

Transgenerational plasticity and bet-hedging: a framework for reaction norm evolution.

1 **Jens Joschinski^{1*}, Dries Bonte¹**

2 ¹ Terrestrial Ecology Unit (TEREC), Department of Biology, Ghent University, Ghent, Belgium.

3 *** Correspondence:**

4 Jens Joschinski

5 Jens.Joschinski@ugent.be

6 **Keywords: Phenotypic plasticity, Trans-generational plasticity, Bet-hedging, Coin-flipping,**
7 **GxE, Canalization, Adaptation, Climate change**

8 **Abstract**

9 Decision-making under uncertain conditions favors bet-hedging (avoidance of fitness variance),
10 whereas predictable environments favor phenotypic plasticity. However, entirely predictable or
11 entirely unpredictable conditions are rarely found in nature. Intermediate strategies are required when
12 the time lag between information sensing and phenotype induction is large (e.g. transgenerational
13 plasticity) and when cues are only partially predictive of future conditions. Nevertheless, current theory
14 regards plasticity and bet-hedging as distinct entities. We here develop a unifying framework: based
15 on traits with binary outcomes like seed germination or diapause incidence we clarify that diversified
16 bet-hedging (risk-spreading among one's offspring) and transgenerational plasticity are mutually
17 exclusive strategies, arising from opposing changes in reaction norms (allocating phenotypic variance
18 among or within environments). We further explain the relationship of this continuum with arithmetic
19 mean maximization vs. conservative bet-hedging (a risk-avoidance strategy), and canalization vs.
20 phenotypic variance in a three-dimensional continuum of reaction norm evolution. We discuss under
21 which scenarios costs and limits may constrain the evolution of reaction norm shapes.

22 **1 Introduction**

23 Changing conditions can promote evolutionary change in various ways (Botero et al., 2015; Tufto,
24 2015). One commonly envisioned mode of evolution is the continuous change of trait means as result
25 of changing mean conditions (Darwin, 1859). Yet, although trait changes in response to novel
26 conditions are widely observed (e.g. due to climate change, Piao et al., 2019), they frequently result
27 from phenotypic plasticity (Boutin and Lane, 2014), i.e. changes of the phenotype in response to an
28 environmental cue. Phenotypic plasticity may provide a short-term relief from changing conditions
29 (Charmantier et al., 2008; Chevin et al., 2010), but also shield a genotype from selection and thereby
30 prevent evolution (Oostra et al., 2018), or it may facilitate evolution via genetic accommodation (Kelly,
31 2019). In any case, phenotypic plasticity is a pervasive evolutionary strategy, and considered a major
32 factor in a rapidly changing climate (Fox et al., 2019).

33 The time scale of phenotypic change depends on the time scale of environmental fluctuation (Rando
34 and Verstrepen, 2007; Stomp et al., 2008). Fluctuations over very rapid timescales can be addressed
35 by reversible plasticity, which includes, for example, the induction of plant defense when herbivores
36 are present (Green and Ryan, 1972). Gradual long-term changes, on the other hand, are addressed by
37 genetic adaptation. Between those extremes lie environmental fluctuations that are roughly on the scale
38 of one life span. When environments change over the course of an organism's development, they can
39 be tackled by irreversible developmental plasticity, i.e. plastic adjustment of developmental pathways
40 that lead to alternative phenotypes (Botero et al., 2015). For example, some *Daphnia* can produce
41 protective phenotypes when chemical cues from predators are sensed during development (Krueger
42 and Dodson, 1981). When environments are constant throughout an organism's life time but change
43 from one generation to the next, phenotypic change can be induced in the offspring generation. These
44 are referred to as anticipatory parental effects (Burgess and Marshall, 2014) or intergenerational
45 inheritance (Perez and Lehner, 2019). For example, aphids that live under crowded conditions may
46 produce winged offspring that can leave the colony and avoid high predation pressure or plant
47 deterioration (Braendle et al., 2006). Lastly, when environmental fluctuations last for several
48 generations, epigenetic modifications may be integrated into the germ line and affect multiple
49 succeeding generations. This is referred to as transgenerational plasticity or non-genetic inheritance
50 (Perez and Lehner, 2019; Adrian-Kalchhauser et al., 2020). For the remainder of the article we will
51 refer to all these irreversible changes simply as phenotypic plasticity, ignoring the potential
52 physiological constrains that may limit their evolution. They all have in common that there is a long
53 delay between information sensing and phenotype induction.

54 Although often assumed, phenotypic plasticity does not need to be adaptive (Ghalambor et al., 2007;
55 Arnold et al., 2019). Plasticity requires some environmental cue on which the induction of phenotypic
56 change is based, and uncertainty around the future environmental state may turn plasticity maladaptive
57 (Burgess and Marshall, 2014; Donelson et al., 2018). Such unpredictable conditions instead favor bet-
58 hedging, which refers to the reduction of fitness variance (Cohen, 1966; Seger and Brockmann, 1987;
59 Starrfelt and Kokko, 2012). Bet-hedging can be achieved by avoiding risky investments (conservative
60 bet-hedging), or by spreading the risk among one's offspring (diversified bet-hedging), i.e. producing
61 offspring with varying phenotypes (Seger and Brockmann, 1987; Starrfelt and Kokko, 2012). Although
62 empirical evidence is difficult to obtain (Simons, 2011), bet-hedging is a likely explanation for high
63 trait variance or unexpected trait means in many systems, such as the seed dormancy of desert annuals
64 (Cohen, 1966), diapausing strategies of insects (Hopper, 1999) and annual killifish (Furness et al.,
65 2015), wing dimorphisms (Grantham et al., 2016), facultative sexual reproduction (Gerber and Kokko,
66 2018), dispersal and partial migration (Goossens et al., 2020).

67 At fluctuations of intermediate time scales where there is a delay between information sensing and
68 phenotype induction, both phenotypic plasticity (e.g. Baker et al., 2019) and bet-hedging (e.g. Venable,
69 2007) may be expected to evolve. Various theoretical studies have clarified the conditions that may
70 lead to one or the other (Botero et al., 2015; Tufto, 2015), but although occurring potentially
71 simultaneously, bet-hedging and plasticity are nevertheless often treated independently (Donelson et
72 al., 2018). Moreover, when diversified bet-hedging and plasticity are considered jointly, there is no
73 clear consensus about their exact relationship. Adaptive offspring variance that is needed for diversified
74 bet-hedging might be either established by developmental instability (Simons and Johnston, 1997;
75 Kærn et al., 2005; Veening et al., 2008; Woods, 2014; Dueck et al., 2016; Perrin, 2016) or by overly
76 relying on cues with little predictive power ("microplasticity", Simons and Johnston, 2006;
77 "hyperplasticity", Scheiner and Holt, 2012). With this article we aim to clarify the relationship between
78 bet-hedging and plasticity, with special attention to readers that are familiar with plasticity but less
79 familiar with bet-hedging theory. We will first use one simple numerical example (insect diapause) to
80 explain the relationship of diversified bet-hedging, conservative bet-hedging and arithmetic mean
81 maximization in detail. We will then extend the consideration to a range of environments whose state
82 is partially predictable, thereby adding the potential for phenotypic plasticity. Lastly, we generalize
83 from our example and describe a method to quantify phenotypic plasticity and bet-hedging based on
84 reaction norm shapes.

85 2 **An example**

86 Common examples of bet-hedging are transgenerational biphenisms, i.e. the parent decides among two
87 possible physiological states of the offspring in the face of uncertainty (e.g. Cohen, 1966; Grantham et
88 al., 2016; Maxwell and Magwene, 2017; see Simons, 2011 for further examples). One of these
89 examples is the timing of insect diapause (Halkett et al., 2004; Péliesson et al., 2013), which we will
90 use to illustrate the theory throughout this article.

91 Multivoltine insects benefit from exponential population growth throughout the growing season, but
92 need to produce an overwintering (diapausing) generation before the onset of cold weather (Kivela et
93 al., 2016). Aphids, for example, reproduce by parthenogenesis during summer, which enables
94 particularly quick population growth; in autumn they invest in sexual offspring that produce diapausing
95 eggs, as frost kills the soft-bodied insects and only eggs survive (Simon et al., 2002). The struggle to
96 keep the growing season long on one hand and to avoid death on the other hand puts diapause timing
97 under intense selection pressure. If the onset of frost would be invariant, day length could be used as
98 reliable cue of impending winter, so plasticity in response to day length is expected to evolve. However,
99 if just one generation faces early frosts, all offspring may simultaneously die and the genotype is driven
100 to extinction, regardless of their otherwise high growth rates. Under unpredictable or only partially
101 predictable conditions, bet-hedging strategies may therefore be expected to evolve (Halkett et al.,
102 2004).

103 For the remainder of this article we will use examples that are loosely based on aphid overwintering.
104 We will assume that parthenogenetic offspring (P_1) may produce four offspring when environmental
105 conditions are mild, but face a 90% mortality rate when conditions change. In contrast, diapausing
106 offspring (P_2) only replace themselves with 1 offspring in either environment. Hence we assign
107 phenotype P_1 a fitness value of 4 in E_1 (summer), but only 0.1 in E_2 (winter), whereas phenotype P_2
108 achieves 1 fitness in either environment. We assume that the evolution of these growth rates is
109 constrained, so only the proportion of each phenotype may evolve.

110

141 plays it safe (Starrfelt and Kokko, 2012). This strategy is similar to investing in a portfolio of stocks
142 rather than a single stock and is called diversified bet-hedging (DBH).

143 The geometric mean can be calculated for any phenotype proportion p (proportion of P_2) between 0
144 and 100% (Fig.1A, solid blue line), showing that actually neither of the three strategies (AMM, CBH,
145 DBH) is optimal. Instead, $p = 0.61$, i.e. a mix of CBH and DBH, yields the highest geometric mean
146 fitness (Table 1). Starrfelt and Kokko (2012) explored the relationship among AMM, CBH and DBH
147 in great detail, and explained fitness optimization as a three-way trade-off between maximizing the
148 arithmetic mean, reducing fitness variance, and reducing fitness correlation among the offspring.
149 However, as outlined in our example, this three-way relationship breaks down to a simple linear
150 gradient when there are exactly two phenotypes to choose from.

151 The same principles also apply when the two environments do not occur with equal frequency, e.g.
152 when the probability of E_2 (winter) is reduced to 20%. In this case the arithmetic mean fitness of P_1
153 and P_2 needs to be weighted by the frequencies of E_1 and E_2 . Nevertheless, arithmetic mean fitness is
154 still a linear function of the phenotype proportion p (Fig. 1A, dashed orange line), and increasing the
155 proportion of P_2 constitutes a change from AMM towards DBH or CBH. In this example with only
156 occasionally adverse conditions, the optimum lies at $p = 0.17$ (solid orange line), i.e. much closer to an
157 AMM strategy. If the frequency of E_2 is raised to 70%, on the other hand, the optimal strategy moves
158 with $p = 0.90$ close to pure CBH (not shown). The optimal strategy thus strongly depends on the
159 environmental frequency.

160 We wish to complete this description of fitness maximization in a single environment with two last
161 special cases. First, we consider the production of two specialist phenotypes, in which P_1 achieves a
162 fitness of 4 in E_1 , but none in E_2 , while P_2 achieves 0 fitness in E_1 but 4 fitness in E_2 (thus deviating
163 from the aphid example). With these parameters geometric mean fitness peaks at $p = 0.5$ (Fig. 1B, blue
164 solid line), so a strategy that maximizes developmental instability is optimal. Yet, the mixed production
165 of offspring does not constitute DBH, because the diversification does not come at the cost of
166 arithmetic mean fitness (i.e. the dashed blue line is flat). If, however, the growth rates of the two
167 phenotypes are slightly uneven, e.g. reduced to 3.9 for P_2 in E_2 , the same investment in P_2 would lower
168 arithmetic mean fitness (dotted grey lines), and hence technically classify as a diversified bet-hedging
169 strategy. This borderline example shows that the classification of bet-hedging strategies is not only a
170 question of whether arithmetic mean fitness is reduced, but rather by how much. The second special
171 case concerns very high probabilities of adverse conditions. When the frequency of E_2 is raised to 0.9,
172 it carries so much weight that the arithmetic mean fitness does not decrease, but increase with the

173 proportion of P_2 (Fig. 1B, dashed orange line). The strategy that avoids variance is hence also the one
174 which maximizes arithmetic mean fitness, so increasing geometric mean fitness (solid orange line)
175 does not come at the cost of arithmetic mean fitness and CBH becomes impossible. In general, the
176 linear gradient from AMM over DBH to CBH (and, in fact, the occurrence of bet-hedging) breaks
177 down, when there is no conflict between arithmetic mean maximization and reduction of fitness
178 variance. We will avoid these special situations in the remainder of the article.

179 4 Calculating optimal reaction norm shapes

180 We so far discussed the optimal phenotype proportion in a single, isolated environment. However, the
181 benefit of diapause lies in adapting to a continually changing environment. Like in many other insects,
182 aphid diapause is mainly governed by night length. Aphids exclusively reproduce by parthenogenesis
183 under long-day (short night) conditions, but transition to the production of sexual forms under long-
184 night conditions (Marcovitch, 1923). The diapause decision can hence be visualized as a biphenic
185 reaction norm, in which the x-axis represents a continuous night length and the y-axis represents a
186 probability (or, from the mother's perspective, a proportion) of diapause induction between 0 and
187 100%. This reaction norm to night length generally follows a logit-curve that ranges from a probability
188 of zero under short nights to a probability of 1 under long nights, and the inflection point at which half
189 of the offspring are diapausing forms is called critical day length (Danilevskii, 1965). The night length
190 response is additionally modulated by temperature (warm temperatures delay diapause), but we ignore
191 the additional plasticity in response to temperature in our considerations.

192 We will now use the diapause example to illustrate how to calculate optimal reaction norm shapes.
193 Imagine an environment in which winter onsets over many years always occur at 14 h night length.
194 Obviously night length would be a reliable cue and plasticity in response to night length can be
195 expected to evolve. Conversely, night length is useless as cue for a plastic response if winter onset
196 fluctuates randomly. Between those extremes lies an only partially reliable cue, i.e. there is between-
197 years variation in the relationship of night length and winter onset. For example, winter onset may in
198 some years coincide with a night length of 14 h, but fall in other years on an earlier (13.8 h) or later
199 (14.5 h) date, which can be described by a normal distribution with a mean of 14 h and some standard
200 deviation. We now use three different scenarios of how environmental conditions (winter onset) may
201 vary: 1) Winter onset fluctuates according to a normal distribution $N_1(14, 1)$ with a mean cue value of
202 14 h and standard deviation 1; 2) Winter onset follows a normal distribution $N_2(14, 4)$ with a mean cue
203 value of 14 h and standard deviation 4, thus simulating lower predictability by night length; 3) Winter

204 onset fluctuates according to a normal distribution $N_3(14, 2)$ with standard deviation 2, but half of the
205 winters are mild enough that offspring of type P_1 (e.g. parthenogenetic offspring) can survive.

206 The cumulative distribution function of N describes the probability that winter will occur at a night
207 length of c or lower (Fig. 2A). If, for example, an aphid lives in an environment of exactly 14 hours
208 night length, it can expect that the offspring will experience winter conditions with a 50% probability
209 (the optimal phenotype proportion is then 0.61, see table 1). At 13 hours night length winter onset is
210 less probable (18%) for environment N_1 (blue line) than for N_2 (41%, orange line), because winter onset
211 variability is lower. In N_3 the probability distribution must be multiplied by 0.5, i.e. with the chance
212 that winter is mild (green line). This reduces the probability of winter onset at $c = 13$ h to 16%. Given
213 these environmental frequencies and the fitness values introduced earlier (parthenogenesis: 4/0.1;
214 diapause: 1/1; in summer/winter conditions, respectively), one can now calculate the optimal
215 proportion p as described in section 3. This proportion is 0.47 (nearly pure DBH) in scenario 1, as there
216 is considerable risk of unfavorable conditions, but in scenarios 2 and 3 the ratios drop to 0.12 and 0.11,
217 respectively. Thus, DBH is favored over pure AMM with increasing probability of winter conditions.
218 The same calculations can be performed along the whole range of c , so the complete optimal reaction
219 norm can be calculated if mean and standard deviation of the environment-cue relationship are known
220 (Fig. 2B, C).

221 With these considerations we explained the reaction norm shape as a series of binary decisions. In each
222 of these decisions, phenotype proportions may range from AMM to CBH, with DBH in between. The
223 overall degree of bet-hedging is hence defined by the reaction norm shape, and in our specific examples
224 mostly correlates with the reaction norm slope (Fig. 2B, orange and blue lines) and range (green line).
225 However, as indicated by the skew in the orange line towards the lower range of c (AMM is
226 discouraged even under low risk) in Fig. 2C, more complex shapes are also possible and the relative
227 contribution of each strategy is difficult to quantify. Furthermore, our examples are based on
228 cumulative densities of normal distributions, but depending on the environmental cue, other shapes
229 (e.g. bimodal, sinusoid) are possible. We hence require summary statistics that adequately describe the
230 reaction norm shape.

231

232 5 Classification of reaction norm shapes

233 In this section we will describe some typical reaction norm shapes and discuss useful summary
 234 statistics to describe the overall degree of plasticity, arithmetic mean maximization, conservative bet-
 235 hedging and diversified bet-hedging. First, let us assume a “plastic” reaction norm (Fig. 3A, dark blue
 236 line). A step function describes a sudden switch from one phenotype (AMM) to the other (CBH), and
 237 the number of environments in which a mix of phenotypes is produced is minimized. This function
 238 maximizes the standard deviation of phenotype proportions p across environments. We refer to the
 239 variance of p as σ_{among}^2 . The opposite of a step function is one in which the mother’s decision is
 240 entirely independent of the environmental cue, i.e. left to developmental instability, and both
 241 phenotypes are produced in equal measure (DBH; Fig. 3A, light blue line). While σ_{among}^2 is zero, there
 242 is variance in phenotypes within each environment (σ_{within}^2). The trait choice is a Bernoulli draw and
 243 the variance of each p is calculated as $p * (1 - p)$, so we define σ_{within}^2 across environments as the mean
 244 Bernoulli variance. The two variance components (among and within environments) complement each
 245 other, and we define their sum $s = \sigma_{among}^2 + \sigma_{within}^2$ as the phenotypic variance of the genotype. It
 246 is not possible to maximize both σ_{among}^2 (steep slope, high range) and σ_{within}^2 (minimal departure from
 247 50%) at once, but intermediate reaction norms with mixed contributions of σ_{among}^2 and σ_{within}^2 are
 248 possible (solid and dashed medium blue lines). The trade-off between σ_{among}^2 and σ_{within}^2 can be
 249 described by the ratio $r = \frac{\sigma_{among}^2}{\sigma_{within}^2}$. r thus describes the degree of developmental (in)stability across
 250 environments.

251 The variance composition is not the only parameter in which reaction norms may vary. Reaction norms
 252 may, for example, be flat ($r = 0$), but the proportion of P_2 (p) might be zero (Fig. 3B, light orange line),
 253 0.8 (dark orange) or 1 (darkest line) in all environments. These reaction norms differ in the mean
 254 frequency of phenotype P_2 across environments, which we denote as f . A frequency of zero indicates
 255 a pure AMM strategy, while $f = 1$ is a pure CBH strategy. A mean frequency of 0.5 indicates a reaction
 256 norm with maximal phenotypic variance (s), enabling the aforementioned gradient from phenotypic
 257 plasticity to DBH (Fig. 3A, solid lines). As with Fig. 3A, intermediate reaction norm shapes are also
 258 possible: a reaction norm may, for example, range from $p = 0$ to $p = 0.3$ or from $p = 0.7$ to $p = 1$ (Fig
 259 3B, dashed lines). Reaction norms can thus vary from complete canalization to high phenotypic
 260 variance, and we express their shape by mean frequency of phenotype P_2 and by the variance
 261 composition. A canalized reaction norm may be only expressing risk-averse phenotypes, or only

262 expressing arithmetic mean optimizers, whereas high phenotypic variance may indicate steep plastic
263 reaction norms or DBH.

264 The two shape parameters f and r reflect the reaction norm shape to a reasonable extent, but as
265 summarizing statistics they cannot sufficiently describe all its features. For example, the reaction norms
266 in Fig. 3C both share the same mean frequency (0.5) and variance composition (0.47), but the strategies
267 under environments that correspond to a low cue c differ considerably. In our aphid example these two
268 strategies differ in the mean timing of diapause induction, which is an important consideration when
269 the onset of seasons is under directional change (IPCC, 2014). This mean timing can be assessed by
270 calculating the inflection point (called critical day length for diapause reaction norms), but for non-
271 logistic reaction norms or more complicated reaction norm shapes a different approach, e.g. based on
272 autocorrelation patterns, is required.

273 In summary we discussed three important parameters that describe a reaction norm shape: The
274 frequency f , the variance composition r (among:within environments), and (for logistic reaction norms)
275 the inflection points. These three parameters are partially interdependent of one another, and can be
276 drawn as three perpendicular axes (Fig. 3D; see also supplementary figure S1 for an alternative
277 representation). The resulting parameter space has three distinct ends which conform to maximum
278 plasticity (i.e. a step-function, dark blue dot), CBH (dark orange), and AMM (light orange). Parameters
279 outside these bounds are not possible, e.g. DBH and plasticity cannot occur in canalized reaction
280 norms, and on the other hand mean frequencies of 0.5 necessarily imply phenotypic variance by DBH
281 or plasticity.

282

283 6 Reaction norm evolution

284 So far we described optimal strategies in a single environment (Section 3), calculated optimal reaction
285 norm shapes (Section 4), and explored which reaction norm shapes are generally possible (Section 5).
286 We now return to our aphid diapause example to illustrate how optimal reaction norms change when
287 environmental conditions and fitness functions are altered. We will cover cases with more frost-
288 resistant parthenogenetic forms (i.e., higher fitness of P_1 in E_2), harsher summer environments (lower
289 fitness of P_1 in E_1), and three forms of change in the environment that are directly relevant for aphid
290 biology: first, mean winter onset may vary with latitude, with earlier winter onset at high latitudes
291 (Danilevskii, 1965). Secondly, winter onset dates may vary among years, which is the condition that
292 should lead to bet-hedging in diapause timing (Halkett et al., 2004). Lastly, aphid populations in

293 warmer climates frequently lost the ability to produce sexual forms and reproduce by parthenogenesis
 294 throughout the year (anholocyclic life cycles, Simon et al., 2002). The preparation for winter makes
 295 only sense if there is sufficient change in environmental conditions, so this kind of canalization
 296 (obligate development) is expected at southern latitudes.

297 We start with environments that vary in among-years predictability. Using night length (in hours) as a
 298 cue c , we consider scenarios where winter onset is normally distributed with a mean cue c of 14 and
 299 standard deviations ranging from 0 to 10. In our standard example with growth rates of 4/0.1
 300 (parthenogenetic) and 1/1 (diapausing), the optimal mean frequency f of risk-averse (diapausing)
 301 phenotypes increases with environmental variance (Fig. 4A, blue solid line), while the variance ratio r
 302 (among : within environments) decreases (Fig. 4C, blue solid line). Thus, a greater tendency towards
 303 DBH and CBH is expected to evolve across environments in unpredictable conditions (see also Fig.
 304 4B, blue lines). With decreasing growth rate of P_1 in E_1 (parthenogenesis in summer) the optimal ratio
 305 decreases less sharply and the frequency of P_2 (diapause) increases more strongly (solid orange and
 306 green lines in Fig. 4A, green lines in Fig. 4B). Here the riskier strategy pays off less, and the balance
 307 is shifted towards CBH. When the growth rate of P_1 in E_2 (winter) is raised to 0.33 (frost tolerance)
 308 both r and f change less steeply with environmental unpredictability (dashed lines), i.e. the optimal
 309 reaction norms tend towards AMM. Increasing the growth rate in E_2 further to 0.66 (dash-dotted lines)
 310 leads to a strategy that ignores environmental risk, except when the chance of mild (summer) conditions
 311 is very low. The range of environments that feature a sufficiently low chance of P_1 decreases with
 312 increasing environmental variance, causing a drop of both f and r as a sign of canalization to AMM
 313 (Fig. 4D). Overall, both CBH and DBH can be expected under unpredictable conditions, but their
 314 relative benefits vary depending on the arithmetic mean fitness of risk-averse and risk-prone
 315 phenotypes.

316 We now simulate global changes in the probability of events, for instance increased or decreased
 317 probabilities of severe winters. For the latter, we multiply the normal distribution by 0.5, overall
 318 halving the probability of being in the harsh environment E_2 (see also Fig. 2A). This discourages risk-
 319 aversion and, for example, having all offspring diapausing is no longer beneficial (Fig. 5). When the
 320 growth rate of P_1 is either 4 (summer) or 0.1 (winter), the frequency f stagnates at 0.2 to 0.25, while
 321 the ratio r decreases from 0.47 to 0.17 (Fig. 5A and C, solid blue line). This is because the reaction
 322 norm range is constrained (Fig. 5B). A lower growth rate of P_1 in E_1 restores phenotypic variance (Fig.
 323 5A and C, orange and green lines), as it reduces its arithmetic mean fitness and makes the alternative

324 phenotype again more profitable (Fig. 5B, green lines). Lowering the environmental risk further
325 increases the benefit of arithmetic mean maximization (dashed lines) and eventually leads to AMM
326 under all environmental conditions (dash-dotted lines). Overall, Fig. 5 shows that a global reduction of
327 the probability for E_2 may discourage CBH, and instead favor AMM. For example, a lower risk of
328 freezing in winter may explain the existence of anholocyclic lines.

329 A third axis of environmental variation concerns changes in mean environments. Moving the
330 distribution of environments to a mean c of 9 h simulates the change of winter onset with latitude, as
331 well as the effects of a changing climate. Although highly relevant for the optimization of fitness, the
332 changes in optimal reaction norm shapes are trivial to describe. We refer to supplementary Material S2
333 for further exploration.

334 In general, we find that r and f evolve with changes in environmental predictability (Fig. 4, solid lines),
335 leading to CBH and DBH in unpredictable environments. Changes in the fitness function (growth rates
336 in our example) may, however, affect the balance of AMM and CBH, and very low rewards for CBH
337 instead lead to the evolution of risky strategies that seek to maximize the arithmetic mean (Fig. 4, dash-
338 dotted lines). When the probability of adverse conditions is globally lowered across the range of
339 environments (e.g. mild winters), the reaction norm range can become constricted, which further
340 affects the balance of the fitness maximization strategies. Lastly, f additionally depends strongly on
341 the mean environment (e.g. winter onset, Supp. S2), but within reasonable limits the general shape of
342 the reaction norms is not affected.

343

344 7 Discussion

345 Phenotypic plasticity can help organisms adapt to changing conditions (Fox et al., 2019), but this
346 requires a predictable cue (Bonamour et al., 2019). Especially for transgenerational plasticity cues are
347 not entirely predictable (Burgess and Marshall, 2014; Donelson et al., 2018), which, at least under
348 some conditions, favors bet-hedging instead (Botero et al., 2015; Tufto, 2015). Nevertheless, the value
349 of bet-hedging strategies as alternatives to plasticity is frequently overlooked.

350 Starrfelt and Kokko (2012) have explained bet-hedging, including its mathematical foundation, in great
351 detail. The main finding was that arithmetic mean fitness maximization, diversified bet-hedging and
352 conservative bet-hedging form a three-way trade-off of conflicting strategies. However, it was difficult
353 to see how these strategies play out in practice (Haaland et al., 2020). We provided a simple, detailed
354 calculation of fitness based on insect diapause as example. Based on this system with only two possible

355 phenotypes (biphenisms) we described how a conflict between arithmetic and geometric mean
 356 optimization can result in bet-hedging (Fig. 1A, B). We explained that the three strategies form a
 357 gradient, in which arithmetic mean maximization (AMM) and conservative bet-hedging (CBH) are
 358 represented by distinct phenotypes, and diversified bet-hedging (DBH) by a mixture of the two
 359 extremes. We also extended the concept by adding a cue the organisms can respond to, thereby
 360 incorporating reaction norms and the potential for phenotypic plasticity. We identified the mean
 361 phenotype frequency f and the variance composition r as two summary statistics of reaction norms that
 362 allow distinguishing between AMM, CBH, DBH and plasticity, and the sum s of the variance
 363 components as a measure of phenotypic variance. Moreover, for logistic reaction norm shapes we
 364 discuss the inflection point as a third useful summary statistic.

365 *Arithmetic mean maximization vs. conservative bet-hedging*

366 In section 3 we described AMM, DBH and CBH as a linear gradient of strategies to cope with a single
 367 environment. When extended to multiple environments, a flat reaction norm at $p = 0$ (Fig. 3B, light
 368 orange line) maximises arithmetic mean fitness (see also Fig. 1A), and any adaptive deviation from
 369 this line incorporates some bet-hedging (in the cases we consider; see Fig. 1B for exceptions). Thus,
 370 the mean phenotype frequency f is a direct measure of the degree of CBH in a reaction norm shape.
 371 We illustrated that f correlates with the frequency of the harsh environment E_2 (compare fig. 4A and
 372 S1, panel A), but f also changes with the degree of environmental variance: higher environmental risk
 373 shifts optimal reaction norms towards DBH and CBH (Fig. 4A, solid lines; Fig. 4B, dark blue vs. light
 374 blue lines), in line with expectations from other studies (Simons, 2011; Tufto, 2015). This shift is
 375 particularly noticeable when the potential fitness gain from a risk-prone strategy is low (Fig. 4B green
 376 lines; Fig. 2C, orange lines). If, on the other hand, the risk is reduced and the potential pay-off high
 377 (Fig. 4A, dashed and dot-dashed lines; Fig. 5), the optimal reaction norm shapes are shifted towards
 378 risk-prone (AMM) strategies (Halkett et al., 2004). Thus our framework made clear that arithmetic
 379 mean maximization and variance avoidance form exact opposites on a gradient of strategies that is
 380 reflected by f (Fig. 3D, y-axis).

381 We have illustrated that frequencies or means of reaction norms that mismatch with environmental
 382 means might serve a function. Recent climate change imposes novel environmental conditions, and
 383 species or populations whose trait means do not evolve in concert with environmental means are often
 384 considered as under risk (e.g. Charmantier and Gienapp, 2014), ignoring that this phenotype-
 385 environment mismatch may in fact be due to an adaptive CBH strategy. This is not to say that CBH

386 can be invoked whenever environmental variance is observed (Simons, 2011), but any combination of
387 mean maximization and variance avoidance (f) has the potential to be adaptive depending on life
388 history and environmental variance.

389 *Phenotypic plasticity vs diversified bet-hedging*

390 Reaction norms that are not entirely canalized exhibit some degree of phenotypic plasticity and/or
391 diversified bet-hedging (Fig. 3 A,B,C), and we expressed their relative contribution with the variance
392 ratio r . When environmental cues convey reliable information, a high r is adaptive, i.e. phenotypes
393 change with the environmental cues, but vary only little for any given cue (solid dark blue lines in Fig.
394 2B, 3A and 4B)(Botero et al., 2015; Tufto, 2015). This reaction norm pattern is commonly referred to
395 as phenotypic plasticity, or, when the offspring phenotype is dictated by the (grand-) parental
396 environment, as inter- or transgenerational plasticity (Perez and Lehner, 2019). A low r , on the other
397 hand, corresponds to DBH across the range of possible environments (orange line in Fig. 2B, solid
398 light blue lines in Fig. 3A and 4B), and occurs predominantly when cues convey little information
399 about the optimal phenotype (Cohen, 1966). Our simple models based on aphid diapause illustrate such
400 a negative relationship between r and cue variance for all but the most extreme growth rate functions
401 (Fig. 4C, 5C). We therefore see phenotypic plasticity and diversified bet-hedging as a continuum of
402 evolutionary strategies that is based on the reaction norm shape (Fig. 3D, x-axis).

403 This definition extends classical concepts of bet-hedging and transgenerational plasticity. Plasticity has
404 a long history of being related to reaction norm shapes (Woltereck, 1913; Bradshaw, 1965), but
405 diversified bet-hedging is not as easily visualized, nor is the relationship with plasticity entirely clear.
406 On the one hand, developmental instability has been seen as a cause of diversified bet-hedging (Simons
407 and Johnston, 1997; Kærn et al., 2005; Woods, 2014; Dueck et al., 2016; Perrin, 2016). Low copy
408 numbers e.g. of transcriptional regulators (Volfson et al., 2006) cause sampling errors that ultimately
409 lead to expression of alternative phenotypes. On the other hand, DBH might be produced by a reaction
410 norm to noise (“microplasticity”, Simons and Johnston, 2006; “hyperplasticity”, Scheiner and Holt,
411 2012). For example, Maxwell and Magwene (2017) engineered a yeast model that evolved a response
412 to estradiol, a compound that was entirely unrelated to fitness but ensured phenotypic variance in a
413 fluctuating environment. Accordingly, the relationship between diversified bet-hedging and plasticity
414 might be perceived as nested or as one of two competing strategies. We instead distinguish them as the
415 two extremes on a continuum of strategies, that correspond to a continuum of reaction norm shapes.

416

417 *Fixed vs. flexible development*

418 The phenotype frequency f and the variance composition r are not entirely independent (Fig. 3D),
419 because phenotypic variance s , i.e. the sum of variance among and within environments, is a quadratic
420 function of f : when f is zero (pure AMM or CBH, Fig. 3B) there is no phenotypic variance and hence
421 no potential for DBH or phenotypic plasticity. When f is 0.5, on the other hand, DBH, phenotypic
422 plasticity, or a mix of the two strategies is necessarily required (Fig. 3A, D).

423 In section 6 we altered the amplitude between summer and winter conditions, both by changing the
424 fitness of the phenotypes (Fig. 4, green and orange lines) and by affecting the global probability of E_2
425 (Fig. 5). Reductions in the difference between summer and winter led to a reduction of phenotypic
426 variance, i.e. to a decrease in f towards canalization (Fig. 4A, Fig. 5 B, D), illustrating that phenotypic
427 variance is not beneficial when environments are stable. The relationship between the variance
428 composition r and environmental variance was, however, maintained (Fig. 5C, dark vs. light lines in
429 Fig. 5B). The benefits of plasticity and DBH under predictable and unpredictable conditions,
430 respectively, were thus also apparent under partially canalizing conditions.

431 Our examples clarified that phenotypic variance is a function of f in binomial reaction norms, and as
432 such it is equally related to both phenotypic plasticity and diversified bet-hedging. The opposite of
433 phenotypic variance (i.e., of plasticity and DBH) in our models is environmental canalization, a term
434 which so far has been used ambiguously (Debat and David, 2001), as it was considered either the
435 opposite of plasticity (Waddington, 1942; Van Buskirk and Steiner, 2009) or of developmental noise
436 (Gibson and Wagner, 2000; Zhang and Hill, 2005) alone. Phenotypic plasticity is regarded an essential
437 component of climate change adaptation (Fox et al., 2019), precisely because of the variance it entails;
438 moreover, de-canalization by phenotypic plasticity may accelerate evolution through genetic
439 accommodation (Kelly, 2019). We argue that the same mechanisms may apply for all modes of
440 phenotypic variance, including diversified bet-hedging.

441 *The importance of mean timing*

442 We introduced the inflection point as additional important reaction norm shape parameter (Fig. 3C, z -
443 axis in Fig. 3D; Fig. S1). In our example the inflection point determined the mean timing of phenotypic
444 change (i.e. the phenology), and clearly depended on the mean timing of environmental change (Fig.
445 S2). The inflection point (called critical day length in diapause reaction norms) is known to change
446 with latitude (Danilevskii, 1965; Bradshaw, 1976), and questions regarding its evolution are highly

447 important under climate change (Saikkonen et al., 2012; Zohner et al., 2016). While limited to logistic
448 reaction norms, we think the inflection point as reaction norm shape parameter deserves special
449 attention, because many phenological traits are of binary nature (e.g. bird arrival, migration onset, plant
450 germination and flowering) and hence modelled as logistic reaction norms.

451 *Outlook*

452 The world is simultaneously changing in climate means, variability and predictability (IPCC, 2014;
453 Lenton et al., 2017; Bathiany et al., 2018), and there are many phenomenological studies on responses
454 to climate change (Parmesan and Yohe, 2003; Badeck et al., 2004; Cohen et al., 2018). However, only
455 few detailed case-studies on the mechanisms of adaptation (Nussey et al., 2005; Gienapp et al., 2013;
456 Lane et al., 2018) exist, and one cannot assume that a matching mean timing or a high level of plasticity
457 is always adaptive (Boutin and Lane, 2014), just like one cannot assume CBH or DBH to be an optimal
458 solution (Simons, 2011) – but one can analyse reaction norm shapes with the proposed shape
459 parameters to decide whether it has the *potential* for adaptive tracking, arithmetic mean maximization,
460 plasticity, bet-hedging or canalization (Joschinski and Bonte, 2020).

461 There is ample room to extend our framework. First of all, we focussed only on the *optimal* reaction
462 norm shape. This ignores that CBH and DBH are often nearly equally suited strategies to cope with
463 environmental uncertainty (Starrfelt and Kokko, 2012), i.e. the shape and curvature of the geometric
464 mean fitness curve (Fig. 1 A) requires further consideration. Secondly, we have restricted our
465 arguments to binary trans-generationally inherited traits, as these are commonly treated both
466 empirically (Venable, 2007; Maxwell and Magwene, 2017; Scholl et al., 2020) and theoretically
467 (Cohen, 1966; Halkett et al., 2004; Starrfelt and Kokko, 2012; Kivela et al., 2016; Gerber and Kokko,
468 2018). For continuous traits, e.g. offspring size (Marshall et al., 2008), our calculations may not apply,
469 because AMM, DBH and CBH need not lie on a linear gradient (i.e. intermediate trait values need not
470 incur highest trait variance). Nevertheless, theory regarding Gaussian functions arrives at a similar
471 conclusion: that offspring variance evolves to the amount of environmental mismatch that is not already
472 covered by phenotypic plasticity (Tufto, 2015). This is equivalent to our finding that only the variance
473 composition (r) changes with environmental variability, whereas the degree of phenotypic variance
474 remains relatively constant (e.g. Fig. 5B). Other possible extensions would include plastic responses
475 that take place within an individual's life time. The opportunity for both within- and transgenerational
476 plasticity may not only make one strategy obsolete (Luquet and Turiel, 2016), but also lead to complex
477 interactions among the two (Fuxjäger et al., 2019). Similarly fitness may include multiplicative
478 instances within an individual's lifetime (e.g. iteroparity), shifting the balance from DBH towards CBH

479 strategies, or conversely sum across generations (“fine-grained” environments), moving the balance
480 towards AMM strategies (Haaland et al., 2020). Lastly, there are also potential bet-hedging strategies
481 that appear entirely unrelated to transgenerational plasticity. These include, for example, an iteroparous
482 life history (Garcia-Gonzalez et al., 2015), hotspots for genetic mutations (“contingency loci”, Rando
483 and Verstrepen, 2007), and sexual reproduction in general (Li et al., 2017). A unification with these
484 alternative strategies might lead to a better understanding of adaptation to rapid climate change.

485 **8 Conclusion**

486 In this review we rephrased reaction norm evolution as a complex trade-off among four strategies. It
487 is increasingly recognized that changes in climate extremes and in predictability are as important as
488 changes in means (IPCC, 2014; Donelson et al., 2018) – focusing only on strategies to match the mean
489 is hence not fruitful. For example, failure to shift mean phenology with climate change (Gienapp et al.,
490 2013) is not problematic per se – it could be mitigated by concurrent changes in phenotypic variance.
491 Similarly, the lack of both phenotypic plasticity and mean change may not have severe fitness
492 consequences, if the lack of plasticity is mitigated by diversified bet-hedging. It is the combination
493 along all three axes that defines fitness in a given environment.

494 **9 Acknowledgments**

495 JJ was financially supported by a DFG research fellowship. DB is funded by FWO project G018017N.

496 **10 Author contributions**

497 JJ and DB developed the theory and JJ wrote the first draft. Both authors contributed to the final version
498 of the manuscript. DB supervised the work.

499 **11 Conflict of interest**

500 The authors declare that the research was conducted in the absence of any commercial or financial
501 relationships that could be construed as a potential conflict of interest.

502 **12 Data availability statement**

503 All datasets generated for this study are included in the supplementary file S3.

504

505 **13 References**

- 506 Adrian-Kalchhauser, I., Sultan, S. E., Shama, L. N. S., Spence-Jones, H., Tiso, S., Valsecchi, C. I. K.,
507 et al. (2020). Understanding “Non-genetic” Inheritance: Insights from Molecular-
508 Evolutionary Crosstalk. *Trends Ecol Evol* 0. doi:10.1016/j.tree.2020.08.011.
- 509 Arnold, P. A., Nicotra, A. B., and Kruuk, L. E. B. (2019). Sparse evidence for selection on
510 phenotypic plasticity in response to temperature. *Phil Trans R Soc B* 374, 20180185.
511 doi:10.1098/rstb.2018.0185.
- 512 Badeck, F. W., Bondeau, A., Bottcher, K., Doktor, D., Lucht, W., Schaber, J., et al. (2004).
513 Responses of spring phenology to climate change. *New Phytol* 162, 295–309.
514 doi:10.1111/j.1469-8137.2004.01059.x.
- 515 Baker, B. H., Sultan, S. E., Lopez-Ichikawa, M., and Waterman, R. (2019). Transgenerational effects
516 of parental light environment on progeny competitive performance and lifetime fitness. *Phil*
517 *Trans R Soc B* 374, 20180182. doi:10.1098/rstb.2018.0182.
- 518 Bathiany, S., Dakos, V., Scheffer, M., and Lenton, T. M. (2018). Climate models predict increasing
519 temperature variability in poor countries. *Sci Adv* 4, eaar5809. doi:10.1126/sciadv.aar5809.
- 520 Bonamour, S., Chevin, L.-M., Charmantier, A., and Teplitsky, C. (2019). Phenotypic plasticity in
521 response to climate change: the importance of cue variation. *Phil. Trans. R. Soc. B* 374,
522 20180178. doi:10.1098/rstb.2018.0178.
- 523 Botero, C. A., Weissing, F. J., Wright, J., and Rubenstein, D. R. (2015). Evolutionary tipping points
524 in the capacity to adapt to environmental change. *PNAS* 112, 184–189.
525 doi:10.1073/pnas.1408589111.
- 526 Boutin, S., and Lane, J. E. (2014). Climate change and mammals: evolutionary versus plastic
527 responses. *Evol Appl* 7, 29–41. doi:10.1111/eva.12121.
- 528 Bradshaw, A. D. (1965). Evolutionary significance of phenotypic plasticity in plants. *Adv Gen* 13,
529 115–155. doi:10.1016/S0065-2660(08)60048-6.
- 530 Bradshaw, W. E. (1976). Geography of photoperiodic response in diapausing mosquito. *Nature* 262,
531 384–386. doi:10.1038/262384b0.
- 532 Braendle, C., Davis, G. K., Brisson, J. A., and Stern, D. L. (2006). Wing dimorphism in aphids.
533 *Heredity* 97, 192–199. doi:10.1038/sj.hdy.6800863.
- 534 Burgess, S. C., and Marshall, D. J. (2014). Adaptive parental effects: the importance of estimating
535 environmental predictability and offspring fitness appropriately. *Oikos* 123, 769–776.
536 doi:10.1111/oik.01235.
- 537 Charmantier, A., and Gienapp, P. (2014). Climate change and timing of avian breeding and
538 migration: evolutionary versus plastic changes. *Evol Appl* 7, 15–28. doi:10.1111/eva.12126.

- 539 Charmantier, A., McCleery, R. H., Cole, L. R., Perrins, C., Kruuk, L. E. B., and Sheldon, B. C.
 540 (2008). Adaptive Phenotypic Plasticity in Response to Climate Change in a Wild Bird
 541 Population. *Science* 320, 800–803. doi:10.1126/science.1157174.
- 542 Chevin, L.-M., Lande, R., and Mace, G. M. (2010). Adaptation, Plasticity, and Extinction in a
 543 Changing Environment: Towards a Predictive Theory. *PLOS Biology* 8, e1000357.
 544 doi:10.1371/journal.pbio.1000357.
- 545 Cohen, D. (1966). Optimizing reproduction in a randomly varying environment. *J Theor Biol* 12,
 546 119–129. doi:10.1016/0022-5193(66)90188-3.
- 547 Cohen, J. M., Lajeunesse, M. J., and Rohr, J. R. (2018). A global synthesis of animal phenological
 548 responses to climate change. *Nature Clim Change* 8, 224–228. doi:10.1038/s41558-018-
 549 0067-3.
- 550 Danilevskii, A. S. (1965). *Photoperiodism and seasonal development of insects*. 1st ed. Edinburgh:
 551 Oliver & Boyd.
- 552 Darwin, C. (1859). *On the Origin of Species by Means of Natural Selection, or the Preservation of*
 553 *Favoured Races in the Struggle for Life*. London: J. Murray.
- 554 Debat, V., and David, P. (2001). Mapping phenotypes: canalization, plasticity and developmental
 555 stability. *Trends Ecol Evol* 16, 555–561. doi:10.1016/S0169-5347(01)02266-2.
- 556 Donelson, J. M., Salinas, S., Munday, P. L., and Shama, L. N. S. (2018). Transgenerational plasticity
 557 and climate change experiments: Where do we go from here? *Glob Change Biol* 24, 13–34.
 558 doi:10.1111/gcb.13903.
- 559 Dueck, H., Eberwine, J., and Kim, J. (2016). Variation is function: Are single cell differences
 560 functionally important? *BioEssays* 38, 172–180. doi:10.1002/bies.201500124.
- 561 Fox, R. J., Donelson, J. M., Schunter, C., Ravasi, T. C., and Gaitán-Espitia, J. D. (2019). Beyond
 562 buying time: the role of plasticity in phenotypic adaptation to rapid environmental change.
 563 *Phil Trans R Soc B* 374, 20180174. doi:10.1098/rstb.2018.0174.
- 564 Furness, A. I., Lee, K., and Reznick, D. N. (2015). Adaptation in a variable environment: Phenotypic
 565 plasticity and bet-hedging during egg diapause and hatching in an annual killifish. *Evolution*
 566 69, 1461–1475. doi:10.1111/evo.12669.
- 567 Fuxjäger, L., Wanzenböck, S., Ringler, E., Wegner, K. M., Ahnelt, H., and Shama, L. N. S. (2019).
 568 Within-generation and transgenerational plasticity of mate choice in oceanic stickleback
 569 under climate change. *Phil. Trans. R. Soc. B* 374, 20180183. doi:10.1098/rstb.2018.0183.
- 570 Garcia-Gonzalez, F., Yasui, Y., and Evans, J. P. (2015). Mating portfolios: bet-hedging, sexual
 571 selection and female multiple mating. *Proc R Soc B* 282, 20141525.
 572 doi:10.1098/rspb.2014.1525.

- 573 Gerber, N., and Kokko, H. (2018). Abandoning the ship using sex, dispersal or dormancy: multiple
574 escape routes from challenging conditions. *Phil Trans R Soc B* 373, 20170424.
575 doi:10.1098/rstb.2017.0424.
- 576 Ghalambor, C. K., McKay, J. K., Carroll, S. P., and Reznick, D. N. (2007). Adaptive versus non-
577 adaptive phenotypic plasticity and the potential for contemporary adaptation in new
578 environments. *Funct Ecol* 21, 394–407. doi:https://doi.org/10.1111/j.1365-
579 2435.2007.01283.x.
- 580 Gibson, G., and Wagner, G. (2000). Canalization in evolutionary genetics: a stabilizing theory?
581 *BioEssays* 22, 372–380. doi:10.1002/(SICI)1521-1878(200004)22:4<372::AID-
582 BIES7>3.0.CO;2-J.
- 583 Gienapp, P., Lof, M., Reed, T. E., McNamara, J., Verhulst, S., and Visser, M. E. (2013). Predicting
584 demographically sustainable rates of adaptation: can great tit breeding time keep pace with
585 climate change? *Phil Trans R Soc B* 368, 20120289. doi:10.1098/rstb.2012.0289.
- 586 Goossens, S., Wybouw, N., Van Leeuwen, T., and Bonte, D. (2020). The physiology of movement.
587 *Mov Ecol* 8, 5. doi:10.1186/s40462-020-0192-2.
- 588 Grantham, M. E., Antonio, C. J., O’Neil, B. R., Zhan, Y. X., and Brisson, J. A. (2016). A case for a
589 joint strategy of diversified bet hedging and plasticity in the pea aphid wing polyphenism.
590 *Biol Lett* 12, 20160654. doi:10.1098/rsbl.2016.0654.
- 591 Green, T. R., and Ryan, C. A. (1972). Wound-Induced Proteinase Inhibitor in Plant Leaves: A
592 Possible Defense Mechanism against Insects. *Science* 175, 776–777.
593 doi:10.1126/science.175.4023.776.
- 594 Haaland, T. R., Wright, J., and Ratikainen, I. I. (2020). Generalists versus specialists in fluctuating
595 environments: a bet-hedging perspective. *Oikos* 129, 879–890. doi:10.1111/oik.07109.
- 596 Halkett, F., Harrington, R., Hullé, M., Kindlmann, P., Menu, F., Risper, C., et al. (2004). Dynamics of
597 production of sexual forms in aphids: theoretical and experimental evidence for adaptive
598 “coin-flipping” plasticity. *Am Nat* 163, E112–E125. doi:10.1086/383618.
- 599 Hopper, K. R. (1999). Risk-spreading and bet-hedging in insect population biology. *Annu Rev*
600 *Entomol* 44, 535–560. doi:10.1146/annurev.ento.44.1.535.
- 601 IPCC (2014). *Climate Change 2014: Synthesis report. Contribution of Working Groups I, II and III*
602 *to the fifth assessment report of the Intergovernmental Panel on Climate Change*. Geneva,
603 Switzerland.
- 604 Joschinski, J., and Bonte, D. (2020). Diapause is not selected as a bet-hedging strategy in insects: a
605 meta-analysis of reaction norm shapes. *bioRxiv*, 752881, version 3 peer-reviewed and
606 recommended by Peer Community in Ecology, doi: 10.24072/pci.ecology.100040.
607 doi:10.1101/752881.
- 608 Kærn, M., Elston, T. C., Blake, W. J., and Collins, J. J. (2005). Stochasticity in gene expression: from
609 theories to phenotypes. *Nat Rev Genet* 6, 451–464. doi:10.1038/nrg1615.

- 610 Kelly, M. (2019). Adaptation to climate change through genetic accommodation and assimilation of
611 plastic phenotypes. *Phil Trans R Soc B* 374, 20180176. doi:10.1098/rstb.2018.0176.
- 612 Kivela, S. M., Valimaki, P., and Gotthard, K. (2016). Evolution of alternative insect life histories in
613 stochastic seasonal environments. *Ecol Evol* 6, 5596–5613. doi:10.1002/ece3.2310.
- 614 Krueger, D. A., and Dodson, S. I. (1981). Embryological induction and predation ecology in *Daphnia*
615 *pulex*. *Limnol Oceanogr* 26, 219–223. doi:10.4319/lo.1981.26.2.0219.
- 616 Lane, J. E., McAdam, A. G., McFarlane, S. E., Williams, C. T., Humphries, M. M., Coltman, D. W.,
617 et al. (2018). Phenological shifts in North American red squirrels: disentangling the roles of
618 phenotypic plasticity and microevolution. *J Evol Biol* 31, 810–821. doi:10.1111/jeb.13263.
- 619 Lenton, T. M., Dakos, V., Bathiany, S., and Scheffer, M. (2017). Observed trends in the magnitude
620 and persistence of monthly temperature variability. *Sci Rep* 7, 5940. doi:10.1038/s41598-017-
621 06382-x.
- 622 Li, X.-Y., Lehtonen, J., and Kokko, H. (2017). Sexual reproduction as bet-hedging [preprint].
623 *bioRxiv*, 103390. doi:10.1101/103390.
- 624 Luquet, E., and Tariel, J. (2016). Offspring reaction norms shaped by parental environment:
625 interaction between within- and trans-generational plasticity of inducible defenses. *BMC*
626 *Evolutionary Biology* 16, 209. doi:10.1186/s12862-016-0795-9.
- 627 Marcovitch, S. (1923). Plant lice and light exposure. *Science* 58, 537–538.
628 doi:10.1126/science.58.1513.537-a.
- 629 Marshall, D. J., Bonduriansky, R., and Bussière, L. F. (2008). Offspring size variation within broods
630 as a bet-hedging strategy in unpredictable environments. *Ecology* 89, 2506–2517.
631 doi:10.1890/07-0267.1.
- 632 Maxwell, C. S., and Magwene, P. M. (2017). When sensing is gambling: An experimental system
633 reveals how plasticity can generate tunable bet-hedging strategies. *Evolution* 71, 859–871.
634 doi:10.1111/evo.13199.
- 635 Nussey, D. H., Postma, E., Gienapp, P., and Visser, M. E. (2005). Selection on heritable phenotypic
636 plasticity in a wild bird population. *Science* 310, 304–306. doi:10.1126/science.1117004.
- 637 Oostra, V., Saastamoinen, M., Zwaan, B. J., and Wheat, C. W. (2018). Strong phenotypic plasticity
638 limits potential for evolutionary responses to climate change. *Nature Communications* 9.
639 doi:10.1038/s41467-018-03384-9.
- 640 Parmesan, C., and Yohe, G. (2003). A globally coherent fingerprint of climate change impacts across
641 natural systems. *Nature* 421, 37. doi:10.1038/nature01286.
- 642 Pélişson, P.-F., Bernstein, C., François, D., Menu, F., and Venner, S. (2013). Dispersal and dormancy
643 strategies among insect species competing for a pulsed resource. *Ecological Entomology* 38,
644 470–477. doi:10.1111/een.12038.

- 645 Perez, M. F., and Lehner, B. (2019). Intergenerational and transgenerational epigenetic inheritance in
646 animals. *Nature Cell Biology* 21, 143–151. doi:10.1038/s41556-018-0242-9.
- 647 Perrin, N. (2016). Random sex determination: When developmental noise tips the sex balance.
648 *BioEssays* 38, 1218–1226. doi:10.1002/bies.201600093.
- 649 Piao, S., Liu, Q., Chen, A., Janssens, I. A., Fu, Y., Dai, J., et al. (2019). Plant phenology and global
650 climate change: Current progresses and challenges. *Global Change Biology* 25, 1922–1940.
651 doi:10.1111/gcb.14619.
- 652 Rando, O. J., and Verstrepen, K. J. (2007). Timescales of genetic and epigenetic inheritance. *Cell*
653 128, 655–668. doi:10.1016/j.cell.2007.01.023.
- 654 Saikkonen, K., Taulavuori, K., Hyvönen, T., Gundel, P. E., Hamilton, C. E., Vänninen, I., et al.
655 (2012). Climate change-driven species' range shifts filtered by photoperiodism. *Nat Clim*
656 *Change* 2, 239–242. doi:10.1038/nclimate1430.
- 657 Scheiner, S. M., and Holt, R. D. (2012). The genetics of phenotypic plasticity. X. Variation versus
658 uncertainty. *Ecol Evol* 2, 751–767. doi:10.1002/ece3.217.
- 659 Scholl, J. P., Calle, L., Miller, N., and Venable, D. L. (2020). Offspring polymorphism and bet
660 hedging: a large-scale, phylogenetic analysis. *Ecology Letters* 23, 1223–1231.
661 doi:10.1111/ele.13522.
- 662 Seger, J., and Brockmann, H. J. (1987). “What is bet-hedging?,” in *Oxford surveys in evolutionary*
663 *biology* (4) (Oxford, UK: Oxford University Press), 182–211.
- 664 Simon, J.-C., Rispé, C., and Sunnucks, P. (2002). Ecology and evolution of sex in aphids. *Trends*
665 *Ecol Evol* 17, 34–39. doi:10.1016/S0169-5347(01)02331-X.
- 666 Simons, A. M. (2011). Modes of response to environmental change and the elusive empirical
667 evidence for bet hedging. *Proc R Soc B* 278, 1601–1609. doi:10.1098/rspb.2011.0176.
- 668 Simons, A. M., and Johnston, M. O. (1997). Developmental instability as a bet-hedging strategy.
669 *Oikos* 80, 401–406. doi:10.2307/3546608.
- 670 Simons, A. M., and Johnston, M. O. (2006). Environmental and genetic sources of diversification in
671 the timing of seed germination: implications for the evolution of bet hedging. *Evolution* 60,
672 2280–2292. doi:10.1111/j.0014-3820.2006.tb01865.x.
- 673 Starrfelt, J., and Kokko, H. (2012). Bet-hedging--a triple trade-off between means, variances and
674 correlations. *Biol Rev* 87, 742–755. doi:10.1111/j.1469-185X.2012.00225.x.
- 675 Stomp, M., van Dijk, M. A., van Overzee, H. M. J., Wortel, M. T., Sigon, C. A. M., Egas, M., et al.
676 (2008). The timescale of phenotypic plasticity and its impact on competition in fluctuating
677 environments. *Am Nat* 172, E169–E185. doi:10.1086/591680.
- 678 Tufto, J. (2015). Genetic evolution, plasticity, and bet-hedging as adaptive responses to temporally
679 autocorrelated fluctuating selection: A quantitative genetic model. *Evolution* 69, 2034–2049.
680 doi:10.1111/evo.12716.

- 681 Van Buskirk, J., and Steiner, U. K. (2009). The fitness costs of developmental canalization and
682 plasticity. *J Evol Biol* 22, 852–860. doi:10.1111/j.1420-9101.2009.01685.x.
- 683 Veening, J.-W., Smits, W. K., and Kuipers, O. P. (2008). Bistability, epigenetics, and bet-hedging in
684 bacteria. *Annu Rev Microbiol* 62, 193–210. doi:10.1146/annurev.micro.62.081307.163002.
- 685 Venable, D. L. (2007). Bet hedging in a guild of desert annuals. *Ecology* 88, 1086–1090.
686 doi:10.1890/06-1495.
- 687 Volfson, D., Marciniak, J., Blake, W. J., Ostroff, N., Tsimring, L. S., and Hasty, J. (2006). Origins of
688 extrinsic variability in eukaryotic gene expression. *Nature* 439, 861–864.
689 doi:10.1038/nature04281.
- 690 Waddington, C. H. (1942). Canalization of development and the inheritance of acquired characters.
691 *Nature* 150, 563–565. doi:10.1038/150563a0.
- 692 Woltereck, R. (1913). Weitere experimentelle untersuchungen über Artänderung, speziell über das
693 Wesen quantitativer Artunterschiede bei Daphniden. *Mol Gen Genet* 9, 146–146.
694 doi:10.1007/BF01876686.
- 695 Woods, H. A. (2014). Mosaic physiology from developmental noise: within-organism physiological
696 diversity as an alternative to phenotypic plasticity and phenotypic flexibility. *J Exp Bio* 217,
697 35–45. doi:10.1242/jeb.089698.
- 698 Zhang, X.-S., and Hill, W. G. (2005). Evolution of the environmental component of the phenotypic
699 variance: stabilizing selection in changing environments and the cost of homogeneity.
700 *Evolution* 59, 1237–1244. doi:10.1111/j.0014-3820.2005.tb01774.x.
- 701 Zohner, C. M., Benito, B. M., Svenning, J.-C., and Renner, S. S. (2016). Day length unlikely to
702 constrain climate-driven shifts in leaf-out times of northern woody plants. *Nat Clim Change*
703 6, 1120–1123. doi:10.1038/nclimate3138.

704

705

706 **14 Figure legends**

707 Fig. 1. Geometric (solid lines) and arithmetic mean fitness (dashed lines) when a genotype can express
 708 two discrete phenotypes in a two-state environment. A) Conflict between geometric and arithmetic
 709 mean maximization. Environment E_2 (e.g. winter) occurs with frequencies of 0.5 (blue) or 0.2 (orange).
 710 Phenotype P_2 represents a risk-averse phenotype (e.g. diapausing offspring) with 1 fitness in either
 711 environment, the alternative phenotype is a phenotype with higher arithmetic mean fitness (4 fitness in
 712 E_1 , 0.1 in E_2). B) No or little conflict between arithmetic and geometric mean maximization. Blue line:
 713 E_2 occurs with frequency 0.5 and P_1 and P_2 are specialists for E_1 and E_2 , respectively (4 fitness if
 714 matched, 0 fitness if mismatched); grey: same as blue line, but P_2 has 3.9 fitness in E_2 ; orange: fitness
 715 is the same as in panel A, but E_2 occurs with frequency 0.8. Colored dots represent the maxima of the
 716 respective functions.

717 Fig. 2 Panel A) Probability of encountering environment E_2 (winter conditions) for different values of
 718 an environmental cue c (e.g. night length). E_2 fluctuates around c according to three normal
 719 distributions $N_1(14,1)$, $N_2(14,4)$ and $0.5 * N_3(14,2)$ (blue, orange, green). Shown are cumulative
 720 probability functions of the three distributions. B) Optimal reaction norm shapes (e.g. proportion p of
 721 diapausing offspring for different night lengths) under the three scenarios of environmental uncertainty
 722 introduced in panel A. As in the main text, fitness of P_1 (parthenogenesis) is 4 in E_1 and 0.1 in E_2 ,
 723 whereas fitness of P_2 is always 1. C) optimal reaction norm shapes when fitness of P_1 is 4/0 and fitness
 724 of P_2 is 1.8/1.8 in E_1/E_2 , respectively. Dotted lines represent $c = 14$ h, small coloured dots refer to the
 725 examples given in the main text.

726 Fig. 3: Example reaction norm shapes. A) Four reaction norm shapes that exhibit high phenotypic
 727 variance. Variance may occur exclusively among environments (dark blue), exclusively within each
 728 environment (light blue), or as a mix of both variance components (medium blue, solid and dashed).
 729 We refer to the ratio of the variance components (among : within) as r . B) Three different reaction
 730 norms with $r = 0$ (solid lines), and two different reaction norms with $r = 0.14$ (dashed). The reaction
 731 norms differ in the mean frequency f of phenotype P_2 , which also affects the phenotypic variance s (i.e.
 732 the sum of variance among and within environments). Reaction norms with $f = 0$ (light orange) and f
 733 $= 1$ (dark orange) are canalized ($s = 0$), and phenotypic variance is maximized at $f = 0.5$ (see panel A).
 734 C) Two logistic reaction norms with the same f and r , but different inflection points. D) possible
 735 parameter space of r, f and inflection points. Grey dots depict sample reaction norms across the range

736 of possible parameters (darkness scales with z-axis), colored dots indicated samples from panel A and
737 B and C in their respective color.

738 Fig. 4: Optimal reaction norm shapes for various growth rate functions and different levels of
739 environmental predictability. Environments are normally distributed around a cue c with a mean of 14.
740 Mean frequency f of phenotype P_2 (Panel A) and variance composition r (Panel C) are plotted against
741 standard deviation of the environment. Growth rates of P_2 (diapause) are always 1 for both
742 environments (summer and winter); growth rates of P_1 (parthenogenesis) in E_1/E_2 are 4/0.1 (blue,
743 solid), 3/0.1 (orange, solid), 2/0.1 (green, solid); 4/0.33, 3/0.33, 2/0.33 (dashed blue, orange and green
744 lines); and 4/0.66, 3/0.66, 2/0.66 (dash-dotted blue, orange and green lines). Panels B and D show
745 optimal reaction norms for environments with standard deviations of 2(darker shade) and 8 (lighter
746 shade) in the according line styles and colors.

747 Fig. 5: Optimal reaction norm shapes for various growth rate functions and different levels of
748 environmental predictability. Environments are normally distributed around a cue c with a mean of 14,
749 but multiplied by 0.5. Growth rates, coloring and line styles are the same as Fig. 4.

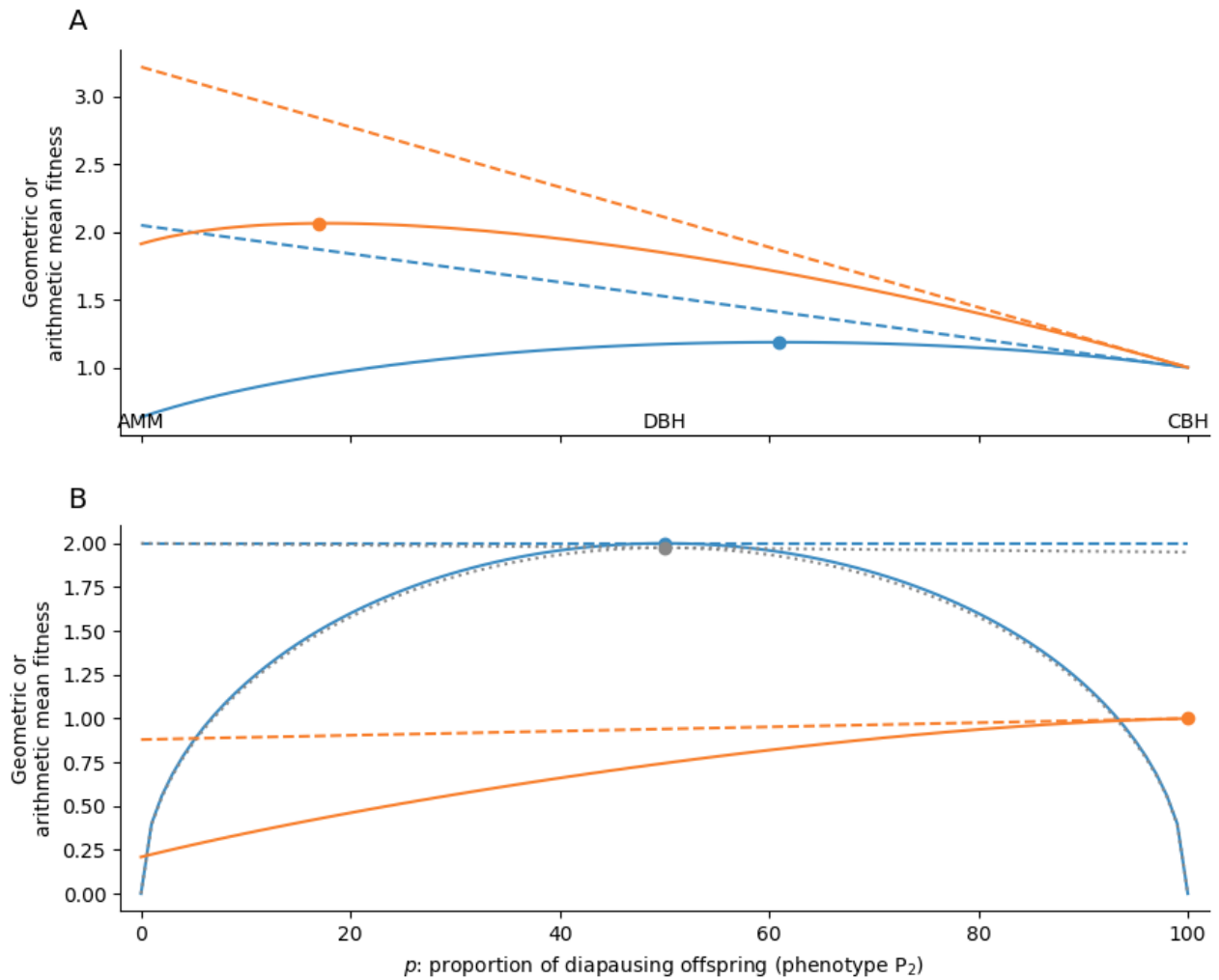
750

751 **Table 1:** Growth rate calculations for various phenotype proportions in a two-environment system. A
 752 genotype may invest in two different phenotypes, P_1 and P_2 , with a fixed proportion p . P_1 has four
 753 offspring if in environment E_1 , but 0.1 if in E_2 ; P_2 achieves 1 fitness in either environment. We show
 754 arithmetic and geometric mean fitness across environments (Environments E_1 and E_2 are chosen with
 755 probability 0.5), as well as their calculation (italics).

	Proportion of P_2 (p)			
	0	0.5	1	0.61
E_1	4 <i>(0 * 1 + 1 * 4)</i>	2.5 <i>(0.5 * 1 + 0.5 * 4)</i>	1 <i>(1 * 1 + 0 * 4)</i>	2.17 <i>(0.61 * 1 + 0.39 * 4)</i>
E_2	0.1 <i>(0 * 1 + 1 * 0.1)</i>	0.55 <i>(0.5 * 1 + 0.5 * 0.1)</i>	1 <i>(1 * 1 + 0 * 0.1)</i>	0.65 <i>(0.61 * 1 + 0.39 * 0.1)</i>
Arithmetic mean	2.05	1.53	1	1.41
Geometric mean	0.63	1.17	1	1.19

756

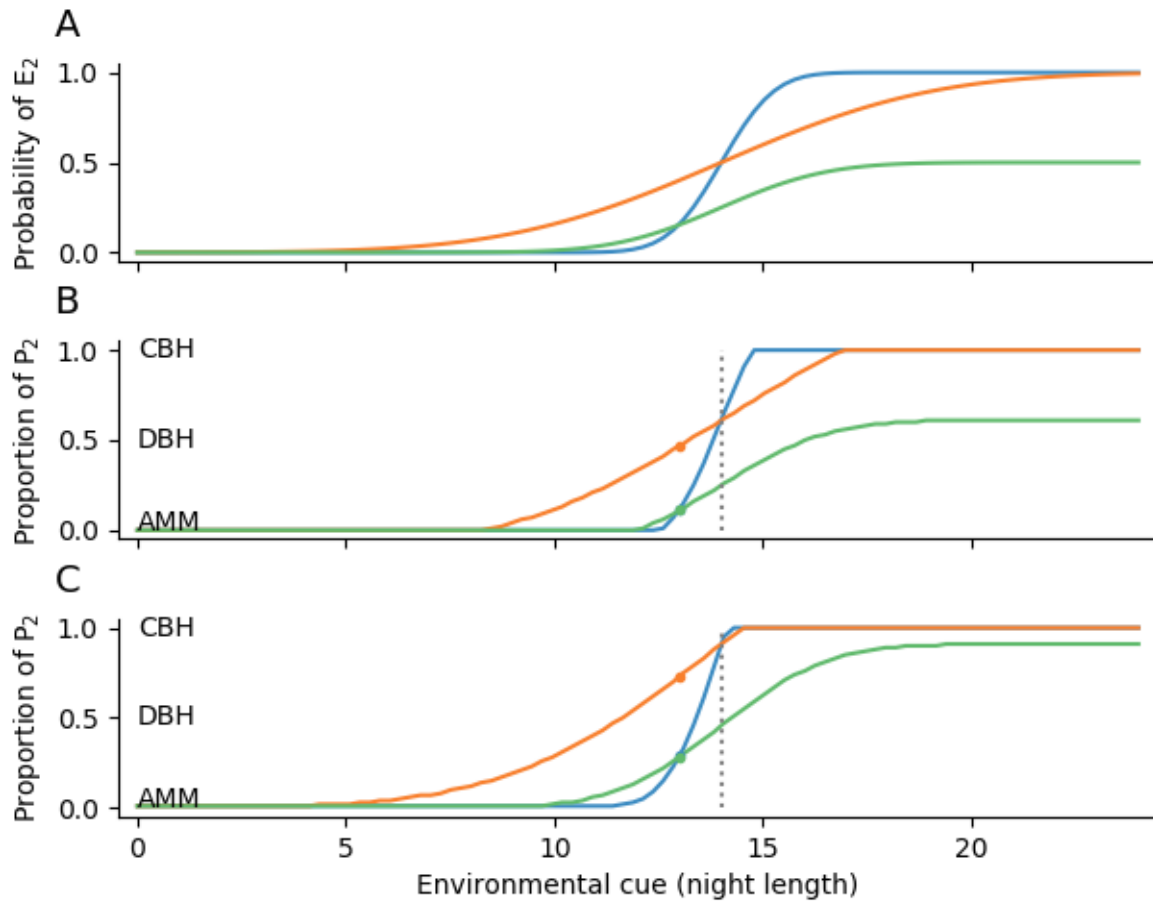
757



758

759 Fig. 1. Geometric (solid lines) and arithmetic mean fitness (dashed lines) when a genotype can express
 760 two discrete phenotypes in a two-state environment. A) Conflict between geometric and arithmetic
 761 mean maximization. Environment E_2 (e.g. winter) occurs with frequencies of 0.5 (blue) or 0.2 (orange).
 762 Phenotype P_2 represents a risk-averse phenotype (e.g. diapausing offspring) with 1 fitness in either
 763 environment, the alternative phenotype is a phenotype with higher arithmetic mean fitness (4 fitness in
 764 E_1 , 0.1 in E_2). B) No or little conflict between arithmetic and geometric mean maximization. Blue line:
 765 E_2 occurs with frequency 0.5 and P_1 and P_2 are specialists for E_1 and E_2 , respectively (4 fitness if
 766 matched, 0 fitness if mismatched); grey: same as blue line, but P_2 has 3.9 fitness in E_2 ; orange: fitness
 767 is the same as in panel A, but E_2 occurs with frequency 0.8. Colored dots represent the maxima of the
 768 respective functions.

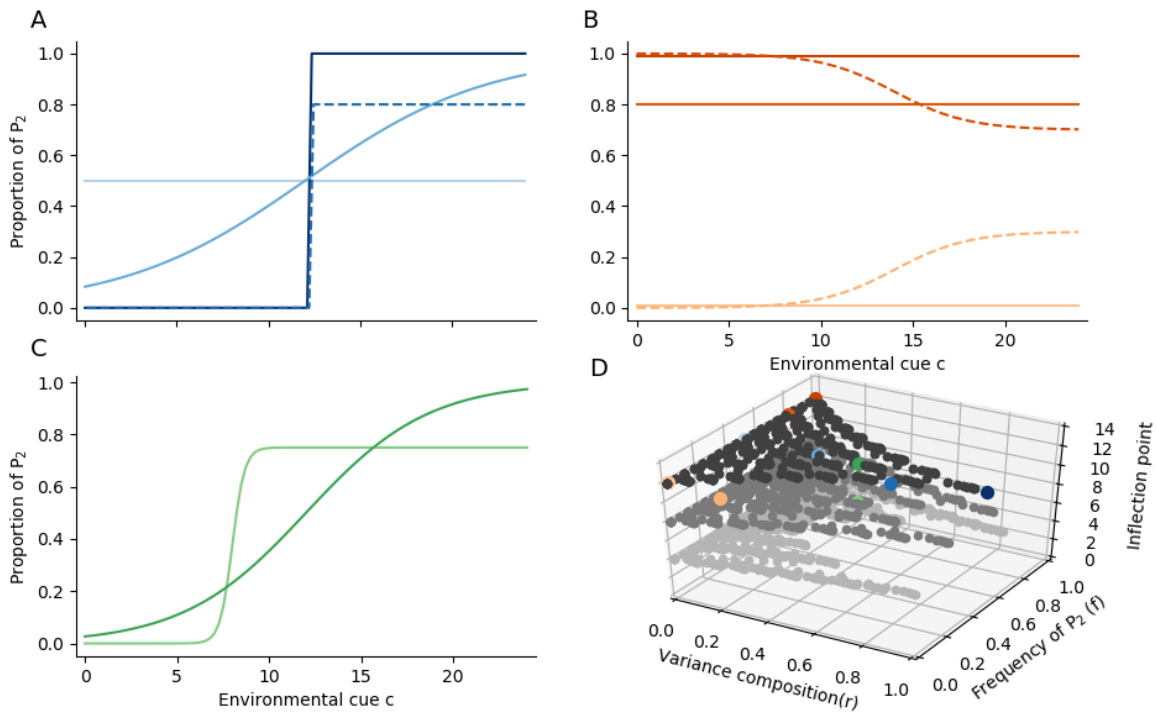
769



770

771 Fig. 2 Panel A) Probability of encountering environment E_2 (winter conditions) for different values of
 772 an environmental cue c (e.g. night length). E_2 fluctuates around c according to three normal
 773 distributions $N_1(14,1)$, $N_2(14,4)$ and $0.5 * N_3(14,2)$ (blue, orange, green). Shown are cumulative
 774 probability functions of the three distributions. B) Optimal reaction norm shapes (e.g. proportion p of
 775 diapausing offspring for different night lengths) under the three scenarios of environmental uncertainty
 776 introduced in panel A. As in the main text, fitness of P_1 (parthenogenesis) is 4 in E_1 and 0.1 in E_2 ,
 777 whereas fitness of P_2 is always 1. C) optimal reaction norm shapes when fitness of P_1 is 4/0 and fitness
 778 of P_2 is 1.8/1.8 in E_1/E_2 , respectively. Dotted lines represent $c = 14$ h, small coloured dots refer to the
 779 examples given in the main text.

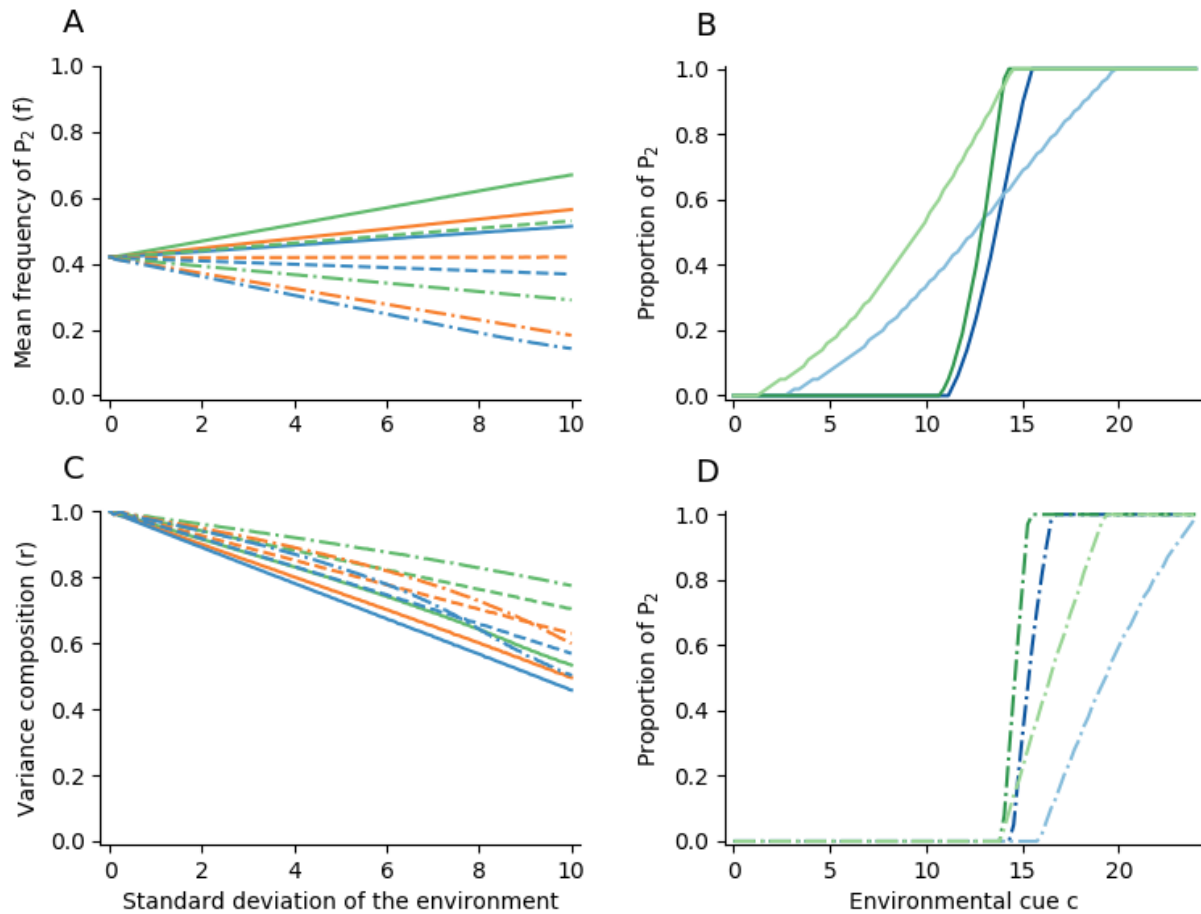
780



781

782 Fig. 3: Example reaction norm shapes. **A)** Four reaction norm shapes that exhibit high phenotypic
 783 variance. Variance may occur exclusively among environments (dark blue), exclusively within each
 784 environment (light blue), or as a mix of both variance components (medium blue, solid and dashed).
 785 We refer to the ratio of the variance components (among : within) as r . **B)** Three different reaction
 786 norms with $r = 0$ (solid lines), and two different reaction norms with $r = 0.14$ (dashed). The reaction
 787 norms differ in the mean frequency f of phenotype P_2 , which also affects the phenotypic variance s (i.e.
 788 the sum of variance among and within environments). Reaction norms with $f = 0$ (light orange) and f
 789 $= 1$ (dark orange) are canalized ($s = 0$), and phenotypic variance is maximized at $f = 0.5$ (see panel A).
 790 **C)** Two logistic reaction norms with the same f and r , but different inflection points. **D)** Possible
 791 parameter space of r , f and inflection points. Grey dots depict sample reaction norms across the range
 792 of possible parameters (darkness scales with z-axis), colored dots indicated samples from panel A and
 793 B and C in their respective color.

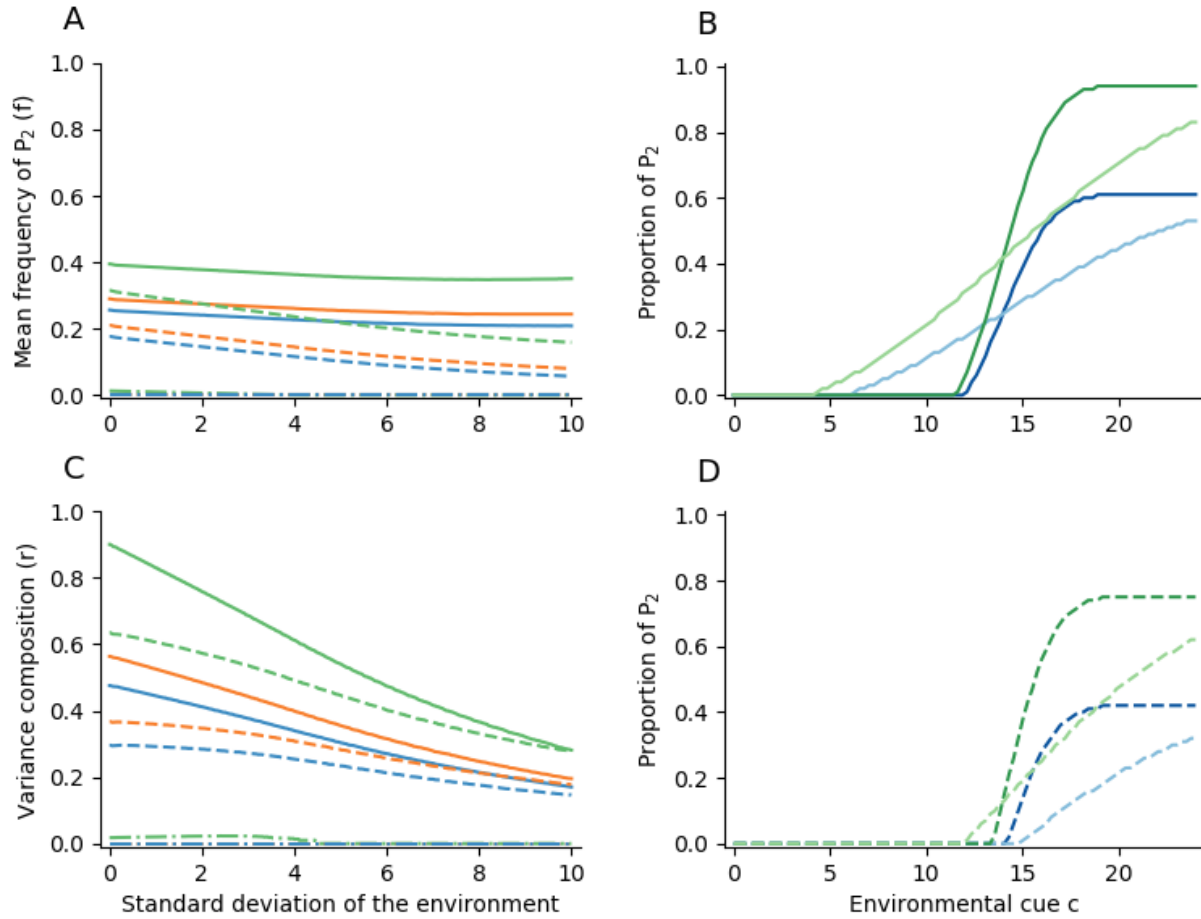
794



795

796 Fig. 4: Optimal reaction norm shapes for various growth rate functions and different levels of
 797 environmental predictability. Environments are normally distributed around a cue c with a mean of 14.
 798 Mean frequency f of phenotype P_2 (Panel A) and variance composition r (Panel C) are plotted against
 799 standard deviation of the environment. Growth rates of P_2 (diapause) are always 1 for both
 800 environments (summer and winter); growth rates of P_1 (parthenogenesis) in E_1/E_2 are 4/0.1 (blue,
 801 solid), 3/0.1 (orange, solid), 2/0.1 (green, solid); 4/0.33, 3/0.33, 2/0.33 (dashed blue, orange and green
 802 lines); and 4/0.66, 3/0.66, 2/0.66 (dash-dotted blue, orange and green lines). Panels B and D show
 803 optimal reaction norms for environments with standard deviations of 2(darker shade) and 8 (lighter
 804 shade) in the according line styles and colors.

805



806

807 Fig. 5: Optimal reaction norm shapes for various growth rate functions and different levels of
 808 environmental predictability. Environments are normally distributed around a cue c with a mean of 14,
 809 but multiplied by 0.5. Growth rates, coloring and line styles are the same as Fig. 4.

810