The role of selection and evolution in changing parturition date in a red deer population

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Abstract

Changing environmental conditions cause changes in the distributions of phenotypic traits in natural populations. However, determining the mechanisms responsible for these changes and, in particular, the relative contributions of phenotypic plasticity vs evolutionary responses, is difficult. To date, to our knowledge no study has reported evidence that evolutionary change underlies the most widely-reported phenotypic 5 response to climate change: the advancement of breeding times. In a wild population of 6 red deer, average parturition date has advanced by nearly two weeks in four decades, and within-individual phenotypic plasticity in response to warming temperatures explains a minor part of this advance. Here we show that parturition date is also heritable and under selection towards earlier dates, and that genetic changes likely also 10 played a role in the shift towards earlier parturition dates. The observed rate of 11 evolution matched the predicted response to selection, and was less likely to be due to 12 genetic drift. Our study provides a rare example of observed rates of genetic change 13 matching theoretical predictions, although the match would not have been detected 14 with a solely phenotypic analysis. It also provides, to our knowledge, the first evidence 15 of both evolution and phenotypic plasticity contributing to advances in phenology in a 16 changing climate. 17

Introduction

Climate change affects various aspects of biodiversity across the planet (e.g., [1,2]). In 19 particular, shifts in phenotypic distributions within populations are widely reported, for a variety of morphological, phenological or life-history traits [2–4]. Surprisingly, however, 21 little is still known about the relative contributions of mechanisms underlying these 22 shifts [5]. Within a population, phenotypic distributions may change due to a change in 23 population structure (e.g., age-structure or sex-ratio), due to phenotypic plasticity 24 (within or between individuals), and due to genetic change [6-8]. The exact mixture of 25 mechanisms driving phenotypic change will determine the future of a population facing 26 a prolonged change in environmental conditions [9], for several reasons. First, the 27 consequences of changing population structure are variable and may be idiosyncratic (e.g., [8,10]). Second, phenotypic plasticity can provide an efficient way to cope with a changing environment but its effect may be short-lived and even maladaptive [11-13]. 30 Third, genetic evolution, when driven by natural selection, can improve population 31 growth rate, potentially contributing to long-term population persistence [12]. 32

In wild populations the respective contributions of plasticity vs evolution remain 33 unknown for the vast majority of documented phenotypic changes [14, 15] (note that by 34 evolution we mean genetic change, here and in the rest of the manuscript). To date, 35 most of the evidence for evolutionary responses to climate change comes from plants [16]. In contrast, despite numerous examples of phenotypic changes apparently related to 37 climate, there have been surprisingly few examples demonstrating unambiguously that a vertebrate population is evolving in response to climate change (see discussions 30 in [17-20]). This lack of evidence may in part be due to the question not being 40 prioritized [14,15]. However it probably also reflects the substantial challenges inherent 41 in testing for adaptive evolution, in terms of requirements for appropriate data and 42 statistical methods. For wild populations in which experimental manipulations are not 43 feasible, the most plausible means of testing for the genetic basis of phenotypic changes 44 is to use long-term pedigree data to test for changes in 'breeding values', the estimated 45 genetic merit of individuals as ascertained from the phenotypes of their relatives [21]. This needs to be done with care, as trends in predicted breeding values can be 47 confounded with environmental trends unless appropriately controlled for [22], and 48 precision of estimates of evolutionary rates can be inflated if the correlation structure of breeding value estimates is not properly handled [23]. To date, three recent studies of wild vertebrate populations using methods that account for uncertainty in breeding value predictions have found evidence of a genetic change underlying phenotypic change in morphology, all in line with selection pressures changing with climate: plumage colouration in collared flycatchers [20], and body size in Siberian Jays [24] and in snow voles [25]. However only with more empirical studies explicitly testing for evolution will it become possible to say whether the current lack of evidence also reflects a general slow rate of adaptation to environmental change in natural populations.

Climate change may have impacts on numerous aspects of an organisms' biology, but 58 phenology (i.e., the seasonal timing of life-history events) appears to have been 59 particularly affected [3,26,27]. Dramatic changes of phenologies in response to earlier 60 onset of spring are particularly well documented in mid- and high-latitude passerines, 61 where breeding times are occurring earlier in numerous populations and species [18,28]. 62 The study of avian systems in particular has shown that a fine tuning of phenology to 63 the climate is crucial in determining individual fitness. Mismatches between mean 64 breeding date and a fitness optimum that shifts with climate may re-shape selective pressures and hence potentially reduce population growth rate [29], although establishing the link between individual-level and population-level processes is not trivial [30,31]. The effects of climate change on mammalian phenology are less well documented than those of birds, and may be even more complex because mammals' long gestation times may make their breeding phenology sensitive to climate across a 70 longer time-frame [17]. Finally, despite the extensive evidence for phenotypic shifts in 71 phenology, the few studies that test for a genetic basis to changes in phenology in wild 72 populations have not found evidence of changes in breeding values [32-35]. 73

In a population of red deer (*Cervus elaphus*, Linnaeus 1758) on the Isle of Rum, NW Scotland, parturition date has advanced at a rate of 4.2 days per decade since 1980, a change that has been linked to temperatures and other weather conditions in the year preceding parturition, around the time of conception [36,37]. Previous studies of this population have shown that phenotypic plasticity and population structure explain a substantial proportion (23%) of the advance in parturition dates [37], and also that within-individual plasticity is sufficient to explain the relationship between temperature and parturition date [38]. However, the documented plasticity does not explain the majority of the observed phenotypic change, leaving room for processes that have not been investigated as of yet. It is plausible that evolution plays a role because the observed phenotypic change is qualitatively consistent with a genetic response to selection: parturition date is heritable in this population [39] and also under selection for earlier dates [40].

In this study we use quantitative genetic animal models [21, 41] to estimate the rate 87 of evolution in parturition date and the contribution of plastic and demographic processes to the observed shift in phenology in the Rum red deer study population. We 89 start by considering the response to selection that might be expected from the observed strength of selection and heritability of parturition date, based on a simple "breeder's 91 equation" prediction [42]. One of the most striking conclusions from the recent 92 application of quantitative genetic theory in evolutionary ecology has been the failure of 93 univariate breeder's equation predictions to capture trait dynamics in wild 94 populations [43, 44]. This may be for multiple reasons, foremost of which is likely to be the unrealistic assumption that only the focal trait is relevant. We therefore also 06 consider a multivariate breeder's equation [45], and ask how selection on offspring size and the genetic correlation between parturition date and size alters the expected 98 evolutionary response. However a second, less well-explored reason for the failure of the theory is the comparison of predicted response to observed rates of phenotypic change, 100 which will obviously be affected by other processes, rather than with estimates of the 101 underlying rates of genetic change. As the central analysis of this work, we use trends in 102 breeding values to estimate the rate of evolution in parturition date and to test whether 103 it is compatible with the response to selection predicted by either the univariate or 104 multivariate breeder's equation, or with genetic drift. Finally we quantify the effect of 105 non-genetic processes contributing to phenotypic change along with evolution. 106

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Methods

Study population

We used data from a long-term study of the unmanaged population of red deer living in the North Block of the Isle of Rum, Scotland $(57^{\circ}01' \text{ N}, 6^{\circ}17' \text{ W})$, for the years 1972-2016. Within the ca. 12 Km² of the study area, calves are marked with ear tags (and a collar for females) shortly after birth, in order to record detailed life-histories of individuals throughout their lives [46]. DNA was obtained from ear punches, post-mortem tissue and cast antlers. The population pedigree was reconstructed from single nucleotide polymorphisms as in [47], using the R package SEQUOIA [48].

We studied the selection and genetics of parturition date, the date on which a female 116 gave birth to a calf in a given year. We therefore focus on females, because males do not 117 express the trait of parturition date —though they may affect it, in both genetic and 118 non-genetic ways. Males were retained in the pedigree and contributed to the 119 calculation of quantitative genetic parameters by informing the relatedness between 120 individuals. Selection was estimated from the association between a trait and individual 121 lifetime breeding success, where lifetime breeding success was the number of offspring 122 produced by an individual across their lifetime, whether or not they survived to 123 breeding and therefore also had parturition records (further details below). Parturition 124 date being a sex-limited trait, selection differentials on parturition date were divided by 125 2 after estimation (i.e., half the population was assumed not to be selected for that 126 trait). We included females that are still alive, even though their lifetime fitness is still 127 unknown, in order not to introduce a fraction of individuals missing not at random with 128 respect to fitness and parturition date. However, excluding living females from the 129 analysis gave indistinguishable results. 130

There has been no culling in the study area since 1973, but individuals are occasionally shot when they visit areas surrounding the study area. Mortality due to culling may exert a kind of artificial selection that studies of natural selection may want to exclude. However, our goal here was to understand the causes of phenotypic change, be they natural or artificial. We therefore retained culled females in our main analyses. These shot females represented a small but non-trivial portion (15%) of the data set (Table 1). Therefore, in a sub-analysis, we also considered selection only among females

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who died of natural causes (i.e., excluding both shot females and females still living), and discuss the consequences of culling for selection and evolution in this system.

Parturition date was measured as the number of days after May 1, because virtually 140 no parturition occurs before that date. Values were (natural-)log-transformed in order 141 to obtain residuals with distributions close to Gaussian. The logged values were 142 multiplied by 100 for reporting convenience (in particular, variance components would 143 have been of the order 10^{-5} without this second step). The working phenotype in all 144 models was therefore $z = 100\log(B)$, where B is the parturition date in number of days 145 after May 1^{st} . Results were converted back to days (see SI 1 for details of the 146 back-transformation process) to facilitate biological understanding. We report results 147 using untransformed data in SI 5. 148

Data type	Number of	Excluding shot	Shot	Total
Parturition date	Individuals Records	$582 \\ 2921$	$\begin{array}{c} 158 \\ 463 \end{array}$	$\begin{array}{c} 740\\ 3384 \end{array}$
Lifetime breeding success	Individuals	1614	282	1896

Table 1. Sample sizes for lifetime breeding success (LBS) and parturition data for years 1972-2016. All numbers refer to females; parturition date is measured repeatedly on individuals. LBS is measured over a lifetime, and there is only one measure per individual. All females with a unique ID have an LBS record, including those that died as calves and therefore did not breed and do not have records for parturition date.

Quantitative genetic analysis

Univariate animal model

We fitted a univariate animal model of (transformed) female parturition date in order to 151 estimate heritability and change in breeding values [21, 41]. The fixed effects in the 152 model were: the sex of the offspring; the female's 'reproductive status', which can take 153 five values to represent different recent reproductive history: 'naive', 'true yeld', 154 'summer yeld', 'winter yeld', 'milk hind' [38]; the female's age in years (first and second 155 order polynomial); a contribution of 'genetic group', to model gene flow into the 156 population, estimated as her expected level of immigrant vs resident genes (see SI 2); 157 the calf's birth year as continuous variables, see next section for details; and the 158 female's pedigree-based inbreeding coefficient [49,50] calculated using the R package 159 MCMCglmm [51]. Population density, estimated as the number of resident adult 160

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females in a given year, had a significant effect on (log) parturition date at the beginning of the study period (e.g., between 1974 and 1987, [46]), but we found no effect in the full data-set (slope -0.38, standard error 0.63) and so did not include density in the final models.

The random effects decomposed the variance not accounted for by fixed effects into six components: additive genetic variance; 'permanent environment' variance (estimable from the repeated measures of the same females across multiple years; [52]); maternal effects variance (i.e. associated with the mother of the breeding female, and hence grand-mother of the new calf); variance associated with the calf's birth year; variance associated with the breeding female's (i.e. mother of the calf) birth year; and residual variance.

Thus, the model of (log-transformed) parturition date (z) of female *i* in year *j* can be written

$$z_{ij} = \mu + X^T b + a_i + p_i + m_i + c_j + y_i + r_{ij}$$
(1)

where μ is an intercept, X is a vector of fixed predictors (including calf's birth year), **b** 174 is a vector of fixed effects, a, p, m, c and y are random effects with which to estimate 175 the variance associated with additive genetic values (i.e., breeding values), permanent 176 environment, maternal (i.e. grand-mother of calf), calf year of birth, and female's year 177 of birth, respectively, and r is the residual. The breeding values (a) are normally 178 distributed as $(a_1, \ldots, a_n)^T \sim N(0, \sigma_A^2(z)\mathbf{A})$, where $\sigma_A^2(z)$ is the additive genetic 179 variance for (\log) parturition date, n is the number of females, and A is the relatedness 180 matrix between individuals. The heritability of (log) parturition date was estimated as 181 $\sigma_A^2(z)$ divided by the sum of all the variance components. 182

We used this animal model to estimate the individual-level repeatability (in addition to the heritability) of parturition date, as the sum of the proportions of variance explained by all effects that are constant for an individual: inbreeding, female's cohort and genetic group (all of which are fixed effects) and additive genetic variance, permanent environment variance, maternal variance and female's cohort variance (that is, all random effects but offspring birth year and residual).

We ran all models in MCMCglmm [51] with Gaussian errors for (log-transformed) ¹⁸⁹ parturition date. We report posterior modes and 95% highest posterior density credible ¹⁹⁰

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intervals. For this univariate model, we used the default inverse gamma priors for variance components, with shape and rate parameters both equal to 0.001 (equivalent to a variance and degree of belief of 1 and 0.002, respectively). We run models for 130000 Markov chain Monte Carlo iterations, with a burn-in of 30000 and thinning of 100.

Selection

We estimated selection acting on parturition date by assessing the association between a female's parturition dates and her fitness. We measured fitness as *lifetime breeding* success (LBS in the text, W in equations), which is the number of offspring produced by an individual, calculated for all females in the database, whether or not they survived to breeding and therefore also had parturition records. 200

Selection was estimated using a model of the covariance between parturition date $_{201}$ and fitness. We used a bivariate generalized linear mixed model, with LBS modelled as $_{202}$ an over-dispersed Poisson trait (with log link function) and log-transformed parturition $_{203}$ date (z) modelled as a Gaussian trait. This model can be written as $_{204}$

$$[z, W] \sim Xb + D_1m + D_2y + D_3c + D_4p + Ir$$
(2)

where Xb represents fixed effects (the same fixed effects for parturition date as above, and only an intercept and genetic group for fitness), m, y, c, p are random effects associated with maternal effects (the identity of the mother of the breeding female), the year of calving, the female's cohort (i.e. her year of birth), and the individual female's identity (or 'permanent environment' effect, because of the repeated measures), respectively. D-matrices link random effect levels to observations, and Ir represents the residuals.

Note that W is only measured once for each individual, unlike the repeated measures on parturition date (z). For W, variance components are therefore null for y (the calf's year of birth) and p (the permanent environment component of a trait, derived from repeated measures). MCMCglmm accommodates the difference in replication between the use two traits by allowing the individual-level random effect p for the replicated trait (parturition date) to covary with the residual variance r of the non-replicated trait (fitness), thus providing a covariance between the repeatable part of an individual's 218

parturition date and her fitness (for a comparable example, see [53]).

The selection differential on parturition date was calculated as the sum of this 220 individual-level covariance, plus the maternal-effect covariance between parturition date 221 and fitness (i.e. covariance among effects of the breeding females' mothers on their 222 daughters' parturition dates and fitness). Selection differentials characterize the 223 within-generation change in phenotype due to selection. We therefore standardized this 224 value by generation time (8 years) to be expressed in rate of change per year, or in total 225 change over the study period. As stated above, the selection differentials were also 226 divided by two, because the covariances were estimated from females only. Males do not 227 express the trait but nevertheless carry genes relevant to parturition date in females. 228 Selection on parturition acts on only half of the population, and the expected response 229 to selection is half that predicted from the strength of selection in females. We also 230 estimated a selection gradient [55], calculated as the selection differential divided by the 231 corresponding variance (that is, the sum of the individual-level and mother-level 232 variance components for parturition date). 233

When expected fitness follows a log-normal distribution, selection parameters can be equivalently calculated on the scale of the data using relative fitness, or on the log-scale using absolute fitness [54]. Because our model uses a log-link function for absolute LBS, its parameter estimates are on the latent scale, but these are directly interpretable as selection differentials and selection gradients relating to relative fitness on the data scale. 236

For multivariate models we used parameter-expanded priors for variance components 239 (with working mean of 0 and variance of 1000). We ran these models for 260000 Markov 240 chain Monte Carlo iterations, with a burn-in of 60000 and thinning interval of 200. 241

Univariate and multivariate predictions of evolution

The response to selection (the per-generation change in the mean value of the trait) was predicted as the product of the heritability in parturition date and the selection differential, following the univariate breeder's equation [42]. The equation was applied constrained from the model of log-transformed parturition date data, and the predicted response was subsequently back transformed to days. Calculations were done on the MCMC posterior distributions of the heritability and the selection differential, in order to propagate the uncertainty in these two parameters. The univariate breeder's equation is the selection in the selection is the selection differential in the selection differential in the selection differential.

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equation ignores the fact that the adaptive evolution of a focal trait depends not only 250 on direct selection on that trait, but also on selection on those traits that are genetically 251 correlated with the focal trait [45]. This assumption may explain in part the common 252 mismatch between predicted and observed evolution in natural populations [44], but it 253 can partly be relaxed by incorporating analyses of relevant associated traits and 254 estimating multivariate selection and genetic covariances: the multivariate response to 255 selection can then be predicted as the product of the genetic variance-covariance matrix 256 **G** and the vector of multivariate selection gradients $\boldsymbol{\beta}$ ($\Delta \boldsymbol{Z} = \boldsymbol{G}\boldsymbol{\beta}$) [45,55]. 257

In the Rum red deer study population, a calf's birth date is correlated with its birth ²⁵⁸ weight [37, 46], a trait also under selection [40]. We therefore applied a bivariate ²⁵⁹ breeder's equation to parturition date and calf birth weight to estimate the effect of ²⁶⁰ indirect selection on the predicted evolutionary response of parturition date to selection. ²⁶¹

We extended the animal model of parturition date (eq. 1) to a bivariate animal 262 model of parturition date and offspring birth weight, using the same fixed effects and 263 random effects for both traits. Note that in this model, the calf's birth date and birth 264 weight (bw) are both being treated as the phenotype of the mother; the treatment of 265 offspring birth weight as a trait of the mother is justified by the observation that more 266 than 90% of the genetic variance in birth weight is maternal-genetic variance rather 267 than direct-genetic variance [52]. This model estimated an additive genetic covariance 268 between the two traits, $\sigma_A(z, bw)$, which can be divided by the square root of the 269 product of the two additive genetic variances, $\sigma_A^2(z)$ and $\sigma_A^2(bw)$, to obtain a genetic 270 correlation. 271

Finally, we extended the bivariate selection model (eq. 2) to a trivariate model also 272 including offspring birth weight (along with parturition date and LBS). For birth weight 273 we used the same fixed and random effects as described above for parturition date. We 274 summed the appropriate covariances and divided by the corresponding variance 275 parameters to estimate β_z , the direct selection gradient on parturition date corrected for 276 the indirect selection on birth weight, and β_{bw} , the direct selection gradient on birth 277 weight corrected for the indirect selection on parturition date. The response to selection 278 could then be calculated as $\beta_z \sigma_A^2(z) + \beta_{bw} \sigma_A(z, bw)$ [55]. 279

We also expressed predicted rates of evolution in Haldanes, that is, in units of standard deviation per generation [56]. We did not express the results in Darwins because parturition dates have no natural zero point, and therefore mean-standardisation is not meaningful.

Components of change

Trend in breeding values

Using the univariate animal model of parturition date containing year as a covariate (see below), we fitted a linear regression of best linear unbiased predictors (i.e., model 287 predictions for the values of a random effect levels, BLUPs hereafter) for individual 288 females' breeding values against the mean birth year of their offspring to each posterior 289 sample. This generates a posterior distribution for the slope of change in mean breeding 290 value [23]. In addition, to visualize potential non-linearity in genetic change, we fitted a 291 smoothing spline function of female cohort year to the BLUPs for individual breeding 292 values for every posterior sample, thus generating the posterior distribution of the 293 time-dynamic of breeding values among cohorts [57]. Changes in breeding values may 294 indicate a response to directional selection, but they can also be produced by random 295 fluctuations under non-directional evolutionary models, such as genetic drift. To assess 296 this possibility, we also compared the posterior distribution of the estimated change in 297 breeding values to the change possible under genetic drift alone, using simulations as 298 described in [23]. 299

In general, breeding values predicted by animal model BLUPs are not equal to the 300 true breeding values, but are influenced by environmental random deviations [22]. As a 301 consequence, a linear regression of BLUPs may confound genetic and non-genetic (e.g. 302 plastic) change and may produce a biased estimate of evolution. This issue can be 303 addressed by including year as a covariate in the animal model used to obtain BLUPs. 304 Unfortunately, the solution is conservative, because the animal model ascribes some of 305 the genetic change to the year effect [22]. We opted to report primarily conservative 306 estimates of evolution, based on an animal model that did contain calf cohort as a 307 covariate (see above). Nevertheless, we also re-fitted the animal model without a fixed 308 effect for cohort and re-calculated the change in BLUPs for breeding values estimated 309 this way; we report this second estimate in the discussion. 310

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Other contributions to phenotypic change

Finally we estimated the contributions of several other terms in equation (1) to the 312 trend in parturition date. We used Geber's method [6,8] on model predictions to 313 estimate the independent contribution of changes in the class structure of age and 314 reproductive status, and the independent contribution of changes in levels of inbreeding 315 (as assessed from the pedigree inbreeding coefficient) and gene flow (as assessed by the 316 genetic groups effect) to the phenotypic change in parturition date. Briefly, this method 317 estimates the contribution of change in a parameter mean (\bar{k}) to change in a trait mean 318 (\bar{z}) as the product of the partial derivative of z on k $(\frac{\partial \bar{z}}{\partial \bar{k}})$ and the slope of k on time 319 $(\frac{\Delta \bar{k}}{\Delta t})$. We applied the equation to each sample of the posterior distributions in order to 320 propagate the uncertainty in the estimated trends. In addition to calculating the net 321 effect through the study period, we calculated $\frac{\partial \bar{z}}{\partial \bar{k}} \bar{k}_t$ for each year t to visualize the 322 dynamic of changes in effects through time graphically. 323

We did not use random effects to estimate non-genetic components of change 324 because random effects other than additive genetic effects are linearly independent of 325 years by construction, and any change in females' maternal effects or permanent 326 environment should be absorbed into the fixed effect of offspring cohort. 327

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Results

Phenotypic change

Average parturition dates became later from 1972 to 1980 (probably reflecting increased population density, [46]), after which they advanced at an apparently constant rate (Fig. 1). A linear regression estimates the change in parturition date to be a total of -12.3 (95%CI [14.6; 10.1]) days over the 45-year study period (from 1972 to 2016). 333



Fig 1. Phenotypic trend in red deer parturition dates, in days after May 1st. Large black dots represent annual means, small grey dots represent individual parturition dates, with the darker shades indicating more calves being born on a given day. About 4% of individual parturition dates fall outside the plotted region (10th May - 12 July; note these are still included in the analyses). The red lines represent the slope and associated 95% confidence interval of a linear regression of all individual parturition dates on year of parturition. Note that the years 1972-1975 have very negative residuals and that the rate of change over 1980-2016 is slightly underestimated by the linear regression being fitted over all years.

Sources of parturition date variation

Parturition date was influenced by a female's reproductive status and age, but there was no clear evidence for effects of inbreeding, of offspring sex or of the proportion of

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immigrant genetic ancestry (SI Table 4.1). Parturition date was heritable, with additive 337 genetic variance accounting for 18% (95%CI [10%; 23%]) of phenotypic variation 338 corrected for fixed effects and variation among years and among cohorts (SI Table 4.2). 330 The individual-level repeatability of parturition date (estimated as the sum of 340 proportions of all variance components except offspring birth year and residual) was 341 19%, of which additive genetic variance was most important, with permanent 342 environment effects and maternal effects both accounting for less than 1% of total 343 phenotypic variation (SI Table 4.2). The random effect for offspring birth year (which 344 captures the variance corrected for the temporal trend) accounted for about 8% of the 345 phenotypic variance (SI Table 4.2). Note that proportions are essentially invariant 346 under monotonic transformation and that these proportions of variances are equivalent 347 on the transformed and on the data scale. 348

Univariate selection and predicted response

Females with earlier parturition dates had, on average, higher lifetime breeding success: ³⁵⁰ the selection differential of parturition dates estimated with LBS was -9.08 days (³⁵¹ 95%CI [-14.91; -3.81]). Given the heritability of parturition date of 0.16, the³⁵² univariate breeder's equation predicts a total response to selection of -1.37 days over³⁵³ the 45-year study period (95%CI [-3.01; -0.60]) (Fig. 4A). This corresponds to -0.031³⁵⁴ [-0.068; -0.014] days per year, -0.25 [-0.55; -0.11] days per generation, or -0.019³⁵⁵ [-0.042; -0.008] Haldanes.³⁵⁶

Selection was stronger among females died of natural causes than among females ³⁵⁷ that were culled (SI Fig.S3). Using the subset of females who died of natural causes, the ³⁵⁸ univariate breeder's equation predicts a response of -2.04 [-3.37; -0.95] days over the ³⁵⁹ study period. In contrast, using the subset of females who were culled, the univariate ³⁶⁰ breeder's equation predicts a response of 0.11 [-0.64; 0.93] days over the study period. ³⁶¹

Bivariate selection and predicted response

Conditional on the fixed effects affecting each trait, the phenotypic correlation between $_{363}$ parturition date and birth weight was positive but weak (correlation = 0.12, 95 %CI $_{364}$ [0.05; 0.16]). The gradient of direct selection on parturition date was negative (mode $_{365}$

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 $\beta_z = -0.0003, 95 \%$ CI [-0.0004; -0.0002]), and that on birth weight was positive ($\beta_{bw} =$ 366 0.0138, 95% CI[0.009; 0.017]). There was also additive genetic variance in offspring birth 367 weight (0.68, 95%CI [0.57; 0.90), corresponding to a heritability of 0.46 (95%CI 368 [0.37; 0.62]). The additive genetic covariance between parturition date and offspring 369 birth weight was -1.78 (95%CI [-4.38; 0.56]), corresponding to a weak negative genetic 370 correlation of -0.16 (95%CI [-0.32; 0.05]). The multivariate breeder's equation predicts 371 a rate of evolution of -1.31 days (95%CI [-2.46; 0.10]) over the study period, which is 372 actually very similar to the univariate breeder's equation prediction of -1.37 days 373 (difference = -0.15 days, 95% CI [-1.59; 1.03], Fig. 4B)374

Genetic contribution to phenotypic change

The slopes of the linear regressions of BLUPs for parturition date breeding values on 376 birth year, integrated over the posterior distribution, suggests an advance in breeding 377 values, with the slope estimated at -0.10, 95% [-0.23; 0.03] per year on the 378 log-transformed scale. Time-splines fitted on the posterior distribution of the BLUPs 379 visually support a linear decrease in breeding values (Fig. 2). This rate of evolution 380 corresponds to a total change over the study period of -2.1 days, 95%CI [-4.5; 0.7] due 381 to genetic change (Fig. 2 and 4C). This is equivalent to -0.045 [-0.100; 0.018] days per 382 year, -0.36 [-0.79; 0.14] days per generation, or -0.028 [-0.062; 0.01] Haldanes. 383

9% of the simulations of genetic drift generated an advance as large or larger than 384 the change estimated from the BLUP linear regression (using the posterior mode for the 385 BLUPs trend as a point of comparison, see Fig. 4D). Inbreeding tended to delay 386 parturition date (SI 4.1), and given that the estimated pedigree inbreeding inevitably 387 increased over time with increasing pedigree depth [49], there was marginal evidence 388 from our model of inbreeding postponing parturition date by 0.38 days (95%CI 389 [-0.04; 1.01]) over the study period, thus opposing the phenotypic trend. However this 390 prediction may be spurious, because the increase in inbreeding coefficient was an 391 artifact of estimating inbreeding from a pedigree [49]. Re-running the model without 392 inbreeding led to almost identical estimates for all other parameters. The effect of gene 393 flow (proportion of immigrant genotype) was very uncertain (SI 4.1) and its overall 394 predicted effect over the study period was a change of 0.15 days (95% CI [-0.34; 0.72]). 395



Fig 2. Trend in breeding values for parturition date. Each black line was obtained from a different MCMC posterior sample, by fitting a spline to the mean of estimated breeding values among individuals living in the same year. The y-axis was centered on the mean breeding values in 1972 to help interpretation.

Non-genetic contributions to phenotypic change

As in previous work [37], we found that mature females tended to give birth earlier than 397 younger females, but very old females gave birth the latest. The effects on mean 398 parturition dates of changes in the age structure tended to be in the opposite direction 399 to the observed phenotypic change: during the first ten years of the study, the mean age 400 of females in the study increased steadily, pushing towards earlier mean parturition 401 dates (-3.68 days, 95% CI [-5.63; -1.92] from 1972 to 1981). For the rest of the study, 402 the change in age structure tended to delay mean parturition date slightly (0.57 days)403 [0.39; 0.71] from 1982 to 2016). Over the study period the change in age structure had a 404 predicted net effect of -0.58 days [-1.67; 0.40] (Fig. 3A). Changes in female 405 reproductive status had a fluctuating effect on parturition date (Fig. 3B), with a 406



Fig 3. Predicted effect of (A) age-structure and (B) female reproductive status on parturition date across years. The origin of the y-axis is arbitrarily set to the predicted effect in the first year. The red thick dashed lines represent the net effect of changes in age structure and female reproductive status on parturition date reported in the text. The thin dotted lines in (A) represent the effect of changes in age structure before, and after 1981, respectively.

resulting total effect over the study period of -0.32 days (95%CI [-0.87; 0.17]).

Offspring sex had no clear effect on parturition date, and since sex-ratio at birth 408 remained stable over the study period (despite an early decline in the proportion of 409 males [58]), this parameter is predicted to have had no noticeable effect (-0.04 days, 410 95%CI [-0.18; 0.04]). 411

Finally, the fixed covariate of calf birth year (SI Table 4.1) captured trends across 412 years that should ideally be unrelated to genetic change, age, and reproductive status, 413 although it will inevitably be partly confounded with genetic change (see Discussion). It 414 may also, similarly, capture persistent changes in maternal effects, permanent 415 environment effects, and various plastic processes. The coefficient (back-transformed) 416 corresponds to a change of -9.3 days (95%CI [-11.9; -2.22]) over the study period, 417 showing that most of the observed change remains unexplained by our model. However, 418 note that some effects (e.g., inbreeding and gene flow) opposed the phenotypic change, 419 and that the model explained more than the difference between the birth year effect and 420 the total phenotypic change. 421



Evolution (days)

Fig 4. Posterior distributions of the parameter estimates for change in parturition date over the study period: (A), (B) the predicted evolutionary response to selection from the univariate and bivariate breeder's equations respectively; (C) the estimated contribution of evolution (from the trend in predicted breeding values); and (D) evolutionary change possible due to genetic drift only. The distributions all have the same area. The dashed line indicates the mode of the distribution (C), the contribution of evolution. The response to selection was estimated using univariate and bivariate breeder's equations, where phenotypic multivariate models gave selection gradients, and animal models gave additive genetic variance-covariances of parturition date and birth weight. A univariate animal model was used to estimate the amount of evolution as the temporal trend in BLUPs for breeding values, and to simulate evolution by genetic drift. Parturition date was modeled using a log-transformation, and all estimates were subsequently converted to change in days over the study period (see SI 1). Parameter estimates are summarized in SI 4.

Discussion

In the Isle of Rum red deer study population, average parturition dates have advanced 423 12.3 days over the last 35 years. Previous research has identified the contribution of 424 plastic changes in response to warming temperatures to this change [38]. Here we have 425 shown that adaptive evolution likely played a role too. Below we discuss the significance 426 of this result for the red deer population, and also the strengths and challenges 427 associated with the quantitative genetic study of evolution in wild populations. 428

Moyes et al. [36] identified the trend toward earlier parturition dates in the Isle of 429 Rum red deer population, and related a substantial component of it to local climate 430 warming. In addition, within-individual plasticity is sufficient to explain the relationship 431 between temperature and parturition date, and plasticity in response to increasing 432 temperature explains a change of -2.8 days over the study period, which is equivalent 433 to 23% of the total phenotypic change [38]. There is little evidence of variation among 434 females in their plastic responses to temperature [38]. Therefore the plastic response to 435 temperature is unlikely to have changed (by genetic evolution or other means) over the 436 study period, and a change in the shape of individuals' plastic responses (reaction 437 norms) probably did not contribute to the change in mean parturition dates. 438

The present work thus reveals a major new aspect of the complex picture of the 439 dynamics of parturition date in this population, by identifying a role for evolution 440 concurrent with the previously-identified plastic responses. We estimated that evolution 441 for parturition date accounted for a total change of -2.1 days (95% CI [-4.5; 0.7 days])442 over the study period. This estimate relies on the modern and conservative version of 443 BLUP-regression, which accounts for criticisms made in [23] and [22], in particular by 444 including calf' year of birth as a covariate. Taking this approach yielded a conservative 445 estimate of genetic change that accounted for 15% of the observed phenotypic change. 446 As expected, the less conservative alternative of not including year as a covariate gave a 447 more rapid estimate of evolution: -2.4 days (95% CI [-4.9; -0.2 days]). The true rate 448 of evolution probably lies between the conservative and the less conservative estimates. 449 We obtained almost identical results from animal models fitted to the untransformed 450 data (see SI 5), although these models performed relatively poorly (skewed residuals 451 and poor MCMC mixing) which may impair the reliability of estimates. 452

Our results suggest modest roles for changes in demographic structure (and 453 essentially no role for changes in offspring sex-ratio). Shifting proportions of females of 454 different reproductive status and ages had a predicted combined effect of -0.9 days 455 (about 7% of the phenotypic change). These effects were also identified in [37]. Changes 456 among individuals, other than change in breeding values, therefore probably explains 457 only a small (but non-negligible) fraction of the observed phenotypic change. However 458 summing the effects of genetic change, plasticity in response to temperature [38] and 459 changing demographic structure still leaves 55% of the change unexplained. Plastic 460 responses to other environmental variables likely account for some of the remaining 461 change, since the calculation in [38] does not consider the response to any variables 462 other than mean temperature during a five month period. In particular, other climatic 463 variables such as average temperatures during other times of the year, temperature 464 variability, rainfall and wind speeds probably affect reproductive traits in the red 465 deer [37]. In addition, the evolution of indirect genetic effects [59] may play a role. 466

The indication of evolution towards earlier parturition dates is consistent with 467 previous work, which found the trait to be heritable [39] and under selection for earlier 468 dates [40] in this population. Under ideal conditions, the product of heritability and 469 strength of selection predicts the evolutionary response to selection [42, 60]. However, 470 this "breeder's equation" frequently fails to give reliable predictions in wild 471 populations [43, 60]. Simultaneous selection on genetically correlated traits is likely to 472 be a major cause of this failure, because fitness is generally causally affected by many 473 traits and genetic correlations are common [44]. Here, however, we obtained a close 474 match between the estimated rate of evolution and the response to selection predicted 475 by the breeder's equation, both in its univariate and in its bivariate forms. We cannot 476 discard the possibility that this close match might be in part a coincidence, for instance 477 if the indirect response to selection on a trait not included in the analysis pulled 478 evolution in one direction but genetic drift pulled it back to match the observed rate of 479 evolution. Nevertheless, our results are consistent with selection acting on parturition 480 date directly (i.e., it was not significantly affected by selection on birth weight), so that 481 its evolutionary trajectory can be predicted from a univariate breeder's equation [60]. 482

We estimated evolution and selection averaged over the study period to obtain the total evolution and response to selection expected over the period. However if an increase in temperature explains selection for earlier parturition, it is possible that 485 selection has intensified in more recent years, and that selection was strongest in 486 warmer years (e.g., [61]). The multivariate models we used to estimate selection allowed 487 the estimation of selection by correcting for fixed and random effects in both parturition 488 date and fitness traits, but are not well suited to estimate changes in selection. Future 489 work could verify the selective scenario by estimating the interaction between 490 parturition date and temperature in a generalized linear model of fitness, but care 491 should then be taken to correct for the effect of time or other selectively irrelevant 492 aspects of variation in parturition date. 493

A changing climate is probably not the only selective agent relevant to the evolution 494 of parturition date in this red deer population. Indeed, selection was stronger among 495 females who died of natural causes (with a predicted response to selection of -2.0 days) 496 than among the whole population, which includes shot females (with a predicted 497 response of -1.37 days), and especially so among shot females only (+0.10 days). 498 Culling may alter selection on parturition date, possibly by removing females from the 499 population at random with respect to their potential parturition dates, thus diluting 500 natural selection. Alternatively, culling may not be random with respect to parturition 501 date, but somehow exert a type of artificial selection for later parturition dates which 502 thus effectively opposes natural selection. Either way, culling may slow down the 503 adaptive response to natural selection in the population. If confirmed, this result would 504 add to the list of evolutionary consequences of culling [15, 57]. 505

Conclusion

The breeder's equation's prediction corresponds closely to the estimate of the 507 evolutionary rate obtained from the trend in breeding values in the deer population, but 508 it is important to highlight that this genetic change is much less than the observed 509 phenotypic change. This mismatch is not surprising given that several mechanisms of 510 phenotypic change, with a genetic basis or not, have been identified on the Rum red 511 deer population (in our analyses presented here as well as also [37,38]). More generally, 512 our results illustrate how phenotypic change can be simultaneously due to both plastic 513 and genetic changes [6, 8, 43]. Plastic changes in response to climate change appear 514

common, but that does not exclude concurrent evolutionary change in response to 515 climate change [14]. Evolutionary changes are more difficult to infer than plastic 516 changes, and few tests of evolution have been performed [14, 18, 43]. Moreover, here as 517 in other systems, non-trivial contributions of evolution may represent only a fraction of 518 the overall phenotypic trend [57, 62]. Evidence for plastic responses should not be taken 519 as reason to dismiss a role for genetic change [63, e.g.], nor the other way around. As 520 another side of the same coin, our results highlight the insights that a quantitative 521 genetic perspective brings to the study of phenotypic trait dynamics. As outlined above, 522 the breeder's equation often fails to predict phenotypic change in the wild. One possible 523 explanation for this failure is 'cryptic evolution', where genetic change is hidden by 524 plastic changes [43]. Our results illustrate that a simple application of the breeder 525 equation can work, but it should be tested by comparison to estimates of genetic 526 changes, not of phenotypic changes. 527

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References

- Mc Carty JP. Ecological Consequences of Recent Climate Change. Conserv Biol. 537 2001;15(2):320–331.
- Parmesan C. Ecological and Evolutionary Responses to Recent Climate Change. 539
 Annu Rev Ecol Evol Syst. 2006;37(1):637–669. 540

528

- Menzel A, Sparks T, Estrella N, Koch E, Aaasa A, Ahas R, et al. European
 phenological response to climate change matches the warming pattern. Glob
 Chang Biol. 2006;12(10):1969–1976.
- Gardner JL, Peters A, Kearney MR, Joseph L, Heinsohn R. Declining body size: 544 a third universal response to warming? Trends Ecol Evol. 2011;26(6):285–291. 545
- O'Connor MI, Selig ER, Pinsky ML, Altermatt F. Toward a conceptual synthesis for climate change responses. Glob Ecol Biogeogr. 2012;21(7):693–703.
- Hairston NG, Ellner SP, Geber Ma, Yoshida T, Fox Ja. Rapid evolution and the convergence of ecological and evolutionary time. Ecol Lett. 2005;8(10):1114–1127.
- Pemberton JM. Evolution of quantitative traits in the wild: mind the ecology.
 Philos Trans R Soc B. 2010;365(1552):2431–2438.
- van Benthem KJ, Bruijning M, Bonnet T, Jongejans E, Postma E, Ozgul A.
 Disentangling evolutionary, plastic and demographic processes underlying trait
 dynamics: a review of four frameworks. Methods Ecol Evol. 2017;8(1):75–85.
- Visser ME. Keeping up with a warming world; assessing the rate of adaptation to climate change. Proc Roy Soc B. 2008;275(1635):649–59.
- Ozgul A, Tuljapurkar S, Benton TG, Pemberton JM, Clutton-Brock TH, Coulson
 T. The dynamics of phenotypic change and the shrinking sheep of St. Kilda.
 Science. 2009;325(5939):464–7.
- Buskirk JV, Steiner UK. The fitness costs of developmental canalization and plasticity. J Evol Biol. 2009;22(4):852–860.
- Chevin LM, Lande R, Mace GM. Adaptation, plasticity, and extinction in a changing environment: towards a predictive theory. PLoS Biol.
 2010;8(4):e1000357.
- Duputié A, Rutschmann A, Ronce O, Chuine I. Phenological plasticity will not help all species adapt to climate change. Glob Chang Biol. 2015;21:3062–3073.
- Merilä J, Hendry AP. Climate change, adaptation, and phenotypic plasticity: 567
 The problem and the evidence. Evol Appl. 2014;7(1):1–14. 568

15	5. Pelletier F, Coltman DW. Will human influences on evolutionary dynamics in the	569
	wild pervade the Anthropocene? BMC Biology. 2018;16(1):7.	570
	doi:10.1186/s12915-017-0476-1.	571
16	6. Franks SJ, Weber JJ, Aitken SN. Evolutionary and plastic responses to climate	572
	change in terrestrial plant populations. Evol Appl. 2014;7(1):123–139.	573
17	7. Boutin S, Lane JE. Climate change and mammals: evolutionary versus plastic	574
	responses. Evol Appl. 2014;7(1):29–41.	575
18	3. Charmantier A. Gienapp P. Climate change and timing of avian breeding and	576
	migration: evolutionary versus plastic changes. Evol Appl. 2014;7(1):15–28.	577
10) Crozier I.C. Hutchings IA Plastic and evolutionary responses to climate change	570
10	in fish Evol Appl 2014.7(1).68–87	578
		010
20). Evans SR, Gustafsson L. Climate change upends selection on ornamentation in a	580
	wild bird. Nature Ecol Evol. $2017;1(2):1-5$.	581
21	. Kruuk LEB. Estimating genetic parameters in natural populations using the	582
	"animal model". Philos Trans R Soc B. 2004;359(1446):873–90.	583
22	2. Postma E. Implications of the difference between true and predicted breeding	584
	values for the study of natural selection and micro-evolution. J Evol Biol.	585
	2006;19:309–320.	586
23	B. Hadfield JD, Wilson AJ, Garant D, Sheldon BC, Kruuk LEB. The misuse of	587
	BLUP in ecology and evolution. Am Nat. 2010;175(1):116–25.	588
24	4. Gienapp P, Merilä J. Disentangling plastic and genetic changes in body mass of	589
	Siberian jays. J Evo Bio. 2014; p. 1–10.	590
25	S Bonnet T. Postma F. Fluctuating selection and its (elusive) evolutionary	501
20	consequences in a wild rodent population. J Evol Biol. 2018:31(4):572–586.	591
		592
26	5. Parmesan C, Yohe G. A globally coherent fingerprint of climate change impacts	593
	across natural systems. Nature. 2003;421(6918):37–42.	594

27.	Durant JM, Hjermann DØ, Ottersen G, Stenseth NC. Climate and the match or mismatch between predator requirements and resource availability. Clim Res. 2007;33:271–283.	595 596 597
28.	Visser M, Lambrechts MM. Global climate change leads to mistimed avian reproduction. Adv Ecol Res. 2004;35:89–110.	598 599
29.	Potti J. Advanced breeding dates in relation to recent climate warming in a Mediterranean montane population of Blue Tits Cyanistes caeruleus. J Ornithol. 2009;150(4):893–901.	600 601 602
30.	Saccheri I, Hanski I. Natural selection and population dynamics. Trends Ecol Evol. 2006;21(6):341–347.	603 604
31.	Reed TE, Grøtan V, Jenouvrier S, Sæther BE, Visser ME. Population growth in a wild bird is buffered against phenological mismatch. Science. 2013;340:488–491.	605 606
32.	Sheldon BC, Kruuk LEB, Merila J. Natural Selection and Inheritance of Breeding Time and Clutch Size in the Collared Flycatcher. Evolution. 2003;57(2):406–420.	607 608
33.	Teplitsky C, Mills JA, Yarrall JW, Merilä J. Indirect genetic effects in a sex-limited trait: the case of breeding time in red-billed gulls. J Evol Biol. 2010;23(5):935–944.	609 610 611
34.	Gienapp P, Postma E, Visser ME. Why breeding time has not responded to selection for earlier breeding in a songbird population. Evolution. 2006;60(11):2381–2388.	612 613 614
35.	Lane JE, McAdam AG, McFarlane SE, Williams CT, Humphries MM, Coltman DW, et al. Phenological shifts in North American red squirrels: disentangling the roles of phenotypic plasticity and microevolution. J Evol Biol. 2018;31(6):810–821.	615 616 617
36.	Moyes K, Nussey DH, Clements MN, Guinness FE, Morris A, Morris S, et al. Advancing breeding phenology in response to environmental change in a wild red deer population. Glob Chang Biol. 2011;17:2455–2469.	618 619 620
37.	Stopher KV, Bento AI, Clutton-Brock TH, Pemberton JM, Kruuk LEB. Multiple pathways mediate the effects of climate change on maternal reproductive traits in a red deer population. Ecology. 2014;95(11):3124–3138.	621 622 623

38.	Froy H, Martin J, Walling K, Clutton-Brock TH, Pemberton JM, Kruuk LEB. Consistent within-individual plasticity is sufficient to explain temperature	624 625
	responses in red deer reproductive traits. J Evol Biol. 2019;(X):X.	626
39.	Clements MN, Cluttont-Brock TH, Guinness FE, Pemberton JM, Kruuk LEB. Variances and Covariances of Phenological Traits in a Wild Mammal Population. Evolution. 2010;65(3):788–801.	627 628 629
40.	Coulson T, Kruuk LEB, Tavecchia G, Pemberton JM. Estimating selection on neonatal traits in red deer using elasticity path analysis. Evolution. 2003;57(12):2879–2892.	630 631 632
41.	Henderson CR. Estimation of genetic parameters. Ann Math Stat. 1950;21:309–310.	633 634
42.	Lush J. Animal breeding plans. Ames, Iowa: Iowa State College Press; 1937.	635
43.	Merilä J, Sheldon BC, Kruuk LEB. Explaining stasis : microevolutionary studies in natural populations. Genetica. 2001;112:199–222.	636 637
44.	Brookfield JFY. Why are estimates of the strength and direction of natural selection from wild populations not congruent with observed rates of phenotypic change? BioEssays. 2016;38:1–8.	638 639 640
45.	Lande R. Quantitative Genetic Analysis of Multivariate Evolution , Applied to Brain : Body Size Allometry. Evolution. 1979;33(1):402–416.	641 642
46.	Clutton-Brock TH, Guinness FE, Albon SD. Red deer: behavior and ecology of two sexes. University of Chicago press; 1982.	643 644
47.	Huisman J, Kruuk LEB, Ellis PA, Clutton-brock T, Pemberton JM. Inbreeding depression across the lifespan in a wild mammal population. Proc Natl Acad Sci USA. 2016;113(13):3585–3590.	645 646 647
48.	Huisman J. Pedigree reconstruction from SNP data: parentage assignment, sibship clustering and beyond. Mol Ecol Resour. 2017;17(5):1009–1024.	648 649
49.	Keller L, Waller D. Inbreeding effects in wild populations. T Ecol Evol. 2002;17(5):19–23.	650 651

50.	Pemberton JM. Wild pedigrees: the way forward. Proc Roy Soc B. 2008;275(1635):613–21.	652 653
51.	Hadfield JD. MCMC Methods for Multi-Response Generalized Linear Mixed Models: The MCMCglmm R Package. J Stat Soft. 2010;33(2):1–22.	654 655
52.	Kruuk LEB, Hadfield JD. How to separate genetic and environmental causes of similarity between relatives. J Evol Biol. 2007;20(5):1890–903.	656 657
53.	Morrissey MB, Parker DJ, Korsten P, Pemberton JM, Kruuk LEB, Wilson AJ. The prediction of adaptive evolution: empirical application of the secondary theorem of selection and comparison to the breeder's equation. Evolution. 2012;66(8):2399–2410.	658 659 660 661
54.	Morrissey MB, Bonnet T. Analogues of the fundamental and secondary theorems of selection, assuming a log-normal distribution of expected fitness. J Hered. 2019;in press.	662 663 664
55.	Lande R, Arnold SJ. The Measurement of Selection on Correlated Characters. Evolution. 1983;37(6):1210–1226.	665 666
56.	Hendry AP, Kinnison MT. Perspective: the pace of modern life: measuring rates of contemporary microevolution. Evolution. 1999;53(6):1637–1653.	667 668
57.	Pigeon G, Festa-Bianchet M, Coltman DW, Pelletier F. Intense selective hunting leads to artificial evolution in horn size. Evol Appl. 2016;9(4):521–530. doi:10.1111/eva.12358.	669 670 671
58.	Kruuk LEB, Clutton-Brock TH, Albon SD, Pemberton JM, Guinness FE. Population density affects sex ratio variation in red deer. Nature. 1999;399(6735):459–461.	672 673 674
59.	Bijma P. The quantitative genetics of indirect genetic effects: a selective review of modelling issues. Heredity. 2014;112(1):61–9.	675 676
60.	Morrissey MB, Kruuk LEB, Wilson aJ. The danger of applying the breeder's equation in observational studies of natural populations. J Evol Biol. 2010;23(11):2277–88.	677 678 679

61.	Marrot P, Charmantier A, Blondel J, Garant D. Current spring warming as a	680
	driver of selection on reproductive timing in a wild passerine. J Anim Ecol.	681
	2018;87(3):754-764. doi:10.1111/1365-2656.12794.	682
62.	Coltman DW, O'Donoghue P, Jorgenson JT, Hogg JT, Strobeck C,	683
	Festa-Bianchet M. Undesirable evolutionary consequences of trophy hunting.	684
	Nature. 2003;426(December):655–658. doi:10.1038/nature02187.1.	685
63.	Kardos M, Luikart G, Allendorf FW. Predicting the Evolutionary Effects of	686
	Hunting Requires an Understanding of genetics. J Wildlife Manage.	687
	2018;82(5):889-891.	688

Supplementary information for "The role of selection and evolution in changing parturition date in a red deer population"

689

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¹ 1 Back-transformation of change to days

We used a linear regression of z on years to predict the initial mean $z(\bar{z}_0)$ in 1972 (as-2 suming a linear change) and extracted the residual variance of that regression $(\sigma^2(\epsilon))$. 3 Assuming that the residuals are normally distributed and their variance constant, Bfollows a log-normal distribution conditional on time. The mean of a log-normal distribution is $\exp(\mu + \frac{\sigma^2}{2})$ where μ and σ^2 are the mean and variance on the log-scale. Therefore 6 we predicted the change in mean particition dates as $\Delta \bar{B} = \exp\left(\frac{\bar{z}_t}{100} + \frac{\sigma^2(\epsilon)/10000}{2}\right) - \frac{1}{2}$ 7 $\exp\left(\frac{\bar{z_0}}{100} + \frac{\sigma^2(\epsilon)/10000}{2}\right)$, where $\bar{z_t}$ is a model prediction of z on the log-scale at the end of 8 the study period (year 2016). This back-transformation is imperfect because the relation-9 ship between z and years is not perfectly linear, but it approximately recovers the observed 10 phenotypic change (non-transformed data : -12.32 days 95%CI [-14.53; -10.10] versus 11 back-transformed estimate : -12.11 days 95%CI [-13.77; -10.35]). 12

¹³ 2 Definition of genetic groups

Animal models measure genetic parameters relative to a base population, that is, the set 14 of individuals that have no known parents [1]. In open natural populations, immigrants 15 will have unknown parents and will be included in the base population by default, irre-16 spective of their birth year. This conflation of cohorts within the base population can 17 blur the estimation of genetic change across cohorts. In addition, changes in the number 18 of immigrants might bias the estimation of evolution if immigrants differ genetically, for 19 instance because they come from a population locally adapted to an environment different 20 to that of the focal population [2, 3]. To account for these potential problems, we included 21 genetic groups [4] in our animal models. We modeled genetic groups using the explicit 22 fixed effect specification [3], and considered individuals with unknown parents to form 23 two groups: local base population and immigrants. We defined the two genetic groups 24 with the following rules: 1) The 'local base population' was defined as the 172 individuals 25 with two unknown parents who were born before 1970 (when the intensive monitoring 26 of the study population started), plus the fathers of calves born from a known mother 27 and an unknown father (fathers contribute to genetic group values through the pedigree, 28 although males have no phenotypes for parturition dates); 2) 'Immigrants' were defined 29 as the 594 individuals born after 1970 from two unknown parents, and also unknown 30 mothers of individuals born in any year with an unknown mother and a known father. 31 We considered missing fathers as local because 2/3 of missing fathers are those of calves 32 without a genetic sample, suggesting that the fathers are missing only because they could 33 not possibly be inferred. Moreover, for the other third of calves with missing fathers, 34 but with a genetic sample, the lack of paternity assignment must be due to the father 35 lacking a genetic sample. We might therefore expect only $\frac{1}{3}\frac{1}{3} = \frac{1}{9}$ of calves without known 36 fathers to be born from fathers who are truly immigrants. On the other hand, the lack of 37 a maternal assignment (i.e. an unknown mother) almost certainly indicates an individual 38 of immigrant origin. 39

3 Selection among individuals culled or not culled



Figure S3.1: Posterior probability densities for selection differential estimated from the same model fitted to three datasets: (i) total population which includes culled individuals, (ii) total population excluding culled individuals, (iii) culled individuals only. Vertical lines highlight posterior modes.

41 4 Supplementary tables

42 4.1 Animal model estimates

Table S4.1: Fixed effects from the univariate animal model of log-transformed parturition date (eq. 1).

		Estimate	95% CI
Intercept*		406.82	[390.79; 420.52]
Female's inbreeding coefficient		89.39	[-7.22; 196.58]
Offspring Sex: Male		1.09	[-1.05; 3.20]
Female's Reproductive Status:	Naive	-13.53	[-19.26; -7.65]
	Summer Yeld	-27.01	[-31.46; -22.00]
	True Yeld	-21.56	[-25.43; -17.82]
	Winter Yeld	-0.60	[-6.19; 4.19]
Female's age		-6.64	[-9.20; -3.76]
Female's age squared		0.38	[0.24; 0.51]
Genetic group: Immigrant		4.38	[-6.97; 16.48]
Calf birth year		-0.47	[-0.80; -0.11]

Notes: *The intercept is defined for the year 1972.

Table S4.2: Random intercept variance components from the univariate animal model of log-transformed parturition date (eq. 1).

	Estimate	95% CI
Additive genetic	208.3	[140.8; 282.9]
Permanent environment	5.9	[0; 40.6]
Focal female's mother	1.9	[0; 10.5]
Offspring birth year	112.3	[59.0; 189.1]
Female cohort	1.9	[0.0 ; 9.7]
Residuals	994.4	[931.4; 1056.0]

⁴³ 4.2 Values for parameters in Figure 4

Table S4.3: Posterior modes and 95% credibility intervals for the parameters presented in Fig. 4. All parameters relate to the estimated or predicted rate of evolution over the study period and are expressed in number of days.

Model	Parameter	Mode	95%CI
Univariate	Evolution (BLUPs)	-2.1	[-4.5; 0.7]
Univariate	Drift	0.00	[-2.998; 3.219]
Univariate	Breeder's equation	-1.37	[-3.01; -0.60]
Bivariate	Breeder's equation	-1.31	[-2.46; 0.10]

44 5 Estimate of evolution on untransformed data

We re-fitted the univariate animal model (equation 1) to the raw data of parturition date and re-estimated heritability and the rate of evolution based on BLUPs regression. Heritability was estimated to 0.10 [0.06; 0.13]. The change in breeding values is estimated to -1.7 [-4.0; 0.7] using the most conservative method and to -2.3 days [-5.7; 0.4] using the less conservative method. 693

50 References

- [1] Kruuk LEB. Estimating genetic parameters in natural populations using the "animal model". Philos Trans R Soc B. 2004;359(1446):873–90.
- [2] Hadfield JD, Wilson AJ, Garant D, Sheldon BC, Kruuk LEB. The misuse of BLUP
 in ecology and evolution. Am Nat. 2010;175(1):116–25.
- ⁵⁵ [3] Wolak ME, Reid JM. Accounting for genetic differences among unknown parents
 ⁵⁶ in microevolutionary studies: how to include genetic groups in quantitative genetic
 ⁵⁷ animal models. J Anim Ecol. 2017;86(1):7–20.
- ⁵⁸ [4] Quaas RL. Additive Genetic Model with Groups and Relationships. J Dairy Sci.
 ⁵⁹ 1988;71(5):1338-1345.