

Investigating sex differences in genetic relatedness in great-tailed grackles in Tempe, Arizona to infer potential sex biases in dispersal

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Abstract

In most bird species, females disperse prior to their first breeding attempt, while males remain closer to the place they hatched for their entire lives. Explanations for such female bias in natal dispersal have focused on the resource-defense based monogamous mating system that is prevalent in most birds. In this system, males are argued to benefit from philopatry because knowing the local environment can help them to establish territories to attract females, while females are argued to benefit from dispersing because they can find suitable unrelated mates. However, theoretical, field, and comparative studies highlight that the factors shaping dispersal decisions are often more complex. Studying species with different social and mating systems can help illuminate the relative role of various factors in the evolution of sex biased dispersal. Here, we use genetic approaches to determine whether females and/or males disperse in great-tailed grackles (*Quiscalus mexicanus*), which have a mating system where the males hold breeding territories that multiple females might choose to place their nest in, but females forage independently of these breeding territories across a wider area. First, we find that, for individuals caught at a single site in Arizona, the average relatedness among all female dyads is higher than average relatedness among other individuals at the site, whereas average relatedness among all males dyads is not. Second, we find that female close relatives are

32 found within shorter distances from each other than pairs of unrelated females, whereas male close relatives
33 are found at larger distances from each other than pairs of unrelated males. Third, we find a decline in
34 relatedness with increasing spatial distances for females, but not for males. Our results indicate sex biases
35 in relatedness structure that differ from most other bird species. Female great-tailed grackles associate with
36 close genetic relatives, presumably by remaining close to where they hatched which would lead to them
37 remaining close to their mothers and sisters. Males are not found close to genetic relatives, suggesting
38 that they disperse away from their fathers and brothers. Our findings show that great-tailed grackles offer
39 a relevant study system to further understand the factors shaping natal philopatry and dispersal, given this
40 reversal of the usual sex-bias in dispersal in line with their divergent social and mating system.

41 Introduction

42 Maturing birds face a decision about where to establish themselves for breeding. In the majority of avian
43 species, the potential costs and benefits of breeding movement decisions appear to differ between the sexes,
44 with males remaining in the area they hatched while females move to breed elsewhere (Greenwood 1980).
45 The main theory proposed to explain this sex bias towards male philopatry has focused on the resource-
46 defense based monogamous mating system found in most bird species (Greenwood 1980; Trochet et al.
47 2016). In monogamous systems, males tend to stay philopatric to defend an area they know to provide
48 resources to attract females, whereas females disperse to avoid the risk of inbreeding with close relatives
49 who dominate reproduction in the area. However, alternative hypotheses about the benefits and costs of
50 philopatry or dispersal could equally apply to explain the dominant female bias in dispersal among species
51 with resource defense based monogamy. In general, it is likely that, in both sexes, decisions of whether to
52 remain in the area or to move short or substantial distances to new breeding grounds are influenced by an
53 interplay of the potential costs of movement, resource availability and competition, and the potential benefits
54 or costs of interacting with close relatives (Mabry et al. 2013; Trochet et al. 2016; Li and Kokko 2019). One
55 way toward a better understanding of the relative role of the various factors that potentially explain breeding
56 movement decisions of both female and male birds is to study dispersal in species with different social and
57 mating systems.

58 Studying dispersal outside of well established study systems is difficult, which means that there is only lim-
59 ited information from bird species with unusual social and mating systems. It is challenging to set up studies
60 that span a large geographical area where the identity of many individuals can be established and followed.
61 As such, the fate of individuals who leave the area often remains unknown and it is unclear whether new indi-
62 viduals found in the area have moved to the area or were simply not observed previously (Walters 2000). To
63 overcome these challenges, genetic approaches are now incorporated to identify dispersal patterns (Law-
64 son Handley and Perrin 2007; Banks and Peakall 2012). In particular, to identify potential sex biases in
65 dispersal, two approaches are used. The first approach relies on determining the spatial distribution of vari-
66 ants of genetic markers that have a sex-specific inheritance (Lawson Handley and Perrin 2007). The second
67 approach uses data from a large number of genetic markers spread across the genome to determine how
68 the similarity across these markers changes with increasing spatial distances among males and females
69 (Banks and Peakall 2012). Studies based on the second approach have increased in recent years because
70 the costs of generating genotypes for a large sample of individuals have rapidly decreased (Harrison, York,
71 and Young 2014; Weinman, Solomon, and Rubenstein 2015; and Thrasher et al. 2018).

72 Here, we investigate SNP (single nucleotide polymorphism) genotype data for a sample of great-tailed
73 grackle (*Quiscalus mexicanus*) females and males at a single site. Great-tailed grackles differ in several
74 aspects from the majority of bird species in which dispersal has been investigated thus far, which might
75 make them a relevant study system to gain further insights into the factors shaping the dispersal decisions
76 of females and males. Great-tailed grackles are a highly social passerine bird found in the Americas. Individ-
77 uals forage year-round in small fission-fusion groups in areas that are not obviously defended against other
78 individuals and at night they roost in large associations (Johnson and Peer 2001), unlike most other bird
79 species where, at least during the breeding season, pairs or families defend foraging territories (Cockburn
80 2006). This could indicate that resource competition might be lower in great-tailed grackles, potentially re-
81 ducing pressure to remain in or move to high quality areas. Essentially everywhere they occur, great-tailed
82 grackles live in human-modified environments (MacGregor-Fors et al. 2009) and their wide range of forag-
83 ing habits routinely includes exploiting human foods (King 2012). In these environments, they can occur
84 in large numbers and at high densities (Escobar-Ibáñez, Rueda-Hernández, and MacGregor-Fors 2020).

85 Great-tailed grackles have recently extensively expanded their geographic range (Wehtje 2003), indicating
86 that they are highly mobile. Great-tailed grackles are sexually dimorphic, with males being larger than fe-
87 males and differing in plumage. During the mating season, some males defend territories around suitable
88 breeding habitats and mate with females who build their nests in these territories. Holding a territory leads
89 to higher reproductive success for these males, but females also mate with roaming males, leading to a
90 polygamous mating system (Johnson et al. 2000). This resembles the mating system observed in many
91 mammalian species, where males disperse to areas with the highest number of potential mates (e.g. Höner
92 et al. 2007). Previously, great-tailed grackle females were assumed to perform all activities related to off-
93 spring care, from building the nest through incubating and feeding the hatchlings, but observations indicate
94 that at least some males partake in these activities (Selander 1970; Folsom et al. 2020). Both the mating and
95 the social system are accordingly different from the resource-defense based monogamous system found in
96 the majority of birds, which might lead to a deviation from female-biased dispersal. Determining patterns
97 of philopatry and dispersal in great-tailed grackles can offer further insights into the potential association
98 between dispersal decisions and the various factors that might shape them.

99 **Hypotheses**

100 Our main hypothesis assumes that great-tailed grackles show a pattern of female-bias in dispersal. It is
101 our main hypothesis because this dispersal pattern predominates across birds and dispersal patterns are
102 often retained from a common ancestor; in addition, the factors that shape this pattern might still operate
103 in great-tailed grackles. Our alternative hypotheses expect that some of the differences in the social and
104 mating system of great-tailed grackles might lead to a deviation from this dispersal pattern. We set these as
105 alternative hypotheses because it is unclear which factors might be important. With the setup of our study,
106 we cannot infer why or how dispersal patterns might have changed, therefore we present these hypotheses
107 simply as alternatives.

108 **Hypothesis** There are sex differences in the natal dispersal rate and distance among individuals in great-
109 tailed grackles (*Quiscalus mexicanus*) with males remaining close to where they hatched and females mov-
110 ing away from where they hatched. Males are expected to remain close to the area where they hatched,
111 therefore a large number of the males on the Arizona State University (ASU) campus are expected to have
112 hatched within the area of the study site and stay close to their relatives. In contrast, females are expected
113 to move before their first breeding attempt (Greenwood 1980), therefore females on campus are likely to
114 come from areas outside of campus in the surrounding area, having moved away from relatives.

115 **Alternative hypothesis 1** Males disperse away from where they hatched, while females remain where they
116 hatched.

117 **Alternative hypothesis 2** Individuals of both sexes remain close to where they hatched.

118 **Alternative hypothesis 3** Individuals of both sexes disperse away from where they hatched.

119 We expect that the movement of individuals will influence the spatial distribution of genetic relatives (Aguillon
120 et al. 2017). Individuals of the sex that remains close to where they hatched are expected to be close to
121 genetic relatives, while individuals of the sex that disperses are expected to be more distant from genetic
122 relatives. We also expect that the further the distance an individual moves, the less likely they are to be
123 even distantly related to another individual within the study area. Our hypotheses generate specific predic-
124 tions about contrasts in the levels of relatedness and the spatial distribution of genetic relatives according to
125 whether individuals are philopatric or disperse. The first analysis (analysis i: average levels of relatedness

126 among individuals in our sample) focuses on whether individuals disperse beyond the trapping area and
127 compares one average value of relatedness per sex. Here, we predict higher levels of average related-
128 ness among all individuals of the philopatric sex than among all individuals of the sex that disperses. This
129 follows if some dispersing emigrants move outside of the trapping area, away from parents and siblings,
130 while immigrants can come from a variety of areas outside of the trapping area and therefore consist of
131 unrelated individuals. The second analysis (analysis ii: geographic distances between individuals that are
132 close genetic relatives) focuses on the distances among close relatives of the same sex that are trapped
133 within our trapping area and investigates the pairwise distances among individuals of the same sex who are
134 closely related. Here, we predict that there are sex biases in the average distances between trapping sites
135 for relatives compared to non-relatives because philopatric individuals will remain close to same-sex parents
136 and siblings while individuals that disperse within the trapping area will end up in different locations than
137 their same-sex parents and siblings. The third analysis (analysis iii: spatial autocorrelation) focuses on how
138 relatedness among pairs of same sex individuals changes as the distance between them increases and
139 investigates correlations among all estimates of pairwise relatedness and pairwise geographic distances
140 among individuals of the same sex. Here, we predict a decline in levels of relatedness as distances among
141 individuals increase to indicate that individuals have remained philopatric such that close relatives are found
142 in close geographic proximity. In contrast, we predict no structure of relatedness in geographic space for
143 individuals who disperse because relatives will be found both close and far from each other.

144 **Deviations from the preregistration**

145 We preregistered our hypotheses, methods, and analysis plans. Analyses began in March 2020 after the
146 preregistration passed pre-study peer review at *Peer Community In Ecology* in November 2019. The final
147 version of the preregistration can be found here <http://corinalogan.com/Preregistrations/gdispersal.html>.

148 We made the following changes and additions to the analyses (explained in detail in the Methods section
149 below):

- 150 - we adjusted the sample to exclude juveniles and reflect sex composition
- 151 - we used a more stringent filter for the ddRadSeq single nucleotide polymorphism
- 152 - we excluded one genotype with implausible levels of heterozygosity
- 153 - we chose the Queller & Goodnight estimator to calculate pairwise relatedness
- 154 - we added an analysis to assess whether the geographic distances among closely related females were
155 shorter than those in a random sample of females
- 156 - we added a configuration to the spatial correlogram analyses, based on the observed distances among
157 related individuals

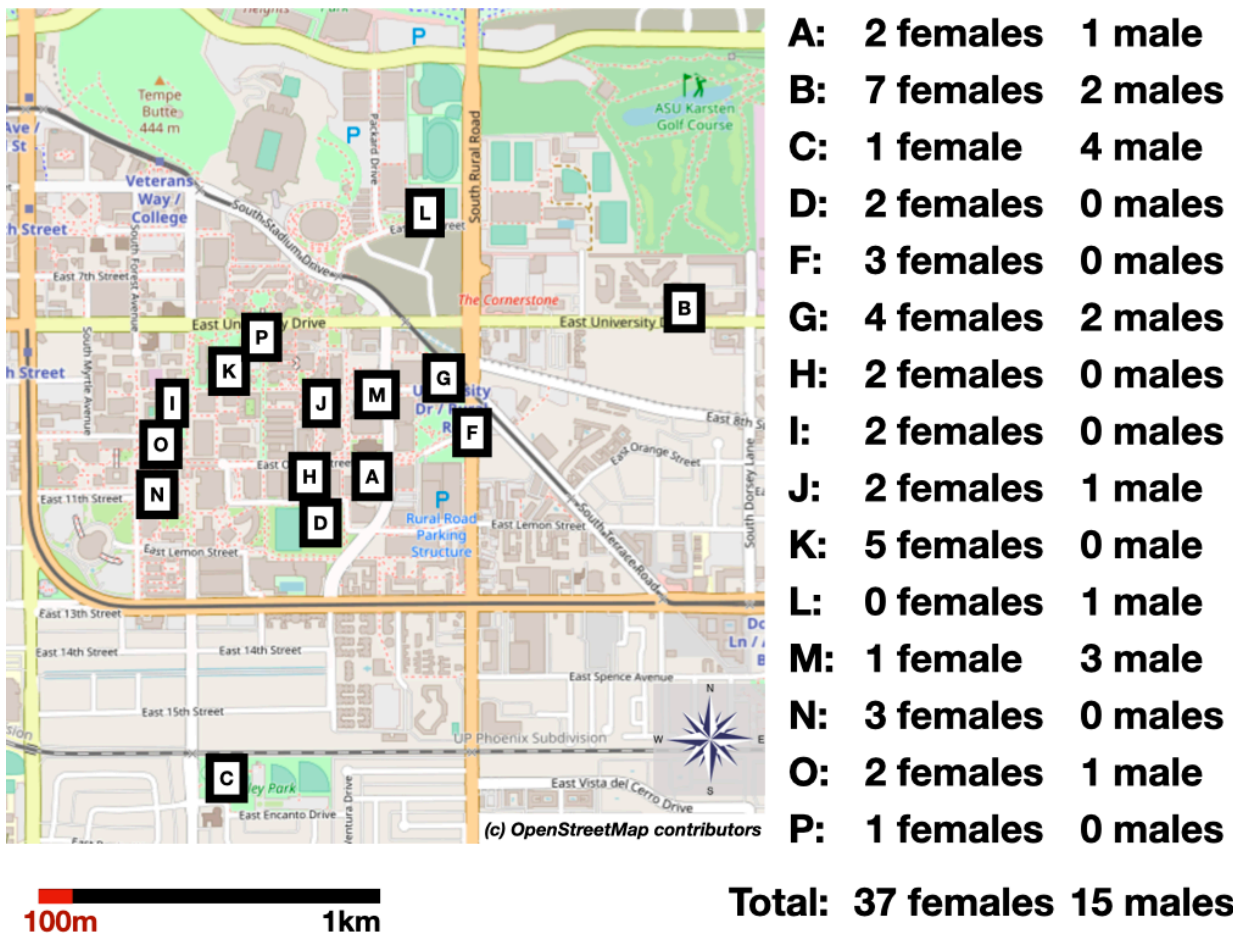
158 **Methods**

159 The methods below are based on the preregistration, with small changes summarized in the Deviations from
 160 the preregistration section above and further explained here (in italics).

161 **Data**

162 **Sample** DNA from 57 great-tailed grackles was obtained from wild individuals caught in Tempe, Arizona,
 163 USA (see Figure 4 for a map showing the trap locations and sample sizes for the individuals included in
 164 the analyses). These individuals were either immediately released, or temporarily brought into aviaries for
 165 behavioral testing and then released back to the wild.

166 *Deviation from preregistration: During the preparation of the analyses, we noticed that we made a mistake*
 167 *when calculating the sex composition in the sample: different from what was written in the preregistration,*
 168 *the sample for our genetic analyses consists of 41 (not 40) females and 16 (not 17) males. In addition, we*
 169 *realized that the sample included some juvenile individuals (<1 yr of age). We excluded these four juveniles*
 170 *from the main analyses because they might have been pre-dispersal at the time of capture. In addition, we*
 171 *excluded one individual after genotyping (see below). The dataset for the all analyses therefore consisted*
 172 *of 37 adult females and 15 adult males.*



173
 174 **Figure 1.** Map displaying the sampling locations of grackles on the Arizona State University campus and
 175 the number of great-tailed grackles trapped at each location as part of this research.

176 The larger number of females than males in our sample appears to reflect the adult sex ratio at this study site.
177 To estimate the sex ratio at the field site, we counted the number of females and males that were trapped
178 in mist nets since the beginning of our study (September 2017 - October 2019). This trapping method likely
179 does not elicit a sex bias in terms of which sex is caught because the nets are invisible. Therefore, if one
180 sex is more neophobic than the other, both sexes are likely to be trapped using this method. A total of 26
181 females and 11 males were trapped using mist nets (a ratio of 2.36 females per 1 male), which is very similar
182 to the sex ratio in our sample consisting of 37 adult females and 15 adult males (2.47 females per 1 male).

183 Females were caught at all but one site, such that comparisons are possible of the genetic relatedness of
184 pairs of females trapped at various distances from each other. Males were not caught at all trap sites, but
185 there are several sites at which multiple males were caught and sufficient sites for comparisons of males
186 that were caught close to each other, and at intermediate and large distances from each other.

187 **Sample size rationale** The sample size presented was the largest one possible (due to the time required
188 for trapping) by July 2019 when the DNA were sequenced using ddRADseq.

189 **Data collection stopping rule** We analyzed all blood samples that were collected through June 2019,
190 which was the end of the trapping season.

191 **Open data** All data necessary for the analyses are available at <https://doi.org/10.5063/F1W66J48> (Lukas
192 2020) and at GitHub (the code included in the R markdown file will load these files directly from GitHub).
193 The raw genetic data is available at <https://www.ncbi.nlm.nih.gov/sra?term=SRP278340>.

194 **Randomization and counterbalancing** No randomization or counterbalancing is involved in this study.

195 **Blinding of conditions during analysis** Experimenters were blind to the sex of the bird when processing
196 samples using ddRADseq (only the alphanumeric bird ID was visible on the tube and no team member who
197 handled the samples memorized which ID goes with which bird because we give the birds names).

198 **Blood collection** Whole blood samples were collected from individual birds by brachial or medial
199 metatarsal venipuncture. Blood was collected and stored in one of two ways until DNA extraction:

- 200 1) At the beginning of the project (2018), 70uL of whole blood was added to silicone-coated micro-blood
201 collection tubes containing 280uL of lysis buffer (White and Densmore 1992, 50–51) and stored at
202 room temperature for up to a year before DNA extraction.
- 203 2) In 2018 a different method was implemented, using DNA from packed red blood cells: 150uL of blood
204 was collected from trapped great-tailed grackles and stored for a minimum of 30 minutes and a max-
205 imum of 60 minutes at room temperature or 3 hours on ice. Samples were then centrifuged at 15x
206 gravity for 10 minutes to separate the serum from the cellular fraction. After the serum layer was re-
207 moved and stored, 600uL lysis buffer (White and Densmore 1992, 50–51) was added to the remaining
208 packed cells. Tubes containing packed cells and lysis buffer were stored at room temperature for up
209 to 1 year before extraction.

210 **DNA extraction and quantification** Some samples were extracted at Arizona State University by Rowney
211 (samples through Dec 2018), while others were shipped with ice packs to Washington State University
212 for extraction by Blackwell and his lab (samples collected Jan-Jun 2019). DNA was extracted from the
213 above samples using the DNeasy Blood and Tissue kit (Qiagen) with slight modifications from the manu-
214 facturer's protocol (see details in Thrasher et al. (2018) Supporting Information, page 7). Approximately
215 100ul of blood/lysis mixture was mixed with 20ul Proteinase K, 150ul PBS, and 200ul buffer AL, then incu-
216 bated overnight at 64C while shaking. Samples were mixed with 200ul ethanol and added to spin columns.
217 Columns were centrifuged and washed according to kit protocol using buffers AW1 and AW2. DNA was
218 eluted into 50ul of RNase and DNase free water at 64C after a 5-10 min incubation on columns. DNA quan-
219 tification was then performed on a Qubit 4.0 Fluorometer (Fisher Scientific) following the manufacturer's
220 protocol for broad range dsDNA. The average yield of samples used for sequencing was 34ng/ul. Extracted
221 DNA samples were shipped with ice packs to the Cornell Lab of Ornithology for ddRAD sequencing in July
222 2019.

223 **ddRAD sequencing** We generated single-nucleotide polymorphism (SNP; where at a given position in
224 the genome two different bases, alleles, can occur) genotypes for 57 individuals from our study site in Ari-
225 zona (we excluded the above mentioned 5 individuals later). The DNA was processed using ddRADseq by
226 Sevchik and Bronwyn Butcher (Cornell University) following methods in Thrasher et al. (2018). Each of the
227 samples' DNA concentrations was measured using the Qubit dsDNA BR Assay Kit and the Qubit Fluorome-
228 ter following the manufacturer's protocol. For this particular experiment, the necessary DNA concentrations
229 were between 5-50ng/ul and so any sample outside of this range needed to be normalized. Those samples
230 with a concentration higher than 50ng/ul were diluted to approximately 25ng/ul with nuclease-free water. For
231 those samples with concentrations lower than 5ng/ul, both elutions were pooled and the DNA concentrated
232 by evaporation using an Eppendorf Vacufuge. The DNA extracts are then run through a PCR thermocycler
233 where the fragments are digested with a combination of two restriction enzymes (SbfI-HF and MspI) and
234 20 different adapters attached to the end of the DNA pieces. A 1% agarose gel is run to ensure the proper
235 digestion and ligation of the DNA samples. The samples are then cleaned up using MagNA beads and size
236 selected using BluePippin for a prespecified length (between 400-700 base pairs). After the samples return
237 from size selection, they are amplified using a low-cycle PCR process and pooled together to be sent in to
238 be sequenced. Sequencing was performed on an Illumina NextSeq500 (using a mid-output kit and run with
239 Illumina PhiX control (15%) to aid sequence alignment) to generate 150 bp single end reads at the Core
240 Facilities of the Cornell Institute of Biotechnology.

241 **SNP processing** These data were post-processed to generate SNP data for relatedness analyses as in
242 Thrasher et al. (2018). After filtering reads for quality and demultiplexing to assign sequences back to
243 specific individuals, genetic loci were assembled *de novo* because no reference genome exists for great-
244 tailed grackles.

245 *Deviation from preregistration: For the ddRadSeq single nucleotide polymorphism (SNP) filtering, two sets*
246 *of restrictions were applied to maximize power by focusing the analysis on the most informative SNPs while*
247 *reducing the potential risk of noise. The first filter was based on the parameters set forth by Thrasher et*
248 *al. (2018), which they showed to provide a set of loci with strong power to discern relationships among*
249 *individuals. Loci were only considered if they were present in 80% of the samples (*r* parameter of the stacks*
250 *pipeline) and had a minimum frequency of the minor allele of 0.05 (min maf parameter of the stacks pipeline).*
251 *This meant that the rare variant at a locus was present in at least 5% of the samples and it resulted in 3647*

252 *acceptable SNPs for analyses. The second filter applied more stringent conditions for the loci to be retained.*
253 *Loci were only considered if they were present in 95% of the samples (r) and had a minimum minor allele*
254 *frequency of 0.05 (min maf). This resulted in 635 acceptable SNPs; 3012 SNPs fewer than in the first,*
255 *less restrictive filtering, but still more than in the study by Thrasher et al. (2018) because each individual*
256 *in our sample had been sequenced to a greater depth. We decided to use the resulting genotypes from*
257 *the second, more restrictive setting for the relatedness analyses because of our small sample size (e.g.,*
258 *if some individuals had a lower quality sample, their relatedness to other individuals might consistently be*
259 *misclassified) and because these settings still provided an effective number of SNPs for analyses. The*
260 *more restrictive filtering reduces noise from missing data and retains high power by selecting loci with high*
261 *heterozygosity (their heterozygosity approaches the maximum of 0.5) [Morin et al. 2004].*

262 *Addition to preregistration: The preregistration had focused on analyses to address the hypotheses. We*
263 *realized that we should also provide descriptive statistics of the genotype data we generated specifically for*
264 *this study. We used functions in the R packages ‘adegenet’ (Jombart 2008), ‘pegas’ (Paradis 2010), and*
265 *‘popgenutils’ (Tourvas 2020) to edit the genotype data and to calculate, based on the allele frequencies in the*
266 *data and assuming random mating, the expected heterozygosity (average chance of finding two different*
267 *alleles across loci across individuals) and probability of identity (chance that two individuals will have the*
268 *same set of alleles across all loci).*

269 **Relatedness estimation** Genetic relatedness between all pairs of individuals was calculated using the
270 package “related” (Pew et al. 2015) in R, following methods in Thrasher et al. (2018). We used the frequen-
271 cies of the alleles at the generated SNPs to calculate relatedness among pairs of individuals, with individuals
272 being classified as related if they share more alleles than what is expected based on random chance given
273 the frequencies of variants in the population (relatedness (R) >0) and as unrelated if they share as many
274 ($R=0$) or fewer genetic variants than expected ($R<0$). We planned to use the function ‘compareestimators’
275 to assess which relatedness estimator appears to perform the best given the characteristics of our data.

276 *Deviation from preregistration: The ‘compareestimator’ function in the R package ‘related’ caused fatal*
277 *errors on multiple computers. We therefore calculated pairwise relatedness using two estimators: 1) the*
278 *Wang estimator (Wang 2002), following the observation in Thrasher et al. (2018) of the suitability of this*
279 *estimator for ddRadSeq data, and 2) the Queller-Goodnight estimator (Queller and Goodnight 1989), which*
280 *is the standard in several studies. With both sets of relatedness estimates, all of our inferences (high levels*
281 *of average relatedness among females, shorter distances among closely related females, spatial structure*
282 *among female genotypes) were similar. We present the results based only on the estimator by Queller &*
283 *Goodnight (1989) because we noticed that, with our data, the estimator by Wang (2002) appeared to be*
284 *more influenced by missing data in the genotypes. We calculated average relatedness between all pairs*
285 *of individuals within one sex: the arithmetic mean of the estimated relatedness based on sharing of SNP*
286 *alleles among all female dyads and all male dyads.*

287 *Additional note: Our preregistration did not include plans to perform pedigree reconstructions as an alter-*
288 *native way to assess relatedness among the individuals for three reasons. First, we have a cross-sectional*
289 *sample, which does not contain longitudinal information from tracking juveniles seen with their potential par-*
290 *ents into adulthood. Second, adults are of unknown age, so for any related individuals who share an allele*
291 *at (almost) all loci we would not be able to determine which is the parent and which is the offspring. Third,*
292 *grackles are not expected to have large clusters of siblings (Johnson et al. 2000), as for example in fish*
293 *species, making it highly unlikely that our sample contains extended families.*

294 **Additional variables**

- 295 1) Sex (female, male): the sex of the individuals assigned based on morphological features
296 2) Distance between trap sites (meters): straight line distance (assuming earth is an ellipsoid) between
297 all pairs of trapping locations based on the longitude and latitude of each site.

298 **Analyses**

299 Analyses were conducted in R (current version 4.0.3; R Core Team (2017)).

300 We did not plan to exclude any data. We did not have to exclude individuals because more than half of their
301 genotype was unknown.

302 *Deviation from preregistration: After obtaining the SNP data, we noticed that one individual, female A053PS,*
303 *was indicated to be related to all other individuals in the sample. When we checked the genotype of this*
304 *individual, it had much higher levels of heterozygosity, independent of how we filtered the data. The genotype*
305 *did not show signs of shifting of alleles between loci (e.g., showing a genotype that lists allele 2 from locus*
306 *1 with allele 1 from locus 2) or have issues with missing data or the dropout of alleles. Instead, the increase*
307 *in heterozygosity was always close to the square of the heterozygosity observed among the remaining*
308 *individuals, suggesting that this genotype might be constructed from two different individuals. We were not*
309 *able to retrace where such an error might have been introduced, therefore we decided to exclude individual*
310 *A053PS from the analyses.*

311 **Ability to detect actual effects** Birds the size of a grackle (~100-150 grams) are expected to show a me-
312 dian natal dispersal distance of about 250-300 meters (Sutherland et al. 2000). Our 15 trap locations were
313 located within a ~1000m radius circle, suggesting that if there are dispersers in our sample, these individuals
314 will have most likely come from areas outside of the trapping circle. In turn, if individuals remain close to
315 their natal area, they would only move distances much shorter than this, suggesting that the pairwise dis-
316 tances between non-dispersed relatives would be shorter than the random distance between any two birds
317 we caught. However, we do not know the average distance that either philopatric or dispersing individuals
318 move. The scale of our sampling area might be so small that individuals of the sex that disperses the least
319 are likely to have hatched outside of this area. In addition, there could be variation among either females
320 or males in the distances individuals move, with potentially also a small proportion of individuals of the pre-
321 dominantly philopatric sex dispersing, which could obscure patterns in the small sample of individuals in our
322 study. Accordingly, we might not be able to detect differences in average relatedness between females and
323 males (analysis i), but we still might expect a sex bias in the geographic distances among relatives (analysis
324 ii).

325 We restricted our sample to adults to focus on the distribution of individuals after any potential natal dis-
326 persal (Goudet, Perrin, and Waser 2002). We only have individuals from within a single site, so we did
327 not use methods that rely on assigning individuals to a source population or measure the relative distribu-
328 tion of genetic variation within versus among populations (Fst or similar measures). We therefore relied
329 on measuring genetic relatedness between pairs of individuals. Approaches relying on spatial analyses of
330 multi-locus genotypes have been shown to be able to detect even modest sex biased dispersal in fine-scale
331 spatial distribution, in particular analyses of spatial autocorrelation (Banks and Peakall 2012). However, our
332 sample size is small, meaning that we might have only limited power to detect potential differences between
333 females and males (Goudet, Perrin, and Waser 2002). For the spatial distribution of relatives (Analysis ii),

334 the number of related individuals in our sample might be too small to detect a strong pattern of the relatives
335 of one sex being more geographically closer to each other than relatives of the other sex. For the isolation-
336 by-distance leading to a change in relatedness within the range of our sampling locations, the signal might
337 be too weak in either or both sexes to make inferences about sex differences (Analysis iii). However, for the
338 comparison of average relatedness (Analysis i), given that we have a large number of SNP loci, we expect
339 that we should have sufficient power to obtain a qualitative assessment of whether relatives are present in
340 our sample (Wang and Santure 2009) and accordingly whether dispersal is more prevalent in either females
341 or males. Examples of empirical studies that detected a signal with small sample sizes include Hofmann et
342 al. (2012); Quaglietta et al. (2013); Gour et al. (2013); and Botero-Delgadillo et al. (2017).

343 **Analysis i: average relatedness and sex** We compared the average and variance in relatedness among
344 all females to that among all males. Since average relatedness tends to decrease as the number of individ-
345 uals in the sample increases (regression to the mean), we performed a permutation analysis to investigate
346 whether the average relatedness among the males or among the females in our sample is higher than what
347 would be expected for a random sample of the same number of females or of individuals of both sexes.

348 *Deviation from preregistration: We adjusted the permutations to reflect the actual sex composition in our*
349 *sample of 37 adult females and 15 adult males, rather than drawing random samples of 40 and 17 individuals*
350 *as planned in the preregistration. We compared (i) the observed average relatedness among the 37 females*
351 *in our sample with the relatedness in the 10,000 random samples of 37 individuals from both sexes; (ii) the*
352 *observed average relatedness among the 15 males in our sample with the relatedness in the 10,000 random*
353 *samples of 15 individuals from both sexes; (iii) the observed average relatedness among the 15 males in*
354 *our sample with the relatedness in the 10,000 random samples of 15 females. We report the proportion of*
355 *10,000 random samples with lower relatedness than the observed values and, for comparison with other*
356 *approaches, assess whether the observed relatedness is higher than the relatedness calculated for 95% of*
357 *all random draws.*

358 **Analysis ii: distances among genetic relatives** Based on the calculations of pairwise genetic related-
359 ness, we selected the subset of pairs of individuals who are estimated to be more closely related than
360 cousins ($r \geq 0.125$) or half-siblings ($r \geq 0.25$). For this subset of closely related individuals, we first determined
361 whether the pairwise geographic distances are shorter than expected for males or for females (Coulon et al.
362 2006).

363 *Deviation from preregistration: In addition to the permutation to assess whether the difference in the average*
364 *distance among closely related females and the average distance among closely related males was larger*
365 *than expected, we performed a permutation to assess whether the average distance among closely related*
366 *female dyads ($r > 0.2499$) was shorter than the average distance among a random sample of the same*
367 *number of female dyads. We added this analysis of just females because the number of closely related*
368 *males was very small.*

369 We performed 10,000 draws of 12 (reflecting $r \geq 0.125$) random pairs of females and calculated the average
370 geographic distance among them. We assessed whether the observed difference in geographic distances
371 was larger than the majority of random samples and, for comparison with other approaches, determined
372 whether the observed distance was larger than that calculated for 95% of all random draws. We repeated
373 these comparisons for the more closely related females ($r \geq 0.25$), randomly drawing seven females 10,000
374 times. For the males, we randomly drew three pairs of individuals (reflecting the number of dyads related

375 at $r \geq 0.125$) and additionally randomly drew one pair of individuals (reflecting the one dyad related at $r \geq 0.25$)
376 10,000 times and compared the geographic distances among the randomly sampled dyads to the geographic
377 distances among the pairs of related males. Next, we performed 10,000 random draws of pairs of males
378 and pairs of females matching the numbers of closely related dyads for their sex. We calculated the differ-
379 ences between the average geographic distances among the subset of males and the average geographic
380 distances among the subset of females. We compared this to the observed difference in the distance among
381 closely related males and among closely related females.

382 **Analysis iii: spatial autocorrelation** To test whether males and females show different patterns of ge-
383 netic isolation by geographic distance, we followed analyses as in Aguillon et al. (2017). For the analysis,
384 we initially created 11 distance bins separated by 200m between 0m-2000m (the maximum distance be-
385 tween trapping sites). The 200m bin size was chosen because there are roosting trees that are ~50m apart
386 suggesting that dispersal might be occurring below this scale and also to maximize the number of pairs in
387 each distance class. The individuals in our sample were caught at one of 15 trap sites, and the resulting
388 105 pairwise distances among individuals was assigned to one of the 11 bins. In addition, we adjusted the
389 distances covered by each bin to have shorter distances for the first few bins to increase the chance of
390 detecting relatives within the smallest bins (changing from 11 equally sized 200m bins to, for example, 9
391 bins at varying distances such as 0-50m, 50m-100m, 100m-150m, 150m-200m, 200m-500m, 500m-750m,
392 750m-1000m, 1000m-1500m, 1500m-2000m) (following Peakall, Ruibal, and Lindenmayer 2003).

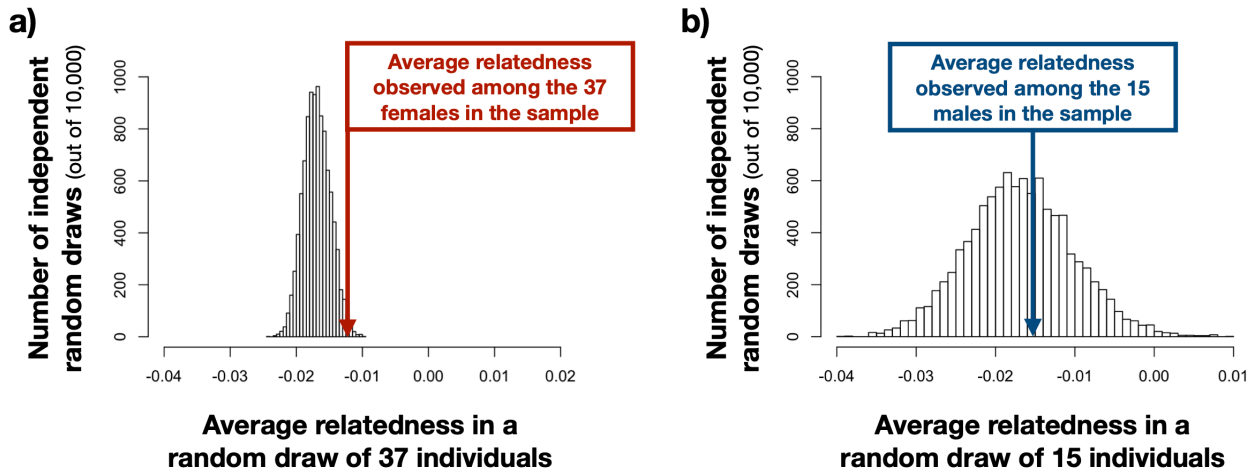
393 *Deviation from preregistration: For the correlogram analyses with set distance classes, we added a config-*
394 *uration where we set the distance classes using information on the average distance among close genetic*
395 *relatives from analysis ii. We had planned for this in the preregistration and decided to add the analyses*
396 *because the observed distances among close genetic relatives did not match the distance classes we had*
397 *initially planned. We spaced the distance classes such that the observed average distance among close*
398 *female kin (~330m) and among close male kin (~670m) fell about halfway between the breakpoints for the*
399 *distance classes (set at 0-150m, 150-450m, 450-900m, 900-1500m, 1500-2000m).*

400 For males and females separately, we linked the matrices of average relatedness and of geographic dis-
401 tance between all pairs of individuals by first plotting genetic relatedness against geographic distance and
402 next by assessing the strength of their association using Mantel correlograms. We used the function 'man-
403 tel.correlog' in the vegan package (Oksanen et al. 2013) in R, performing 10,000 permutations to assess
404 the strength of the association. This approach relies on the establishment of the multivariate Mantel correlo-
405 gram by Legendre and Legendre (2012). The approach involves partitioning the geographic locations into a
406 series of discrete distance classes. The result of this set of analyses is a Mantel's correlogram analogous to
407 an autocorrelation function but performed on a set of distance matrices. For each distance class, a separate
408 matrix is generated and codes whether a given geographic distance between a pair of individuals falls within
409 that range. A normalized Mantel statistic is calculated using permutations for each distance class. The per-
410 mutation statistics, plotted against distance classes, produce a multivariate correlogram. These analyses
411 are performed separately for each sex to determine whether isolation-by-distance might occur and indicate
412 dispersal of the individuals of that sex. A stronger negative correlation between genetic relatedness and
413 spatial distance for males than for females would indicate that males disperse shorter distances than fe-
414 males, and in particular we expect that males captured at the same trapping site will be much more closely
415 related to each other than females captured at the same trapping site.

416 **Results**

417 **Genotyping** After filtering the sequencing data to include only the most useful SNPs, our dataset consists
418 of 635 SNPs for 52 individuals. Data is missing for 2.7% of all alleles (individuals missing information for
419 either one or both of their chromosomes for that particular position), with no individual or SNP showing a
420 particular underrepresentation of information. All SNPs have 2 alleles and the observed heterozygosity (in-
421 dividuals carrying one copy each of the two bases) is 0.48, which is slightly higher than the heterozygosity
422 expected in a population with the same allele frequencies and random mating (0.46). The increased het-
423 erozygosity potentially reflects that inbreeding is rare, likely because individuals of one sex disperse prior to
424 breeding. The probability of identity for siblings, the chance that two siblings will show the same genotypes
425 given the allele frequencies across these 635 loci and random mating among individuals, is less than 10^{-139} .

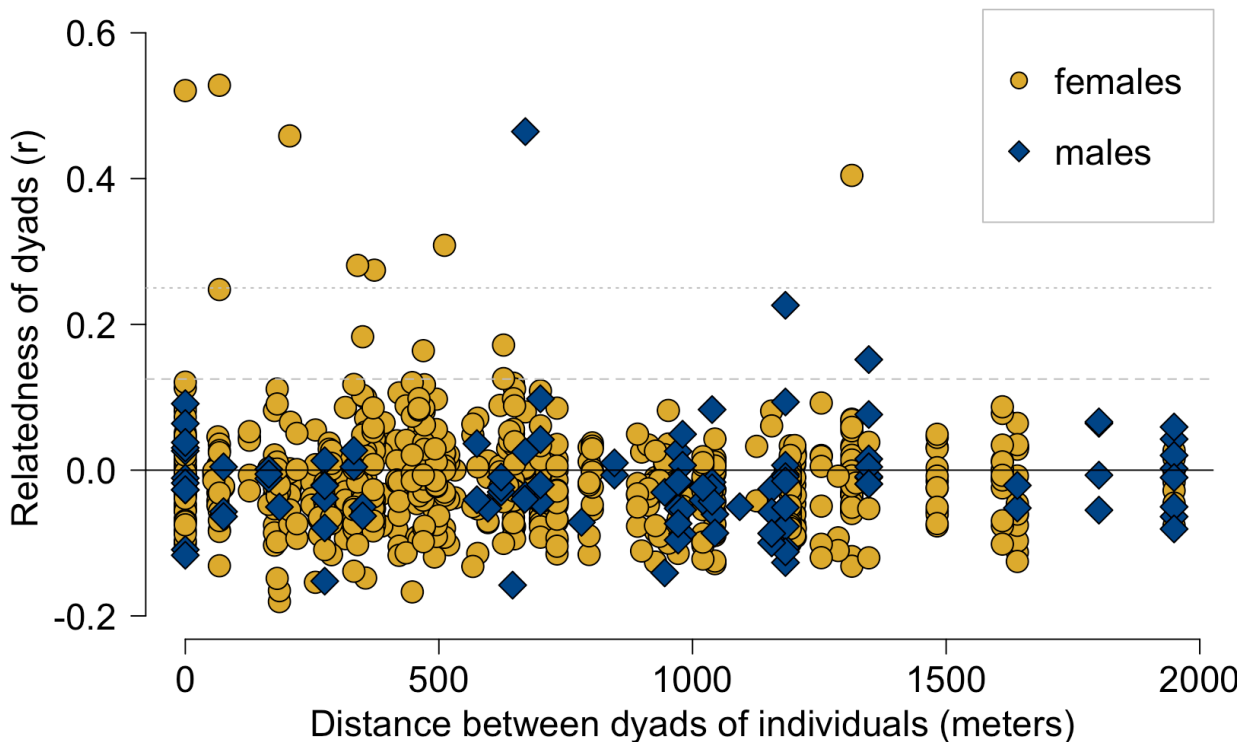
426 **Analysis i: average relatedness and sex** The average relatedness among the 37 adult females (666
427 dyads) is -0.013 (standard deviation, SD=0.07), the average relatedness among the 15 adult males (105
428 dyads) is -0.015 (SD=0.08), and the average relatedness among all 52 adult individuals in our sample (1326
429 dyads) is -0.017 (SD=0.07). To assess whether the average relatedness among females is higher than
430 expected, we compare it to the average relatedness calculated from random draws of 37 individuals from
431 all 52 individuals. In less than 4% of these draws is the level of relatedness as high as or higher than that
432 observed in our sample of females (Figure 1a). Therefore, although the difference in the level of average
433 relatedness among females compared to among all individuals is small (0.004), it is higher than expected
434 by chance. The average relatedness observed among the 15 males is not different from that expected by
435 chance among 15 randomly drawn individuals from the total 52 (40% of random samples give a value as low
436 as or lower than what we found in our sample of males; Figure 1b) or among 15 randomly drawn individuals
437 from the 37 females (61% of random samples give a value as low or lower than the male value). Of the eight
438 close genetic relatives (relatedness of 0.25 or higher), seven are female dyads and one is a male dyad, and
439 the majority of dyads are not related to each other (658/666 female dyads are not close relatives; 104/105
440 male dyads are not close relatives).



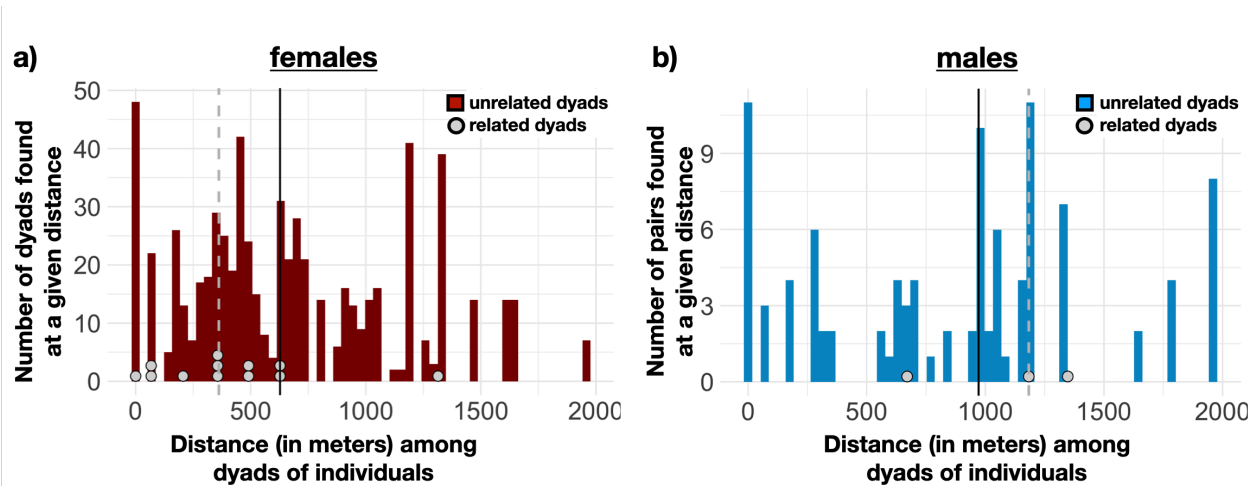
441

442 **Figure 2.** Females are more related than expected by random chance, whereas males are not. a) In less
443 than 4% of 10,000 repetitions is the average relatedness among the 37 randomly drawn individuals (of
444 both sexes) as high as or higher than the observed relatedness among the 37 females in our sample. b)
445 In contrast, average relatedness among 15 randomly drawn individuals (of both sexes) is higher than the
446 observed relatedness among the 15 males in our sample in 38% of 10,000 draws.

447 **Analysis ii: distances among genetic relatives** Close female genetic relatives are found to have been
 448 trapped in close spatial proximity to each other (Figure 2). The median distance between the eight female
 449 dyads related at 0.25 or higher is 340m (SD=440m) and between the twelve female dyads related at 0.125
 450 or higher is 360m (SD=354m), compared to a median of 620m (SD=464m) among all dyads of females
 451 (Figure 3). A median distance as short as or shorter than 340m is observed in less than 6% of all random
 452 samples of 7 female dyads and a median distance of 360m or shorter is observed in less than 4% of all
 453 random samples of 12 female dyads. The distance among the one pair of males related at higher than
 454 0.25 is 670m, and the median distance among the three male dyads related at 0.125 or higher is 1183m
 455 (SD=353m). This compares to a median of 972m (SD=569m) among all dyads of males, with about 40% of
 456 male dyads being 670m or less apart. The difference in distances among the 12 related females ($r \geq 0.125$,
 457 on average 360m apart) compared to the three related males ($r \geq 0.125$, on average 1183m apart) is 823m.
 458 This difference in distance (and larger differences in distance) is present in only 2% of 10,000 random draws
 459 comparing average distances among 12 random females and three random males.



460
 461 **Figure 3.** Change in genetic relatedness as geographic distance among dyads increases. Each dot reflects
 462 a single dyad, a pair of female individuals (yellow) or a pair of male individuals (blue). There are very
 463 few close male relatives who are found at larger distances. The small number of close female relatives
 464 are all found within relatively short distances of each other. The dotted horizontal line indicates the level of
 465 relatedness for half-siblings ($r=0.25$), the dashed line indicates the level of relatedness for cousins ($r=0.125$).



466

467 **Figure 4.** The geographic distance among dyads of closely related individuals (relatedness of 0.125 or
 468 higher; light circles) compared to the distance among dyads of unrelated individuals (colored bars). a)
 469 Among females, twelve closely related individuals were trapped at locations near each other (median distance
 470 indicated by dotted grey line), with eleven of the twelve closely related female dyads at distances as
 471 near as or nearer than the median of unrelated female dyads (vertical black line). b) In contrast, only one
 472 of the three closely related male pairs was trapped at locations that were as near as or nearer than the
 473 median distance among the unrelated males (vertical black line). The distances among the closely related
 474 males were about three times larger (median indicated by dotted grey line) than the distances among closely
 475 related females.

476 **Analysis iii: spatial autocorrelation** Correlogram analyses linking genetic relatedness and spatial distance
 477 for females showed negative values when females are in close spatial proximity and positive values
 478 when they are far apart (the corrected probability values for females are different than expected by chance
 479 in two of the five distance classes), suggesting that as spatial distance among females increases the relatedness
 480 among them decreases (Table 1). Correlogram analyses for males showed no consistent relationships
 481 between genetic relatedness and spatial distance, with values fluctuating around zero (none of the corrected
 482 probability values for males are different than expected by chance in any of the five distance classes; Table
 483 1).

484 **Table 1.** Output of correlogram analyses linking pairwise relatedness to pairwise distances. The values
 485 represent the correlations between relatedness and distances for males and females across trapping sites
 486 binned into distance classes, with the probability of observing the values by chance corrected for the multiple
 487 tests across distances classes (based on the Holm-Bonferroni method).

Distance class	Females: correlation	Females: corrected probability	Males: correlation	Males: corrected probability
0-150m	-0.10	0.01	-0.01	0.39
150-450m	0.02	0.32	0.09	0.37
450-900m	-0.05	0.25	-0.13	0.21
900-1600m	0.10	0.04	0.09	0.55
1600-2000m	0.01	0.66	-0.05	0.73

488 Discussion

489 Our results show that in great-tailed grackles, unlike in most other bird species but in line with their divergent
490 social and mating system, the majority of males are not philopatric because individuals are not found in close
491 proximity to fathers or brothers. In contrast, several female great-tailed grackles are found in close proximity
492 to genetic kin. The most likely explanation for this assortment of kin in space is that at least some females
493 remain close to where they hatched. Overall, the findings support the first alternative hypothesis that males
494 disperse more than females. We find that the mean level of average genetic relatedness is slightly lower
495 among males compared to females in our sample and that females are more closely related to each other
496 than expected by chance, while males are not (analysis i); the mean geographic distance between pairs of
497 individuals that are close genetic relatives is higher among males compared to females (analysis ii); and
498 there is no spatial relationship between genetic relatedness and geographic distance for males, while there
499 is a negative spatial autocorrelation signal indicating a negative relationship between genetic relatedness
500 and geographic distance for females (analysis iii).

501 Our small sample (we estimate that we trapped ~25-30% of all grackles within this area, which is continuously
502 connected to other areas in which grackles reside) and the limited number of genetic relatives we found,
503 restrict the inferences we can draw. The consistency of the results across the three types of analyses,
504 showing that at least some female great-tailed grackles remain close to relatives and most males disperse,
505 is reassuring and supports the inference that males disperse more than females. Previous studies relying
506 on spatial analyses of multi-locus genotypes were also able to detect even modest sex biased dispersal in
507 fine-scale spatial distribution (examples of empirical studies that detected a signal with small sample sizes
508 include Hofmann et al. 2012; Quaglietta et al. 2013; Gour et al. 2013; Botero-Delgadillo et al. 2017). In
509 particular, the effective number of SNP loci we have for each individual likely increased our power to obtain
510 a qualitative assessment of whether relatives are present in our sample and, accordingly, whether dispersal
511 is more prevalent in females or males based on spatial autocorrelation (Banks and Peakall 2012). However,
512 we cannot infer how substantial this sex bias is (our comparison of average relatedness between sexes is
513 inconclusive), what percentage of females and males might disperse, or how far they might move. In addition,
514 because we only have information for a small number of individuals from within a single site, we could not
515 use methods that rely on assigning individuals to a source population or measure the relative distribution of
516 genetic variation within versus among populations (F_{st} or similar measures). Such approaches could reveal
517 the proportion of individuals who do disperse and the distances they might move, something which we are
518 planning to investigate in the future (see Logan et al. 2020).

519 Our findings indicate that great-tailed grackles are a species that might help us better understand the factors
520 influencing the dispersal decisions of female and male birds. The reversal of the sex bias in great-tailed
521 grackles compared to what is observed in most other avian species is in line with the main hypothesis that
522 has been put forward to explain the contrast in sex biases in dispersal between birds and mammals: that
523 in polygynous species, males disperse to search for mating opportunities, while in monogamous species,
524 males remain philopatric to defend resources for high quality partners. However, given that the link between
525 the mating system and dispersal is more ambiguous than sometimes assumed (Li and Kokko 2019) and
526 given the limitations of our study, we cannot determine the underlying reasons for why males disperse or
527 why females apparently remain close to where they hatched. We only observe a general pattern of bias, but
528 we do not have sufficiently detailed information on the experiences of particular individuals that might have
529 shaped their dispersal behavior. Individual-based studies are needed to investigate resource and mating
530 competition and whether the patterning of relatives in space relates to kin-based social interactions and
531 inbreeding. In addition, information on dispersal patterns from different sites might help elucidate how much

532 the sex bias we detect at this site in the city center of Tempe is shaped by local factors or whether it is linked
533 to general features of the biology of great-tailed grackles.

534 Code

535 All data necessary for the analyses are available at <https://doi.org/10.5063/F1W66J48> (Lukas 2020) and at
536 github (the provided code will load these files directly from github).

```
options(width=60)

library(related)
library(tidyr)
library(dplyr)
library(vegan)
library(geosphere)
library(DataCombine)
library(data.table)
library(readr)

# SNP data, processed to calculate pairwise relatedness
input<-readgenotypedata("https://raw.githubusercontent.com/
                        corinalogan/grackles/master/Files/Preregistrations/
                        gDispersal_GrackleGenotypesForRelatedness.txt")

# Individual level data, listing the sex (M ale or F female), age (A dult or J uvenile), and latitude an
gracklelocations<-read_csv(url("https://raw.githubusercontent.com/
                              corinalogan/grackles/master/Files/Preregistrations/
                              gDispersal_GrackleIndividualInformationForRelatedness.csv"))
gracklelocations<-data.frame(gracklelocations)
```

537 Load data

```
options(width=60)
input$gdata$V1<-as.character(gracklelocations$Individual)
gracklelocations<-filter(gracklelocations,Individual != "AF_053PS")
adults<-filter(gracklelocations,Age %in% "A")[,]$Individual
adultgracklelocations<-filter(gracklelocations,Individual %in% adults)
```

538 Subset data to relevant sample, excluding juveniles and outlier genotype

```

options(width=60)

#Plot pairwise distances among all females and among all males in the sample
#Calculate all pairwise distances
all_pairwise_distances <- distm(adultgracklelocations[,c('Lon','Lat')], adultgracklelocations[,c('Lon',
rownames(all_pairwise_distances)<-adultgracklelocations$Individual
colnames(all_pairwise_distances)<-adultgracklelocations$Individual
diag(all_pairwise_distances)<-NA

#Calculate pairwise distances among all the females
female_pairwise_distances <- distm(adultgracklelocations[adultgracklelocations$Sex=="F",c('Lon','Lat')],
rownames(female_pairwise_distances)<-adultgracklelocations[adultgracklelocations$Sex=="F",]$Individual
colnames(female_pairwise_distances)<-adultgracklelocations[adultgracklelocations$Sex=="F",]$Individual
diag(female_pairwise_distances)<-NA

#Calculate pairwise distances among all the males
male_pairwise_distances <- distm(adultgracklelocations[adultgracklelocations$Sex=="M",c('Lon','Lat')],
rownames(male_pairwise_distances)<-adultgracklelocations[adultgracklelocations$Sex=="M",]$Individual
colnames(male_pairwise_distances)<-adultgracklelocations[adultgracklelocations$Sex=="M",]$Individual
diag(male_pairwise_distances)<-NA

#plot distributions of pairwise distances
hist(all_pairwise_distances,col="grey75",border="black",breaks=10)
hist(female_pairwise_distances,col="grey75",border="black",breaks=10)
hist(male_pairwise_distances,col="grey75",border="black",breaks=10)

```

539 **Calculate pairwise distances from trapping site locations**

```

options(width=60)

#Analysis 1: Assess whether average relatedness is higher among females or among males
#Calculate pairwise relatedness, here choosing the relatedness method developed by Wang and Queller & G
outfile<-coancestry(input$gdata,wang=1,quellergt=1)

#extract the relevant information from the file
pairwise_r<-outfile$relatedness

# We now exclude the individual with the dubious genotype and the juvenile individuals
pairwise_r<-filter(pairwise_r,ind1.id != "AF_053PS")
pairwise_r<-filter(pairwise_r,ind2.id != "AF_053PS")

# Next, we exculde all juvenile individuals
pairwise_r<-filter(pairwise_r,ind1.id %in% adults)
pairwise_r<-filter(pairwise_r,ind2.id %in% adults)

#This leaves us with 1326 pairwise relatedness values among the 52 remaining individuals

#identify which individuals are female and which are male
females<-filter(gracklelocations,Sex %in% "F",Age %in% "A")[,]$Individual
males<-filter(gracklelocations,Sex %in% "M",Age %in% "A")[,]$Individual

#Calculate average of and variance in relatedness among all individuals, all females, and all males
# First using the relatedness estimates based on the method by Wang
mean(filter(pairwise_r,ind1.id %in% females,ind2.id %in% females)$wang)
mean(filter(pairwise_r,ind1.id %in% males,ind2.id %in% males)$wang)
mean(pairwise_r$wang)

var(filter(pairwise_r,ind1.id %in% females,ind2.id %in% females)$wang)
var(filter(pairwise_r,ind1.id %in% males,ind2.id %in% males)$wang)
var(pairwise_r$wang)

# Next using the relatedness estimates based on the method by Queller and Goodnight
mean(filter(pairwise_r,ind1.id %in% females,ind2.id %in% females)$quellergt)
mean(filter(pairwise_r,ind1.id %in% males,ind2.id %in% males)$quellergt)
mean(pairwise_r$quellergt)

var(filter(pairwise_r,ind1.id %in% females,ind2.id %in% females)$quellergt)
var(filter(pairwise_r,ind1.id %in% males,ind2.id %in% males)$quellergt)
var(pairwise_r$quellergt)

```

```

#Perform a simulation to assess whether average relatedness among males is different from what we would
#First based on the relatedness estimates based on the method by Wang
simulatedrelatedness<-matrix(ncol=1,nrow=10000)
for (i in 1:10000) {
  currentset<-sample(adults,length(males))
  simulatedrelatedness[i,1]<-mean(filter(pairwise_r,ind1.id %in% currentset,ind2.id %in% currentset)$wang)
}
hist(simulatedrelatedness)
#This value is similar to a p-value, it reflects the probability that the average relatedness observed
sum(simulatedrelatedness>mean(filter(pairwise_r,ind1.id %in% males,ind2.id %in% males)$wang))/10000

#Perform a simulation to assess whether average relatedness among females is different from what we would
simulatedrelatedness<-matrix(ncol=1,nrow=10000)
for (i in 1:10000) {
  currentset<-sample(adults,length(females))
  simulatedrelatedness[i,1]<-mean(filter(pairwise_r,ind1.id %in% currentset,ind2.id %in% currentset)$wang)
}
hist(simulatedrelatedness)
#This value is similar to a p-value, it reflects the probability that the average relatedness observed
sum(simulatedrelatedness>mean(filter(pairwise_r,ind1.id %in% females,ind2.id %in% females)$wang))/10000

#Next based on the relatedness estimates based on the method by Queller & Goodnight
#Perform a simulation to assess whether average relatedness among males is different from what we would
simulatedrelatedness<-matrix(ncol=1,nrow=10000)
for (i in 1:10000) {
  currentset<-sample(adults,length(males))
  simulatedrelatedness[i,1]<-mean(filter(pairwise_r,ind1.id %in% currentset,ind2.id %in% currentset)$queller)
}
hist(simulatedrelatedness)
#This value is similar to a p-value, it reflects the probability that the average relatedness observed
sum(simulatedrelatedness>mean(filter(pairwise_r,ind1.id %in% males,ind2.id %in% males)$queller))/10000

#Perform a simulation to assess whether average relatedness among females is different from what we would
simulatedrelatedness<-matrix(ncol=1,nrow=10000)
for (i in 1:10000) {
  currentset<-sample(adults,length(females))
  simulatedrelatedness[i,1]<-mean(filter(pairwise_r,ind1.id %in% currentset,ind2.id %in% currentset)$queller)
}
hist(simulatedrelatedness)
#This value is similar to a p-value, it reflects the probability that the average relatedness observed
sum(simulatedrelatedness>mean(filter(pairwise_r,ind1.id %in% females,ind2.id %in% females)$queller))/10000

```



```

options(width=60)

#Analysis 2: Assess whether distances among closely related females are shorter than distances among cl
#First define close relatives as all pairs of individuals who are related by a level of 0.25 or higher
close_relatives_females<-filter(pairwise_r,wang >0.2499,ind1.id %in% females,ind2.id %in% females)
close_relatives_females_individuals<-c(close_relatives_females$ind1.id,close_relatives_females$ind2.id)

#Alternatively, select close relatives as pairs of individuals who are related at a level of 0.25 of hi
close_relatives_females<-filter(pairwise_r,quellergt >0.2499,ind1.id %in% females,ind2.id %in% females)
close_relatives_females_individuals<-c(close_relatives_females$ind1.id,close_relatives_females$ind2.id)

#Pick one of the two estimators before proceeding with the following analyses

#Next subset the the distance matrix to only include these individuals

females_pairwise_distances_matrix<-as.data.frame(female_pairwise_distances)
close_relatives_females_pairwise_distances<-matrix(nrow=nrow(close_relatives_females),ncol=1)

for (i in 1:nrow(close_relatives_females)) {
  ind1<-close_relatives_females[i,]$ind1.id
  ind2<-close_relatives_females[i,]$ind2.id
  pair_distance<-females_pairwise_distances_matrix[ind1,ind2]
  close_relatives_females_pairwise_distances[i,]<-pair_distance
}

median(close_relatives_females_pairwise_distances)

hist(close_relatives_females_pairwise_distances)

#repeat the same for the males
close_relatives_males<-filter(pairwise_r,wang >0.2499,ind1.id %in% males,ind2.id %in% males)
close_relatives_males_individuals<-c(close_relatives_males$ind1.id,close_relatives_males$ind2.id)

#Again, the alternative with the Queller & Goodnight method, pick only one of the two
close_relatives_males<-filter(pairwise_r,quellergt >0.2499,ind1.id %in% males,ind2.id %in% males)
close_relatives_males_individuals<-c(close_relatives_males$ind1.id,close_relatives_males$ind2.id)

#Next subset the the distance matrix to only include these individuals

males_pairwise_distances_matrix<-as.data.frame(male_pairwise_distances)
close_relatives_males_pairwise_distances<-matrix(nrow=nrow(close_relatives_males),ncol=1)

```

```

for (i in 1:nrow(close_relatives_males)) {
  ind1<-close_relatives_males[i,]$ind1.id
  ind2<-close_relatives_males[i,]$ind2.id
  pair_distance<-males_pairwise_distances_matrix[ind1,ind2]
  close_relatives_males_pairwise_distances[i,]<-pair_distance
}

median(close_relatives_males_pairwise_distances)

hist(close_relatives_males_pairwise_distances)

#calculate difference between the distances among males and among females
observeddifferenceindistances<-median(close_relatives_males_pairwise_distances,na.rm=T)-median(close_relatives_females_pairwise_distances,na.rm=T)

#perform simulation to generate random draws of matching numbers of individuals to assess whether the s
number_close_relatives_females<-nrow(close_relatives_females)
number_close_relatives_males<-nrow(close_relatives_males)

simulateddifferenceindistances<-matrix(ncol=1,nrow=10000)
simulateddfemaleindistances<-matrix(ncol=1,nrow=10000)
simulateddmaleindistances<-matrix(ncol=1,nrow=10000)

for (i in 1:10000) {
  simulated_close_relatives_females<-sample_n(pairwise_r, number_close_relatives_females, replace = TRUE)

  subset_relatives_females_pairwise_distances<-matrix(nrow=nrow(simulated_close_relatives_females),ncol=1)

  for (j in 1:nrow(simulated_close_relatives_females)) {
    ind1<-simulated_close_relatives_females[j,]$ind1.id
    ind2<-simulated_close_relatives_females[j,]$ind2.id
    pair_distance<-all_pairwise_distances[ind1,ind2]
    subset_relatives_females_pairwise_distances[j,]<-pair_distance
  }

  simulated_close_relatives_males<-sample_n(pairwise_r, number_close_relatives_males, replace = TRUE)

  subset_relatives_males_pairwise_distances<-matrix(nrow=nrow(simulated_close_relatives_males),ncol=1)

  for (k in 1:nrow(simulated_close_relatives_males)) {
    ind1<-simulated_close_relatives_males[k,]$ind1.id
    ind2<-simulated_close_relatives_males[k,]$ind2.id
    pair_distance<-all_pairwise_distances[ind1,ind2]
    subset_relatives_males_pairwise_distances[k,]<-pair_distance
  }
}

```



```
simulateddfemaleindistances[i,1]<-median(subset_relatives_females_pairwise_distances,na.rm=T)
simulateddmaleindistances[i,1]<-median(subset_relatives_males_pairwise_distances,na.rm=T)
simulateddifferencesindistances[i,1]<-median(subset_relatives_males_pairwise_distances,na.rm=T)-median(
  }

sum(simulateddfemaleindistances<median(close_relatives_females_pairwise_distances))/10000
sum(simulateddmaleindistances>median(close_relatives_males_pairwise_distances))/10000
sum(simulateddifferencesindistances>observeddifferenceindistances)/10000
```

541 ***Analysis ii: distances among genetic relatives***

```

options(width=60)

#Analysis 3: Correlogram to assess change of relatedness with distances

#have each value only once in the distance matrix
for (i in 1:ncol(all_pairwise_distances)) {
  all_pairwise_distances[i,i:ncol(all_pairwise_distances)]<-NA
}

#turn pairwise_r$wang into a matrix
all_relatedness<-select(pairwise_r,ind1.id,ind2.id,wang)
relatedness_matrix<-spread(all_relatedness,"ind1.id","wang")
relatedness_matrix <- cbind(relatedness_matrix,AF_061PR="NA")
relatedness_matrix<-arrange(relatedness_matrix,ind2.id)
relatedness_matrix <- InsertRow(data=relatedness_matrix, NewRow=rep("NA",53), RowNum=1)
relatedness_matrix[1,1] <- "AF_001YP"
rownames(relatedness_matrix)<-relatedness_matrix[,1]

#turn pairwise_r$quellert into a matrix
all_relatedness<-select(pairwise_r,ind1.id,ind2.id,quellert)
relatedness_matrix<-spread(all_relatedness,"ind1.id","quellert")
relatedness_matrix <- cbind(relatedness_matrix,AF_061PR="NA")
relatedness_matrix<-arrange(relatedness_matrix,ind2.id)
relatedness_matrix <- InsertRow(data=relatedness_matrix, NewRow=rep("NA",53), RowNum=1)
relatedness_matrix[1,1] <- "AF_001YP"
rownames(relatedness_matrix)<-relatedness_matrix[,1]

relatedness_matrix<-relatedness_matrix[1:52,2:53]

female_relatedness_matrix<-relatedness_matrix[rownames(relatedness_matrix) %in% females,colnames(relatedness_matrix) %in% females]
male_relatedness_matrix<-relatedness_matrix[rownames(relatedness_matrix) %in% males,colnames(relatedness_matrix) %in% males]

#perform the correlogram analysis
#first way, defining the distance classes
female_correlogram_setdistances<-mantel.correlog(D.eco=female_relatedness_matrix,D.geo=female_pairwise_distances)
male_correlogram_setdistances<-mantel.correlog(D.eco=male_relatedness_matrix,D.geo=male_pairwise_distances)
#second way, setting the number of distance classes
female_correlogram_classes<-mantel.correlog(D.eco=female_relatedness_matrix,D.geo=female_pairwise_distances,n=5)
male_correlogram_classes<-mantel.correlog(D.eco=male_relatedness_matrix,D.geo=male_pairwise_distances,n=5)

#additional way, with the distance classes based on the inferred distance among relatives from analysis
female_correlogram_setdistances<-mantel.correlog(D.eco=female_relatedness_matrix,D.geo=female_pairwise_distances)

```

```
male_correlogram_setdistances<-mantel.correlog(D.eco=male_relatedness_matrix,D.geo=male_pairwise_distan  
female_correlogram_setdistances  
male_correlogram_setdistances
```

542 ***Analysis iii: spatial autocorrelation***

543 **Ethics**

544 This research is carried out in accordance with permits from the:

- 545 1) US Fish and Wildlife Service (scientific collecting permit number MB76700A-0,1,2)
- 546 2) US Geological Survey Bird Banding Laboratory (federal bird banding permit number 23872)
- 547 3) Arizona Game and Fish Department (scientific collecting license number SP594338 [2017], SP606267
548 [2018], and SP639866 [2019])
- 549 4) Institutional Animal Care and Use Committee at Arizona State University (protocol number 17-1594R)
- 550 5) University of Cambridge ethical review process (non-regulated use of animals in scientific procedures:
551 zoo4/17 [2017])

552 **Author contributions**

553 **Sevchik:** Hypothesis development, sample processing, data analysis and interpretation, write up, revis-
554 ing/editing.

555 **Logan:** Hypothesis development, data analysis and interpretation, write up, revising/editing, materi-
556 als/funding.

557 **McCune:** Blood collection, data analysis and interpretation, revising/editing.

558 **Blackwell:** Hypothesis development, DNA extraction, revising/editing.

559 **Rowney:** Blood collection, DNA extraction, sample processing, write up, revising/editing.

560 **Lukas:** Hypothesis development, data analysis and interpretation, write up, revising/editing, materi-
561 als/funding.

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566 We, the authors, declare that we have no financial conflicts of interest with the content of this article. Corina
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References

- 578
579 Aguillon, Stefanie M, John W Fitzpatrick, Reed Bowman, Stephan J Schoech, Andrew G Clark, Graham
580 Coop, and Nancy Chen. 2017. “Deconstructing Isolation-by-Distance: The Genomic Consequences of
581 Limited Dispersal.” *PLoS Genetics* 13 (8): e1006911.
- 582 Banks, Sam C, and ROD Peakall. 2012. “Genetic Spatial Autocorrelation Can Readily Detect Sex-Biased
583 Dispersal.” *Molecular Ecology* 21 (9): 2092–2105.
- 584 ———. 2012. “Genetic Spatial Autocorrelation Can Readily Detect Sex-Biased Dispersal.” *Molecular Ecology*
585 21 (9): 2092–2105.
- 586 Botero-Delgado, Esteban, Verónica Quirici, Yanina Poblete, Élfego Cuevas, Sylvia Kuhn, Alexander Girg,
587 Kim Teltscher, Elie Poulin, Bart Kempnaers, and Rodrigo A Vásquez. 2017. “Variation in Fine-Scale
588 Genetic Structure and Local Dispersal Patterns Between Peripheral Populations of a South American
589 Passerine Bird.” *Ecology and Evolution* 7 (20): 8363–78.
- 590 Cockburn, Andrew. 2006. “Prevalence of Different Modes of Parental Care in Birds.” *Proceedings of the
591 Royal Society B: Biological Sciences* 273 (1592): 1375–83.
- 592 Coulon, A, J-F Cosson, N Morellet, J-M Angibault, B Cargnelutti, M Galan, S Aulagnier, and AJM Hewison.
593 2006. “Dispersal Is Not Female Biased in a Resource-Defence Mating Ungulate, the European Roe
594 Deer.” *Proceedings of the Royal Society B: Biological Sciences* 273 (1584): 341–48.
- 595 Escobar-Ibáñez, Juan Fernando, Rafael Rueda-Hernández, and Ian MacGregor-Fors. 2020. “The Greener
596 the Better! Avian Communities Across a Neotropical Gradient of Urbanization Density.” *Frontiers in
597 Ecology and Evolution* 8: 285.
- 598 Folsom, MA, M MacPherson, D Lukas, KB McCune, L Bergeron, A Bond, A Blackwell, C Rowney, and CJ
599 Logan. 2020. “Investigating the Rare Behaviour of Male Parental Care in Great-Tailed Grackles.” *PCI
600 Ecology* <http://corinalogan.com/Preregistrations/gmalecare.html>.
- 601 Goudet, Jérôme, Nicolas Perrin, and Peter Waser. 2002. “Tests for Sex-Biased Dispersal Using
602 Bi-Parentally Inherited Genetic Markers.” *Molecular Ecology* 11 (6): 1103–14.
- 603 Gour, Digpal Singh, Jyotsna Bhagavatula, Maradani Bhavanishankar, Patlolla Anuradha Reddy, Jaya A
604 Gupta, Mriganka Shekhar Sarkar, Shaik Mohammed Hussain, Segu Harika, Ravinder Gulia, and Sisinthy
605 Shivaji. 2013. “Philopatry and Dispersal Patterns in Tiger (*Panthera Tigris*.)” *PLoS One* 8 (7): e66956.
- 606 Greenwood, Paul J. 1980. “Mating Systems, Philopatry and Dispersal in Birds and Mammals.” *Animal
607 Behaviour* 28 (4): 1140–62.
- 608 Harrison, Xavier A, Jennifer E York, and Andrew J Young. 2014. “Population Genetic Structure and Direct
609 Observations Reveal Sex-Reversed Patterns of Dispersal in a Cooperative Bird.” *Molecular Ecology* 23
610 (23): 5740–55.
- 611 Hofmann, Sylvia, Peter Fritzsche, Torstein Solhøy, Tsering Dorge, and Georg Miede. 2012. “Evidence of
612 Sex-Biased Dispersal in *Thermophilis Baileyi* Inferred from Microsatellite Markers.” *Herpetologica* 68 (4):
613 514–22.
- 614 Höner, Oliver P, Bettina Wachter, Marion L East, WJ Streich, K Wilhelm, T Burke, and H Hofer. 2007.
615 “Female Mate-Choice Drives the Evolution of Male-Biased Dispersal in a Social Mammal.” *Nature* 448
616 (7155): 798–801.

- 617 Johnson, Kristine, Emily DuVal, Megan Kiehl, and Colin Hughes. 2000. "Male Mating Strategies and the
618 Mating System of Great-Tailed Grackles." *Behavioral Ecology* 11 (2): 132–41.
- 619 Johnson, Kristine, and Brian D Peer. 2001. *Great-Tailed Grackle: Quiscalus Mexicanus*. Birds of North
620 America, Incorporated.
- 621 Jombart, T. 2008. "Adegenet: A R Package for the Multivariate Analysis of Genetic Markers." *Bioinformatics*
622 24: 1403–5. <https://doi.org/10.1093/bioinformatics/btn129>.
- 623 King, Corey. 2012. "Habitat Use and Behavior of Great-Tailed Grackles (*Quiscalus Mexicanus*) in Urban
624 and Peri-Urban Habitats of San Marcos, Hays County, Texas."
- 625 Lawson Handley, LJ, and N Perrin. 2007. "Advances in Our Understanding of Mammalian Sex-Biased
626 Dispersal." *Molecular Ecology* 16 (8): 1559–78.
- 627 Legendre, Pierre, and Loic FJ Legendre. 2012. *Numerical Ecology*. Vol. 24. Elsevier.
- 628 Li, Xiang-Yi, and Hanna Kokko. 2019. "Sex-Biased Dispersal: A Review of the Theory." *Biological Reviews*
629 94 (2): 721–36.
- 630 ———. 2019. "Sex-Biased Dispersal: A Review of the Theory." *Biological Reviews* 94 (2): 721–36.
- 631 Logan, CJ, McCune KB, Breen A, Chen N, and D Lukas. 2020. "Implementing a Rapid Geographic Range
632 Expansion - the Role of Behavior and Habitat Changes." *In Principle Acceptance by PCI Ecology of the*
633 *Version on 6 Oct 2020*. <http://corinalogan.com/Preregistrations/gxpopbehaviorhabitat.html>.
- 634 Lukas, D. 2020. "Individual and Relatedness Data for Great Tailed Grackles from Tempe, AZ, USA 2019."
635 Knowledge Network for Biocomplexity. <https://doi.org/10.5063/F1W66J48>.
- 636 Mabry, Karen E, Erin L Shelley, Katie E Davis, Daniel T Blumstein, and Dirk H Van Vuren. 2013. "Social
637 Mating System and Sex-Biased Dispersal in Mammals and Birds: A Phylogenetic Analysis." *PloS One*
638 8 (3): e57980.
- 639 MacGregor-Fors, Ian, Leopoldo Vázquez, Jorge H Vega-Rivera, and Jorge E Schondube. 2009. "Non-
640 Exotic Invasion of Great-Tailed Grackles *Quiscalus Mexicanus* in a Tropical Dry Forest Reserve." *Ardea*
641 97 (3): 367–69.
- 642 Oksanen, Jari, F Guillaume Blanchet, Roeland Kindt, Pierre Legendre, Peter R Minchin, RB O'hara, Gavin
643 L Simpson, et al. 2013. "Package 'Vegan'." *Community Ecology Package, Version 2* (9): 1–295.
- 644 Paradis, E. 2010. "Pegas: An R Package for Population Genetics with an Integrated–Modular Approach."
645 *Bioinformatics* 26: 419–20.
- 646 Peakall, Rod, Monica Ruibal, and David B Lindenmayer. 2003. "Spatial Autocorrelation Analysis Offers
647 New Insights into Gene Flow in the Australian Bush Rat, *Rattus Fuscipes*." *Evolution* 57 (5): 1182–95.
- 648 Pew, Jack, Paul H Muir, Jinliang Wang, and Timothy R Frasier. 2015. "Related: An R Package for Analysing
649 Pairwise Relatedness from Codominant Molecular Markers." *Molecular Ecology Resources* 15 (3): 557–
650 61.
- 651 Quaglietta, Lorenzo, Vania C Fonseca, Petra Hájková, António Mira, and Luigi Boitani. 2013. "Fine-Scale
652 Population Genetic Structure and Short-Range Sex-Biased Dispersal in a Solitary Carnivore, *Lutra Lutra*."
653 *Journal of Mammalogy* 94 (3): 561–71.

- 654 Queller, David C, and Keith F Goodnight. 1989. "Estimating Relatedness Using Genetic Markers." *Evolution*
655 43 (2): 258–75.
- 656 R Core Team. 2017. *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foun-
657 dation for Statistical Computing. <https://www.R-project.org>.
- 658 Selander, Robert K. 1970. "Parental Feeding in a Male Great-Tailed Grackle." *The Condor* 72 (2): 238–38.
- 659 Sutherland, Glenn D, Alton S Harestad, Karen Price, and Kenneth P Lertzman. 2000. "Scaling of Natal
660 Dispersal Distances in Terrestrial Birds and Mammals." *Conservation Ecology* 4 (1).
- 661 Thrasher, Derrick J, Bronwyn G Butcher, Leonardo Campagna, Michael S Webster, and Irby J Lovette. 2018.
662 "Double-Digest RAD Sequencing Outperforms Microsatellite Loci at Assigning Paternity and Estimating
663 Relatedness: A Proof of Concept in a Highly Promiscuous Bird." *Molecular Ecology Resources* 18 (5):
664 953–65.
- 665 Tourvas, N. 2020. "PopGenUtils: An R Package of Helper Functions for the Analysis of Population Genetics
666 Data." <https://Github.com/Nikostourvas/PopGenUtils/>.
- 667 Trochet, Audrey, Elodie A Courtois, Virginie M Stevens, Michel Baguette, Alexis Chaine, Dirk S Schmeller,
668 Jean Clobert, and John J Wiens. 2016. "Evolution of Sex-Biased Dispersal." *The Quarterly Review of*
669 *Biology* 91 (3): 297–320.
- 670 Walters, Jeffrey R. 2000. "Dispersal Behavior: An Ornithological Frontier." *The Condor* 102 (3): 479–81.
- 671 Wang, Jinliang. 2002. "An Estimator for Pairwise Relatedness Using Molecular Markers." *Genetics* 160 (3):
672 1203–15.
- 673 ———. 2002. "An Estimator for Pairwise Relatedness Using Molecular Markers." *Genetics* 160 (3): 1203–
674 15.
- 675 Wang, Jinliang, and Anna W Santure. 2009. "Parentage and Sibship Inference from Multilocus Genotype
676 Data Under Polygamy." *Genetics* 181 (4): 1579–94.
- 677 Wehtje, Walter. 2003. "The Range Expansion of the Great-Tailed Grackle (*Quiscalus Mexicanus* Gmelin) in
678 North America Since 1880." *Journal of Biogeography* 30 (10): 1593–1607.
- 679 Weinman, Lucia R, Joseph W Solomon, and Dustin R Rubenstein. 2015. "A Comparison of Single Nu-
680 cleotide Polymorphism and Microsatellite Markers for Analysis of Parentage and Kinship in a Coopera-
681 tively Breeding Bird." *Molecular Ecology Resources* 15 (3): 502–11.
- 682 White, P. S., and L. D. Densmore. 1992. "Mitochondrial DNA Isolation." In *Molecular Genetic Analysis of*
683 *Populations: A Practical Approach*, 50–51. IRL Press at Oxford University Press.