

1 **Resurrection approach shows evolutionary changes in drought responses in**
2 **European plant species after two decades of climate change**

3 Robert Rauschkolb^{1,*}, Lisa Henres¹, Caroline Lou², Sandrine Godefroid³, Lara Dixon⁴,
4 Oliver Bossdorf¹, Andreas Ensslin⁵, J.F. Scheepens^{1,6}

5
6 ¹ Plant Evolutionary Ecology, Institute of Evolution and Ecology, University of Tübingen,
7 Auf der Morgenstelle 5, 72076 Tübingen, Germany

8 ² Department of Biology, McGill University, Montreal, Quebec, H3A 1B1 Canada

9 ³Meise Botanic Garden, Nieuwelaan 38, 1860 Meise, Belgium

10 ⁴Conservatoire Botanique National Méditerranéen de Porquerolles, 34 avenue
11 Gambetta, 83400 Hyères, France

12 ⁵ Conservatory and Botanic Garden of the City of Geneva, 1296 Chambésy, Geneva,
13 Switzerland

14 ⁶ Plant Ecology, Institute of Ecology, Evolution and Diversity, Goethe University
15 Frankfurt, Max-von-Laue-Str. 13, 60438 Frankfurt am Main, Germany

16 * Corresponding author: robert.rauschkolb@uni-tuebingen.de, +49 7071 2974248

17 ORCID IDs: R.R. 0000-0002-8569-8705; S.G. 0000-0002-1893-9249; O.B. 0000-0001-
18 7504-6511; A.E. 0000-0003-2922-5310; J.F.S. 0000-0003-1650-2008

19 **Running Title:** Resurrection approach shows rapid evolution of drought responses in
20 plants

21 **Keywords:** common garden experiments, multi-species experiments, phenotypic
22 plasticity, rapid evolution, seed banks, watering treatments

23 **Words:** 4355; two figures and two tables

24 **Supplementary material:** one figure and raw data as two txt-files

25 **Abstract**

26 Plant populations must continuously adapt to the impacts of ongoing global climate
27 change, including warmer temperatures and more extreme weather events. We can
28 detect such evolutionary changes within plant populations through the resurrection
29 approach whereby plants grown from seeds stored in seed banks (“ancestors”) are
30 compared to freshly collected seeds from the same populations (“descendants”) in
31 common garden experiments. In this study we used the resurrection approach in two
32 multi-species experiments to investigate changes in phenotypic traits and drought
33 tolerance of European plant species from two biogeographic regions. In the seedling
34 survival experiment using seedlings of four Mediterranean species, watering was
35 ceased and day of mortality recorded. We found that descendants survived significantly
36 longer without any watering but these seedlings were smaller than the ancestral
37 seedlings. In the watering response experiment we investigated phenotypic responses
38 to drought in adult plants of nine species originating from temperate climatic regions in
39 Europe. We found that descendant plants were significantly taller under well-watered
40 conditions but smaller under drought than their ancestors, thus showing stronger
41 plasticity. Our study suggests that plants have already evolved phenotypically, including
42 through changes in trait means and plasticity, within the last decades. The observed
43 evolutionary changes are consistent with adaptation to increased drought. More
44 generally, the resurrection approach proved to be a useful tool to study rapid
45 evolutionary processes in plants under climate change. Future studies should include
46 fitness measures and comparative transplantations of descendants and ancestors into

47 their original habitat to disentangle adaptive from non-adaptive responses to recent
48 climate change.

49

50 **Introduction**

51 Climate change has increased dramatically over the last decades (IPCC 2018), and
52 plant populations are already responding (Peñuelas and Filella 2001, Parmesan and
53 Yohe 2003). During the 21st century annual precipitation sums will further increase in
54 northern Europe and decrease in the south (IPCC 2013). For central and western
55 Europe, precipitation is expected to increase in winter and decrease in summer (IPCC
56 2013), leading to more droughts in the growing season. Moreover, the anticipated
57 higher temperatures will lead to higher evapotranspiration (Feng and Fu 2013). These
58 changes in environmental conditions will likely increase the frequency, duration and
59 geographic extent of drought events in Southern and Central Europe (Ruosteenoja et al
60 2018; Samaniego et al 2018; Spinoni et al 2018). Changes in water availability and
61 more frequent droughts are especially strong stressors for plants (Jaleel et al 2009;
62 Fleta-Soriano and Munné-Bosch 2016), and many plant populations may not be well
63 enough adapted to these novel environmental conditions (Anderson et al 2012; Shaw
64 and Etterson 2012). To avoid extinction, plant populations need to migrate to track
65 suitable conditions and so they may respond through phenotypic plasticity or adaptive
66 evolution, or both (Holt 1990; Hoffmann and Sgrò 2011). For populations to survive
67 drought events *in situ*, they may plastically or genetically adjust functional traits, which
68 could involve reduced growth (Kusaka et al 2005; Shao et al 2008), increased root-
69 shoot-ratio (Sharp and LeNoble 2002; Aroca 2012) or changes in the ratio of chlorophyll
70 a/b and carotenoids (Farooq et al 2009).

71 Since drought events are periodic, the ability to change functional trait values
72 through phenotypic plasticity may be a better strategy than to evolve constitutive

73 changes in mean traits (Sultan and Spencer 2002, Alpert and Simms 2002, Gianoli and
74 Valladares 2012), especially in environments with strong climatic variability. Studies on
75 the effects of climate change on plant populations often only consider changes in mean
76 climate conditions (Bertrand et al 2011), but there is strong evidence for increased
77 climatic variability – specifically more heavy rain events followed by longer dry periods
78 in many regions (Kharin et al 2007). It is known that changes in environmental variability
79 may constitute distinct selection pressures and result in adaptive evolution of plasticity.
80 For example, Lázaro-Nogal et al (2015) showed in a common garden study with several
81 origins of *Senna candolleana* that populations originating from climatically more variable
82 heterogeneous environments had a higher plasticity in growth traits. A similar
83 observation was made by Gianoli and Gonzáles-Teuber (2005) who showed that
84 plasticity in leaf area, leaf shape, leaf area ratio (LAR), and foliar trichome density in
85 *Convolvulus chilensis* was highest for plants from the population with the highest
86 interannual variation in precipitation. Thus, increased climatic variability appears to be
87 associated with systematic, and presumably adaptive, changes in phenotypic plasticity.
88 The fate of plant populations will also depend on their ability to adapt to altered climatic
89 variability.

90 A powerful method to test for recent evolution - whether in trait means or in their
91 plasticity - is to compare ancestors with their descendants by using stored propagules
92 such as seeds (Orsini et al 2013; Merilä and Hendry 2014; Franks et al 2018). If
93 ancestors can be revived, the resulting plants can be compared to individuals raised
94 from propagules sampled from the same population today. Growing ancestors and
95 descendants together under common conditions then allows to directly test for heritable

96 trait differentiation among temporally separated populations (Franks et al 2007, 2008).
97 Understanding how populations responded evolutionarily in the past is extremely
98 valuable for making predictions for future population responses to environmental
99 change (Orsini et al 2013; Franks et al 2018).

100 An increasing number of studies have used this so-called “resurrection approach”
101 to compare plants grown from seeds collected before and after drought events. They
102 have shown that plants adapted their phenology through an early start of flowering in
103 order to avoid temporary droughts (Franks et al 2007; Nevo et al 2012; Vigouroux et al
104 2011; Thomann et al 2015). For growth traits, results appear to be more species-
105 specific. For example, in an experiment with *Mimulus laciniatus* by Dickmann (2016)
106 drought-adapted plants grew bigger, whereas Vigouroux and colleagues (2011) showed
107 opposite results in a study with *Pennisetum glaucum*.

108 Here, we investigated whether single populations of multiple plant species from
109 Mediterranean and temperate regions of Europe have already evolved their drought
110 tolerance over the last decades as the expected response to more frequent and longer
111 drought events (Met office 2011; DWD 2018; IRM 2020). To investigate this, we
112 conducted two common garden experiments in which we applied drought treatments to
113 plants raised from seeds stored for at least 20 years in three different seed banks
114 (ancestors) and from seeds that we collected from the same populations in 2018
115 (descendants). In the first experiment (“seedling survival experiment”), we used four
116 Mediterranean species to test whether seedlings of the descendants survived longer
117 without watering than the seedlings of their ancestors. Seedling establishment is a
118 crucial process for population survival (Grubb_1977), and seedlings are especially

119 susceptible to drought (Moles and Westoby_2004). Therefore seedling drought
120 resistance should be under high selection pressure in increasingly dry and more
121 variable environments (Schupp 1995; Fenner and Kitajima 1999). In the second
122 experiment (“watering response experiment”) we worked with adult plants from nine
123 temperate European species. We subjected ancestors and descendants to well-watered
124 vs. dry conditions and compared their growth responses to test the hypothesis that
125 populations evolved phenotypic traits, and/or their plasticities, to cope with increased
126 droughts.

127

128 **Material and Methods**

129 **Seed collection**

130 For the seedling survival experiment we obtained seeds of Mediterranean species from
131 the seed bank at the Conservatoire Botanique National Méditerranéen de Porquerolles
132 (CBNMed, Hyères, France). The seed material for the watering response experiment
133 was provided by the Meise Botanic Garden (Belgium) and the botanical garden of
134 Osnabrück (Germany). For both experiments, we only used seeds with precise
135 sampling dates and location records, and which had been stored for at least 20 years.
136 We only selected species with a short life cycle as they are expected to respond more
137 quickly to selection and are therefore more likely to show rapid evolutionary changes
138 between ancestors and descendants. Finally, we specifically chose species with
139 relatively isolated (but large) populations to reduce outcrossing with other populations.
140 From the CBNMed (seedling survival experiment) we used seeds of four species that
141 met these requirements (Table 1): *Anthemis maritima*, *Matthiola tricuspidata*, *Medicago*

142 *marina* and *Plantago subulata*. The seeds of these four species had been collected
143 between 1992 and 1997 in the region of Hyères, Southern France, where average
144 temperatures have increased by 1.5 °C and precipitation decreased during the last 30
145 years (Met office 2011). After ancestor seed collection, the seeds of *A. maritima*, *M.*
146 *tricuspidata* and *M. marina* were cleaned, dried and stored at 5°C, whereas seeds of
147 *Plantago subulata* were ultra-dessicated and stored at 17°C at the CBNMed until
148 November 2018. For the watering response experiment we used seeds of a total of nine
149 species that met our requirements (Table 1): *Centaureum erythraea*, *Clinopodium*
150 *vulgare*, *Dianthus carthusianorum*, *Digitalis lutea*, *Leontodon hispidus*, *Melica ciliata*,
151 *Pimpinella saxifraga*, *Sedum album*, *Teucrium chamaedrys*. The seeds of these nine
152 species had been collected between 1992 and 1995 in Belgium and close to Osnabrück,
153 Lower-Saxony (Germany). In Belgium, summer temperatures have increased by 0.37°C
154 per decade since 1981 (IRM 2020) and in northern Germany mean annual temperature
155 increased by 0.7°C when comparing 1961-1990 to 1981-2010 (DWD 2018). After
156 ancestor seed collection, the seeds had been cleaned, dried at 15% relative humidity
157 and then stored at -20°C at Meise Botanic Garden and the botanical garden of
158 Osnabrück until November 2018. To obtain the descendants, seeds of all species were
159 collected from the same populations in spring (Mediterranean species) and summer
160 2018 (temperate species).

161

162 **Seedling survival experiment**

163 In order to break physical seed dormancy, we scarified seeds of *M. marina* by softly
164 scrubbing them with sandpaper. To reduce the growth of bacteria or fungi during

165 germination, we surface-sterilized all seeds with 3% Sodium Hypochlorite (NaOCl) and
166 two drops of Tween20 per 200ml solution, and washed them three times with sterilized
167 water. We germinated all seeds on 1% water agar in 90 mm Petri dishes. After one
168 week of cold-dark stratification at 5°C we transferred the Petri dishes to a walk-in growth
169 chamber (light intensity = $230 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, 50% relative humidity) with a light/dark cycle
170 of 8/16 hours and temperatures of 23/18°C.

171 For each species we filled one seedling tray (96-cell QuickPot®, 3.8 x 3.8 cm cells) with
172 a standard potting soil (Einheitserde®, BioLine, Topfsubstrat Öko torffrei) and planted
173 seedlings into every other cell so that seedlings did not grow directly next to each other.
174 We planted the ancestors and descendants in an even pattern, but to identify the
175 seedlings, we did not use any labels but noted their position separately from the tray in
176 order to reduce observer bias. The trays were placed in a walk-in growth chamber with
177 a light/dark cycle of 12/12 hours and 23/18°C (light intensity = $230 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, 50%
178 relative humidity). To reduce insect damage, sticky traps were distributed throughout
179 the room. The seedlings were watered regularly for 2-3 weeks (depending on the
180 species) to allow their establishment, and after that, we stopped watering to simulate a
181 temporary drought. We recorded mortality due to desiccation at least every other day. A
182 seedling was scored as dead when it was completely dry and all leaves had lost their
183 green colour. We cut each dead seedling 1 mm above ground, dried it at 60°C for 48 h,
184 and weighed it.

185 We used linear models to analyse how seedling origin (ancestor vs. descendant)
186 affected mean time to desiccation and aboveground biomass. To account for species
187 differences, we included species identity in the model as a fixed factor. For the analyses

188 of time to desiccation we included days between transplanting and start of the
189 experiment and for the analyses of aboveground biomass the total lifespan of the
190 seedling as covariables. We additionally analysed the data separately for each species,
191 using the same model but excluding species identity. Finally, we used linear models to
192 test whether the mean time to desiccation of a plant individual is explained by its
193 aboveground biomass while correcting for the total lifespan of the seedling. For all
194 models we checked the residuals visually for normality and heteroscedasticity.

195

196 **Watering response experiment**

197 We germinated seeds in trays filled with a standard potting soil (Einheitserde®, BioLine,
198 Pikiersubstrat), with each origin sown into a separate tray. In order to break seed
199 dormancy we kept the soil moist and cold-dark stratified the seeds at 5°C for two
200 months. In March 2019, we transferred the trays to the greenhouse and allowed the
201 seeds to germinate at 20°C and a natural spring daylight regime. We kept the seedlings
202 in the trays for three months before the start of the experiment

203 We filled 9 x 9 cm pots with a 3:1 mixture of potting soil (Einheitserde®, BioLine,
204 Topfsubstrat Öko torffrei) and sand (0-2 mm play sand, WECO GmbH). We
205 transplanted the seedlings into the pots within one week in June 2019, and we always
206 planted pairs of ancestor and descendant seedlings that were approximately of equal
207 size. Right after transplantation we measured shoot length or rosette diameter
208 (henceforth referred to as plant size) and the number of leaves or shoots, depending on
209 the species (Table 1). After two weeks we separated all plants into a well-watered
210 control group and a drought-suffering group, each with at least seven replicates per

211 origin (ancestor vs. descendant) and species (Table 1). When five of the pots of a
212 species had a dry soil surface, all plants of that species were watered, with control
213 plants receiving 60 mL and drought plants receiving 30 mL water at each watering event.
214 We re-randomized all pots in the greenhouse weekly. After eight weeks we repeated the
215 growth trait measurements and subsequently harvested the plants and determined their
216 aboveground biomass after oven-drying at 60°C for three days.

217 In order to be able to compare various measured traits across nine different species, we
218 transformed all data by dividing the measured value for a trait by the mean value of this
219 trait for a species. We then analysed the variation in plant size, number of leaves or
220 shoots, and aboveground biomass with linear models that included origin (ancestor vs.
221 descendant), treatment (drought vs. control) and species, and all possible interactions,
222 as explanatory variables. A two-way interaction between plant origin and treatment
223 would indicate that plants have evolved a different response to drought, and a three-
224 way interaction between plant origin, treatment and species would show that species
225 vary in their evolutionary responses to drought. We subsequently used linear models
226 including only origin, treatment and their interaction to analyse the data for each species
227 separately. As the sizes of transplanted seedling differed, we corrected for this by
228 including the initial size measurement as a covariate in all our models. For all models
229 we visually checked the residuals for normality and heteroscedasticity.

230 All analyses were done in R using the packages *plyr* (Wickham 2011) and *ggplot2*
231 (Wickham 2016). All data is accessible as supplements in Tables S1 and S2.

232

233 **Results**

234 **Seedling survival experiment**

235 In the seedling survival experiment, seedlings from descendants survived on average
236 almost two days longer than seedlings from ancestors (Fig.1a, $F_{1,208} = 12.99$, $p < 0.001$).
237 We also found species differences in mean survival ability (Fig. 1a, $F_{3,208} = 255.21$, $p <$
238 0.001) and an interaction between species and origin (ancestors vs. descendants; Fig.
239 1a, $F_{3,208} = 2.74$, $p = 0.04$). The overall effect of temporal origin is strongly influenced by
240 a single species, *A. maritima*, since only descendants from this species showed a
241 significantly longer survival than their ancestors in individual species analyses (Fig. 1a,
242 $F_{1,60} = 6.01$, $p = 0.017$). On average, seedlings from descendants had a significantly
243 lower biomass than those from ancestors (Fig. 1b, $F_{1,204} = 19.92$, $p < 0.001$). Again,
244 there was an interaction between species and origin (Fig. 1b, $F_{3,204} = 3.57$, $p = 0.015$),
245 with the overall effect largely driven by *A. maritima* as only this species showed a
246 significant biomass difference between origins in individual species analyses (Fig. 1b,
247 $F_{1,59} = 6.08$, $p = 0.016$). Across species, plants with a lower biomass generally survived
248 longer ($F_{1,200} = 12.43$, $p = <0.001$, $r^2=0.46$). However, at the species level we observed
249 such a significant negative correlation between biomass and survival only for *A.*
250 *maritima* ($F_{4,58} = 4.03$, $p = 0.006$, $r^2=0.16$), whereas for *M. tricuspidata* ($F_{3,59} = 4.01$, $p =$
251 0.012 , $r^2=0.13$) and *M. marina* ($F_{4,46} = 7.71$, $p <0.001$, $r^2=0.35$) there were positive
252 correlations, i.e. larger plants survived longer.

253

254 **Watering response experiment**

255 In the watering response experiment, the drought treatment had a significant effect on
256 all three measured traits. Across all nine species, plants grown under drought conditions

257 grew smaller, produced less branches or leaves and had a lower aboveground biomass
258 (Fig. 2a-c). These observations were also consistent at the species level: wherever a
259 significant effect occurred, the drought treatment lowered plant performance (Table 2).
260 There were no significant interactions between species and treatment. The temporal
261 origin did not affect plant size (shoot length and rosette diameter) in any of the studied
262 species, but we found a significant difference in aboveground biomass between the
263 ancestors and descendants of three species: In *C. erythraea* and *M. ciliata* ancestors
264 produced significantly less biomass, whereas in *D. carthusianorum* the opposite was
265 true (Table 2). Across species, there was a significant drought by origin interaction for
266 plant size (Fig. 2a): While ancestral plants showed only a slight decrease of plant size in
267 response to drought, the descendants strongly decreased plant size under drought. This
268 observation is quite consistent across species, as there was no significant three-way
269 interaction among treatment, origin and species in our model ($F_{8,329} = 0.45$, $p = 0.890$).

270

271 **Discussion**

272 **Seedling survival experiment**

273 In our seedling survival experiment with four Mediterranean plant species, we showed
274 that descendant seedlings produced less aboveground biomass but survived longer
275 under drought than their ancestors. The four studied species varied in their survival
276 ability and growth, and only for *A. maritima* smaller size was associated with longer
277 survival. Similar results using a multi-species approach with ten grassland herbs were
278 found by Harrison and LaForgia (2019). In their experiment, species with small
279 seedlings and low specific leaf area had a higher survival ability under drought. A

280 possible explanation for this is reduced evapotranspiration through a decreased leaf
281 number, leaf size and branching, and lower plant biomass (Aroca 2012). These
282 observations also correspond with the notion that plants in dry conditions often
283 decrease aboveground biomass production and allocate more biomass to roots leading
284 to a higher root-shoot ratio (Martin and Stephens 2006; Villagra and Cavagnaro 2006;
285 Erice et al 2007). Of course, other traits can also be responsible for an increased
286 seedling drought tolerance as root structures such as hypocotyl hairs (Aronne and De
287 Micco 2004) or seed size (Fenner and Kitajima 1999). Furthermore, according to the
288 predictions of climate change for the Mediterranean region, with decreased rainfall and
289 more frequent droughts, high plasticity in seedling traits could also be advantageous
290 (Gimeno et al 2009). For example, Padilla and Pugnaire (2007) showed that a rooting
291 depth responding plastically to water availability could be more important than a
292 constitutively higher root:shoot ratio. In this experiment it seems likely that the reduction
293 in aboveground biomass for *A. maritima* is an adaptation (Erice et al 2010) to increased
294 drought frequency in the Mediterranean region over the last decades (Met office 2011).
295 For the three other studied species we could not observe the above mentioned pattern.
296 Possible reasons for that could be that these species have not really evolved yet, the
297 differences cannot be seen at seedling stage or these species established one of the
298 other mentioned or even alternative strategies to cope with drought during seedling
299 stage.

300 Altogether, we show that seedling survival under drought has evolved in the last 21-26
301 years in four Mediterranean plant species. As those changes have evolved in a very
302 short time (supposedly less than 10 generations assuming cycles more than two years

303 per generation), our study suggests high selection pressures under the increasingly
304 heat-wave prone Mediterranean climate.

305

306 **Watering response experiment**

307 In our watering response experiment with nine species from temperate Europe we
308 subjected adult plants to drought which generally led to decreased plant sizes and
309 aboveground biomasses. We found no general cross-species differences in mean traits
310 between descendants and ancestors, but there was a general difference between
311 ancestors and descendants in the plasticity of plant size in response to drought, with a
312 much stronger decrease of size in the descendant plants. Since plant biomass and
313 number of leaves or shoots were not affected, this stronger shift in plant size under dry
314 conditions could be accompanied by changes in leaf morphology such as leaf thickness
315 or shape which are known to be highly plastic traits (Gianoli and Gonzáles-Teuber 2005;
316 Lázaro-Nogal et al 2015). A reduction of leaf area accompanied by increasing leaf
317 thickness and/or more pubescent leaves may reduce evapotranspiration (Gianoli and
318 Gonzáles-Teuber 2005) and can therefore be a successful strategy under drought (La
319 Riva et al 2016).

320 In general plant responses to drought are very complex as drought affects plants at
321 various development stages and different tissues (Yordanov et al 2000). Our
322 experiment does not allow us to identify the processes underlying the observed patterns
323 which could include increased resource allocation to roots (Martin and Stephens 2006;
324 Villagra and Cavagnaro 2006; Erice et al 2007), reduced evapotranspiration (Aroca

325 2012), reduced photosynthesis rate and increased oxidative stress (Zlatev and Lidon
326 2012) or a combination of these and other factors.

327 We also found larger plant size in the control treatment for the descendants compared
328 to ancestors, which may be an adaptation of the species' life history. As flowering onset
329 is often related to plant size (Vile et al 2006; Sun and Frelich 2011), we argue that
330 plants grow and develop fast when water supply is satisfactory to avoid potential
331 drought stress later on in their life cycle (Greene et al 2011).

332 In general higher environmental heterogeneity should favour greater phenotypic
333 plasticity (Alpert and Simms 2002; Bradshaw and Holzapfel 2006; Matesanz et al 2010).

334 In the regions of origin of the study species, drought frequency has increased over the
335 last 20 years (Spinoni et al 2018), and environmental conditions thus became more
336 unpredictable (Altvater et al 2011) and may thus have favoured evolution of plasticity. It
337 is common that, as in our experiment, differentiation in phenotypic plasticity only occurs
338 in some traits (Heschel et al 2002), and of course the mere observation of higher
339 plasticity in a phenotypic trait does not necessarily mean that this is an adaptive change.

340 Nevertheless, it is known that plasticity, like mean trait values, may evolve by natural
341 selection (Ackerly et al 2000; Richards et al 2006). To test whether the observed higher
342 plasticity in plant size of the descendant plants is an adaptive change to increased
343 drought frequencies requires further experiments that include longer-term
344 measurements of plant fitness (Richards et al 2006). Ideally these experiments would
345 take place at the species' sites of origin and incorporate a large number of populations
346 with different magnitudes of climate change during the past decades.

347 We acknowledge that in both experiments our results could have been influenced by
348 unintentional selection during sampling (Hay and Smith 2003; Hoban and Schlarbaum
349 2014) or by potential selection during storage (Crawford et al 2007; Godefroid et al
350 2010). Furthermore, we could not rule out maternal effects, which could have been
351 passed on over multiple generations (Gimeno et al 2009). However, we expect that
352 random processes or unintentional selection are unlikely to have a stronger effect than
353 those exerted by the drought treatments, as seedling recruitment and responses to
354 drought are under high selection pressure (Schupp 1995; Fenner and Kitajima 1999).
355 Moreover, the observed higher plasticity in descendants was consistent for seven out of
356 the nine studied species (Fig. S1), which is highly unlikely if part or all of this effect was
357 due to random processes or unintentional selection on each species separately.

358

359 **Conclusions**

360

361 Ongoing climate change is expected to influence the evolution of plant populations, but
362 so far experimental tests of this are rare. Our multi-species experiments using species
363 from two different biogeographic regions in Europe indicate that plants have indeed
364 evolved within the last decades, possibly in response to increased drought frequencies.
365 The observed evolutionary changes involve both changes in trait means and in plasticity
366 in response to drought. Given the increased occurrence of drought events, our results
367 suggest that climate change may have already influenced the evolutionary trajectory of
368 plant species in different regions of Europe. Our study also demonstrates that the
369 resurrection approach is a powerful tool for studying rapid evolutionary changes in
370 plants. Future studies should corroborate our results across other species, ideally with
371 multiple populations for each, using seeds from plants that underwent one generation in

372 a common garden to exclude possible maternal effects. To disentangle adaptive from
373 non-adaptive responses to recent climate change, these experiments should
374 incorporate fitness measures and comparative transplantations of descendants and
375 ancestors into their original habitat.

376

377 **Acknowledgments**

378 We thank Nicolai Friesen and Jaroslav Kloster (University of Osnabrück) for their help
379 with the seed collections in Osnabrück, all the authorities in France, Belgium and
380 Germany for issuing the permits for seed sampling and Madalin Parepa (University of
381 Tübingen) for discussion on data analyses.

382 **Funding**

383 This research was financially supported by a PhD scholarship of the Deutsche
384 Bundesstiftung Umwelt (DBU) to R.R. and a RISE scholarship of the German Academic
385 Exchange Service (DAAD) to C.L..

386 **References**

387 Ackerly DD, Dudley SA, Sultan SE, Schmitt J, Coleman JS, Linder CR et al (2000) The
388 evolution of plant ecophysiological traits: recent advances and future directions. *Int J*
389 *Plant Sci* 50(11):979-995

390 Alpert P, Simms EL (2002) The relative advantages of plasticity and fixity in different
391 environments: when is it good for a plant to adjust? *Evol Ecol* 16(3):285–297

392 Altvater S, van de Sandt K, Marinova N, de Block D, Klostermann J, Swart R, Bouwma I,
393 McCallum S, Dworak T, Osberghaus D (2011) Assessment of the most significant
394 threats to the EU posed by the changing climate in the short, medium and long term
395 Task 1 report. Ecologic, Berlin

396 Anderson JT, Panetta AM, Mitchell-Olds T (2012) Evolutionary and ecological
397 responses to anthropogenic climate change: update on anthropogenic climate change.
398 *Plant Physiol* 160(4):1728–1740

399 Aroca R (2012) Plant responses to drought stress: from morphological to molecular
400 features. Springer, Berlin Heidelberg

401 Aronne G, de Micco V (2004) Hypocotyl features of *Myrtus communis* (Myrtaceae) A
402 many-sided strategy for possible enhancement of seedling establishment in the
403 Mediterranean environment. *Bot J Linn Soc* 145(2):195–202

404 Bertrand R, Lenoir J, Piedallu C, Riofrío-Dillon G, de Ruffray P, Vidal C, Pierrat JC,
405 Gégout JC (2011) Changes in plant community composition lag behind climate warming
406 in lowland forests. *Nature* 479:517-520

407 Bradshaw WE, Holzapfel CM (2006) Climate change evolutionary response to rapid
408 climate change. *Science* 312(5779):1477–1478

409 Crawford AD, Steadman KJ, Plummer JA, Cochrane A, Probert RJ (2007) Analysis of
410 seed-bank data confirms suitability of international seed-storage standards for the
411 Australian flora. *Aust J Bot* 55(1):18-29

412 DWD (2018) Klimareport Niedersachsen. Deutscher Wetterdienst, Offenbach am Main
413 (Deutschland), 2018

414 Dickman EE (2016) Phenotypic responses of a Sierra Nevada monkeyflower to climate
415 variation and severe drought. Master Thesis, University of California (Merced)

416 Erice G, Irigoyen JJ, Sánchez-Díaz M, Avice JC, Ourry A (2007) Effect of drought,
417 elevated CO₂ and temperature on accumulation of N and vegetative storage proteins
418 (VSP) in taproot of nodulated alfalfa before and after cutting. *Plant Sci* 172(5):903–912

419 Erice G, Louahlia S, Irigoyen JJ, Sanchez-Diaz M, Avice JC (2010) Biomass partitioning,
420 morphology and water status of four alfalfa genotypes submitted to progressive drought
421 and subsequent recovery. *J Plant Physiol* 167(2):114–120

422 Farooq M, Wahid A, Kobayashi N, Fujita D, Basra SMA (2009) Plant drought stress:
423 effects, mechanisms and management. *Agron Sustain Dev* 29(1):185–212

424 Feng S, Fu Q (2013) Expansion of global drylands under a warming climate. *Atmos*
425 *Chem Phys Discuss* 13(6):14637–14665

426 Fenner M, Kitajima K. (1999) Seed and seedling ecology. In: Pugnaire FI, Valladares F
427 (ed) *Handbook of functional plant ecology*, Marcel Dekker, New York Basel pp589–621

428 Fleta-Soriano E, Munné-Bosch S (2016) Stress memory and the inevitable effects of
429 drought: a physiological perspective. *Front Plant Sci* 7:143

430 Franks SJ, Sim S, Weis AE (2007) Rapid evolution of flowering time by an annual plant
431 in response to a climate fluctuation. *P Natl Acad Sci USA* 104(4):1278–1282

432 Franks SJ, Weis AE (2008) A change in climate causes rapid evolution of multiple life-
433 history traits and their interactions in an annual plant. *J Evolution Biol* 21(5):1321–1334

434 Franks SJ, Hamann E, Weis AE (2018) Using the resurrection approach to understand
435 contemporary evolution in changing environments. *Evol Appl* 11(1):17–28

436 Gianoli E, González-Teuber M (2005) Environmental heterogeneity and population
437 differentiation in plasticity to drought in *Convolvulus chilensis* (Convolvulaceae). *Evol*
438 *Ecol* 19(6):603–613

439 Gianoli E, Valladares F (2012) Studying phenotypic plasticity: the advantages of a broad
440 approach. *Biol J Linn Soc* 105(1):1–7

441 Gimeno TE, Pías B, Lemos-Filho JP, Valladares F (2009) Plasticity and stress tolerance
442 override local adaptation in the responses of Mediterranean holm oak seedlings to
443 drought and cold. *Tree Physiol* 29(1):87–98

444 Godefroid S, van de Vyver A, Vanderborght T (2010) Germination capacity and viability
445 of threatened species collections in seed banks. *Biodivers Conserv* 19(5):1365–1383

446 Grene R, Vasquez-Robinet C, Bohnert HJ (2011) Molecular biology and physiological
447 genomics of dehydration stress. In: Lüttge U, Beck E, Bartels D (ed) *Plant Desiccation*
448 *Tolerance*, vol 215. Springer, Berlin Heidelberg pp255–287

449 Grubb PJ (1977) The maintenance of species-richness in plant communities: the
450 importance of the regeneration niche. *Biol Rev* 52(1):107–145

451 Harrison S, LaForgia M (2019) Seedling traits predict drought-induced mortality linked to
452 diversity loss. *P Natl Acad Sci USA* 116(12):5576–5581

453 Hay FR, Smith RD (2003) Seed maturity: when to collect seeds from wild plants. In:
454 Smith RD, Dickie JB, Linington S, Pritchard HW, Probert RJ (ed) *Seed Conservation –*
455 *Turning Science into Practice*, Royal Botanic Gardens Kew, Kew, pp97–133

456 Heschel MS, Donohue K, Hausmann N, Schmitt J (2002) Population differentiation and
457 natural selection for water-use efficiency in *Impatiens capensis* (Balsaminaceae). *Int J*
458 *Plant Sci* 163(6):907–912

459 Hoban S, Schlarbaum S (2014) Optimal sampling of seeds from plant populations for
460 ex-situ conservation of genetic biodiversity, considering realistic population structure.
461 Biol Conserv 177:90–99

462 Hoffmann AA, Sgrò CM (2011) Climate change and evolutionary adaptation. Nature
463 470(7335): 479–485

464 Holt RD (1990) The microevolutionary consequences of climate change. Trends Ecol
465 Evol 5(9):311–315

466 IPCC (2013) Climate change 2013: the physical science basis. Contribution of Working
467 Group I to the 5th assessment report of the intergovernmental panel on climate change.
468 Stocker TF, Qin D, Plattner GK, Tignor M, Allen SK, Boschung J, Nauels A, Xia Y, Bex
469 V, Midgley PM (ed), University Press Cambridge, United Kingdom and New York

470 IPCC (2018) Global warming of 1.5°C. An IPCC special report on the impacts of global
471 warming of 1.5°C above pre-industrial levels and related global greenhouse gas
472 emission pathways, in the context of strengthening the global response to the threat of
473 climate change, sustainable development, and efforts to eradicate poverty. Masson-
474 Delmotte V, Zhai P, Pörtner HO, Roberts D, Skea J, Shukla PR, Pirani A, Moufouma-
475 Okia W, Péan C, Pidcock R, Connors S, Matthews JBR, Chen Y, Zhou X, Gomis MI,
476 Lonnoy E, Maycock T, Tignor M, Waterfield T (ed), In Press

477 IRM (2020) Tendances observées en Belgique. Institut Royal Météorologique, Bruxelles,
478 Belgium. Available from [https://www.meteo.be/fr/climat/tendances-climatiques-
479 observees/en-belgique](https://www.meteo.be/fr/climat/tendances-climatiques-observees/en-belgique) (accessed January 2020)

480 Jaleel CA, Manivannan P, Wahid A, Farooq M, Al-Juburi HJ, Somasundaram R,
481 Panneerselvam, R (2009) Drought stress in plants: a review on morphological
482 characteristics and pigments composition. Int J Agric Biol 11(1)100–105

483 Kharin VV, Zwiers FW, Zhang Xm, Hegerl GC (2007) Changes in temperature and
484 precipitation extremes in the IPCC ensemble of global coupled model simulations. J
485 Climate 20(8):1419–1444

486 Kusaka M, Ohta M, Fujimura T (2005) Contribution of inorganic components to osmotic
487 adjustment and leaf folding for drought tolerance in pearl millet. *Physiol Plantarum*
488 125(4):474–489

489 La Riva EG de, Olmo M, Poorter H, Ubera JL, Villar R (2016) Leaf mass per area (LMA)
490 and its relationship with leaf structure and anatomy in 34 Mediterranean woody species
491 along a water availability gradient. *PLOS ONE* 11(2), e0148788

492 Lázaro-Nogal A, Matesanz S, Godoy A, Pérez-Trautman F, Gianoli E, Valladares F
493 (2015) Environmental heterogeneity leads to higher plasticity in dry-edge populations of
494 a semi-arid Chilean shrub: insights into climate change responses. *J Ecol* 103(2):338–
495 350

496 Martin PJ, Stephens W (2006) Willow growth in response to nutrients and moisture on a
497 clay landfill cap soil. I. Growth and biomass production. *Bioresource Technol* 97(3):437–
498 448

499 Matesanz S, Gianoli E, Valladares F (2010) Global change and the evolution of
500 phenotypic plasticity in plants. *Ann NY Acad Sci* 1206:35–55

501 Merilä J, Hendry AP (2014) Climate change, adaptation, and phenotypic plasticity: the
502 problem and the evidence. *Evol Appl* 7:1–14

503 Met Office (2011) *Climate: Observations, projections and impacts (France)*. Met Office,
504 Exeter, United Kingdom

505 Moles AT, Westoby M (2004) Seedling survival and seed size: a synthesis of the
506 literature. *J Ecol* 92(3):372–383

507 Nevo E, Fu YB, Pavlicek T, Khalifa S, Tavasi M, Beiles A (2012) Evolution of wild
508 cereals during 28 years of global warming in Israel. *P Natl Acad Sci USA* 109(9):3412–
509 3415

510 Orsini L, Schwenk K, Meester L de, Colbourne JK, Pfrender ME, Weider LJ (2013) The
511 evolutionary time machine: using dormant propagules to forecast how populations can
512 adapt to changing environments. *Trends Ecol Evol* 28(5):274–282

513 Padilla FM, Pugnaire FI (2007) Rooting depth and soil moisture control Mediterranean
514 woody seedling survival during drought. *Funct Ecol* 21(3):489–495

515 Parmesan C, Yohe G (2003) A globally coherent fingerprint of climate change impacts
516 across natural systems. *Nature* 421(6918):37–42

517 Peñuelas J, Filella I (2001) Responses to a warming world. *Science* 294(5543):793–795

518 Richards CL, Bosdorf O, Muth NZ, Gurevitch J, Pigliucci M (2006) Jack of all trades,
519 master of some? On the role of phenotypic plasticity in plant invasions. *Ecol Lett*
520 9(8):981–993

521 Ruosteenoja K, Markkanen T, Venäläinen A, Räisänen P, Peltola H (2018) Seasonal
522 soil moisture and drought occurrence in Europe in CMIP5 projections for the 21st
523 century. *Clim Dyn* 50(3-4):1177–1192

524 Samaniego L, Thober S, Kumar R, Wanders N, Rakovec O, Pan M et al (2018)
525 Anthropogenic warming exacerbates European soil moisture droughts. *Nature Clim*
526 *Change* 8(5):421–426

527 Schupp EW (1995): Seed-seedling conflicts, habitat choice, and patterns of plant
528 recruitment. *Am J Bot* 82(3) 399-409

529 Shao H, Chu L, Shao M, Jaleel CA, Mi H (2008) Higher plant antioxidants and redox
530 signaling under environmental stresses. *C R Biol* 331(6):433–441

531 Sharp RE, LeNoble ME (2002) ABA, ethylene and the control of shoot and root growth
532 under water stress. *J Exp Bot* 53(366):33–37

533 Shaw RG, Etterson JR (2012) Rapid climate change and the rate of adaptation: insight
534 from experimental quantitative genetics. *New Phytol* 195(4):752–765

535 Spinoni J, Vogt JV, Naumann G, Barbosa P, Dosio A (2018) Will drought events
536 become more frequent and severe in Europe? *Int J Climatol* 38(4):1718-1736

537 Sultan SE, Spencer HG (2002) Metapopulation structure favors plasticity over local
538 adaptation. *Am Nat* 160(2):271–283

539 Sun S, Frelich LE (2011) Flowering phenology and height growth pattern are associated
540 with maximum plant height, relative growth rate and stem tissue mass density in
541 herbaceous grassland species. *J Ecol* 99(4):991–1000

542 Thomann M, Imbert E, Engstrand RC, Cheptou PO (2015) Contemporary evolution of
543 plant reproductive strategies under global change is revealed by stored seeds. *J*
544 *Evolution Biol* 28(4):766-78

545 Vigouroux Y, Mariac C, Mita S de, Pham JL, Gérard B, Kapran I et al (2011) Selection
546 for earlier flowering crop associated with climatic variations in the Sahel. *PLOS ONE*
547 6(5): e19563

548 Vile D, Shipley B, Garnier E (2006) A structural equation model to integrate changes in
549 functional strategies during old-field succession. *Ecology* 87(2):504–517

550 Villagra PE, Cavagnaro JB (2006) Water stress effects on the seedling growth of
551 *Prosopis argentina* and *Prosopis alpataco*. *J Arid Environ* 64(3):390–400

552 Wickham H (2011) Tools for Splitting, Applying and Combining Data. R package version
553 1.8.6. <https://cran.r-project.org/web/packages/plyr/index.html>

554 Wickham H (2016) Create Elegant Data Visualisations Using the Grammar of Graphics.
555 R package version 3.3.1. <https://cran.r-project.org/web/packages/ggplot2/index.html>

556 Yordanov I, Velikova V, Tsonev T (2000) Plant Responses to drought, acclimation, and
557 stress tolerance. *Photosynthetica* 38(2):171–186

558 Zlatev Z, Lidon F (2012) An overview on drought induced changes in plant growth,
559 water relation and photosynthesis. *Emir J Food Agric* 24(1):57-72

560 **Supplementary material**

561 **Fig. S1** Individual reaction norm plots of plant size for all tested species

562 **Table S1** Raw data of the seedling survival experiment

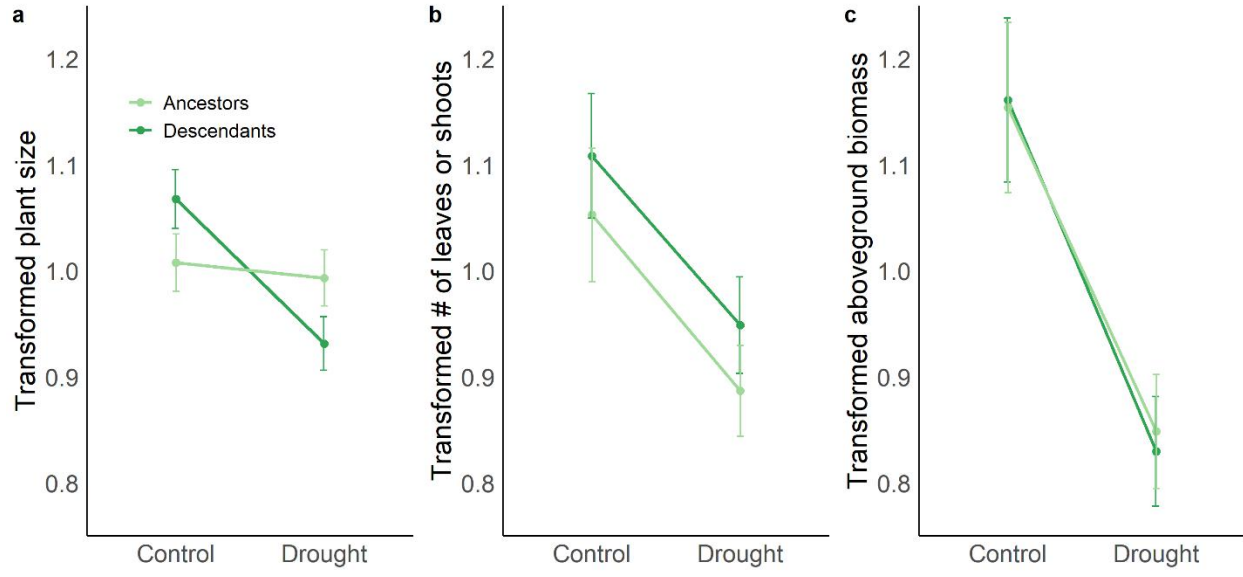
563 **Table S2** Raw data of the watering response experiment

564 **Figures and tables**

565 **Figure captions**

566 **Fig. 1** Mean number of days until mortality after watering ceased (a) and aboveground
567 biomass at harvest (b) of seedlings of four Mediterranean species (*Anthemis maritima*,
568 *Matthiola tricuspidata*, *Medicago marina*, *Plantago subulata*) from two different temporal
569 origins (ancestors vs. descendants). The bars show means and standard errors. * =
570 $p < 0.05$, *** = $p < 0.001$

571 **Fig. 2** Reaction norm plots of plant size (a), number of leaves or shoots (b) and
572 aboveground biomass (c) in the watering response experiment. The data are always
573 transformed and across all nine species from two different origins (ancestors vs.
574 descendants). Error bars show standard errors.



577
578

Fig 2

579 **Table 1** Study species used in the two experiments with details on plant family,
 580 seedbank, country, year of collection, life form, measured traits in the watering response
 581 experiment and the number of replicates used.

Species	Family	Seedbank	Country	Collection year	Measure of		Replicates
					plant size	Number of leaves of shoots	
Seedling survival experiment: Mediterranean species							
<i>Anthemis maritima</i>	Asteraceae	CBNMed	France	1992	-	-	30
<i>Matthiola tricuspidata</i>	Brassicaceae	CBNMed	France	1994	-	-	30
<i>Medicago marina</i>	Fabaceae	CBNMed	France	1992	-	-	20
<i>Plantago subulata</i>	Plantaginaceae	CBNMed	France	1997	-	-	9
Watering response experiment: Temperate species							
<i>Centaureum erythraea</i>	Gentianaceae	Meise	Belgium	1992	diameter	leaves	12
<i>Clinopodium vulgare</i>	Lamiaceae	Meise	Belgium	1992	height	shoots	12
<i>Dianthus carthusianorum</i>	Caryophyllaceae	Osnabrück	Germany	1993	diameter	leaves	7
<i>Digitalis lutea</i>	Plantaginaceae	Meise	Belgium	1992	diameter	leaves	12
<i>Leontodon hispidus</i>	Asteraceae	Meise	Belgium	1995	diameter	leaves	12
<i>Melica ciliata</i>	Poaceae	Meise	Belgium	1992	height	shoots	7
<i>Pimpinella saxifraga</i>	Apiaceae	Meise	Belgium	1992	diameter	shoots	8
<i>Sedum album</i>	Crassulaceae	Meise	Belgium	1992	diameter	shoots	12
<i>Teucrium chamaedrys</i>	Lamiaceae	Meise	Belgium	1992	height	shoots	12

583 **Table 2** F-values of linear model analyses of the watering response experiment, each
 584 testing for effects of treatment (T; drought vs. control), origin (O; ancestors vs.
 585 descendants), and their interaction (T x O). , and, for the all-species model, species as
 586 explanatory factors. The arrows indicate the direction of a significant effect (↓ =
 587 transformed values of the descendants or drought, respectively, are smaller). Shading
 588 indicates the range of the p-value (light gray: p<0.1, medium gray: p<0.05, dark gray:
 589 p<0.001). Degrees of freedom for all tested effects was 1 and varies for the residuals.

	Plant size			Number of leaves or shoots			Aboveground biomass		
	T	O	T x O	T	O	T x O	T	O	T x O
All Species	12.02↓	0.02	5.95	9.84↓	1.44	0.03	27.86↓	0.01	0.01
<i>Centaureum erythraea</i>	7.91↓	3.69	4.04	0.03	3.94	0.17	4.95↓	14.08↑	2.87
<i>Clinopodium vulgare</i>	0.28	0.05	0.68	0.25	0.73	0.52	2.03	0.07	0.38
<i>Dianthus carthusianorum</i>	0.25	0.11	0.53	1.78	0.20	1.31	0.03	8.79↓	3.08
<i>Digitalis lutea</i>	0.73	0.04	0.04	7.23↓	2.87	0.09	3.75	0.13	0.36
<i>Leontodon hispidus</i>	20.25↓	1.64	0.86	7.12↓	3.71	0.02	29.15↓	2.85	0.53
<i>Melica ciliata</i>	0.36	0.19	2.43	1.13	9.58↑	0.60	2.49	22.30↑	1.40
<i>Pimpinella saxifraga</i>	1.50	2.54	0.48	4.40↓	1.75	0.02	3.38	0.05	2.15
<i>Sedum album</i>	5.01 ↓	0.04	0.65	12.24↓	0.95	0.79	38.40↓	0.69	0.48
<i>Teucrium chamaedrys</i>	1.75	0.39	0.97	3.41	0.76	0.60	4.98↓	1.53	0.42