1	Main	carbon	sources	supporting	primary	and	secondary	production	in	a
2	2 disturbed semiarid wetland from central Spain									

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18 Abstract

19 Approximately 95% of the total carbon (C) in wetlands is typically found as particulate 20 organic matter (POM). In this study, we evaluated the main C sources of a semiarid floodplain wetland in central Spain under disturbance. For this, we used stable isotopes (¹³C 21 and ¹⁵N) and the Bayesian mixing model SISUS. We show that the allochthonous C derived 22 from wastewater discharges have distinctive δ^{13} C values compared to the C of sites that did 23 24 not receive wastewater discharges. Wastewater discharges affected the isotopic signatures of dissolved organic carbon as well as δ^{15} N values of primary producers (epiphyton, filamentous 25 algae and *Scirpus maritimus*). δ^{15} N values showed a greater response to the effects of 26 environmental degradation. The salt-tolerant marginal tree Tamarix canariensis was the main 27 28 autochthonous contributor to POM (69 to 84%), while the main source of energy for the 29 secondary production was the submerged macrophyte Ceratophyllum submersum. Tamarix 30 canariensis was also the main component of the diet for macroinvertebrates, followed by 31 POM. Other species such as *Cladium mariscus*, an emblematic aquatic plant in the region, 32 showed a very low contribution to POM (less than 20%), although it represented a relatively 33 important energy source for zooplankton (up to 47%). This relatively low contribution is 34 probably due to the drastic reduction in the abundance of this species during the last 50 years. 35 In contrast, *Phragmites australis*, which has multiplied its abundance by twenty-two since 36 1950, does not seem to contribute significantly to POM or to the diet of invertebrates (less 37 than 22%), although it can represent up to 54% zooplankton's diet. Isotopic results confirm 38 that in this eutrophied and disturbed wetland, the contribution of macrophytes to the food 39 web as autochthonous organic sources is lower than expected based on their biomass.

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41 Keywords: carbon sources; SISUS; Daimiel; primary producers; secondary producers;
42 trophic interactions; wetland

43 **1. Introduction**

Approximately 95% of the total carbon (C) found in wetlands is in the form of particulate 44 45 organic matter (POM), which is generated through the decomposition of litterfall and detrital 46 production (Reddy & DeLaune, 2008). Wetlands can produce their own organic matter (i.e., 47 authoctonous sources, including remains of vegetation like leaves, roots, rhizomes and 48 detrital material from algae, microorganisms, invertebrates, vertebrates, etc.), but they can 49 also receive organic C from allochtonous sources. Allochtonous sources include dissolved 50 and particulate organic C from diffuse sources, including those derived from agriculture and 51 wastewater effluents of different types (Reddy & DeLaune, 2008). Allochthonous 52 contributions are generally very important to support the metabolism of aquatic ecosystems 53 and they are, in most river systems, the largest contributor to the organic C budget (Carpenter 54 et al., 2005), affecting significantly the configuration and regulation of food webs (Polis & 55 Strong, 1996; Huxel & McCann, 1998)

56 Wetland POM consists of a mixture of terrestrial, lacustrine and riverine organic 57 remains, all subject to additional physical, chemical and biological transformations (Hedges, 58 Clark & Come, 1988), with the intensity and direction of water exchange between the river 59 and the floodplain conditioning the balance of organic matter (Albuquerque & Mozeto, 1997; 60 Valett et al., 2005) The role of wetlands in the retention and transformation of organic matter 61 has been shown in numerous studies (Newbold et al., 1982; Puckett et al., 1993; Sánchez 62 Carrillo, Álvarez Cobelas & Angeler, 2001; Valett et al., 2005). However, the balances of 63 organic matter of wetlands are hard to establish, mainly due to the difficulty of identifying the 64 main sources of organic C in these systems. This difficulty resides in the presence of multiple 65 molecular forms and organic matter sources, as well as the high spatiotemporal variability in 66 the activity of these ecosystems (Hedges et al., 1988; Albuquerque & Mozeto, 1997). 67 Different complex markers, such as organic matter fatty acid composition (Mortillaro et al.,

68 2011), the comparative composition of carbohydrates, proteins, lignin, lipids, char and 69 "carbonyl" groups (Nelson & Baldock, 2005; Rodríguez-Murillo, Almendros & Knicker, 70 2011) have been widely used to elucidate the source and destination of the organic matter in 71 aquatic environments and to characterize nutrient utilization by heterotrophic organisms 72 (Teranes & Bernasconi, 2000; Mortillaro et al., 2011). Stable isotopes have also been used to 73 determine the contribution of POM to food webs (Sullivan & Moncreiff, 1990; Fry, 1991; 74 Bunn & Boon, 1993; Keough, Sierszen & Hagley, 1996; Grey, Jones & Sleep, 2001; Bunn, 75 Davies & Winning, 2003; Pace et al., 2004). For example, Bunn & Boon (1993) measured ¹³C and ¹⁵N in Australian floodplain wetlands (Billabongs) to evaluate the contribution of 76 77 POM to three food webs and found that none of the conspicuous organic matter sources of 78 the site (terrestrial, submerged, floating and emergent aquatic macrophytes) were the source 79 of C for most consumers. In contrast, Keough et al. (1996) showed that two food webs in a 80 coastal wetland of Lake Superior depended on the C fixed by phytoplankton, although they 81 also found no evidence of a direct connection between the most abundant macrophytes and the fish fauna analysed. Although δ^{13} C and δ^{15} N have proved useful to distinguish between 82 83 aquatic and terrestrial primary producers and their influence on the metabolism of wetlands, 84 their signals are often masked by the different contributions of organic matter from different photosynthetic sources such as phytoplankton or terrestrial and aquatic C₃/C₄ plants 85 86 (Townsend-Small, McClain & Brandes, 2005).

In recent years, there has been greater use of mixing models using stable isotopes to quantify the diet or the path of organic matter in food webs (Phillips, 2001; Phillips & Gregg, 2003; Benstead *et al.*, 2006; Zeug & Winemiller, 2008). These models are based on mixture sets of two or more stable isotopes (typically δ^{13} C and δ^{15} N) and are a powerful tool to determine the relative contribution of each of the sources in those organisms that have more than one potential source and may themselves be isotopically distinct (Phillips, 2001). In a

93 previous study, (Rodríguez-Murillo et al., 2011) characterized the organic matter in sediments of "Las Tablas de Daimiel" to determine its origin and transformations using $\delta^{13}C$ nuclear 94 magnetic resonance and elemental analysis and applying a molecular mixture model. Their 95 96 results indicate that macrophytes (emergent or submerged) are the main source of organic 97 matter in wetland soils and that their quality depend more on drastic changes, such as the 98 effect of fires, than on external inputs. However, the contribution of different 99 autochthons/allochthonous carbon sources in sustaining the food web of this wetland is 100 unknown.

101 The aim of this study was to determine the main sources of C in "Las Tablas de 102 Daimiel", a semiarid floodplain wetland from central Spain, through the isotopic signatures 103 $(\delta^{13}C \text{ and } \delta^{15}N)$ of plants, phytoplankton, algae and detrital bacteria. We assessed how 104 different forms of biomass (i.e., C sources) are incorporated into both organic matter and 105 primary producers. We also evaluated the effect that organic material from urban wastewater 106 discharge has on the food web of this wetland. Moreover, we also collected information about 107 potential organic sources that support communities in different habitats of the wetland 108 (coastal areas, shallow areas free of emergent vegetation, deep areas and river channels). This 109 information is essential to define the functions that support the food web and may be useful 110 and necessary for mitigation projects and wetland restoration.

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112 **2. Material and methods**

113 1.1 Study site

This study was conducted in the "National Park Las Tablas de Daimiel" (hereafter referred to
as NPTD), a unique wetland in the Euro-Mediterranean region (39° 08' N, 3° 43' W, Fig. 1).
Currently, the ecosystem encompasses the heart of the Biosphere Reserve "La Mancha
Húmeda" (≈8.000 km2), a major wetland region of southern Europe (Sánchez-Carrillo &

Angeler, 2010). NPTD potentially floods an area of 15.9 km² and the maximum average depth is 0.9 m. The flooded area has great annual seasonal fluctuations that lead to drastic changes in the habitats during the dry cycles (Angeler *et al.*, 2001).

121 The National Park is what remains of an extensive floodplain that covered an area of 150-250 km² in the early twentieth century (Sánchez-Carrillo & Angeler, 2010). Since the 122 123 1950s, the extent of the wetland has been drastically reduced and its ecological integrity is seriously threatened (Cirujano, Álvarez-Cobelas & Sánchez-Andrés, 2010; Sánchez-Carrillo 124 125 & Angeler, 2010). The uniqueness and richness of the landscape used to be characterized by 126 the seasonal mixture of surface, sulphate-rich waters with more fresh and stable groundwater 127 (Coronado Castillo et al., 1974); however, currently this system works very differently from 128 the biogeochemical point of view, and many external inputs of organic matter from the 129 watershed (a large fraction of this wastewater is untreated or insufficiently treated) has 130 boosted internal production and turned the wetland into turbid waters (Alvarez-Cobelas et al. 2001; Sánchez-Carrillo et al. 2001). Thus, NPTD has become a hypertrophic wetland, 131 132 receiving both high organic loads from wastewater with insufficient treatment as well as 133 diffused agricultural pollution that reaches the basin through the Gigüela River (Sánchez-134 Carrillo & Angeler, 2010). More information about Las Tablas de Daimiel can be found in (Sánchez-Carrillo & Angeler, 2010). Physical-chemical water conditions during the study 135 136 period in different parts of the wetland are shown in Table 1.

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138 2.2 Collection of potential wetland C sources

The living C reservoirs collected were: seven macrophytes (four emerging: *Cladium mariscus*, *Phragmites australis*, *Typha dominguensis* and *Scirpus maritimus*, and three submerged: *Potamogeton pectinatus*, *Ceratophyllum submersum* and *Chara hispida* var. *major*, the latter
being the most abundant charophyte in the Park), *Tamarix canariensis* (forming large clusters

in coastal wetland areas and flourishing in marginal areas), epiphytic algae, filamentous algae,
phytoplankton and microbial mats. Complete charophytes were collected and cleaned of
sediments. For the rest of plants, we collected green leaves and, when present, rhizomes.
Periphyton was collected by scraping submerged wood substrate, macrophytes and rocks.
Filamentous algae were collected in sites where they were present. The roots from the three
most abundant species of wetland were also analysed (*C. mariscus, P. australis* and *T. dominguensis*).

We separated phytoplankton from the zooplankton by filtration with Nytal filters ($45\mu m$) and then filtering a volume of 10 L with GF/F filters ($0.2 \mu m$). Microbial mats including cyanobacteria such as *Microcoleus chtonoplastes* (Barón, 2011) were collected by carefully removing the protruding layer on the substrate with a spatula and then passing the remaining material through a 1-mm mesh.

155 As the secondary producers from this study, we collected zooplankton and benthic 156 macroinvertebrates. Invertebrates were collected using dredged sediments, storing live 157 specimens without preservatives. Invertebrate species were identified as Crocothemis 158 erytrhaea (Odonata), Naucoris maculatus (Heteroptera) and Chironomus sp. (Diptera). 159 Zooplankton was filtered from 40 L of water through a Nytal filter, and this material was 160 subsequently slurred and filtered with 0.45 µm filters before being oven-dried at 80 °C. 161 Invertebrates were analysed as a composite samples because it was not possible to obtain 162 sufficient mass of each species at each site.

We also collected sediment samples, from which visible vegetation remains were eliminated. The water C pool was estimated as the sum of particulate organic C (POC), dissolved organic C (DOC) and dissolved inorganic C (DIC), which were collected at each site. Samples for δ^{13} C-DIC were taken into 60 ml vials and were acidified to pH <2 with 1 ml of H₂SO₄ 10 N (Carpenter et al., 2005). For analysis of δ^{13} C-POC and DOC, 250 ml of water were filtered through a mesh GF/F (0.7 μ m). For δ^{13} C-POC, we scraped the filter residue, which was then oven-dried at 40 °C. δ^{13} C-DOC was obtained after evaporating the water obtained from the filtrate at 40 °C (Pace *et al.*, 2007).

171 During the summer of 2011 (June-July) samples were taken along a longitudinal 172 gradient to cover the environmental spatial heterogeneity of the wetland (Fig. 1), including 173 inland areas (n = 7 sites; A1: A7), coastal areas (n = 5 sites, B1: B5), channels (n = 3 sites, C1: 174 C3), and deep zones (n = 2 sites, D1: D2; Fig. 1). M1 is the channel where the wastewater 175 from Villarrubia de los Ojos discharged after treatment. The interior areas are shallow (about 176 50-70 cm deep) and emerging macrophyte patches with vegetation-free zones can be found. 177 The coastal areas were usually covered with emergent macrophytes, with reeds and tamarisk 178 predominantly present. These areas were drying up over the dry season. The deepest areas 179 correspond to the terminal part of the wetland, with depths of 3-4 m, with emergent 180 vegetation restricted to the shoreline and without submerged macrophytes.

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182 2.3 Sample preparation and analysis

We removed carbonates from *Chara hispida* var. *major* and *Ceratophyllum submersum* with 5% HCl (Demopoulos, Fry & Smith, 2007). Carbonates of sediment were removed using 1M HCl (Demopoulos *et al.*, 2007). Subsequently, all samples were washed repeatedly with distilled water and successively oven-dried at 40 °C (invertebrates), 60 °C (vegetation; Boon & Bunn 1994) and 80 °C (detritus, phytoplankton and zooplankton). Subsequently all plant, detrital and invertebrate samples were crushed mechanically and manually, with the help of an agate mortar, and were separated using a 1 mm mesh.

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191 2.4 Stable isotope analysis

The analyses of the ratio of ^{12/13}C and ^{14/15}N stable isotopes were determined by Isotope Ratios Mass Spectrometry (IRMS) in the Stable Isotope Laboratory at the University of Arizona, United States (http://www.geo.arizona.edu/research/iso_lab.html) Two standards were used: Peedee belemnite (PDB) for C and atmospheric N₂ for N. The results are expressed in parts per thousand (‰) with respect to differences with the standards according to the following equation: δ^{13} C or δ^{15} N (‰) = [(R_{sample} / R_{standar}) - 1] x 10³, where R = ¹³C /¹²C and ¹⁵N /¹⁴N. The analytical precision of the replicas of the analysis was less than 0.2 ‰.

200 2.5. Statistical analysis

Differences in $\delta^{13}C/\delta^{15}N$ of organic sources were evaluated according to two criteria, for 201 202 which we used ANOVAs. (1) The position of each sampling point was considered to 203 determine if there were differences between the different habitats studied, i.e. the (i) inner 204 zone subjected to the influence of municipal wastewater discharges, (ii) littoral, (iii) channels and (iii) deep zones. (2) To evaluate the effects of wastewater discharges in $\delta^{13}C/\delta^{15}N$ -organic 205 206 sources we considered the following categories: (i) entrance, (ii) discharge area of Gigüela 207 river, (ii) intermediate zone, and (iii) end zone (Navarro bridge dam). Finally, the contribution of autochthonous sources of C in the secondary production was determined using the 208 Bayesian model SISUS in R (Erhardt, 2010). 209

210

211 **3. Results**

212 The results of δ^{13} C and δ^{15} N values of the major C sources from NPTD and the relative 213 position of each organism is represented in Fig. 2.

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215 $\delta^{13}C$ and $\delta^{15}N$ of nonliving reservoirs

Non-living C reservoirs of the wetland showed wide variation in ¹³C. δ^{13} C values of DOC were highly enriched (average -8.83‰ ± 2.78; Fig. 2) and there were no significant differences in terms of their spatial position (df=2, F=3.562, *p*=0.096). Instead, significant differences related to the effect of wastewater discharges were associated with less negative δ^{13} C values of DOC near the entry point (df=3, F=6.053, *p*=0.010). Meanwhile, the δ^{13} C of POC was greatly depleted (-26.89 ± 4.36 ‰), showing very homogeneous signatures around the wetland.

223 DON and PON showed similar isotopic signatures (average: $7.66 \pm 1.98\%$ and $7.82 \pm 3.02\%$ respectively). δ^{15} N values of DON values showed no significant differences according 225 to their spatial position (df=2, F=0.944, *p*=0.440), but did show significant differences related 226 to the presence of wastewater discharge that were related to more enriched values near the 227 entry point (df=3, F=17.035, *p*<0.001).

Sedimental detritus had δ^{13} C values that were less depleted than δ^{13} C of POC (average -17.96 ± 3.52 ‰; Table 2), showing no significant differences according to the criteria studied. However, δ^{15} N of sedimental detritus (average: 7.39±2.44‰) did vary depending on the effect of wastewater discharge, being also more positive near the entry point (df=3, F=3.897, *p*=0.040). δ^{13} C of DIC were on average -7.74±0.87 ‰ and only showed significant differences depending on the spatial location (df=2, F=4.473, *p*=0.020).

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235 3.2 $\delta^{13}C$ and $\delta^{15}N$ values of the living reservoirs

Regarding primary producers, charophyte showed enriched δ^{13} C values (-14.94 ± 0.43 ‰; Table 2), while emerging macrophytes presented depleted δ^{13} C values (\approx -28 ‰). Regarding δ^{15} N values, the lowest average values were those of *T. canariensis* (4.85 ± 1.12 ‰) and the highest by *S. maritimus* (11.03 ± 2.88‰). Only phytoplankton showed significant spatial differences in δ^{15} N (*p*=0.028, Table 3). The effect of wastewater discharge in δ^{15} N of primary producers was not significant, although the δ^{15} N of epiphyton was marginally affected by the distance to the water inlet (R²=0.5174; p=0.057). The effect of wastewater on δ^{13} C of primary producers was not significant. Secondary production was depleted in δ^{13} C, mainly in the case of invertebrates (-27.80 ‰), with zooplankton having slightly more enriched values (average: -23.50±5.43‰). In contrast δ^{15} N of secondary producers was very rich, especially that of zooplankton.

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3.4 Contribution of C sources to wetland organic matter, primary producers and secondary
production

Using SISUS, we showed that *T. canariensis* was the main source contributing to POM (69-84%), followed by the roots of the most abundant emergent macrophytes (\approx 19%) and, with similar contribution, *P. australis*, *C. mariscus* and *T. dominguensis* (\approx 15%; Table 2). According to SISUS, C sources that contribute to wetland secondary production vary widely. *C. submersum* appears as the main diet of zooplankton, although POC and microbial mats also represent a significant contribution (Table 2. *T. canariensis* appeared to be the main diet of invertebrates (Table 2).

257

258 4. Discussion

In this study, the δ^{13} C of both C living and nonliving reservoirs were more dependent on the physical-chemical characteristics of water and sediments than on other habitat features, while the effects of wastewater discharges mainly affected the isotopic signature of the dissolved fraction of organic matter and sediment detritus. Regarding δ^{15} N, primary producers such as epiphyton, filamentous algae and *S. maritimus* did show a clear decrease as we moved away from the entry point as seen in other systems influenced by wastewater discharge (Costanzo *et al.*, 2001). These results suggest that while both δ^{13} C of both C living and nonliving 266 reservoirs are good markers of environmental conditions, $\delta^{15}N$ of primary producers are 267 better indicators of disturbance, thus providing complementary information.

The isotopic fractionation that occurred in phytoplankton during photosynthesis in the 268 PNTD is outside the normal range of observation (-13.4 $\% \delta^{13}$ C in our case, -20 to 30 %; 269 270 Jones et al., 1998), which can be associated with the composition of phytoplankton species, temperature and even the ability to use HCO₃⁻ by algae, with greater affinity for δ^{13} C and, 271 thus, with less fractionation (Bade *et al.*, 2006). Furthermore, δ^{13} C of aquatic plant values 272 273 reveal a distinctive C fixation pathway according to the species: while macrophytes generally are in the range of C₃ plant values, C. hispida var. major and P. pectinatus have C₄ plant 274 signatures (Keeley, 1990). δ^{13} C of C. submersum values show a possible contamination 275 276 because carbonate values should be close to those of C₄ plants (Sensula et al., 2006). 277 Nevertheless, some authors like (Raz-Guzman, A. & L.A., 1992) do not considered it feasible 278 to categorize aquatic vegetation exclusively as C_3 or C_4 because the relationship between δ^{13} C and the photosynthetic mechanism is not as strict as in the case of terrestrial vegetation, 279 280 and also aquatic vegetation reflects HCO_3^- isotopic composition. And this, in turn, depends on 281 the salinity, which can interfere with the signatures and obscure the carboxylation process.

282 In the PNTD, previous studies determined that the emergent vegetation can contribute 283 up to 65% organic matter deposited on the wetland (autochthonous organic matter; Sánchez-284 Carrillo et al. 2001). Our results indicate that the main contribution to POM was from T. 285 canariensis, while emerging and submerged macrophytes represented less than 20%. Since 286 most of the carbon containing macrophytes is recalcitrant (Mann, 1988), it is expected that 287 most of these plant remains entering the detrital pool will eventually be degraded by fungi 288 and bacteria (Kuehn et al., 2000). However, the conditions prevailing in the years prior to 289 sampling (2006-2010) as well as the biomass removed might be masking the actual 290 contribution of emergent macrophytes to the detritus in the system.

291 Some studies show that *Tamarix* sp. has a low litter quality, which tends to reduce the biodiversity of macroinvertebrates and increases its accumulation in sediments (Bailey, 292 Schweitzer & Whitham, 2001). The strong spread of this species in the park during the dry 293 294 years prior to sampling could be masking their actual effects during wet cycles when it is 295 confined to the margins of the wetland (Bailey et al., 2001). POM showed isotopic signatures 296 consistent with macrophytes, which may indicate that their importance probably varies 297 temporarily and needs to be re-evaluated in a long dry-wet cycle. In any case, the importance 298 of *T. canariensis* in secondary production is surprising, despite its low nutritional value.

299 The Chironomidae family, the most abundant of the benthic macroinvertebrates during our study, was found to be related to Tamarix sp. seeds (Campobasso, 2017) and with 300 301 some algae species (Maasri et al., 2008). The proliferation of T. canariensis during the dry 302 years has resulted in that, for many areas, it has been the dominant riparian plant species. 303 This could have increased its importance for secondary production, which should have 304 become dominated by macroinvertebrates that tolerate its low litter quality (Bailey et al., 305 2001). Other species such as C. mariscus, emblematic of Daimiel, show a very low contribution to POM (less than 20%), although they represent a relatively important energy 306 307 source for zooplankton (up to 47.17%), probably as a result of a drastic reduction in coverage suffered during the last 50 years (Cirujano et al., 2010). Instead, P. australis, which has 308 309 increased by twenty-two times since 1950 (Alvarez-Cobelas et al., 2001), but does not 310 contribute significantly to the POM, or invertebrates (less than 22%) can represent up to 54% 311 zooplankton's diet. Gratton & Denno (2006) demonstrated in a coastal wetland that the large 312 biomass of P. australis was not functionally integrated in the local food web compared with 313 other plants such as Spartina. The small contribution of P. australis in Las Tablas is 314 paradoxical considering the great amount of biomass generated annually (M.D. Ribeiro,

315 2005). We speculate that this matter is quickly assimilated by microorganisms, but that it does316 not represent a significant of macroinvertebrates's diet.

317 The direct contribution of phytoplankton to zooplankton was high (up to 68%). POM 318 was the main zooplankton's diet, confirming what has been observed in other aquatic 319 ecosystems, even from other latitudes (Jones et al., 1999; Grey, Jones & Sleep, 2000). 320 However, in the NPTD, δ^{13} C of zooplankton were more enriched than those of POM, 321 contradictory to most freshwater ecosystems (depleted in comparison with the POM; Grey & 322 Jones 1999; Jones et al. 1999). Isotopic relations between zooplankton and food sources can 323 be masked by the dominance of a particular species, as well as the season and the habitat 324 where this is collected (Grey & Jones, 1999; Grey et al., 2000)

325 In this study, we showed that invertebrates trophically depend on the vegetation but 326 not on macroalgae, contrary to the results of Finlay, Khandwala & Power (2002) but similar 327 to Bunn et al. (2003) in an Australian stream. Finlay et al. (2002) found that most consumer invertebrates (e.g., mayflies, caddisflies, blackflies, Hydropsychidae, stoneflies, Hemiptera 328 329 and Odonata) depended on algal production, mainly composed by *Cladophora glomerata*. In 330 NPTD, it is possible that dry cycles modify macroinvertebrate trophic patterns or that in this 331 category of semiarid lentic systems the macroalgae's contribution to secondary production is 332 lower than expected. In any case, our results should be strengthened with more isotopic 333 studies in different hydrological and environmental conditions to capture all the possible 334 variability of sources and trophic processes.

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Cili-u
3-/L µg/L
.01 8.72±4.46
.01 6.00±4.36
.01 22.69±17.24
.00 2.84±2.87
.83 80.62±96.33
$\frac{4}{0}$ $\frac{1}{0}$ $\frac{1}{0}$

Table 1. Physicochemical characteristics of the five sampling zones in June 2011.

Table 2. Range of contribution (%) of the main C sources (primary producers and sediment

detritus) to particulate organic matter (POM) and secondary production in summer 2011,

482 based on SISUS.

	POM		Inverte	ebrates	Zooplankton		
	Min	Max	Min	Max	Min	Max	
Sedimen detritus	0	2.89	0.00	3.87	0.00	46.28	
DOC (intermediate zone)			0.00	2.02	0.00	24.29	
DOC (end zone)			0.00	2.08	0.00	24.92	
DOC (entrance zone)			0.00	1.74	0.00	20.83	
POC			0.00	29.08	0.00	80.64	
Phytoplankton	0.00	4.23	0.00	5.66	0.00	67.90	
Periphyton	0.00	4.97	0.00	6.64	0.00	63.54	
filamentous algae	0.00	5.49	0.00	7.35	0.00	63.83	
Microbial mats	0.00	6.40	0.00	8.55	0.00	77.87	
C. hispida var. major	0.00	2.23	0.00	2.98	0.00	35.71	
C. submersum	0.00	8.60	0.00	11.50	0.00	92.35	
P. pectinatus	0.00	3.14	0.00	4.20	0.00	50.37	
S. maritimus	0.00	14.58	0.00	20.41	0.00	48.06	
T. dominguensis	0.00	15.52	0.00	21.75	0.00	51.21	
C. mariscus	0.00	15.67	0.00	20.95	0.00	47.17	
P. australis	0.00	16.21	0.00	21.66	0.00	54.00	
T. canariensis	69.46	83.74	59.16	78.71	0.00	41.22	
Roots	0.00	19.02					

- **Fig. 1.** Location of "National Park Las Tablas de Daimiel" and sampling sites. Inland areas (n 486 = 7 sites; A1: A7), littorals (n = 5 sites, B1: B5), channels (n = 3 sites, C1: C3), and deep 487 zones (n = 2 sites, D1: D2; Fig. 1). M1 is the channel where the wastewater from "Villarrubia 488 de los Ojos" discharges after treatment.
- 489



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Fig. 2. $\delta^{13}C$ and $\delta^{15}N$ values of the main C sources and secondary production (invertebrates 500 501 and zooplankton) at NPTD. Errors bars are SE for the same group across habitats.

