

1 Deep reticulation: the long legacy of hybridization in vascular plant evolution

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13 14 SUMMARY

15 Hybridization has long been recognized as a fundamental evolutionary process in plants, but our
16 understanding of its phylogenetic distribution and biological significance across deep evolutionary scales
17 has been largely obscure—until recently. Over the past decade, genomic and phylogenomic datasets have
18 revealed, perhaps not surprisingly, that hybridization, often associated with polyploidy, has been common
19 throughout the evolutionary history of plants, particularly in various lineages of flowering plants.

20 However, phylogenomic studies have also highlighted the challenges of disentangling signals of ancient
21 hybridization from other sources of genomic conflict (in particular, incomplete lineage sorting). Here we
22 provide a critical review of ancient hybridization in vascular plants, outlining well-documented cases of
23 ancient hybridization across plant phylogeny as well as the challenges unique to documenting ancient vs.
24 recent hybridization. We provide a definition for ancient hybridization, which, to our knowledge, has not
25 been explicitly attempted before. Further documenting the extent of deep reticulation in plants should
26 remain an important research focus, especially since published examples likely represent the tip of the
27 iceberg in terms of the total extent of ancient hybridization. However, future research should increasingly
28 explore the macroevolutionary significance of this process, in terms of its impact on evolutionary
29 trajectories (e.g., how does hybridization influence trait evolution or the generation of biodiversity over
30 long time scales?), as well as how life history and ecological factors shape, or have shaped, the frequency
31 of hybridization across geologic time and plant phylogeny. Finally, we consider the implications of
32 ubiquitous ancient hybridization for how we conceptualize, analyze, and classify plant phylogeny.

33 Networks, as opposed to bifurcating trees, represent more accurate representations of evolutionary history
34 in many cases, but our ability to infer, visualize, and use networks for comparative analyses is highly
35 limited. Developing improved methods for the generation, visualization, and use of networks represents a
36 critical future direction for plant biology. Current classification systems also do not generally allow for
37 the recognition of reticulate lineages, and our classifications themselves are largely based on evidence
38 from the chloroplast genome. Updating plant classification to better reflect nuclear phylogenies, as well as
39 considering whether and how to recognize hybridization in classification systems, will represent an
40 important challenge for the plant systematics community.

41
42 **Keywords:** Ancient hybridization, angiosperms, gene flow, phylogenomics, polyploidy, reticulation,
43 vascular plants

44

45 INTRODUCTION

46 Hybridization—or crossing between individuals from different species or divergent populations (Stebbins
47 1959; Harrison 1990; Arnold 1997)—is a biological process that has fundamentally shaped the Tree of
48 Life from its tips to some of its deepest branches. The circumstances that give rise to hybridization,
49 control its frequency, and shape its evolutionary outcomes are complex and involve the interplay of
50 ecological, genomic, population genetic, and life history factors. Hybridization can counter divergence by
51 homogenizing previously isolated subpopulations. Intermittent hybridization can facilitate the sharing of
52 adaptive alleles among divergent populations or separate species. Hybridization can also lead to the
53 formation of new (reticulate) lineages through hybrid speciation, when an evolutionarily stable population
54 of hybrid individuals is established in reproductive isolation from its parental species. However, in
55 addition to these “creative” aspects of hybridization (sensu Arnold 1992), hybridization can also break
56 apart co-adaptive gene groups, cause Dobzhansky-Muller incompatibilities, and even erode the genetic
57 integrity of rare species (Todesco et al. 2016).

58 The study of hybridization has a long history in botany (Haartman 1751; Kölreuter 1766; Roberts
59 1929), and it was solidified as a fundamental process in plant evolution during the Modern Synthesis
60 (Anderson and Hubricht 1938; Anderson 1948; Stebbins 1959). The study of hybridization, including
61 hybridization in association with polyploidy (allopolyploidy), has continued to be a central focus of plant
62 biology (e.g., Grant 1971; Soltis and Soltis 2009; Yakimowski and Rieseberg 2014; Alix et al. 2017).
63 Recent advances in genomics and phylogenomics have greatly increased our ability to detect both recent
64 and ancient hybridization and to understand its genetic and evolutionary consequences. It has been
65 estimated that ~25 percent of plant species in the UK flora actively hybridize with relatives (Mallet 2005),
66 and that more than 30% of living plant species are polyploids (Wood et al. 2009). It is also now clear that
67 ancient polyploidy is prevalent across plant phylogeny and that all major vascular plant lineages have
68 polyploidy in their past (Jiao et al. 2011; Amborella Genome Project 2013; Leebens-Mack et al. 2019;
69 Stull et al. 2021). It is unclear how many of these ancient polyploidy events were associated with
70 hybridization (i.e., allopolyploidy), but recent phylogenomic studies have dramatically increased the
71 number of documented cases of ancient hybridization in plants (both homoploid and polyploid). This is
72 perhaps not surprising—given the prevalence of recent hybridization and the appreciation that it can lead to
73 evolutionarily stable lineages (Ungerer et al. 1998; Soltis and Soltis 2009; Katche et al. 2021), we might
74 expect that hybridization has been an important process throughout the entirety of plant evolutionary
75 history, and also that some major lineages of plants could be of hybrid origin (via hybrid speciation) or
76 have experienced varying degrees of introgression with other lineages.

77 In light of mounting evidence for ancient hybridization, and the dramatic recent increase in
78 research on the subject, a critical review of the topic is timely to synthesize current knowledge and guide
79 future work. There are numerous extensive reviews on hybridization in plants (e.g., Mallet 2005; Soltis
80 and Soltis 2009; Abbott et al. 2013; Yakimowski and Rieseberg 2014; Suarez-Gonzalez et al. 2018,
81 Edelman and Mallet 2021), but these generally focus on the study of recent hybridization. Here we
82 provide a critical review focused in particular on *ancient* hybridization (see definition below). We survey
83 well-documented examples of ancient hybridization between major lineages of plants, ranging in
84 temporal scale from thousands of years ago to over 100 million years ago, and outline what we view as
85 compelling evidence for recognizing hybridization that occurred in deep time. We suggest that genomic
86 data should, when possible, be paired with other lines of evidence (e.g., biogeographic and niche
87 reconstructions) in order to evaluate the plausibility of historical hybridization scenarios, but that the
88 feasibility of such approaches is perhaps limited for putative hybridization events predating the Cenozoic.

89 While we assume that hybridization in the distant past is a fundamentally similar process to that
 90 happening in recent time, it is clear that detecting and studying ancient hybridization involves unique
 91 challenges that deserve special attention.

92 The consideration of hybridization across deep time also raises macroevolutionary questions
 93 worthy of discussion. For example, has the prevalence of hybridization been relatively consistent through
 94 time, or have certain geologic periods witnessed, or even facilitated, increased frequencies? What is the
 95 relationship between hybridization and diversification? Do particular traits tend to facilitate hybridization
 96 or the persistence of hybrid populations? Is the propensity to hybridize itself an adaptive life history
 97 strategy? Our discussions here build on previous considerations of these questions (e.g., Grant 1971;
 98 Ellstrand et al. 1996; Whitney et al. 2010; Folk et al. 2018a; Mitchell et al. 2019; Mitchell and Whitney
 99 2021). The prevalence of reticulation has also led some to question whether a bifurcating tree is the most
 100 appropriate way to model the diversification process and conceptualize evolutionary history more
 101 generally (e.g., Funk 1981; Linder and Rieseberg 2004; Baptiste et al. 2013). This has fundamental
 102 implications for the field of evolutionary biology, as well as for biological nomenclature and how we
 103 construct classifications. By reviewing current knowledge about ancient hybridization and discussing
 104 important issues related to its detection and association with broader patterns and processes of plant
 105 evolution, we hope to provide a roadmap for future research on the topic.

106

107 ANCIENT HYBRIDIZATION DEFINED

108 To our knowledge, there have been no *explicit* attempts to establish a definition for “ancient
 109 hybridization” as a phenomenon distinct from “recent hybridization”, despite the widespread use of the
 110 former term in the literature. To some extent,
 111 such a distinction is arbitrary since the same
 112 process is at play, but given that the
 113 population genetic and genomic
 114 consequences of hybridization tend to unfold
 115 in relatively discernable stages (Moran et al.
 116 2021), there might be a point in the
 117 evolutionary trajectory following
 118 hybridization that we can refer to past
 119 hybridization events as “ancient” in a
 120 meaningful sense. We consider hybridization
 121 events to be “ancient” if (a) gene flow
 122 between the lineages in question has ceased,
 123 (b) the genomes in the hybrid populations
 124 have stabilized, and (c) the lineage of hybrid
 125 origin has subsequently diversified or
 126 undergone a prolonged period of anagenesis
 127 following genome stabilization (e.g., Fig. 1).
 128 Regarding (b), genome stabilization can
 129 occur through the rapid purging of much of
 130 one parent’s ancestry (the minority parent) in
 131 cases of introgression (historically called
 132 introgressive hybridization), thus reducing

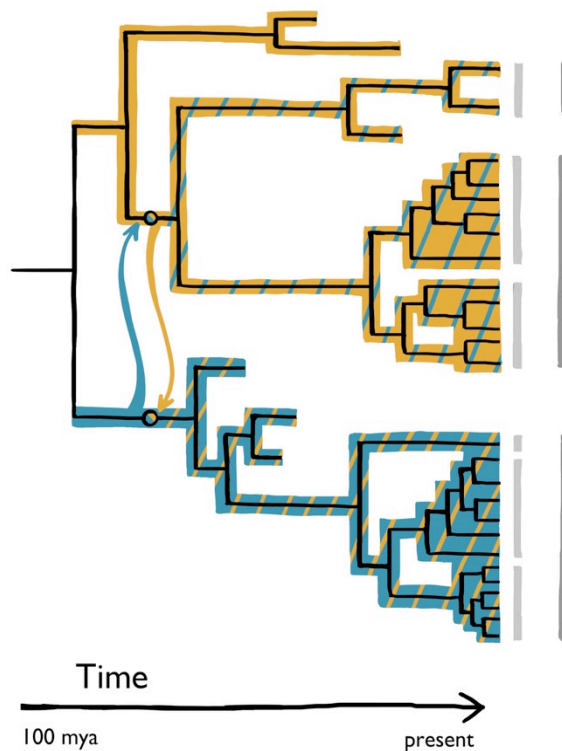


Figure 1. A chronogram of a fictional exemplar clade, with both extinct and extant taxa, in which ancient hybridization occurred between members of sister clades (highlighted in cobalt and gold) 80 million years ago; the reticulation event is marked by circles.

133 heterozygosity; through recombinational hybrid speciation, where chromosomal arrangements are
 134 established that render the hybrids interfertile but incompatible with either parent; and/or through
 135 polyploidization, which results in fixed heterosis. We can call this a “genomic” definition for ancient
 136 hybridization, and we follow this definition here.

137 Another approach to defining hybridization as “ancient” vs. “recent” is in relation to human
 138 history. For example, we can consider hybridization events to be “ancient” if they occurred before the
 139 Holocene (i.e., before 11,600 years ago). We call this a “historical” definition of ancient hybridization,
 140 and this roughly follows the logic of Welch and Rieseberg (2002). We chose the Holocene as the
 141 demarcation point as it has a concrete geologic definition and represents a general turning point in human
 142 history, when activities such as agriculture began to significantly impact Earth’s ecosystems (Larsen
 143 2006); furthermore, the proliferation of agriculture both directly and indirectly promoted widespread
 144 hybridization in plants (Anderson 1948, 1949). In some respects, this definition is more arbitrary and less
 145 biologically meaningful than the “genomic” definition above. For example, we would expect hybrid
 146 genomes to stabilize much more quickly in lineages with short vs. long generation times, so a
 147 hybridization event that occurred 15,000 years ago would be, in an evolutionary/genomic sense, perhaps
 148 more “ancient” in an annual herbaceous lineage than it would be in a woody lineage with much longer
 149 generation times. However, this historical definition is perhaps more straightforward in its application,
 150 and it also brings into focus the significance of anthropogenic activities in facilitating gene flow and the
 151 formation and persistence of hybrids over the last 11,600 years (Ottenburghs 2021), and the importance of
 152 identifying “natural” hybrids that occurred prior to major phases of anthropogenic disturbance and
 153 stewardship (Welch and Rieseberg 2002).

154

155 **HISTORICAL VIEWS ON ANCIENT HYBRIDIZATION**

156 Although the role of hybridization in plant evolution has been recognized for nearly a century,
 157 hybridization has generally been considered on recent timescales, whether viewed via “genomic” or
 158 “historical” perspectives. Ancient hybridization has tended to be associated with polyploidy. For
 159 example, Stebbins (1950) identified several angiosperm families as having ancient polyploid roots, having
 160 arisen via allopolyploidy, i.e. the combined processes of hybridization and chromosome doubling.
 161 (Although Stebbins did not explicitly note allopolyploidy in the origins of these ancient polyploid clades,
 162 he considered autopolyploidy to be ‘rare and maladaptive’ (Stebbins 1950, 1971); thus, his inferences of
 163 ancient polyploidy must have involved hybridization.) The magnoliid families Magnoliaceae,
 164 Myristicaceae, Calycanthaceae, and Lauraceae were proposed as ancient polyploids, as was the rosid
 165 family Salicaceae (Stebbins 1950). These hypotheses have been supported by genetic and genomic data
 166 (e.g., Soltis and Soltis 1990; Cui et al. 2006; Tuskan et al. 2006; Chaw et al. 2019; Chen et al. 2019). In
 167 contrast to ancient polyploidy, explicit hypothesized instances of ancient homoploid hybridization are rare
 168 in the classic botanical literature, perhaps because the effects of hybridization per se seem to be greatly
 169 obscured through time.

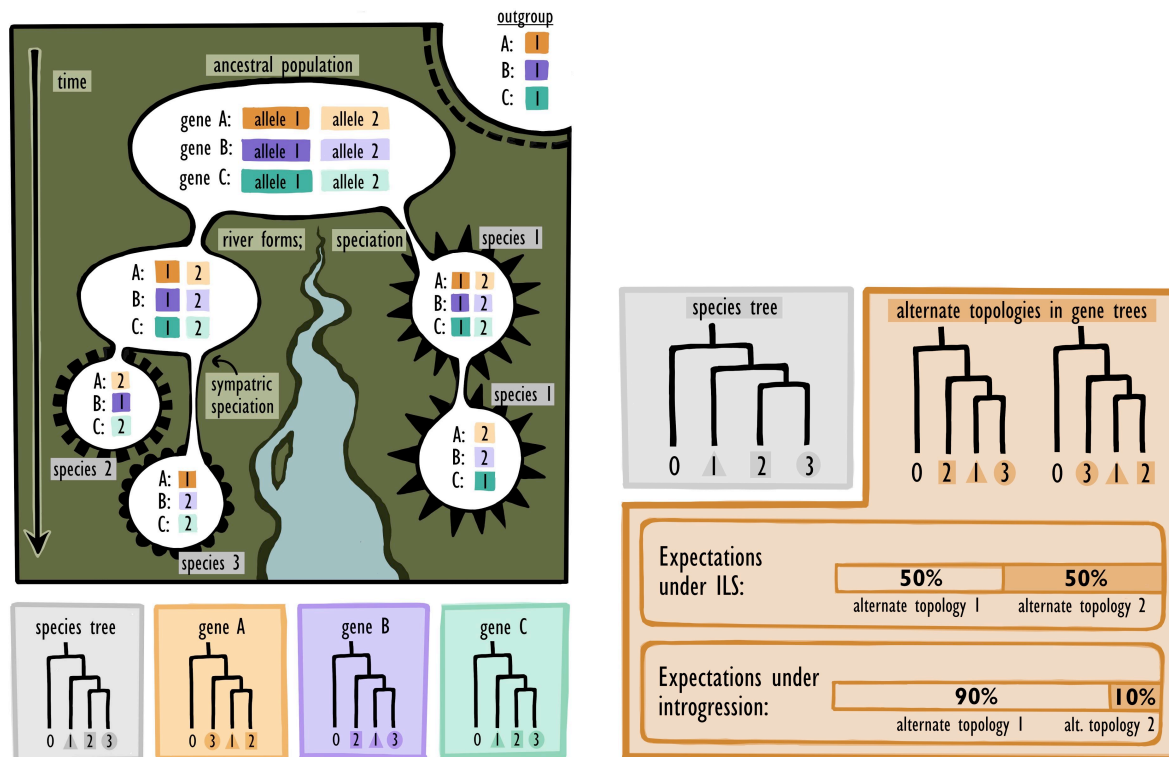
170 The genetic expectations for a stabilized homoploid hybrid species, based on Mendelian
 171 principles, were described by Gallez and Gottlieb (1982). Additivity of parental alleles at a *single locus*
 172 would be expected in a hybrid derivative, for example, allele *a* from parental species A and allele *b* from
 173 parental species B. In addition, because generations of hybridization and backcrossing may lead to loss of
 174 one parental allele or the other at some loci, rendering these loci monomorphic, additivity should be
 175 apparent *across loci* within a population or hybrid species, such that allele *a* from parental species A
 176 would be detected at locus 1 and allele *b* from parental species B would be detected at locus 2, etc. These

177 forms of additivity at specific genetic loci are valuable guides for detecting and documenting homoploid
178 hybridization on recent timescales. However, through time, as alleles from one parent or the other are lost
179 due to drift or selection and new alleles arise via mutation and accumulate, the genetic make-up of the
180 hybrid derivative will deviate from these expectations, and the hybrid origins of the lineage will
181 eventually be obscured. Thus, genetic comparisons, even at the level of genome sequences, may not be
182 useful in identifying cases of ancient hybridization, and alternative approaches to detecting ancient
183 hybridization are needed.

184

185 **HYBRIDIZATION DETECTION**

186 Hybridization is often detected via gene tree discordance, wherein the evolutionary histories of different
187 genes conflict (e.g., Rieseberg et al. 1990; Wendel and Doyle 1998; Sang and Zhong 2000). In plants, this
188 often manifests as cytonuclear discordance, where genes from the chloroplast genome show relationships
189 distinct from those of the nuclear genome due to “chloroplast capture,” a phenomenon in which the
190 chloroplast genome of one lineage becomes fixed in another following hybridization and backcrossing
191 (Rieseberg and Soltis 1991; Soltis and Kuzoff 1995). This occurs because chloroplast genomes typically
192 experience uniparental inheritance, lack recombination, and have small effective population sizes (Palmer
193 1985; Rieseberg and Soltis 1991), which prevents the sharing of genetic material between parental
194 plastomes and can lead to rapid fixation of one parental genome in hybrid populations. Given that
195 chloroplast genomes are typically maternally inherited (with some important exceptions such as paternal
196 inheritance in gymnosperms and biparental inheritance in some angiosperms), chloroplast capture can be
197 helpful for identifying maternal (or seed) contributions to gene flow (Asmussen and Schnabel 1991;
198 McCauley 1994). Discordance among nuclear loci across the genome can also stem from hybridization
199 and introgression, though these signals are generally more complicated to interpret than comparisons
200 between summary nuclear and chloroplast topologies due to biparental inheritance and
201 recombination/segregation in nuclear genomes. For either case, a topology that differs from the overall
202 inferred species tree indicates that the gene in question has a different evolutionary history that may be
203 explained by hybridization and introgression from a different species. However, there are other
204 evolutionary processes that produce patterns of discordance similar to those resulting from gene flow,
205 necessitating tests to differentiate between them.



206
 207 **Figure 2.** A) Diagram illustrating the process of incomplete lineage sorting, depicting an ancestral population where three genes
 208 have two alleles maintained. In subsequent speciation events, the two alleles are maintained in lineages and then randomly fixed,
 209 creating gene trees with topologies inconsistent with the species tree. B) Diagram illustrating expectations for the ABBA-BABA
 210 test under incomplete lineage sorting and introgression. Because incomplete lineage sorting occurs randomly across all genes,
 211 both alternative topologies are expected to occur in equal frequency. Because introgression occurs for specific taxa at specific
 212 loci, one alternative topology is expected to dominate in frequency.

213 The most common confounding factor for detecting hybridization is incomplete lineage sorting
 214 (ILS; Fig. 2), a process by which loci that are polymorphic at the time of species divergence randomly fix
 215 afterwards in a pattern that does not reflect the evolutionary history of the lineage, causing gene tree
 216 discordance (Nei 1986; Doyle 1992; Maddison 1997). When comparing chloroplast topologies to nuclear
 217 topologies, coalescent simulations can help identify when discordance is likely to arise under just ILS
 218 (Joly et al. 2009a; Folk et al. 2017). When comparing topologies of multiple loci across the nuclear
 219 genome, ILS can be distinguished from hybridization by comparing the ratio of sites or genes supporting
 220 topologies discordant with the inferred species tree for the whole genome or a subsection of the genome
 221 (ABBA-BABA test, Green et al. 2010; Patterson's D , Durand et al. 2011; \hat{f}_d , Martin et al. 2015). Under
 222 ILS, an even ratio of all discordant topologies is typically expected, as ILS acts randomly across the
 223 whole genome. In cases of introgression, a bias towards a particular discordant topology is expected, as
 224 introgression typically only results in specific genomic regions displaying discordance. This framework
 225 has been extended to allow inference of not only the presence, but also the directionality, of gene flow in
 226 more complex phylogenetic scenarios (DFOIL; Pease and Hahn 2015). ILS and hybridization can also be
 227 distinguished by examining the distribution of branch lengths for discordant genes (QuIBL, Edelman et
 228 al. 2019), with an exponential distribution expected under ILS and an exponential distribution with
 229 additional modes expected under hybridization.

230 Hybridization can also be detected against a background signal of ILS using phylogenetic
231 network inference—specifically, programs that infer phylogenetic networks using multilocus data and the
232 multispecies coalescent. One such program, PhyloNet (Wen et al. 2018), can use maximum parsimony or
233 maximum likelihood methods on a set of gene trees to estimate a phylogenetic network or use a Bayesian
234 framework directly on a multiple sequence alignment. However, the calculations to infer an entire
235 network can be computationally costly, especially as the number of taxa and hybridization events
236 increases, so the authors suggest subsetting to taxa and putative hybridization events of interest when
237 applying this program (Wen et al. 2018). The other popular program for this type of analysis, SNaQ
238 (Solis-Lemus et al. 2017), takes a different approach. It infers a phylogenetic network from a set of genes
239 or quartet concordance factors (Larget et al. 2010). The implemented algorithm increases scalability of
240 the analysis to larger datasets, but does not have a formal method for evaluating the true number of
241 hybridization events and cannot infer hybridization between sister species (Solis-Lemus et al. 2016).

242 Population genetics approaches offer another window onto hybridization/introgression. Genome-
243 wide markers such as SNPs from RAD-seq, genotyping-by-sequencing, or whole-genome resequencing
244 can be used to identify hybrid individuals and, in some cases, to delimit regions of the genome that have
245 been introgressed. Putative hybrids are genotyped, along with individuals from the parent species thought
246 not to be admixed. Admixture analysis (i.e., in STRUCTURE, Pritchard et al. 2000) is then used to
247 characterize the genetic composition of putative hybrids, estimating the genetic contribution of K source
248 populations to individuals. This admixture analysis can then be used to calculate a hybrid index and
249 estimate the type of hybrid (F1, F2, advanced generation). When marker coverage of the genome is
250 sufficiently dense and genetic distances between markers are known, admixture mapping analysis can be
251 conducted for smaller genomic regions to delimit which are likely of hybrid origin (Chakraborty and
252 Weiss 1988). Because ILS also affects this type of data, Patterson's D (and related statistics) and QuIBL
253 tests are used as well. As a caveat, admixture analysis assigns variants to populations based on existing
254 variation in the dataset and specified K and is therefore sensitive to biased sampling and misspecified K
255 (Lawson et al. 2018), which could lead to misidentification of hybrid individuals and genomic regions.
256 Other metrics that can be used to delimit introgressed genomic regions are relative and absolute
257 divergence (F_{ST} and d_{XY}), with the assumption that regions of the genome of hybrid origin will have a
258 higher divergence value compared to the background rate for the entire genome. Like the other metrics
259 mentioned here, divergence is affected by factors other than hybridization such as local recombination
260 rate (Noor and Bennett 2009). Therefore, researchers usually use many different metrics together to build
261 an overall case for hybrid origin of individuals and genomic regions.

262

263 **CHALLENGES OF DETECTING DEEP RETICULATION**

264 Detecting hybridization becomes an increasing challenge when considering possible events that occurred
265 deeper in evolutionary history. Sources of anecdotal evidence useful for detecting recent or contemporary
266 hybridization such as intermediate morphologies, overlapping ranges of putative parents, and increases in
267 chromosome number (in the case of allopolyploids) are typically no longer available given thousands to
268 millions of years of evolutionary change in phenotype, geographic range, and genome structure. The
269 inference of ancient hybridization therefore relies largely on phylogenomic signal, which also tends to
270 erode with the passage of evolutionary time.

271 On the one hand, with hybridization and subsequent backcrossing, purging of introgressed alleles
272 via recombination can quickly reduce the extent of information available for reconstructing past
273 hybridization (Moran et al. 2021). On the other hand, signal for gene flow (especially when limited) can

274 be very challenging to tease apart from the noise of ILS, a more or less constant backdrop to phylogenetic
275 signal that varies in intensity with factors such as time between divergences and population size. In
276 general, we might expect the circumstances that give rise to extensive ILS (e.g., rapid diversification
277 across heterogeneous landscapes) to also create ample opportunities for hybridization (due to close
278 proximity of closely related species, which may be brought into repeated contact with climatic
279 fluctuations). Rapid diversification will generally leave little signal in gene trees for later inference of
280 discrete evolutionary processes, and when that signal is eroded over time due to saturation or selection,
281 most individual gene trees might offer limited useful information. Taken together, in many cases, we
282 might be left with a limited ability to identify cases of ancient hybridization with certitude. Because signal
283 for past hybridization from the nuclear genome can be lost or obscured over time, well-supported
284 cytonuclear discordance might represent the best remaining evidence for ancient hybridization (Sun et al.
285 2015; Stull et al. 2015, 2020). However, often both nuclear and chloroplast data show uncertainty or
286 conflicting signals for particular deep relationships (e.g., Gitzendanner et al. 2018; Leebens-Mack et al.
287 2019), in which case cytonuclear discordance itself is no longer itself a reliable indicator. Furthermore,
288 many instances of ancient hybridization will not be reflected by cytonuclear discordance given that
289 chloroplast capture does not always accompany introgression.

290 Recent studies on *Amaranthaceae* s.l. (Morales-Briones et al. 2021) and *Malpighiales* (Cai et al.
291 2021) represent excellent examples of the challenges of detecting ancient hybridization in the context of
292 ancient, rapid radiations. In both cases, there are extreme levels of gene tree discordance along the
293 backbone, to the extent that it seems impossible to resolve clear bifurcating relationships among major
294 lineages, or even clear patterns of alternative conflicting relationships. While signal for gene flow does
295 appear to be present (e.g., imbalanced frequencies of alternative topologies), this occurs alongside
296 extensive ILS and gene-tree estimation error. Inferring discrete hybridization events from these complex
297 phylogenomic contexts is beyond the abilities of current methods, but reframing how we think about such
298 phylogenomic challenges might nevertheless offer meaningful biological insights (e.g., Parins-Fukuchi et
299 al. 2021).

300 Extinction represents another roadblock to the detection of ancient hybridization, a point that has
301 received relatively little attention until recently (e.g., Thomas et al. 2017; Tricou et al. 2022). Extinction
302 can obscure the parental lineages involved in ancient hybridization events, and in some cases might lead
303 to incorrect inferences of the parental lineages involved. For example, relatives of the extinct parental
304 lineages might be incorrectly inferred as participating in gene flow in the absence of the parental lineages
305 themselves (Lawson et al. 2018). In the case of polyploidy, instances of allopolyploidy could be
306 incorrectly reconstructed as autopolyploidy in gene-tree reconciliation approaches if both parental
307 lineages have gone extinct (Thomas et al. 2017). Similar problems with inference can also emerge from
308 incomplete sampling of extant lineages, but such problems should be remedied with the addition of more
309 complete data. Extinction, however, will likely represent an intractable barrier to a more complete
310 understanding of ancient hybridization in many cases. While extinction should generally represent a more
311 common confounding factor in the reconstruction of ancient hybridization events, we suggest that it is an
312 important issue to consider even when examining more shallow hybridization events given the extent of
313 recent and ongoing extinction in the Anthropocene.

314 **GENETIC CONSEQUENCES OF HYBRIDIZATION**

315 Although much of this review focuses on the positive (“creative”) potential of hybridization in evolution,
316 it is important to stress that hybridization often, and perhaps predominantly, has negative outcomes,
317

318 ranging from ecological selection against hybrids to more fundamental genetic and/or genomic obstacles.
319 In this section we outline the more immediate genetic consequences of hybridization, both positive and
320 negative, and the different means by which populations and genomes can stabilize after hybridization.

321 Hybridization can result in distinct evolutionary outcomes depending on the parental contributors
322 and the processes involved in the stabilization of hybrid populations (Arnold 1992). A fundamental initial
323 filter for hybrid success and subsequent genetic outcomes concerns whether or not the F₁ and subsequent
324 hybrid generations are viable and fertile, which is influenced by the genetic composition of the parental
325 lineages and the environments in which they live (Yakimowski and Rieseberg 2014; Moran et al. 2021).
326 Ecological selection against F₁ hybrids may occur due to abiotic filtering of unsuitable intermediate
327 phenotypes from either parent's niche (e.g., in *Encelia*, DiVittorio et al. 2020; in *Mimulus*, Kenney and
328 Sweigart 2016). Pollinator and herbivore interactions may also be disrupted in F₁ hybrids (e.g., in
329 *Mimulus*, Schemske and Bradshaw 1999; in *Eucalyptus*, Whitham et al. 1994). Regarding innate
330 reproductive isolation mechanisms, both genetic incompatibilities (reviewed in Bomblies 2010) and the
331 accumulation of deleterious alleles (hybridization load) can negatively impact the viability and fitness of
332 hybrid individuals. Fertility in particular is often compromised by incompatibilities (e.g., in *Mimulus*,
333 Sweigart et al. 2006). However, the extent of hybrid sterility varies greatly, and even with highly limited
334 fertility and rarely viable offspring, there is potential for hybrid success, especially since fertility can
335 increase with subsequent generations and the process of polyploidization can “rescue” hybrids from
336 sterility (Stebbins 1950). Assuming some level of fertility in hybrids, several processes can stabilize F₁
337 hybrids or subsequent hybrid generations and influence evolutionary outcomes. These include
338 polyploidization, introgression, and recombinational speciation (Grant 1971).

339 In the context of hybridization, polyploidy is significant in several ways. With hybrids that are
340 partially or largely sterile due to chromosomal incompatibilities, polyploidy can lead to the recovery of
341 fertility. Polyploidy can also result in reproductive isolation from parental lineages, effectively serving as
342 an “instantaneous” speciation mechanism. Polyploidy is also significant in that it results in fixed
343 heterosis, while other forms of hybrid stabilization typically result in the rapid erosion of heterozygosity.
344 The importance of allopolyploidy (i.e., polyploidy involving hybridization between different species, also
345 termed amphiploidy) in plant evolution has long been emphasized. Stebbins (1959) remarked that
346 “[a]mphiploidy, or the production of stable, true-breeding new species through the doubling of
347 chromosome number of a sterile interspecific hybrid, is now generally recognized as one of the
348 commonest ways in which plant species arise.” A survey by Wood et al. (2009) found that, in
349 angiosperms and ferns, 15% and 31% of speciation events, respectively, are accompanied by ploidy
350 increases, generally underscoring the importance of polyploidy in plant speciation. Understanding the
351 long-term evolutionary consequences of polyploidization is an area of active research, building off recent
352 studies documenting the presence of ancient polyploidy in most major plant lineages (e.g., Jiao et al.
353 2011, Jiao et al. 2014; Li et al. 2015; Leebens-Mack et al. 2019; Stull et al. 2021).

354 Another means by which hybrid populations/generations can stabilize is through recombinational
355 speciation, whereby segregation and recombination of chromosomal variants of the two parents result in
356 hybrids that are both fertile and reproductively isolated from their parental populations. This is perhaps
357 the primary mechanism of homoploid hybrid speciation (Coyne and Orr 2004). As noted by Yakimowski
358 and Rieseberg (2014), this model of hybrid speciation was initially proposed by Stebbins (1957a, 1959)
359 and then more formally developed and named by Grant (1958). While this mode of hybrid speciation
360 appears to be much less common than hybrid polyploid speciation, there are multiple empirical examples
361 (e.g., Stebbins 1957b; Grant 1971, 1981; Rieseberg 1991).

362 Introgression is another primary means by which the products of hybridization can become
363 established and effectively contribute to evolution. Introgression occurs when hybrids, after F₁ formation,
364 subsequently backcross with one or both parental species, with demographic processes and selection on
365 recombinant types quickly winnowing heterozygosity. This process can result in the transfer of a few
366 genes or chromosomal segments from one species to another. It has long been appreciated that
367 introgression could serve as a major source of variation for evolution (Anderson 1949)—a means of
368 generating considerable variation and perhaps novel features more rapidly than could other processes
369 such as mutation (Abbott et al. 2013; Soltis 2013; Suarez-Gonzalez et al. 2018). While adaptive
370 introgression (i.e., the transfer of fitness-increasing alleles from one population to another) is widely
371 recognized as an important process (Edelman and Mallet 2021), we note that introgression can also break
372 up adaptive allele combinations (segregation load) and result in the transfer of deleterious mutations
373 (hybridization load), resulting in fitness decreases (Moran et al. 2021).

374 Introgression can manifest in different ways. In some cases, introgression might result in one
375 widespread species effectively swamping into extinction a second, rare species (Todesco et al. 2016).
376 Hybrid populations could be relatively ephemeral, with a relatively brief phase of backcrossing with one
377 or both parents, such that both parental lineages largely retain evolutionary independence. With hybrid
378 swarms, however, persistent hybrid populations can contribute to ongoing backcrossing and gene flow
379 between the two parental species, perhaps resulting in more blurred species boundaries, at least at
380 geographic points of contact between hybrid and parental populations. Syngameons, groups of related
381 species that engage in frequent hybridization, represent another context where introgression is common
382 and potentially a significant source of adaptive variation (Lotsy 1925; Grant 1971), a topic that will be
383 discussed further below.

384 **EXAMPLES OF ANCIENT HYBRIDIZATION**

386 Below we review examples of well-documented cases of ancient hybridization (in the sense of our
387 genomic definition above) across vascular plant phylogeny that occurred at different points in geologic
388 history. Several clades (e.g., Fagales and Rosaceae) are discussed in greater detail because they have
389 experienced extensive hybridization throughout their evolutionary history. The examples discussed here
390 are not meant to be comprehensive but instead highlight that ancient hybridization spans the phylogenetic
391 breadth of vascular plants and underscore major themes concerning the evolutionary significance of deep
392 reticulation and the challenges of detecting it. We also note that one way to bolster inferences of ancient
393 hybridization is through reconstructions of ancestral ecological niches and paleo-distributions, which can
394 aid in evaluating the plausibility of ancient hybridization events suggested by phylogenomic evidence.
395 This type of approach is exemplified by Folk et al. (2018b), examining ancient hybridization in
396 Saxifragaceae, which we discuss further below. While most studies of this type have focused on putative
397 hybridization events during the Pleistocene (e.g., Ma et al. 2019), there is clearly potential to apply such
398 approaches to deeper time scales as paleoclimatic models permit (Folk et al. 2022).

399 **Rosids**

401 The large rosid clade of angiosperms (~90,000 species; Sun et al. 2020) represents one of the first
402 documented cases of ancient reticulation involving major angiosperm lineages. The placement of the
403 rosid subclade of Celastrales, Oxalidales, and Malpighiales (known as the COM clade, including ~19,000
404 species) has been highly contentious, with different data sets supporting very different placements (Sun et
405 al. 2015; reviewed in Soltis et al. 2018). Chloroplast genes support a relationship between the COM clade

406 and members of Fabidae, one of the two large rosid subclades. In contrast, mtDNA and nuclear genes
 407 both support a relationship between the COM clade and members of Malvidae, the other large rosid
 408 subclade (e.g., Duarte et al. 2010; Soltis et al., 2011; Zhang et al. 2012). Several morphological features,
 409 including an unusual type of ovule and contorted petals, also support the placement of the COM clade
 410 with malvids (Endress and Matthews 2006). Signals for both conflicting topologies (i.e., COM with
 411 fabids vs. COM with malvids) are present in nuclear data and were likely not caused by sampling or
 412 systematic errors, supporting a biological source of conflict such as reticulation or ancient introgression
 413 during the early diversification and rapid radiation of the rosid clade (Sun et al. 2015).

414

415 Saxifragaceae

416 Multiple examples of deep hybridization have been documented in Saxifragaceae. One of the most
 417 noteworthy of these involves the well-supported capture of the chloroplast genome from the ancestor of
 418 two species of *Mitella* (currently with a circumboreal distribution) by the ancestor of a clade of five
 419 species of *Heuchera* from southern Californian (Folk et al. 2018b). Extant members of these two clades
 420 are currently separated by ca. 1,300 km, which, at face value, might suggest that hybridization between
 421 these two lineages is improbable. However, Folk et al. (2018b) used a multifaceted approach employing
 422 niche modeling and estimates of ancestral ranges to show that the two clades potentially shared suitable
 423 habitat during the Pleistocene, when the reconstructed geographic distributions of the clades overlapped
 424 in southern California (Fig. 3). During that time, the analyses revealed that the distribution of the *Mitella*
 425 clade ancestor was pushed well to the south, providing an opportunity for ancient hybridization and gene
 426 flow.

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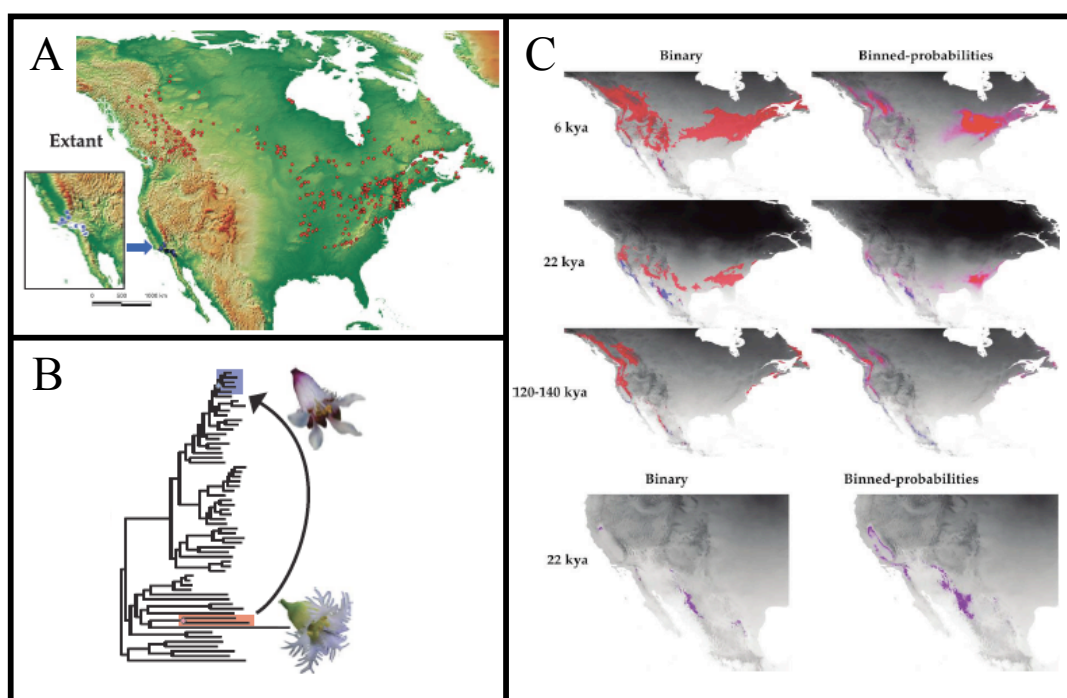
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Figure 3. Adapted from figs. 1 and 5 of Folk et al. (2018). (A) Current distribution of *Heuchera* (blue) and *Mitella* (red). (B) Phylogenetic positions of the focal clades (*Heuchera*, blue; *Mitella*, red). (C) Projections of ancestral geographic ranges based on estimates of ancestral niche space for *Heuchera* (blue) and *Mitella* (red).

450 **Fagales**

451 Oaks (*Quercus*) have long been an exemplar for hybridization, with a reputation for rampant
 452 hybridization and phenotypic variability (Palmer 1948; Hardin 1975). Recent hybridization is thought to
 453 occur only among species within the same taxonomic section (Palmer 1948; Hardin 1975), but evidence
 454 also suggests that ancient hybridization events occurred between the ancestors of different sections of
 455 *Quercus* as well as between those of different genera of subfamily Quercoideae of Fagaceae. Almost
 456 every genus in Fagaceae has been described as sharing chloroplast haplotypes with another (Manos et al.
 457 2008; Xiang et al. 2014; Simeone et al. 2016; Yang et al. 2021), belying nuclear and morphological data
 458 that resolve them as monophyletic (Manos et al. 2001; Oh and Manos 2008). This cytonuclear
 459 incongruence indicates that long-retained signals of either ILS or hybridization have persisted in these
 460 genomes for millions of years. Almost all studies found that haplotypes clustered by geography, with a
 461 North-Central American and a Eurasian clade, similar to chloroplast haplotype clustering by geography
 462 within *Quercus* itself (Whittemore and Schaal 1991; Petit et al. 1993; Dumolin-Lapegue et al. 1997;
 463 Simeone et al. 2016).

464 The extent of ancient
 465 hybridization in oaks and relatives in
 466 Quercoideae revealed within the nuclear
 467 genome has been the focus of several
 468 recent studies. Liu et al. (in review)
 469 tested for signals of hybridization using
 470 hundreds of nuclear loci and full
 471 chloroplast genomes for over 400
 472 species in Fagaceae and recovered
 473 geographic structuring of chloroplast
 474 haplotypes and nuclear evidence of
 475 multiple deep reticulation events
 476 between oaks and other genera in
 477 Fagaceae. Similarly, Zhou et al. (2022;
 478 Fig. 4) used over 2000 nuclear loci and
 479 complete chloroplast genomes to
 480 investigate signals of hybridization for
 481 over 90 species across Fagaceae and also
 482 recovered geographic clustering of
 483 chloroplast genotypes and rampant
 484 cytonuclear discordance at the base of
 485 the clade containing *Castanea*,

486 *Castanopsis*, *Chrysolepis*, *Lithocarpus*, *Notholithocarpus*, and *Quercus*. However, analyses of nuclear
 487 genes did not detect deep reticulation and found strong evidence of hybridization only within *Quercus*.

488 As evidenced by these studies, the chloroplast genome has retained a much stronger signal of
 489 ancient hybridization than the nuclear genome. The geographic clustering evident at even deep
 490 relationships implies that hybridization between the ancestors of *Quercus* and other genera was common
 491 enough to be preserved millions of years later. In contrast, recombination and continued divergence of
 492 introgressed haploblocks in the nuclear genome have made it much harder to identify which regions of the
 493 genome may have been exchanged by the ancestors of these genera. At the shallower scale, it is thought

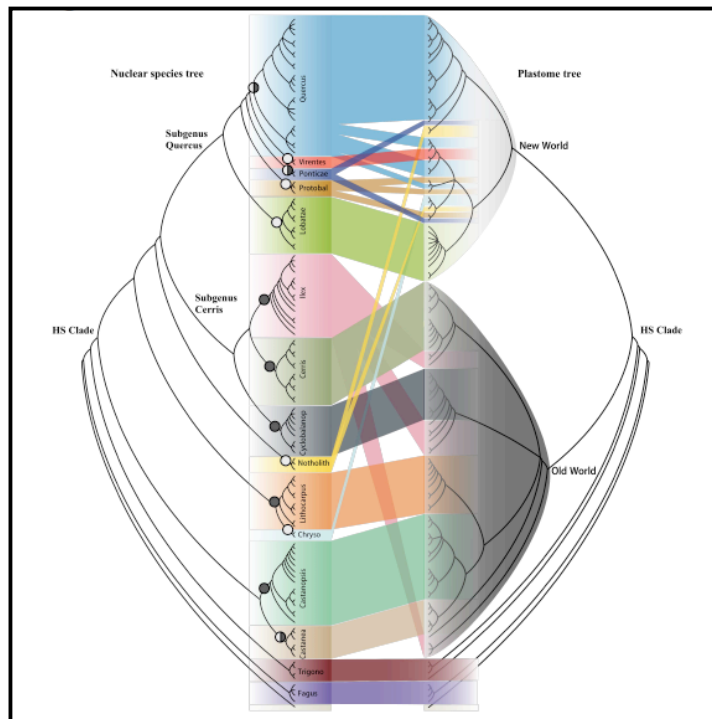


Figure 4. From Zhou et al. (2022; fig. 2) showing conflicts between nuclear and chloroplast phylogenies.

494 that the geographic clustering of chloroplast haplotypes between oak species is a signal of the
 495 confinement of many species to glacial refugia during the Pleistocene, which provided an opportunity for
 496 hybridization, and their subsequent northward recolonization when climates warmed afterwards (Petit et
 497 al. 2002). Given the association of hybridization with rapid changes in climate (e.g., Buggs 2007; Ryan et
 498 al. 2018), it is possible that the deeper events detected in these studies arose from similar circumstances of
 499 environmental upheaval. Oaks and their ancestors seem to undergo periodic cycles of hybridization and
 500 diversification corresponding to fluctuations in their ranges; Cannon and Petit (2020) suggest that these
 501 cycles and the group's ability to hybridize is actually adaptive in itself, allowing for species in the group
 502 to incorporate genetic variation from relatives which could help them to respond to changes in climate.
 503 Given recent evidence for introgression of genes associated with drought, waterlogging, and budbreak in
 504 several species of *Quercus* (Fitz-Gibbon et al. 2017; Leroy et al. 2020), the adaptive function of many
 505 introgressed alleles seems at the very least plausible. However, considering the difficulties of even
 506 identifying signals of ancient hybridization in the nuclear genome, verification of adaptive introgression
 507 of nuclear genes during ancient hybridization may be infeasible in most cases.

508 Ancient hybridization has been reported in other clades of Fagales (Zhang et al. 2019; Cardoni et
 509 al. 2021), with Betulaceae being a salient example. Wang et al. (2022), using genome resequencing to
 510 investigate ancient hybridization in Coryloideae (Betulaceae), found that *Carpinus* sect. *Distegocarpus*
 511 likely originated via homoploid hybrid speciation between the parental lineages *Carpinus* sect. *Carpinus*
 512 and *Ostrya*. Given that similar patterns of genomic admixture are shared between all three species of sect.
 513 *Distegocarpus* and the putative parental lineages, this reticulation event (dated at 17-33 mya) likely
 514 corresponds closely to the origin of the section, which subsequently diversified into three species
 515 (*Carpinus cordata*, *C. fangiana*, and *C. japonica*).

516

517 **Rosaceae**

518 Rosaceae are another excellent example of a clade with extensive hybridization throughout its
 519 evolutionary history, with numerous documented cases between closely related extant species (e.g., in
 520 *Rubus*, *Fragaria*, *Potentilla*, *Rosa*, Spiraea; Šarhanová et al. 2017; Debray et al. 2021; Qiao et al. 2021)
 521 as well as more ancient cases between the ancestors of small to large clades within the family (e.g.,
 522 Morales-Briones et al. 2018; Hodel et al. 2021, 2022). The many examples of hybridization in Rosaceae
 523 illustrate well how the process can function as an evolutionary force, contributing to speciation and the
 524 origins of phenotypic and phylogenetic diversity.

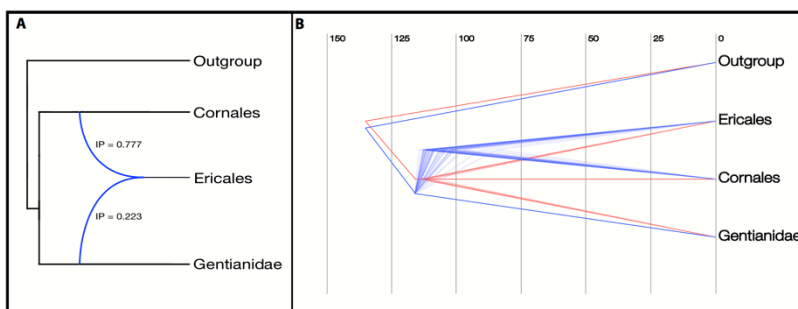
525 Hybridization has long been considered common in *Prunus*, and molecular data reveal that it is
 526 indeed rampant. Cultivated species frequently hybridize with each other as well as with wild species. For
 527 example, the rare Eurasian shrub *Prunus fruticosa* (ground cherry) has hybridized with cultivated *P.*
 528 *cerasus* (Macková et al. 2018), while genomic data suggest extensive natural interspecific hybridization
 529 among sympatric species of *Prunus* (Baek et al. 2018). At a deeper level in *Prunus*, Hodel et al. (2021)
 530 documented a hybridization event involving subgenus *Amygdalus* and subgenus *Prunus* using a
 531 phylogenomic approach. In *Rubus*, hybridization is not only common among close relatives, but is also
 532 one of the driving forces in the formation of naturally occurring apomictic lineages in subgenus *Rubus* in
 533 Europe (Šarhanová et al. 2017). In the Pacific Northwest of the U.S., hybridization between native and
 534 introduced species of *Rubus* is generating novel genotypes that may represent the early stages of newly
 535 formed invasive species. Alice et al. (2001) documented hybridization between distant relatives (i.e., *R.*
 536 *caesius* from subgenus *Rubus* and *R. idaicus* from subgenus *Idaeobatus*); most of the detected hybrids
 537 appeared to be later-generation hybrids, suggesting that hybridization between these distantly related

538 species has the potential for generating new species, or at least genetic diversity. *Alchemilla* also shows
 539 signatures of hybridization at both shallow and deep phylogenetic levels (Morales-Briones et al. 2018).

540 The apple tribe (Maleae) has also long been suspected to have a hybrid and/or allopolyploid
 541 origin given its high base chromosome number ($x = 17$) relative to other Rosaceae lineages (with $x = 5, 7,$
 542 $8,$ or 9), and several alternative hybridization scenarios have been hypothesized to explain Maleae origins,
 543 generally involving ancestors of amygdaloid or spiraeoid lineages (Evans and Campbell 2002). Recent
 544 phylogenomic analyses (Hodel et al. 2022) suggest that the ancestor of the Maleae ($x = 17$) + Gillenieae
 545 ($x = 9$) arose via hybridization between two distantly related Amygdaloideae lineages: (1) an ancestor of
 546 tribe Spiraeae ($x = 9$) and (2) an ancestor of the clade Sorbarieae ($x = 9$) + Exochordeae ($x = 8$) +
 547 Kerrieae ($x = 9$); this was then followed by polyploidy ($x = 18$) and aneuploidy ($x = 17$) in the Maleae
 548 lineage, resulting in the apple tribe's current base chromosome number.

549 Asterids

551 Phylogenetic analyses of asterids based primarily on the chloroplast genome have consistently recovered
 552 Cornales and Ericales as successive sisters to the core asterids (e.g., Albach et al. 2001; Soltis et al. 2011;
 553 Stull et al. 2015), while data from the nuclear genome has supported Cornales and Ericales as sister (e.g.,
 554 Zhang et al. 2012). In an in-
 555 depth comparison of asterid
 556 phylogeny based on chloroplast
 557 and nuclear datasets, Stull et al.
 558 (2020) determined that these
 559 conflicting relationships likely
 560 stem from hybridization during
 561 the early diversification of



562 asterids (Fig. 5). The
 563 predominant nuclear signal ($\frac{2}{3}$ of
 564 well-supported nuclear genes)

565 supports a sister relationship between Cornales and Ericales, but $\frac{1}{3}$ of the nuclear genes support a sister
 566 relationship between Ericales and *Gentianidae* (the chloroplast topology). Along with network analyses
 567 and coalescent simulations, these results suggest the possibility that Ericales is a reticulate lineage
 568 resulting from hybridization between ancestors of Cornales and *Gentianidae*.

570 Caryophyllales

571 Most studies examining ancient whole-genome duplication (WGD) do not specifically address whether
 572 inferred WGD events were caused by auto- or allopolyploidy, but this distinction is critically important
 573 for a better understanding of how polyploidy has shaped plant evolution. Yang et al. (2018), in a
 574 phylotranscriptomic study of Caryophyllales, devised an approach for inferring auto- vs. allopolyploidy
 575 from orthogroup trees, finding strong support for two ancient allopolyploidy events, one in
 576 Amaranthaceae and another in the ancestor of *Schiedea* (Caryophyllaceae). The type of polyploidy (auto-
 577 vs. allo-) responsible for the remaining 24 WGD events, however, appeared to be unclear. The question of
 578 whether most ancient WGD events represent auto- or allopolyploidy is one that deserves greater attention.
 579 It is also worth noting that some cases of presumed ancient homoploid hybridization could represent
 580 allopolyploidy obscured by subsequent diploidization.

581

582 Amaryllidaceae

583 Tribe Hippeastreae (Amaryllidaceae) provides an example of deep reticulation in the monocots. The tribe
584 consists of 10–13 genera and ca. 180 species. Phylogenetic analyses of nuclear and chloroplast regions
585 revealed two well-supported clades within the tribe (Garcia et al. 2014). Within one of these clades, the
586 mainly Neotropical Hippeastrinae, Garcia et al. (2014, 2017) uncovered widespread cytonuclear
587 discordance and rampant non-monophyly of genera. Closer examination of the data revealed that the
588 discordance is likely due to numerous ancient hybridization events (some associated with polyploidy)
589 preceding the radiation of this major clade (Garcia et al. 2017). In contrast, phylogenetic analysis of the
590 second clade of Hippeastreae, subtribe Traubiinae, shows a tree-like pattern, consistent with the apparent
591 absence of hybridization and allopolyploidy in this clade (Garcia et al. 2017).

592

593 Mesangiosperms

594 Despite intensive study, phylogenetic relationships among the five clades (magnoliids, monocots,
595 eudicots, Chloranthales, Ceratophyllales) of *Mesangiospermae* have remained uncertain due to their rapid
596 radiation (Moore et al. 2007). Several phylogenetic analyses have recovered Chloranthaceae sister to
597 magnoliids (reviewed in Soltis et al. 2019). However, incongruence has long been noted between nuclear
598 and chloroplast DNA datasets regarding the placement of magnoliids + Chloranthaceae (or magnoliids
599 alone in those studies in which Chloranthaceae were not included) relative to other mesangiosperms.
600 Chloroplast trees have routinely recovered magnoliids and Chloranthaceae as sister to a clade of
601 monocots plus eudicots; in contrast, phylogenetic analyses of nuclear genes typically placed monocots as
602 the sister to a clade of magnoliids, Chloranthaceae, and eudicots (e.g., Moore et al. 2007; Leebens-Mack
603 et al. 2019; Chaw et al. 2019; Chen et al. 2019; Li et al. 2019; Soltis et al. 2011). Using complete nuclear
604 genome sequences for *Amborella* and representatives of all mesangiosperm lineages, Guo et al. (2021)
605 conducted a variety of phylogenomic analyses to understand the conflict underlying these contentious
606 relationships, finding that ancient hybridization is a plausible cause for the observed conflict concerning
607 the placement of Chloranthales + magnoliids relative to eudicots and monocots.

608

609 Gymnosperms

610 Increasing evidence for WGD (both recent and ancient; Li et al. 2015; Wu et al. 2016; Leebens-Mack et
611 al. 2019; Stull et al. 2021) and (relatively) recent hybridization (Worth et al. 2016; Sullivan et al. 2017;
612 Ma et al. 2019) in various lineages of gymnosperms suggests a possible role for ancient hybridization in
613 gymnosperm evolution, but well-documented examples are relatively few compared to those of flowering
614 plants. However, one example comes from Cupressaceae: Liu et al. (2022) found evidence for ancient
615 hybridization involving the ancestors of two clades of Cupressoideae, one including *Microbiota*,
616 *Platycladus*, and *Tetraclinis* and the other including *Juniperus*, *Cupressus*, *Hesperocyparis*, *Callitropsis*,
617 and *Xanthocyparis*. One intriguing incongruence that deserves further attention concerns the phylogenetic
618 placement of *Ginkgo* (Stull et al. 2021). Most nuclear genes place *Ginkgo* sister to cycads, but an
619 appreciable number of genes support *Ginkgo* sister to conifers, consistent with many morphological
620 phylogenetic analyses of seed plants (e.g., Crane 1985). Phylogenomic analyses also support an ancient
621 WGD in the common ancestor of extant gymnosperms (Stull et al. 2021; Liu et al. 2022), the branch
622 subtending these alternative placements of *Ginkgo*.

623

624

625

626 Ferns

627 Hybridization and allopolyploidy are also common across ferns (Barrington et al. 1989; Otto and Whitton
628 2000; Sigel 2016; Leebens-Mack et al. 2019), and recent emphasis has been placed on examples of “deep
629 hybridization” in ferns. However, these reports generally pertain to recently formed hybrids among
630 distantly related lineages, rather than to instances of hybridization that occurred in the distant past. For
631 example, x*Cystocarpium roskamianum* is a recently formed inter-generic hybrid between *Gymnocarpium*
632 and *Cystopteris*, which diverged ca. 60 mya (Rothfels et al. 2015). Similarly, x*Lindsaeosoria flynnii* is
633 another recently formed inter-generic hybrid involving *Lindsaea* and *Odontosoria*, whose divergence
634 perhaps traces back to the Mesozoic (Lehtonen 2018).

635 Ranker and Sundue (2015) suggested that ferns exhibit slower evolution of reproductive barriers
636 than angiosperms, which should facilitate hybridization in general, including among more distantly
637 related lineages than is typical for other plant clades. If correct, hybridization among fern species should
638 be widespread. But the extent of ancient hybridization in fern evolutionary history remains unclear. The
639 best information from phylogenomics comes from the Leebens-Mack et al. (2019), which provides
640 evidence for shallow as well as some deep polyploidy events, implicating ancient hybridization if
641 allopolyploidy was at play. Additionally, recently published, high-quality fern genomes and
642 transcriptomes provide strong evidence for a few ancient polyploidy events in ferns, with several
643 polyploidy events inferred in the common ancestors of smaller orders and families (Huang et al. 2019,
644 Pelosi et al. 2022), one associated with the origin of core leptosporangiate ferns (Li et al. 2018; Huang et
645 al. 2019; Huang et al. 2022; Marchant et al. 2022) and possibly another associated with much of
646 Polypodiales (Leebens-Mack et al. 2019; Pelosi et al. 2022). However, there remain few robust, large-
647 scale comparisons of fern phylogeny based on comparably sampled nuclear vs. chloroplast datasets.
648 Furthermore, because of their large and complex genomes, complete genome sequencing of ferns, which
649 could reveal such ancient signatures, is in its infancy.

650

651 MACROEVOLUTIONARY SIGNIFICANCE OF HYBRIDIZATION

652 Hybridization can be viewed as a fundamentally microevolutionary process given that it essentially
653 represents a form of gene flow that can either lead to the establishment of hybrid populations or genetic
654 introgression from one species or population to another, and as a consequence, it influences the pool of
655 variation on which selection can operate. However, it may have broader macroevolutionary significance
656 in the sense that it may either influence or accompany other broader patterns in plant evolution. In
657 particular, the reality that hybridization can occur among divergent populations as well as lineages
658 considered taxonomically to be separate species underscores that hybridization is a phenomenon that in
659 some senses transcends evolutionary scales. Given the capacity of hybridization to reconnect diverging
660 lineages and potentially impart lasting influence on evolutionary trajectories, evaluation of the
661 significance of hybridization events whose signals appear across broader evolutionary scales is warranted.
662 It is also worth considering both biotic and abiotic circumstances that might influence the frequency of
663 hybridization among different lineages and at different times in Earth history. In this section, we consider
664 the macroevolutionary significance of hybridization. We do not use the term macroevolution in any
665 specific mechanistic sense (e.g., in reference to species selection), but instead as a general umbrella for
666 considering how hybridization might influence broader patterns of plant evolution, and how different
667 circumstances or traits might influence the frequency of hybridization across space, time, and phylogeny.

668

669

670 **The ecological, geographic, and geologic context of hybridization**

671 One of the significant themes to come out of the last 70 years of hybridization work is the association of
672 hybridization with ecological and range change. Many classic cases of hybridization date to the
673 Pleistocene, as ranges of different species were brought into secondary contact in glacial refugia or
674 subsequently following range expansion during interglacial periods (e.g., Anderson and Stebbins 1954;
675 Petit et al. 1997, Edwards et al. 2006; Joly et al. 2009b). Other periods of significant geologic and
676 environmental change might also have corresponded with increased levels of hybridization. For example,
677 the availability of land bridges in the Northern Hemisphere during the early-mid Eocene may have
678 facilitated oak migrations between Eurasia and the Americas, bringing lineages into contact and resulting
679 in hybridization (Liu et al. in review). Similarly, climate change in the late Miocene to Pliocene might
680 have brought various oak lineages into contact due to range shifts, again resulting in widespread
681 hybridization (Zhou et al. 2022). The Cretaceous-Paleogene (K-Pg) mass extinction may have coincided
682 with (or have been followed by or possibly even driven) a wave of polyploidization (Fawcett et al. 2009;
683 Van de Peer et al. 2021), although further research is needed to evaluate the strength of this possible
684 connection. This suggested wave has been explained both by the enhanced persistence of the polyploids
685 themselves, as well as a possible increase in unreduced gamete production and diploid hybridization
686 caused by the extreme environmental and disturbed ecological conditions present in the wake of the
687 asteroid impact (Levin and Soltis, 2018; Levin 2020). Of course, responses to climate change and
688 ecological disturbance can be lineage-specific and idiosyncratic, and so different geologic periods and
689 environmental contexts might impact lineages very differently with regard to range shifts and patterns of
690 gene flow. But nevertheless, periods of Earth history (and regions of the globe) that experienced bouts of
691 ecological disturbance, climate change, and/or intermittent land connections (with consequent
692 community-level range fluctuations) might have experienced increased frequencies of hybridization.

693 During remarkable phases of diversification of major lineages (e.g., angiosperms), there may
694 have been generally increased opportunities for hybridization as well, simply as a byproduct of the
695 presence of numerous closely related lineages (lacking pre- or postzygotic barriers) in close proximity.
696 For example, the large-scale ecological shift from gymno- to angiosperm dominance in the mid-
697 Cretaceous (at a global scale) might represent a unique period in angiosperm history where diversification
698 rates were elevated, ranges were shifting relatively rapidly, and as a result hybridization was particularly
699 common (Boulter et al. 1988); this is not inconsistent with the numerous angiosperm phylogenies
700 showing considerable genomic conflict at nodes dating to the early-mid Cretaceous. Of course, rapid
701 diversification itself can be a source of conflict (via ILS), and angiosperms have undergone repeated
702 bouts of remarkable diversification (Magallón and Castillo 2009; Tank et al. 2015; Landis et al. 2018), so
703 if there is any merit to the notion that the initial emergence and radiation of major lineages is often
704 accompanied by increased levels of hybridization, we would expect to see periodic bursts of hybridization
705 throughout geologic history at different locations in plant phylogeny.

706 It is important to note that the association of hybridization with large-scale geographic patterns is
707 not new; it could be viewed as an extension of classic theory about ecological control of hybridization.
708 Anderson's "hybridization of the habitat" (Anderson 1948) centered the importance of ecological
709 circumstances in the promotion of hybrid formation and establishment. Visible early-generation hybrids
710 were often spotted in anthropogenically disturbed places and in the transition zones between two types of
711 habitats ("hybrid habitats"), leading Anderson to conclude that hybrids were probably excluded from
712 parental ranges via ecological filtering or competitive exclusion with parents. Rapid environmental
713 changes might in some cases result in "hybrid habitats" due to the establishment of large areas of unique

714 habitat or a transition zone where no parental species is well-adapted, allowing for the persistence of
715 hybrids. Another interpretation is that these “hybrid habitats” are simply points of contact between
716 different species ranges and habitats, and as climates change, these areas of contact will naturally shift as
717 well, creating more opportunities for hybridization.

718

719 **Trait and phylogenetic correlations with hybridization frequency**

720 Several attempts have been made to characterize traits commonly associated with hybridization, as well
721 as to identify lineages in which hybridization is more frequent, perhaps as a consequence of the traits they
722 possess (e.g., Grant 1971; Ellstrand et al. 1996; Whitney et al. 2010; Mitchell et al. 2019). While some
723 traits might generally facilitate hybrid formation, others seem potentially important for facilitating hybrid
724 persistence. Grant (1971) described several different life history paradigms associated with different
725 hybridization patterns, observing that homoploid syngameons (e.g., *Ceanothus*) were usually long-lived
726 woody species with stable chromosome structure, large population size, and often (but not always) wind
727 pollination. Ellstrand et al. (1996) found strong taxonomic clustering of hybridizing taxa, as well as a
728 tendency for hybrids to be outcrossing perennials with the ability to reproduce clonally; however, they did
729 not differentiate between polyploid and homoploid hybrids in their study. We expect chromosomes of
730 homoploid hybrid taxa to be comparatively conserved with regards to structure and sequence; otherwise
731 viable hybrid offspring would be unlikely (without clonality to allow the persistence of hybrids with odd
732 chromosome pairs). This corresponds with Grant’s (1971) observation that trees often form syngameons;
733 their long generation times result in slower rates of evolution, increasing the chance that diverged species
734 will still maintain enough genetic similarity to cross successfully.

735 Wind pollination, as a more promiscuous form of pollen spread than animal-mediated pollination,
736 is often thought to facilitate hybridization. Wind pollination is a feature of many taxa famous for
737 homoploid hybridization (e.g., *Populus*, *Quercus*, *Nothofagus*; Smith and Sytsma 1990; Williams et al.
738 2001; Acosta and Premoli 2018), but many other groups in which hybridization is common are insect-
739 pollinated (e.g., *Helianthus*, *Eucalyptus*; Rieseberg 1991; Van Dijk et al. 2020), indicating that pollinator
740 choice can lead to porous species barriers. In fact, even in cases of tight pollinator specialization, such as
741 figs, hybridization can still occur, perhaps in particular during pollinator-host switches (Wang et al.
742 2021). A study by Mitchell et al. (2021) examined the correlation of 11 different traits with hybridization
743 and found weak positive associations between perenniality, woodiness, outcrossing, abiotic pollination,
744 and larger genomes and increased frequencies of hybridization, but some of these results were scale
745 dependent. Hybridization is a complex phenomenon that appears to require the confluence of several to
746 multiple abiotic and biotic factors (and for populations to persist once formed), and so it is not surprising
747 that particular traits generally show inconsistent relationships with hybridization frequency. For example,
748 traits that facilitate hybrid persistence (e.g., perenniality) will not show a strong relationship with
749 hybridization when paired with other traits that prevent hybrid formation in the first place (e.g., high-
750 fidelity biotic pollination).

751 Through the lens of oak diversity, Cannon and Petit (2020) proposed that the ability to hybridize
752 itself while maintaining species boundaries (i.e., the syngameon strategy) may be adaptive. This
753 hypothesis is intriguing and will require future work to test fully, in particular, to establish which traits
754 would need to be selected to maintain the ability to hybridize and whether these traits are consistent
755 across the Tree of Life. Additional work from a speciation biology paradigm could test the plausibility of
756 maintaining equilibrium between selecting for regions of the genome that preserve the ability to hybridize
757 and other regions that contribute to species boundaries. Another line of investigation could be establishing

758 the timing of diversification and hybridization events in conjunction with selective pressures for the
759 ability to hybridize, which we may even be able to do in real time due to Anthropocene-driven
760 environmental change and ecological disturbance. Of course, these are a few of many possible avenues of
761 investigation regarding this question. Another important problem is obtaining a more complete
762 understanding of how many lineages operate as syngameons. It has been suggested that many diverse
763 tropical tree genera might represent syngameons, but these are more poorly understood compared to
764 temperate systems (but see Larson et al. 2021).

765

766 **Hybridization and species diversification**

767 While perhaps more attention has been paid to hybridization as a vehicle for the evolution of novel
768 phenotypes (Anderson and Stebbins 1954; Seehausen 2004; Suarez-Gonzalez et al. 2018), there has also
769 been interest in whether hybridization might directly influence diversification rates in plants and other
770 organisms. There are several ways that hybridization could influence diversification rates, either
771 positively or negatively; hybridization could also in some cases impact standing levels of biodiversity
772 without any influence on diversification rates per se. When hybridization (and the resulting rich genetic
773 variation) is paired with the right ecological circumstances (e.g., the colonization of an island system by a
774 hybrid population), this could lead to an adaptive radiation (Barrier et al. 1999; Choi et al. 2021) and,
775 presumably, increased rates of speciation in the radiating lineage. Conversely, hybridization could lead to
776 decreased speciation rates (or simply decreased levels of standing diversity) via the merging of recently
777 divergent lineages upon secondary contact. However, there are also deeper theoretical concerns regarding
778 the relationship between diversification and hybridization. Diversification itself might create more
779 opportunities for hybridization—given close geographic proximity of multiple, minimally divergent
780 lineages—and so the observation of species-rich lineages with rampant hybridization is not in itself
781 sufficient evidence that hybridization is the driving force. In other words, disentangling the causal
782 relationship of these two processes is not trivial (Mitchell and Whitney 2021), and perhaps in some cases
783 there is a positive feedback of interrelated causality leading to increases in both. A recent study explicitly
784 examining the relationship between hybridization and diversification (Mitchell and Whitney 2021) overall
785 found a relatively limited relationship between these phenomena at a broad scale, although a stronger
786 positive relationship was observed when taking life history traits (e.g., perenniality and woodiness) into
787 consideration.

788 Diversification rates aside, hybridization (particularly in association with polyploidy) can
789 hypothetically increase standing diversity via a ratchet mechanism (Scarpino et al. 2014), whereby
790 allopolyploidy produces distinct lineages that are reproductively isolated from their parents. In clades
791 where hybridization/allopolyploidy is ubiquitous, such as Rosaceae, this ratchet mechanism can perhaps
792 lead to appreciable increases in species diversity without directly imparting any influence on inherent
793 speciation rates in the conventional sense (Vamosi and Dickinson 2006). However, this raises the
794 question of whether the resulting allopolyploidy lineages themselves might generally have elevated rates
795 of diversification, but prevailing evidence suggests there is generally a limited relationship between
796 polyploidization events and diversification (Tank et al. 2015; Landis et al. 2018; Smith et al. 2018; Stull
797 et al. 2021), although this may, in part, be a data-deficient problem. Overall, the relationship between
798 hybridization and diversification might best be characterized as idiosyncratic. We suggest that future
799 research on this topic focus on understanding, for particular clades, the specific contexts in which (or
800 circumstances by which) hybridization influences diversification rates (either positively or negatively) or
801 standing diversity, instead of searching for a general positive or negative relationship between

802 hybridization and diversification across major lineages. It seems likely that the relationship between
803 hybridization and diversification/diversity is too complex to draw meaningful (simple) generalizations at
804 a broad scale, but through focused case studies, more detailed general principles might emerge on how
805 hybridization influences diversification (and vice versa).

806

807 NOMENCLATURAL IMPLICATIONS

808 The chloroplast genome has been the primary source of data in plant phylogenetics for over three decades
809 given the relative ease of isolating, sequencing, and analyzing chloroplast DNA (e.g., Ritland and Clegg
810 1987; Palmer et al. 1988; Gitzendanner et al. 2018; Li et al. 2019). As a result, our current major
811 classifications (e.g., APG IV 2016; PPG I 2016) are largely based on chloroplast phylogenies. But it was
812 always made clear in the formulation of these classifications that nuclear data would ultimately be
813 required to confirm their accuracy, especially in light of processes such as chloroplast capture that can
814 mislead organismal phylogenies (Rieseberg and Soltis 1991). The surge in availability of nuclear
815 sequence data from target capture, transcriptome, and whole-genome sequencing over the past decade has
816 provided much-needed perspective on vascular plant phylogeny from the nuclear genome. Notably, as
817 reviewed above, many nuclear phylogenomic studies have revealed deep incongruences with plastome
818 phylogenies, in some cases involving the placements of clades recognized as families, orders, or more
819 inclusive named clades (e.g., Folk et al. 2018b; Liu et al. 2022; Leebens-Mack et al. 2019; Stull et al.
820 2020). This raises the important question of how best to treat these chloroplast-nuclear incongruities in
821 classifications moving forward. One solution is to continue with the chloroplast placements, as in the
822 APG treatments to date (APG IV 2016), for reasons of stability, with an asterisk or some other means
823 used to indicate that a lineage stems from reticulation and might have an alternative nuclear phylogenetic
824 placement. However, we argue this is an untenable approach given that the nuclear genome is more
825 representative of organismal phylogeny as a whole, and therefore classifications should be updated to
826 reflect the nuclear phylogeny. Then the issue still remains of how to treat or depict reticulate lineages in
827 classification. One option, made possible by the PhyloCode (Cantino and De Queiroz 2020), is to place
828 reticulate lineages in both parental clades. Thus, in the case of the COM clade (Sun et al. 2015), it would
829 be placed in both Malvaceae (the predominant nuclear signal) and Fabaceae (the predominant chloroplast
830 signal and the APG IV placement), as was done in the PhyloCode treatment of the COM clade (Judd et al.
831 2020a,b). This approach then raises the question: how admixed must a clade be to warrant this treatment?
832 Arguably, the amount of admixture must be significant, but what counts as significant may differ based
833 on the age and size of the clade. This issue is discussed further below.

834 In contrast to the PhyloCode, there currently is no readily available means for designating deep
835 reticulation events in the *International Code of Nomenclature for Algae, Fungi, and Plants* (ICN; Turland
836 et al. 2018). Under the ICN, hybrids between species and between species in different genera can be given
837 names. For example, hybrids between *Magnolia denudata* × *Magnolia liliiflora* have been given the name
838 *Magnolia* × *soulangeana*. However, when a person sees the name *Magnolia* × *soulangeana* in the
839 literature, the parents would not be known to the reader without looking into the hybrid name in more
840 detail. A hybrid can also be designated by using an × between the putative parents. For example, one
841 could use *Magnolia denudata* × *Magnolia liliiflora* (to designate *Magnolia* × *soulangeana*). The code also
842 permits the use of new generic names for hybrids between two species from different genera. The generic
843 name *Sorbopyrus* has been used for hybrids between *Sorbus* and *Pyrus*, and ×*Sorbopyrus auricularis* is a
844 hybrid between *Sorbus aria* and *Pyrus communis*. It is possible that the code of nomenclature could be
845 extended to allow the application of these approaches to deep reticulation events involving named

846 lineages above the generic rank. For example, ×Ericales would designate that the Ericales lineage has
 847 reticulation in its history or is of reticulate origin (Stull et al. 2020). However, this system of naming
 848 would not in itself indicate the putative parents and does not define the alternative clades, as the
 849 PhyloCode permits. This could be done by indicating the involved parental lineages in parentheses:
 850 ×Ericales (Cornales/*Gentianidae*).

851 Whether we are currently ready to overhaul (or update) angiosperm classification (or vascular
 852 plant classification more broadly) based on available nuclear phylogenies is another question. Recently
 853 generated large-scale nuclear datasets are incredibly valuable, but these can also be compromised by
 854 analytical challenges including paralogy, data quality, and low-information gene trees, which can result in
 855 spurious topologies, especially among major lineages. Many areas of angiosperm phylogeny display high
 856 levels of conflict between not only nuclear and chloroplast datasets, but within and among nuclear
 857 datasets themselves. Nuclear phylogenomic studies focused on particular problems in angiosperm
 858 phylogeny—e.g., asterids (Stull et al. 2020; Zhang et al. 2020), Caryophyllales (Walker et al. 2018; Yang
 859 et al. 2018), Caesalpinioideae (Leguminosae; Ringelberg et al. 2022), Commelinales (Zuntini et al. 2021),
 860 Gentianales (Antonelli et al. 2021), Orchidaceae (Pérez-Escobar et al. 2021), Piperales (Jost et al. 2021)—
 861 perhaps represent a more reliable basis for making specific classification or nomenclatural changes
 862 compared with broader analyses of plant phylogeny (Leebens-Mack et al. 2019; Baker et al. 2022).

863 Another fundamental question is what level of hybridization should we consider sufficiently
 864 substantial to be reflected in names and classifications. For example, if lineage A (sister to lineage B)
 865 shows a small proportion of nuclear genes (e.g., <5%) reflecting introgression from lineage C, should we
 866 seek to reflect this in our classifications? What if the proportion of introgressed genes is higher (e.g.,
 867 25%)? Does it matter what relationship the chloroplast genome reflects in such cases? With cytonuclear
 868 discordance, the chloroplast genome might support A and C as sister, even if the preponderance of
 869 nuclear signal supports A and B as sister. What about a case where 60 percent of the nuclear genome
 870 supports A and B as sister, while 40 percent of the nuclear genome (as well as the chloroplast genome)
 871 support A and C as sister? The issues raised above will represent important considerations for the plant
 872 systematics community moving forward.

873

874 **DEPICTING AND ANALYZING A RETICULATE TREE OF LIFE**

875 The concept of the Tree of Life is a wonderful metaphor
 876 for the evolutionary connectivity of all life but is also
 877 problematic biologically in that it assumes a bifurcating
 878 tree. We have known for a long time, in part based on
 879 rampant hybridization among the tips of the plant branch
 880 of the Tree of Life, that this assumption is unrealistic.
 881 With growing evidence for reticulation across the depth
 882 of green plant evolution, this raises the question of how
 883 best to depict plant phylogeny so that it is easy to view
 884 yet biologically realistic, while remaining a useful tool
 885 for teaching students and educating the public.

886 Phylogenetic relationships in Bacteria and Archaea are
 887 generally depicted as a network, given the prevalence of
 888 reticulation via horizontal gene transfer (Dagan et al.,
 889 2008; Dagan and Martin 2009; Gontier, 2015; Kunin et al. 2005; Fig. 6), and perhaps the plant

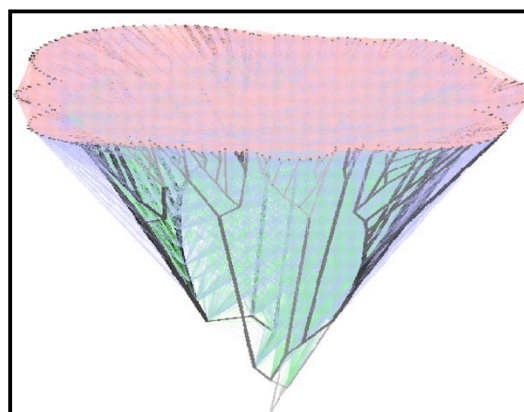


Figure 6. Network representation of prokaryote phylogeny including both vertical inheritance and lateral exchange (from Dagan and Martin 2009; fig. 2).

890 community should increasingly move toward these types of depictions (Fig. 7)—a Net of Life rather than
891 a Tree of Life for plant evolutionary history. Of course, as noted above, network methods are increasingly
892 being used in the phylogenetics community, but these are extremely limited in their scalability and are
893 often wanting in effective communication of evolutionary patterns. Methods development in
894 reconstructing, visualizing, and applying phylogenetic networks for evolutionary inferences is a critical
895 direction in evolutionary biology that is currently in its infancy (e.g., Hibbins and Hahn 2021). It is
896 important to note that the vast majority of comparative methods aimed at inferring divergence times,
897 diversification rates, ancestral states and trait evolution, biogeographic history, and other aspects of
898 evolutionary history typically require or assume a bifurcating tree. Of course, for many applications, the
899 use of strictly bifurcating trees is likely a reasonable simplifying assumption. But in lineages with
900 rampant hybridization, the reconstruction and analyses of phylogenetic networks will ultimately be
901 essential for a more accurate understanding of evolutionary history and processes.

902

903 **SUMMARY AND FUTURE DIRECTIONS**

904 Hybridization has been recognized for nearly a century as an important force in plant evolution, but we
905 are only now beginning to appreciate the extent of reticulation in shaping both genomes and their
906 resulting phenomes across the plant branch of the Tree of Life. Not only does hybridization frequently
907 tangle the tips of the Tree, it is also evident among the deeper branches, revealing past evidence of
908 reticulation that renders plant evolutionary history as a potentially complex network, the extent of which
909 is yet to be uncovered. This shift of perspective from a bifurcating tree to a network is not exactly a
910 surprise, but the phylogenetic depth of the network paradigm as revealed by genomic and phylogenomic
911 studies is reshaping our view of the structure of the Tree and requiring new developments in methods of
912 reconstructing, visualizing, communicating, and computing over plant evolutionary history. The genetic
913 consequences envisioned decades ago for recent hybridization are being reinterpreted to accommodate
914 deep time. Moreover, genome sequences and nuclear phylogenies are revealing the complex, mosaic
915 nature of plant genomes and the possible roles that hybridization (with or without WGD) may have
916 played in catalyzing plant diversity, potentially blurring the boundary between microevolutionary
917 processes and macroevolutionary patterns. We hope that this review will likewise catalyze new studies
918 that cross disciplinary boundaries to clarify the legacy and fate of hybridization in plant evolution.

919

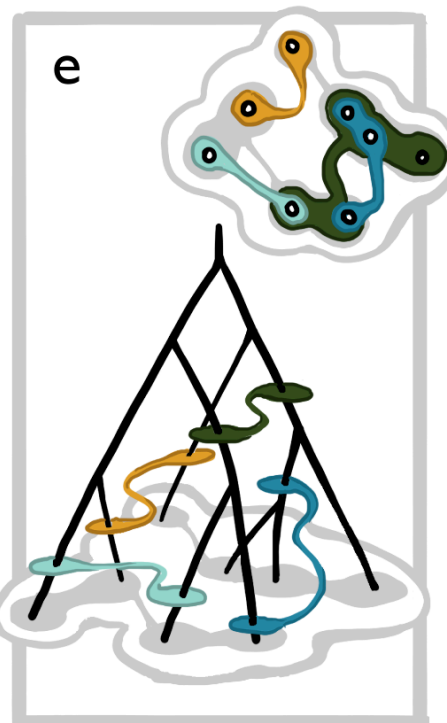
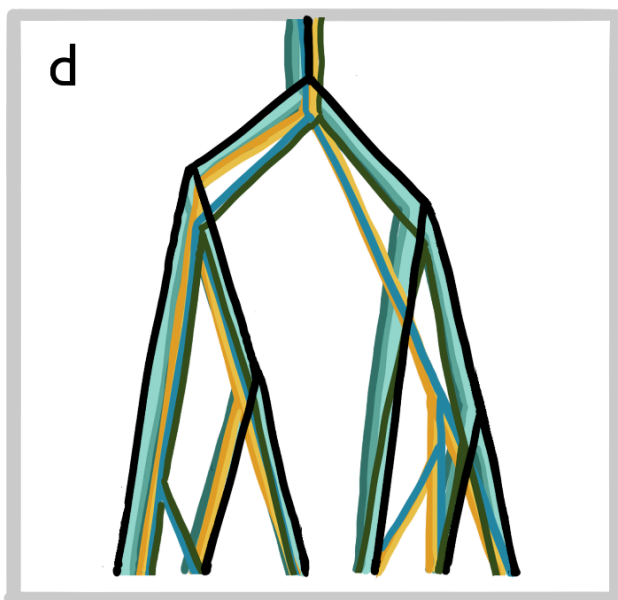
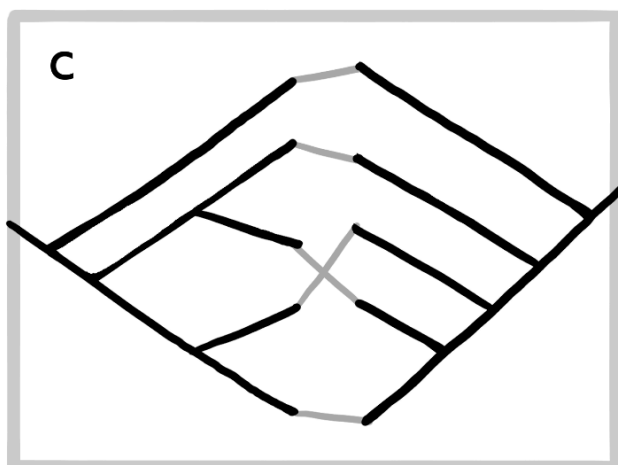
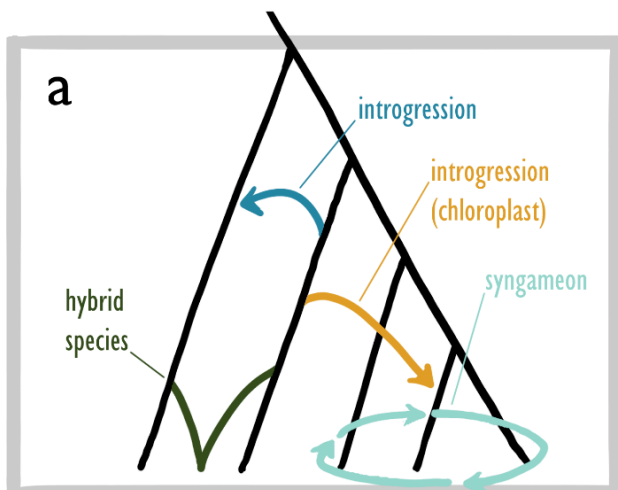


Figure 7. Different options for visual representations of reticulation. (a) Phylogeny with reticulation events depicted by arrows or lines between lineages connected by gene flow. This approach is commonly used to display results of phylogenetic network inference. This format is easy to read and can convey directionality (through arrows) and degree (through line weight), but does not directly show alternative topologies. (b) Main phylogeny in front with alternative topologies behind in different colors, with widths at tips proportional to the number of gene trees supporting the topology for each tip. This format directly conveys the degree of gene flow and alternative topologies and emphasizes as well the mosaic nature of genomes. However, it becomes hard to read when depicting too many alternative topologies or too many tips. (c) Tanglegram, two alternative topologies facing each other, with connections between corresponding tips. This method is commonly used to depict differences between two main topologies from different sources, such as nuclear and chloroplast trees. This format allows for easy visualization of group-wide patterns and alternative topologies, but can only accommodate two topologies at once, and thus may not be able to accommodate complicated multi-event reticulation scenarios or fully capture the extent of gene-tree discordance. (d) Main phylogeny in front with a cloud of alternative topologies plotted behind, one for every locus analyzed. Commonly used for large, genome-wide datasets. This format depicts each alternative topology and its amount of support, making it useful for comparing overall trends in the dataset. However, it can be hard to read, depending on the number and distribution of alternative topologies. (e) Main phylogeny plotted in three dimensions, with reticulation events depicted by lines between lineages connected by gene flow. Tips of the phylogeny are projected onto a two-dimensional cross-section with groupings of shared ancestry in gray and groupings based on reticulation events in colors corresponding to lines in three-dimensional phylogeny. Similar in concept to (a), but the three-dimensional rendering may allow for depictions of more reticulation events on one tree, and the two-dimensional projection quickly shows “main” and “reticulation” clusters of evolutionary similarity. Currently, to our knowledge, there are no tools for depicting phylogenies in three dimensions or tools

955

956

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962

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