

# The role of selection and evolution in changing parturition date in a red deer population

Timothée Bonnet<sup>1\*</sup>, Michael B. Morrissey<sup>2</sup>, Tim H. Clutton-Brock<sup>3</sup>, Josephine M. Pemberton<sup>4</sup>, Loeske E.B. Kruuk<sup>1</sup>

**1** Research School of Biology, 46 Sullivans Creek Road, , The Australian National University, Canberra ACT 2600, Australia.

**2** School of Biology, University of St Andrews, St Andrews, United Kingdom.

**3** Department of Zoology, University of Cambridge, Cambridge, United Kingdom.

**4** Institute of Evolutionary Biology, University of Edinburgh, Edinburgh, United Kingdom.

\* timotheebonnetc@gmail.com

## Abstract

Changing environmental conditions cause changes in the distributions of phenotypic traits in natural populations. However, determining the mechanisms responsible for these changes and, in particular, the relative contributions of phenotypic plasticity vs evolutionary responses, is difficult. To date, to our knowledge no study has reported evidence that evolutionary change underlies the most widely-reported phenotypic response to climate change: the advancement of breeding times. In a wild population of red deer, average parturition date has advanced by nearly two weeks in four decades. Here we quantify the contribution of plastic, demographic and genetic components to this change. In particular, we quantify the role of direct phenotypic plasticity in response to increasing temperatures and the role of changes in the population age and stage structure. Importantly, we show that adaptive evolution likely played a role in the shift towards earlier parturition dates. The observed rate of evolution was consistent with a response to selection, and was less likely to be due to genetic drift. Our study

provides a rare example of observed rates of genetic change being consistent with theoretical predictions, although the consistency would not have been detected with a solely phenotypic analysis. It also provides, to our knowledge, the first evidence of both evolution and phenotypic plasticity contributing to advances in phenology in a changing climate.

## Introduction

Climate change affects various aspects of biodiversity across the planet (e.g., [1, 2]). In particular, shifts in phenotypic distributions within populations are widely reported, for a variety of morphological, phenological or life-history traits [2–4]. Surprisingly, however, little is still known about the relative contributions of mechanisms underlying these shifts [5]. Within a population, phenotypic distributions may change due to a change in population structure (e.g., age-structure or sex-ratio), due to phenotypic plasticity (within or between individuals), and due to genetic change [6–8]. The exact mixture of mechanisms driving phenotypic change will determine the future of a population facing a prolonged change in environmental conditions [9], for several reasons. First, the consequences of changing population structure are variable and may be idiosyncratic (e.g., [8, 10]). Second, phenotypic plasticity can provide an efficient way to cope with a changing environment but its effect may be short-lived and even maladaptive [11–13]. Third, genetic evolution, when driven by natural selection, can improve population growth rate, potentially contributing to long-term population persistence [12].

In wild populations the respective contributions of plasticity vs evolution remain unknown for the vast majority of documented phenotypic changes [14, 15] (note that by evolution we mean genetic change, here and in the rest of the manuscript). To date, most of the evidence for evolutionary responses to climate change comes from plants [16]. In contrast, despite numerous examples of phenotypic changes apparently related to climate, there have been surprisingly few examples demonstrating unambiguously that a vertebrate population is evolving in response to climate change (see discussions in [17–20]). This lack of evidence may in part be due to the question not being prioritized [14, 15]. However it probably also reflects the substantial challenges inherent in testing for adaptive evolution, in terms of requirements for appropriate data

and statistical methods. For wild populations in which experimental manipulations are not feasible, the most plausible means of testing for the genetic basis of phenotypic changes is to use long-term pedigree data to test for changes in ‘breeding values’, the estimated genetic merit of individuals as ascertained from the phenotypes of their relatives [21]. This needs to be done with care, as trends in predicted breeding values can be confounded with environmental trends unless appropriately controlled for [22], and the precision of estimates of evolutionary rates can be inflated if the correlation structure of breeding value estimates is not properly handled [23]. To our knowledge, among the studies of wild vertebrate populations that properly account for uncertainty in breeding value predictions, only three have found evidence of genetic change underlying phenotypic change in line with selection pressures changing with climate: plumage colouration in collared flycatchers [20], and body size in Siberian Jays [24] and snow voles [25]. However only with more empirical studies explicitly testing for evolution will it become possible to say whether the current lack of evidence also reflects a generally slow rate of adaptation to environmental change in natural populations [26].

Climate change may have impacts on numerous aspects of an organism’s biology, but phenology (i.e., the seasonal timing of life-history events) appears to be particularly affected [3, 27–29]. Dramatic changes of phenologies in response to earlier onset of spring are particularly well documented in mid- and high-latitude passerines, where breeding times are occurring earlier in numerous populations and species [18, 30]. The study of avian systems in particular has shown that a fine tuning of phenology to the climate is crucial in determining individual fitness. Mismatches between mean breeding date and a fitness optimum that shifts with climate may re-shape selective pressures and hence potentially reduce population growth rate [31], although establishing the link between individual-level and population-level processes is challenging [32, 33]. The effects of climate change on mammalian phenology are less well documented and less clear than those of birds [29], and may be more complex because mammals’ long gestation times likely make their breeding phenology sensitive to climate across a longer time-frame [17]. Finally, despite the extensive evidence for phenotypic shifts in phenology, the few studies that test for a genetic basis to changes in phenology in wild populations have not found evidence of genetic changes [34–37]. One possible exception is the change of egg hatching date in winter moths [38], where a common garden

experiment suggested a contribution of genetic change.

In a population of red deer (*Cervus elaphus*, Linnaeus 1758) on the Isle of Rum, NW Scotland, parturition date has advanced at a rate of 4.2 days per decade since 1980, a change that has been linked to temperatures and other weather conditions in the year preceding parturition, especially around the time of conception [39, 40]. Previous studies of this population have shown that phenotypic plasticity in response to temperature and population structure explain a substantial proportion (23%) of the advance in parturition dates [40], and also that within-individual plasticity is sufficient to explain the relationship between temperature and parturition date [41]. However, the documented plasticity does not explain the majority of the observed phenotypic change, leaving room for processes that have not been investigated as of yet. It is plausible that evolution plays a role because the observed phenotypic change is qualitatively consistent with a genetic response to selection: parturition date is heritable in this population [42] and also under selection for earlier dates [43].

In this study we use quantitative genetic animal models [21, 44] to estimate the rate of evolution in parturition date and the contribution of plastic and demographic processes to the observed shift in phenology in the Rum red deer study population. We start by considering the response to selection that might be expected from the observed strength of selection and (narrow-sense) heritability of parturition date, based on a simple “breeder’s equation” prediction [45]. One of the most striking conclusions from the recent application of quantitative genetic theory in evolutionary ecology has been the failure of univariate breeder’s equation predictions to capture trait dynamics in wild populations [46, 47]. This may be for multiple reasons, foremost of which is likely to be the unrealistic assumption that only the focal trait is relevant. We therefore also consider a multivariate breeder’s equation [48], and ask how selection on offspring size and the genetic correlation between parturition date and size alters the expected evolutionary response. However there is a second, less well-explored reason for the failure of the theory: predicted genetic responses to selection are often compared to observed rates of phenotypic change rather than of genetic change. Phenotypic changes are generally affected not only by genetic changes but also by numerous non-genetic processes, and thus may poorly reflect underlying genetic changes. As the central analysis of this work, we use trends in breeding values to estimate the rate of evolution

in parturition date and to test whether it is compatible with the response to selection 108  
predicted by either the univariate or multivariate breeder's equation, or with genetic 109  
drift. We also quantify the effect of non-genetic processes contributing to phenotypic 110  
change along with evolution. 111

## Methods 112

### Study population 113

We used data from a long-term study of the unmanaged population of red deer living in 114  
the North Block of the Isle of Rum, Scotland (57°01' N, 6°17' W), for the years 115  
1972-2016. The work takes place under a UK Home Office Project Licence under the 116  
Animals (Scientific Procedures) Act 1986 as amended (licence no 70/8818 held by 117  
J.M.Pemberton), and with the assistance of Scottish Natural Heritage, which manages 118  
the Isle of Rum National Nature Reserve. Within the ca. 12 Km<sup>2</sup> of the study area, 119  
calves are marked with ear tags (and a collar for females) shortly after birth, in order to 120  
record detailed life-histories of individuals throughout their lives [49]. DNA was 121  
obtained from ear punches, post-mortem tissue and cast antlers. The population 122  
pedigree was reconstructed from single nucleotide polymorphisms as in [50], using the R 123  
package SEQUOIA [51]. 124

We studied the selection and genetics of parturition date, the date on which a female 125  
gave birth to a calf in a given year. We therefore focus on females, because males do not 126  
express the trait of parturition date —though they may affect it, in both genetic and 127  
non-genetic ways. We estimated the selection on parturition date among males in 128  
supplementary information S5 Text, and found its direction to be unclear and its 129  
magnitude to be likely weak, so we did not consider it further in the calculations that 130  
follow. However males were retained in the pedigree and contributed to the calculation 131  
of quantitative genetic parameters by informing the relatedness between individuals. We 132  
included females that are still alive, even though their lifetime fitness is still unknown, 133  
in order not to introduce a fraction of individuals missing 'not at random' with respect 134  
to fitness and parturition date [52]. However, excluding living females (12% of 135  
individuals) from the analysis gave slightly stronger estimates of selection but did not 136

affect results qualitatively (see S6 Figure).

Death is usually due to natural causes, as there has been no culling in the study area since 1973, but individuals are occasionally shot when they visit areas surrounding the study area. Mortality due to culling may exert a kind of artificial selection that studies of natural selection may want to exclude. However, our goal here was to understand the causes of phenotypic change, be they natural or artificial. We therefore retained culled females in our main analyses. These shot females represented a small but non-trivial portion (15%) of the data set (Table 1). Therefore, in a sub-analysis, we also considered selection only among females who died of natural causes (i.e., excluding both shot females and females still living), and discuss the consequences of culling for selection and evolution in this system.

Parturition date was measured as the number of days after May 1, because virtually no parturition occurs before that date (except for 2 outliers from late March, which were discarded prior to analysis). Values were (natural-)log-transformed in order to obtain residuals with distributions close to Gaussian. The logged values were multiplied by 100 for reporting convenience (in particular, variance components would have been of the order  $10^{-5}$  without this second step). The working phenotype in all models was therefore  $z = 100\log(B)$ , where  $B$  is the parturition date in number of days after May 1<sup>st</sup>. Results were converted back to days (see SI 1 for details of the back-transformation process) to facilitate biological understanding. We report results using untransformed data in S8 Text. In brief we obtained similar results from animal models fitted to the untransformed parturition date data, but these models performed relatively poorly (skewed residuals and poor MCMC mixing) which may impair the reliability of estimates.

Data type	Number of	Excluding shot	Shot	Total
Parturition date	Individuals	582	158	740
	Records	2921	463	3384
Lifetime breeding success	Individuals	1614	282	1896

**Table 1.** Sample sizes for lifetime breeding success (LBS) and parturition date for years 1972-2016. All numbers refer to females; parturition date is measured repeatedly on individuals. LBS is measured over a lifetime, and there is only one measure per individual. All females in the study population have an LBS record, including those that died as calves and therefore did not breed and do not have records for parturition date.

## Quantitative genetic analysis

161

### Univariate animal model

162

We fitted a univariate animal model of female (log) parturition date in order to estimate heritability and change in breeding values [21,44]. The fixed effects in the model were: the sex of the offspring; the female's 'reproductive status', which can take five values to represent different recent reproductive history: 'naive', 'true yield', 'summer yield', 'winter yield', 'milk hind' [41]; the female's age in years (first and second order polynomial, which provided a good fit to the data, see S2 Text); a continuous covariate of the expected proportion of immigrant vs resident genes in each female to model gene flow into the population ('genetic group', see S3 Text); the offspring birth year as a continuous variable, see next section for details; the female's pedigree-based inbreeding coefficient [53,54] calculated using the R package MCMCglmm [55]; and finally, air temperature around the rut period on the year preceding a parturition (mean daily maximum temperature between 17 July and 20 November, following [41]). The covariate temperature aims to capture the plastic response to temperature, in particular through its effect on the timing of mating, as shown in [41]. The factor sex of the offspring also capture a plastic response, albeit indirect and complex, because sex of the offspring is affected by variation in population density, winter rainfall and presumably other factors related to nutritional stress in females [56]. Population density, estimated as the number of resident adult females in a given year, had a significant effect on (log) parturition date at the beginning of the study period (e.g., between 1974 and 1987, [49]), but we found no effect in the full data-set (slope  $-0.38$ , standard error  $0.63$ ) and therefore excluded this variable from the analyses.

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

The random effects decomposed the variance not accounted for by fixed effects into six components: additive genetic variance; 'permanent environment' variance (estimable from the repeated measures of the same females across multiple years; [57]); maternal effects variance (i.e. associated with the mother of the breeding female, and hence grand-mother of the new calf); variance associated with the offspring birth year; variance associated with the breeding female's (i.e. mother of the calf) cohort; and residual variance.

184

185

186

187

188

189

190

Thus, the model of (log) parturition date ( $z$ ) of female  $i$  in year  $j$  can be written 191

$$z_{ij} = \mu + \mathbf{X}^T \mathbf{b} + a_i + p_i + m_i + y_i + c_j + r_{ij} \quad (1) \quad 192$$

where  $\mu$  is an intercept,  $\mathbf{X}$  is a vector of fixed predictors (including the covariate 192  
offspring birth year),  $\mathbf{b}$  is a vector of fixed effects,  $a$ ,  $p$ ,  $m$ ,  $y$  and  $c$  are random effects 193  
with which to estimate the variance associated with additive genetic values (i.e., 194  
breeding values), permanent environment, maternal identity (i.e. grand-mother of calf), 195  
offspring birth year, and female's cohort, respectively, and  $r$  is the residual. The 196  
breeding values ( $a$ ) are normally distributed as  $(a_1, \dots, a_n)^T \sim N(0, \sigma_A^2(z)\mathbf{A})$ , where 197  
 $\sigma_A^2(z)$  is the additive genetic variance for (log) parturition date,  $n$  is the number of 198  
females, and  $\mathbf{A}$  is the relatedness matrix between individuals. The heritability of (log) 199  
parturition date was estimated as  $\sigma_A^2(z)$  divided by the total phenotypic variance. 200

We used this animal model to estimate the individual-level repeatability (in addition 201  
to the heritability) of (log) parturition date, as the sum of the proportions of variance 202  
explained by all effects that are constant for an individual: inbreeding and genetic group 203  
(which are fixed effects and had the variance they explain calculated as the variance in 204  
the product of their data values by their parameter estimates [58]) and additive genetic 205  
variance, permanent environment variance, maternal variance and female's cohort 206  
variance (that is, all random effects but offspring birth year and residual). 207

We ran all models in the Bayesian framework of the R package MCMCglmm [55] 208  
with Gaussian errors for (log) parturition date. We report posterior modes and 95% 209  
highest posterior density credible intervals. For this univariate model, we used the 210  
default inverse gamma priors for variance components, with shape and rate parameters 211  
both equal to 0.001 (equivalent to a variance and degree of belief of 1 and 0.002, 212  
respectively). We run models for 130000 Markov chain Monte Carlo iterations, with a 213  
burn-in of 30000 and thinning of 100, and checked mixing and convergence by visual 214  
inspection of trace plots for all parameters. All code for analyses is provided in S9. 215

## Selection 216

We estimated selection acting on (log) parturition date by assessing the association 217  
between a female's fitness and her propensity for (log) parturition date corrected for 218



various effects. We measured fitness as *lifetime breeding success* (LBS in the text,  $W$  in equations), which is the number of offspring that have been produced by an individual, calculated for all females in the database, whether or not they survived to breeding and therefore also had parturition records.

Selection was estimated using a model of the covariance between (log) parturition date and fitness. We used a bivariate generalized linear mixed model, with LBS modelled as an over-dispersed Poisson trait (with log link function) and (log) parturition date ( $z$ ) modelled as a Gaussian trait. This model can be written as

$$[z, W] \sim \mathbf{X}\mathbf{b} + \mathbf{D}_1\mathbf{m} + \mathbf{D}_2\mathbf{y} + \mathbf{D}_3\mathbf{c} + \mathbf{D}_4\mathbf{p} + \mathbf{I}\mathbf{r} \quad (2)$$

where  $\mathbf{X}\mathbf{b}$  represents fixed effects (only an intercept and genetic group for fitness, and the same fixed effects for log-parturition date as above, except for temperature),  $\mathbf{m}$ ,  $\mathbf{y}$ ,  $\mathbf{c}$ ,  $\mathbf{p}$  are random effects associated with maternal effects (the identity of the mother of the breeding female), the year of calving, the female's cohort (i.e. her year of birth), and the individual female's identity (or 'permanent environment' effect, because of the repeated measures), respectively.  $\mathbf{D}$ -matrices link random effect levels to observations, and  $\mathbf{I}\mathbf{r}$  represents the residuals.

Note that  $W$  is only measured once for each individual, unlike the repeated measures on parturition date ( $z$ ). For  $W$ , variance components are therefore null for  $\mathbf{y}$  (the year of calving) and  $\mathbf{p}$  (the 'permanent environment' component of a trait, derived from repeated measures on an individual). `MCMCglmm` accommodates this difference in replication between the two traits by allowing the individual-level random effect  $\mathbf{p}$  for the replicated trait (parturition date) to covary with the residual variance  $\mathbf{r}$  of the non-replicated trait (fitness), thus providing a covariance between the repeatable part of an individual's parturition date and her fitness (for a comparable example, see [59]).

The selection differential on (log) parturition date was calculated as the sum of this individual-level covariance, plus the maternal-effect covariance between (log) parturition date and fitness (i.e. covariance among effects of the breeding females' mothers on their daughters' log-parturition dates and fitness). Selection differentials characterize the within-generation change in phenotype due to selection. We therefore standardized this value by generation time (8 years) to be expressed in rate of change per year, or in total

change over the study period. Selection differentials were divided by two, because the covariances were estimated from females only. Males do not express the trait but nevertheless carry genes relevant to parturition date in females. Selection on parturition acts on only half of the population, and the expected response to selection is half that predicted from the strength of selection in females. As outlined above, we considered the possibility that males are nevertheless selected for the trait in S5 Text, but as we did not find support for any selection through males we did not consider it further. We also estimated a selection gradient [60], calculated as the selection differential divided by the corresponding variance (that is, the sum of the individual-level and mother-level variance components for parturition date).

When expected fitness follows a log-normal distribution, selection parameters can be equivalently calculated on the scale of the data using relative fitness, or on the log-scale using absolute fitness [61, 62]. Because our model uses a log-link function for absolute LBS, parameter estimates involving LBS are on the latent scale, but these are directly interpretable as selection differentials and selection gradients relating to *relative* fitness on the data scale.

For multivariate models we used parameter-expanded priors for variance components with degree of belief equal to 0.002 (for random effects fitted for a single trait) or 2 (for random effects fitted to both traits), working mean of 0 and variance of 1000. For residual variances we used the same degrees of belief (0.002 or 2), but did not use parameter-expansion since this is not possible in MCMCglmm. Estimates of selection were identical when using inverse-gamma priors instead of parameter-expanded priors (S4 Figure). We ran these models for 260000 Markov chain Monte Carlo iterations, with a burn-in of 60000 and thinning interval of 200.

### Univariate and multivariate predictions of evolution

The response to selection (the per-generation change in the mean value of the trait) was predicted as the product of the heritability in (log) parturition date and the selection differential, following the univariate breeder's equation [45]. The equation was applied to estimates from the model of (log) parturition date data, and the predicted response was subsequently back transformed to days. Calculations were done on the MCMC posterior distributions of the heritability and the selection differential, in order to propagate the

uncertainty in these two parameters. The univariate breeder’s equation ignores the fact that the adaptive evolution of a focal trait depends not only on direct selection on that trait, but also on selection on any other traits that are genetically correlated with the focal trait [48]. This assumption may explain in part the common mismatch between predicted and observed evolution in natural populations [47], but it can partly be relaxed by incorporating analyses of relevant associated traits and estimating multivariate selection and genetic covariances: the multivariate response to selection can then be predicted as the product of the additive genetic variance-covariance matrix  $\mathbf{G}$  and the vector of multivariate selection gradients  $\boldsymbol{\beta}$  ( $\Delta\mathbf{Z} = \mathbf{G}\boldsymbol{\beta}$ ) [48,60].

In the Rum red deer study population, a calf’s birth date is correlated with its birth weight [40,49], a trait also under selection [43]. We therefore applied a bivariate breeder’s equation to parturition date and calf birth weight to estimate the effect of indirect selection on the predicted evolutionary response of parturition date to selection.

We extended the animal model of (log) parturition date (eq. 1) to a bivariate animal model of (log) parturition date and offspring birth weight, using the same fixed effects and random effects for both traits. Note that in this model, the calf’s birth date and birth weight ( $bw$ ) are both being treated as the phenotype of the mother; the treatment of offspring birth weight as a trait of the mother is justified by the observation that more than 90% of the genetic variance in birth weight is maternal-genetic variance rather than direct-genetic variance [57]. This model estimated an additive genetic covariance between the two traits,  $\sigma_A(z, bw)$ , which can be divided by the square root of the product of the two additive genetic variances,  $\sigma_A^2(z)$  and  $\sigma_A^2(bw)$ , to obtain a genetic correlation.

Finally, we extended the bivariate selection model (eq. 2) to a trivariate model also including offspring birth weight (along with log-parturition date and LBS). For birth weight we used the same fixed and random effects as described above for (log) parturition date. We summed the appropriate covariances of the phenotypes with LBS to obtain a vector of selection differentials  $\mathbf{s}$ . We summed the appropriate variances and covariances for the two phenotypes to obtain a phenotypic variance-covariance matrix  $\mathbf{P}$ . We then applied  $\mathbf{P}^{-1}\mathbf{s}$  to obtain  $\beta_z$ , the direct selection gradient on (log) parturition date corrected for the indirect selection on birth weight, and  $\beta_{bw}$ , the direct selection gradient on birth weight corrected for the indirect selection on (log) parturition date [63].

The response to selection could then be calculated as  $\beta_z \sigma_A^2(z) + \beta_{bw} \sigma_A(z, bw)$  [60]. 311

We also expressed predicted rates of evolution in Haldanes, that is, in units of 312  
standard deviation per generation [64]. We did not express the results in Darwins (i.e., 313  
change in log mean phenotype per million years) because parturition dates have no 314  
natural zero point, and therefore mean-standardisation is not meaningful here. 315

## Components of change 316

### Genetic change 317

Using the univariate animal model of (log) parturition date containing year as a 318  
covariate (see below), we fitted a linear regression of best linear unbiased predictors (i.e., 319  
model predictions for the values of a random effect's levels, BLUPs hereafter) for 320  
individual females' breeding values against the mean birth year of their offspring to each 321  
posterior sample. This generates a posterior distribution for the slope of change in mean 322  
breeding value [23]. In addition, to visualize any potential non-linearity in genetic 323  
change, we fitted a penalized thin plate regression spline (i.e., a smoothing function) of 324  
mean offspring birth year to the BLUPs for individual females' breeding values for every 325  
posterior sample, thus generating the posterior distribution of the time-dynamic of 326  
breeding values among birth years [65]. 327

Changes in breeding values may indicate a response to directional selection, but they 328  
can also be produced by random fluctuations under non-directional evolutionary models, 329  
such as genetic drift. To assess this possibility, we also compared the posterior 330  
distribution of the estimated change in breeding values to the change possible under 331  
genetic drift alone, using simulations conditional on the pedigree and on estimated 332  
additive genetic variance, as described in [23]. In general, breeding values predicted by 333  
animal model BLUPs are not equal to the true breeding values, but are influenced by 334  
environmental random deviations [22]. As a consequence, a linear regression of BLUPs 335  
may confound genetic and non-genetic (e.g. plastic) change and may produce a biased 336  
estimate of evolution. This issue can be addressed by including year as a covariate in 337  
the animal model used to obtain BLUPs. Unfortunately, the solution returns a 338  
conservative estimate of the rate of change in breeding values, because the animal model 339  
ascribes some of the genetic change to the year effect [22]. We opted to report these 340

341 primarily conservative estimates of evolution, based on an animal model that did  
342 contain offspring birth year as a covariate (see above). However we also re-fitted the  
343 animal model without a fixed effect for birth year and re-calculated the change in  
344 BLUPs for breeding values estimated this way.

345 In an alternative approach, we estimated expected genetic change using the  
346 secondary theorem of selection, from the additive genetic covariance between trait and  
347 relative fitness - also known as the Robertson-Price identity [59, 66, 67]. To this end, we  
348 expanded the bivariate model of parturition date and fitness described in equation 2 by  
349 adding a random effect of additive genetic variance-covariance. Assuming log-normality  
350 of LBS, the expected rate of evolution per generation in parturition date can be  
351 estimated directly from the additive genetic covariance between parturition date and  
352 LBS [62], divided by two to account for females making up only half of the population.

### 353 **Other contributions to phenotypic change**

354 Finally we estimated the contributions of several other terms in equation (1) to the  
355 trend in parturition date. We used Geber's method [6, 8] on model predictions to  
356 estimate the independent contribution of changes in the class structure of age and  
357 reproductive status, in plastic responses to temperatures and sex of the offspring, and  
358 the independent contribution of changes in levels of inbreeding (as assessed from the  
359 pedigree inbreeding coefficient) and gene flow (as assessed by the genetic groups effect)  
360 to the phenotypic change in parturition date. Briefly, this method estimates the  
361 contribution of change in a parameter mean ( $\bar{k}$ ) to change in a trait mean ( $\bar{z}$ ) as the  
362 product of the partial derivative of  $z$  on  $k$  ( $\frac{\partial \bar{z}}{\partial k}$ ) and the slope of  $k$  on time ( $\frac{\Delta \bar{k}}{\Delta t}$ ). We  
363 applied the equation to each sample of the posterior distributions in order to propagate  
364 the uncertainty in the estimated trends. In addition to calculating the net effect  
365 through the study period, we calculated  $\frac{\partial \bar{z}}{\partial k} \bar{k}_t$  for each year  $t$  to visualize the dynamic of  
366 changes in effects through time graphically.

367 We did not use random effects to estimate non-genetic components of change  
368 because random effects other than additive genetic effects are linearly independent of  
369 years by construction, and any change in females' maternal effects or permanent  
370 environment effects should be absorbed into the fixed effect of offspring birth year.

## Results

371

### Phenotypic change

372

The average parturition dates for female red deer became later from 1972 to 1980 (probably reflecting increased population density, [49]), after which they advanced at an apparently constant rate (Fig. 1). A linear regression estimates the change in parturition date to be a total of  $-12.3$  (95%CI  $[-14.5; -10.1]$ ) days over the 45-year study period (from 1972 to 2016). The slope was identical ( $-12.3$ ) when data were aggregated among females (i.e. estimated using mean parturition date over mean breeding year for each female), which may be more comparable to results below.

373

374

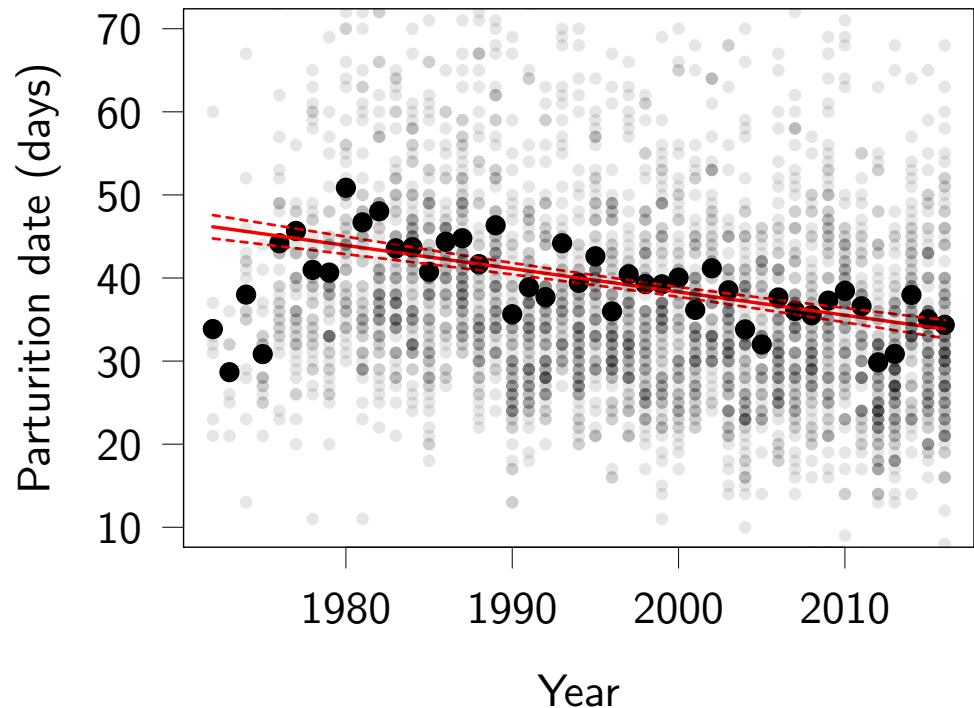
375

376

377

378

379



**Fig 1.** Phenotypic trend in red deer parturition dates, in days after May 1st. Large black dots represent annual means, small grey dots represent individual parturition dates, with the darker shades indicating more calves being born on a given day. About 4% of individual parturition dates fall outside the plotted region (10th May - 12 July; note these are still included in the analyses). The red lines represent the slope and associated 95% confidence interval of a linear regression of all individual parturition dates on year of parturition. Note that the years 1972-1975 have very negative residuals and that the rate of change over 1980-2016 is slightly underestimated by the linear regression being fitted over all years.

## Sources of parturition date variation

Parturition date was influenced by a female's reproductive status and age, but there was no clear evidence for effects of her inbreeding coefficient, of offspring sex or of the proportion of immigrant genetic ancestry (S7 Table 7.1). Parturition date was heritable, with additive genetic variance accounting for 17% (95%CI [11%; 21%]) of phenotypic variation. The individual-level repeatability of parturition date (estimated as the sum of proportions of all variance components except offspring birth year and residual) was 19%, of which additive genetic variance was most important, with permanent environment effects and maternal effects both accounting for less than 1% of total phenotypic variation (S7 Table 7.2). The random effect for offspring birth year (which captures the variance among years over and above that corrected for the temporal linear trend) accounted for about 8% of the phenotypic variance (S7 Table 7.2). Note that proportions are essentially invariant under monotonic transformation and that these proportions of variances are equivalent on the transformed (i.e. log) and on the original data scale.

## Univariate selection and predicted response

Females with earlier parturition dates had, on average, higher lifetime breeding success: the selection differential of parturition dates estimated with LBS was  $-1.37$  days of change within a generation (95%CI  $[-2.43; -0.65]$ , see S7 Tables 7.3 and 7.4 for random effect and fixed effect estimates of the model described in equation 2). Given the heritability of parturition date of 0.17, the univariate breeder's equation predicts a response to selection of  $-0.25$  days per generation (95%CI  $[-0.55; -0.11]$ ), that is  $-1.45$  days over the 44-year study period (95%CI  $[-3.01; -0.60]$ ) (Fig. 3). The predicted response also corresponds to  $-0.031$  days per year (95%CI  $[-0.068; -0.014]$ ) or  $-0.019$  Haldanes (95%CI  $[-0.042; -0.008]$ ).

Selection was weaker among females that were culled than among females that died of natural causes, than among the complete population of females already dead, and than among the complete population of females dead and still alive (S6 Figure). Considering the subset of females that died of natural causes, the univariate breeder's equation predicts a response of  $-2.04$  days (95%CI  $[-3.37; -0.95]$ ) over the study

period. In contrast, using the subset of females who were culled, the univariate breeder's equation predicts a response of 0.11 days (95%CI [-0.64; 0.93]) over the study period.

## Bivariate selection and predicted response

Conditional on the fixed effects affecting each trait, the phenotypic correlation between (log) parturition date and birth weight was positive but weak (correlation = 0.12, 95%CI [0.05; 0.16]). The gradient of direct selection on (log) parturition date was negative (mode  $\beta_z = -0.0003$ , 95 %CI [-0.0004; -0.0002]), and that on birth weight was positive ( $\beta_{bw} = 0.0138$ , 95%CI[0.009; 0.017]). There was also additive genetic variance in offspring birth weight (0.68, 95%CI [0.57; 0.90]), corresponding to a heritability of 0.46 (95%CI [0.37; 0.62]). The additive genetic covariance between (log) parturition date and offspring birth weight was -1.78 (95%CI [-4.38; 0.56]), corresponding to a weak negative genetic correlation of -0.16 (95%CI [-0.32; 0.05]). The multivariate breeder's equation predicts a rate of evolution of -1.41 days (95%CI [-2.70; 0.11]) over the study period, which is actually similar to the univariate breeder's equation prediction of -1.45 days (difference = -0.01 days, 95%CI [-0.71; 0.55], Fig. 3)

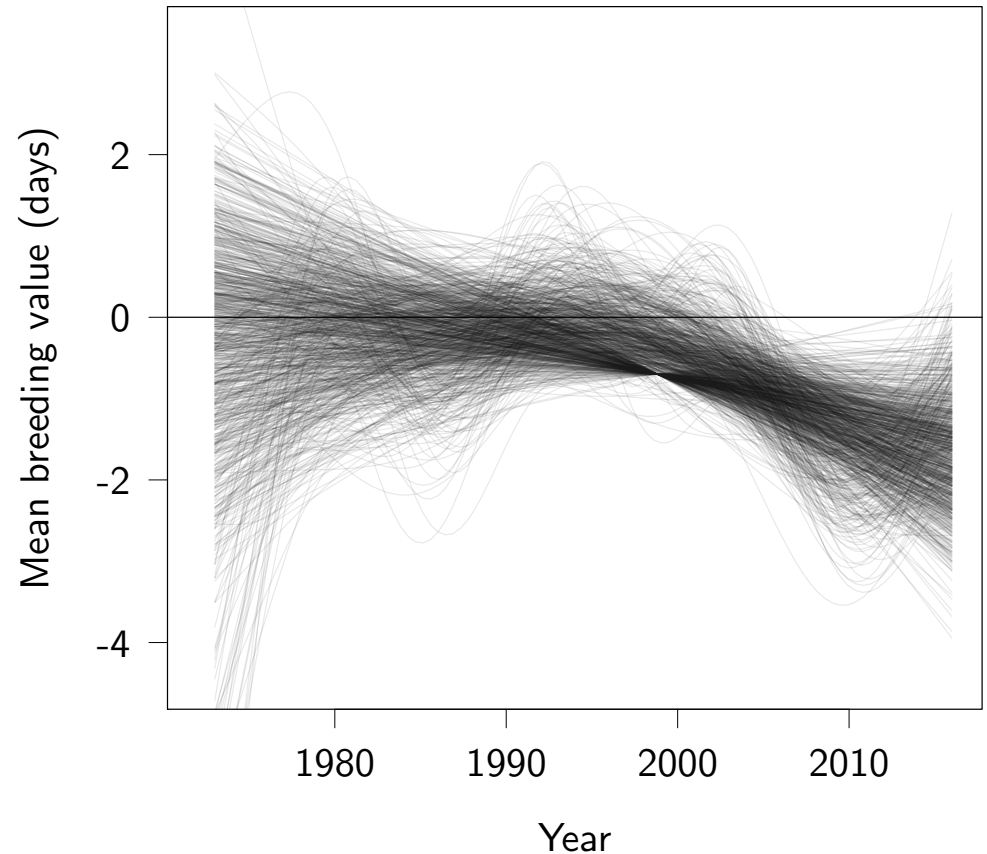
## Genetic contribution to phenotypic change

Using the most conservative method (namely the model including calf birth year as a covariate), the slopes of the linear regressions of BLUPs for parturition date breeding values on mean offspring birth year, integrated over the posterior distribution, suggests an advance in breeding values, with the slope estimated at -0.10, 95%CI [-0.23; 0.03] per year on the log-transformed scale. This result is uncertain: there is a probability of 7% that the change in BLUPs is null or positive. Time-splines fitted on the posterior distribution of the BLUPs visually support a linear decrease in breeding values (Fig. 2). The estimated rate of evolution corresponds to a total change over the study period of -2.1 days, 95%CI [-4.5; 0.7] due to genetic change (Fig. 2 and 3), equivalent to -0.045 days per year (95%CI [-0.100; 0.018]), -0.36 days per generation (95%CI [-0.79; 0.14]), or -0.028 Haldanes (95%CI [-0.062; 0.01]).

The less conservative BLUP regression estimated that evolution contributed -2.4 days (95%CI [-4.9; -0.2 days]) to the phenotypic change over the study period. The



secondary theorem of natural selection estimated a change of  $-4.9$  days (95%CI $[-10.6; -0.7]$ ) over the study period (the estimate is the additive genetic covariance between parturition date and relative LBS, after back-transformation to days. See S7 Table 7.3 for raw estimates on the scale of  $100 \times \log(\text{parturition date})$ ).



**Fig 2.** Trend in breeding values for parturition date. Each black line was obtained from a different MCMC posterior sample, by fitting a spline to the mean of estimated breeding values among individuals living in the same year. The y-axis was centered on the mean breeding values in 1972 to help interpretation. Some lines are straight because the smoother function used penalizes complex polynomials.

9% of the simulations of genetic drift generated an advance as large or larger than the change estimated from the conservative BLUP linear regression (using the posterior mode for the BLUPs trend as a point of comparison, see Fig. 3). Inbreeding tended to delay parturition date (S7 Table 7.1), and given that the estimated pedigree inbreeding inevitably increased over time with increasing pedigree depth [53], there was marginal evidence of inbreeding postponing parturition date by 0.38 days (95%CI  $[-0.04; 1.01]$ ) over the study period, thus opposing the phenotypic trend. However this prediction may

be spurious, because the increase in inbreeding coefficient was an artifact of estimating  
inbreeding from a pedigree [53]. Re-running the model without inbreeding led to almost  
identical estimates for all other parameters. The effect of gene flow (the proportion of  
immigrant genotype) was uncertain (S7 Table 7.1) and its overall predicted effect over  
the study period was a change of 0.15 days (95%CI  $[-0.34; 0.72]$ ).

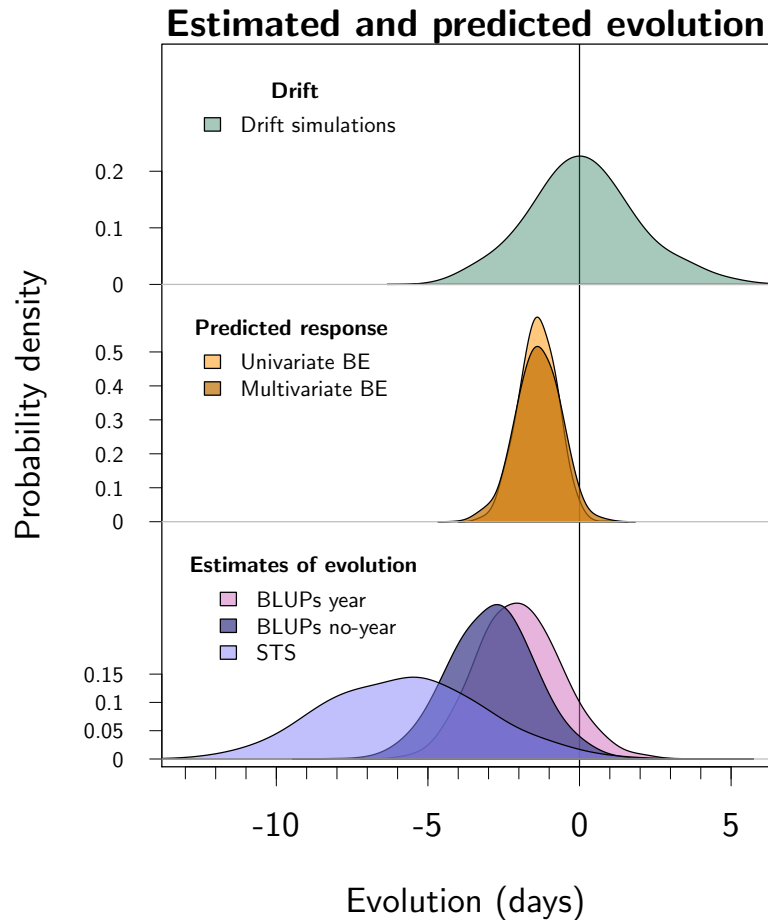
## Non-genetic contributions to phenotypic change

As in previous work [40], we found that mature females tended to give birth earlier than  
younger females, but very old females gave birth the latest (S2 Text). The effects of  
changes in the age structure on mean parturition dates tended to be in the opposite  
direction to the observed phenotypic change: during the first ten years of the study, the  
mean age of females in the study increased steadily, pushing towards earlier mean  
parturition dates ( $-3.68$  days, 95%CI  $[-5.63; -1.92]$  from 1972 to 1981). For the rest of  
the study, the change in age structure tended to delay mean parturition date slightly  
( $0.57$  days, 95%CI  $[0.39; 0.71]$  from 1982 to 2016). Over the study period the change in  
age structure had a predicted net effect of  $-0.58$  days, 95%CI  $[-1.67; 0.40]$  (Fig. 4A).  
Changes in female reproductive status had a fluctuating effect on parturition date (Fig.  
4B), with an uncertain total effect over the study period of  $-0.32$  days (95%CI  
 $[-0.87; 0.17]$ ).

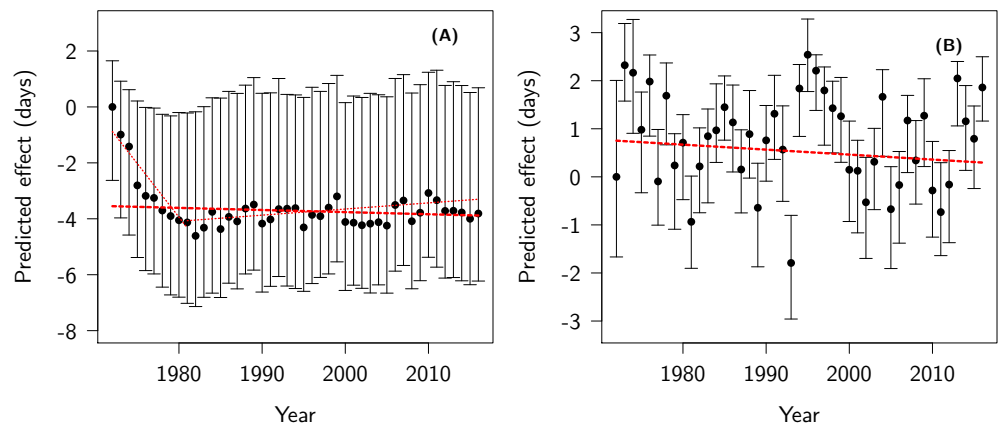
Offspring sex had no clear effect on parturition date, and since sex-ratio at birth  
remained relatively stable over the study period (despite an early decline in the  
proportion of males [56]), this parameter is predicted to have had a small overall effect  
( $-0.04$  days, 95%CI  $[-0.18; 0.04]$ ). Warmer temperatures during the previous rut season  
tended to advance parturition date, with an overall effect of  $-1.40$  days  
(95%CI  $[-3.05; 0.50]$ ) over the study period. The effect is less clear than the  $-2.4$  days  
reported in [41] most likely because our model contains a covariate for year while [41]  
did not. When we remove the year covariate we obtained an estimate of  $-2.56$  days  
(95%CI  $[-5.23; -0.69]$ ).

Fig 5 summarises all the components of change described above. Altogether, these  
effects captured a predicted change of  $-7.98$  days (95%CI  $[-12.85; -3.22]$ ) over the  
study period. This leaves an unexplained change of  $-4.99$  days (95%CI  $[-9.76; -0.13]$ ).

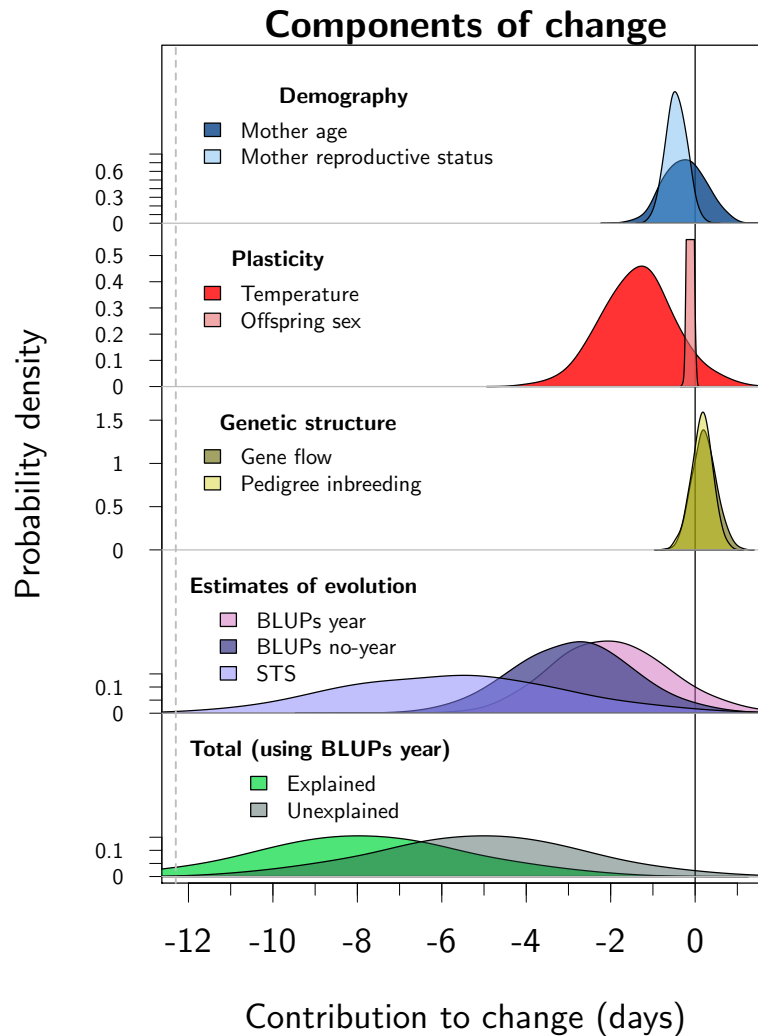
Given the model specification, the unexplained fraction must capture persistent changes 480  
in maternal effects, individual-level (referred to here as 'permanent') environment 481  
effects, and various other effects of phenotypic plasticity (other than that due to sex of 482  
the offspring and mean temperature during the rut period), which were not explicitly 483  
accounted for in the model. 484



**Fig 3.** Posterior distributions for predicted and estimated evolution over the study period, from top to bottom: 1) Evolutionary change possible due to genetic drift. This distribution was generated by simulating random changes conditional on the estimated additive genetic variance and on the pedigree. 2) Predicted evolutionary response to selection from the univariate and bivariate breeder’s equations respectively. The response to selection was estimated using univariate and bivariate breeder’s equations, where phenotypic multivariate models gave selection gradients, and animal models gave additive genetic variance-covariances of parturition date and birth weight. 3) Estimated contribution of evolution estimated in three ways: “BLUPs year” is the conservative BLUP regression with offspring birth year fitted as a fixed effect, “BLUPs no-year” is the BLUP regression without offspring birth year fitted as a fixed effect, “STS” is the secondary theorem of natural selection using the additive genetic covariance between parturition date and fitness. Parturition date was modeled using a log-transformation, and all estimates were subsequently converted to change in days over the study period (see S1 Text). Parameter estimates are summarized in S7 Table 7.1. The distributions all have the same area, but the y-axes scales vary to help visualization.



**Fig 4.** Predicted effect of (A) age-structure and (B) female reproductive status on parturition date across years. The origin of the y-axis is arbitrarily set to the predicted effect in the first year. The red thick dashed lines represent the net effect of changes in age structure and female reproductive status on parturition date reported in the text. The thin dotted lines in (A) represent the effect of changes in age structure before, and after 1981, respectively.



**Fig 5.** Posterior distributions of the components of change in parturition date over the study period. All estimates are derived from the univariate animal model described by equation 1 (except for STS, derived from the bivariate animal model). Unlike other distributions, those in the row “Estimates of evolution” relate to a single component of change, estimated in three different ways: “BLUPs year” is the conservative BLUP regression with offspring birth year fitted as a fixed effect, “BLUPs no-year” is the BLUP regression without offspring birth year fitted as a fixed effect, “STS” is the secondary theorem of natural selection using the additive genetic covariance between parturition date and fitness. To accommodate strong differences in the uncertainty around component estimates, the scale of the y-axis differs among rows, and the density of the component “Offspring Sex” was truncated.

## Discussion

In the Isle of Rum red deer study population, average parturition dates have advanced 12.3 days over the last 35 years. Previous research has identified the contribution of plastic changes in response to warming temperatures to this change [41]. Here we have shown that adaptive evolution likely played a role too (Fig. 3), and we have quantified the relative importance of demographic, plastic and evolutionary changes (Fig. 5). Below we discuss the significance of the results for the red deer population, and also the strengths and challenges associated with the quantitative genetic study of evolution in wild populations.

Moyes et al. [39] identified the trend toward earlier parturition dates in the Isle of Rum red deer population, and related a substantial component of the trend to local climate warming. In addition, within-individual plasticity is sufficient to explain the relationship between temperature and parturition date, and plasticity in response to increasing temperature explains some of the phenotypic change [41]. There is little evidence of variation among females in their plastic responses to temperature [41]. Therefore the plastic response to temperature is unlikely to have changed (by genetic evolution or other means) over the study period, and a change in the strength of individuals' plastic responses (reaction norm slopes) probably did not contribute to the change in mean parturition dates.

The present work thus reveals a major new aspect of the complex picture of the dynamics of parturition date in this population, by identifying a role for evolution concurrent with the previously-identified plastic responses. We estimated that evolution for parturition date accounted for a total change of  $-2.1$  days (95%CI  $[-4.5; 0.7]$ ) over the study period. This estimate relies on the modern and conservative version of BLUP-regression, which accounts for criticisms made in [23] and [22], in particular by including offspring birth year as a covariate. Taking this approach yielded a conservative estimate of genetic change that accounted for 15% of the observed phenotypic change. As expected, the less conservative alternative of not including year as a covariate gave a more rapid estimate of evolution:  $-2.4$  days (95%CI  $[-4.9; -0.2$  days]). As a third method, the secondary theorem of selection (STS) predicted an evolutionary change of  $-4.9$  days (95%CI  $[-10.6; -0.7]$ ), which is more

evolution than estimated by the two BLUP regressions. The STS may be more powerful than BLUP regression because the addition of a second trait (LBS) in the model provides more genetic information compare to a univariate approach. On the other hand, the STS, as applied here, has the disadvantage of assuming a log-normal distribution for fitness (LBS here) [61,62]. Log-normality is violated because LBS is zero-inflated and the variance-covariance components may therefore be inaccurate. The three methods all have possible weaknesses and none is likely to perfectly capture the amount of evolution. Nevertheless the three methods agree qualitatively and the posterior distributions for the role of evolution largely overlap between them (Fig. 3).

Our results suggest modest roles for changes in demographic structure. Shifting proportions of females of different reproductive status and ages had a predicted combined effect of  $-0.9$  days (about 7% of the phenotypic change). These effects were also identified in [40]. Changes among individuals, other than change in breeding values, therefore probably explain only a small (but non-negligible) fraction of the observed phenotypic change. However summing all the effects estimated here still leaves a change of  $-4.99$  days (95%CI  $[-9.76; -0.13]$ ) unexplained. Plastic responses to other environmental variables likely account for some of the remaining change, since we did not consider the response to any variables other than mean temperature during a five month period, and the sex of the offspring (which responds to the biotic and abiotic environment in a complex way [56]). In particular, other climatic variables such as temperatures during other times of the year, temperature variability, rainfall and wind speeds probably affect reproductive traits in the red deer [40]. In addition, the evolution of indirect genetic effects [68] may play a role. For instance, maternal genetic effects [69], which in this study would be genetic effects for how a mother influences the reproductive timing of her daughters, could evolve. However, maternal genetic effects and their possible contribution to phenotypic change are likely small in this system, because total maternal effects (which include both genetic and non-genetic maternal effects) account for less than 1% of phenoytpic variance. Other types of social interactions which influence parturition may have a genetic basis that evolves, but such effects are difficult to study without *a priori* knowledge of the relevant individual interaction mechanisms [68].

The indication of evolution towards earlier parturition dates is consistent with



previous work, which found the trait to be heritable [42] and under selection for earlier 548  
dates [43] in this population. Under ideal conditions, the product of heritability and 549  
strength of selection predicts the evolutionary response to selection [45,70]. However, 550  
this "breeder's equation" frequently fails to give reliable predictions in wild 551  
populations [46,70]. Simultaneous selection on genetically correlated traits is likely to 552  
be a major cause of this failure, because fitness is generally causally affected by many 553  
traits and genetic correlations are common [47]. Here, however, we obtained a 554  
reasonable match between the estimated rate of evolution and the response to selection 555  
predicted by the breeder's equation, both in its univariate and in its bivariate forms. 556  
We cannot discard the possibility that this match might be in part a coincidence, for 557  
instance if the indirect response to selection on a trait not included in the analysis 558  
pulled evolution in one direction but genetic drift pulled it back to match the observed 559  
rate of evolution. Other factors may have biased the prediction of evolution and made 560  
the match coincidental, in particular, an imperfect fitness measure or a missing 561  
fraction [52]. Although lifetime breeding success is widely used as a measure of fitness 562  
in evolutionary ecology, it is generally not exactly the quantity maximized by natural 563  
selection when generations overlap and the environment and the population structure 564  
vary [71,72]. Moreover, we cannot measure parturition date in females that died before 565  
reproducing, which creates a missing fraction in the estimation of selection [52]. It is 566  
possible that those females who died early are not 'missing at random' with respect to 567  
genetic merit for parturition time, meaning that the true response to selection may 568  
differ from that predicted. This second problem is in part solved by the calculation of 569  
the STS (using the additive genetic covariance between parturition date and LBS), 570  
because all individuals have a breeding value for parturition date even if they never 571  
expressed the trait. The STS is therefore estimated with a much smaller missing 572  
fraction, consisting only of local individuals that do not have a LBS record (for instance, 573  
aborted embryos). The STS clearly predicted negative evolution  $-4.9$  days 574  
(95%CI $[-10.6; -0.7]$ ), reinforcing the idea that the true selection on parturition date 575  
favors earlier dates. However, it is possible that the STS is not only related to the direct 576  
selection on parturition date, but also influenced by selection on other traits genetically 577  
correlated to parturition date. In summary, there are several potential factors which 578  
may bias the estimate of selection in one way or another, and so we interpret the 579

estimates with caution. However, our different analyses all point towards a role of selection in advancing parturition date.

We estimated evolution and selection averaged over the study period to obtain the total evolution and response to selection expected over the period. However if an increase in temperature explains selection for earlier parturition, it is possible that selection has intensified in more recent years, and that selection was strongest in warmer years (e.g., [73]). The multivariate models we used to estimate selection allowed the estimation of selection by correcting for fixed and random effects in both parturition date and fitness traits, but are not well suited to estimate changes in selection. Future work could investigate the selective scenario by estimating the interaction between parturition date and temperature in a generalized linear model of fitness, but care should then be taken to correct for the effect of time or other selectively irrelevant aspects of variation in parturition date.

A changing climate is probably not the only selective agent relevant to the evolution of parturition date in this red deer population. Indeed, selection was stronger among females who died of natural causes (with a predicted response to selection of  $-2.0$  days) than among the whole population, which includes shot females (with a predicted response of  $-1.45$  days), and especially so among shot females only ( $+0.10$  days). Culling may alter selection on parturition date, possibly by removing females from the population at random with respect to their potential parturition dates, thus diluting natural selection. Alternatively, culling may not be random with respect to parturition date, but somehow exert a type of artificial selection for later parturition dates which thus effectively opposes natural selection. Either way, culling may be slowing down the adaptive response to natural selection in the population. If confirmed, this result would add to the list of evolutionary consequences of culling [15, 65].

## Conclusion

The breeder's equation's prediction for the response to selection corresponds to the estimate of the evolutionary rate obtained from the trend in breeding values in the deer population, but it is important to highlight that this genetic change is much less than the observed phenotypic change. The mismatch is not surprising given that several

mechanisms of phenotypic change, with a genetic basis or not, have been identified on  
the Rum red deer population (in our analyses presented here as well as in [40, 41]).  
More generally, our results illustrate how phenotypic change can be simultaneously due  
to both plastic and genetic changes [6, 8, 46]. Plastic changes in response to climate  
change appear common in natural populations, but that does not preclude concurrent  
evolutionary change in response to climate change [14]. Evolutionary changes are  
substantially more difficult to infer than plastic changes, and to date few tests of  
evolution have been performed [14, 18, 38, 46]. Moreover, here as in other systems, large  
contributions of evolution may represent only part of the overall phenotypic  
trend [65, 74]. Thus evidence for plastic responses should not be taken as reason to  
dismiss a role for genetic change [75, e.g.], nor the other way around. As another side of  
the same coin, our results highlight the insights that a quantitative genetic perspective  
brings to the study of phenotypic trait dynamics. As outlined above, the breeder's  
equation often fails to predict phenotypic change in the wild. One possible explanation  
for this failure is 'cryptic evolution', where genetic change is hidden by plastic  
changes [46]. Our results illustrate that a simple application of the breeder equation can  
work, but that it should be tested by comparison with estimates of genetic changes, not  
of phenotypic changes.

## Acknowledgments

Thanks to Scottish Natural Heritage for permission to work on the Isle of Rum National  
Nature Reserve, and their staff on Rum for support. Thanks to everybody involved  
with the red deer project across the decades, in particular to Ali Morris, Sean Morris,  
Martin Baker, Fiona Guinness, Ian Stevenson, Dan Nussey and Craig Walling. Thanks  
to Lauren Harrison and John Stinchcombe for comments on the manuscript and to  
Jarrod Hadfield for help with selection models.

## References

1. Mc Carty JP. Ecological Consequences of Recent Climate Change. *Conserv Biol.*  
2001;15(2):320–331.

2. Parmesan C. Ecological and Evolutionary Responses to Recent Climate Change. *Annu Rev Ecol Evol Syst.* 2006;37(1):637–669. 638  
639
3. Menzel A, Sparks T, Estrella N, Koch E, Aasa A, Ahas R, et al. European phenological response to climate change matches the warming pattern. *Glob Chang Biol.* 2006;12(10):1969–1976. 640  
641  
642
4. Gardner JL, Peters A, Kearney MR, Joseph L, Heinsohn R. Declining body size: a third universal response to warming? *Trends Ecol Evol.* 2011;26(6):285–291. 643  
644
5. O'Connor MI, Selig ER, Pinsky ML, Altermatt F. Toward a conceptual synthesis for climate change responses. *Glob Ecol Biogeogr.* 2012;21(7):693–703. 645  
646
6. Hairston NG, Ellner SP, Geber Ma, Yoshida T, Fox Ja. Rapid evolution and the convergence of ecological and evolutionary time. *Ecol Lett.* 2005;8(10):1114–1127. 647  
648
7. Pemberton JM. Evolution of quantitative traits in the wild: mind the ecology. *Philos Trans R Soc B.* 2010;365(1552):2431–2438. 649  
650
8. van Benthem KJ, Bruijning M, Bonnet T, Jongejans E, Postma E, Ozgul A. Disentangling evolutionary, plastic and demographic processes underlying trait dynamics: a review of four frameworks. *Methods Ecol Evol.* 2017;8(1):75–85. 651  
652  
653
9. Visser ME. Keeping up with a warming world; assessing the rate of adaptation to climate change. *Proc Roy Soc B.* 2008;275(1635):649–59. 654  
655
10. Ozgul A, Tuljapurkar S, Benton TG, Pemberton JM, Clutton-Brock TH, Coulson T. The dynamics of phenotypic change and the shrinking sheep of St. Kilda. *Science.* 2009;325(5939):464–7. 656  
657  
658
11. Buskirk JV, Steiner UK. The fitness costs of developmental canalization and plasticity. *J Evol Biol.* 2009;22(4):852–860. 659  
660
12. Chevin LM, Lande R, Mace GM. Adaptation, plasticity, and extinction in a changing environment: towards a predictive theory. *PLoS Biol.* 2010;8(4):e1000357. 661  
662  
663
13. Duputié A, Rutschmann A, Ronce O, Chuine I. Phenological plasticity will not help all species adapt to climate change. *Glob Chang Biol.* 2015;21:3062–3073. 664  
665

14. Merilä J, Hendry AP. Climate change, adaptation, and phenotypic plasticity: The problem and the evidence. *Evol Appl.* 2014;7(1):1–14. 666  
667
15. Pelletier F, Coltman DW. Will human influences on evolutionary dynamics in the wild pervade the Anthropocene? *BMC Biology.* 2018;16(1):7. 668  
doi:10.1186/s12915-017-0476-1. 670
16. Franks SJ, Weber JJ, Aitken SN. Evolutionary and plastic responses to climate change in terrestrial plant populations. *Evol Appl.* 2014;7(1):123–139. 671  
672
17. Boutin S, Lane JE. Climate change and mammals: evolutionary versus plastic responses. *Evol Appl.* 2014;7(1):29–41. 673  
674
18. Charmantier A, Gienapp P. Climate change and timing of avian breeding and migration: evolutionary versus plastic changes. *Evol Appl.* 2014;7(1):15–28. 675  
676
19. Crozier LG, Hutchings JA. Plastic and evolutionary responses to climate change in fish. *Evol Appl.* 2014;7(1):68–87. 677  
678
20. Evans SR, Gustafsson L. Climate change upends selection on ornamentation in a wild bird. *Nature Ecol Evol.* 2017;1(2):1–5. 679  
680
21. Kruuk LEB. Estimating genetic parameters in natural populations using the "animal model". *Philos Trans R Soc B.* 2004;359(1446):873–90. 681  
682
22. Postma E. Implications of the difference between true and predicted breeding values for the study of natural selection and micro-evolution. *J Evol Biol.* 2006;19:309–320. 683  
685
23. Hadfield JD, Wilson AJ, Garant D, Sheldon BC, Kruuk LEB. The misuse of BLUP in ecology and evolution. *Am Nat.* 2010;175(1):116–25. 686  
687
24. Gienapp P, Merilä J. Disentangling plastic and genetic changes in body mass of Siberian jays. *J Evo Bio.* 2014; p. 1–10. 688  
689
25. Bonnet T, Wandeler P, Camenisch G, Postma E. Bigger Is Fitter? Quantitative Genetic Decomposition of Selection Reveals an Adaptive Evolutionary Decline of Body Mass in a Wild Rodent Population. *PLoS Biol.* 2017;15(1):e1002592. 690  
691  
692

26. Ramakers JJC, Gienapp P, Visser ME. Phenological mismatch drives selection on elevation, but not on slope, of breeding time plasticity in a wild songbird. *Evolution*. 2019;73(2):175–187.
27. Parmesan C, Yohe G. A globally coherent fingerprint of climate change impacts across natural systems. *Nature*. 2003;421(6918):37–42.
28. Durant JM, Hjermmann DØ, Ottersen G, Stenseth NC. Climate and the match or mismatch between predator requirements and resource availability. *Clim Res*. 2007;33:271–283.
29. Radchuk V, Reed T, Teplitsky C, Pol Mvd, Charmantier A, Hassall C, et al. Adaptive responses of animals to climate change are most likely insufficient. *Nature Communications*. 2019;10(1):1–14.
30. Visser M, Lambrechts MM. Global climate change leads to mistimed avian reproduction. *Adv Ecol Res*. 2004;35:89–110.
31. Potti J. Advanced breeding dates in relation to recent climate warming in a Mediterranean montane population of Blue Tits *Cyanistes caeruleus*. *J Ornithol*. 2009;150(4):893–901.
32. Saccheri I, Hanski I. Natural selection and population dynamics. *Trends Ecol Evol*. 2006;21(6):341–347.
33. Reed TE, Grøtan V, Jenouvrier S, Sæther BE, Visser ME. Population growth in a wild bird is buffered against phenological mismatch. *Science*. 2013;340:488–491.
34. Sheldon BC, Kruuk LEB, Merila J. Natural Selection and Inheritance of Breeding Time and Clutch Size in the Collared Flycatcher. *Evolution*. 2003;57(2):406–420.
35. Teplitsky C, Mills JA, Yarrall JW, Merilä J. Indirect genetic effects in a sex-limited trait: the case of breeding time in red-billed gulls. *J Evol Biol*. 2010;23(5):935–944.
36. Gienapp P, Postma E, Visser ME. Why breeding time has not responded to selection for earlier breeding in a songbird population. *Evolution*. 2006;60(11):2381–2388.

37. Lane JE, McAdam AG, McFarlane SE, Williams CT, Humphries MM, Coltman DW, et al. Phenological shifts in North American red squirrels: disentangling the roles of phenotypic plasticity and microevolution. *J Evol Biol.* 2018;31(6):810–821.
38. van Asch M, Salis L, Holleman LJM, van Lith B, Visser ME. Evolutionary response of the egg hatching date of a herbivorous insect under climate change. *Nature Climate Change.* 2013;3(3):244–248. doi:10.1038/nclimate1717.
39. Moyes K, Nussey DH, Clements MN, Guinness FE, Morris A, Morris S, et al. Advancing breeding phenology in response to environmental change in a wild red deer population. *Glob Chang Biol.* 2011;17:2455–2469.
40. Stoper KV, Bento AI, Clutton-Brock TH, Pemberton JM, Kruuk LEB. Multiple pathways mediate the effects of climate change on maternal reproductive traits in a red deer population. *Ecology.* 2014;95(11):3124–3138.
41. Froy H, Martin J, Walling K, Clutton-Brock TH, Pemberton JM, Kruuk LEB. Consistent within-individual plasticity is sufficient to explain temperature responses in red deer reproductive traits. *J Evol Biol.* 2019;(In press).
42. Clements MN, Clutton-Brock TH, Guinness FE, Pemberton JM, Kruuk LEB. Variances and Covariances of Phenological Traits in a Wild Mammal Population. *Evolution.* 2010;65(3):788–801.
43. Coulson T, Kruuk LEB, Tavecchia G, Pemberton JM. Estimating selection on neonatal traits in red deer using elasticity path analysis. *Evolution.* 2003;57(12):2879–2892.
44. Henderson CR. Estimation of genetic parameters. *Ann Math Stat.* 1950;21:309–310.
45. Lush J. *Animal breeding plans.* Ames, Iowa: Iowa State College Press; 1937.
46. Merilä J, Sheldon BC, Kruuk LEB. Explaining stasis : microevolutionary studies in natural populations. *Genetica.* 2001;112:199–222.
47. Brookfield JFY. Why are estimates of the strength and direction of natural selection from wild populations not congruent with observed rates of phenotypic change? *BioEssays.* 2016;38:1–8.

48. Lande R. Quantitative Genetic Analysis of Multivariate Evolution , Applied to Brain : Body Size Allometry. *Evolution*. 1979;33(1):402–416. 750  
751
49. Clutton-Brock TH, Guinness FE, Albon SD. Red deer: behavior and ecology of two sexes. University of Chicago press; 1982. 752  
753
50. Huisman J, Kruuk LEB, Ellis PA, Clutton-brock T, Pemberton JM. Inbreeding depression across the lifespan in a wild mammal population. *Proc Natl Acad Sci USA*. 2016;113(13):3585–3590. 754  
755  
756
51. Huisman J. Pedigree reconstruction from SNP data: parentage assignment, sibship clustering and beyond. *Mol Ecol Resour*. 2017;17(5):1009–1024. 757  
758
52. Hadfield JD. Estimating evolutionary parameters when viability selection is operating. *Proc Roy Soc B*. 2008;275(1635):723–34. 759  
760
53. Keller L, Waller D. Inbreeding effects in wild populations. *T Ecol Evol*. 2002;17(5):19–23. 761  
762
54. Pemberton JM. Wild pedigrees: the way forward. *Proc Roy Soc B*. 2008;275(1635):613–21. 763  
764
55. Hadfield JD. MCMC Methods for Multi-Response Generalized Linear Mixed Models: The MCMCglmm R Package. *J Stat Soft*. 2010;33(2):1–22. 765  
766
56. Kruuk LEB, Clutton-Brock TH, Albon SD, Pemberton JM, Guinness FE. Population density affects sex ratio variation in red deer. *Nature*. 1999;399(6735):459–461. 767  
768  
769
57. Kruuk LEB, Hadfield JD. How to separate genetic and environmental causes of similarity between relatives. *J Evol Biol*. 2007;20(5):1890–903. 770  
771
58. Nakagawa S, Schielzeth H. A general and simple method for obtaining  $R^2$  from generalized linear mixed-effects models. *Methods in Ecology and Evolution*. 2013;4(2):133–142. 772  
773  
774
59. Morrissey MB, Parker DJ, Korsten P, Pemberton JM, Kruuk LEB, Wilson AJ. The prediction of adaptive evolution: empirical application of the secondary 775  
776



- theorem of selection and comparison to the breeder's equation. *Evolution*. 2012;66(8):2399–2410. 777  
778
60. Lande R, Arnold SJ. The Measurement of Selection on Correlated Characters. *Evolution*. 1983;37(6):1210–1226. 779  
780
61. Bonnet T, Morrissey MB, Kruuk LEB. Estimation of Genetic Variance in Fitness, and Inference of Adaptation, When Fitness Follows a Log-Normal Distribution. *Journal of Heredity*. 2019;110(4):383–395. 781  
782  
783
62. Morrissey MB, Bonnet T. Analogues of the fundamental and secondary theorems of selection, assuming a log-normal distribution of expected fitness. *J Hered*. 2019;110:396–402. 784  
785  
786
63. Stinchcombe JR, Simonsen AK, Blows MW. Estimating uncertainty in multivariate responses to selection. *Evolution*. 2014;68(4):1188–96. 787  
788
64. Hendry AP, Kinnison MT. Perspective: the pace of modern life: measuring rates of contemporary microevolution. *Evolution*. 1999;53(6):1637–1653. 789  
790
65. Pigeon G, Festa-Bianchet M, Coltman DW, Pelletier F. Intense selective hunting leads to artificial evolution in horn size. *Evol Appl*. 2016;9(4):521–530. 791  
792  
793  
doi:10.1111/eva.12358.
66. Robertson A. A mathematical model of the culling process in dairy cattle. *Anim Prod*. 1966;8:95–108. 794  
795
67. Price GR. Selection and covariance. *Nature*. 1970;227:520–521. 796
68. Bijma P. The quantitative genetics of indirect genetic effects: a selective review of modelling issues. *Heredity*. 2014;112(1):61–9. 797  
798
69. Wilson AJ, Coltman DW, Pemberton JM, Overall ADJ, Byrne KA, Kruuk LEB. Maternal genetic effects set the potential for evolution in a free-living vertebrate population. *Journal of Evolutionary Biology*. 2005;18(2):405–414. 799  
800  
801  
802  
doi:10.1111/j.1420-9101.2004.00824.x.

70. Morrissey MB, Kruuk LEB, Wilson aJ. The danger of applying the breeder's  
equation in observational studies of natural populations. *J Evol Biol.*  
2010;23(11):2277–88. 803  
804  
805
71. Grafen A. On the uses of data on lifetime reproductive success. In:  
Clutton-Brock TH, editor. *Reproductive success*. The University of Chicago  
Press; 1988. p. 454–471. 806  
807  
808
72. Sæther Be, Engen S. The concept of fitness in fluctuating environments. *Trends*  
*Ecol Evol.* 2015;30(5):1–9. 809  
810
73. Marrot P, Charmantier A, Blondel J, Garant D. Current spring warming as a  
driver of selection on reproductive timing in a wild passerine. *J Anim Ecol.*  
2018;87(3):754–764. doi:10.1111/1365-2656.12794. 811  
812  
813
74. Coltman DW, O'Donoghue P, Jorgenson JT, Hogg JT, Strobeck C,  
Festa-Bianchet M. Undesirable evolutionary consequences of trophy hunting.  
*Nature.* 2003;426(December):655–658. doi:10.1038/nature02187.1. 814  
815  
816
75. Kardos M, Luikart G, Allendorf FW. Predicting the Evolutionary Effects of  
Hunting Requires an Understanding of genetics. *J Wildlife Manage.*  
2018;82(5):889–891. 817  
818  
819