The aerobic scope protection hypothesis: a mechanism explainingreduced growth of ectotherms in warming environments?

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Jutfelt F¹, Norin T², Åsheim ER^{1,3}, Rowsey LE⁴, Andreassen AH¹, Morgan R¹, Clark TD⁵,
Speers-Roesch B⁴

8 ¹Department of Biology, Norwegian University of Science and Technology, 7491 Trondheim, Norway.

9 ²DTU Aqua: National Institute of Aquatic Resources, Technical University of Denmark, 2800 Kgs.

- 10 Lyngby, Denmark.
- ³Organismal and Evolutionary Biology Research Program, University of Helsinki, 00014 Helsinki, Finland.
- ⁴Department of Biological Sciences, University of New Brunswick, Saint John, NB, Canada, E2L 4L5.
- ⁵Deakin University, School of Life and Environmental Sciences, Geelong, Victoria 3216, Australia.
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17 Abstract

18 Temperature has a dramatic effect on the physiology of ectothermic animals, impacting most 19 of their biology. When temperatures increase above optimal for an animal, their growth rate 20 tends to decrease. The mechanism behind this growth rate reduction is unknown. Here, we 21 suggest the aerobic scope protection hypothesis as a mechanistic explanation for the reduction 22 in growth. After a meal, metabolic rate, and hence oxygen consumption rate, transiently 23 increases in a process called specific dynamic action (SDA). At warmer temperatures, the SDA 24 response becomes temporally compressed, leading to a higher peak oxygen consumption rate. 25 This peak in oxygen consumption rate takes up much of the animal's aerobic scope (the 26 difference between maximum and resting rates of oxygen consumption), leaving little residual 27 aerobic scope for other functions. We propose that animals will actively protect their 28 postprandial residual aerobic scope by reducing meal sizes in order to regulate the peak SDA 29 response. This hypothesis is consistent with the published literature and we suggest further 30 predictions to test it.

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43 Temperature and oxygen delivery capacity

44 The body temperature of ectothermic animals generally follows the ambient temperature. As 45 biochemical reaction rates are temperature sensitive, biological rates in these animals are partly 46 under the control of the environment. Animals have evolved to function across a finite range of 47 suitable temperatures, above and below which physiological functions can suffer. Climate 48 warming is causing increased temperatures worldwide and is threatening the function of ectothermic animals (Pinsky et al., 2019). One common observation is that when temperatures 49 50 increase above suitable, growth rate and other performance metrics gradually or abruptly 51 decline (Huey and Stevenson, 1979); Gräns et al., 2014). The precise mechanisms behind these 52 decreases in performance are, however, subject to active debate (Clark et al., 2013; Jutfelt et 53 al., 2014; Jutfelt et al., 2018; Pörtner et al., 2017; Schulte, 2015).

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55 Thermal constraint on oxygen delivery to the body has been proposed as a function that can 56 limit performance, particularly in water-breathing animals (Brett, 1972; Fry, 1947; Pörtner and 57 Knust, 2007; Pörtner and Farrell, 2008), a hypothesis called the 'Fry paradigm' or 'oxygen- and 58 capacity-limited thermal tolerance' (OCLTT). As temperatures increase above suitable, the 59 standard (maintenance) metabolic rate (SMR) is suggested to increase faster than the aerobic maximum metabolic rate (MMR) leading to a reduction in the difference between them (i.e. the 60 61 aerobic scope). Animals without sufficient aerobic scope are faced with trade-offs in their 62 allocation of energy to activity, digestion, growth, and reproduction (Weiner, 1992), and may 63 even be forced into unsustainable anaerobic metabolism when aerobic scope is most 64 compromised, which is suggested to cause unfavourable downstream consequences in 65 functions such as growth (Pörtner and Farrell, 2008; Pörtner and Knust, 2007).

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67 As the OCLTT hypothesis has been suggested to explain the mechanisms of climate change 68 impacts on water-breathers (Pörtner and Farrell, 2008), it has received great interdisciplinary 69 interest. Several tests of predictions derived from the hypothesis have, however, failed to find 70 support for it (Jutfelt et al., 2018; Verberk et al., 2016). One prediction, that tissue hypoxia 71 restricts growth (Pörtner and Farrell, 2008; Pörtner and Knust, 2007) and therefore that markers 72 of anaerobic metabolism should be detectable at supra-optimal temperatures, has only limited 73 support, as markers for hypoxia generally only appear at or close to lethal temperatures (Pörtner 74 and Knust, 2007; Verberk et al., 2016). Tissue hypoxia is therefore unlikely to explain growth 75 reductions that appear at temperatures far below lethal (Clark et al., 2013; Jutfelt et al., 2018). 76 This mismatch between hypothesis and empirical data remains a major impediment in thermal 77 biology, hampering our ability to model and predict the impacts of climate change on 78 ectothermic animals.

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Here, we propose a possible resolution to the mismatch between the OCLTT hypothesis and the lack of empirical support for some of its predictions about growth. In fishes, reductions in growth at supra-optimal temperatures have long been ascribed to increases in SMR combined with reduced appetite (Jobling, 1997). We hypothesise that this reduction in appetite occurs voluntarily in order to conserve a portion of the aerobic scope that is available to the animal on top of the cost of digestion and assimilation. This 'aerobic scope protection hypothesis' has been briefly alluded to in the literature (e.g. Jobling, (1997); Norin and Clark, (2017)), but it
has not been fully articulated or explored.

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90 Aerobic scope protection by reduction of food intake

At supra-optimal acclimation temperatures, most fishes exhibit a decrease in food consumption 91 92 (Figure 1), likely due to a loss of appetite. The reduced food intake, combined with elevated SMR at high temperatures, results in reduced growth (Jobling, 1997). To investigate the nature 93 94 of this temperature-appetite relationship, we compiled data from studies investigating the thermal effect on voluntary food intake in fishes. The emerging pattern revealed large 95 96 interspecific variation in thermal sensitivity (Figure 1), likely related to different acclimation 97 capacities of the different species. However, there was a clear pattern of slowly increasing food 98 intake up to an optimum temperature, reflecting the species' maximum food intake, followed 99 by sharply reduced food intake at temperatures above optimal. While the increasing food intake 100 can be attributed to a temperature-induced increase in maintenance metabolism (i.e. SMR), the 101 reason for the loss of appetite at supra-optimal acclimation temperatures is likely a deficit or 102 declining functioning of one or several physiological systems, but the precise system or 103 mechanism has remained elusive.





Peak-centred temperature (°C)

106 Figure 1. Food intake as a function of temperature. Food intake temperature profiles normalised as percent 107 of maximum intake, compiled from 14 studies on 11 species of fish from tropical to cold temperate thermal 108 environments (A). To be able to compare shapes of the food intake temperature profiles between species, all 109 profiles were centred around the temperature of peak food intake (highest data point for each species) for 110 each species (A and B). Overlapping points at maximum food intake are replaced with a black square. Data 111 for each species were fitted with a second-degree polynomial regression in (A). In (B) all data are combined, 112 and a third-degree polynomial regression shows the shape of the influence of temperature on food intake. 113 Data are from (Fernández-Montero et al., 2017; Fonds et al., 1992; Handeland et al., 2008; Hofmann and 114 Fischer, 2003; Kinne, 1960; Koskela et al., 1997; Mesa et al., 2013; Ojanguren et al., 2001; Remen et al., 115 2016; Schram et al., 2013; Sun and Chen, 2009; Sun and Chen, 2014; Xie et al., 2010; Zanuzzo et al., 2019).

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117 When animals eat a meal, their metabolic rate initially increases sharply as part of the digestive 118 response and remains elevated for an extended period (Figure 2), before eventually returning 119 to baseline levels (i.e. SMR in a resting ectotherm). This increase in post-prandial oxygen 120 consumption rate is termed 'specific dynamic action' (SDA) and is considered a product of 121 increased activity in the gut as well as increased biochemical nutrient processing and tissue 122 assimilation of nutrients (Secor, 2009). The SDA response is temperature sensitive and can last 123 a week or two in cold-living Antarctic fishes (Johnston and Battram, 1993); (Boyce and Clarke, 124 1997), but is much shorter in duration at warmer temperatures in animals eating similar-sized 125 meals (Secor, 2009; Secor, 2011). As the total amount of energy expended to digest and 126 assimilate a meal of a certain size (the area under the SDA curve) often is independent of 127 temperature (Secor, 2009), the shorter duration of the SDA response at warm temperatures 128 results in a greater post-prandial peak in metabolic rate (Figure 2) (Secor, 2009; Secor, 2011). 129 This peak can occupy the majority of an animal's aerobic scope. For example, in the short-horn 130 sculpin (Myoxocephalus scorpius), a week of warming from 10 to 16°C did not change the 131 aerobic scope but elevated the post-prandial peak in metabolic rate up to MMR (Sandblom et 132 al., 2014), leaving no aerobic scope for activities other than SDA. As these fish were gavage 133 fed a meal corresponding to 5% of their body mass, they did not have the option to regulate 134 their meal size.





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138 Figure 2. Conceptual illustration of the effect of temperature on metabolic rate and SDA in ectotherms. 139 Standard metabolic rate (SMR) and maximum metabolic rate (MMR) increase with temperature. The 140 metabolic increase after a same-sized meal (SDA) is shown as a time profile at three different temperatures 141 ('suitable' in blue, 'warm' in purple, and 'hot' in red). Increasing temperature temporally compresses the 142 SDA response while giving rise to a higher post-prandial peak metabolic rate. At the warm temperature, the 143 post-prandial residual aerobic scope (PRAS) is reduced compared to the suitable temperature, potentially 144 limiting other oxygen-demanding processes. At the hot temperature, PRAS is gone and the post-prandial 145 peak metabolic rate overshoots the maximum oxygen delivery capacity, forcing the animal to exploit its 146 anaerobic metabolism (hatched area above MMR) and creating a costly oxygen debt. We propose that water-147 breathers actively avoid this latter situation by reducing meal sizes to maintain a sufficient PRAS for activities 148 other than digestion.

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150 Meal size is a primary determinant of the magnitude of the SDA response (Norin and Clark, 151 2017; Secor, 2009), which provides a potential mechanism for fishes and other animals to 152 voluntarily regulate their SDA magnitude by eating relatively large or small meals. Previous 153 research has shown that fish eating larger meals grow more efficiently but also incur a relatively 154 high post-prandial peak in metabolic rate during digestion of the larger meals (Norin and Clark, 155 2017), which reduces the amount of aerobic scope available for other oxygen-demanding 156 activities such as swimming and collective behaviour (McLean et al., 2018). These findings 157 indicate that, ideally, animals should regulate their food intake based on their need to maintain 158 aerobic scope under challenging conditions. However, the interacting effects of temperature 159 and aerobic scope protection on meal size selectivity have not been explored. We propose that 160 animals use this mechanism of reduced food intake at elevated temperatures to conserve their 161 post-prandial residual aerobic scope (PRAS) within a species- and context-specific limit 162 (Figure 3).





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Figure 3. Aerobic scope protection by meal size reduction. At hot temperatures, the SDA after a large meal can comprise the full aerobic scope, and potentially force anaerobic metabolism (hatched area above MMR). By voluntarily reducing meal size, fishes and other ectothermic animals may reduce their post-prandial peak metabolic rate and thus maintain a sufficient post-prandial residual aerobic scope (PRAS) for other aerobic processes.

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173 Testing the aerobic scope protection hypothesis

174 The prediction derived from the OCLTT hypothesis, that tissue hypoxia and hypoxic markers 175 should occur at supra-optimal temperatures, has not been clearly supported and is even 176 contradicted by experimental data (Gräns et al., 2014; Pörtner and Knust, 2007). One resolution 177 to the lack of support for this OCLTT prediction may be that fish generally do not allow their 178 aerobic scope to be reduced to the point of insufficient oxygenation of their tissues, and the 179 aerobic scope protection hypothesis presented here could be the mechanism behind this. 180 Avoidance of tissue hypoxia, however, is more difficult to detect experimentally than actual 181 realised tissue hypoxia. Below, we summarise observations consistent with the aerobic scope 182 protection hypothesis, and we suggest paths for experimentally testing it.

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184 A review of the relationship between growth rate of fishes and their aerobic capacities found185 no consistent correlation (Blier et al., 1997). This seemingly contradicts our aerobic scope

protection hypothesis and indicates that aerobic capacity is not a major constraint to fish growth [but see (Auer et al., 2015)]. The investigation into the relationship between growth rate and aerobic capacity, however, did not include the full temperature range (sub- to supra-optimal) for the species included (Blier et al., 1997). We predict that temperature is a major interacting factor in this correlation due to its effect on the SDA response (cf. Figure 2), and that the benefit of having a higher oxygen transport capacity only becomes apparent at supra-optimal temperatures. This prediction would be interesting to test.

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194 If reductions in growth occur through a voluntary reduction in food intake that is implemented 195 to reserve a portion of the aerobic scope available for activities other than SDA, as proposed 196 here, it is predicted that the appetite of an animal would be more sensitive to water hypoxia in 197 warmer than in colder temperatures, since hypoxia decreases MMR while warmer temperatures 198 increase the post-prandial peak metabolic rate. Indeed, when channel catfish (Ictalurus 199 punctatus) were exposed to water hypoxia, their food intake was only marginally affected at 200 colder temperatures. However, at the warmest temperatures, even mild hypoxia (80-90% air 201 saturation) caused a large decrease in voluntary food intake, suggesting that the fish were highly attentive to the water oxygen levels (Buentello et al., 2000). Likewise, appetite in hypoxia was 202 203 highly sensitive to temperature in Atlantic salmon (Salmo salar); the fish maintained stable 204 maximum food intake from normoxia down to hypoxia (40% air saturation) at 7°C, but food 205 intake started to decline at oxygen levels of 75% air saturation at 19°C (Figure 4). This 206 demonstrates increasing oxygen limitation on appetite with increasing temperature, which is 207 consistent with our proposed aerobic scope protection mechanism. 208





Figure 4. Water oxygen levels required to sustain maximum food intake. While food intake gradually increased with increasing temperature in Atlantic salmon (*Salmo salar*) within the species' suitable temperature range (A), varying oxygen levels did not markedly affect feeding until a breakpoint oxygen level, which occurred closer to normoxia at higher temperatures (i.e. food intake was more insensitive to water oxygen levels at lower temperatures) (A, B). Redrawn from Remen et al., (2016).

217 If the reduction in PRAS becomes restrictive to food intake only at higher temperatures, then218 factors that increase aerobic scope can be predicted to also increase food intake. Hyperoxia has

219 been shown to increase the aerobic scope in fish; oxygen levels of 200% air saturation 220 effectively doubled the aerobic scope (Brijs et al., 2015) and increased venous oxygen partial 221 pressure and cardiac stroke volume in European perch (Perca fluviatilis) (Ekström et al., 2016). 222 As a higher aerobic scope would allow for a higher post-prandial peak metabolic rate during 223 the SDA response without markedly restricting PRAS or tissue oxygenation, we predict that 224 water hyperoxia would increase appetite. Additionally, this effect of hyperoxia would be 225 temperature sensitive, with no effect at suitable temperatures and increasing effect with 226 increasing supra-optimal temperatures. Few studies have tested this prediction: Rainbow trout 227 fed various diets at hypoxia (50% air saturation), normoxia (100% air saturation), and hyperoxia 228 (200% air saturation) varied in their weight gain in accordance with the prediction for 229 increasing appetite with increasing oxygen availability (Dabrowski et al., 2004), although 230 another study on rainbow trout did not find the same benefit of hyperoxia (Caldwell and 231 Hinshaw, 1994). Notably, the growth experiment by Dabrowski et al. (2004) was conducted at 232 20-22°C, which is above the temperature range for optimal growth in rainbow trout (Hokanson 233 et al., 1977). This supports our prediction of an increasing effect of hyperoxia at supra-optimal 234 temperatures, but temperature has not yet been systematically included as a factor.

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With supra-optimal temperatures and temporal compression of the SDA response, animals may
face a trade-off between using their aerobic scope for SDA or for mobility. This trade-off can
be quantified and would be predicted to be more obvious at warmer temperatures. The tradeoff could be detected as both a decrease in spontaneous activity and/or a decrease in meal size.

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241 Beyond its dependence on meal size, the SDA response is also highly variable between species 242 (Secor, 2009). An animal's lifestyle influences the amount of aerobic scope taken up by its 243 SDA. For example, among fishes, active swimmers tend to show a less pronounced SDA 244 response than sit-and-wait ambush predators (Clark et al., 2013; Fu et al., 2009a; Fu et al., 245 2009b; Secor, 2009). Therefore, it could be predicted that ambush predators, with their larger 246 SDA, would be more likely to reduce their food intake to conserve PRAS at elevated 247 temperatures, compared to more active fishes. However, ambush predators also require less 248 PRAS than active swimmers (Clark et al. 2013), so, in the absence of experimental work, it is 249 difficult to predict which lifestyle, if any, would show the more pronounced decrease in 250 appetite.

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252 Anecdotal, yet interesting, evidence for immediate PRAS limitation comes from an observation 253 by Claireaux et al. (Claireaux et al., 2000) who placed recently fed Atlantic cod (Gadus 254 morhua) in respirometers and gradually reduced water oxygen saturation. At the point where 255 the cod's PRAS approached zero, they terminated digestion, as colourfully described by the 256 authors: "At this point, scope for activity being almost nil, all the fish in the measuring chamber 257 simultaneously regurgitated their food, presumably to reduce their immediate oxygen 258 requirements. Then, as soon as water S_{O2} [water oxygen saturation] was restored to above 45%, 259 approximately 5 min later, the fish recovered enough metabolic potential to resume digestion 260 and re-swallowed all the food." This suggests that fish monitor not only external oxygen levels, 261 but also somehow are attentive to the magnitude of their aerobic scope and SDA, and that they 262 act to avoid an aerobic deficit (cf. Figure 3) by controlling their SDA.

263 264 The predictions derived from the aerobic scope protection hypothesis presented here are similar 265 to what the OCLTT hypothesis predicts, in that oxygen transport is limiting thermal 266 performance. It could even be argued that the mechanism suggested here is inherent to the larger 267 OCLTT framework. The mechanism of voluntary meal size reductions, however, has not 268 previously been explicitly proposed, and there are several differences between OCLTT and the 269 aerobic scope protection hypothesis that can be experimentally tested and therefore used to 270 discriminate between the two hypotheses. The OCLTT hypothesis has a focus on 271 cardiorespiratory function, oxygen delivery to the tissues, and (in extreme conditions) tissue 272 hypoxia as the limiting constraints to performance at higher temperatures. The difference 273 proposed here is that cardiorespiratory oxygen limitations only develop at relatively high 274 temperatures if the animal allows its SDA to occupy the majority of its aerobic scope and, to 275 avoid this, the animal should reduce its appetite at temperatures where the postprandial residual 276 aerobic scope may become limited. This loss of appetite is the main mechanism that can be 277 used to differentiate our aerobic scope protection hypothesis from the OCLTT hypothesis. In 278 Figure 5, we present a range of predictions that would be consistent with voluntary food intake 279 restriction and aerobic scope protection. These can be readily tested, and some of them already 280 have some support in the literature as outlined above. The figures show the main direction of 281 change in a suite of attributes related to feeding or physical activity, but the exact shape of the 282 illustrated relationships will be more complex and context-dependent. A more specific 283 prediction is made in Figure 5I, where the shape of the temperature-appetite function from 284 Figure 1 is used (green line) and the predicted effect of hyperoxic water is added (blue line). 285

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Figure 5. Predictions derived from the aerobic scope protection hypothesis. (A) Food intake decreases 290 with increasing supra-optimal temperature in normoxic water, but less so in hyperoxic water as it can increase aerobic scope (Brijs et al., 2015) and thus PRAS. (B) Growth rate is suggested to show a similar pattern to 292 food intake at supra-optimal temperatures, as increased food intake in hyperoxia is assumed to result in higher 293 growth rate. (C) Voluntary activity is predicted to decline with increasing food rations at high temperatures, 294 but hyperoxia may ameliorate this decline by increasing aerobic scope and thus PRAS. (D) Voluntary activity 295 often increases with temperature (up to a point), but activity competes with digestion for the available aerobic 296 scope, leading to a lower increase in activity with temperature in fed compared to unfed animals. (E) Animals 297 that are force-fed a large meal should show reduced locomotor performance compared to unfed or voluntarily 298 feeding animals at supra-optimal temperature but not at sub-optimal temperature. (F) Animals may estimate 299 their need for activity in the near future (e.g. escaping predators, migrating, navigating currents), and 300 increased perceived risk for activity may reduce food intake to maintain a larger PRAS at supra-optimal 301 temperature. (G) Aerobic capacity generally is not considered to correlate with growth rate (but see (Auer et 302 al., 2015)), although this should be temperature-dependent and a positive correlation is predicted at warmer 303 temperatures. (H) The minimum oxygen level required for maintaining maximum food intake or appetite 304 (Min O_{2sat} for max app) increases with temperature (cf. Figure 4). If the animal predicts an increased risk of 305 physical activity, the increase with temperature should be higher than in the absence of risk of activity. (I) 306 The peak-centred shape of the temperature-food intake relationship (cf. Figure 1) should differ between 307 normoxia (green line) and hyperoxia (blue line).

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309 Finally, we caution that the aerobic scope protection hypothesis is likely not a silver bullet 310 mechanism to understand performance limitation at high temperature, but only one potential 311 explanation among many for the observed patterns of thermal performance in ectothermic 312 animals. For example, it can be assumed that the gastrointestinal tract is adapted to suit the food 313 type and amount associated with the species' niche. With warming, the increased energy 314 expenditure and therefore food throughput requirement may cause the gastrointestinal capacity 315 to limit nutrient uptake and thus growth (McLeod and Clark, 2016). The immune system is 316 tailored to the risk of infection and cell damage (e.g. through oxidative stress or thermal increase 317 in turnover), but these functions may become restrictive at higher temperatures (Dittmar et al., 318 2014; Miest et al., 2019; Wegner et al., 2008). Nerve function is optimised for a certain thermal 319 range through ion channel function and membrane fluidity (Cossins et al., 1977). Gonads can 320 be more thermally sensitive than other organs (Pankhurst and King, 2010). Mitochondrial 321 proton leak increases with temperature (Leo et al., 2017), meaning more food and oxygen is 322 consumed per unit of ATP produced. Such limitations can likely occur for every physiological 323 organ system. Therefore, oxygen delivery is just one permissive factor that needs to be in place 324 for the organism to function at higher temperatures. There may be many such permissive 325 factors, both physiological and ecological, any of which can become limiting for fitness in 326 certain contexts (e.g. limitations in food, water, ions, refuges). Such permissive factors may 327 become saturated at some high level (e.g. more food will not increase growth over a certain 328 maximum food-handling level). Likewise, oxygen delivery is one permissive factor to tissue 329 performance among many that may or may not become limiting under thermal challenges, and 330 whether it does is context specific. The task for ecophysiologists now is to figure out ways of 331 testing which factors are limiting which animals and in which contexts; a task that will likely 332 keep us busy for some time.

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350 Author contribution

F.J. conceived of the initial ideas and wrote the first manuscript draft. F.J., B.S.R., L.E.R., and
E.R.Å. collected data and produced figures. All authors discussed the ideas and helped write

- the manuscript.
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355 References

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- Auer, S. K., Salin, K., Anderson, G. J. and Metcalfe, N. B. (2015). Aerobic scope explains
 individual variation in feeding capacity. *Biol. Lett.* 11, 20150793–3.
- Blier, P. U., Pelletier, D. and Dutil, J. D. (1997). Does aerobic capacity set a limit on fish growth rate? *Rev. Fish. Sci.* 5, 323–340.
- Brijs, J., Jutfelt, F., Clark, T. D., Grans, A., Ekström, A. and Sandblom, E. (2015).
 Experimental manipulations of tissue oxygen supply do not affect warming tolerance of European perch. J. Exp. Biol. 218, 2448–2454.
- Buentello, J. A., Gatlin, D. M., III and Neill, W. H. (2000). Effects of water temperature
 and dissolved oxygen on daily feed consumption, feed utilization and growth of channel
 catfish (Ictalurus punctatus). *Aquaculture* 182, 339–352.
- 367 Caldwell, C. A. and Hinshaw, J. (1994). Physiological and haematological responses in
 368 rainbow trout subjected to supplemental dissolved oxygen in fish culture. *Aquaculture* 369 126, 183–193.
- 370 Claireaux, G., Webber, D. M. and Lagardère, J. P. (2000). Influence of water temperature
 and oxygenation on the aerobic metabolic scope of Atlantic cod (Gadus morhua). *Journal* 372 of Sea Research 44, 257–265.
- 373 Clark, T. D., Sandblom, E. and Jutfelt, F. (2013). Aerobic scope measurements of fishes in
 an era of climate change: respirometry, relevance and recommendations. *J. Exp. Biol.*375 216, 2771–2782.
- 376 Cossins, A. R., Friedlander, M. J. and Prosser, C. L. (1977). Correlations between
 377 behavioral temperature adaptations of goldfish and the viscosity and fatty acid
 378 composition of their synaptic membranes. *Journal of Comparative Physiology* A 120,
 379 109–121.
- 380 Dabrowski, K., Lee, K. J., Guz, L., Verlhac, V. and Gabaudan, J. (2004). Effects of
 381 dietary ascorbic acid on oxygen stress (hypoxia or hyperoxia), growth and tissue vitamin
 382 concentrations in juvenile rainbow trout (Oncorhynchus mykiss). *Aquaculture* 233, 383–392.
- 384 Ekström, A., Brijs, J., Clark, T. D., Gräns, A., Jutfelt, F. and Sandblom, E. (2016).
 385 Cardiac oxygen limitation during an acute thermal challenge in the European perch:
 386 effects of chronic environmental warming and experimental hyperoxia. *American Journal*387 of Physiology Regulatory, Integrative and Comparative Physiology 311, R440–R449.
- Fernández-Montero, A., Caballero, M. J., Torrecillas, S., Tuset, V. M., Lombarte, A.,
 Ginés, R. R., Izquierdo, M., Robaina, L. and Montero, D. (2017). Effect of
 temperature on growth performance of greater amberjack (SERIOLA DUMERILIRisso
 1810) Juveniles. Aquac Res 49, 908–918.
- Fonds, M., CRONIE, R., VETHAAK, A. D. and VANDERPUYL, P. (1992). Metabolism,
 Food-Consumption and Growth of Plaice (Pleuronectes-Platessa) and Flounder

- (Platichthys-Flesus) in Relation to Fish Size and Temperature. *Netherlands Journal of Sea Research* 29, 127–143.
- Fu, S. J., Zeng, L. Q., Li, X. M., Pang, X., Cao, Z. D., Peng, J. L. and Wang, Y. X.
 (2009a). The behavioural, digestive and metabolic characteristics of fishes with different foraging strategies. *J. Exp. Biol.* 212, 2296–2302.
- Fu, S.-J., Zeng, L.-Q., Li, X.-M., Pang, X., Cao, Z.-D., Peng, J.-L. and Wang, Y.-X.
 (2009b). Effect of meal size on excess post-exercise oxygen consumption in fishes with different locomotive and digestive performance. *Journal of Comparative Physiology B:* ... 179, 509–517.
- Handeland, S. O., Imsland, A. K. and Stefansson, S. O. (2008). The effect of temperature
 and fish size on growth, feed intake, food conversion efficiency and stomach evacuation
 rate of Atlantic salmon post-smolts. *Aquaculture* 283, 36–42.
- 406 Hofmann, N. and Fischer, P. (2003). Impact of temperature on food intake and growth in
 407 juvenile burbot. *Journal of Fish Biology* 63, 1295–1305.
- 408 Hokanson, K., KLEINER, C. F. and THORSLUND, T. W. (1977). Effects of Constant
 409 Temperatures and Diel Temperature-Fluctuations on Specific Growth and Mortality410 Rates and Yield of Juvenile Rainbow-Trout, Salmo-Gairdneri. *Journal of the Fisheries*411 *Research Board of Canada* 34, 639–648.
- 412 Huey, R. B. and Stevenson, R. D. (1979). Integrating thermal physiology and ecology of
 413 ectotherms: A discussion of approaches. *Integrative and Comparative Biology* 19, 357–
 414 366.
- Johnston, I. A. and Battram, J. (1993). Feeding energetics and metabolism in demersal fish
 species from Antarctic, temperate and tropical environments. *Mar Biol* 115, 7–14.
- Jutfelt, F., Gräns, A., Jönsson, E., Wiklander, K., Seth, H., Olsson, C., Dupont, S.,
 Ortega-Martinez, O., Sundell, K., Axelsson, M., et al. (2014). Response to 'How and how not to investigate the oxygen and capacity limitation of thermal tolerance (OCLTT) and aerobic scope—remarks on the article by Gräns et al.'. *J. Exp. Biol.* 217, 4433–4435.
- Jutfelt, F., Norin, T., Ern, R., Overgaard, J., Wang, T., McKenzie, D. J., Lefevre, S.,
 Nilsson, G. E., Metcalfe, N. B., Hickey, A. J. R., et al. (2018). Oxygen- and capacitylimited thermal tolerance: blurring ecology and physiology. *J. Exp. Biol.* 221.
- 424 Kinne, O. (1960). Growth, Food Intake, and Food Conversion in a Euryplastic Fish Exposed
 425 to Different Temperatures and Salinities. *Physiological zoology* 33, 288–317.
- 426 Koskela, J., Pirhonen, J. and Jobling, M. (1997). Feed intake, growth rate and body
 427 composition of juvenile Baltic salmon exposed to different constant temperatures.
 428 Aquaculture International 5, 351–360.
- Leo, E., Kunz, K. L., Schmidt, M., Storch, D., Pörtner, H. O. and Mark, F. C. (2017).
 Mitochondrial acclimation potential to ocean acidification and warming of Polar cod
 (Boreogadus saida) and Atlantic cod (Gadus morhua). *Front Zool* 14, 21.

- 432 McLean, S., Persson, A., Norin, T. and Killen, S. S. (2018). Metabolic Costs of Feeding
 433 Predictively Alter the Spatial Distribution of Individuals in Fish Schools. *Current Biology*434 28, 1144–1149.e4.
- 435 McLeod, I. M. and Clark, T. D. (2016). Limited Capacity for Faster Digestion in Larval
 436 Coral Reef Fish at an Elevated Temperature. *PLoS ONE* 11, e0155360–13.
- 437 Mesa, M. G., Weiland, L. K., Christiansen, H. E., Sauter, S. T. and Beauchamp, D. A.
- 438 (2013). Development and Evaluation of a Bioenergetics Model for Bull Trout.
 439 *Transactions of the American Fisheries Society* 142, 41–49.
- 440 Norin, T. and Clark, T. D. (2017). Fish face a trade-off between 'eating big' for growth
 441 efficiency and 'eating small' to retain aerobic capacity. *Biol. Lett.* 13, 20170298–4.
- 442 Ojanguren, A. F., Reyes-Gavilan, F. G. and Brana, F. (2001). Thermal sensitivity of
 443 growth, food intake and activity of juvenile brown trout. *Journal of thermal biology* 26,
 444 165–170.
- Pankhurst, N. W. and King, H. R. (2010). Temperature and salmonid reproduction:
 implications for aquaculture. *Journal of Fish Biology* 76, 69–85.
- Pinsky, M. L., Eikeset, A. M., McCauley, D. J., Payne, J. L. and Sunday, J. M. (2019).
 Greater vulnerability to warming of marine versus terrestrial ectotherms. *Nature* 1–20.
- 449 Pörtner, H. O. and Farrell, A. P. (2008). Physiology and climate change. *Science* 322, 690–
 450 692.
- 451 Pörtner, H. O. and Knust, R. (2007). Climate change affects marine fishes through the
 452 oxygen limitation of thermal tolerance. *Science* 315, 95–97.
- 453 Pörtner, H. O., Bock, C. and Mark, F. C. (2017). Oxygen- and capacity-limited thermal
 454 tolerance: bridging ecology and physiology. *J. Exp. Biol.* 220, 2685–2696.
- 455 Remen, M., Sievers, M., Torgersen, T. and Oppedal, F. (2016). The oxygen threshold for
 456 maximal feed intake of Atlantic salmon post-smolts is highly temperature-dependent.
 457 *Aquaculture* 464, 582–592.
- 458 Sandblom, E., Gräns, A., Axelsson, M. and Seth, H. (2014). Temperature acclimation rate
 459 of aerobic scope and feeding metabolism in fishes: implications in a thermally extreme
 460 future. *Proc. Biol. Sci.* 281, 20141490–20141490.
- 461 Schram, E., Bierman, S., Teal, L. R., Haenen, O., van de Vis, H. and Rijnsdorp, A. D.
 462 (2013). Thermal Preference of Juvenile Dover Sole (Solea solea) in Relation to Thermal
 463 Acclimation and Optimal Growth Temperature. *PLoS ONE* 8, e61357–15.
- 464 Schulte, P. M. (2015). The effects of temperature on aerobic metabolism: towards a
 465 mechanistic understanding of the responses of ectotherms to a changing environment. *J.*466 *Exp. Biol.* 218, 1856–1866.
- 467 Secor, S. M. (2009). Specific dynamic action: a review of the postprandial metabolic
 468 response. *Journal of Comparative Physiology B:* ... 179, 1–56.

- 469 Sun, L. and Chen, H. (2009). Effects of ration and temperature on growth, fecal production,
 470 nitrogenous excretion and energy budget of juvenile cobia (Rachycentron canadum).
 471 Aquaculture 292, 197–206.
- 472 Sun, L. and Chen, H. (2014). Effects of water temperature and fish size on growth and
 473 bioenergetics of cobia (Rachycentron canadum). *Aquaculture* 426-427, 172–180.
- 474 Verberk, W. C. E. P., Overgaard, J., Ern, R., Bayley, M., Wang, T., Boardman, L. and
- 475 Terblanche, J. S. (2016). Does oxygen limit thermal tolerance in arthropods? A critical
 476 review of current evidence. *Comparative Biochemistry and Physiology Part A:* ... 192,
 477 64–78.
- Weiner, J. (1992). Physiological limits to sustainable energy budgets in birds and mammals:
 Ecological implications. *Trends in Ecology & Evolution* 7, 384–388.
- 480 Xie, S., ZHENG, K., CHEN, J., ZHANG, Z., ZHU, X. and YANG, Y. (2010). Effect of
 481 water temperature on energy budget of Nile tilapia, Oreochromis niloticus. *Aquaculture*
- 482 *Nutrition* 17, e683–e690.
- 483 Zanuzzo, F. S., Bailey, J. A., Garber, A. F. and Gamperl, A. K. (2019). The acute and
- 484 incremental thermal tolerance of Atlantic cod (Gadus morhua) families under normoxia
- and mild hypoxia. *Comparative Biochemistry and Physiology Part A:* ... 233, 30–38.