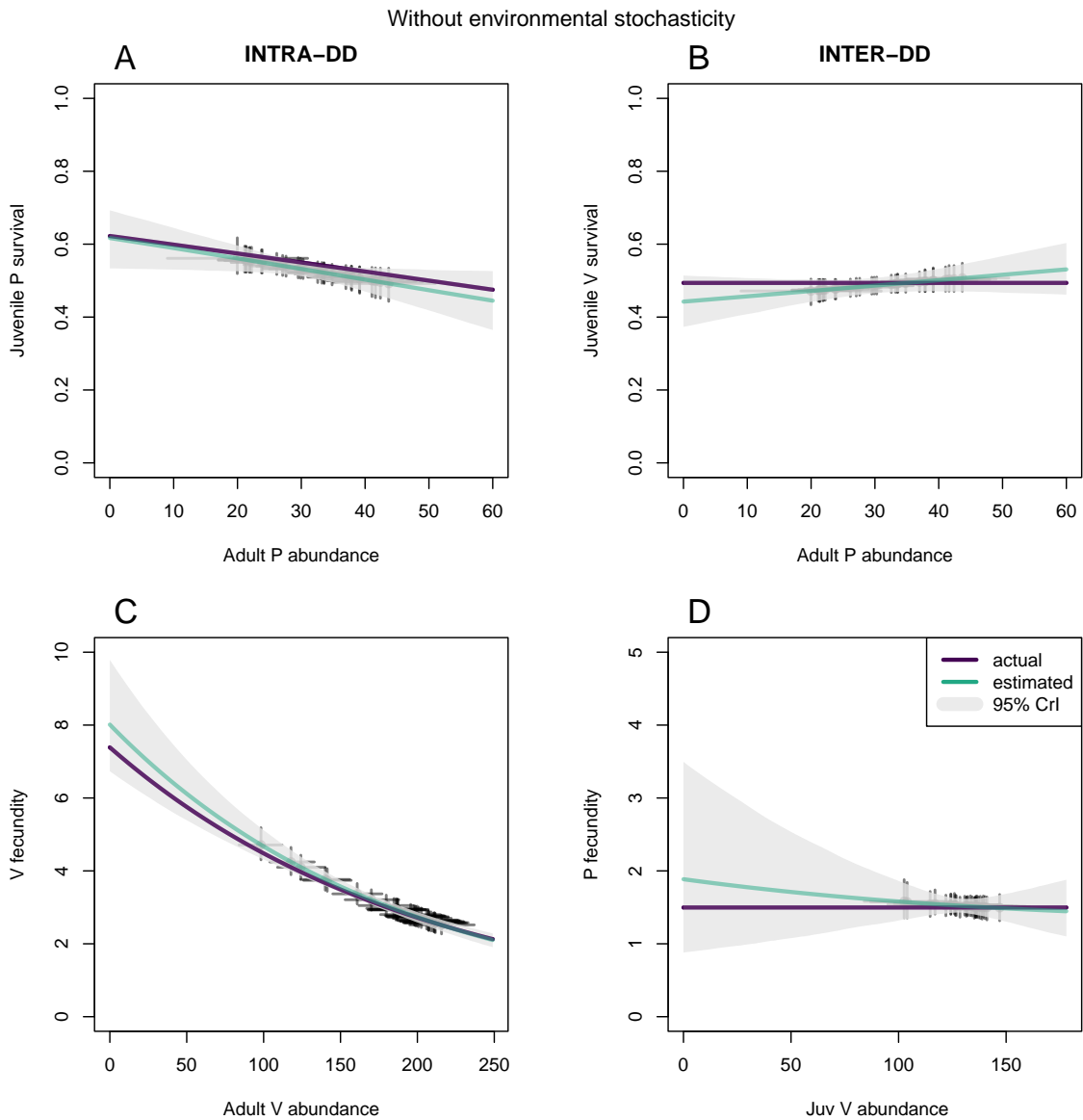


Figure 2: Example of posterior mean (blue-green line) and 95% Credible Intervals (grey polygons) of density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities estimated by one of the 100 models run in the scenario without random time variation. Purple lines indicate the simulated (true) relationships. Points represent estimated mean demographic parameter each year plotted against estimated yearly abundance values and vertical and horizontal error bars their respective 95% Credible Intervals.

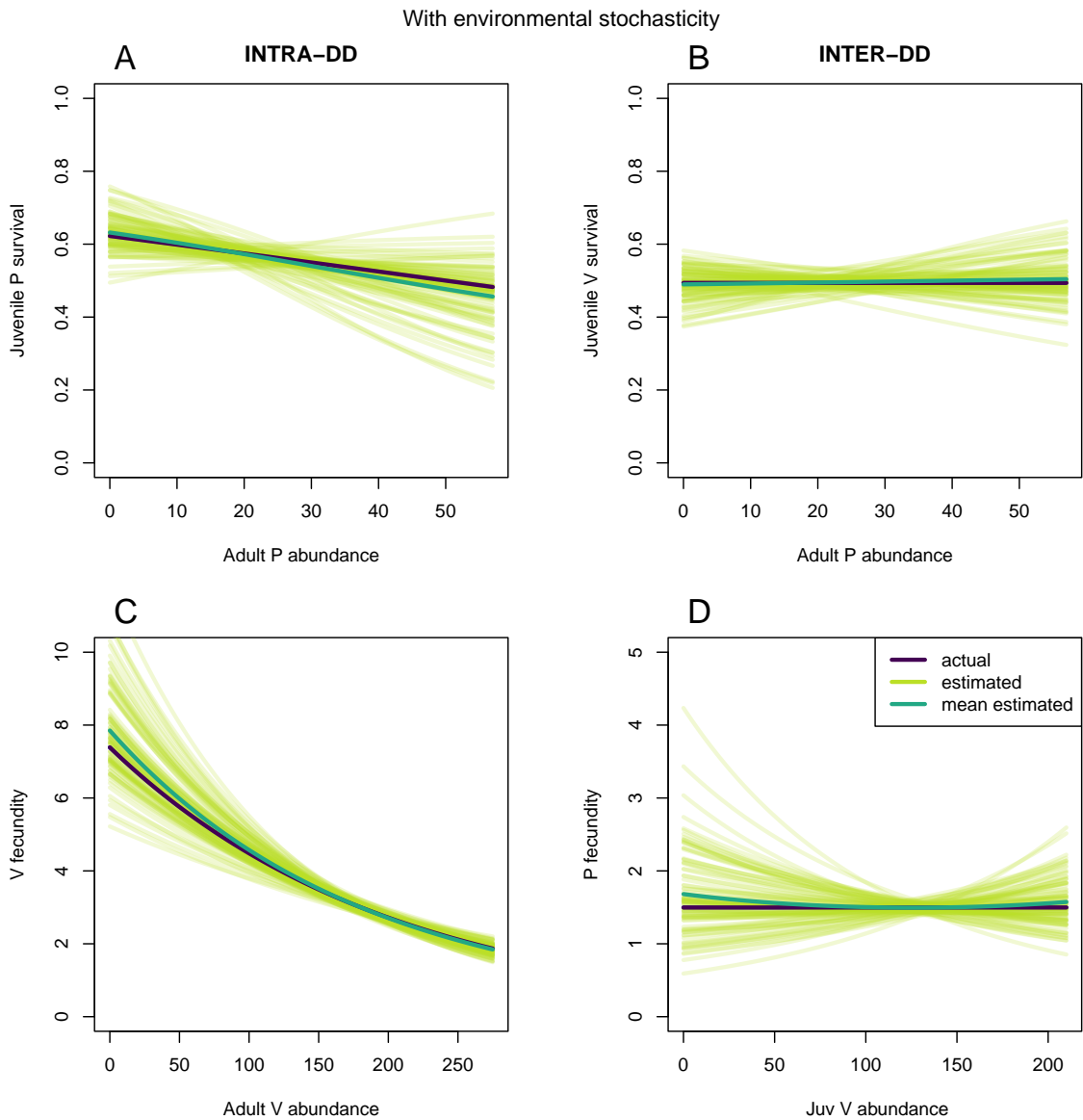


Figure 3: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario with random time variation. Purple: simulated relationships, light green: posterior mean relationships for the 94 fitted models that appear to converge satisfactorily, dark green: average of the posterior mean relationships. True inter species density-dependencies (right panels) were set to be absent.

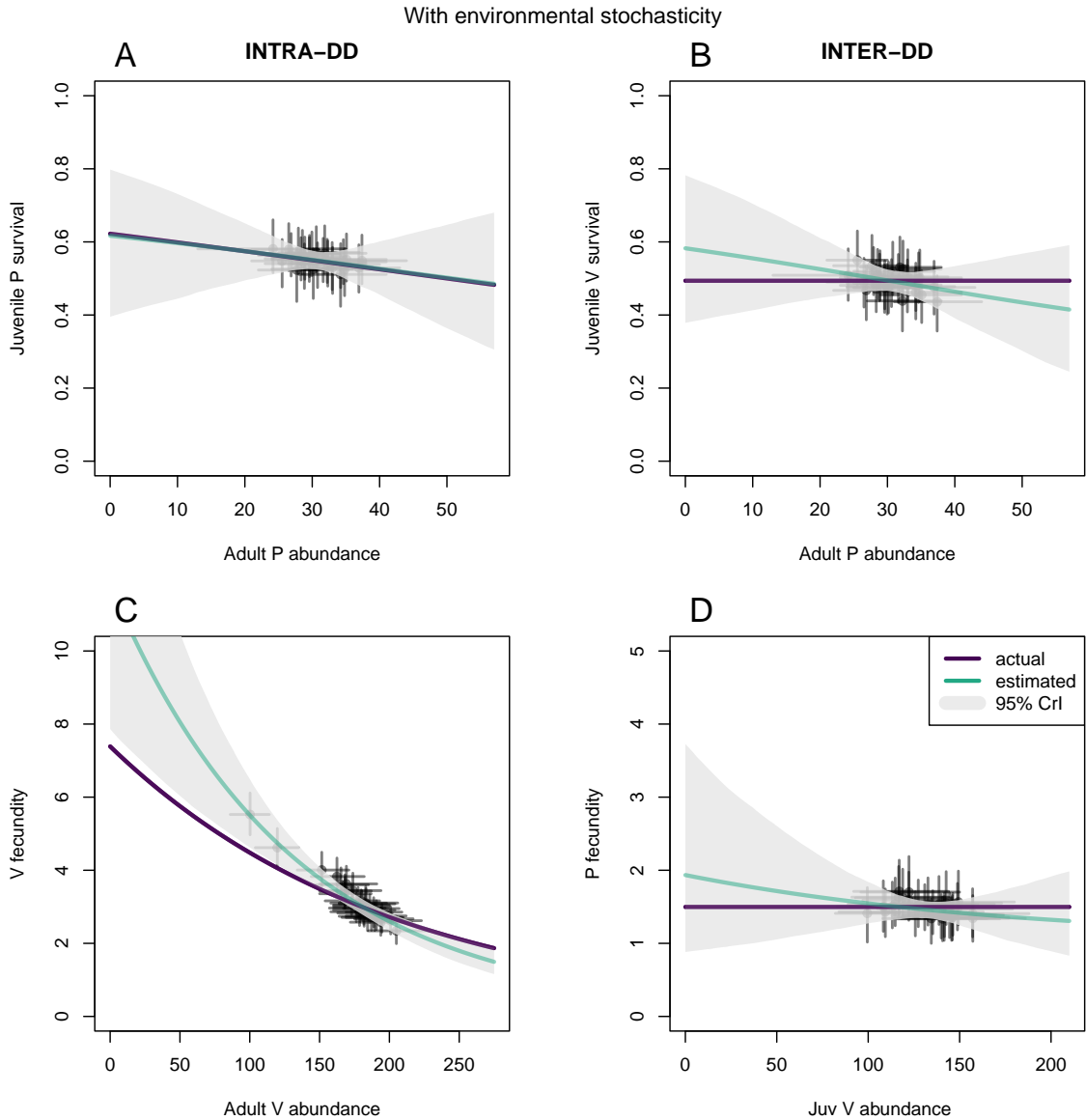


Figure 4: Example of posterior mean (blue-green line) and 95% Credible Intervals (grey polygons) of density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities estimated by one of the 100 models run in the scenario with random time variation. Purple lines indicate the simulated (true) relationships. Points represent estimated mean demographic parameter each year plotted against estimated yearly abundance values and vertical and horizontal error bars their respective 95% Credible Intervals.

228 We did not detect more false positive species interactions than expected by chance when inves-
 229 tigating the coverage probability of the species interaction parameters at 95% (i.e., the proportion
 230 of simulations where 95% CrI of estimated parameter includes the true parameter value). In the
 231 scenario with 100 juveniles marked each year for 30 years and no interspecific density dependence
 232 nor temporal random variation, this probability was 0.95 for α_4 and 0.92 for α_6 (cf Table 2, see
 233 Figure 2 for an example of estimated mean and pointwise 95% CrI density dependent curves).

234 Coverage probabilities were also satisfactory when interspecific interactions were simulated to be
235 nonzero (0.94 and 0.93). Species interactions parameters were still estimable with no noticeable
236 bias in the presence of random time variation (Figures 3 and 4), in which case the coverage prob-
237 abilities of the species interaction parameters α_4 and α_6 at 95% were 0.99 and 0.98 respectively in
238 absence of interspecific interactions (Table 2). In the presence of interspecific interactions, coverage
239 values were both 0.96. Moreover, the addition of random time variation did not noticeably alter the
240 precision of the species interaction parameters, both in absence and presence of species interactions
241 (Figure S3, Table 2).

Table 2: Summary table of parameter estimates. Value refers to the true values used to simulate the data and values of the interspecific density dependent parameters are highlighted in bold. Estimate (95% quantiles) are the mean and the 95% quantiles of the posterior mean estimates. Coverage 95% is the proportion of 95% Credible Intervals that included the true parameter values.

Scenario	Param.	Value	Estimate (95% quantiles)	Coverage 95%
30 years 100 ind. marked/year No temporal noise No interspecies DD	α_1	0.29	0.304 (0.166; 0.482)	0.97
	α_2	-0.01	-0.013 (-0.032; 0.005)	0.95
	α_3	-0.025	-0.033 (-0.144; 0.083)	0.99
	α_4	0	0.001 (-0.017; 0.019)	0.95
	α_5	0.404	0.413 (0.199; 0.635)	0.93
	α_6	0	0 (-0.008; 0.007)	0.92
	α_7	1.24	1.243 (1.198; 1.287)	0.97
	α_8	-0.005	-0.005 (-0.006; -0.004)	0.96
30 years 100 ind. marked/year Temporal noise No interspecies DD	α_1	0.29	0.281 (0.124; 0.428)	0.947
	α_2	-0.01	-0.013 (-0.038; 0.003)	0.947
	α_3	-0.025	-0.02 (-0.173; 0.122)	0.947
	α_4	0	0.001 (-0.012; 0.017)	0.989
	α_5	0.404	0.399 (0.236; 0.532)	0.968
	α_6	0	0 (-0.005; 0.004)	0.979
	α_7	1.24	1.244 (1.17; 1.319)	0.968
	α_8	-0.005	-0.005 (-0.007; -0.004)	0.968
30 years 100 ind. marked/year No temporal noise Interspecies DD	α_1	0.29	0.282 (0.122; 0.427)	0.97
	α_2	-0.01	-0.011 (-0.027; 0.004)	0.98
	α_3	-0.025	-0.019 (-0.156; 0.125)	0.99
	α_4	-0.025	-0.026 (-0.042; -0.009)	0.94
	α_5	0.404	0.395 (0.264; 0.502)	0.94
	α_6	0.004	0.004 (-0.001; 0.01)	0.93
	α_7	1.24	1.241 (1.195; 1.281)	0.95
	α_8	-0.005	-0.005 (-0.006; -0.004)	0.96
30 years 100 ind. marked/year Temporal noise Interspecies DD	α_1	0.29	0.29 (0.143; 0.459)	0.967
	α_2	-0.01	-0.01 (-0.026; 0.007)	0.989
	α_3	-0.025	-0.014 (-0.176; 0.145)	0.967
	α_4	-0.025	-0.025 (-0.041; -0.007)	0.957
	α_5	0.404	0.403 (0.276; 0.523)	0.957
	α_6	0.004	0.004 (-0.001; 0.009)	0.957
	α_7	1.24	1.237 (1.179; 1.311)	0.913
	α_8	-0.005	-0.005 (-0.008; -0.004)	0.924

242 4 Discussion

243 Building on the multispecies integrated predator-prey model of [Barraquand & Gimenez \(2019\)](#),
244 we investigated here whether multispecies IPMs could output interactions where there are in fact
245 none. We did so by modelling functions relating vital rates to stage-specific species densities, whose
246 slope parameters are used to model species interactions. We found that when those slopes were
247 simulated as zero, the estimates were centered on zero and therefore unbiased. There was also a
248 good coverage probability of interaction parameters (close to 0.95 for 95% CrIs). We also found
249 that adding temporal variability to these multispecies density-dependent link functions did not
250 alter these results. This confirms that multispecies IPMs are a promising way to estimate species
251 interactions, and in particular, that they could be used to infer whether two species interact or not
252 when such information is missing.

253 These results are encouraging, though some readers might find our sample sizes relatively large
254 (see [Appendix B](#) for slightly lower sample sizes). In a previous version of this work, we inadvertently
255 omitted the θ^A CMR array in the code, which transformed the model into a capture-removal model
256 (i.e., individuals were re-captured only once and then removed from the population, as in hunting
257 or fishing data). In this configuration, the lower amount of data on survival and detection provided
258 proper estimation of all quantities for the main text data design but not those of [Appendix B](#)
259 for which convergence was not always reached. With live capture-recapture data, all data designs
260 (main text and [Appendix B](#)) now provide satisfactory convergence and estimation. Moreover, in
261 field population studies, additional types of data available are likely to improve the estimation of
262 species interactions and we give three examples below. First, when age classes can be determined
263 during the count observation process, including such information explicitly in the model (see e.g.,
264 [Weegman *et al.*, 2016](#); [Paquet *et al.*, 2019](#)) will increase identifiability and/or precision of survival
265 parameters and age specific abundances, and therefore will likely improve the estimation of density
266 dependence parameters as well. This stage-specific abundance information may also allow, in some
267 cases where counts are provided with little error, to remove the observation process, which we
268 cannot do in our current model formulation because the observed population size sums adult and
269 juvenile densities, and this sum has to arise from a probability distribution (Equations (2) and (3)).
270 Second, integrating dead prey recovery data is likely to give extra information on the strength of
271 predator-prey interactions. Dead recoveries are classically implemented in capture-mark-recovery
272 models ([Seber, 1972](#); [North & Morgan, 1979](#)) which in some cases can be combined with CMR

273 data (Barker, 1999) and counts (Reynolds *et al.*, 2009). Since the probability to find a dead prey
274 is likely affected by predation rates in the population (e.g., in some systems prey eaten will not be
275 recovered, in others dead recoveries may present signs of predation), taking the predation process
276 into account in the dead recoveries data-generation mechanism could improve the estimation of the
277 strength of predator-prey interactions. Finally, the spatial structure of the data should contain
278 additional information that may help to estimate parameters. The extension to spatially explicit
279 IPMs (Chandler & Clark, 2014; Zhao, 2020) for interacting populations represents a promising way
280 forward for the estimation of species interactions.

281 We commented above on the amount of data and possible additional data types. However, the
282 efficiency of multispecies IPMs in estimating species interactions may also depend on the parameter
283 set, and thus on the ecological features of the populations studied. For example, the parameters
284 considered here correspond well to vertebrate predator-prey systems with a stable equilibrium in
285 absence of environmental perturbations. Faster life histories, different stage or age structure, and
286 multiple factors contributing to altering the quantity of information encapsulated in the various
287 data streams may alter the sample sizes required for efficient inferences. When applying these
288 models to new systems with different life history parameters and density-dependent structures
289 (e.g., predators also eating adult prey), simulated datasets with plausible ecological features for the
290 empirical system considered (and similar data designs), will help confirm that parameter values can
291 be recovered without bias and with sufficient precision. Tools such as JAGS (Plummer *et al.*, 2003)
292 or Nimble (de Valpine *et al.*, 2017) make it particularly handy to both simulate and fit data with
293 complex dynamic models.

294 Finally, while using the same model to simulate and fit the data is a necessary first step to (i)
295 assess the identifiability of model parameters (and assess the amount and type of data needed for
296 practical identifiability), (ii) evaluate the coverage of parameter estimates, and (iii) check for bias
297 in the estimates that can still occur, notably because of limited sample sizes (Paquet *et al.*, 2021),
298 an important next step will be to evaluate the sensitivity of multi-species IPM estimates to model
299 mis-specifications (Plard *et al.*, 2021). For example, different functions than the log and logit links
300 chosen here may be used to fit or to simulate intra- and inter-specific density-dependencies. Hence,
301 we encourage future work to try fitting a broader range of plausible models that differ from the
302 model used to simulate the data (or conversely, to simulate from more mechanistic models) in order
303 to assess such sensitivity.

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308 **Data and code**

309 Computer R code used to generate and analyze the data is available at [https://github.com/](https://github.com/MatthieuPaquet/multi_species)
310 [MatthieuPaquet/multi_species](https://github.com/MatthieuPaquet/multi_species) and the generated data and figures are available at [https://](https://osf.io/xf6e/)
311 osf.io/xf6e/ and linked to the code repository.

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314 **Conflict of interest**

315 The authors of this preprint declare that they have no financial conflict of interest with respect to
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317 Ecology.

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402 **Supplementary Information**

403 **A Results from the scenarios in presence of species interactions**

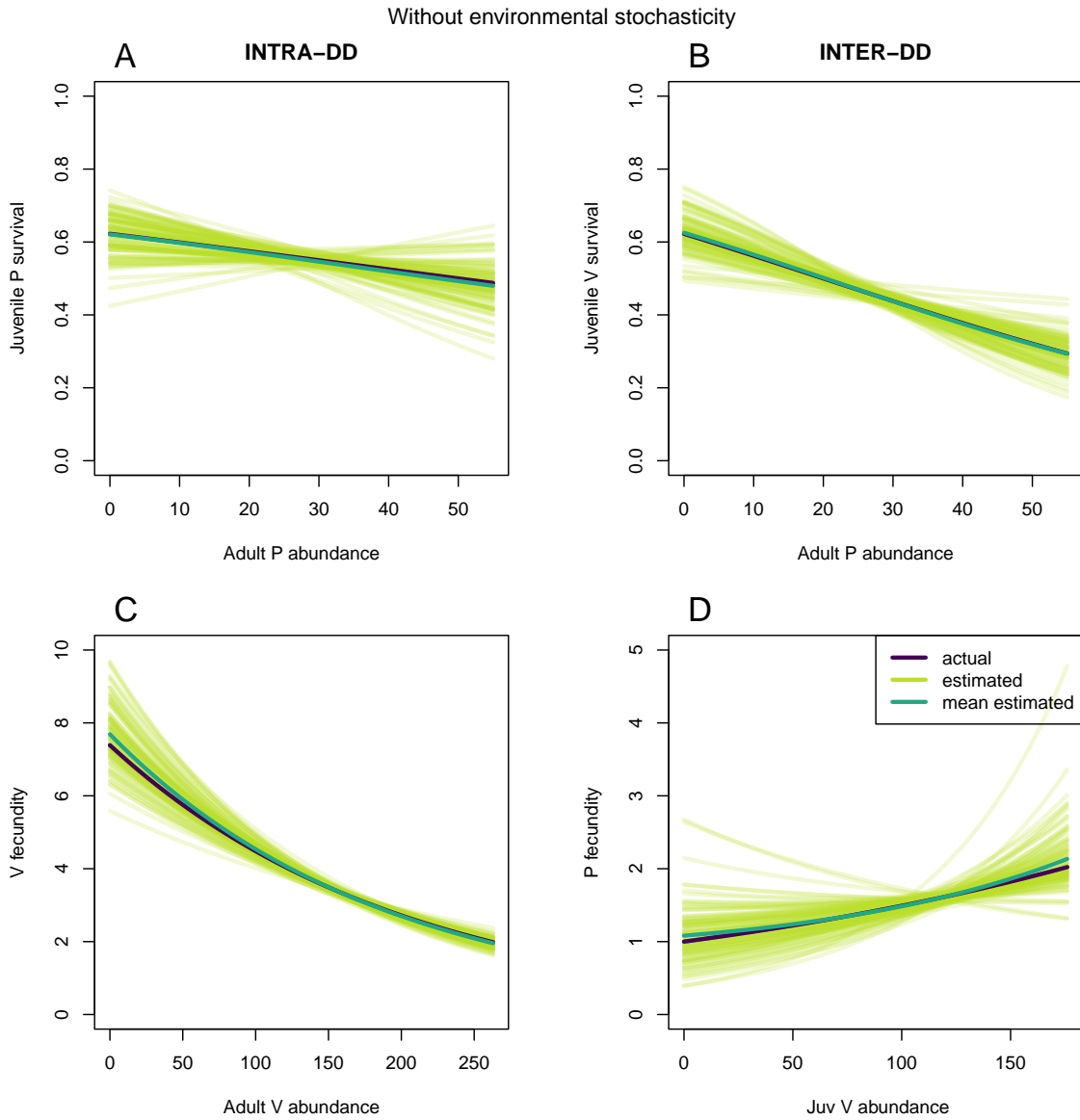


Figure S1: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario without random time variation in presence of true inter species density-dependencies. Purple: simulated relationships, light green: posterior mean relationships for the 100 fitted models that appear to converge satisfactorily, dark green: average of the posterior mean relationships.

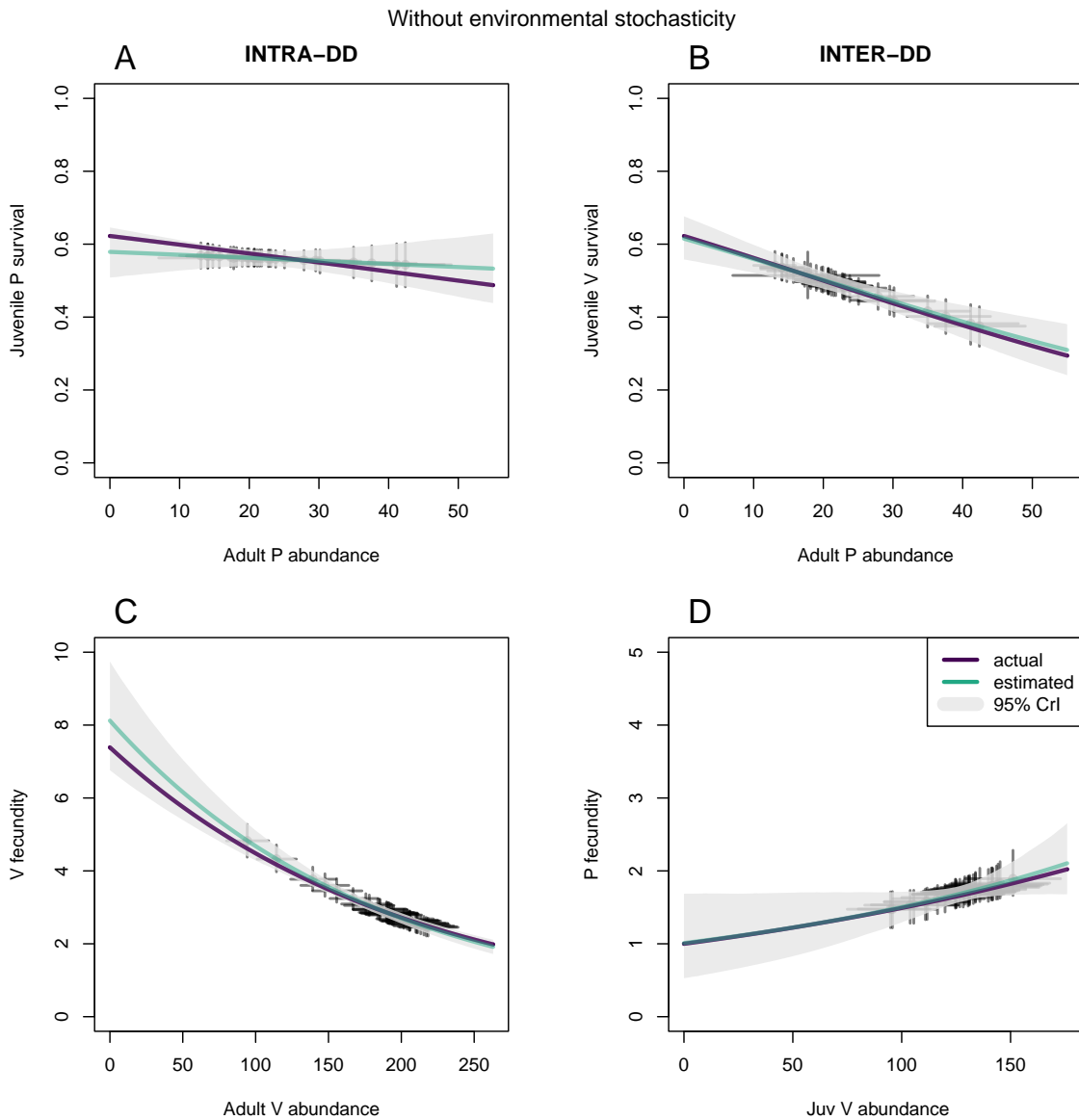


Figure S2: Example of posterior mean (blue-green line) and 95% Credible Intervals (grey polygons) of density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities estimated by one of the 100 models run in the scenario without random time variation in presence of true inter species density-dependencies. Purple lines indicate the simulated (true) relationships. Points represent estimated mean demographic parameter each year plotted against estimated yearly abundance values and vertical and horizontal error bars their respective 95% Credible Intervals.

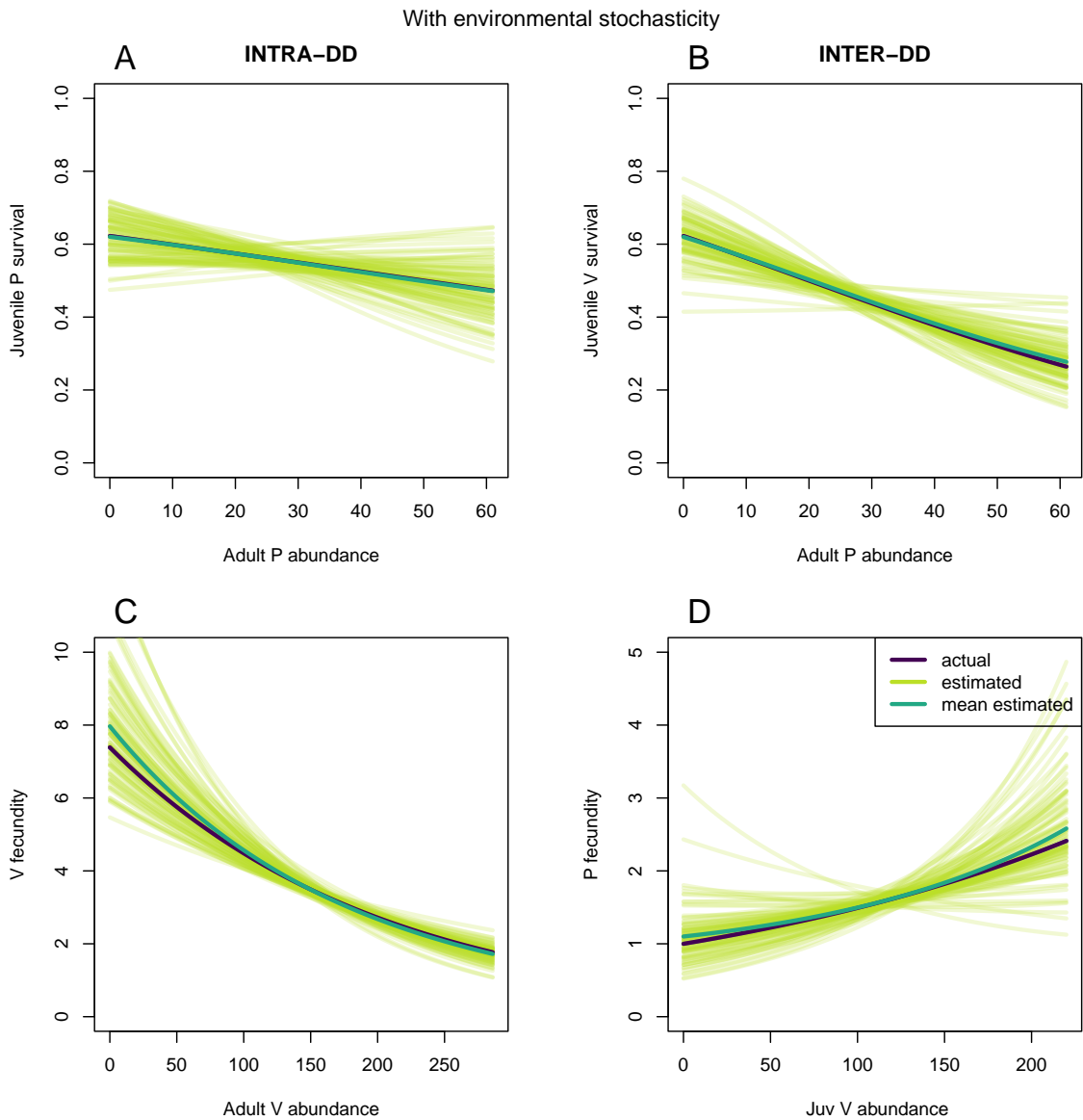


Figure S3: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario with random time variation in presence of true inter species density-dependencies. Purple: simulated relationships, light green: posterior mean relationships for the 92 fitted models that appear to converge satisfactorily, dark green: average of the posterior mean relationships.

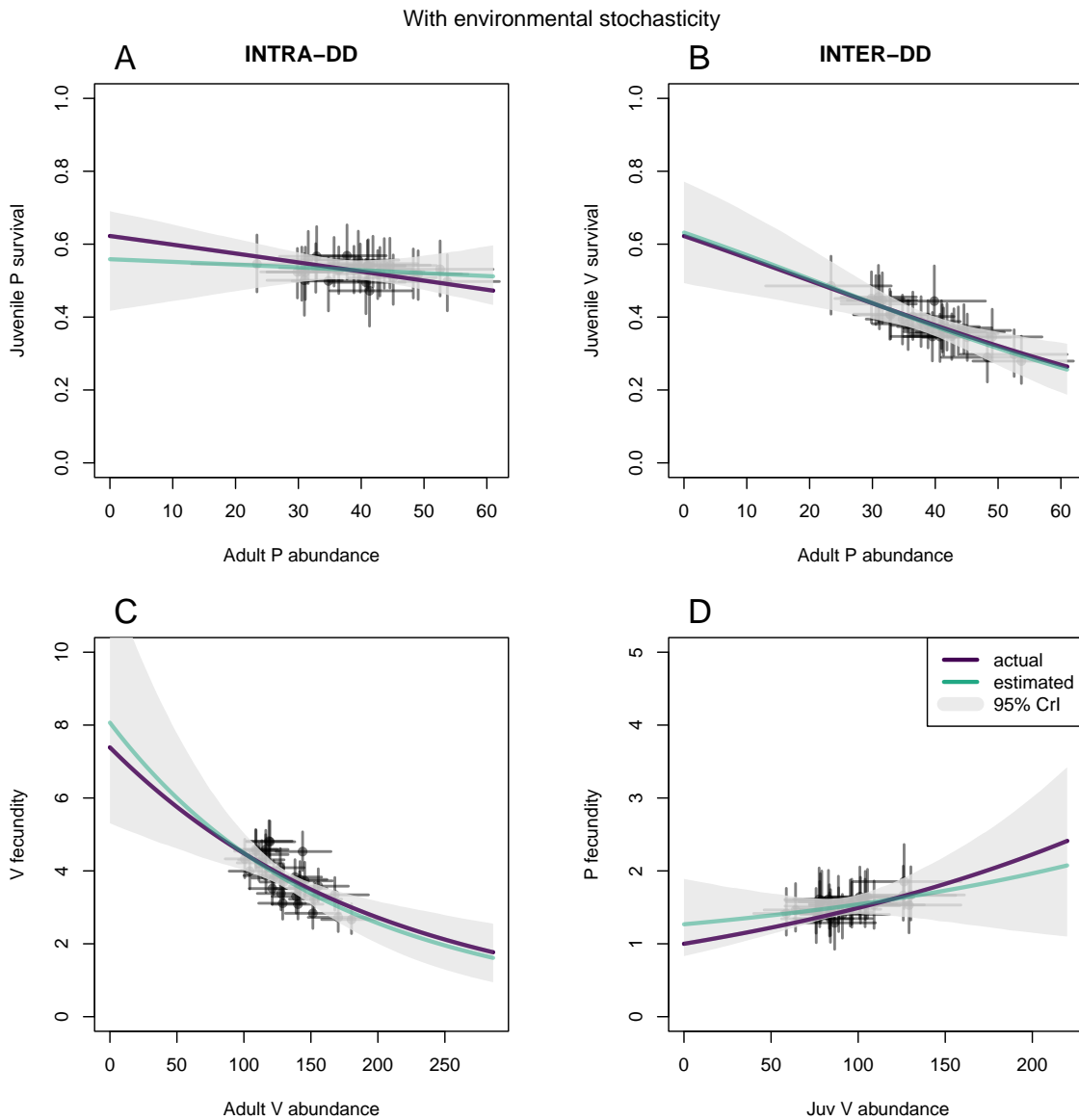


Figure S4: Example of posterior mean (blue-green line) and 95% Credible Intervals (grey polygons) of density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities estimated by one of the 100 models run in the scenario with random time variation in presence of true inter species density-dependencies. Purple lines indicate the simulated (true) relationships. Points represent estimated mean demographic parameter each year plotted against estimated yearly abundance values and vertical and horizontal error bars their respective 95% Credible Intervals.

404 **B Results from the scenarios with 100 juveniles of each species marked each year**
 405 **for 10 years, and 20 juveniles of each species marked for 30 years, without**
 406 **centering abundances in the link functions**

407 The results presented below follow the data design of [Barraquand & Gimenez \(2019\)](#).

Table S1: Value refers to the true values used to simulate the data and values of the interspecific density dependent parameters are highlighted in bold. Estimate (95% quantiles) are the mean and the 95% quantiles of the posterior mean estimates. Coverage 95% is the proportion of 95% Credible Intervals that included the true parameter values.

Scenario	Param.	Value	Estimate (95% quantiles)	Coverage 95%
10 years 100 ind. marked/year No temporal noise	α_1	0.5	0.458 (-0.545; 1.293)	0.98
	α_2	-0.01	-0.008 (-0.041; 0.031)	0.99
	α_3	-0.025	-0.045 (-0.78; 0.563)	0.99
	α_4	0	0.001 (-0.026; 0.033)	0.98
	α_5	0.404	0.286 (-0.577; 1.098)	0.949
	α_6	0	0.001 (-0.005; 0.008)	0.96
	α_7	2	1.998 (1.773; 2.227)	0.98
	α_8	-0.005	-0.005 (-0.006; -0.004)	0.97
10 years 100 ind. marked/year Temporal noise	α_1	0.5	0.278 (-0.361; 0.874)	0.99
	α_2	-0.01	-0.001 (-0.028; 0.029)	0.99
	α_3	-0.025	-0.023 (-0.594; 0.614)	1
	α_4	0	0 (-0.025; 0.023)	1
	α_5	0.404	0.323 (-0.379; 0.918)	0.99
	α_6	0	0.001 (-0.004; 0.006)	1
	α_7	2	1.911 (1.467; 2.323)	0.948
	α_8	-0.005	-0.004 (-0.007; -0.002)	0.927
30 years 20 ind. marked/year No temporal noise	α_1	0.5	0.56 (-0.155; 1.278)	0.99
	α_2	-0.01	-0.013 (-0.045; 0.021)	0.99
	α_3	-0.025	-0.031 (-0.529; 0.436)	0.97
	α_4	0	0.001 (-0.024; 0.023)	0.97
	α_5	0.404	0.329 (-0.305; 1.009)	0.97
	α_6	0	0.001 (-0.005; 0.006)	0.98
	α_7	2	2.009 (1.832; 2.194)	0.96
	α_8	-0.005	-0.005 (-0.006; -0.004)	0.97
30 years 20 ind. marked/year Temporal noise	α_1	0.5	0.527 (-0.275; 1.206)	0.968
	α_2	-0.01	-0.011 (-0.041; 0.019)	0.968
	α_3	-0.025	-0.007 (-0.528; 0.533)	0.979
	α_4	0	0 (-0.02; 0.017)	0.989
	α_5	0.404	0.316 (-0.191; 0.823)	0.968
	α_6	0	0.001 (-0.004; 0.004)	0.968
	α_7	2	1.954 (1.702; 2.247)	0.957
	α_8	-0.005	-0.005 (-0.006; -0.003)	0.957

10 years, without environmental stochasticity

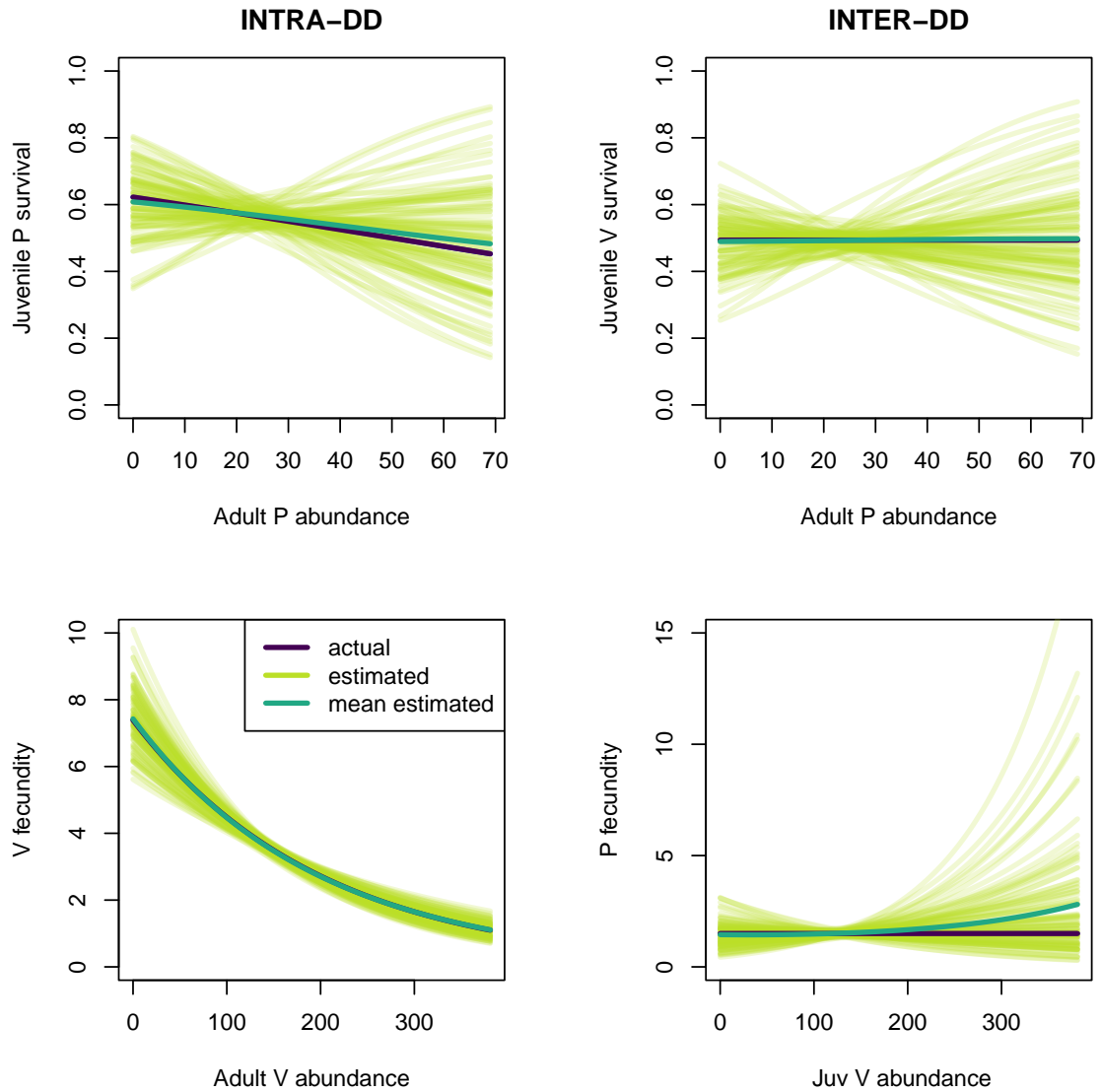


Figure S5: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario with 100 juveniles per species marked each year for 10 years without random time variation in absence of true inter species density-dependencies. Purple: simulated relationships, light green: posterior mean relationships for the 99 fitted models that appear to converge satisfactorily, dark green: average of the posterior mean relationships.

10 years, with environmental stochasticity

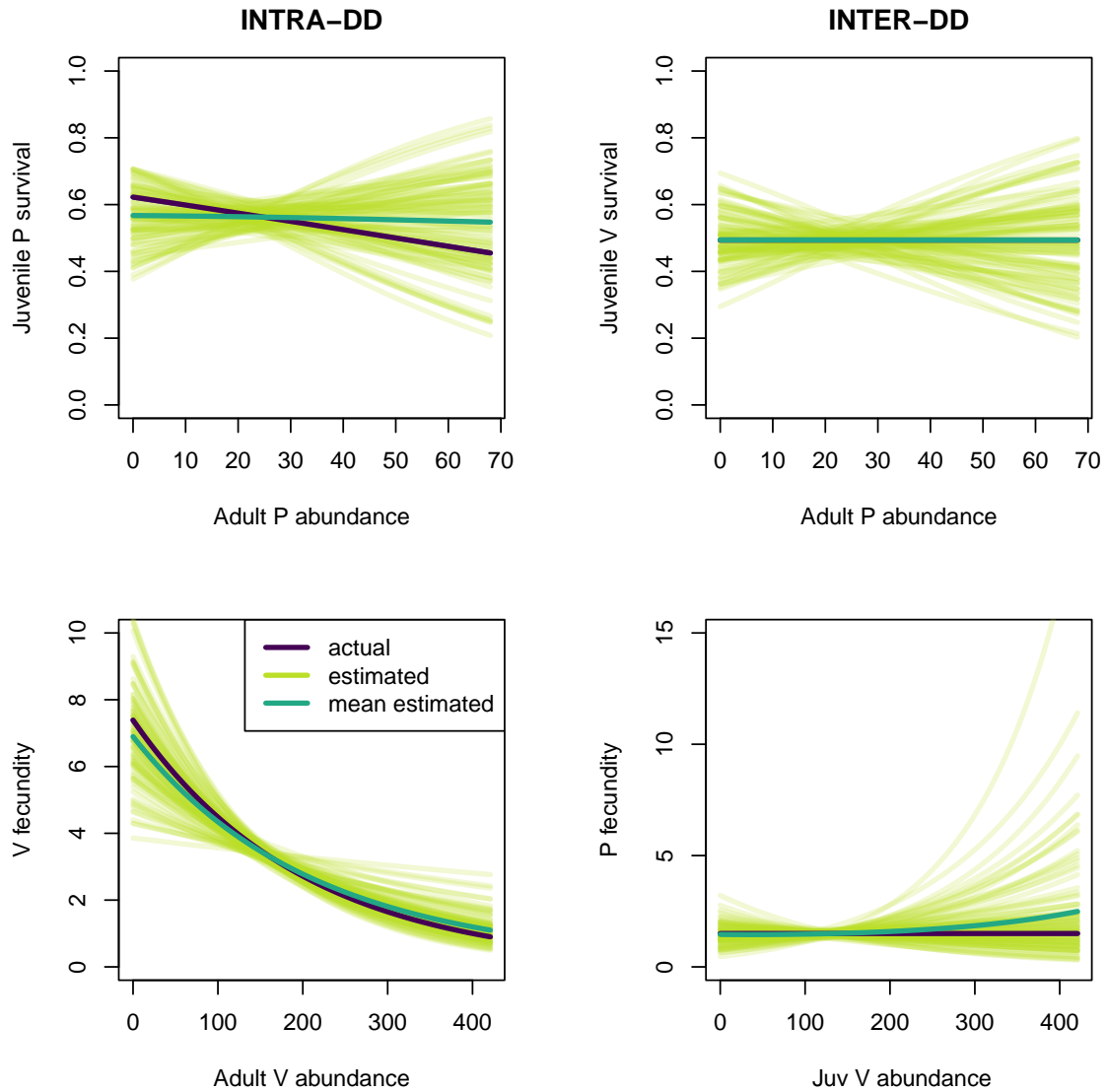


Figure S6: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario with 100 juveniles per species marked each year for 10 years with random time variation in absence of true inter species density-dependencies. Purple: simulated relationships, light green: posterior mean relationships for the 96 fitted models that appear to converge satisfactorily, dark green: average of the posterior mean relationships.

30 years, without environmental stochasticity

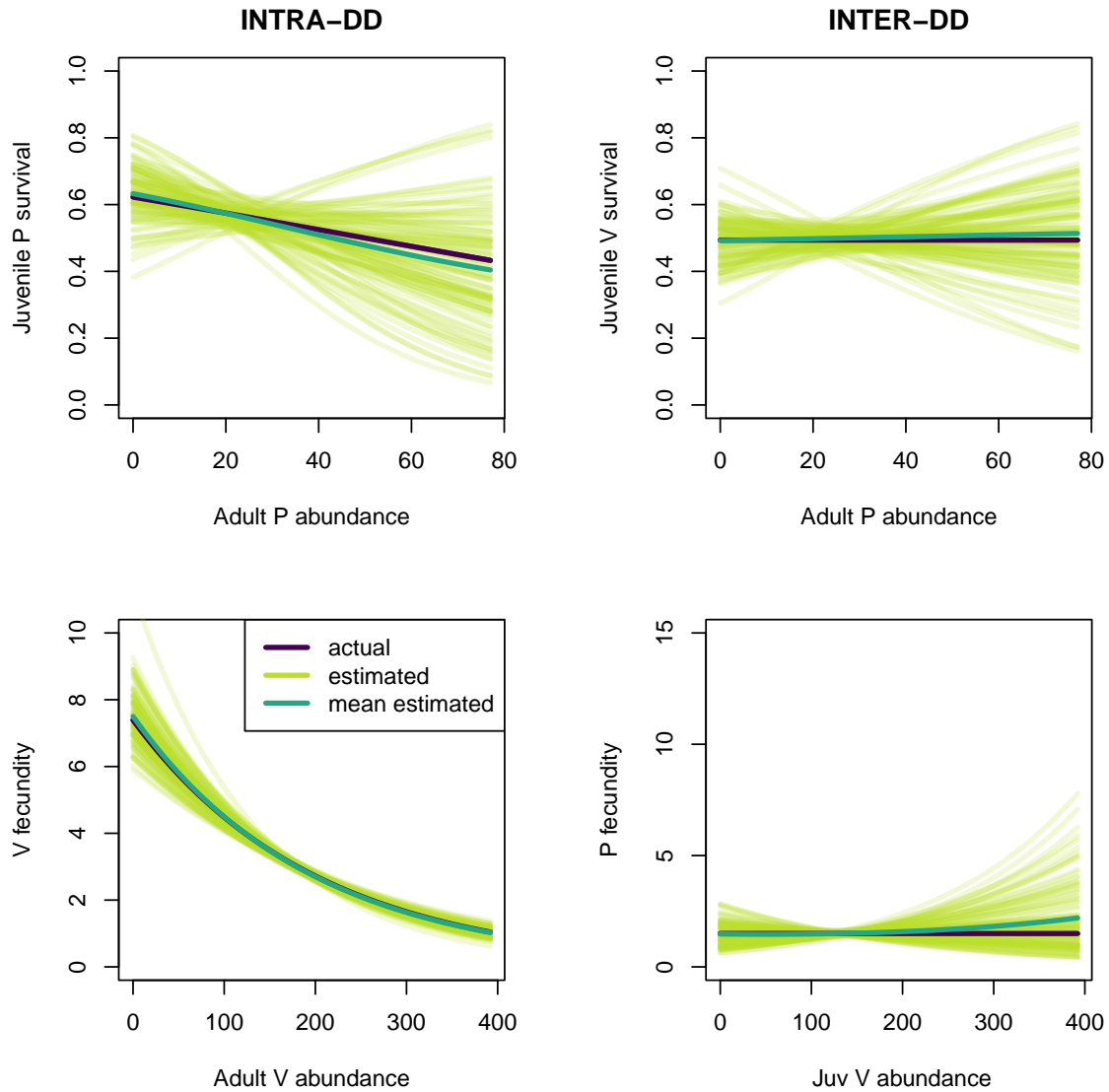


Figure S7: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario with 20 juveniles per species marked each year for 30 years without random time variation in absence of true inter species density-dependencies. Purple: simulated relationships, light green: posterior mean relationships for the 100 fitted models that appear to converge satisfactorily, dark green: average of the posterior mean relationships.

30 years, with environmental stochasticity

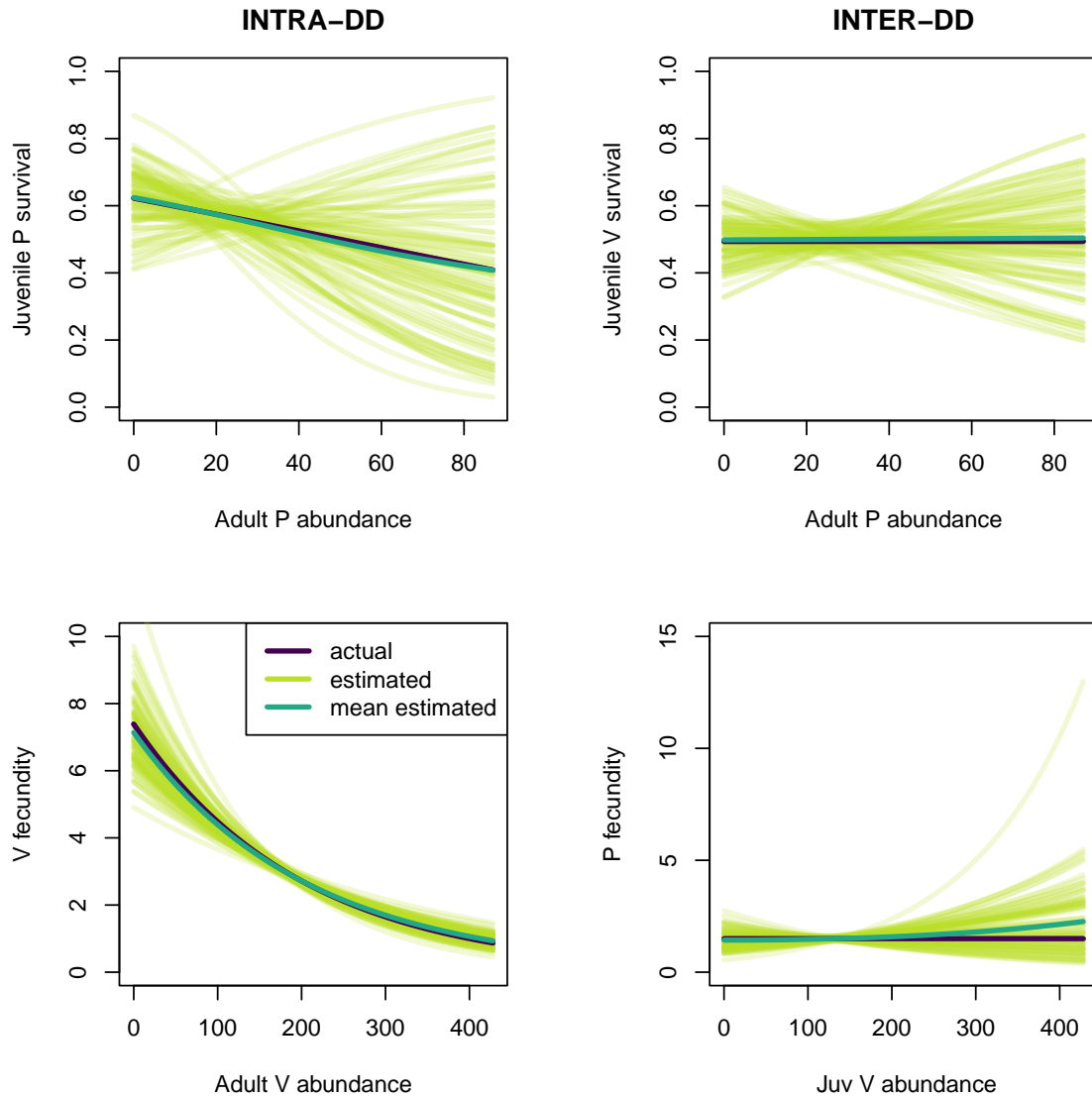


Figure S8: Density-dependencies for juvenile survival rates (**A** for predator and **B** for prey) as well as prey (**C**) and predator (**D**) fecundities in the scenario with 20 juveniles per species marked each year for 30 years with random time variation in absence of true inter species density-dependencies. Purple: simulated relationships, light green: posterior mean relationships for the 94 fitted models that appear to converge satisfactorily, dark green: average of the posterior mean relationships.

408 C Sensitivity of parameter estimation to the choice of initial values

409 To assess whether the accuracy of the estimation of density dependent parameters was condi-
410 tioned by the fact that we used true parameter values as initial values, we also ran the MCMC
411 using values that substantially deviated from the true value and expected posterior distributions.
412 For this study, we used data (and the corresponding model) without temporal random noise and
413 without true interspecific interactions. We chose one simulated dataset for which the true values
414 of α_2 , α_4 , α_6 and α_8 fell well within the 95% credible intervals of the posterior samples when
415 using the true value as initial value (see script https://github.com/MatthieuPaquet/multi_
416 [species/blob/main/script_initial_values.R](https://github.com/MatthieuPaquet/multi_species/blob/main/script_initial_values.R) for more details on the procedure). We then
417 simulated 100 sets of initial values that deviated from the true values by 4 standard deviations
418 estimated from the posterior samples when the true values were used as initial values (hereafter
419 $SD_{\hat{\alpha}_i}$). For the parameters for which negative density dependence was expected, we simulated
420 the 100 initial values as $\alpha_i^{init} \sim \mathcal{N}(\alpha_i - 4SD_{\hat{\alpha}_i}, SD_{\hat{\alpha}_i})$ whereas for α_8 , which was a potentially
421 positive prey \rightarrow predator link (and would have been assumed positive in an empirical analysis),
422 we used $\alpha_8^{init} \sim \mathcal{N}(\alpha_8 + 4SD_{\hat{\alpha}_8}, SD_{\hat{\alpha}_8})$. We used true parameter values as initial values for all
423 other model parameters. Preliminary runs showed that convergence was reached very quickly
424 (typically after a couple of iterations) with efficient mixing. We then ran 2 chains for 1200 it-
425 erations and discarded the first 200 as burn-in and did not use thinning. For comparison we
426 also run 2 MCMC chains once, under the same settings, using the true values as initial val-
427 ues (see script https://github.com/MatthieuPaquet/multi_species/blob/main/script_MCMC_
428 [simulated_initial_values_out_of_posterior.R](https://github.com/MatthieuPaquet/multi_species/blob/main/script_MCMC_simulated_initial_values_out_of_posterior.R)). The results showed no sign of influence of the
429 initial value chosen on the parameter estimates (Figure S9).

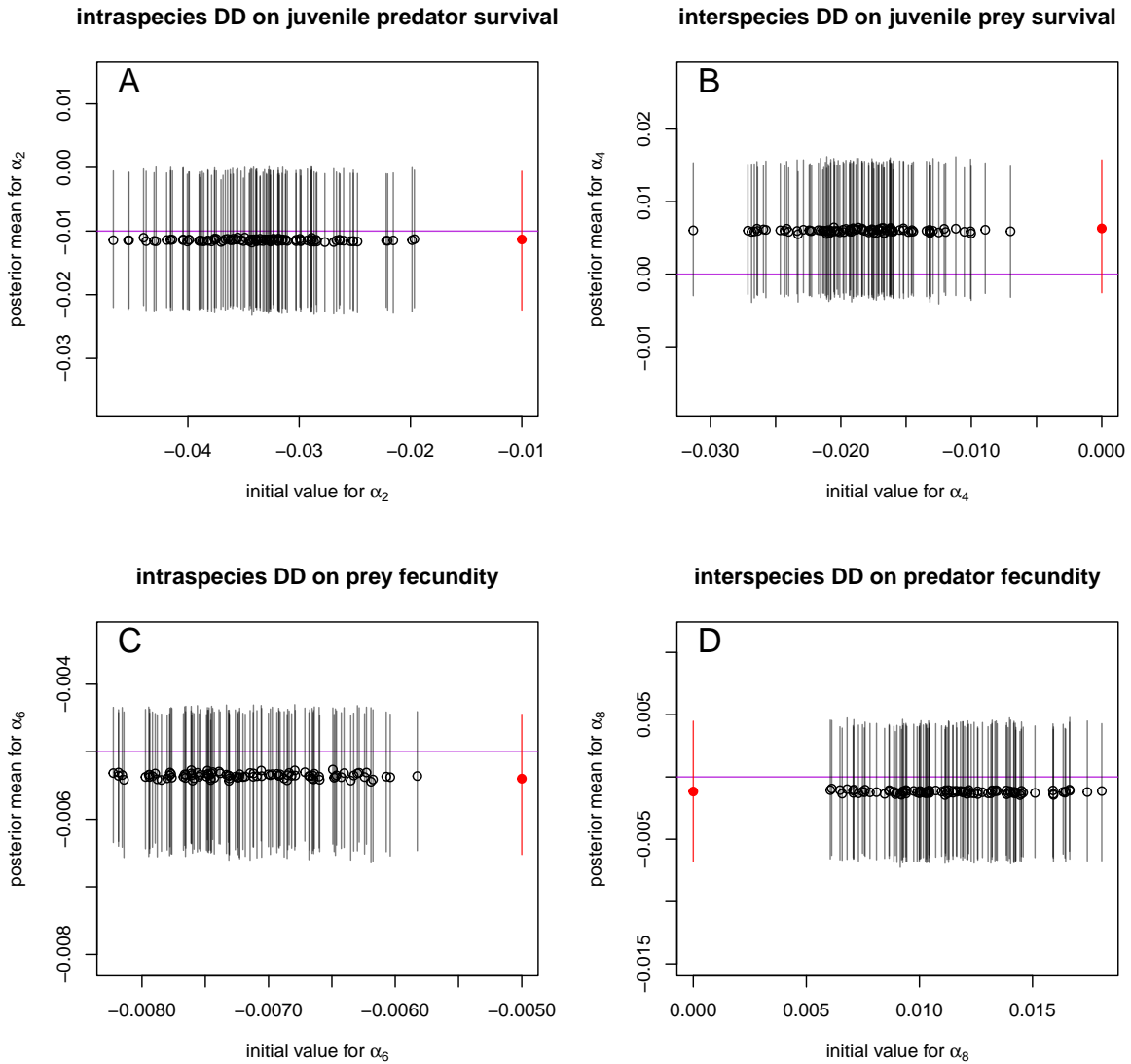


Figure S9: Estimation of density dependent parameter values (α_2 in panel **A**, α_4 in panel **B**, α_6 in panel **C** and α_8 in panel **D**) in relation to the initial values chosen to start the MCMC chains. Dots show the posterior means and vertical lines the 95% credible intervals. Purple horizontal lines highlight the value used to simulate the data. Red dots and intervals show the case where the true values are used as initial values.