

# The role of selection and evolution in changing parturition date in a red deer population

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## Abstract

Changing environmental conditions cause changes in the distributions of phenotypic 1  
traits in natural populations. However, determining the mechanisms responsible for 2  
these changes and, in particular, the relative contributions of phenotypic plasticity vs 3  
evolutionary responses, is difficult. To date, to our knowledge no study has reported 4  
evidence that evolutionary change underlies the most widely-reported phenotypic 5  
response to climate change: the advancement of breeding times. In a wild population of 6  
red deer, average parturition date has advanced by nearly two weeks in four decades, 7  
and within-individual phenotypic plasticity in response to warming temperatures 8  
explains a minor part of this advance. Here we show that parturition date is also 9  
heritable and under selection towards earlier dates, and that genetic changes likely also 10  
played a role in the shift towards earlier parturition dates. The observed rate of 11  
evolution matched the predicted response to selection, and was less likely to be due to 12  
genetic drift. Our study provides a rare example of observed rates of genetic change 13  
matching theoretical predictions, although the match would not have been detected 14  
with a solely phenotypic analysis. It also provides, to our knowledge, the first evidence 15  
of both evolution and phenotypic plasticity contributing to advances in phenology in a 16

## Introduction

Climate change affects various aspects of biodiversity across the planet (e.g., [1,2]). In particular, shifts in phenotypic distributions within populations are widely reported, for a variety of morphological, phenological or life-history traits [2–4]. Surprisingly, however, little is still known about the relative contributions of mechanisms underlying these shifts [5]. Within a population, phenotypic distributions may change due to a change in population structure (e.g., age-structure or sex-ratio), due to phenotypic plasticity (within or between individuals), and due to genetic change [6–8]. The exact mixture of mechanisms driving phenotypic change will determine the future of a population facing a prolonged change in environmental conditions [9], for several reasons. First, the consequences of changing population structure are variable and may be idiosyncratic (e.g., [8,10]). Second, phenotypic plasticity can provide an efficient way to cope with a changing environment but its effect may be short-lived and even maladaptive [11–13]. Third, genetic evolution, when driven by natural selection, can improve population growth rate, potentially contributing to long-term population persistence [12].

In wild populations the respective contributions of plasticity vs evolution remain unknown for the vast majority of documented phenotypic changes [14,15] (note that by evolution we mean genetic change, here and in the rest of the manuscript). To date, most of the evidence for evolutionary responses to climate change comes from plants [16]. In contrast, despite numerous examples of phenotypic changes apparently related to climate, there have been surprisingly few examples demonstrating unambiguously that a vertebrate population is evolving in response to climate change (see discussions in [17–20]). This lack of evidence may in part be due to the question not being prioritized [14,15]. However it probably also reflects the substantial challenges inherent in testing for adaptive evolution, in terms of requirements for appropriate data and statistical methods. For wild populations in which experimental manipulations are not feasible, the most plausible means of testing for the genetic basis of phenotypic changes is to use long-term pedigree data to test for changes in ‘breeding values’, the estimated genetic merit of individuals as ascertained from the phenotypes of their relatives [21].

This needs to be done with care, as trends in predicted breeding values can be confounded with environmental trends unless appropriately controlled for [22], and precision of estimates of evolutionary rates can be inflated if the correlation structure of breeding value estimates is not properly handled [23]. To date, three recent studies of wild vertebrate populations using methods that account for uncertainty in breeding value predictions have found evidence of a genetic change underlying phenotypic change in morphology, all in line with selection pressures changing with climate: plumage colouration in collared flycatchers [20], and body size in Siberian Jays [24] and in snow voles [25]. However only with more empirical studies explicitly testing for evolution will it become possible to say whether the current lack of evidence also reflects a general slow rate of adaptation to environmental change in natural populations.

Climate change may have impacts on numerous aspects of an organisms' biology, but phenology (i.e., the seasonal timing of life-history events) appears to have been particularly affected [3, 26, 27]. Dramatic changes of phenologies in response to earlier onset of spring are particularly well documented in mid- and high-latitude passerines, where breeding times are occurring earlier in numerous populations and species [18, 28]. The study of avian systems in particular has shown that a fine tuning of phenology to the climate is crucial in determining individual fitness. Mismatches between mean breeding date and a fitness optimum that shifts with climate may re-shape selective pressures and hence potentially reduce population growth rate [29], although establishing the link between individual-level and population-level processes is not trivial [30, 31]. The effects of climate change on mammalian phenology are less well documented than those of birds, and may be even more complex because mammals' long gestation times may make their breeding phenology sensitive to climate across a longer time-frame [17]. Finally, despite the extensive evidence for phenotypic shifts in phenology, the few studies that test for a genetic basis to changes in phenology in wild populations have not found evidence of changes in breeding values [32–35].

In a population of red deer (*Cervus elaphus*, Linnaeus 1758) on the Isle of Rum, NW Scotland, parturition date has advanced at a rate of 4.2 days per decade since 1980, a change that has been linked to temperatures and other weather conditions in the year preceding parturition, around the time of conception [36, 37]. Previous studies of this population have shown that phenotypic plasticity and population structure explain a

substantial proportion (23%) of the advance in parturition dates [37], and also that within-individual plasticity is sufficient to explain the relationship between temperature and parturition date [38]. However, the documented plasticity does not explain the majority of the observed phenotypic change, leaving room for processes that have not been investigated as of yet. It is plausible that evolution plays a role because the observed phenotypic change is qualitatively consistent with a genetic response to selection: parturition date is heritable in this population [39] and also under selection for earlier dates [40].

In this study we use quantitative genetic animal models [21,41] to estimate the rate of evolution in parturition date and the contribution of plastic and demographic processes to the observed shift in phenology in the Rum red deer study population. We start by considering the response to selection that might be expected from the observed strength of selection and heritability of parturition date, based on a simple “breeder’s equation” prediction [42]. One of the most striking conclusions from the recent application of quantitative genetic theory in evolutionary ecology has been the failure of univariate breeder’s equation predictions to capture trait dynamics in wild populations [43,44]. This may be for multiple reasons, foremost of which is likely to be the unrealistic assumption that only the focal trait is relevant. We therefore also consider a multivariate breeder’s equation [45], and ask how selection on offspring size and the genetic correlation between parturition date and size alters the expected evolutionary response. However a second, less well-explored reason for the failure of the theory is the comparison of predicted response to observed rates of phenotypic change, which will obviously be affected by other processes, rather than with estimates of the underlying rates of genetic change. As the central analysis of this work, we use trends in breeding values to estimate the rate of evolution in parturition date and to test whether it is compatible with the response to selection predicted by either the univariate or multivariate breeder’s equation, or with genetic drift. Finally we quantify the effect of non-genetic processes contributing to phenotypic change along with evolution.

# Methods

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## Study population

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We used data from a long-term study of the unmanaged population of red deer living in the North Block of the Isle of Rum, Scotland (57°01' N, 6°17' W), for the years 1972-2016. The work takes place under a UK Home Office Project Licence under the Animals (Scientific Procedures) Act 1986 as amended. Licence no 70/8818 held by J.M. Pemberton. The project takes place with the permission of Scottish Natural Heritage which manages the Isle of Rum National Nature Reserve. Within the ca. 12 Km<sup>2</sup> of the study area, calves are marked with ear tags (and a collar for females) shortly after birth, in order to record detailed life-histories of individuals throughout their lives [46]. DNA was obtained from ear punches, post-mortem tissue and cast antlers. The population pedigree was reconstructed from single nucleotide polymorphisms as in [47], using the R package SEQUOIA [48].

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We studied the selection and genetics of parturition date, the date on which a female gave birth to a calf in a given year. We therefore focus on females, because males do not express the trait of parturition date —though they may affect it, in both genetic and non-genetic ways. Males were retained in the pedigree and contributed to the calculation of quantitative genetic parameters by informing the relatedness between individuals. Selection was estimated from the association between a trait and individual lifetime breeding success, where lifetime breeding success was the number of offspring produced by an individual across their lifetime, whether or not they survived to breeding and therefore also had parturition records (further details below). Parturition date being a sex-limited trait, selection differentials on parturition date were divided by 2 after estimation (i.e., half the population was assumed not to be selected for that trait). We included females that are still alive, even though their lifetime fitness is still unknown, in order not to introduce a fraction of individuals missing not at random with respect to fitness and parturition date. However, excluding living females from the analysis gave indistinguishable results.

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There has been no culling in the study area since 1973, but individuals are occasionally shot when they visit areas surrounding the study area. Mortality due to culling may exert a kind of artificial selection that studies of natural selection may want

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to exclude. However, our goal here was to understand the causes of phenotypic change, be they natural or artificial. We therefore retained culled females in our main analyses. These shot females represented a small but non-trivial portion (15%) of the data set (Table 1). Therefore, in a sub-analysis, we also considered selection only among females who died of natural causes (i.e., excluding both shot females and females still living), and discuss the consequences of culling for selection and evolution in this system.

Parturition date was measured as the number of days after May 1, because virtually no parturition occurs before that date. Values were (natural-)log-transformed in order to obtain residuals with distributions close to Gaussian. The logged values were multiplied by 100 for reporting convenience (in particular, variance components would have been of the order  $10^{-5}$  without this second step). The working phenotype in all models was therefore  $z = 100\log(B)$ , where  $B$  is the parturition date in number of days after May 1<sup>st</sup>. Results were converted back to days (see SI 1 for details of the back-transformation process) to facilitate biological understanding. We report results using untransformed data in SI 5.

Data type	Number of	Excluding shot	Shot	Total
Parturition date	Individuals	582	158	740
	Records	2921	463	3384
Lifetime breeding success	Individuals	1614	282	1896

**Table 1.** Sample sizes for lifetime breeding success (LBS) and parturition data for years 1972-2016. All numbers refer to females; parturition date is measured repeatedly on individuals. LBS is measured over a lifetime, and there is only one measure per individual. All females with a unique ID have an LBS record, including those that died as calves and therefore did not breed and do not have records for parturition date.

## Quantitative genetic analysis

### Univariate animal model

We fitted a univariate animal model of (transformed) female parturition date in order to estimate heritability and change in breeding values [21, 41]. The fixed effects in the model were: the sex of the offspring; the female’s ‘reproductive status’, which can take five values to represent different recent reproductive history: ‘naive’, ‘true yeld’, ‘summer yeld’, ‘winter yeld’, ‘milk hind’ [38]; the female’s age in years (first and second order polynomial); a contribution of ‘genetic group’, to model gene flow into the

population, estimated as her expected level of immigrant vs resident genes (see SI 2);  
the calf's birth year as continuous variables, see next section for details; and the  
female's pedigree-based inbreeding coefficient [49, 50] calculated using the R package  
MCMCglmm [51]. Population density, estimated as the number of resident adult  
females in a given year, had a significant effect on (log) parturition date at the  
beginning of the study period (e.g., between 1974 and 1987, [46]), but we found no  
effect in the full data-set (slope  $-0.38$ , standard error  $0.63$ ) and so did not include  
density in the final models.

The random effects decomposed the variance not accounted for by fixed effects into  
six components: additive genetic variance; 'permanent environment' variance (estimable  
from the repeated measures of the same females across multiple years; [52]); maternal  
effects variance (i.e. associated with the mother of the breeding female, and hence  
grand-mother of the new calf); variance associated with the calf's birth year; variance  
associated with the breeding female's (i.e. mother of the calf) birth year; and residual  
variance.

Thus, the model of (log-transformed) parturition date ( $z$ ) of female  $i$  in year  $j$  can  
be written

$$z_{ij} = \mu + \mathbf{X}^T \mathbf{b} + a_i + p_i + m_i + c_j + y_i + r_{ij} \quad (1)$$

where  $\mu$  is an intercept,  $\mathbf{X}$  is a vector of fixed predictors (including calf's birth year),  $\mathbf{b}$   
is a vector of fixed effects,  $a$ ,  $p$ ,  $m$ ,  $c$  and  $y$  are random effects with which to estimate  
the variance associated with additive genetic values (i.e., breeding values), permanent  
environment, maternal (i.e. grand-mother of calf), calf year of birth, and female's year  
of birth, respectively, and  $r$  is the residual. The breeding values ( $a$ ) are normally  
distributed as  $(a_1, \dots, a_n)^T \sim N(0, \sigma_A^2(z) \mathbf{A})$ , where  $\sigma_A^2(z)$  is the additive genetic  
variance for (log) parturition date,  $n$  is the number of females, and  $\mathbf{A}$  is the relatedness  
matrix between individuals. The heritability of (log) parturition date was estimated as  
 $\sigma_A^2(z)$  divided by the sum of all the variance components.

We used this animal model to estimate the individual-level repeatability (in addition  
to the heritability) of parturition date, as the sum of the proportions of variance  
explained by all effects that are constant for an individual: inbreeding, female's cohort  
and genetic group (all of which are fixed effects) and additive genetic variance,

permanent environment variance, maternal variance and female’s cohort variance (that is, all random effects but offspring birth year and residual).

We ran all models in MCMCglmm [51] with Gaussian errors for (log-transformed) parturition date. We report posterior modes and 95% highest posterior density credible intervals. For this univariate model, we used the default inverse gamma priors for variance components, with shape and rate parameters both equal to 0.001 (equivalent to a variance and degree of belief of 1 and 0.002, respectively). We run models for 130000 Markov chain Monte Carlo iterations, with a burn-in of 30000 and thinning of 100.

## Selection

We estimated selection acting on parturition date by assessing the association between a female’s parturition dates and her fitness. We measured fitness as *lifetime breeding success* (LBS in the text,  $W$  in equations), which is the number of offspring produced by an individual, calculated for all females in the database, whether or not they survived to breeding and therefore also had parturition records.

Selection was estimated using a model of the covariance between parturition date and fitness. We used a bivariate generalized linear mixed model, with LBS modelled as an over-dispersed Poisson trait (with log link function) and log-transformed parturition date ( $z$ ) modelled as a Gaussian trait. This model can be written as

$$[z, W] \sim \mathbf{X}\mathbf{b} + \mathbf{D}_1\mathbf{m} + \mathbf{D}_2\mathbf{y} + \mathbf{D}_3\mathbf{c} + \mathbf{D}_4\mathbf{p} + \mathbf{I}\mathbf{r} \quad (2)$$

where  $\mathbf{X}\mathbf{b}$  represents fixed effects (the same fixed effects for parturition date as above, and only an intercept and genetic group for fitness),  $\mathbf{m}$ ,  $\mathbf{y}$ ,  $\mathbf{c}$ ,  $\mathbf{p}$  are random effects associated with maternal effects (the identity of the mother of the breeding female), the year of calving, the female’s cohort (i.e. her year of birth), and the individual female’s identity (or ‘permanent environment’ effect, because of the repeated measures), respectively.  $\mathbf{D}$ -matrices link random effect levels to observations, and  $\mathbf{I}\mathbf{r}$  represents the residuals.

Note that  $W$  is only measured once for each individual, unlike the repeated measures on parturition date ( $z$ ). For  $W$ , variance components are therefore null for  $\mathbf{y}$  (the calf’s year of birth) and  $\mathbf{p}$  (the permanent environment component of a trait, derived from



repeated measures). `MCMCglmm` accommodates the difference in replication between the two traits by allowing the individual-level random effect  $\mathbf{p}$  for the replicated trait (parturition date) to covary with the residual variance  $\mathbf{r}$  of the non-replicated trait (fitness), thus providing a covariance between the repeatable part of an individual's parturition date and her fitness (for a comparable example, see [53]).

The selection differential on parturition date was calculated as the sum of this individual-level covariance, plus the maternal-effect covariance between parturition date and fitness (i.e. covariance among effects of the breeding females' mothers on their daughters' parturition dates and fitness). Selection differentials characterize the within-generation change in phenotype due to selection. We therefore standardized this value by generation time (8 years) to be expressed in rate of change per year, or in total change over the study period. As stated above, the selection differentials were also divided by two, because the covariances were estimated from females only. Males do not express the trait but nevertheless carry genes relevant to parturition date in females. Selection on parturition acts on only half of the population, and the expected response to selection is half that predicted from the strength of selection in females. We also estimated a selection gradient [55], calculated as the selection differential divided by the corresponding variance (that is, the sum of the individual-level and mother-level variance components for parturition date).

When expected fitness follows a log-normal distribution, selection parameters can be equivalently calculated on the scale of the data using relative fitness, or on the log-scale using absolute fitness [54]. Because our model uses a log-link function for absolute LBS, its parameter estimates are on the latent scale, but these are directly interpretable as selection differentials and selection gradients relating to relative fitness on the data scale.

For multivariate models we used parameter-expanded priors for variance components (with working mean of 0 and variance of 1000). We ran these models for 260000 Markov chain Monte Carlo iterations, with a burn-in of 60000 and thinning interval of 200.

### **Univariate and multivariate predictions of evolution**

The response to selection (the per-generation change in the mean value of the trait) was predicted as the product of the heritability in parturition date and the selection differential, following the univariate breeder's equation [42]. The equation was applied

to estimates from the model of log-transformed parturition date data, and the predicted response was subsequently back transformed to days. Calculations were done on the MCMC posterior distributions of the heritability and the selection differential, in order to propagate the uncertainty in these two parameters. The univariate breeder's equation ignores the fact that the adaptive evolution of a focal trait depends not only on direct selection on that trait, but also on selection on those traits that are genetically correlated with the focal trait [45]. This assumption may explain in part the common mismatch between predicted and observed evolution in natural populations [44], but it can partly be relaxed by incorporating analyses of relevant associated traits and estimating multivariate selection and genetic covariances: the multivariate response to selection can then be predicted as the product of the genetic variance-covariance matrix  $\mathbf{G}$  and the vector of multivariate selection gradients  $\boldsymbol{\beta}$  ( $\Delta\mathbf{Z} = \mathbf{G}\boldsymbol{\beta}$ ) [45, 55].

In the Rum red deer study population, a calf's birth date is correlated with its birth weight [37, 46], a trait also under selection [40]. We therefore applied a bivariate breeder's equation to parturition date and calf birth weight to estimate the effect of indirect selection on the predicted evolutionary response of parturition date to selection.

We extended the animal model of parturition date (eq. 1) to a bivariate animal model of parturition date and offspring birth weight, using the same fixed effects and random effects for both traits. Note that in this model, the calf's birth date and birth weight ( $bw$ ) are both being treated as the phenotype of the mother; the treatment of offspring birth weight as a trait of the mother is justified by the observation that more than 90% of the genetic variance in birth weight is maternal-genetic variance rather than direct-genetic variance [52]. This model estimated an additive genetic covariance between the two traits,  $\sigma_A(z, bw)$ , which can be divided by the square root of the product of the two additive genetic variances,  $\sigma_A^2(z)$  and  $\sigma_A^2(bw)$ , to obtain a genetic correlation.

Finally, we extended the bivariate selection model (eq. 2) to a trivariate model also including offspring birth weight (along with parturition date and LBS). For birth weight we used the same fixed and random effects as described above for parturition date. We summed the appropriate covariances of the phenotypes with LBS to obtain a vector of selection differentials  $\mathbf{s}$ . We summed the appropriate variances and covariances for the two phenotypes to obtain a phenotypic variance-covariance matrix  $\mathbf{P}$ . We then applied

$\mathbf{P}^{-1}\mathbf{s}$  to obtain  $\beta_z$ , the direct selection gradient on parturition date corrected for the indirect selection on birth weight, and  $\beta_{bw}$ , the direct selection gradient on birth weight corrected for the indirect selection on parturition date. The response to selection could then be calculated as  $\beta_z\sigma_A^2(z) + \beta_{bw}\sigma_A(z, bw)$  [55].

We also expressed predicted rates of evolution in Haldanes, that is, in units of standard deviation per generation [56]. We did not express the results in Darwins because parturition dates have no natural zero point, and therefore mean-standardisation is not meaningful.

## Components of change

### Trend in breeding values

Using the univariate animal model of parturition date containing year as a covariate (see below), we fitted a linear regression of best linear unbiased predictors (i.e., model predictions for the values of a random effect levels, BLUPs hereafter) for individual females' breeding values against the mean birth year of their offspring to each posterior sample. This generates a posterior distribution for the slope of change in mean breeding value [23]. In addition, to visualize potential non-linearity in genetic change, we fitted a smoothing spline function of female cohort year to the BLUPs for individual breeding values for every posterior sample, thus generating the posterior distribution of the time-dynamic of breeding values among cohorts [57]. Changes in breeding values may indicate a response to directional selection, but they can also be produced by random fluctuations under non-directional evolutionary models, such as genetic drift. To assess this possibility, we also compared the posterior distribution of the estimated change in breeding values to the change possible under genetic drift alone, using simulations as described in [23].

In general, breeding values predicted by animal model BLUPs are not equal to the true breeding values, but are influenced by environmental random deviations [22]. As a consequence, a linear regression of BLUPs may confound genetic and non-genetic (e.g. plastic) change and may produce a biased estimate of evolution. This issue can be addressed by including year as a covariate in the animal model used to obtain BLUPs. Unfortunately, the solution is conservative, because the animal model ascribes some of

the genetic change to the year effect [22]. We opted to report primarily conservative estimates of evolution, based on an animal model that did contain calf cohort as a covariate (see above). Nevertheless, we also re-fitted the animal model without a fixed effect for cohort and re-calculated the change in BLUPs for breeding values estimated this way; we report this second estimate in the discussion.

### Other contributions to phenotypic change

Finally we estimated the contributions of several other terms in equation (1) to the trend in parturition date. We used Geber’s method [6, 8] on model predictions to estimate the independent contribution of changes in the class structure of age and reproductive status, and the independent contribution of changes in levels of inbreeding (as assessed from the pedigree inbreeding coefficient) and gene flow (as assessed by the genetic groups effect) to the phenotypic change in parturition date. Briefly, this method estimates the contribution of change in a parameter mean ( $\bar{k}$ ) to change in a trait mean ( $\bar{z}$ ) as the product of the partial derivative of  $z$  on  $k$  ( $\frac{\partial \bar{z}}{\partial \bar{k}}$ ) and the slope of  $k$  on time ( $\frac{\Delta \bar{k}}{\Delta t}$ ). We applied the equation to each sample of the posterior distributions in order to propagate the uncertainty in the estimated trends. In addition to calculating the net effect through the study period, we calculated  $\frac{\partial \bar{z}}{\partial \bar{k}} \bar{k}_t$  for each year  $t$  to visualize the dynamic of changes in effects through time graphically.

We did not use random effects to estimate non-genetic components of change because random effects other than additive genetic effects are linearly independent of years by construction, and any change in females’ maternal effects or permanent environment should be absorbed into the fixed effect of offspring cohort.

## Results

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### Phenotypic change

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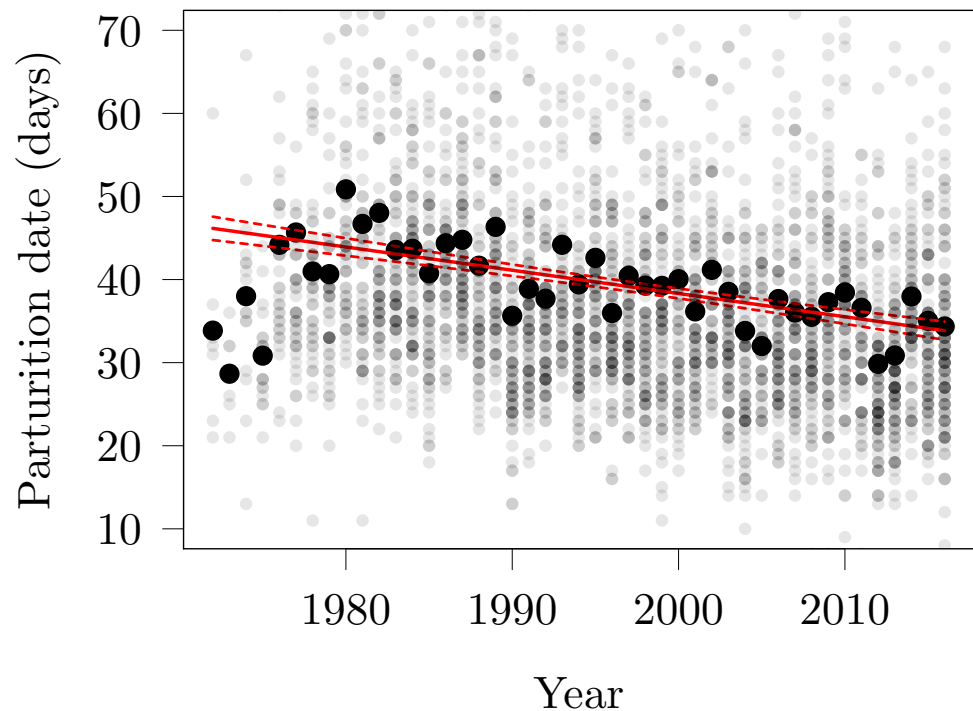
Average parturition dates became later from 1972 to 1980 (probably reflecting increased population density, [46]), after which they advanced at an apparently constant rate (Fig. 1). A linear regression estimates the change in parturition date to be a total of  $-12.3$  (95%CI [14.6;10.1]) days over the 45-year study period (from 1972 to 2016).

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**Fig 1.** Phenotypic trend in red deer parturition dates, in days after May 1st. Large black dots represent annual means, small grey dots represent individual parturition dates, with the darker shades indicating more calves being born on a given day. About 4% of individual parturition dates fall outside the plotted region (10th May - 12 July; note these are still included in the analyses). The red lines represent the slope and associated 95% confidence interval of a linear regression of all individual parturition dates on year of parturition. Note that the years 1972-1975 have very negative residuals and that the rate of change over 1980-2016 is slightly underestimated by the linear regression being fitted over all years.

### Sources of parturition date variation

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Parturition date was influenced by a female's reproductive status and age, but there was no clear evidence for effects of inbreeding, of offspring sex or of the proportion of

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immigrant genetic ancestry (SI Table 4.1). Parturition date was heritable, with additive genetic variance accounting for 18% (95%CI [10%; 23%]) of phenotypic variation corrected for fixed effects and variation among years and among cohorts (SI Table 4.2). The individual-level repeatability of parturition date (estimated as the sum of proportions of all variance components except offspring birth year and residual) was 19%, of which additive genetic variance was most important, with permanent environment effects and maternal effects both accounting for less than 1% of total phenotypic variation (SI Table 4.2). The random effect for offspring birth year (which captures the variance corrected for the temporal trend) accounted for about 8% of the phenotypic variance (SI Table 4.2). Note that proportions are essentially invariant under monotonic transformation and that these proportions of variances are equivalent on the transformed and on the data scale.

## Univariate selection and predicted response

Females with earlier parturition dates had, on average, higher lifetime breeding success: the selection differential of parturition dates estimated with LBS was  $-9.08$  days (95%CI  $[-14.91; -3.81]$ ). Given the heritability of parturition date of 0.16, the univariate breeder's equation predicts a total response to selection of  $-1.37$  days over the 45-year study period (95%CI  $[-3.01; -0.60]$ ) (Fig. 4A). This corresponds to  $-0.031$   $[-0.068; -0.014]$  days per year,  $-0.25$   $[-0.55; -0.11]$  days per generation, or  $-0.019$   $[-0.042; -0.008]$  Haldanes.

Selection was stronger among females died of natural causes than among females that were culled (SI Fig.S3). Using the subset of females who died of natural causes, the univariate breeder's equation predicts a response of  $-2.04$   $[-3.37; -0.95]$  days over the study period. In contrast, using the subset of females who were culled, the univariate breeder's equation predicts a response of  $0.11$   $[-0.64; 0.93]$  days over the study period.

## Bivariate selection and predicted response

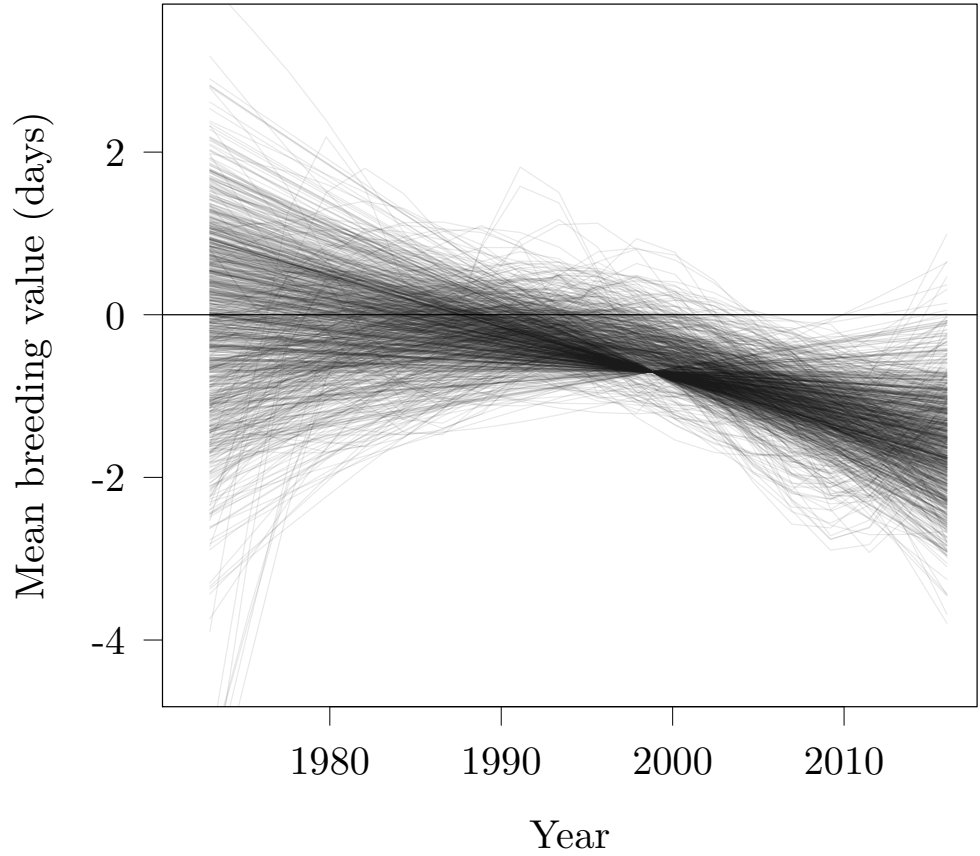
Conditional on the fixed effects affecting each trait, the phenotypic correlation between parturition date and birth weight was positive but weak (correlation = 0.12, 95 %CI  $[0.05; 0.16]$ ). The gradient of direct selection on parturition date was negative (mode

$\beta_z = -0.0003$ , 95 %CI  $[-0.0004; -0.0002]$ ), and that on birth weight was positive ( $\beta_{bw} = 0.0138$ , 95%CI $[0.009; 0.017]$ ). There was also additive genetic variance in offspring birth weight (0.68, 95%CI  $[0.57; 0.90]$ ), corresponding to a heritability of 0.46 (95%CI  $[0.37; 0.62]$ ). The additive genetic covariance between parturition date and offspring birth weight was  $-1.78$  (95%CI  $[-4.38; 0.56]$ ), corresponding to a weak negative genetic correlation of  $-0.16$  (95%CI  $[-0.32; 0.05]$ ). The multivariate breeder's equation predicts a rate of evolution of  $-1.31$  days (95%CI  $[-2.46; 0.10]$ ) over the study period, which is actually very similar to the univariate breeder's equation prediction of  $-1.37$  days (difference =  $-0.15$  days, 95%CI  $[-1.59; 1.03]$ , Fig. 4B)

## Genetic contribution to phenotypic change

The slopes of the linear regressions of BLUPs for parturition date breeding values on birth year, integrated over the posterior distribution, suggests an advance in breeding values, with the slope estimated at  $-0.10$ , 95%  $[-0.23; 0.03]$  per year on the log-transformed scale. Time-splines fitted on the posterior distribution of the BLUPs visually support a linear decrease in breeding values (Fig. 2). This rate of evolution corresponds to a total change over the study period of  $-2.1$  days, 95%CI  $[-4.5; 0.7]$  due to genetic change (Fig. 2 and 4C). This is equivalent to  $-0.045$   $[-0.100; 0.018]$  days per year,  $-0.36$   $[-0.79; 0.14]$  days per generation, or  $-0.028$   $[-0.062; 0.01]$  Haldanes.

9% of the simulations of genetic drift generated an advance as large or larger than the change estimated from the BLUP linear regression (using the posterior mode for the BLUPs trend as a point of comparison, see Fig. 4D). Inbreeding tended to delay parturition date (SI 4.1), and given that the estimated pedigree inbreeding inevitably increased over time with increasing pedigree depth [49], there was marginal evidence from our model of inbreeding postponing parturition date by 0.38 days (95%CI  $[-0.04; 1.01]$ ) over the study period, thus opposing the phenotypic trend. However this prediction may be spurious, because the increase in inbreeding coefficient was an artifact of estimating inbreeding from a pedigree [49]. Re-running the model without inbreeding led to almost identical estimates for all other parameters. The effect of gene flow (proportion of immigrant genotype) was very uncertain (SI 4.1) and its overall predicted effect over the study period was a change of 0.15 days (95%CI  $[-0.34; 0.72]$ ).

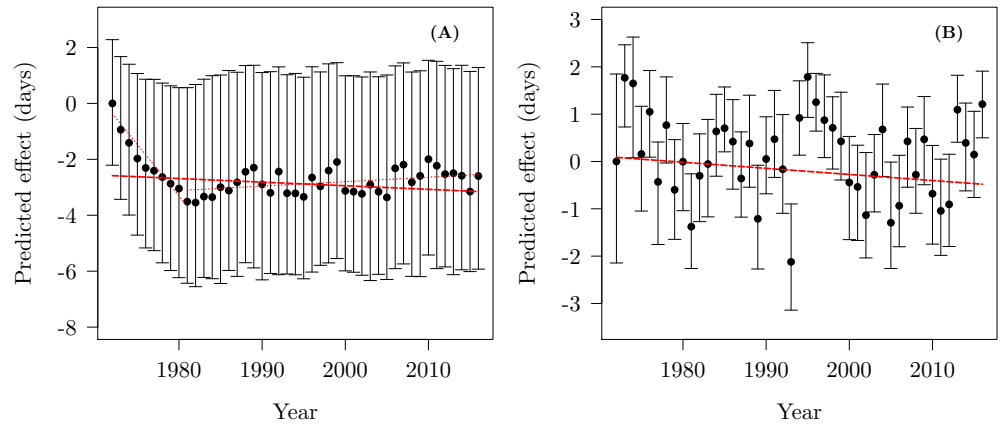


**Fig 2.** Trend in breeding values for parturition date. Each black line was obtained from a different MCMC posterior sample, by fitting a spline to the mean of estimated breeding values among individuals living in the same year. The y-axis was centered on the mean breeding values in 1972 to help interpretation.

### Non-genetic contributions to phenotypic change

As in previous work [37], we found that mature females tended to give birth earlier than younger females, but very old females gave birth the latest. The effects on mean parturition dates of changes in the age structure tended to be in the opposite direction to the observed phenotypic change: during the first ten years of the study, the mean age of females in the study increased steadily, pushing towards earlier mean parturition dates ( $-3.68$  days, 95%CI  $[-5.63; -1.92]$  from 1972 to 1981). For the rest of the study, the change in age structure tended to delay mean parturition date slightly ( $0.57$  days  $[0.39; 0.71]$  from 1982 to 2016). Over the study period the change in age structure had a predicted net effect of  $-0.58$  days  $[-1.67; 0.40]$  (Fig. 3A). Changes in female reproductive status had a fluctuating effect on parturition date (Fig. 3B), with a



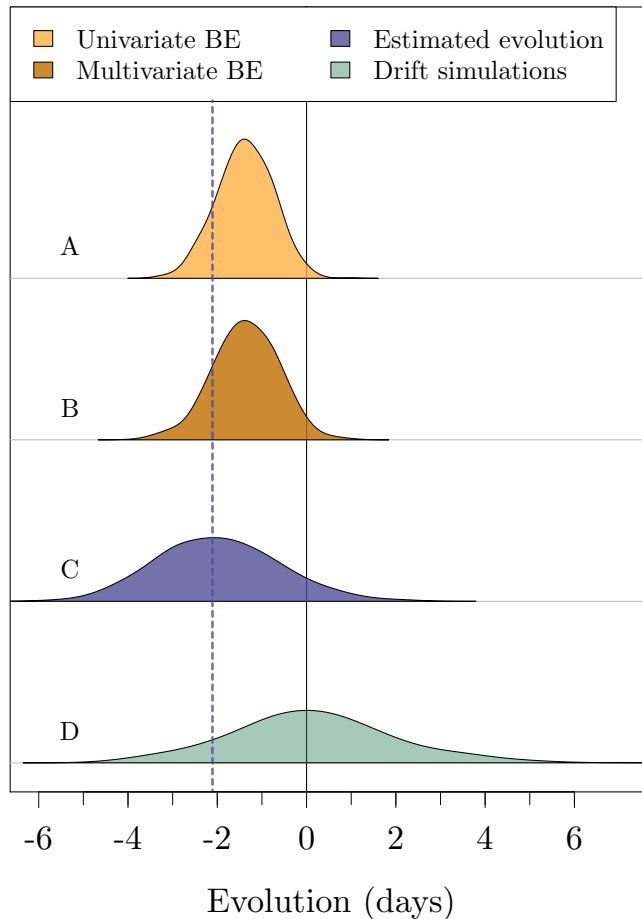


**Fig 3.** Predicted effect of (A) age-structure and (B) female reproductive status on parturition date across years. The origin of the y-axis is arbitrarily set to the predicted effect in the first year. The red thick dashed lines represent the net effect of changes in age structure and female reproductive status on parturition date reported in the text. The thin dotted lines in (A) represent the effect of changes in age structure before, and after 1981, respectively.

resulting total effect over the study period of  $-0.32$  days (95%CI  $[-0.87; 0.17]$ ). 413

Offspring sex had no clear effect on parturition date, and since sex-ratio at birth 414  
remained stable over the study period (despite an early decline in the proportion of 415  
males [58]), this parameter is predicted to have had no noticeable effect ( $-0.04$  days, 416  
95%CI  $[-0.18; 0.04]$ ). 417

Finally, the fixed covariate of calf birth year (SI Table 4.1) captured trends across 418  
years that should ideally be unrelated to genetic change, age, and reproductive status, 419  
although it will inevitably be partly confounded with genetic change (see Discussion). It 420  
may also, similarly, capture persistent changes in maternal effects, permanent 421  
environment effects, and various plastic processes. The coefficient (back-transformed) 422  
corresponds to a change of  $-9.3$  days (95%CI  $[-11.9; -2.22]$ ) over the study period, 423  
showing that most of the observed change remains unexplained by our model. However, 424  
note that some effects (e.g., inbreeding and gene flow) opposed the phenotypic change, 425  
and that the model explained more than the difference between the birth year effect and 426  
the total phenotypic change. 427



**Fig 4.** Posterior distributions of the parameter estimates for change in parturition date over the study period: (A), (B) the predicted evolutionary response to selection from the univariate and bivariate breeder’s equations respectively; (C) the estimated contribution of evolution (from the trend in predicted breeding values); and (D) evolutionary change possible due to genetic drift only. The distributions all have the same area. The dashed line indicates the mode of the distribution (C), the contribution of evolution. The response to selection was estimated using univariate and bivariate breeder’s equations, where phenotypic multivariate models gave selection gradients, and animal models gave additive genetic variance-covariances of parturition date and birth weight. A univariate animal model was used to estimate the amount of evolution as the temporal trend in BLUPs for breeding values, and to simulate evolution by genetic drift. Parturition date was modeled using a log-transformation, and all estimates were subsequently converted to change in days over the study period (see SI 1). Parameter estimates are summarized in SI 4.

## Discussion

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In the Isle of Rum red deer study population, average parturition dates have advanced 12.3 days over the last 35 years. Previous research has identified the contribution of plastic changes in response to warming temperatures to this change [38]. Here we have shown that adaptive evolution likely played a role too. Below we discuss the significance of this result for the red deer population, and also the strengths and challenges associated with the quantitative genetic study of evolution in wild populations.

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Moyes et al. [36] identified the trend toward earlier parturition dates in the Isle of Rum red deer population, and related a substantial component of it to local climate warming. In addition, within-individual plasticity is sufficient to explain the relationship between temperature and parturition date, and plasticity in response to increasing temperature explains a change of  $-2.8$  days over the study period, which is equivalent to 23% of the total phenotypic change [38]. There is little evidence of variation among females in their plastic responses to temperature [38]. Therefore the plastic response to temperature is unlikely to have changed (by genetic evolution or other means) over the study period, and a change in the shape of individuals' plastic responses (reaction norms) probably did not contribute to the change in mean parturition dates.

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The present work thus reveals a major new aspect of the complex picture of the dynamics of parturition date in this population, by identifying a role for evolution concurrent with the previously-identified plastic responses. We estimated that evolution for parturition date accounted for a total change of  $-2.1$  days (95%CI  $[-4.5; 0.7$  days]) over the study period. This estimate relies on the modern and conservative version of BLUP-regression, which accounts for criticisms made in [23] and [22], in particular by including calf<sup>7</sup> year of birth as a covariate. Taking this approach yielded a conservative estimate of genetic change that accounted for 15% of the observed phenotypic change. As expected, the less conservative alternative of not including year as a covariate gave a more rapid estimate of evolution:  $-2.4$  days (95%CI  $[-4.9; -0.2$  days]). The true rate of evolution probably lies between the conservative and the less conservative estimates. We obtained almost identical results from animal models fitted to the untransformed data (see SI 5), although these models performed relatively poorly (skewed residuals and poor MCMC mixing) which may impair the reliability of estimates.

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Our results suggest modest roles for changes in demographic structure (and essentially no role for changes in offspring sex-ratio). Shifting proportions of females of different reproductive status and ages had a predicted combined effect of  $-0.9$  days (about 7% of the phenotypic change). These effects were also identified in [37]. Changes among individuals, other than change in breeding values, therefore probably explains only a small (but non-negligible) fraction of the observed phenotypic change. However summing the effects of genetic change, plasticity in response to temperature [38] and changing demographic structure still leaves 55% of the change unexplained. Plastic responses to other environmental variables likely account for some of the remaining change, since the calculation in [38] does not consider the response to any variables other than mean temperature during a five month period. In particular, other climatic variables such as average temperatures during other times of the year, temperature variability, rainfall and wind speeds probably affect reproductive traits in the red deer [37]. In addition, the evolution of indirect genetic effects [59] may play a role.

The indication of evolution towards earlier parturition dates is consistent with previous work, which found the trait to be heritable [39] and under selection for earlier dates [40] in this population. Under ideal conditions, the product of heritability and strength of selection predicts the evolutionary response to selection [42,60]. However, this "breeder's equation" frequently fails to give reliable predictions in wild populations [43,60]. Simultaneous selection on genetically correlated traits is likely to be a major cause of this failure, because fitness is generally causally affected by many traits and genetic correlations are common [44]. Here, however, we obtained a close match between the estimated rate of evolution and the response to selection predicted by the breeder's equation, both in its univariate and in its bivariate forms. We cannot discard the possibility that this close match might be in part a coincidence, for instance if the indirect response to selection on a trait not included in the analysis pulled evolution in one direction but genetic drift pulled it back to match the observed rate of evolution. Nevertheless, our results are consistent with selection acting on parturition date directly (i.e., it was not significantly affected by selection on birth weight), so that its evolutionary trajectory can be predicted from a univariate breeder's equation [60].

We estimated evolution and selection averaged over the study period to obtain the total evolution and response to selection expected over the period. However if an

increase in temperature explains selection for earlier parturition, it is possible that selection has intensified in more recent years, and that selection was strongest in warmer years(e.g., [61]). The multivariate models we used to estimate selection allowed the estimation of selection by correcting for fixed and random effects in both parturition date and fitness traits, but are not well suited to estimate changes in selection. Future work could verify the selective scenario by estimating the interaction between parturition date and temperature in a generalized linear model of fitness, but care should then be taken to correct for the effect of time or other selectively irrelevant aspects of variation in parturition date.

A changing climate is probably not the only selective agent relevant to the evolution of parturition date in this red deer population. Indeed, selection was stronger among females who died of natural causes (with a predicted response to selection of  $-2.0$  days) than among the whole population, which includes shot females (with a predicted response of  $-1.37$  days), and especially so among shot females only ( $+0.10$  days). Culling may alter selection on parturition date, possibly by removing females from the population at random with respect to their potential parturition dates, thus diluting natural selection. Alternatively, culling may not be random with respect to parturition date, but somehow exert a type of artificial selection for later parturition dates which thus effectively opposes natural selection. Either way, culling may slow down the adaptive response to natural selection in the population. If confirmed, this result would add to the list of evolutionary consequences of culling [15, 57].

## Conclusion

The breeder's equation's prediction corresponds closely to the estimate of the evolutionary rate obtained from the trend in breeding values in the deer population, but it is important to highlight that this genetic change is much less than the observed phenotypic change. This mismatch is not surprising given that several mechanisms of phenotypic change, with a genetic basis or not, have been identified on the Rum red deer population (in our analyses presented here as well as also [37, 38]). More generally, our results illustrate how phenotypic change can be simultaneously due to both plastic and genetic changes [6, 8, 43]. Plastic changes in response to climate change appear

common, but that does not exclude concurrent evolutionary change in response to  
climate change [14]. Evolutionary changes are more difficult to infer than plastic  
changes, and few tests of evolution have been performed [14, 18, 43]. Moreover, here as  
in other systems, non-trivial contributions of evolution may represent only a fraction of  
the overall phenotypic trend [57, 62]. Evidence for plastic responses should not be taken  
as reason to dismiss a role for genetic change [63, e.g.], nor the other way around. As  
another side of the same coin, our results highlight the insights that a quantitative  
genetic perspective brings to the study of phenotypic trait dynamics. As outlined above,  
the breeder's equation often fails to predict phenotypic change in the wild. One possible  
explanation for this failure is 'cryptic evolution', where genetic change is hidden by  
plastic changes [43]. Our results illustrate that a simple application of the breeder  
equation can work, but it should be tested by comparison to estimates of genetic  
changes, not of phenotypic changes.

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